

REVIEW

The long non-coding RNA landscape in triple-negative breast cancer

Wenwen Zhang¹  | Xiaoxiang Guan² | Jinhai Tang³

¹Department of Oncology, Nanjing First Hospital, Nanjing Medical University, Nanjing, China

²Department of Oncology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China

³Department of General Surgery, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China

Correspondence

Wenwen Zhang, Department of Oncology, Nanjing First Hospital, Nanjing Medical University, 68 Changle Road, Nanjing 210006, China.
Email: wwzhang1022@hotmail.com

Jinhai Tang, Department of General Surgery, The First Affiliated Hospital of Nanjing Medical University, 300 Guangzhou Road, Nanjing 210029, China.
Email: jhtang@njmu.edu.cn

Funding information

Key International Cooperation of the National Natural Science Foundation of China, Grant/Award Number: 81920108029; Natural Science Foundation of Jiangsu Province, Grant/Award Number: BK20180133; National Natural Science Foundation of China, Grant/Award Number: 81773102 and 81802667

Abstract

Triple-negative breast cancer (TNBC) is a type of breast cancer that has a higher risk of distant recurrence and metastasis, leading to a relatively aggressive biological behaviour and poor outcome. So far, the clinical management of TNBC is challenging because of its heterogeneity and paucity of specific targeted therapy. Recently, various studies have identified a lot of differently expressed long non-coding RNAs (lncRNAs) in TNBC. Those lncRNAs have been reported to play important roles in the multistep process of TNBC tumorigenesis. Here, we review the biological characteristics of lncRNAs, and present the current state of knowledge concerning the expression, function and regulation of lncRNAs in TNBC. Accumulating studies explored the potential lncRNAs-based therapeutics in TNBC, including the techniques of genetic modification using antisense oligonucleotides, locked nucleic acid and RNA nanotechnology. In current review, we also discuss the future prospects of studies about lncRNAs in TNBC and development of lncRNA-based strategies for clinical TNBC patients.

1 | INTRODUCTION

Breast cancer (BC) is the most frequently diagnosed malignancy and the leading cause of cancer death in females worldwide.¹ Triple-negative breast cancer (TNBC) is a subgroup of breast cancers that lack the expression of oestrogen receptor (ER), progesterone receptor (PR) and human epidermal growth factor receptor 2 (HER-2).² The risk of distant recurrence and metastasis in TNBC patients is substantially higher than in non-TNBC patients.³ The

clinical management of TNBC is challenging because of the relatively aggressive biologic behaviour and paucity of specific targeted therapy.⁴ Thus, a better understanding of the regulations and mechanisms of tumorigenesis in TNBC cells and the identification of effective biomarkers for diagnosis and prognosis of TNBC patients are consequently keenly awaited.

lncRNAs, with a length exceeding 200 nucleotides, are non-protein-coding transcripts.⁵ Accumulating studies report that lncRNAs expression is dysregulated in various types of cancer

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Cell Proliferation* Published by John Wiley & Sons Ltd.

including breast cancer, ovarian cancer, hepatocellular carcinoma and many others.⁶⁻¹⁰ Moreover, several lncRNAs have been reported to play crucial roles in various biological processes, including cell proliferation, apoptosis, invasion, differentiation and development.¹¹⁻¹⁴ In TNBC, various studies have identified a lot of dysregulated lncRNAs that play important roles in the process of tumorigenesis through diverse mechanisms. For instance, lncRNAs can act as miRNA 'sponges' and compete miRNA-targeted mRNAs, thereby affecting the miRNA-mediated gene regulation.^{15,16} This competing endogenous RNAs (ceRNA) mechanisms and network construction, by sequestering miRNAs and sparing their protein-coding counterparts from post-translational regulation, have been mainly studied to act as the main molecular mechanism of lncRNA biological function.¹⁵ Some lncRNAs were reported to assemble with mRNAs to protect them from miRNA action and increase their stability. Some lncRNAs are named scaffold lncRNAs, which could serve as a central platform to assemble with different molecular components such as proteins and RNAs and promote their intermolecular interactions. Moreover, signal lncRNAs have also been reported to interact with transcription factors (TFs) or histone-modifying enzymes to *cis*-regulate or *trans*-regulate the expression of their target genes.⁸ Thus, lncRNAs promise potential diagnostic and prognostic biomarkers, therapeutic targets and improve the clinical benefits for TNBC patients.

Accumulating studies have explored the potential lncRNAs-based therapeutics in TNBC, including the techniques of genetic modification using antisense oligonucleotides (ASOs), locked nucleic acid (LNA) and RNA nanotechnology. Such as, Jin et al designed eight ASOs targeting lncRNA TROJAN and transfected TNBC cells with ASOs without using any transfection reagents to simulate *in vivo* conditions. They observed that lung metastasis nodules were significantly reduced in anti-TROJAN ASO-treated group than the control group, and the ASO toxicity was limited after detecting the murine blood biochemical indexes.¹⁷ Hu et al reported that treatment with LINK-A LNAs could repress cell proliferation in TNBC cells and increase the sensitivity of mammary gland tumours to immunotherapy.¹⁸ In current review, we accumulated literature to the understanding of lncRNAs biogenesis and function, as well as the latest findings of novel lncRNAs-based therapeutics in TNBC. We also present the current state of knowledge concerning the expression and regulation of lncRNAs in TNBC, and discuss the future development of lncRNA-based strategies for clinical TNBC patients.

2 | BIOLOGICAL CHARACTERISTICS OF LNCRNAS

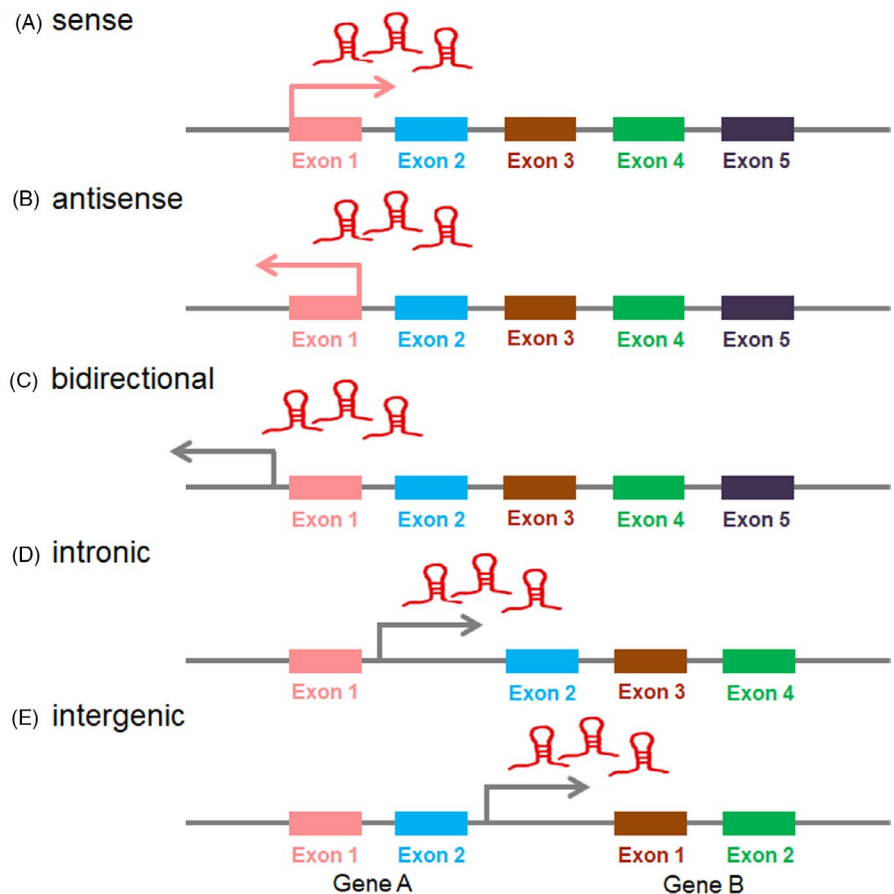
lncRNAs are functionally defined as transcripts >200 nt in length with no protein-coding potential, many of which are uniquely expressed in differentiated tissues or specific cancer types.¹⁹ Distinguishing lncRNAs from other protein-coding mRNAs is not a

trivial process. H19, the first lncRNA reported by Brannan et al in 1990, was just defined as not a classical mRNA, and the product of H19 gene was described to be an RNA molecule.²⁰ In fact, lncRNAs were first described during the large-scale sequencing of full-length cDNA libraries in the mouse.²¹ The number of lncRNAs was reported to outnumber protein-coding genes, and their sequences cover a larger fraction of the human genome.²² lncRNAs may be located within nuclear or cytosolic fractions, and are overlapping with, or interspersed between, multiple coding and non-coding transcripts.^{23,24} Based on their genomic proximity to neighbouring transcripts, they are classified five categories (Figure 1): (a) sense, overlapping one or more exons of a protein-coding gene on the same strand; (b) antisense, overlapping one or more exons of a protein-coding gene on the opposite strand; (c) bidirectional, initiating its expression in close genomic proximity at <1000 base pairs away to a neighbouring coding transcript on the opposite strand; (d) intronic, deriving from an intron of a second transcript; or (e) intergenic, acting as an independent unit within the genomic interval between two genes.^{25,26} lncRNAs were initially thought to be the products of an inconsequential transcription resulting from low RNA polymerase fidelity.²⁷ It is now widely recognized that lncRNAs could identify cellular pathologies such as cancer, provide prognostic value, or even inform therapeutic options for cancer patients, by serving as signals of specific cellular states or readouts of active cellular programmes.²⁸ Recent studies have shown that lncRNAs can regulate gene expression at different levels, including chromatin modification, transcription and post-transcriptional regulation.²⁹ lncRNAs were reported to regulate several biological processes such as cell proliferation, apoptosis, cell cycle, cell invasion and metastasis, cellular differentiation, chromatin modification and nuclear-cytoplasmic trafficking.³⁰ It has been suggested that the involvement of lncRNAs in human diseases could be far more prevalent than previously known.³¹ Recently, lncRNAs-related studies in cancer increased dramatically and have become one of the hottest topics in RNA biology.

3 | PROFILES OF LNCRNAS EXPRESSION IN TNBC

Recently, abnormal expression of many lncRNAs has been found in almost all tumours in humans, including TNBC. However, our understanding of lncRNAs remains significantly less mature than mRNAs, or even miRNAs. Next generation sequencing (NGS) is a DNA sequencing technology, which could perform sequencing of millions of small fragments of DNA in parallel. These fragments are then pieced together by mapping the individual reads to the human reference genome.³² NGS is now used to sequence entire genomes or constrained to specific areas of interest to get the population genomic and gene expression differences in a large array of organisms.³² Thus, NGS technologies may help researchers to accelerate the identification and characterization of important, yet-to-be-annotated functional lncRNAs in TNBC.

