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## Targeting GPCR Signaling for Idiopathic Pulmonary Fibrosis Therapies

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### Abstract

A variety of G-protein coupled receptors (GPCRs) have been implicated in the pathogenesis of pulmonary fibrosis, largely through their promotion of profibrotic fibroblast activation. In contrast, recent work has highlighted the beneficial effects G $\alpha$ s-coupled GPCRs exert on reducing fibroblast activation and fibrosis. This review highlights how fibrosis promoting and inhibiting GPCR signaling converges on downstream signaling and transcriptional effectors, and how the diversity and dynamics of GPCR expression challenge efforts to identify effective therapies for IPF. Next generation strategies to overcome these challenges, focusing on target selection, polypharmacology and personalized medicine approaches, are discussed as a path toward more effective GPCR-targeted therapies for pulmonary fibrosis.

### Keywords

G-protein coupled receptor; ROCK; YAP; TAZ; MRTF; fibrosis

### GPCRs as targets for pulmonary fibrosis

**G-protein Coupled Receptors** (GPCRs)(see Glossary) are a class of over 800 receptors, making up one of the largest and most diverse families of proteins in the genome[1]. The basic function of GPCRs is to communicate extracellular cues into intracellular signals. The transduction of extracellular stimuli is mediated by the interaction of each receptor with one or more of four major unique **G-protein** families: G $\alpha$ i/o, G $\alpha$ q/11, G $\alpha$ 12/13, and G $\alpha$ s [2, 3] (Fig. 1). Physiological appropriate responses are ensured by coordinated cell-specific expression of receptors [4, 5] and presentation of their endogenous ligands. Their diverse expression, along with their well-defined binding pockets and cell membrane expression, have made them frequent targets for therapeutic development [6], accounting for ~35% of all FDA approved drugs [7]. Multiple prior and ongoing drug discovery campaigns and clinical trial efforts have targeted GPCRs for treatment of **idiopathic pulmonary fibrosis** (IPF) (Table 1).

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IPF is a progressive chronic lung disorder characterized by uncontrolled deposition of fibrous connective tissue, notably collagen I, predominantly affecting the alveolar interstitium where it results in a replacement of healthy gas exchange tissue with fibrotic scar, leading to respiratory failure and eventual death [8]. IPF affects ~3 million people worldwide with a median life expectancy after diagnosis of 2–4 years and an increasing annual incidence of 3 to 18 cases per 100,000 people in the United States and Europe [8, 9]. **Nintedanib** and **Pirfenidone** were approved by the FDA in 2014 as the first pharmaceutical treatments for IPF based on their ability to reduce the decline of pulmonary function [10, 11] however both drugs show only a modest improvement in mortality [11]. The prevailing hypothesis for the initiation and propagation of pulmonary fibrosis is that repeated **epithelial** insult and impaired repair leads to recruitment or activation of **extracellular matrix** (ECM) depositing **fibroblasts** [12–14], potentially supported by additional disease-associated cells such as macrophages [15–17]. This can be modeled experimentally by introducing an epithelial injury in mice by a onetime intratracheal administration of bleomycin which promotes inflammation, injury, and lung fibrosis [18]. Although fibroblasts are transiently activated following tissue injury, in pulmonary fibrosis they maintain an active state, driven by **profibrotic** soluble factors, mechanosignaling, altered metabolism, and cellular aging [14, 19]. This review focuses on the roles of GPCR signaling in progression and resolution of pulmonary fibrosis and as targets for therapeutic intervention. While not a focus of this review, the parallels to GPCR signaling in asthma, another chronic lung disease, are striking, and include similar challenges with receptor redundancy and desensitization [20, 21]. Consideration of the approaches used there, including multi-drug and personalized strategies, may provide a useful path for GPCR targeting in IPF.

### **Gai/o, Gaq/11, Gα12/13 are pro-fibrotic fibroblast regulators**

Multiple GPCR ligand/receptor pairs have been recognized as potential drivers of fibrogenic fibroblast activation in vitro, and fibrosis progression in vivo. The specific ligands implicated in pulmonary fibroblast activation and fibrosis include: endothelin (ET-1) [22–24], lysophosphatidic acid (LPA) [25–29], serotonin (5-HT) [30], sphingosine-1-phosphate (S1P) [31] and angiotensin [32]. Although each ligand can interact with multiple cognate receptor subtypes, specific receptors including LPA receptor 1, endothelin receptor A, serotonin receptors 2A and 2B, and sphingosine-1-phosphate receptor 1 have been identified as primary drivers of ligand-mediated fibrogenic activation of human lung fibroblasts. These receptors are capable of activating Gai/o, Gaq/11, and Gα12/13 subclasses of G-proteins [33]. The remarkable diversity of these upstream GPCRs all capable of engaging very similar patterns of fibroblast activation strongly suggests the presence of convergent downstream mechanisms common to all three receptor families. While a diverse array of signaling and transcriptional programs is activated by these receptors [34], recent work has identified a common intersecting effect on activation of Rho GTPases and actin cytoskeletal assembly [35] (Fig. 1). Together the activation of these pathways promotes nuclear translocation of **myocardin-related transcription factors** (MRTF-A/B) and the Hippo pathway effectors **yes-associated protein** (YAP) and **transcriptional coactivator with PDZ-binding motif** (TAZ) [36, 37], both of which are essential to the activation of fibroblasts to the contractile and matrix synthetic states that drive fibrosis [38–40]. Interestingly, mechanosensitive signaling through integrin and focal adhesion mediated

pathways also promote fibroblast activation through the same convergent pathways [41]. Genetic and pharmacological approaches have confirmed the central roles for MRTFA/B and YAP/TAZ in experimental pulmonary fibrosis [38–40, 42], highlighting the potential for identifying and targeting specific upstream GPCRs driving these effects. Additionally, MRTFA/B and YAP/TAZ cooperate and crosstalk with SMAD3 downstream of TGF $\beta$  [43, 44], further highlighting their attractiveness as targets for pulmonary fibrosis. Multiple clinical trials have already been conducted or are ongoing to test antagonists of specific GPCRs (Table 1). Below we detail some of the challenges that this approach faces, and later we propose some opportunities for identifying more effective GPCR targeted therapies for IPF.

### G $\alpha$ s is an anti-fibrotic fibroblast regulator

Receptors that couple to G $\alpha$ s activate adenylyl cyclase, elevating intracellular levels of **cyclic adenosine monophosphate** (cAMP). Through the use of the cAMP enhancing pharmacologic tool forskolin, elevated cAMP was identified as a means to block **transforming growth factor beta** (TGF $\beta$ ) induced fibroblast activation [45]. Later it was recognized that elevation of cAMP through G $\alpha$ s coupled GPCRs inhibits fibroblast proliferation, expression of ECM proteins, and cellular contractility [46, 47]. Downstream, cAMP interacts with two main effector proteins, protein kinase A (PKA) and exchange factor directly activated by cAMP 1/2 (EPAC1/2) each sharing unique responsibilities for **antifibrotic** effects [45]. PKA activation causes phosphorylation and activation of the transcription factor cAMP response element-binding protein (CREB) which is itself an antifibrotic mediator [48] (Fig. 1). In harmony with these findings phosphodiesterase inhibitors, which enhance intracellular cAMP, block fibroblast activation and reduce lung fibrosis in vivo [49]. In contrast to all other classes of G-proteins discussed above, G $\alpha$ s/cAMP signaling promotes phosphorylation and inhibition of YAP/TAZ nuclear translocation [37, 50], as well as reduced MRTF nuclear localization [51]. Ligands and agonists that stimulate G $\alpha$ s coupled GPCRs promote matrix degradation in multiple tissues [52, 53], and exert protective effects against lung fibroblast activation and fibrosis [46, 47]. Thus treatments that can selectively enhance G $\alpha$ s signaling in fibroblasts should have promise for anti-fibrotic therapies, and we discuss this opportunity in detail below.

