

Protective Effects of Baicalin on Arsenic Trioxide-induced Oxidative Damage and Apoptosis in Human Umbilical Vein Endothelial Cells

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Abstract. *Background/Aim:* Arsenic trioxide (As_2O_3) is an environmental pollutant. However, the detailed mechanisms about As_2O_3 -induced loss of endothelial integrity are unknown. This study aimed at investigating how As_2O_3 causes endothelial dysfunction and whether baicalin can reverse such dysfunction. *Materials and Methods:* Human umbilical vein endothelial cells (HUVECs) were used to examine As_2O_3 -induced oxidative stress, and apoptosis. The influence of baicalin on As_2O_3 -induced endothelial dysfunction were investigated. *Results:* The viability of HUVECs was inhibited by As_2O_3 and cells underwent apoptosis. As_2O_3 treatment increased NADPH oxidase activity, and elevated the level of reactive oxygen species (ROS). Formamidopyrimidine DNA-glycosylase- and endonuclease III-digestible adducts were accumulated. Baicalin reversed As_2O_3 -induced apoptosis and As_2O_3 -suppressed cell viability. Baicalin caused a decrease in NADPH oxidase activity, and re-balanced the ROS level. As_2O_3 -induced formamidopyrimidine DNA-glycosylase- and endonuclease III-digestible adducts were down-regulated.

Conclusion: Baicalin was found to have the potential capacity to protect endothelial cells from As_2O_3 -induced cytotoxicity.

Arsenic has been recognized as a major inorganic pollutant, and drinking contaminated groundwater is a major source of arsenic toxicity for humans. The level of inorganic arsenic in contaminated groundwater has been found to be as high as 5300 $\mu\text{g/l}$ in Latin America and Asian countries (1-5). From the viewpoint of epidemiology, mounting evidence has shown that chronic exposure to inorganic arsenic from drinking water is closely related to many human disorders, including atherosclerosis (6), diabetes mellitus (7), hypertension (8), ischemic heart disease (9, 10) peripheral vascular disease (11), and cancer (12, 13). However, extremely little information is available in the literature on the effects of inorganic arsenic exposure on the vascular system. Translational scientists are also interested in potential anti-toxic traditional Chinese medicine for protection against these disorders caused by chronic exposure to arsenic. Endothelial cells in blood vessels are the key targets of vasculopathy caused by inorganic arsenic exposure. Arsenic has been reported to increase vascular oxidative stress, which has been reported to promote cell apoptosis and dysfunction of endothelial cells (14). Oxidative stress is thought to be a major part of the intracellular mechanism of arsenic toxicity (15, 16), while its effects on DNA has, as far as we are aware, never been studied.

Scutellaria baicalensis Georgi has been used as traditional Chinese medicine for more than 2,000 years, and has been reported to have antioxidant, anti-inflammatory, antibacterial, and immune-enhancing properties (17-21). Baicalin is a flavonoid representing the major bioactive component from the

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roots of *Scutellaria baicalensis* Georgi and is reported to have a series of pharmacological activities, including antioxidant, anti-inflammatory, antiviral, anti-human immunodeficiency virus and antitumor properties (17, 22-27). However, to our knowledge, it has never been studied whether it may protect from arsenic toxicity. The aim of the present study was to evaluate the effects of As₂O₃ exposure on endothelial apoptosis and oxidative stress, and to investigate whether baicalin can effectively prevent such oxidative endothelial damage.

Materials and Methods

Cell culture and chemicals. Human umbilical vein endothelial cells (HUVECs) (CRL-1730; American Type Culture Collection, Manassas, VA, USA) were maintained in RPMI-1640 Medium (Hyclone, South Logan, UT, USA), supplemented with 10% fetal calf serum. Baicalin, As₂O₃, dimethyl sulfoxide (DMSO), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) and propidium iodide (PI) were obtained from Sigma Chemical Co. (St. Louis, MO, USA) and Aldrich Chemical Co. (Milwaukee, WI, USA). Baicalin was freshly dissolved in DMSO and the final concentration of DMSO in any experimental treatment was less than 0.05%.