FIGURE 1 Classification of lncRNAs based on their genomic proximity to neighbouring transcripts



In recent years, researchers got a lot of abnormally expressed lncRNAs in TNBC patients or cells using public databases based on the NGS technologies. Tian et al found a total of 1034 dysregulated lncRNAs in the two TNBC microarrays from the Gene Expression Omnibus (GEO) database. Among them, 537 lncRNAs were significantly correlated with 451 protein-coding genes, which were mainly enriched in terms including cell division, cell cycle, and involved in PI3K-Akt, MAPK, ErbB family and p53 signalling pathways.³³ In addition, further literatures related to the lncRNA expressions profiles also detected a series of dysregulated lncRNAs in TNBC.³⁴⁻⁴⁰

Previous studies have shown that lncRNAs can act as miRNA 'sponges' and compete miRNA-targeted mRNAs, thereby affecting the miRNA-mediated gene regulation.^{15,16} This crosstalk forms a complex post-transcriptional regulatory network including mRNAs, lncRNAs, called competing endogenous RNAs (ceRNA) network.^{15,16} ceRNA-mediated regulatory mechanisms are reported to be an important pathway for lncRNAs-modulated post-transcriptional regulation in TNBC. Such as, Le et al developed a complex ceRNA network in TNBC using microarray mRNA and lncRNA expression data obtained from The Cancer Genome Atlas (TCGA) database and two GEO databases.⁴¹ As a result, they identified differentially expressed 4565 mRNAs, 427 miRNAs and 4852 lncRNAs, and constructed ceRNA network using 37 lncRNAs, 28 miRNAs and 16 mRNAs. On the basis of establishing the ceRNA network, they found that two mRNAs expression are correlated with prognosis of TNBC patients.⁴¹ Similarly, Liu et al also

constructed a ceRNA network based on analysis of differentially expressed RNAs between 150 TNBC tissues and 823 non-TNBC tissues downloaded from TCGA database.⁴² They identified 190 differentially expressed lncRNAs, 48 differentially expressed mRNAs and 13 differentially expressed miRNAs in this ceRNA network. They concluded that eight lncRNAs and one mRNA could act as prognostic factors in TNBC, using survival analysis and receiver operating characteristic (ROC) curve creation in the network.⁴² Additionally, they found that lncRNA OSTN-AS1 was primarily related to immunologic function, including immune cell infiltration and immune-related markers co-expression.⁴² Song et al also constructed a ceRNA network of TNBC using TCGA database and revealed 686 mRNAs, 26 miRNAs and 50 lncRNAs as key molecules for high risk of TNBC.⁴³ At the same time, the ceRNA crosstalk network of TNBC constructed by Yuan et al contains 22 hub mRNAs, and 14 key differentially expressed lncRNAs.⁴⁴ Jiang et al developed an integrated ceRNA network signature based on three mRNA (FCGR1A, RSAD2, CHRDL1) and two lncRNA (HIF1A-AS2 and AK124454), using transcriptome microarrays for 33 paired TNBC and adjacent normal breast tissue.⁴⁵ They also found that the prognostic and predictive accuracy of this ceRNA signature was better than clinicopathological parameters to predict tumour recurrence and the benefit of taxane chemotherapy in TNBC.⁴⁵ Taken together, the ceRNA co-regulatory network could help us understand the potential characteristics of biological function and pathological roles of lncRNAs in the development and progression of TNBC.

4 | ROLES OF LNCRNAs IN TNBC

To date, numerous lncRNAs have been identified to dysregulated express and play an important role in the biological function of TNBC, including cellular proliferation, apoptosis, cell cycle, migration, invasion, angiogenesis and drug resistance (Table 1). In this chapter, we will provide an overview of lncRNA biological function in TNBC (Figure 2).

4.1 | lncRNAs involved in the regulation of cell proliferation and apoptosis

Cancer has been considered to be the result of accumulated gene mutations, which led to uncontrolled cell proliferations. And deregulated cell proliferation and inhibition of cell apoptosis lie at the heart of tumour development.⁴⁶ The role of lncRNAs in the regulation of TNBC cell proliferation and apoptosis has also been widely investigated. ceRNA mechanisms and network construction, by sequestering miRNAs and sparing their protein-coding counterparts from post-translational regulation, have been mainly studied to act as the main molecular mechanism of lncRNA biological function.¹⁵ For example, lncRNA GAS5 was reported to promote apoptosis and inhibit proliferation of TNBC cells by targeting miR-196a-5p and miR-378a-5p/SUFU signalling.⁴⁷⁻⁴⁹ LINC00096 promoted cell proliferation by sponging miR-383-5p and regulating RBM3 expression in TNBC.⁵⁰ lncRNA HEIH was shown to regulate cell proliferation and apoptosis through miR-4458/SOCS1 axis in TNBC.⁵¹ lncRNA HCP5 could also promote cell proliferation and inhibit cell apoptosis as a ceRNA to regulate BIRC3 by sponging miR-219a-5p.⁵² LINC00339 promoted cell proliferation and inhibited cell apoptosis through miR-377-3p/HOXC6 signalling pathway.⁵³ Knockdown of lncRNA HOST2 could inhibit the proliferation of TNBC cells via regulation of the let-7b/CDK6 axis.⁵⁴

There are also other molecular mechanisms of lncRNAs. Some lncRNAs assemble with mRNAs to protect them from miRNA action and increase their stability. Some lncRNAs are named scaffold lncRNAs, which could serve as a central platform to assemble with different molecular components such as proteins and RNAs and promote their intermolecular interactions. Signal lncRNAs have also been reported to interact with transcription factors (TFs) or histone-modifying enzymes to *cis*-regulate or *trans*-regulate the expression of their target genes.⁸ For instance, in TNBC, Tariq et al revealed that LINC02095 promotes breast cancer proliferation by facilitating the expression of oncogenic transcription factor, SOX9.⁵⁵ lncRNA DANCR was reported to bind with RXRA and increase its serine 49/78 phosphorylation, leading to activating PI3K/Akt signalling and TNBC cell proliferation.⁵⁶ Shen et al demonstrated that LINC00152 obviously enhanced NEDD4-1-mediated ubiquitination and degradation of PTEN protein in TNBC.⁵⁷ Meanwhile, Wu et al also revealed that LINC00152 could enhance TNBC tumorigenesis by inactivation of the BRCA1/PTEN through DNA methyltransferases.⁵⁸ Besides, lncRNA PDCD4-AS1 was reported to stabilize

PDCD4 RNA by forming RNA duplex and regulate the interaction between PDCD4 RNA and RNA decay promoting factors such as HuR.⁵⁹

Several studies have indicated that lncRNAs could also play an important role in the TNBC cell proliferation and apoptosis process by regulating other lncRNAs. lncRNA DRHC was shown to inhibit TNBC cells proliferation by down-regulating the expression of lncRNA HOTAIR, while HOTAIR did not affect the expression level of DRHC.⁶⁰ Similarly, lncRNA PTCSC3 overexpression led to down-regulated lncRNA H19 in TNBC cells, while H19 overexpression did not affect PTCSC3 expression.⁶¹ lncRNA NRON overexpression inhibited cancer cell proliferation and down-regulated lncRNA snaR in TNBC, while snaR overexpression did not significantly affect NRON expression.⁶² There are other lncRNAs involved in the process of TNBC cell proliferation and apoptosis, including AWPPH, LUCAT1, HAND2-AS1, POU3F3, MALAT1, ANRIL.⁶³⁻⁶⁸ These lncRNAs could be potential targets for further mechanistic studies to establish their functional role in TNBC cell proliferation and apoptosis.

Cell cycle progression is regulated by cyclin-dependent kinases (CDKs), which are activated by cyclin binding and inhibited by CDK inhibitors.⁶⁹ p27, an inhibitor of CDK, binds not only to the cyclin E/CDK2 complex, but also to the cyclin D/CDK4,6 complexes, involving in the regulation of the cell cycle.^{70,71} lncRNA MIR100HG was reported to inhibit cell arrest in the G₁ phase, through binding to p27 to form RNA-DNA triplex structures at 275-352 nt, 462-557 nt and 2635-2688 nt.⁷² It was showed that lncRNA LUCAT1 plays a key role in cell cycle G₁ arrest by regulating the expression of cyclin D1, CDK4 in clear cell renal cell carcinoma (ccRCC)⁷³ and the expression of p21, p57 in non-small-cell lung cancer (NSCLC).⁷⁴ In TNBC, LUCAT1 was also shown to contribute to accelerate cell cycle progression through modulating miR-5702.⁶⁴ Besides, Wang et al reported that LINC00339 inhibited cell cycle arrest at G₀/G₁ phase by sponging to miR-377-3p and activating miR-377-3p/HOXC6 signalling pathway in TNBC.⁵³ Shin et al revealed that lncRNA NEAT1 conferred oncogenic role by regulating cell cycle progression in TNBC cells.⁷⁵ lncRNA RMST was also shown to induce the block of G₀/G₁ phase in TNBC.⁷⁶ Taken together, researches about lncRNAs in the regulation of cell cycle in TNBC are preliminary (Figure 3). Maybe, lncRNAs profiles to identify the abnormally expressed lncRNAs and further mechanistic studies to investigate the role of lncRNAs in the regulation of cell cycle progression in TNBC are needed.

4.2 | lncRNAs involved in the regulation of cell invasion and metastasis

Metastasis is the major cause of cancer-related deaths. It has been increasingly recognized that lncRNAs play important roles in tumour invasiveness and metastasis. Overexpression of GAS5 was shown to undermine the tumour promotion effect induced by ectopic expression of miR-196a-5p, including cell invasion and FOXO1/PI3K/Akt signal pathway activation.⁴⁹ TROJAN can bind to metastasis-repressing factor, ZMYND8 and increase its

TABLE 1 Identified lncRNAs in TNBC

lncRNAs	Expression	Biological function	Potential Targets	References
GAS5	Down	Inhibit cell proliferation and invasion; Promote cell apoptosis; Inhibit paclitaxel, cisplatin, adriamycin and PI3K/mTOR inhibitor resistance	miR-378a-5p/SUFU, miR-196a-5p	47-49,108
LINC02095	Up	Promote cell proliferation	SOX9	55
HOTAIR	Up	Promote cell migration and invasion; imatinib and lapatinib resistance	miR-148a; LEF1/TCF4	92,93,109
WT1-AS	Down	Inhibit cell migration and invasion	TGF- β 1	96
LINC00096	Up	Promote cell proliferation and invasion	miR-383-5p/RBM3	50
DRHC	Down	Inhibit cell proliferation	HOTAIR	60
HEIH	Up	Promote cell proliferation Inhibit cell apoptosis	miR-4458/SOCS1	51
LUCAT1	Up	Promote cell proliferation, cell cycle progression and metastasis; Inhibit cell apoptosis	miR-5702	64
CCAT1	Up	Promote cell proliferation, migration, and invasion	miR-218/ZFX	77
ASRPS	Down	Inhibit angiogenesis	STAT3	125
HAND2-AS1	Down	Inhibit cell proliferation	RUNX2	65
LINC01133	Up	Promote cell stem cell (CSC)-like phenotypic traits	KLF4	119
LINC01096	Up	Promote cell proliferation, migration, and invasion; Inhibit cell apoptosis	miR-3130-3p	97
PAPAS	Up	Promote cell migration and invasion	miR-34a	83
HCP5	Up	Promote cell proliferation; Inhibit cell apoptosis	miR-219a-5p/BIRC3	52
NRAD1	Up	Promote cell proliferation and CSC-like phenotypic traits	-	122
LINK-A	Up	Promote immunotherapy resistance; AKT inhibitors resistance; glycolysis reprogramming	PI3K/GPCR; Akt; HIF1 α	18,110,127
MIR503HG	Down	Inhibit cell migration and invasion	miR-103/OLFM4	78
AWPPH	Up	Promote cell proliferation; Promote carboplatin resistance	miR-21; FZD7	63,112
PTCSC3	Down	Inhibit cell proliferation	H19	61
NRON	Down	Inhibit cell proliferation	snaR	62
sONE	Down	Inhibit cell proliferation, migration, and invasion	miR-34a/15a/16, let-7a, TP53/c-Myc; NOS3	84,85
NAMPT-AS	Up	Promote cell metastasis	miR-548b-3p/ NAMPT	79
DANCR	Up	Promote cell proliferation and invasion, and CSC-like phenotypic traits	miR-216a-5p; RXRA; EZH2, CD44, ABCG2; Nanog, SOX2, and OCT4	56,98,120
NEAT1	Up	Inhibit cell apoptosis; Promote cell cycle progression; Promote cisplatin and paclitaxel resistance and cancer stemness	-	75
TROJAN	Up	Promote cell proliferation and invasion	ZMYND8	17
POU3F3	Up	Promote cell proliferation; Inhibit cell apoptosis	Caspase-9	66
NEF	Down	Inhibit cell migration and invasion	miR-155	99
ZEB2-AS1	Up	Promote cell proliferation, metastasis and EMT	ZEB2	88
LncKLHDC7B	Down	Inhibit cell migration and invasion; Promote cell apoptosis	KLHDC7B	100
HIF1A-AS2	Up	Promote cell migration and invasion	-	101
LINC00339	Up	Promote cell proliferation; Inhibit cell cycle arrest, apoptosis	miR-377-3p/HOXC6	53
LINC00152	Up	Promote cell proliferation and invasion; Inhibit cell apoptosis	PTEN, BRCA1	57,58