## Challenges

### Redundancy

As already outlined above, GPCRs are a large class of receptors, several of which are capable of activating similar profibrotic features in pulmonary fibroblasts. Confirmed expression of multiple profibrotic GPCR ligands in human disease and mouse models of pulmonary fibrosis further supports the potential for widespread redundancy in fibroblast activating mechanisms in vivo. For example, LPA has been shown to be increased in the **bronchial alveolar lavage fluid** (BALF) [29], and **exhaled breath condensate** (EBC) [25] from patients with idiopathic pulmonary fibrosis, and in rodent models of pulmonary fibrosis [26–28]. Similar reports have been observed for ET-1 [22–24], serotonin [30] and S1P [31]. While mouse models have reported beneficial effects of therapies targeting individual GPCRs in these studies, the complex environment in human IPF may not be as

amenable to such an approach. Beyond GPCR ligands, additional biochemical and biomechanical signals contribute to fibroblast activation. These results strongly suggest pulmonary fibrosis is promoted by a profibrotic ligand and extracellular matrix milieu of diverse molecules with overlapping and convergent effects on fibroblast activation, challenging the concept that efficacy in this disease be achieved by blockade of an individual GPCR.

### Loss of G $\alpha$ s coupled GPCRs

An alternative strategy to antagonizing profibrotic signaling would be to agonize antifibrotic signaling, as already highlighted above through G $\alpha$ s -coupled elevation of cAMP. Strikingly however, widespread repression of Gs coupled GPCRs has been documented in pulmonary fibrosis, posing a challenge to this approach. The most well characterized example is the prostaglandin family of receptors. In cultured human lung fibroblasts prostaglandin E2 stimulates antifibrotic responses [47]. However, in fibroblasts derived from patients with IPF the anti-fibrotic efficacy of prostaglandin E2 is muted by reduced expression of the prostaglandin receptor *PTGER2* [54–56]. This phenomenon is also observed in experimental lung fibrosis [57]. Similarly, the relaxin receptor *RXFP1* has also been reported to be repressed in IPF patient samples [58]. Data from an RNAseq analysis of TGF $\beta$ 1-stimulated IPF fibroblasts (GSE136534) also documents a pervasive repression of G $\alpha$ s coupled GPCRs [59]. Together these findings suggest a coordinated transcriptional repression of G $\alpha$ s/cAMP signaling is a central feature of pathogenic fibroblast activation. While genomic datasets support this mechanism, detailed functional studies, as already conducted for prostaglandin E2, are still necessary to broadly validate receptor downregulation and its roles in IPF.

### Ubiquitous Receptor Expression

Another potential challenge in the development of GPCR regulators for the treatment of pulmonary fibrosis is the widespread expression of these receptors. In many cell types and multiple organs, receptors for endothelin, LPA, serotonin, angiotensin, S1P, and prostaglandins are some of the most highly expressed GPCRs [4, 60, 61]. At a systemic level targeting these receptors could cause dose limiting deleterious effects that preclude beneficial effects in the lung. Even within the lung there are likely to be confounding roles for some of these receptors in specific cell types important for the treatment of fibrosis. A prominent example coming into focus is the alveolar epithelium. Alveolar injury is a hallmark of IPF and repair of the epithelium is likely to be essential for successful resolution [13]. Following alveolar injury, alveolar type II (AEII) cells proliferate and differentiate into alveolar type I (AEI) cells, a process essential to lung repair [13]. The same pathway identified above as a desirable target in fibroblasts (YAP/TAZ) is essential to epithelial repair [62]. Specifically TAZ is required for epithelial differentiation following injury and genetic deletion of TAZ in AEII cells worsens fibrosis in the lungs [63]. In another example, G $\alpha$ q/11 genetic deletion in AEII cells causes pulmonary inflammation and alveolar enlargement consistent with emphysema [64]. The abundant overlap in GPCR expression between alveolar epithelium and fibroblasts [65], thus poses a substantial challenge for GPCR therapeutics in IPF. Similarly, overlap between the GPCRs expressed in fibroblasts and additional disease relevant cell types such as endothelium and macrophages adds further complexity that is only now beginning to be appreciated.

## Next Generation Strategies

### Solutions to Redundancy: Target downstream, polypharmacology, personalized medicine

As discussed above a multitude of receptors may be activated in IPF that all have the capacity to promote fibroblast activation. However, these receptors appear to rely on common intracellular pathways for this effect (Fig. 1). Thus, one strategy would be to target the common downstream pathways. Gα<sub>q</sub> is one of the major profibrotic heterotrimeric G-proteins activated by LPA, ET-1, ATII, and 5-HT signaling. Synthetic and naturally derived Gα<sub>q</sub> inhibitors have been investigated for efficacy in experimental models of asthma but have yet to be tested in pulmonary fibrosis [66]. **Rho-associated coiled-coil containing kinases** (ROCK1/2) are downstream effectors to Rho GTPases, and ROCK1/2 inhibitors effectively block fibroblast activation and pulmonary fibrosis in rodent models [67]. However the pleotropic roles of ROCK kinases and the well-known side effects of their inhibition, has raised concerns regarding their potential as therapeutic targets [68]. A recent investigation has found ROCK1- or ROCK2-haploinsufficient mice are equally resistant to pulmonary fibrosis, and targeting ROCK1 may have a beneficial effect in preventing alveolar epithelial apoptosis [69], opening the door to developing ROCK1/2 selective inhibitors that could offer greater safety while still effectively targeting lung fibrosis. Continuing downstream, another opportunity could be in targeting transcriptional regulators. Multiple small-molecule inhibitors of the MRTFA/B pathway have been developed and tested in models of tissue fibrosis [39, 70–72]. Notably, CCG-203971 reduced collagen lung content and enhanced apoptosis of activated fibroblasts in two rodent models of pulmonary fibrosis [42]. Similar approaches have been pursued in developing inhibitors of YAP/TAZ transcriptional activity. The first focused effort in developing a YAP/TAZ inhibitor resulted in the identification of porphyrin family rings, specifically verteporfin, as a feasible mechanism to therapeutically inhibit YAP/TAZ function [73]. Later studies showed effective antifibrotic effects of verteporfin in a silicosis model of pulmonary fibrosis [74]. Additionally, dihydrotanshinone I, a natural compound was found to reduce collagen expression through disruption of YAP/TAZ nuclear localization and transcriptional activity, shows efficacy in rodent models of liver [75] and lung [74] fibrosis. Finally, substantial effort has focused on targeting YAP and TAZ through modulating RhoGTPase membrane association via mevalonate metabolism, one effect of inhibiting HMG-CoA reductase using statin drugs [76]. Although this is a means of indirectly targeting YAP/TAZ, statins have already shown beneficial effect in rodent models of pulmonary fibrosis [77, 78] and in clinical data emerging from retrospective analysis [79, 80]. Together these data support the potential of targeting downstream of GPCRs to solve the problem of activating receptor redundancy. However, the potential challenge inherent in the widespread expression and function of these pathways, addressed below, remains.