Cell viability assay. Cell viability were performed by tetrazolium 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) colorimetric assay as previously published (28, 29). Briefly, HUVECs were cultured on 96-well plates at a density of 3×10^4 cells/well, and then exposed to 0-20 μM As₂O₃, with or without pre-treatment of 0-200 $\mu\text{g}/\text{ml}$ baicalin for 1 h prior to As₂O₃ exposure. After As₂O₃ exposure, the cells were re-incubated with 0.5 mg/ml MTT. The plates were further incubated in the dark for 4 h at 37°C. The color intensity was then measured at 570 nm using a Multiskan MS ELISA reader (Labsystems, Helsinki, Finland). Each experiment was repeated at least in triplicate.

Cell apoptosis measurement. HUVECs were seeded in 10-cm dishes at $2 \times 10^6/\text{ml}$ and treated with 0-20 μM As₂O₃, with or without 0-200 $\mu\text{g}/\text{ml}$ baicalin pre-treatment, and cells were incubated for 24 h. Then the cells were fixed with 70% ethanol and incubated with PI buffer [4 $\mu\text{g}/\text{ml}$ PI, 0.5 $\mu\text{g}/\text{ml}$ RNase, and 1% Triton X-100 in phosphate-buffered saline (PBS)] for 30 min in the dark. After the fixing and staining processes, the HUVECs were filtered through a 40- μm nylon filter and 10,000 HUVECs were analyzed for appearance of sub-G₁ phase cells using a FACS Calibur instrument (BD Biosciences, San Jose, CA, USA) equipped with Cell Quest software as described elsewhere (30, 31). The sub-G₁ population was deemed to be representative of the percentage of cells in apoptosis. Each experiment was repeated at least in triplicate.

Measurement of ROS production. Cells were plated at a density of 2×10^5 cells/well into 12-well plates and incubated with 0-20 μM As₂O₃ alone, 0-200 $\mu\text{g}/\text{ml}$ baicalin alone, or pre-treatment 0-200 $\mu\text{g}/\text{ml}$ of baicalin followed by co-treatment for 12 h. The cells were harvested, re-suspended in 10 μM 2',7'-dichlorodihydrofluorescein diacetate, incubated at 37°C for 30 min and then analyzed by flow cytometry (FACS Calibur; BD Bioscience). Results are expressed as a fold of the value of the control and each experiment was repeated at least in triplicate.

Detection of NADPH oxidase activity. HUVECs were plated at a density of 2×10^5 cells/well into 12-well plates and incubated with 0-20 μM As₂O₃ alone, 0-200 $\mu\text{g}/\text{ml}$ baicalin alone, or pre-treatment 0-200 $\mu\text{g}/\text{ml}$ of baicalin followed by co-treatment for 24 h, and were washed with PBS. Then cells were scraped from the plate in 1 ml of PBS and centrifuged for 10 min at 4°C at $9,100 \times g$. The supernatant was discarded and the pellet was resuspended to measure the activity of NADPH oxidase. The reaction buffer contained 100 mM of Tris-HCl, 1 mM of EDTA and 0.2 mM of NADPH. NADPH oxidase activity was calculated by monitoring the alteration in absorbance at 340 nm. Each experiment was repeated at least in triplicate.

Formamidopyrimidine DNA-glycosylase (FPG) and endonuclease III-incorporated comet assay. A glass microscope slide was placed on a hot plate at 60°C, and 100 μl of 1% normal-melting point agarose at 65°C was firstly applied. A coverslip was applied immediately and the slide was put on ice. Then the coverslips were removed. A second layer of agarose containing HUVECs in a volume of 80 μl was applied. This second layer was prepared by mixing 400 μl of 1.5% low-melting point agarose with 200 μl of PBS containing HUVECs. A coverslip was applied to each slide with HUVECs. After the gel had solidified, the coverslips were removed. In a similar manner as the first layer, 100 μl of 1.5% low-melting-point agarose was then applied as a third layer.