(Continues)

TABLE 1 (Continued)

lncRNAs	Expression	Biological function	Potential Targets	References
AFAP1-AS1	Up	Promote EMT	Wnt/ β -catenin	81
PDCD4-AS1	Down	Inhibit cell proliferation and migration	PDCD4	59
HOST2	Down	Inhibit cell proliferation	let-7b/CDK6	54
BORG	Up	Promote doxorubicin resistance	RPA1	115
PVT1	Up	Promote cell proliferation and migration, EMT	p21, KLF5/ β -catenin	89,90
H19	Up	Promote paclitaxel resistance and CSC-like phenotypic traits	Akt	114,123
TP73-AS1	Up	Promote cell vasculogenic mimicry	miR-490-3p/TWIST1	126
TUG1	Down	Enhance cisplatin sensitivity	miR-197/NLK	113
MIR100HG	Up	Promote cell proliferation, Inhibit cell cycle arrest	p27	72
LINC01638	Up	Promote cell proliferation, metastasis and CSC-like phenotypic traits	c-Myc	121
ARNILA	Down	Promote EMT, invasion and metastasis	miR-204/SOX4	91
LINC-ZNF469-3	Up	Promote cell invasion, stemness properties and lung metastasis	miR-574-5p/ZEB1	87
ROR	Up	Promote cell invasion and metastasis	miR-145/MUC1; miR-145/ARF6	102,103
AIRN	Down	Inhibit cell migration and invasion	Wnt/ β -catenin/mTOR/PI3K	104
MANCR	Up	Promote cell proliferation, Inhibit DNA damage	-	128
RMST	Down	Inhibit cell proliferation, invasion and migration; Promote cell apoptosis, and regulate cell cycle.	-	76
SKA1BC	Up	Promote cell migration and invasion	KAI1	105
ANRIL	Up	Promote cell proliferation, Inhibit cell apoptosis	miR-199a	67
MALAT1	Up	Promote cell proliferation, cell cycle arrest, and invasion; Inhibit cell apoptosis	miR-129-5p; miR-1/Slug; miR-448/KDM5B	68,106,149
SNHG12	Up	Promote cell proliferation and migration, Inhibit cell apoptosis	MMP13	94
HULC	Up	Promote cell metastasis	MMP-2, MMP-9	107
SNAR	Up	Promote cell proliferation, migration and invasion	-	95
LINP1	Up	Promote DNA DSB repair, and radiotherapy resistance	Ku80	117
PCAT6	Up	radiotherapy resistance	miR-185-5p/TPD52	118

degradation through the ubiquitin-proteasome pathway.¹⁷ Han et al reported that lncRNA CCAT1 could promote TNBC cells migration and invasion by suppressing miR-218/ZFX signalling.⁷⁷ In addition, lncRNA MIR503HG was reported to inhibit cell migration and invasion via miR-103/OLFM4 axis in TNBC.⁷⁸ lncRNA NAMPT-AS promoted TNBC cell metastasis and regulated autophagy, through epigenetically regulating NAMPT expression. NAMPT-AS could recruit POU2F2 to activate the transcription of NAMPT, or serve as a ceRNA to rescue NAMPT degradation from miR-548b-3p in TNBC.⁷⁹

Epithelial-mesenchymal transition (EMT) has been involved in carcinogenesis and confers metastatic properties upon cancer cells by enhancing cell mobility, invasion and resistance to apoptotic stimuli.⁸⁰ Zhang et al revealed that AFAP1-AS1 could activate Wnt/ β -catenin pathway to promote tumorigenesis and cell invasion by inducing the expression of c-myc and EMT-related molecules in TNBC.⁸¹ MiR-34a

was reported to implicate in certain EMT-associated signal pathways to repress tumorigenesis, cancer progression and metastasis.⁸² lncRNA PAPAS was shown to promote migration and invasion of TNBC cells by down-regulating miR-34a.⁸³ lncRNA sONE was also reported to repress endothelial nitric oxide synthase (eNOS)-induced nitric oxide (NO) production, regulating TP53 and c-Myc proteins levels and finally altering the levels of a panel of tumour-suppressor miRNAs, including miR-34a, miR-15, miR-16 and let-7a.⁸⁴ Besides, sONE was also shown to inhibit H₂S-induced TNBC cell migration and invasion through activating sONE/NOS3/NO signalling axis.⁸⁵ ZEB1 and ZEB2 belong to the ZEB family transcription factors, which play a pivotal role in the process of EMT.⁸⁶ lncRNA LINC-ZNF469-3 was reported to enhance invasion ability and stemness properties, and promote lung metastasis through miR-574-5p/ZEB1 axis in TNBC.⁸⁷ lncRNA ZEB2-AS1 was shown to promote TNBC cell metastasis by positively regulating ZEB2 expression and activating the EMT via the PI3K/Akt/GSK3 β /ZEB2

FIGURE 2 The biological function of lncRNAs in TNBC

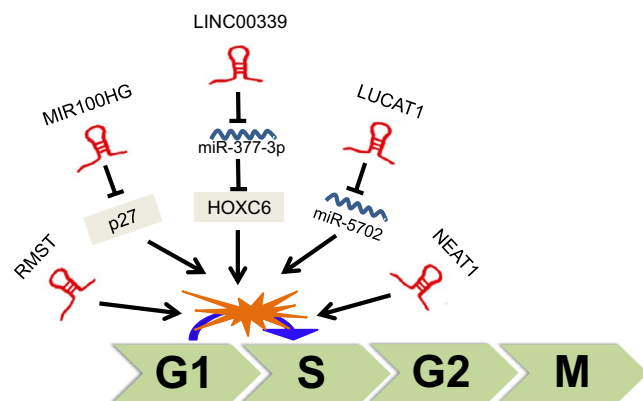
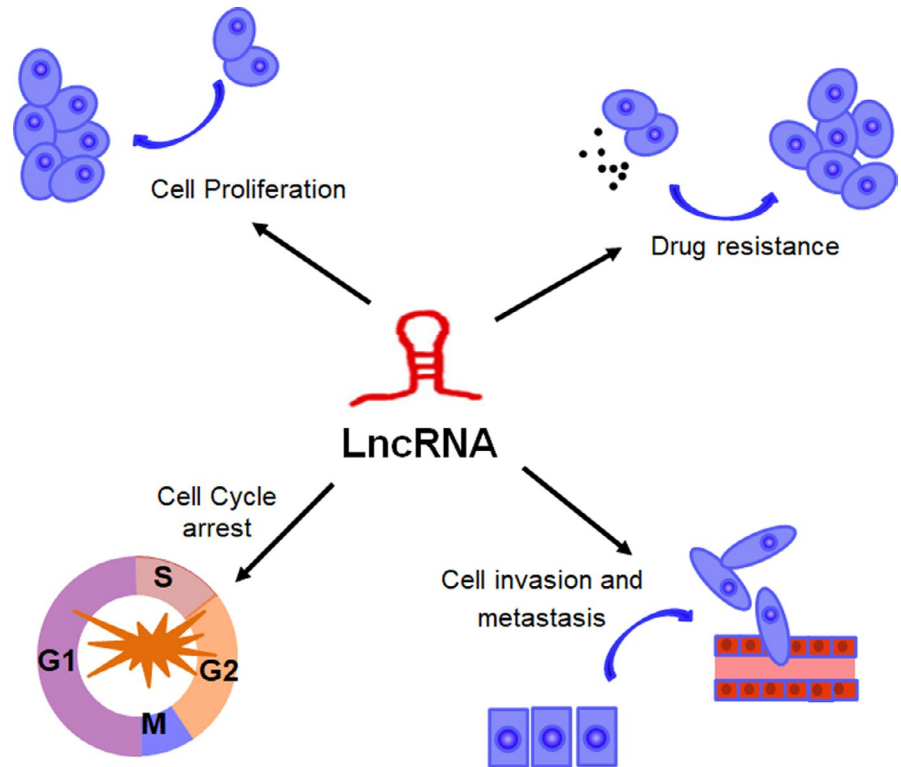


FIGURE 3 lncRNAs involved in the regulation of TNBC cell cycle

signalling pathway.⁸⁸ Another lncRNA, PVT1, was reported to promote EMT and cell migration via regulating p21 and KLF5/ β -catenin signalling in TNBC.^{89,90}

The results of our studies also showed that lncRNA androgen receptor (AR) negatively induced long non-coding RNA (ARNILA) could promote EMT, invasion and metastasis of TNBC, by functioning as a ceRNA for miR-204 to facilitate expression of its target gene Sox4.⁹¹ There are other lncRNAs involved in the process of TNBC invasion and metastasis, including HOTAIR, SNHG12, SNAR, WT1-AS, LINC01096, DANCR, NEF, HIF1A-AS2, LncKLHDC7B, ROR, AIRN, RMST, MALAT1, SKAI1BC, HULC.^{76,92-107} Taken together, these studies revealed that lncRNAs play an important role in the regulation of cell invasion and metastasis in TNBC (Figure 4).

4.3 | lncRNAs involved in the regulation of drug resistance

Emerging evidences suggest that lncRNAs could implicate in regulation of drug resistance by targeting different genes in TNBC (Figure 5). lncRNA growth-stasis-specific transcript 5 (GAS5) is the most widely studied lncRNA involved in the regulation of various drug resistance. The expression level of GAS5 in TNBC patients was reported to associate with tumour resistance to several chemotherapeutic drugs, including adriamycin, paclitaxel and cisplatin.^{47,48} In addition, GAS5 expression could reduce the sensitivity to not only mTORC1 inhibitor rapalogues, but also dual mTORC1/mTORC2 inhibitor AZD8055.¹⁰⁸ Nevertheless, they displayed a significant increase in response to the dual PI3K/mTOR inhibitor, BEZ235.¹⁰⁸ Considering the important role of GAS5 in the sensitivity of multiple drugs, GAS5 may be a potential biomarker for monitoring prognosis of TNBC patients.