**Polypharmacology** is the design and development of pharmaceutical agents with the ability to simultaneously interact with multiple targets and signaling pathways. Recently, polypharmacology has gained interest for its potential to generate higher efficacy agents with more predictable pharmacokinetic profile and reduced drug resistance [81]. Aided by advancements in GPCR structural and molecular biology, compounds can be rationally designed to target multiple receptors as agonist and antagonist [82]. Intriguingly, one of the

two drugs approved for the treatment of IPF is a clear example of a polypharmacology. Nintedanib is a tyrosine kinase inhibitor initially designed to target proangiogenic pathways. Nintedanib acts as an ATP-competitive inhibitor of fibroblast growth factor receptor (FGFR)-1, vascular endothelial growth factor receptor (VEGFR)-2, platelet-derived growth factor receptors (PDGFRs), Flt-3 and members of the Src-family, such as Src, Lyn and Lck [83],[84]. Two new GPCR ligands in clinical trials for IPF may also elicit their effects through polypharmacology (Table 1). RP5063 is a modulator of dopamine and serotonin receptors developed primarily for the treatment of schizophrenia and neuropsychiatric disorders. It has a partial agonism and potent binding affinity with dopamine D<sub>2</sub>, D<sub>3</sub>, D<sub>4</sub>, and serotonin 5-HT<sub>1A</sub> and 5-HT<sub>2A</sub> receptors, and antagonist activity at the serotonin 5-HT<sub>2B</sub>, 5-HT<sub>6</sub> and 5-HT<sub>7</sub> receptors. Moreover, it exhibits moderate binding affinity for 5-HT<sub>2C</sub>,  $\alpha_2$ -adrenergic, and muscarinic acetylcholine receptors [85]. PBI-4050 is a synthetic analog of a medium-length chain fatty acid recognized as an agonist of GPR40 and an antagonist of GPR84, both Free Fatty Acid Receptors (FFAR). As it exerts anti-inflammatory and anti-fibrotic properties, it is being studied for the treatment of Alström Syndrome, Liver and Lung fibrosis, and Chronic Kidney Diseases [86]. PBI-4050 diminishes bleomycin-induced pulmonary fibrosis in mice and reduces TGF- $\beta$  induced fibroblast activation in vitro [87]. The recent enthusiasm for polypharmacology is a potentially powerful solution to the complexity of GPCR actions in IPF. However, an inherent challenge to this strategy is how much “polypharmacology” is enough, especially when considering the diversity of ligands present in IPF. For example it is unlikely that one molecule could antagonize serotonin receptors (small-molecule) and endothelin receptors (peptide). Polypharmacology will likely benefit from an individualized patient strategy.

Personalized medicine involves a customized approach to disease, identifying the specific characteristics of a patient and tailoring their therapy accordingly. To date, clinical trials testing GPCR ligands for the treatment of IPF have taken a non-personalized approach, one compound for all enrolled patients (Table 1). More broadly, trials in IPF patients have thus far not considered “**endotypes**”, or subtypes of a disease with distinct pathophysiological or molecular mechanisms [88], a strategy embraced in other lung diseases including asthma and COPD [89, 90]. Several methods are available to ascertain levels of molecules in patients with IPF, including exhaled breath condensate, and serum sampling [91, 92]. These techniques could be used to identify the relative expression of potential GPCR ligands and tailor therapies based on molecular profiles. Going one step further, patient derived cells can be studied to rapidly identify suitable therapeutic strategies. For example, patient derived multicellular “pulmospheres” have shown responses that were predictive of therapeutic response to approved IPF drugs pirfenidone and nintedanib [93]. Future clinical trials may divide patient populations based on their respective endotypes, and analysis of ongoing and past trials of GPCR targeted therapeutics may provide critical insight into molecular classifiers that identify patients most likely to benefit from specific therapeutic approaches. Given the diversity of GPCRs implicated in IPF, such an approach seems warranted.



### Solutions to Loss of G $\alpha$ s Receptors: Rescue downregulated receptors, or target those that remain

Agonists of G $\alpha$ s receptors display impressive anti-fibrotic effects [46, 47, 52, 53]. However, decreased expression of receptors of this class in diseased tissue or in TGF $\beta$ -stimulated fibroblasts provides a challenge to an agonist based therapeutic. Significant efforts have been devoted to understanding the reduced responsiveness of IPF fibroblasts to prostaglandin E2 and have identified epigenetic repression of the *PTGER2* receptor through promoter hypermethylation as critical [54]. Inhibitors of PI3K and Akt reduce methylation of the *PTGER2* promoter and enhance its transcription, suggesting a mechanism to restore receptor expression and function [54]. In a recent study, TGF $\beta$  induced repression of GPCR encoding genes including prostanoid receptors (*PTGER2*, *PTGER4*, and *PTGIR*), adenosine receptors (*ADORA2A* and *ADORA2B*), as well as the beta-2 adrenergic receptor (*ADRB2*). This repression is dependent on **histone deacetylase** (HDAC) activity, and treatment with an investigational HDAC inhibitor pracinostat (SB939) enhanced expression of these G $\alpha$ s coupled receptors [59]. MicroRNAs have also been identified to function in TGF $\beta$  induced repression of G $\alpha$ s coupled receptors. Expression of miR-144-3p is enhanced in lungs of patients with IPF and stimulated by TGF $\beta$  in cultured lung fibroblasts. Expression of miR-144-3p causes reduced expression of the G $\alpha$ s coupled receptor, RFXP1, whereas blocking miR-144-3p with a targeted antagomir enhances expression of the receptor and reduces fibroblast activation [94]. These studies identify potential mechanisms to rescue G $\alpha$ s coupled receptors in patients with IPF by targeting the signaling pathways, epigenetic regulators, or microRNAs that repress their presentation.

A simpler approach is to target G $\alpha$ s coupled receptors not repressed in IPF. The calcitonin-receptor-like receptor (CRLR), which is activated by adrenomedullin and couples to G $\alpha$ s to promote cAMP, is actually increased in expression by TGF $\beta$  in fibroblasts and in the mouse lung following bleomycin induced fibrosis [95]. Likewise, the dopamine receptor D1 (*DRD1*) couples to G $\alpha$ s, and is not decreased in IPF patient fibroblasts or in freshly sorted fibroblasts from bleomycin injured mice [65]. More broadly, RNA-seq analysis of TGF $\beta$  stimulated fibroblasts identified three receptors that exclusively couple to G $\alpha$ s that were not repressed by TGF $\beta$ ; dopamine receptor D1, G-protein bile acid receptor 1, and an orphan GPCR, GPR3. Further identification of GPCRs preserved in disease settings, such as through emerging single cell RNA-seq analyses [17], will help to define the G $\alpha$ s coupled receptors available for targeting in IPF patients.

### Solutions to Ubiquitous Receptor Expression: Allosteric modulators, target selectively expressed receptors

The final and perhaps most challenging aspect of GPCR targeting is their widespread expression and function. One potential solution to this challenge is the use of **allosteric modulators**, molecules that bind to a unique site on a receptor to influence the activity of the ligand that binds to the orthosteric, or principle binding site [96]. This approach takes advantage of the potentially unique tissue and compartment-specific expression of endogenous ligands, such that it modulates GPCR function only where these ligands are present. This limits some of the problems that may occur with direct GPCR agonists and antagonists that will function throughout the organism [96]. Importantly, allosteric

molecules can enhance (positive allosteric modulators) or repress (negative allosteric modulators) the activity of ligand-receptor signaling. Two allosteric modulators have been FDA approved and there are several more in clinical trials [97]. Although not yet investigated for utility in pulmonary fibrosis, multiple allosteric modulators have been identified and developed for receptors of interest in IPF including serotonin, adenosine, adrenergic, dopamine, endothelin, free fatty acid and prostaglandin receptors [98–100]. The toolbox of allosteric modulators may thus provide unique opportunities to target widely expressed GPCRs safely and effectively for IPF.