For enzyme digestion, as performed previously (32, 33), slides were washed after lysis treatment with distilled water then incubated at 37°C for 30 min in enzyme reaction buffer. Then one unit of FPG or one unit of endonuclease III in 10 μl of enzyme buffers supplied by Trevigen (Gaithersburg, MD, USA) [FDG: 10 mM HEPES-KOH (pH 7.4), 100 mM KCl; endonuclease III: 10 mM EDTA, 0.1 mg/ml bovine serum albumin; and 10 mM HEPES-KOH (pH 7.4), 100 mM KCl, and 10 mM EDTA] was added. A coverslip was applied and the slides were further incubated at 37°C for 2 h.

After removing coverslips, slides were immersed in cell lysis solution (2.5 M NaCl, 100 mM EDTA and 10 mM Tris, pH adjusted to 10.0 with NaOH, and 1% *N*-lauroylsarcosine, 1% Triton X-100, and 10% DMSO added immediately before use) at 4°C for 18 h. DNA was totally denatured by incubating in 0.3 M NaOH, 1 mM EDTA pH 13.4 for 20 min. Electrophoresis was performed at 25 V, 300 mA for 25 min using a tank. The slides were stained by adding 12.5 μl of 200x SYBR Green I. A coverslip was applied and the comets were observed under a fluorescence microscope (wavelength 450-490 nm, farb teiler 510 nm, long-pass filter 520 nm). The comet moment was calculated using the formula $\sum_{0 \rightarrow n} [(amount \text{ of DNA at distance } X) \times (distance \text{ } X)] / total \text{ DNA}$.

Statistical analysis. The results were expressed as the mean \pm standard error of the mean of at least three independent experiments in which triplicate samples were performed. Statistical significance was assessed using Student's *t*-test and one-way ANOVA followed by *post hoc* test using SPSS (version 15.0) software (SPSS Inc., Chicago, IL, USA). A value of $p < 0.05$ was considered statistically significant.

Results

As₂O₃-induced cytotoxicity and apoptosis were suppressed by baicalin in HUVECs. Originally, we found that As₂O₃ induced cell cytotoxicity, with dose-dependent reduction of cell