Besides, HOTAIR expression was shown to be transcriptionally repressed by the combined treatment of EGFR/HER-2 inhibitor lapatinib plus c-ABL inhibitor imatinib. Enforced expression of HOTAIR conferred increased resistance to the dual treatment by recruitment of β -catenin to the HOTAIR promoter at the LEF1/TCF4-binding site.¹⁰⁹ Another lncRNA, LINK-A (long intergenic non-coding RNA for kinase activation, also called LINC01139), could directly interact with the AKT pleckstrin homology (PH) domain and PIP₃, facilitating AKT-PIP₃ interaction and consequent enzymatic activation.¹¹⁰ LINK-A-dependent AKT hyperactivation led to resistance to AKT inhibitors, while genomic deletions of the LINK-A PIP₃-binding motif dramatically sensitized TNBC cells to AKT inhibitors.¹¹⁰

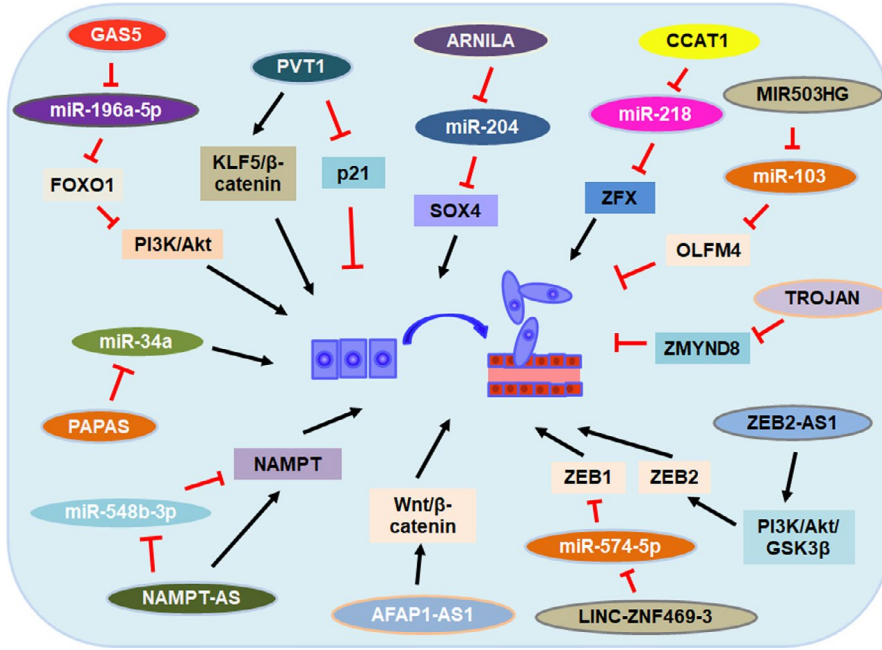


FIGURE 4 lncRNAs involved in the regulation of TNBC cell invasion and metastasis

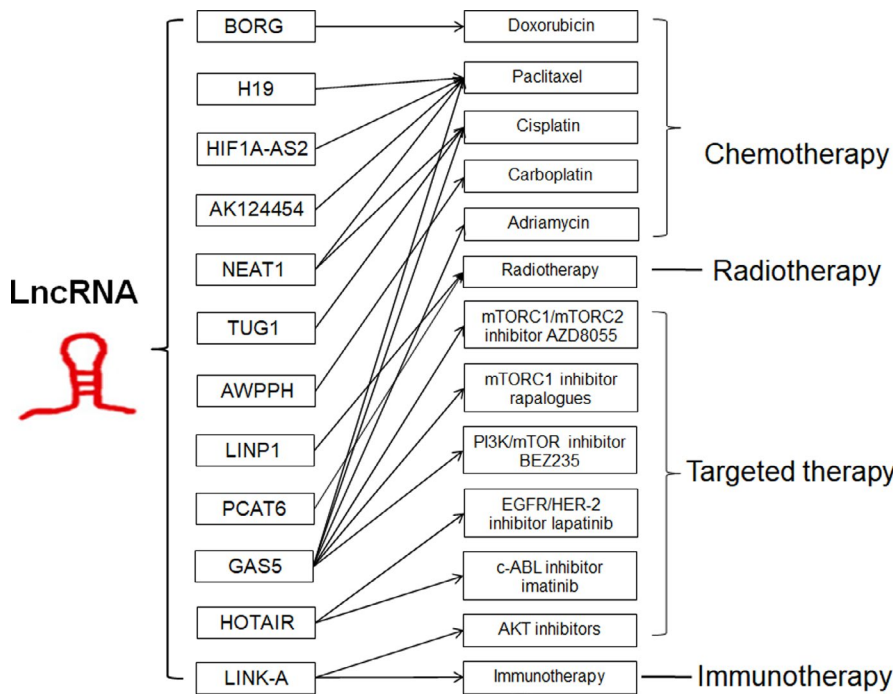


FIGURE 5 lncRNAs involved in the regulation of drug resistance in TNBC

Immunotherapy, including programmed cell death protein-1 and programmed death ligand-1 (PD-1/PD-L1) blockade, has been demonstrated to inhibit cancer progression and validated with the clinical success for the treatment of a variety of human cancers.¹¹¹ Hu and colleagues demonstrated that LINK-A could also regulate the immunosurveillance in TNBC via LINK-A-PKA-TRIM71 signalling axis.¹⁸ Patients with PD-1 blockade-resistant TNBC exhibited elevated LINK-A levels, and LINK-A locked nucleic acids treatment sensitized mammary gland tumours to immune checkpoint blockers.¹⁸

There are other lncRNAs involved in the regulation of drug resistance, including chemotherapy drugs paclitaxel, doxorubicin, cisplatin, carboplatin and radiotherapy resistance. For example,

lncRNA AWPPH could improve cancer cell viability under carboplatin treatment, while lncRNA AWPPH small interfering RNA (siRNA) silencing led to increased chemosensitivity.¹¹² HIF1A-AS2 and AK124454 were showed to contribute paclitaxel resistance in TNBC cells.⁴⁵ lncRNA TUG1 was shown to sponge miR-197, induce expression of NLK and inactivate WNT signalling pathway, thus increasing cisplatin sensitivity of TNBC cells.¹¹³ lncRNA H19 was also reported to confer paclitaxel resistance, while knock-down of H19 might restore the chemosensitivity in paclitaxel-resistant TNBC by mediating the AKT signalling pathway.¹¹⁴ lncRNA NEAT1 was shown to mediate paclitaxel and cisplatin resistance in TNBC.⁷⁵ Besides, lncRNA BORG led to doxorubicin resistance via

binding to RPA1 and activating the NF- κ B signalling axis.¹¹⁵ DNA repair is a series of processes by which damaged DNA is identified and corrected in cells. This process is essential to genomic integrity and is involved in tumorigenesis.¹¹⁶ LncRNA LINP1 was reported to enhance repair of DNA double-strand breaks (DSB) by acting as a scaffold linking Ku80 and DNA-dependent protein kinase catalytic subunit (DNA-PKcs), thereby coordinating the NHEJ pathway, a key determinant of ionizing radiation (IR) resistance.¹¹⁷ Importantly, blocking LINP1 could increase the sensitivity of the tumour-cell response to radiotherapy in TNBC.¹¹⁷ Additionally, knockdown of lncRNA PCAT6 promoted the radiosensitivity of TNBC cells through regulating miR-185-5p/TPD52 axis.¹¹⁸ Taken together, these studies evoke the potential of altering lncRNAs expression in future to represent a novel therapeutic approach to reverse drug resistance or radiotherapy resistance in TNBC patients. However, further studies and mechanistic investigations of the regulation mechanism of lncRNAs-mediated drug resistance in TNBC are needed in the future.

4.4 | Others

Several recent studies also demonstrated that lncRNAs could implicate in other malignant processes, including angiogenesis and cancer stemness. For example, it was reported that mesenchymal stem/stromal cells (MSCs) strongly induced the lncRNA LINC01133 in neighbouring TNBC cells.¹¹⁹ LINC01133 promoted phenotypic and growth characteristics of cancer stem cell-like cells, and that it was a direct mediator of the MSC-triggered miR-199a/FOXP2 pathway and pluripotency-determining gene Kruppel-Like Factor 4 (KLF4) in TNBC models.¹¹⁹ LncRNA DANCR was shown to promote the expression of TNBC cancer stem cell markers through repressing the binding of EZH2 on the promoters of CD44 and ABCG2.¹²⁰ LINC01638 was reported to maintain the mesenchymal traits of TNBC cells, including an enriched EMT signature and cancer stem cell-like state, through interacting with c-Myc to prevent SPOP-mediated c-Myc degradation, and activate MTDH/Twist1 signalling.¹²¹ Furthermore, there are other lncRNAs reported to involve in the regulation of TNBC stemness, including NEAT1, LINC-ZNF469-3, NRAD1, H19.^{75,87,122,123} TNBC patients demonstrate enhanced angiogenesis when compared with non-TNBC patients.¹²⁴ Wang et al recently reported that lncRNA ASRPS could directly bind to STAT3 in the coiled coil domain (CCD) and inhibit STAT3 phosphorylation, leading to reduced expression of VEGF and reduced angiogenesis.¹²⁵ Vasculogenic mimicry (VM), a malignant tumour-specific non-endothelial vascular network, provides oxygen and nutrients to tumour cells and facilitate tumour progression. Tao et al showed that lncRNA TP73-AS1 was upregulated in VM positive TNBC tissues and involved in TNBC VM formation, by binding to miR-490-3p and activating the miR-490-3p/TWIST1 axis.¹²⁶ Besides, lncRNAs are also reported to implicate in other process of TNBC cells. Such as, LINK-A was identified to promote TNBC glycolysis reprogramming by mediating HIF1 α phosphorylation at Tyr 565 and Ser 797.¹²⁷ Tracy

et al revealed that lncRNA MANCR significantly inhibited DNA damage and regulated genomic stability of TNBC.¹²⁸

5 | LNCRNA ACTS AS BIOMARKER FOR DIAGNOSIS AND PROGNOSIS IN TNBC

Since various lncRNAs have been found to be differentially expressed in TNBC, there is increasing evidence to show lncRNAs have diagnostic or prognostic potential for clinical TNBC patients. To study the role of lncRNAs in the diagnosis and prognosis of TNBC patients, numerous researchers analysed the lncRNAs expression levels (even epigenetic level) in TNBC versus non-TNBC patients or healthy controls from tissues specimens, plasma (circulating lncRNA), exosome lncRNA, or micropeptide, and investigated the association with the prognosis of TNBC patients, including overall survival, disease-free survival, lymph node metastasis, and distant metastasis. For instance, Fan et al implemented a comprehensive analysis of lncRNA expression profiles and clinical data of 1097 breast cancer samples from TCGA database. They detected 1510 differentially expressed lncRNAs in normal and TNBC samples, and 672 differentially expressed lncRNAs between non-TNBC and TNBC samples.¹²⁹ They identified three lncRNAs (AC091043.1, AP000924.1 and FOXCUT) maybe have strong diagnostic value for TNBC diagnosis. They also found that other three lncRNAs (AC010343.3, AL354793.1 and FGF10-AS1) expression levels were associated with the clinical prognosis of TNBC patients.¹²⁹ Liu et al compared the differential lncRNAs expression in the plasma of TNBC patients (n=25), non-TNBC patients (n=35) and healthy controls. At last, they found that the expression levels of three lncRNAs, ANRIL, HIF1A-AS2 and UCA1 were significantly increased in the plasma of TNBC patients, suggesting that those three lncRNAs expression may serve as TNBC-specific diagnostic biomarkers.¹³⁰ A recent meta-analysis summarized the prognostic value of 24 lncRNAs from a total of 2803 TNBC patients and demonstrated that expression of nine lncRNAs (SNHG12, MALAT1, HOTAIR, HIF1A-AS2, HULC, LINC00096, ZEB2-AS1, LUCAT1 and LINC000173) showed a marked correlation with positive lymph node metastasis, while lncRNA MIR503HG, GAS5, TCONS_I2_00002973 showed the opposite effect.¹³¹ The authors also found high expression level of another seven lncRNAs (MALAT1, HIF1A-AS2, HULC, LINC00096, ADPGK-AS1, ZEB2-AS1, LUCAT1) was positively correlated with distant metastasis, while patients with a high lncRNA MIR503HG expression level have lower rate of distant metastasis.¹³¹