A simpler solution, when available, is to identify and target receptors expressed uniquely in the cell type or compartment of interest. For example, the “GPCRome” of fibroblasts and alveolar epithelial cells displays considerable overlap, but there are multiple receptors uniquely expressed in each cell type [65]. The D1 dopamine receptor is the most preferentially expressed receptor in fibroblasts compared to alveolar epithelial cells and lung endothelial cells, and it can be selectively activated in vitro and in vivo without apparent effects on either lung epithelial or endothelial cells. D1 agonism in the mouse lung reduces lung collagen abundance after bleomycin injury, and in vitro stimulation of the D1 receptor prompts fibroblasts to take on a pro-resolution phenotype by enhancing gene expression of matrix degrading enzymes, reducing expression of matrix crosslinking genes, and promoting fibroblasts to produce a less stiff ECM. This approach to selectively target G $\alpha$ s in lung fibroblasts shows early promise as a targeted therapy for IPF. Expansion of this approach to include analysis of systematic datasets, including single cell RNA-seq profiles, may further refine our understanding of GPCR and ligand expression patterns across cell populations in health and disease, allowing for identification of additional selective targets for therapeutic intervention. The number of drugs which target GPCRs is very large; however the number of distinct GPCRs targeted by those drugs is actually only a small percentage of the more than 800 unique GPCRs [101]. Until recently, investigations into GPCR signaling in disease was focused on non-chemosensory, class A receptors (almost all of the receptors mentioned in this review). However enthusiasm has shifted towards lesser known adhesion receptors, orphan receptors, and chemosensory receptors [102], paving the way for new discoveries in pulmonary fibrosis.

## Concluding Remarks

GPCR targeting approaches have thus far yielded disappointing results as IPF therapies (Table 1). Despite this, several promising therapies continue to enter and progress through clinical trials, raising hopes for future success. Several challenges have emerged as outlined above, and major questions remain to be addressed (see Outstanding Questions) as we progress toward next generation therapeutic targets and candidates. The widespread expression and complex cell-specific effects of GPCR modulators will clearly need to be considered in refining our approaches to target identification and early testing. Single cell RNA-seq datasets will provide unique perspectives on receptor distribution [17, 103], while more complex human cellular model systems such as organoids and organ on chip models may provide useful early testing strategies for identifying promising directions [104, 105]. Personalized approaches that target molecular endotypes may split the patient population into smaller segments better served by specific candidate therapies. Candidate molecules



that employ polypharmacology or target common downstream mechanisms may overcome the challenge of redundant fibrosis-promoting signals. Finally, G $\alpha$ s targeting strategies, particularly those focused on receptors uniquely expressed on fibrosis-promoting activated fibroblasts may provide an effective new approach to IPF treatment. Collectively, these advances in understanding offer a fresh perspective on the challenges and opportunities in targeting GPCRs in IPF, and will underpin efforts to identify new strategies that offer therapeutic benefit.

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## Glossary

### **Allosteric Modulators**

Molecules that affect protein activity by binding to a secondary, allosteric, site rather than a primary, orthosteric, site

### **Alveolar and Alveolus**

Refers to an alveolus (plural: alveoli), a small, balloon-shaped air cavity arranged in clusters at the end of the respiratory tree in the lung parenchyma

### **Bronchial Alveolar Lavage Fluid (BALF)**

Fluid collected following the insertion of a device that can infuse solutions into specific parts of the lung

### **Cyclic Adenosine Monophosphate (cAMP)**

Second messenger molecule which is elevated following G $\alpha$ s activation

### **Endotypes**

A subtype of a condition defined by a distinct pathophysiological mechanism

### **Exhaled Breath Condensate (EBC)**

Breath is condensed following exhalation and allows the determination of biomolecules present in respiratory compartments

### **Extracellular Matrix (ECM)**

Non-cellular component that provides structural support to cells and organs

### **Fibroblasts**

Spindle-shape cells in the connective tissue responsible for the synthesis and degradation of extracellular matrix

### **Fluorescence-Activated Cell Sorting (FACS)**

The process of sorting cells for a desired population through the use of metrics such as size, protein fused fluorescent proteins, and cell surface markers

**G-protein Coupled Receptors (GPCRs)**

Largest family of cell membrane receptors that mediate physiological responses through the transduction of extracellular

**Histone Heacetylase (HDAC)**

A class of enzymes which removes acetyl groups from histones. This removal causes DNA to bind more tightly, and an overall decrease in gene expression

**Idiopathic Pulmonary Fibrosis (IPF)**

A progressive lung disease characterized by the uncontrolled scarring of the lung connective tissue by activated fibroblasts

**Myocardin-Related Transcription Factor (MRTFA/B)**

Transcriptional coactivators which are regulated by cytoskeletal dynamics, regulate expression of profibrotic genes

**Nintedanib**

An approved small molecule, therapeutic for the treatment of IPF. It has been shown to elicit antifibrotic activity through the inhibition of a wide range of tyrosine kinase receptors

**Pirfenidone**

An approved small molecule, therapeutic for the treatment of IPF. It has proven anti-fibrotic activity, but an unknown mechanism of action

**Polypharmacology**

Design and development of pharmaceutical agents that simultaneously interact with multiple targets

**Profibrotic and Antifibrotic**

A property that promotes or reduces fibrosis - the pathological accumulation of fibrous proteins in an organ

**Rho-associated coiled-coil containing kinases (ROCK1/2)**

Are a pair of serine/threonine kinases that are important for a wide range of processes in IPF

**Transforming Growth Factor-Beta (TGF $\beta$ )**

A profibrotic cytokine involved in fibroblast proliferation and recruitment, cell differentiation, and matrix regulation

**Yes-Associated Protein (YAP)/Transcriptional Coactivator with PDZ-binding Motif (TAZ)**

Transcriptional coactivators which are regulated by G-protein and mechanosignaling, regulate expression of profibrotic genes

**References**

1. Kroeze WK, Sheffler DJ, and Roth BL, G-protein-coupled receptors at a glance. *Journal of Cell Science*, 2003 116(24): p. 4867–4869. [PubMed: 14625380]
2. Vass M, et al., Chemical Diversity in the G Protein-Coupled Receptor Superfamily. *Trends in Pharmacological Sciences*, 2018 39(5): p. 494–512. [PubMed: 29576399]