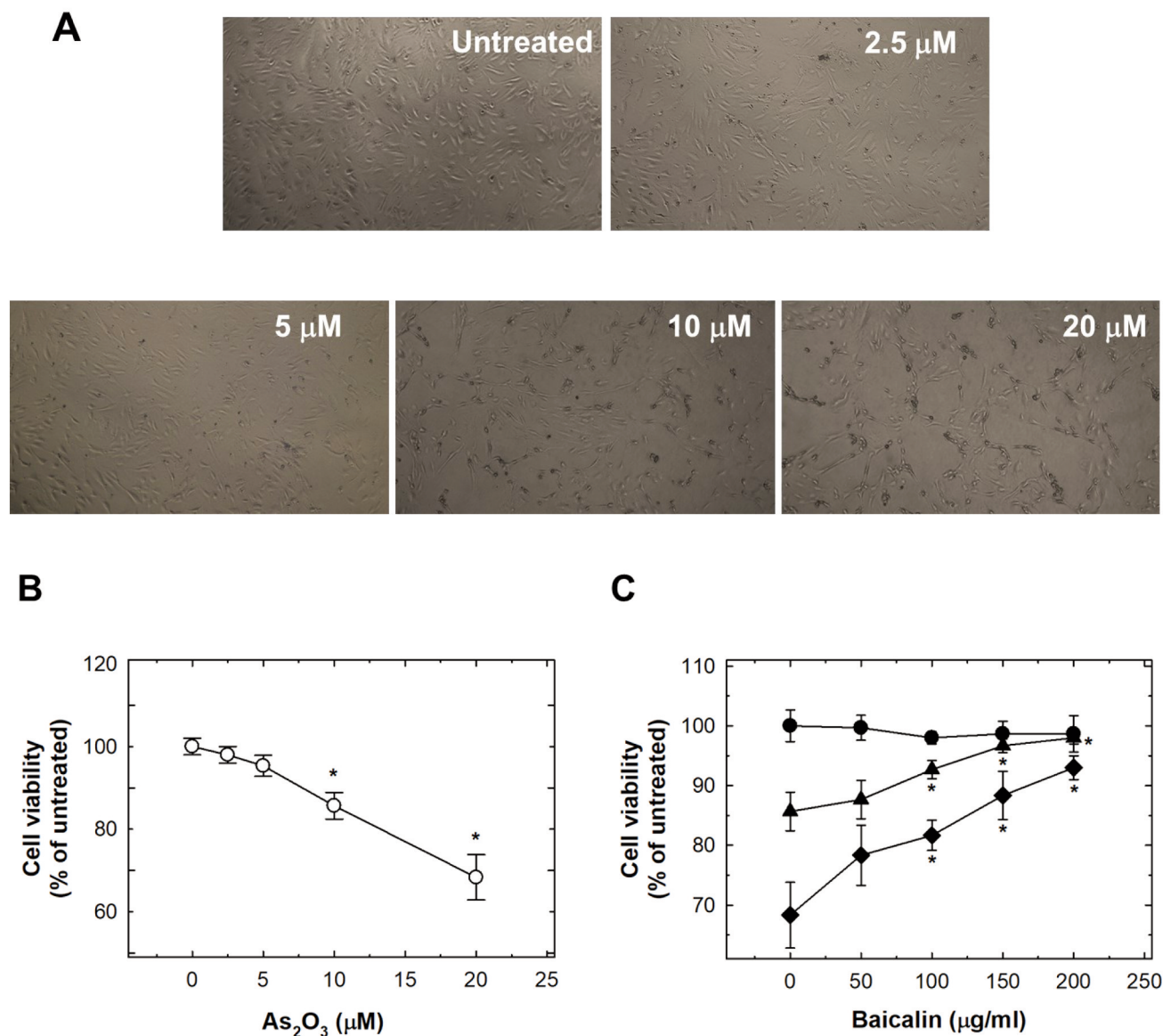


Figure 1. The effects of As₂O₃ and baicalin on viability of human umbilical vein endothelial cells (HUVECs). A: The effects of 24-h treatment of HUVECs with 0-20 μM As₂O₃. HUVECs were photographed under a microscope at ×40, representative images are shown. B: The quantification of the effect of 0-20 μM As₂O₃ treatment for 24 h in HUVECs. C: The effect on HUVEC viability after pretreatment with baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▲) or 0 μM (●) As₂O₃ for 24 h. Data are the mean±SD for three independent experiments. *Significantly different at $p < 0.05$ compared with untreated control cells.

viability (Figure 1A). Treatment of HUVECs with 10 μM and 20 μM reduced viability by 14.3% and 31.7%, respectively (Figure 1B). Pre-treatment with baicalin restored cell viability to almost normal levels (Figure 1C).

Treatment with As₂O₃ at 5 μM or more for 24 h was capable of inducing a significant sub G1 population of cells, *i.e.* significant apoptosis (Figure 2A). Pre-treatment with baicalin abrogated the activation of apoptosis dose-dependently (Figure 2B).

As₂O₃ induced oxidative stress in HUVECs which was suppressed by baicalin. As₂O₃ treatment dose-dependently induced significant ROS production at 15 min (Figure 3A). The peak ROS level was reached at 8 h, and plateau at 12 h (Figure 3A). The effect of As₂O₃ treatment at 10 and 20 μM on ROS was suppressed by pre-treatment with baicalin (Figure 3B). Exposure to As₂O₃ increased NADPH oxidase activity dose-dependently (Figure 4A). Pre-treatment with baicalin abrogated As₂O₃-induced activation of NADPH oxidase (Figure 4B).