DNA methylation is the best-studied mechanism of epigenetic gene regulation.¹³² The aberrant DNA methylation statuses play an essential role in the pathological process of many cancers. It was demonstrated that TNBC tumours are genome-wide hypomethylation compared with other subtypes and normal breast control tissues and the hypomethylation is associated with worse overall survival (OS).¹³³⁻¹³⁵ Plenty of evidence have revealed that cancer cells utilize DNA methylation as a strategy to abnormally silence a variety of genes including lncRNAs. Bermejo et al conducted an

epigenome-wide association study (EWAS) and identified that LINC00299 is high methylated in TNBC patients' peripheral blood, making hypermethylation of LINC00299 a useful circulating biomarker for TNBC patients.¹³⁶

LncRNAs were also reported to predict responses to therapy, including chemotherapy, radiotherapy and immunotherapy. One study has showed that circulating lncRNA H19 was high expressed and could predict the response to neoadjuvant chemotherapy (NAC) in TNBC patients.¹³⁷ They found patients with a pathological complete response (pCR) had lower pre-therapeutic levels of lncRNA H19 compared with the non-complete responders. Meanwhile, patients with higher degree of downstaging of initial tumours had lower baseline levels of lncRNA H19 among non-complete responders.¹³⁷ Those data suggested that circulating lncRNA H19 may be a useful marker for predicting the response to neoadjuvant chemotherapy. Another study determined that lncRNA LINK-A could predict immunosuppression and immunotherapy resistance.¹⁸ TNBC patients who responded to pembrolizumab (anti-PD-1 immunotherapy) exhibited relatively lower expression of LINK-A and higher CD8⁺ T-cell infiltration compared with non-responders. CD8⁺ T-cell infiltration in this cohort of patients with TNBC negatively correlated with LINK-A expression.¹⁸ These results implicated the potential for lncRNA LINK-A to serve as biomarker for predicting the outcome of TNBC patients treated with immune checkpoint inhibitors. Recently, Bi et al reported that higher lncAFAP1-AS1 expression was detected in the patients with local recurrence, using the surgically resected tumour tissues of TNBC patients receiving post-operative radiotherapy.¹³⁸ They also found higher lncAFAP1-AS1 expression was correlated with poor disease-free survival and overall survival of TNBC patients.¹³⁸ These results demonstrated

that high lncAFAP1-AS1 expression is associated with radio-resistance of TNBC patients, and the expression level of lncAFAP1-AS1 in tumour tissues could be used to predict the outcome of TNBC radiotherapy.

In conclude, it was noticeable that lncRNAs might be more reliable diagnostic and prognostic biomarkers for TNBC patients as a result of its aberrant expression in tumorigenesis (Figure 6). However, in the future, lncRNA diagnosis and prognosis biomarker studies will need to specify more focus on the serum circulating lncRNA and predicting the response to therapy, including chemotherapy, radiotherapy, targeted therapy and immunotherapy. Additionally, further investigation of a larger patient population is necessary to confirm the diagnostic and prognostic evaluation of lncRNAs in TNBC patients.

6 | LNCRNA-BASED THERAPEUTICS IN TNBC

Since lncRNAs play crucial roles in the biological processes and tumorigenesis and abnormal expression of lncRNAs occur in multiple human cancers, this presents with lncRNA-based therapeutics possibility to correct this dysregulation. Recently, accumulating studies indicating the significance of lncRNAs in the regulation of TNBC development and drug resistance accelerated the investigation to explore the potential lncRNAs-based therapeutics in TNBC (Figure 6).

Antisense oligonucleotides (ASOs) were able to modulate RNA processing and protein expression through different mechanisms, making them able to serve as a variety of molecular targets.¹³⁹ Recently, advancements of ASO structure and chemical

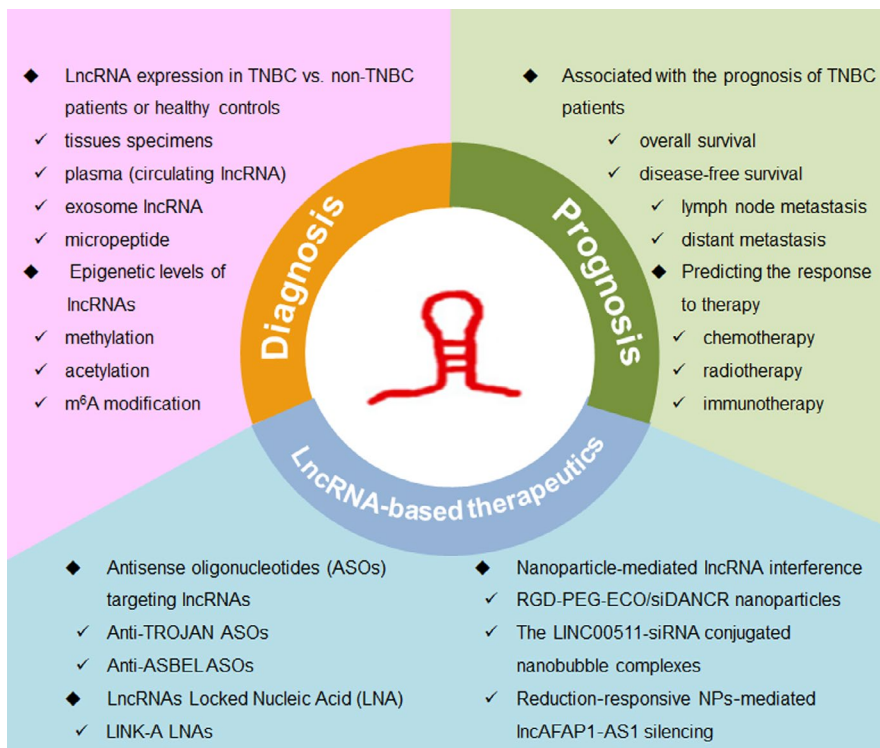


FIGURE 6 lncRNA act as biomarker for diagnosis and prognosis in TNBC

modifications greatly improved the advantage and effectiveness of ASOs to act as precious tools to understand disease mechanisms and as valuable therapeutics for disease intervention.¹⁴⁰ Moreover, many ASOs are undergoing clinical trials, taking advantage of the various mechanisms and synthetic structures now available for the design of ASOs-based therapies.¹⁴⁰ ASOs also were used to inhibit the expression of lncRNAs for lncRNA-based therapeutics in TNBC.

lncRNA TROJAN was shown to promote TNBC proliferation and metastasis, and correlated with poor patient survival.¹⁷ Jin et al designed eight ASOs targeting TROJAN and transfected TNBC cells with ASOs without using any transfection reagents to simulate *in vivo* conditions. After using anti-TROJAN targeted therapy in an intravenous xenograft mouse model, they observed that lung metastasis nodules were significantly reduced in ASO-treated group than the control group.¹⁷ Meanwhile, they also found that the ASO toxicity was limited after detecting the murine blood biochemical indexes.¹⁷ Taken together, these results demonstrated that modification of lncRNA TROJAN by ASO treatment maybe a novel therapeutic approach for TNBC clinical patients.

lncRNA ASBEL has been identified as an antisense transcript of BTG3 gene, which encodes an anti-proliferation protein and is remarkable down-regulated in TNBC. A number of single-stranded modified ASOs were designed, synthesized and screened for specific lncRNA ASBEL knockdown. And anti-ASBEL ASOs were reported to play a significant tumour suppressive role in TNBC by effective down-regulating lncRNA ASBEL.¹⁴¹ Besides, Vidovic et al revealed that ASO targeting lncRNA NRAD1 could reduce tumour growth and inhibit tumour cells to acquire and maintain stem cell characteristics in TNBC.¹²²

Locked Nucleic Acid (LNA) is a novel, third generation DNA analogue that has the potential to impact strongly on the future development of a diversity of nucleic acid-based technologies.¹⁴² lncRNA LINK-A has been characterized as an oncogenic lncRNA in TNBC by activating HIF1 α .¹²⁷ Recently, Hu et al reported that treatment with LINK-A LNAs could repress cell proliferation in TNBC cells, but not non-TNBC cells.¹⁸ They also found that LINK-A LNAs-treated MMTV-Tg mice exhibited inhibited tumour growth and reduced lung metastasis compared with scramble LNAs-treated mice.¹⁸ Besides, treatment with LINK-A LNAs could improve the protein stability of the antigen peptide-loading complex (PLC) components and major histocompatibility complex (MHC) class I complex, resulting in sensitization of mammary gland tumours to immunotherapy. And LINK-A LNAs treatment could improve CD8⁺/CD3⁺ T-cell infiltration and cytotoxicity, while tumour growth was synergistically suppressed by a combinatorial treatment of LINK-A LNAs and immune checkpoint blockers (ICBs).¹⁸ Therefore, LINK-A may act as a powerful biomarker for predicting the prognosis of TNBC patients who received immunotherapy, and targeting LINK-A could further sensitize TNBC to immune checkpoint inhibitors.

RNA nanotechnology is a rapidly evolving field that has emerged as a novel vector system for targeted therapy in various human diseases.¹⁴³⁻¹⁴⁵ RNA nanoparticle-based targeted therapy through inhibition of non-coding RNA has also been reported in the treatment

for human cancer.¹⁴⁶ lncRNA DANCR was reported to be significantly overexpressed and promote cell proliferation, invasion, and CSC-like phenotypic traits in TNBC.^{56,98,120} Vaidya et al formulated tumour-targeting RGD-PEG-ECO/siDANCR nanoparticles via self-assembly of multifunctional amino lipid ECO, cyclic RGD peptide-PEG and siDANCR for systemic delivery.¹⁴⁷ The nanoparticle-mediated RNA interference (RNAi) of the oncogenic lncRNA DANCR demonstrated effective TNBC therapy. They found that DANCR expression was 80%-90% knockdown after treatment with the therapeutic RGD-PEG-ECO/siDANCR nanoparticles in TNBC cells, indicating efficient intracellular siRNA delivery and sustained target silencing. Moreover, the RGD-PEG-ECO/siDANCR nanoparticles mediated significant reduction in TNBC cell proliferation, invasion, migration, survival and tumour spheroid formation, suggesting excellent *in vitro* therapeutic efficacy. Furthermore, the RGD-PEG-ECO/siDANCR nanoparticles TNBC xenografts in nude mouse model also led to suppression of TNBC progression with no overt toxic side-effects, which demonstrated the efficacy and safety of the nanoparticle therapy. Similarly, Wu et al structured a novel theranostic agent loaded with LINC00511-siRNA to deliver siRNA, and detected the responses of drug sensitivity in TNBC.¹⁴⁸ They demonstrated that the combination of low-frequency ultrasound (LFUS) irradiation and nanobubble complexes was regarded as an efficient and safe method for siRNA transfection.¹⁴⁸ Recently, another study engineered a reduction-responsive nanoparticle (NP) platform for effective lncAFAP1-AS1 siRNA (siAFAP1-AS1) delivery, and reported that systemic delivery of siAFAP1-AS1 with the reduction-responsive NPs can synergistically reverse radio-resistance by scavenging intracellular glutathione, leading to a dramatically enhanced radiotherapy effect in both xenograft and metastatic TNBC tumour models.¹³⁸ Overall, these results demonstrate that this RNA nanoparticle-based targeted therapy by nanoparticle-mediated modulation of onco-lncRNAs is a promising approach that utilizes chemically modified RNPs for tumour-specific targeting and lncRNA inhibition that will be beneficial in TNBC and other cancers setting where lncRNA knockdown is desired for a better clinical output.^{138,147,148}

7 | CONCLUSIONS AND FUTURE PROSPECTS

Overall, recent evidences suggest that many lncRNAs were abnormal expressed and characterized as biomarkers for diagnosis and prognosis in TNBC. lncRNAs have been identified to involve in the regulation of pathological and physiological processes of TNBC cells, including cell proliferation, apoptosis, EMT, metastasis and therapy resistance. The functional lncRNAs and their regulators hold the potential for development of novel lncRNA-based therapeutics in clinical TNBC treatment, using ASOs, LNA or RNA nanotechnology targeting lncRNA.