3. Milligan G. and Kostenis E, Heterotrimeric G-proteins: a short history. *British Journal of Pharmacology*, 2006 147: p. S46–S55. [PubMed: 16402120]
4. Insel PA, et al., GPCR expression in tissues and cells: Are the optimal receptors being used as drug targets? *British Journal of Pharmacology*, 2012 165(6): p. 1613–1616. [PubMed: 21488863]
5. Insel PA, et al., GPCRomics: An Approach to Discover GPCR Drug Targets. *Trends in Pharmacological Sciences*, 2019 40(6): p. 378–387. [PubMed: 31078319]
6. Hauser AS, et al., Pharmacogenomics of GPCR Drug Targets. *Cell*, 2018 172(1–2): p. 41–+. [PubMed: 29249361]
7. Sriram K. and Insel PA, G Protein-Coupled Receptors as Targets for Approved Drugs: How Many Targets and How Many Drugs? *Molecular Pharmacology*, 2018 93(4): p. 251–258. [PubMed: 29298813]
8. Martinez FJ, et al., Idiopathic pulmonary fibrosis. *Nature Reviews Disease Primers*, 2017 3.
9. Richeldi L, Collard HR, and Jones MG, Idiopathic pulmonary fibrosis. *Lancet*, 2017 389(10082): p. 1941–1952. [PubMed: 28365056]
10. Canestaro WJ, et al., Drug Treatment of Idiopathic Pulmonary Fibrosis: Systematic Review and Network Meta-Analysis. *Chest*, 2016 149(3): p. 756–66. [PubMed: 26836914]
11. Maher TM and Streck ME, Antifibrotic therapy for idiopathic pulmonary fibrosis: time to treat. *Respir Res*, 2019 20(1): p. 205. [PubMed: 31492155]
12. Winters NI, et al., Epithelial Injury and Dysfunction in the Pathogenesis of Idiopathic Pulmonary Fibrosis. *Am J Med Sci*, 2019 357(5): p. 374–378. [PubMed: 31010463]
13. Kulkarni T, et al., Alveolar epithelial disintegrity in pulmonary fibrosis. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 2016 311(2): p. L185–L191. [PubMed: 27233996]
14. King TE Jr., Pardo A, and Selman M, Idiopathic pulmonary fibrosis. *Lancet*, 2011 378(9807): p. 1949–61. [PubMed: 21719092]
15. Misharin AV, et al., Monocyte-derived alveolar macrophages drive lung fibrosis and persist in the lung over the life span. *J Exp Med*, 2017 214(8): p. 2387–2404. [PubMed: 28694385]
16. Morse C, et al., Proliferating SPP1/MERTK-expressing macrophages in idiopathic pulmonary fibrosis. *European Respiratory Journal*, 2019 54(2).
17. Reyfman PA, et al., Single-Cell Transcriptomic Analysis of Human Lung Provides Insights into the Pathobiology of Pulmonary Fibrosis. *American Journal of Respiratory and Critical Care Medicine*, 2019 199(12): p. 1517–1536. [PubMed: 30554520]
18. Moore BB and Hogaboam CM, Murine models of pulmonary fibrosis. *Am J Physiol Lung Cell Mol Physiol*, 2008 294(2): p. L152–60. [PubMed: 17993587]
19. Pardo A. and Selman M, Lung Fibroblasts, Aging, and Idiopathic Pulmonary Fibrosis. *Ann Am Thorac Soc*, 2016 13 Suppl 5: p. S417–S421. [PubMed: 28005427]
20. Deshpande DA and Penn RB, Targeting G protein-coupled receptor signaling in asthma. *Cell Signal*, 2006 18(12): p. 2105–20. [PubMed: 16828259]
21. Wendell SG, Fan H, and Zhang C, G Protein-Coupled Receptors in Asthma Therapy: Pharmacology and Drug Action. *Pharmacol Rev*, 2020 72(1): p. 1–49. [PubMed: 31767622]
22. Ugucioni M, et al., Endothelin-1 in Idiopathic Pulmonary Fibrosis. *Journal of Clinical Pathology*, 1995 48(4): p. 330–334. [PubMed: 7615852]
23. Mutsaers SE, et al., Increased endothelin-1 and its localization during the development of bleomycin-induced pulmonary fibrosis in rats. *Am J Respir Cell Mol Biol*, 1998 18(5): p. 611–9. [PubMed: 9569231]
24. Saleh D, et al., Elevated expression of endothelin-1 and endothelin-converting enzyme-1 in idiopathic pulmonary fibrosis: possible involvement of proinflammatory cytokines. *Am J Respir Cell Mol Biol*, 1997 16(2): p. 187–93. [PubMed: 9032126]
25. Montesi SB, et al., Docosatetraenoyl LPA is elevated in exhaled breath condensate in idiopathic pulmonary fibrosis. *Bmc Pulmonary Medicine*, 2014 14.
26. Cong CC, et al., Regulation of silicosis formation by lysophosphatidic acid and its receptors. *Experimental Lung Research*, 2014 40(7): p. 317–326. [PubMed: 24926730]

27. Black KE, et al., Autotaxin activity increases locally following lung injury, but is not required for pulmonary lysophosphatidic acid production or fibrosis. *Faseb Journal*, 2016 30(6): p. 2435–2450. [PubMed: 27006447]
28. Tager A, et al., The lysophosphatidic acid receptor LPA1 links pulmonary fibrosis to lung injury by mediating fibroblast recruitment and vascular leak. *Inflammation Research*, 2007 56: p. S347–S347.
29. Tager AM, et al., The lysophosphatidic acid receptor LPA1 links pulmonary fibrosis to lung injury by mediating fibroblast recruitment and vascular leak. *Nat Med*, 2008 14(1): p. 45–54. [PubMed: 18066075]
30. Fabre A, et al., Modulation of bleomycin-induced lung fibrosis by serotonin receptor antagonists in mice. *Eur Respir J*, 2008 32(2): p. 426–36. [PubMed: 18321937]
31. Huang LS and Natarajan V, Sphingolipids in pulmonary fibrosis. *Adv Biol Regul*, 2015 57: p. 55–63. [PubMed: 25446881]
32. Tan WSD, et al., Targeting the renin-angiotensin system as novel therapeutic strategy for pulmonary diseases. *Current Opinion in Pharmacology*, 2018 40: p. 9–17. [PubMed: 29288933]
33. Flock T, et al., Selectivity determinants of GPCR-G-protein binding. *Nature*, 2017 545(7654): p. 317–+. [PubMed: 28489817]
34. Radeff-Huang J, et al., G protein mediated signaling pathways in lysophospholipid induced cell proliferation and survival. *J Cell Biochem*, 2004 92(5): p. 949–66. [PubMed: 15258918]
35. Maekawa M, et al., Signaling from rho to the actin cytoskeleton through protein kinases ROCK and LIM-kinase. *Science*, 1999 285(5429): p. 895–898. [PubMed: 10436159]
36. Yu OM, Miyamoto S, and Brown JH, Myocardin-Related Transcription Factor A and Yes-Associated Protein Exert Dual Control in G Protein-Coupled Receptor- and RhoA-Mediated Transcriptional Regulation and Cell Proliferation. *Mol Cell Biol*, 2016 36(1): p. 39–49. [PubMed: 26459764]
37. Yu FX, et al., Regulation of the Hippo-YAP Pathway by G-Protein-Coupled Receptor Signaling. *Cell*, 2012 150(4): p. 780–791. [PubMed: 22863277]
38. Liu F, et al., Mechanosignaling through YAP and TAZ drives fibroblast activation and fibrosis. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 2015 308(4): p. L344–L357. [PubMed: 25502501]
39. Tsou PS, et al., Cellular mechanisms of tissue fibrosis. 8. Current and future drug targets in fibrosis: focus on Rho GTPase-regulated gene transcription. *Am J Physiol Cell Physiol*, 2014 307(1): p. C2–13. [PubMed: 24740541]
40. Bernau K, et al., Megakaryoblastic leukemia-1 is required for the development of bleomycin-induced pulmonary fibrosis. *Respiratory Research*, 2015 16.
41. Tschumperlin DJ, et al., Mechanosensing and fibrosis. *J Clin Invest*, 2018 128(1): p. 74–84. [PubMed: 29293092]
42. Sisson TH, et al., Inhibition of myocardin-related transcription factor/serum response factor signaling decreases lung fibrosis and promotes mesenchymal cell apoptosis. *Am J Pathol*, 2015 185(4): p. 969–86. [PubMed: 25681733]
43. Speight P, et al., Context-dependent switch in chemo/mechanotransduction via multilevel crosstalk among cytoskeleton-regulated MRTF and TAZ and TGFbeta-regulated Smad3. *Nat Commun*, 2016 7: p. 11642. [PubMed: 27189435]
44. Miranda MZ, et al., TGF-beta1 regulates the expression and transcriptional activity of TAZ protein via a Smad3-independent, myocardin-related transcription factor-mediated mechanism. *J Biol Chem*, 2017 292(36): p. 14902–14920. [PubMed: 28739802]
45. Insel PA, et al., cAMP and Epac in the regulation of tissue fibrosis. *Br J Pharmacol*, 2012 166(2): p. 447–56. [PubMed: 22232328]
46. Della Latta V, et al., The role of the adenosinergic system in lung fibrosis. *Pharmacological Research*, 2013 76: p. 182–189. [PubMed: 23994158]
47. Bozyk PD and Moore BB, Prostaglandin E2 and the pathogenesis of pulmonary fibrosis. *Am J Respir Cell Mol Biol*, 2011 45(3): p. 445–52. [PubMed: 21421906]