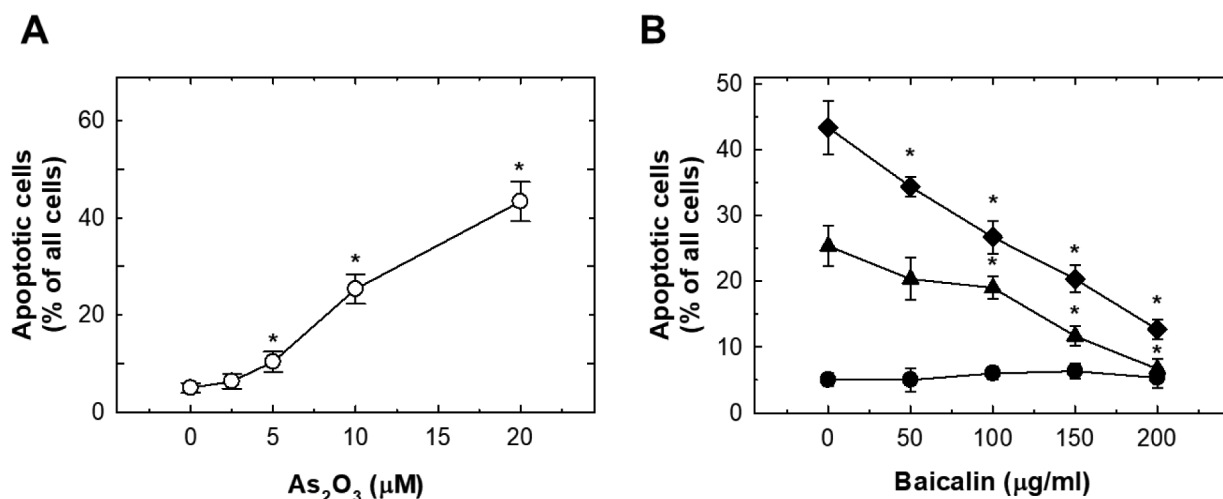


Figure 2. The effects of As₂O₃ and baicalin on apoptosis of human umbilical vein endothelial cells (HUVECs). A: The effects of 24-h treatment with 0-20 μM As₂O₃ on HUVECs. Apoptotic cells were detected by flow cytometry by the appearance of a sub-G₁ cell population. B: The effect on apoptosis of HUVECs after pretreatment with baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▲) or 0 μM (●) As₂O₃ for 24 h. Data are the mean±SD for three independent experiments. *Significantly different at p<0.05 compared with untreated cells.