In the future, (a) lncRNA diagnosis and prognosis biomarker studies will need to specify more focus on the serum circulating

lncRNA and predicting the response to therapy, including chemotherapy, radiotherapy, targeted therapy and immunotherapy. (b) high throughput next generation sequencing (NGS) used for lncRNA profiling has identified a lot of differential lncRNAs in TNBC versus non-TNBC tissues. However, further comprehensive functional studies of the identified TNBC-related lncRNAs are needed. (c) Since lncRNAs play important roles in the multiple process of TNBC development, the mechanism of the regulation of abnormally expressed lncRNAs should also need to be investigated in the future, (d) almost all of the lncRNAs-related studies in TNBC are focused on cell lines. Future studies can play more attention in the clinical TNBC patients or animal models. (e) lncRNA-based therapeutics in TNBC have been developed by numerous researchers. However, the technology used for current lncRNA-based therapeutics is focus on antisense oligonucleotides to inhibit the expression of onco-lncRNAs. More technology, such as RNA nanotechnology, may hold the potential for development of novel therapeutics in clinical TNBC treatment. Furthermore, other emerging technology through lncRNA replacement therapy to restore levels of tumour-suppressor lncRNAs could also be developed in the future.

As lncRNAs play significant roles in the TNBC tumorigenesis, further characterization of this category of molecules to uncover their potential roles as therapeutic targets, diagnosis and prognosis biomarkers for TNBC are an important priority for the clinical TNBC treatment.

ACKNOWLEDGEMENTS

This work was supported by National Natural Science Foundation of China (No. 81773102, 81802667), Natural Science Foundation of Jiangsu Province (BK20180133) and Key International Cooperation of the National Natural Science Foundation of China (No. 81920108029).

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

AUTHORS' CONTRIBUTIONS

WZ, XG and JT conceived the study and wrote the manuscript.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Wenwen Zhang  <https://orcid.org/0000-0003-1285-3118>

REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2018;68(6):394-424.
2. Perou CM, Sørlie T, Eisen MB, et al. Molecular portraits of human breast tumours. *Nature*. 2000;406.
3. Rakha EA, El-Sayed ME, Green AR, Lee AH, Robertson JF, Ellis IO. Prognostic markers in triple-negative breast cancer. *Cancer*. 2007;109(1):25-32.
4. Tan AR, Swain SM. Therapeutic strategies for triple-negative breast cancer. *Cancer J*. 2008;14(6):343-351.
5. Batista PJ, Chang HY. Long noncoding RNAs: cellular address codes in development and disease. *Cell*. 2013;152(6):1298-1307.
6. Hansji H, Leung EY, Baguley BC, Finlay GJ, Askarian-Amiri ME. Keeping abreast with long non-coding RNAs in mammary gland development and breast cancer. *Front Genet*. 2014;5:379.
7. Tripathi MK, Doxtater K, Keramatnia F, et al. Role of lncRNAs in ovarian cancer: defining new biomarkers for therapeutic purposes. *Drug Discovery Today*. 2018;23(9):1635-1643.
8. Wong CM, Tsang FH, Ng IO. Non-coding RNAs in hepatocellular carcinoma: molecular functions and pathological implications. *Nat Rev Gastroenterol Hepatol*. 2018;15(3):137-151.
9. Alvarez-Dominguez JR, Lodish HF. Emerging mechanisms of long noncoding RNA function during normal and malignant hematopoiesis. *Blood*. 2017;130(18):1965-1975.
10. Bhan A, Soleimani M, Mandal SS. Long Noncoding RNA and Cancer: A New Paradigm. *Can Res*. 2017;77(15):3965-3981.
11. Evans JR, Feng FY, Chinnaiyan AM. The bright side of dark matter: lncRNAs in cancer. *J Clin Invest*. 2016;126(8):2775-2782.
12. Xiong XD, Ren X, Cai MY, Yang JW, Liu X, Yang JM. Long non-coding RNAs: an emerging powerhouse in the battle between life and death of tumor cells. *Drug Resist Updat*. 2016;26:28-42.
13. Schmitt AM, Chang HY. Long noncoding RNAs in cancer pathways. *Cancer Cell*. 2016;29(4):452-463.
14. Huarte M. The emerging role of lncRNAs in cancer. *Nat Med*. 2015;21(11):1253-1261.
15. Poliseno L, Salmena L, Zhang J, Carver B, Haveman WJ, Pandolfi PP. A coding-independent function of gene and pseudogene mRNAs regulates tumour biology. *Nature*. 2010;465(7301):1033-1038.
16. Cesana M, Cacchiarelli D, Legnini I, et al. A long noncoding RNA controls muscle differentiation by functioning as a competing endogenous RNA. *Cell*. 2011;147(2):358-369.
17. Jin X, Xu XE, Jiang YZ, et al. The endogenous retrovirus-derived long noncoding RNA TROJAN promotes triple-negative breast cancer progression via ZMYND8 degradation. *Sci Adv*. 2019;5(3):eaat9820.
18. Hu Q, Ye Y, Chan LC, et al. Oncogenic lncRNA downregulates cancer cell antigen presentation and intrinsic tumor suppression. *Nat Immunol*. 2019;20(7):835-851.
19. Iyer MK, Niknafs YS, Malik R, et al. The landscape of long noncoding RNAs in the human transcriptome. *Nat Genet*. 2015;47(3):199-208.
20. Brannan CI, Dees EC, Ingram RS, Tilghman SM. The product of the H19 gene may function as an RNA. *Mol Cell Biol*. 1990;10(1):28-36.
21. Okazaki Y, Furuno M, Kasukawa T, et al. Analysis of the mouse transcriptome based on functional annotation of 60,770 full-length cDNAs. *Nature*. 2002;420(6915):563-573.
22. Derrien T, Johnson R, Bussotti G, et al. The GENCODE v7 catalog of human long noncoding RNAs: analysis of their gene structure, evolution, and expression. *Genome Res*. 2012;22(9):1775-1789.
23. Consortium EP, Birney E, Stamatoyannopoulos JA, et al. Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature*. 2007;447(7146):799-816.
24. Carninci P, Kasukawa T, Katayama S, et al. The transcriptional landscape of the mammalian genome. *Science*. 2005;309(5740):1559-1563.
25. Ponting CP, Oliver PL, Reik W. Evolution and functions of long noncoding RNAs. *Cell*. 2009;136(4):629-641.

26. Ma L, Bajic VB, Zhang Z. On the classification of long non-coding RNAs. *RNA Biol.* 2013;10(6):925-933.
27. Louro R, Smirnova AS, Verjovski-Almeida S. Long intronic non-coding RNA transcription: expression noise or expression choice? *Genomics.* 2009;93(4):291-298.
28. Wang KC, Chang HY. Molecular mechanisms of long noncoding RNAs. *Mol Cell.* 2011;43(6):904-914.
29. Mercer TR, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nat Rev Genet.* 2009;10(3):155-159.
30. Wilusz JE, Sunwoo H, Spector DL. Long noncoding RNAs: functional surprises from the RNA world. *Genes Dev.* 2009;23(13):1494-1504.
31. Wapinski O, Chang HY. Long noncoding RNAs and human disease. *Trends Cell Biol.* 2011;21(6):354-361.
32. Hardwick SA, Deveson IW, Mercer TR. Reference standards for next-generation sequencing. *Nat Rev Genet.* 2017;18(8):473-484.
33. Tian T, Gong Z, Wang M, et al. Identification of long non-coding RNA signatures in triple-negative breast cancer. *Cancer Cell Int.* 2018;18:103.
34. Koduru SV, Tiwari AK, Leberfinger A, et al. A comprehensive NGS data analysis of differentially regulated miRNAs, piRNAs, lncRNAs and sn/snoRNAs in triple negative breast cancer. *J Cancer.* 2017;8(4):578-596.
35. Yang F, Liu YH, Dong SY, et al. Co-expression networks revealed potential core lncRNAs in the triple-negative breast cancer. *Gene.* 2016;591(2):471-477.
36. Liu YR, Jiang YZ, Xu XE, et al. Comprehensive transcriptome analysis identifies novel molecular subtypes and subtype-specific RNAs of triple-negative breast cancer. *Breast Cancer Res.* 2016;18(1):33.
37. Lv M, Xu P, Wu Y, et al. LncRNAs as new biomarkers to differentiate triple negative breast cancer from non-triple negative breast cancer. *Oncotarget.* 2016;7(11):13047-13059.
38. Liu YR, Jiang YZ, Xu XE, Hu X, Yu KD, Shao ZM. Comprehensive transcriptome profiling reveals multigene signatures in triple-negative breast cancer. *Clin Cancer Res.* 2016;22(7):1653-1662.
39. Shen X, Xie B, Ma Z, et al. Identification of novel long non-coding RNAs in triple-negative breast cancer. *Oncotarget.* 2015;6(25):21730-21739.
40. Chen C, Li Z, Yang Y, Xiang T, Song W, Liu S. Microarray expression profiling of dysregulated long non-coding RNAs in triple-negative breast cancer. *Cancer Biol Ther.* 2015;16(6):856-865.
41. Le K, Guo H, Zhang Q, et al. Gene and lncRNA co-expression network analysis reveals novel ceRNA network for triple-negative breast cancer. *Sci Rep.* 2019;9(1):15122.
42. Liu Z, Mi M, Li X, Zheng X, Wu G, Zhang L. lncRNA OSTN-AS1 May Represent a Novel Immune-Related Prognostic Marker for Triple-Negative Breast Cancer Based on Integrated Analysis of a ceRNA Network. *Front Genet.* 2019;10:850.
43. Song X, Zhang C, Liu Z, Liu Q, He K, Yu Z. Characterization of ceRNA network to reveal potential prognostic biomarkers in triple-negative breast cancer. *PeerJ.* 2019;7:e7522.
44. Yuan N, Zhang G, Bie F, et al. Integrative analysis of lncRNAs and miRNAs with coding RNAs associated with ceRNA cross-talk network in triple negative breast cancer. *Oncotargets Ther.* 2017;10:5883-5897.
45. Jiang YZ, Liu YR, Xu XE, et al. Transcriptome analysis of triple-negative breast cancer reveals an integrated mRNA-lncRNA signature with predictive and prognostic value. *Can Res.* 2016;76(8):2105-2114.
46. Evan GI, Vousden KH. Proliferation, cell cycle and apoptosis in cancer. *Nature.* 2001;411(6835):342-348.
47. Zheng S, Li M, Miao K, Xu H. lncRNA GAS5-promoted apoptosis in triple-negative breast cancer by targeting miR-378a-5p/SUFU signaling. *J Cell Biochem.* 2019;121(3):2225-2235.
48. Li J, Li L, Yuan H, Huang XW, Xiang T, Dai S. Up-regulated lncRNA GAS5 promotes chemosensitivity and apoptosis of triple-negative breast cancer cells. *Cell Cycle.* 2019;18(16):1965-1975.
49. Li S, Zhou J, Wang Z, Wang P, Gao X, Wang Y. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. *Biomed Pharmacother.* 2018;104:451-457.
50. Tian Y, Xia S, Ma M, Zuo Y. LINC00096 promotes the proliferation and invasion by sponging miR-383-5p and regulating RBM3 expression in triple-negative breast cancer. *Oncotargets Ther.* 2019;12:10569-10578.
51. Li P, Zhou B, Lv Y, Qian Q. lncRNA HEIH regulates cell proliferation and apoptosis through miR-4458/SOCS1 axis in triple-negative breast cancer. *Hum Cell.* 2019;32(4):522-528.
52. Wang L, Luan T, Zhou S, et al. lncRNA HCP5 promotes triple negative breast cancer progression as a ceRNA to regulate BIRC3 by sponging miR-219a-5p. *Cancer Med.* 2019;8(9):4389-4403.
53. Wang X, Chen T, Zhang Y, et al. Long noncoding RNA linc00339 promotes triple-negative breast cancer progression through miR-377-3p/HOXC6 signaling pathway. *J Cell Physiol.* 2019;234(8):13303-13317.
54. Zhang Y, Zhang H, Kang H, Huo W, Zhou Y, Zhang Y. Knockdown of long non-coding RNA HOST2 inhibits the proliferation of triple negative breast cancer via regulation of the let-7b/CDK6 axis. *Int J Mol Med.* 2019;43(2):1049-1057.
55. Tariq A, Hao Q, Sun Q, et al. lncRNA-mediated regulation of SOX9 expression in basal sub-type breast cancer cells. *RNA.* 2019;26:175-185.
56. Tang J, Zhong G, Zhang H, et al. lncRNA DANCR upregulates PI3K/AKT signaling through activating serine phosphorylation of RXRA. *Cell Death Dis.* 2018;9(12):1167.
57. Shen X, Zhong J, Yu P, Zhao Q, Huang T. YY1-regulated LINC00152 promotes triple negative breast cancer progression by affecting on stability of PTEN protein. *Biochem Biophys Res Commun.* 2019;509(2):448-454.
58. Wu J, Shuang Z, Zhao J, et al. linc00152 promotes tumorigenesis by regulating DNMTs in triple-negative breast cancer. *Biomed Pharmacother.* 2018;97:1275-1281.
59. Jadhavi M, Gholamalamdari O, Tang W, et al. A natural antisense lncRNA controls breast cancer progression by promoting tumor suppressor gene mRNA stability. *PLoS Genet.* 2018;14(11):e1007802.
60. Yu F, Wang L, Zhang B. Long non-coding RNA DRHC inhibits the proliferation of cancer cells in triple negative breast cancer by downregulating long non-coding RNA HOTAIR. *Oncol Lett.* 2019;18(4):3817-3822.
61. Wang N, Hou M, Zhan Y, Sheng X. lncRNA PTCSC3 inhibits triple-negative breast cancer cell proliferation by downregulating lncRNA H19. *J Cell Biochem.* 2019;120(9):15083-15088.
62. Niu L, Fan Q, Yan M, Wang L. lncRNA NRON down-regulates lncRNA snaR and inhibits cancer cell proliferation in TNBC. *Biosci Rep.* 2019;39(5):BSR20190468.
63. Wang K, Li X, Song C, Li M. lncRNA AWPPH promotes the growth of triple-negative breast cancer by up-regulating frizzled homolog 7 (FZD7). *Biosci Rep.* 2018;38(6):BSR20181223.
64. Mou E, Wang H. lncRNA LUCAT1 facilitates tumorigenesis and metastasis of triple-negative breast cancer through modulating miR-5702. *Biosci Rep.* 2019;39(9):BSR20190489.
65. Wei M, Liu L, Wang Z. Long non-coding RNA hsa1 and neural crest derivatives expressed 2-antisense RNA 1 overexpression inhibits the proliferation of cancer cells by reducing RUNX2 expression in triple-negative breast cancer. *Oncol Letters.* 2019;18(6):6775-6780.
66. Yang J, Meng X, Yu Y, Pan L, Zheng Q, Lin W. lncRNA POU3F3 promotes proliferation and inhibits apoptosis of cancer cells in triple-negative breast cancer by inactivating caspase 9. *Biosci Biotechnol Biochem.* 2019;83(6):1117-1123.