48. Liu Y, et al., Dibutyryl-cAMP attenuates pulmonary fibrosis by blocking myofibroblast differentiation via PKA/CREB/CBP signaling in rats with silicosis. *Respiratory Research*, 2017 18.
49. Sisson TH, et al., Phosphodiesterase 4 inhibition reduces lung fibrosis following targeted type II alveolar epithelial cell injury. *Physiological Reports*, 2018 6(12).
50. Zmajkovicova K, et al., The Antifibrotic Activity of Prostacyclin Receptor Agonism Is Mediated through Inhibition of YAP/TAZ. *American Journal of Respiratory Cell and Molecular Biology*, 2019 60(5): p. 578–591. [PubMed: 30537446]
51. Penke LR, et al., Prostaglandin E2 inhibits alpha-smooth muscle actin transcription during myofibroblast differentiation via distinct mechanisms of modulation of serum response factor and myocardin-related transcription factor-A. *J Biol Chem*, 2014 289(24): p. 17151–62. [PubMed: 24802754]
52. Neumann E, Khawaja K, and Muller-Ladner U, G protein-coupled receptors in rheumatology. *Nature Reviews Rheumatology*, 2014 10(7): p. 429–436. [PubMed: 24798574]
53. Samuel CS, et al., Anti-fibrotic actions of relaxin. *British Journal of Pharmacology*, 2017 174(10): p. 962–976. [PubMed: 27250825]
54. Huang SK, et al., Hypermethylation of PTGER2 confers prostaglandin E2 resistance in fibrotic fibroblasts from humans and mice. *Am J Pathol*, 2010 177(5): p. 2245–55. [PubMed: 20889571]
55. Huang SK, et al., Variable prostaglandin E2 resistance in fibroblasts from patients with usual interstitial pneumonia. *Am J Respir Crit Care Med*, 2008 177(1): p. 66–74. [PubMed: 17916807]
56. Mukherjee S, et al., Prostaglandin E2 inhibits profibrotic function of human pulmonary fibroblasts by disrupting Ca(2+) signaling. *Am J Physiol Lung Cell Mol Physiol*, 2019 316(5): p. L810–L821. [PubMed: 30758990]
57. Moore BB, et al., Bleomycin-induced E prostanoid receptor changes alter fibroblast responses to prostaglandin E2. *J Immunol*, 2005 174(9): p. 5644–9. [PubMed: 15843564]
58. Tan JN, et al., Expression of RXFP1 Is Decreased in Idiopathic Pulmonary Fibrosis Implications for Relaxin-based Therapies. *American Journal of Respiratory and Critical Care Medicine*, 2016 194(11): p. 1392–1402. [PubMed: 27310652]
59. Jones DL, et al., TGFbeta-induced fibroblast activation requires persistent and targeted HDAC-mediated gene repression. *J Cell Sci*, 2019.
60. Insel PA, et al., GPCRomics: An Approach to Discover GPCR Drug Targets. *Trends Pharmacol Sci*, 2019 40(6): p. 378–387. [PubMed: 31078319]
61. Snead AN and Insel PA, Defining the cellular repertoire of GPCRs identifies a profibrotic role for the most highly expressed receptor, protease-activated receptor 1, in cardiac fibroblasts. *Faseb Journal*, 2012 26(11): p. 4540–4547. [PubMed: 22859370]
62. LaCanna R, et al., Yap/Taz regulate alveolar regeneration and resolution of lung inflammation. *Journal of Clinical Investigation*, 2019 129(5): p. 2107–2122.
63. Sun TH, et al., TAZ is required for lung alveolar epithelial cell differentiation after injury. *Jci Insight*, 2019 4(14).
64. John AE, et al., Loss of epithelial Gq and G11 signaling inhibits TGFbeta production but promotes IL-33-mediated macrophage polarization and emphysema. *Sci Signal*, 2016 9(451): p. ra104. [PubMed: 27811142]
65. Haak AJ, et al., Selective YAP/TAZ inhibition in fibroblasts via dopamine receptor D1 agonism reverses fibrosis. *Sci Transl Med*, 2019 11(516).
66. Carr R 3rd, et al., Interdicting Gq Activation in Airway Disease by Receptor-Dependent and Receptor-Independent Mechanisms. *Mol Pharmacol*, 2016 89(1): p. 94–104. [PubMed: 26464325]
67. Knipe RS, Tager AM, and Liao JK, The Rho kinases: critical mediators of multiple profibrotic processes and rational targets for new therapies for pulmonary fibrosis. *Pharmacol Rev*, 2015 67(1): p. 103–17. [PubMed: 25395505]
68. Feng YB, et al., Rho Kinase (ROCK) Inhibitors and Their Therapeutic Potential. *Journal of Medicinal Chemistry*, 2016 59(6): p. 2269–2300. [PubMed: 26486225]
69. Knipe RS, et al., The Rho Kinase Isoforms ROCK1 and ROCK2 Each Contribute to the Development of Experimental Pulmonary Fibrosis. *American Journal of Respiratory Cell and Molecular Biology*, 2018 58(4): p. 471–481. [PubMed: 29211497]

70. Johnson LA, et al., Novel Rho/MRTF/SRF inhibitors block matrix-stiffness and TGF-beta-induced fibrogenesis in human colonic myofibroblasts. *Inflamm Bowel Dis*, 2014 20(1): p. 154–65. [PubMed: 24280883]
71. Haak AJ, et al., Targeting the Myofibroblast Genetic Switch: Inhibitors of Myocardin-Related Transcription Factor/Serum Response Factor-Regulated Gene Transcription Prevent Fibrosis in a Murine Model of Skin Injury. *Journal of Pharmacology and Experimental Therapeutics*, 2014 349(3): p. 480–486.
72. Kahl DJ, et al., 5-Aryl-1,3,4-oxadiazol-2-ylthioalkanoic Acids: A Highly Potent New Class of Inhibitors of Rho/Myocardin-Related Transcription Factor (MRTF)/Serum Response Factor (SRF)-Mediated Gene Transcription as Potential Antifibrotic Agents for Scleroderma. *J Med Chem*, 2019 62(9): p. 4350–4369. [PubMed: 30951312]
73. Liu-Chittenden Y, et al., Genetic and pharmacological disruption of the TEAD-YAP complex suppresses the oncogenic activity of YAP. *Genes & Development*, 2012 26(12): p. 1300–1305. [PubMed: 22677547]
74. Li SY, et al., Targeting Mechanics-Induced Fibroblast Activation through CD44-RhoA-YAP Pathway Ameliorates Crystalline Silica-Induced Silicosis. *Theranostics*, 2019 9(17): p. 4993–5008. [PubMed: 31410197]
75. Ge M, et al., The anti-hepatic fibrosis effects of dihydrotanshinone I are mediated by disrupting the yes-associated protein and transcriptional enhancer D2 complex and stimulating autophagy. *Br J Pharmacol*, 2017 174(10): p. 1147–1160. [PubMed: 28257144]
76. Sorrentino G, et al., Metabolic control of YAP and TAZ by the mevalonate pathway. *Nature Cell Biology*, 2014 16(4): p. 357–+. [PubMed: 24658687]
77. Tulek B, et al., Effects of simvastatin on bleomycin-induced pulmonary fibrosis in female rats. *Biological Research*, 2012 45(4): p. 345–350. [PubMed: 23558989]
78. Zhu B, et al., Atorvastatin attenuates bleomycin-induced pulmonary fibrosis via suppressing iNOS expression and the CTGF (CCN2)/ERK signaling pathway. *Int J Mol Sci*, 2013 14(12): p. 24476–91. [PubMed: 24351828]
79. Kreuter M, et al., Effect of statins on disease-related outcomes in patients with idiopathic pulmonary fibrosis. *Thorax*, 2017 72(2): p. 148–153. [PubMed: 27708114]
80. Kreuter M, et al., Statin Therapy and Outcomes in Trials of Nintedanib in Idiopathic Pulmonary Fibrosis. *Respiration*, 2018 95(5): p. 317–326. [PubMed: 29414827]
81. Anighoro A, Bajorath J, and Rastelli G, Polypharmacology: Challenges and Opportunities in Drug Discovery. *Journal of Medicinal Chemistry*, 2014 57(19): p. 7874–7887. [PubMed: 24946140]
82. Jacobson KA, New paradigms in GPCR drug discovery. *Biochem Pharmacol*, 2015 98(4): p. 541–55. [PubMed: 26265138]
83. Roth GJ, et al., Nintedanib: from discovery to the clinic. *J Med Chem*, 2015 58(3): p. 1053–63. [PubMed: 25474320]
84. Wollin L, et al., Mode of action of nintedanib in the treatment of idiopathic pulmonary fibrosis. *Eur Respir J*, 2015 45(5): p. 1434–45. [PubMed: 25745043]
85. Rajagopal L, et al., RP5063, an atypical antipsychotic drug with a unique pharmacologic profile, improves declarative memory and psychosis in mouse models of schizophrenia. *Behavioural Brain Research*, 2017 332: p. 180–199. [PubMed: 28373127]
86. Li Y, et al., Fatty acid receptor modulator PBI-4050 inhibits kidney fibrosis and improves glycemic control. *Jci Insight*, 2018 3(10).
87. Nguyen QT, et al., PBI-4050 Reduces Pulmonary Hypertension, Lung fibrosis and Right Ventricular Dysfunction in Heart Failure. *Cardiovasc Res*, 2019.
88. Goodwin AT and Jenkins G, Molecular Endotyping of Pulmonary Fibrosis. *Chest*, 2016 149(1): p. 228–237. [PubMed: 26356594]
89. Svenningsen S. and Nair P, Asthma Endotypes and an Overview of Targeted Therapy for Asthma. *Front Med (Lausanne)*, 2017 4: p. 158. [PubMed: 29018800]
90. Garudadri S. and Woodruff PG, Targeting Chronic Obstructive Pulmonary Disease Phenotypes, Endotypes, and Biomarkers. *Ann Am Thorac Soc*, 2018 15(Suppl 4): p. S234–S238. [PubMed: 30758998]