67. Xu ST, Xu JH, Zheng ZR, et al. Long non-coding RNA ANRIL promotes carcinogenesis via sponging miR-199a in triple-negative breast cancer. *Biomed Pharmacother.* 2017;96:14-21.
68. Zuo Y, Li Y, Zhou Z, Ma M, Fu K. Long non-coding RNA MALAT1 promotes proliferation and invasion via targeting miR-129-5p in triple-negative breast cancer. *Biomed Pharmacother.* 2017;95:922-928.
69. Sherr CJ, Roberts JM. CDK inhibitors: positive and negative regulators of G1-phase progression. *Genes Dev.* 1999;13(12):1501-1512.
70. Lee J, Kim SS. The function of p27 KIP1 during tumor development. *Exp Mol Med.* 2009;41(11):765-771.
71. He W, Wang X, Chen L, Guan X. A crosstalk imbalance between p27(Kip1) and its interacting molecules enhances breast carcinogenesis. *Cancer Biother Radiopharm.* 2012;27(7):399-402.
72. Wang S, Ke H, Zhang H, et al. LncRNA MIR100HG promotes cell proliferation in triple-negative breast cancer through triplex formation with p27 loci. *Cell Death Dis.* 2018;9(8):805.
73. Zheng Z, Zhao F, Zhu D, et al. Long non-coding RNA LUCAT1 promotes proliferation and invasion in clear cell renal cell carcinoma through AKT/GSK-3beta signaling pathway. *Cell Physiol Biochem.* 2018;48(3):891-904.
74. Sun Y, Jin SD, Zhu Q, et al. Long non-coding RNA LUCAT1 is associated with poor prognosis in human non-small lung cancer and regulates cell proliferation via epigenetically repressing p21 and p57 expression. *Oncotarget.* 2017;8(17):28297-28311.
75. Shin VY, Chen J, Cheuk IW, et al. Long non-coding RNA NEAT1 confers oncogenic role in triple-negative breast cancer through modulating chemoresistance and cancer stemness. *Cell Death Dis.* 2019;10(4):270.
76. Wang L, Liu D, Wu X, et al. Long non-coding RNA (LncRNA) RMST in triple-negative breast cancer (TNBC): expression analysis and biological roles research. *J Cell Physiol.* 2018;233(10):6603-6612.
77. Han C, Li X, Fan Q, Liu G, Yin J. CCAT1 promotes triple-negative breast cancer progression by suppressing miR-218/ZFX signaling. *Aging.* 2019;11(14):4858-4875.
78. Fu J, Dong G, Shi H, et al. LncRNA MIR503HG inhibits cell migration and invasion via miR-103/OLFM4 axis in triple negative breast cancer. *J Cell Mol Med.* 2019;23(7):4738-4745.
79. Zhang H, Zhang N, Liu Y, et al. Epigenetic regulation of NAMPT by NAMPT-AS drives metastatic progression in triple-negative breast cancer. *Can Res.* 2019;79(13):3347-3359.
80. Mittal V. Epithelial mesenchymal transition in tumor metastasis. *Annu Rev Pathol.* 2018;13:395-412.
81. Zhang K, Liu P, Tang H, et al. AFAP1-AS1 promotes epithelial-mesenchymal transition and tumorigenesis through wnt/beta-catenin signaling pathway in triple-negative breast cancer. *Front Pharmacol.* 2018;9:1248.
82. Nie D, Fu J, Chen H, Cheng J, Fu J. Roles of microRNA-34a in epithelial to mesenchymal transition, competing endogenous RNA sponging and its therapeutic potential. *Int J Mol Sci.* 2019;20(4):861.
83. Kong Y, Geng C, Dong Q. LncRNA PAPAS may promote triple-negative breast cancer by downregulating miR-34a. *J Int Med Res.* 2019;47(8):3709-3718.
84. Youness RA, Hafez HM, Khallaf E, Assal RA, Abdel Motaal A, Gad MZ. The long noncoding RNA sONE represses triple-negative breast cancer aggressiveness through inducing the expression of miR-34a, miR-15a, miR-16, and let-7a. *J Cell Physiol.* 2019;234(11):20286-20297.
85. Youness RA, Assal RA, Abdel Motaal A, Gad MZ. A novel role of sONE/NOS3/NO signaling cascade in mediating hydrogen sulphide bilateral effects on triple negative breast cancer progression. *Nitric Oxide.* 2018;80:12-23.
86. Fardi M, Alivand M, Baradaran B, Farshdousti Hagh M, Solali S. The crucial role of ZEB2: from development to epithelial-to-mesenchymal transition and cancer complexity. *J Cell Physiol.* 2019;234(9):14783-14799.
87. Wang PS, Chou CH, Lin CH, et al. A novel long non-coding RNA linc-ZNF469-3 promotes lung metastasis through miR-574-5p-ZEB1 axis in triple negative breast cancer. *Oncogene.* 2018;37(34):4662-4678.
88. Zhang G, Li H, Sun R, et al. Long non-coding RNA ZEB2-AS1 promotes the proliferation, metastasis and epithelial mesenchymal transition in triple-negative breast cancer by epigenetically activating ZEB2. *J Cell Mol Med.* 2019;23(5):3271-3279.
89. Tang J, Li Y, Sang Y, et al. LncRNA PVT1 regulates triple-negative breast cancer through KLF5/beta-catenin signaling. *Oncogene.* 2018;37(34):4723-4734.
90. Wang L, Wang R, Ye Z, et al. PVT1 affects EMT and cell proliferation and migration via regulating p21 in triple-negative breast cancer cells cultured with mature adipogenic medium. *Acta Biochim Biophys Sin (Shanghai).* 2018;50(12):1211-1218.
91. Yang F, Shen Y, Zhang W, et al. An androgen receptor negatively induced long non-coding RNA ARNILA binding to miR-204 promotes the invasion and metastasis of triple-negative breast cancer. *Cell Death Differ.* 2018;25(12):2209-2220.
92. Liang H, Huang W, Wang Y, Ding L, Zeng L. Overexpression of MiR-146a-5p upregulates lncRNA HOTAIR in triple-negative breast cancer cells and predicts poor prognosis. *Technol Cancer Res Treat.* 2019;18:1533033819882949.
93. Tao S, He H, Chen Q. Estradiol induces HOTAIR levels via GPER-mediated miR-148a inhibition in breast cancer. *J Transl Med.* 2015;13:131.
94. Wang O, Yang F, Liu Y, et al. C-MYC-induced upregulation of lncRNA SNHG12 regulates cell proliferation, apoptosis and migration in triple-negative breast cancer. *Am J Transl Res.* 2017;9(2):533-545.
95. Lee J, Jung JH, Chae YS, et al. Long noncoding RNA snaR regulates proliferation, migration and invasion of triple-negative breast cancer cells. *Anticancer Res.* 2016;36(12):6289-6295.
96. Wang J, Xi C, Yang X, et al. LncRNA WT1-AS inhibits triple-negative breast cancer cell migration and invasion by downregulating transforming growth factor beta1. *Cancer Biother Radiopharm.* 2019;34(10):671-675.
97. Wang GP, Mou ZL, Xu YY, Liu GX, Wang DM, Zhang HP. LINC01096 knockdown inhibits progression of triple-negative breast cancer by increasing miR-3130-3p. *Eur Rev Med Pharmacol Sci.* 2019;23(17):7445-7456.
98. Tao W, Wang C, Zhu B, Zhang G, Pang D. LncRNA DANCR contributes to tumor progression via targeting miR-216a-5p in breast cancer: lncRNA DANCR contributes to tumor progression. *Biosci Rep.* 2019;39(4).
99. Song X, Liu Z, Yu Z. LncRNA NEF is downregulated in triple negative breast cancer and correlated with poor prognosis. *Acta Biochim Biophys Sin (Shanghai).* 2019;51(4):386-392.
100. Beltran-Anaya FO, Romero-Cordoba S, Rebollar-Vega R, et al. Expression of long non-coding RNA ENSG00000226738 (LncKLHDC7B) is enriched in the immunomodulatory triple-negative breast cancer subtype and its alteration promotes cell migration, invasion, and resistance to cell death. *Mol Oncol.* 2019;13(4):909-927.
101. Wang Y, Zhang G, Han J. HIF1A-AS2 predicts poor prognosis and regulates cell migration and invasion in triple-negative breast cancer. *J Cell Biochem.* 2019;120(6):10513-10518.
102. Ma J, Yang Y, Huo D, et al. LincRNA-RoR/miR-145 promote invasion and metastasis in triple-negative breast cancer via targeting MUC1. *Biochem Biophys Res Commun.* 2018;500(3):614-620.
103. Eades G, Wolfson B, Zhang Y, Li Q, Yao Y, Zhou Q. lincRNA-RoR and miR-145 regulate invasion in triple-negative breast cancer via targeting ARF6. *Mol Cancer Res.* 2015;13(2):330-338.

104. Liu L, Yu D, Shi H, Li J, Meng L. Reduced lncRNA Aim enhances the malignant invasion of triple-negative breast cancer cells mainly by activating Wnt/beta-catenin/mTOR/PI3K signaling. *Pharmazie*. 2017;72(10):599-603.
105. Aram R, Dotan I, Hotz-Wagenblatt A, Canaani D. Identification of a novel metastasis inducing lncRNA which suppresses the KAI1/CD82 metastasis suppressor gene and is up-regulated in triple-negative breast cancer. *Oncotarget*. 2017;8(40):67538-67552.
106. Jin C, Yan B, Lu Q, Lin Y, Ma L. Reciprocal regulation of Hsa-miR-1 and long noncoding RNA MALAT1 promotes triple-negative breast cancer development. *Tumour Biol*. 2016;37(6):7383-7394.
107. Shi F, Xiao F, Ding P, Qin H, Huang R. Long noncoding RNA highly up-regulated in liver cancer predicts unfavorable outcome and regulates metastasis by MMPs in triple-negative breast cancer. *Arch Med Res*. 2016;47(6):446-453.
108. Pickard MR, Williams GT. Regulation of apoptosis by long non-coding RNA GAS5 in breast cancer cells: implications for chemotherapy. *Breast Cancer Res Treat*. 2014;145(2):359-370.
109. Wang YL, Overstreet AM, Chen MS, et al. Combined inhibition of EGFR and c-ABL suppresses the growth of triple-negative breast cancer growth through inhibition of HOTAIR. *Oncotarget*. 2015;6(13):11150-11161.
110. Lin A, Hu Q, Li C, et al. The LINK-A lncRNA interacts with PtdIns(3,4,5)P3 to hyperactivate AKT and confer resistance to AKT inhibitors. *Nat Cell Biol*. 2017;19(3):238-251.
111. Chen L, Han X. Anti-PD-1/PD-L1 therapy of human cancer: past, present, and future. *J Clin Investig*. 2015;125(9):3384-3391.
112. Liu AN, Qu HJ, Gong WJ, Xiang JY, Yang MM, Zhang W. lncRNA AWPPH and miRNA-21 regulates cancer cell proliferation and chemosensitivity in triple-negative breast cancer by interacting with each other. *J Cell Biochem*. 2019;120(9):14860-14866.
113. Tang T, Cheng Y, She Q, et al. Long non-coding RNA TUG1 sponges miR-197 to enhance cisplatin sensitivity in triple negative breast cancer. *Biomed Pharmacother*. 2018;107:338-346.
114. Han J, Han B, Wu X, et al. Knockdown of lncRNA H19 restores chemo-sensitivity in paclitaxel-resistant triple-negative breast cancer through triggering apoptosis and regulating Akt signaling pathway. *Toxicol Appl Pharmacol*. 2018;359:55-61.
115. Gooding AJ, Zhang B, Gunawardane L, Beard A, Valadkhan S, Schiemann WP. The lncRNA BORG facilitates the survival and chemoresistance of triple-negative breast cancers. *Oncogene*. 2019;38(12):2020-2041.
116. Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature*. 2012;481(7381):287-294.
117. Zhang Y, He Q, Hu Z, et al. Long noncoding RNA LINP1 regulates repair of DNA double-strand breaks in triple-negative breast cancer. *Nat Struct Mol Biol*. 2016;23(6):522-530.
118. Shi R, Wu P, Liu M, Chen B, Cong L. Knockdown of lncRNA PCAT6 enhances radiosensitivity in triple-negative breast cancer cells by regulating miR-185-5p/TPD52 axis. *OncoTargets Ther*. 2020;13:3025-3037.
119. Tu Z, Schmollerl J, Cuiffo BG, Karnoub AE. Microenvironmental regulation of long noncoding RNA LINC01133 promotes cancer stem cell-like phenotypic traits in triple-negative breast cancers. *Stem Cells*. 2019;37(10):1281-1292.
120. Sha S, Yuan D, Liu Y, Han B, Zhong N. Targeting long non-coding RNA DANCR inhibits triple negative breast cancer progression. *Biol Open*. 2017;6(9):1310-1316.
121. Luo L, Tang H, Ling L, et al. LINC01638 lncRNA activates MTDH-Twist1 signaling by preventing SPOP-mediated c-Myc degradation in triple-negative breast cancer. *Oncogene*. 2018;37(47):6166-6179.
122. Vidovic D, Huynh TT, Konda P, et al. ALDH1A3-regulated long non-coding RNA NRAD1 is a potential novel target for triple-negative breast tumors and cancer stem cells. *Cell Death Differ*. 2020;27(1):363-378.
123. Shima H, Kida K, Adachi S, et al. lnc RNA H19 is associated with poor prognosis in breast cancer patients and promotes cancer stemness. *Breast Cancer Res Treat*. 2018;170(3):507-516.
124. Ribatti D, Nico B, Ruggieri S, Tamma R, Simone G, Mangia A. Angiogenesis and antiangiogenesis in triple-negative breast cancer. *Transl Oncol*. 2016;9(5):453-457.
125. Wang Y, Wu S, Zhu X, et al. lncRNA-encoded polypeptide ASRPS inhibits triple-negative breast cancer angiogenesis. *J Exp Med*. 2020;217(3).
126. Tao W, Sun W, Zhu H, Zhang J. Knockdown of long non-coding RNA TP73-AS1 suppresses triple negative breast cancer cell vasculogenic mimicry by targeting miR-490-3p/TWIST1 axis. *Biochem Biophys Res Commun*. 2018;504(4):629-634.
127. Lin A, Li C, Xing Z, et al. The LINK-A lncRNA activates normoxic HIF1alpha signalling in triple-negative breast cancer. *Nat Cell Biol*. 2016;18(2):213-224.
128. Tracy KM, Tye CE, Ghule PN, et al. Mitotically-associated lncRNA (MANCR) affects genomic stability and cell division in aggressive breast cancer. *Mol Cancer Res*. 2018;16(4):587-598.
129. Fan CN, Ma L, Liu N. Comprehensive analysis of novel three-long noncoding RNA signatures as a diagnostic and prognostic biomarkers of human triple-negative breast cancer. *J Cell Biochem*. 2019;120(3):3185-3196.
130. Liu M, Xing LQ, Liu YJ. A three-long noncoding RNA signature as a diagnostic biomarker for differentiating between triple-negative and non-triple-negative breast cancers. *Medicine*. 2017;96(9):e6222.
131. Zhang S, Ma F, Xie X, Shen Y. Prognostic value of long non-coding RNAs in triple negative breast cancer: a PRISMA-compliant meta-analysis. *Medicine*. 2020;99(37):e21861.
132. Jones PA. Functions of DNA methylation: islands, start sites, gene bodies and beyond. *Nat Rev Genet*. 2012;13(7):484-492.
133. Yu J, Zayas J, Qin B, Wang L. Targeting DNA methylation for treating triple-negative breast cancer. *Pharmacogenomics*. 2019;20(16):1151-1157.
134. Cancer Genome Atlas N. Comprehensive molecular portraits of human breast tumours. *Nature*. 2012;490(7418):61-70.
135. Fang F, Turcan S, Rimmer A, et al. Breast cancer methylomes establish an epigenomic foundation for metastasis. *Sci Transl Med*. 2011;3(75):75ra25.
136. Bermejo JL, Huang G, Manoochchri M, et al. Long intergenic non-coding RNA 299 methylation in peripheral blood is a biomarker for triple-negative breast cancer. *Epigenomics*. 2019;11(1):81-93.
137. Ozgur E, Ferhatoglu F, Sen F, Saip P, Gezer U. Circulating lncRNA H19 may be a useful marker of response to neoadjuvant chemotherapy in breast cancer. *Cancer Biomark*. 2019;27:11-17.
138. Bi Z, Li Q, Dinglin X, et al. Nanoparticles (NPs)-mediated lncRNA AFAP1-AS1 silencing to block Wnt/beta-catenin signaling pathway for synergistic reversal of radioresistance and effective cancer radiotherapy. *Adv Sci (Weinh)*. 2020;7(18):2000915.
139. Rinaldi C, Wood MJA. Antisense oligonucleotides: the next frontier for treatment of neurological disorders. *Nat Rev Neurol*. 2018;14(1):9-21.
140. Schoch KM, Miller TM. Antisense oligonucleotides: translation from mouse models to human neurodegenerative diseases. *Neuron*. 2017;94(6):1056-1070.
141. Xia Y, Xiao X, Deng X, et al. Targeting long non-coding RNA ASBEL with oligonucleotide antagonist for breast cancer therapy. *Biochem Biophys Res Commun*. 2017;489(4):386-392.
142. Grunweller A, Hartmann RK. Locked nucleic acid oligonucleotides: the next generation of antisense agents? *BioDrugs*. 2007;21(4):235-243.

143. Shu Y, Pi F, Sharma A, et al. Stable RNA nanoparticles as potential new generation drugs for cancer therapy. *Adv Drug Deliv Rev.* 2014;66:74-89.
144. Guo P. The emerging field of RNA nanotechnology. *Nat Nanotechnol.* 2010;5(12):833-842.
145. Guo P, Haque F, Hallahan B, Reif R, Li H. Uniqueness, advantages, challenges, solutions, and perspectives in therapeutics applying RNA nanotechnology. *Nucleic Acid Ther.* 2012;22(4):226-245.
146. Lee TJ, Yoo JY, Shu D, et al. RNA nanoparticle-based targeted therapy for glioblastoma through inhibition of oncogenic miR-21. *Mol Ther.* 2017;25(7):1544-1555.
147. Vaidya AM, Sun Z, Ayat N, et al. Systemic delivery of tumor-targeting siRNA nanoparticles against an oncogenic lncRNA facilitates effective triple-negative breast cancer therapy. *Bioconjug Chem.* 2019;30(3):907-919.
148. Wu B, Yuan Y, Han X, et al. Structure of LINC00511-siRNA-conjugated nanobubbles and improvement of cisplatin sensitivity on triple negative breast cancer. *FASEB J.* 2020;34(7):9713-9726.
149. Bamodu OA, Huang WC, Lee WH, et al. Aberrant KDM5B expression promotes aggressive breast cancer through MALAT1 overexpression and downregulation of hsa-miR-448. *BMC Cancer.* 2016;16:160.

How to cite this article: Zhang W, Guan X, Tang J. The long non-coding RNA landscape in triple-negative breast cancer. *Cell Prolif.* 2021;54:e12966. <https://doi.org/10.1111/cpr.12966>