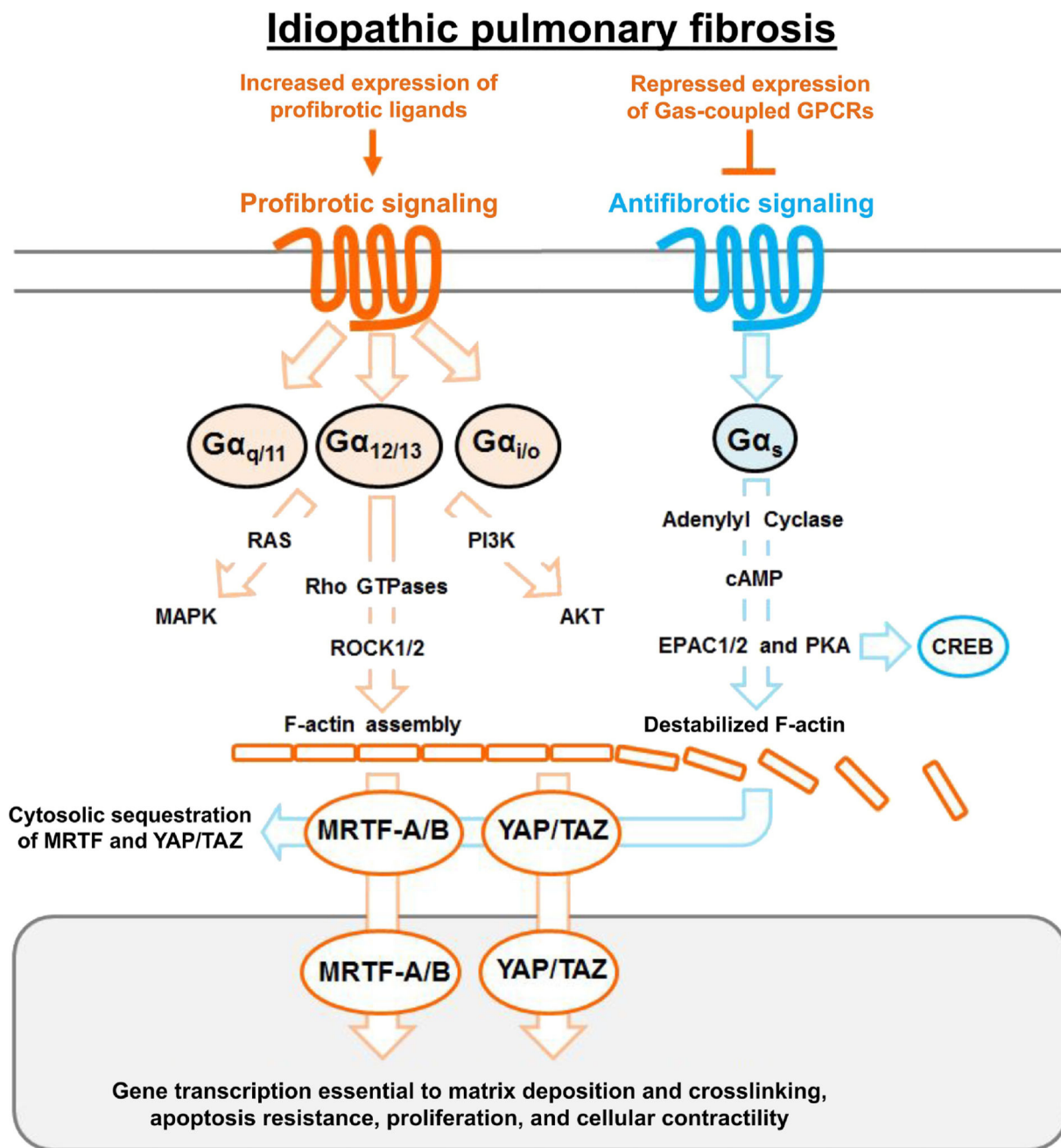
91. Hayton C, et al., Breath biomarkers in idiopathic pulmonary fibrosis: a systematic review. *Respir Res*, 2019 20(1): p. 7. [PubMed: 30634961]
92. Spagnolo P, Tzouveleakis A, and Maher TM, Personalized medicine in idiopathic pulmonary fibrosis: facts and promises. *Current Opinion in Pulmonary Medicine*, 2015 21(5): p. 470–478. [PubMed: 26132817]
93. Surolia R, et al., 3D pulmospheres serve as a personalized and predictive multicellular model for assessment of antifibrotic drugs. *Jci Insight*, 2017 2(2).
94. Bahudhanapati H, et al., MicroRNA-144–3p targets relaxin/insulin-like family peptide receptor 1 (RXFP1) expression in lung fibroblasts from patients with idiopathic pulmonary fibrosis. *J Biol Chem*, 2019 294(13): p. 5008–5022. [PubMed: 30709904]
95. Kach J, et al., Regulation of myofibroblast differentiation and bleomycin-induced pulmonary fibrosis by adrenomedullin. *Am J Physiol Lung Cell Mol Physiol*, 2013 304(11): p. L757–64. [PubMed: 23585227]
96. Christopoulos A, Advances in G protein-coupled receptor allosteric modulation: from function to structure. *Mol Pharmacol*, 2014 86(5): p. 463–78. [PubMed: 25061106]
97. Hauser AS, et al., Trends in GPCR drug discovery: new agents, targets and indications. *Nat Rev Drug Discov*, 2017 16(12): p. 829–842. [PubMed: 29075003]
98. Gentry PR, Sexton PM, and Christopoulos A, Novel Allosteric Modulators of G Protein-coupled Receptors. *J Biol Chem*, 2015 290(32): p. 19478–88. [PubMed: 26100627]
99. Jiang JX, et al., Discovery of 2-Piperidinyl Phenyl Benzamides and Trisubstituted Pyrimidines as Positive Allosteric Modulators of the Prostaglandin Receptor EP2. *Acs Chemical Neuroscience*, 2018 9(4): p. 699–707. [PubMed: 29292987]
100. Luderman KD, et al., Identification of Positive Allosteric Modulators of the D-1 Dopamine Receptor That Act at Diverse Binding Sites. *Molecular Pharmacology*, 2018 94(4): p. 1197–1209. [PubMed: 30068735]
101. Wacker D, Stevens RC, and Roth BL, How Ligands Illuminate GPCR Molecular Pharmacology. *Cell*, 2017 170(3): p. 414–427. [PubMed: 28753422]
102. Hauser AS, et al., Trends in GPCR drug discovery: new agents, targets and indications. *Nature Reviews Drug Discovery*, 2017 16(12): p. 829–842. [PubMed: 29075003]
103. Xie T, et al., Single-Cell Deconvolution of Fibroblast Heterogeneity in Mouse Pulmonary Fibrosis. *Cell Rep*, 2018 22(13): p. 3625–3640. [PubMed: 29590628]
104. Gkatzis K, et al., Use of three-dimensional organoids and lung-on-a-chip methods to study lung development, regeneration and disease. *European Respiratory Journal*, 2018 52(5).
105. Clevers H, Modeling Development and Disease with Organoids. *Cell*, 2016 165(7): p. 1586–1597. [PubMed: 27315476]
106. King TE Jr., et al., BUILD-1: a randomized placebo-controlled trial of bosentan in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med*, 2008 177(1): p. 75–81. [PubMed: 17901413]
107. King TE Jr., et al., BUILD-3: a randomized, controlled trial of bosentan in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med*, 2011 184(1): p. 92–9. [PubMed: 21474646]
108. Raghu G, et al., Treatment of idiopathic pulmonary fibrosis with ambrisentan: a parallel, randomized trial. *Ann Intern Med*, 2013 158(9): p. 641–9. [PubMed: 23648946]
109. Raghu G, et al., Macitentan for the treatment of idiopathic pulmonary fibrosis: the randomised controlled MUSIC trial. *Eur Respir J*, 2013 42(6): p. 1622–32. [PubMed: 23682110]
110. Palmer SM, et al., Randomized, Double-Blind, Placebo-Controlled, Phase 2 Trial of BMS-986020, a Lysophosphatidic Acid Receptor Antagonist for the Treatment of Idiopathic Pulmonary Fibrosis. *Chest*, 2018 154(5): p. 1061–1069. [PubMed: 30201408]
111. Khalil N, et al., Phase 2 clinical trial of PBI-4050 in patients with idiopathic pulmonary fibrosis. *European Respiratory Journal*, 2019 53(3).
112. Couluris M, et al., Treatment of idiopathic pulmonary fibrosis with losartan: a pilot project. *Lung*, 2012 190(5): p. 523–7. [PubMed: 22810758]
113. Wright CE, et al., Inhaled beclomethasone/formoterol in idiopathic pulmonary fibrosis: a randomised controlled exploratory study. *ERJ Open Res*, 2017 3(4).

### Outstanding Questions

- Can targeting a single GPCR effectively treat most patients with IPF?
- When identifying GPCR-based targets for antifibrotic therapy, when and how should receptor expression and function in diverse cell types be considered?
- Is the field too focused on molecular target identification and not enough on understanding the complex multicellular interactions and integrated effects of GPCR-based therapeutic candidates?

### Highlights

- Multiple GPCR ligand receptor pairs are implicated in IPF, and clinical trials are currently underway targeting GPCR pathways for the treatment of IPF.
- Individual GPCRs can promote profibrotic or antifibrotic phenotypes in lung fibroblasts, depending on the receptor class and downstream signaling pathways.
- The convergence of downstream pathways on common signaling and transcriptional mechanisms integrates diverse GPCR effects and may provide a path to overcome redundancy.
- Signaling programs downstream of GPCR signaling are also essential to alveolar epithelial regeneration and repair, highlighting the need to identify strategies that account for this complexity.



Trends in Pharmacological Sciences

**Fig. 1. GPCR Signaling Promotes profibrotic and antifibrotic signaling.**

In IPF there is increased abundance of profibrotic ligands, notably: LPA, endothelin, and serotonin that activate receptors coupled to  $G_{\alpha i/o}$ ,  $G_{\alpha q/11}$ ,  $G_{\alpha 12/13}$  promoting multiple pathways including Rho and ROCK which regulates the actin cytoskeleton. MRTF-A/B and YAP/TAZ are cytoskeletal sensitive profibrotic transcription co-factors essential to fibroblast activation. Known transcript targets for MRTF and YAP/TAZ are profibrotic genes: *COL1A1*, *COL1A2*, *CTGF*, and *ACTA2*. GPCRs which couple to  $G_{\alpha s}$  are antifibrotic and negatively regulate MRTF-A/B and YAP/TAZ, but are often repressed in IPF patient

fibroblasts. Profibrotic signaling is shown in orange and antifibrotic signaling is show in blue.

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**Table 1.**

Summary of GPCR Targeting Clinical Trials for IPF

Receptor activity	Drug	Outcomes	Status	Trial Number Citation
ET <sub>A</sub> and ET <sub>B</sub> antagonist	Bosentan	No significant Improvements.	Phase 2/3 (completed)	<a href="#">NCT00071461</a> [106]
ET <sub>A</sub> and ET <sub>B</sub> antagonist	Bosentan	No significant Improvements.	Phase 3 (completed)	<a href="#">NCT00391443</a> [107]
ET <sub>A</sub> antagonist	Ambrisentan	More patients experienced disease progression and death in treatment group compared to placebo.	Phase 3 (terminated)	<a href="#">NCT00768300</a> [108]
ET <sub>A</sub> and ET <sub>B</sub> antagonist	Macitentan	No significant Improvements.	Phase 2 (completed)	<a href="#">NCT00903331</a> [109]
LPA <sub>1</sub> antagonist	BMS-986020	Improvement in forced vital capacity for the 600 mg/bid group compared to placebo.	Phase 2 (completed)	<a href="#">NCT01766817</a> [110]
GPR40 agonist and GPR84 antagonist	PBI-4050	PBI-4050 alone or in combination was well tolerated.	Phase 2 (completed)	<a href="#">NCT02538536</a> [111]
AT <sub>1</sub> antagonist	Losartan	Improvement in forced vital capacity.	Pilot Study	<a href="#">NCT00879879</a> [112]
β <sub>2</sub> Adrenergic agonist	Formoterol	Treatment significantly improved forced expiratory volume and flow.	Pilot Study	EudraCT: 2013-004404-19 [113]
Leukotriene antagonist	Tipelukast	-	Phase 2 (recruiting)	<a href="#">NCT02503657</a>
Prostanoid antagonist	Treprostinil	-	Phase 2 (terminated)	<a href="#">NCT00703339</a>
Prostanoid antagonist	Treprostinil	-	Phase 2 (completed)	<a href="#">NCT00705133</a>
GPR84 antagonist	GLPG1205	-	Phase 2 (recruiting)	<a href="#">NCT03725852</a>
Smoothened antagonist	Vismodegib	-	Phase 1 (completed) Phase 2 (terminated)	<a href="#">NCT02648048</a> <a href="#">NCT02168530</a>
Serotonergic and Dopaminergic ligand	RP5063	-	Phase 2 (planning)	-