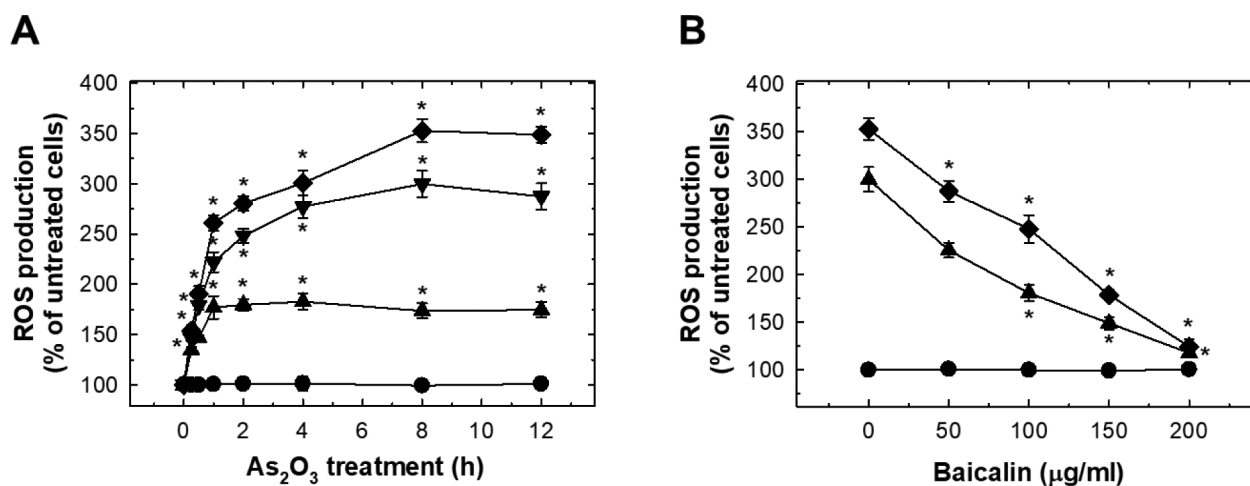


Figure 3. The effects of As₂O₃ and baicalin on reactive oxygen species (ROS) production in human umbilical vein endothelial cells (HUVECs). A: HUVECs were treated with 0, 5, 10, and 20 μM As₂O₃ for the indicated time, and ROS were detected with 2',7'-dichlorodihydrofluorescein diacetate. B: The effect on ROS production in HUVECs after pretreatment with baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▼), 5 μM (▲) or 0 μM (●) As₂O₃ for 12 h. Data are the mean±SD for three independent experiments. *Significantly different at p<0.05 compared with untreated cells.

As₂O₃ induced oxidative FPG- and endonuclease III-digestible DNA adducts in HUVECs which was suppressed by baicalin. As₂O₃ treatment induced a significant level of FPG- and endonuclease III-digestible adducts (Figures 5A and 6A). The levels of these oxidative adducts were effectively reduced by pre-treatment with baicalin but did not reach basal levels (Figures 5B and 6B).

Discussion

Vascular endothelium not only serves as a physical monolayer between blood and tissue but plays an important role in detection and reaction to blood-borne stresses. Impaired endothelial function maybe an irreversible process in the initiation event of various cardiovascular diseases and

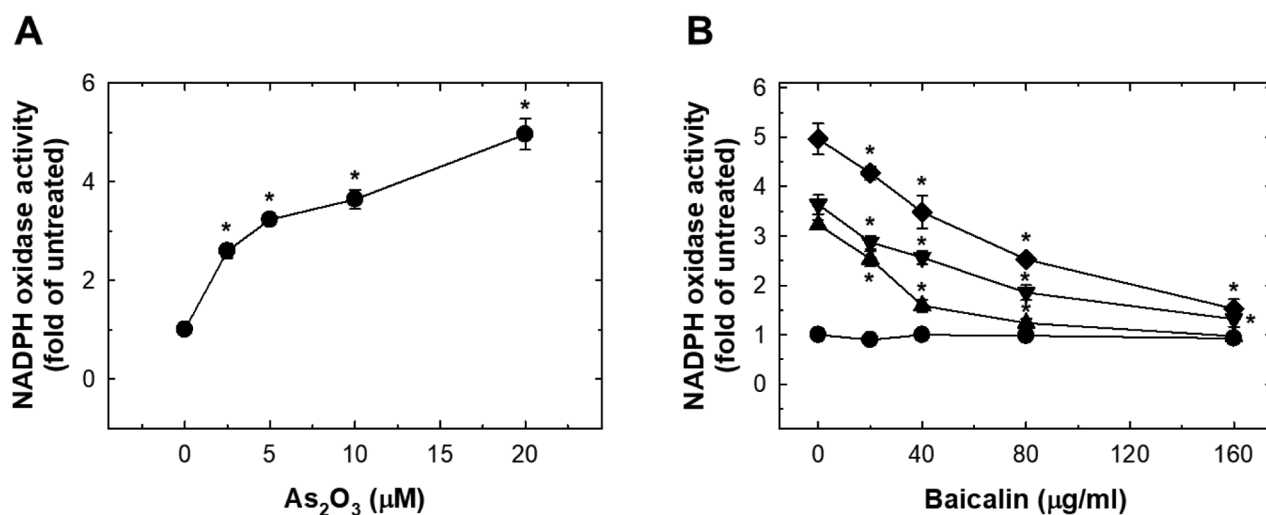


Figure 4. The effects of As₂O₃ and baicalin on NADPH oxidase activity in HUVECs. A: HUVECs were treated with 0-20 μM As₂O₃ for 24 h and NADPH oxidase activity was evaluated as fold that of untreated cells. B: The effect on NADPH oxidase activity in HUVECs after pretreatment of baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▼), 5 μM (▲) or 0 μM (●) As₂O₃ for 24 h. Data are the mean±SD for three independent experiments. *Significantly different at $p < 0.05$ compared with untreated cells.

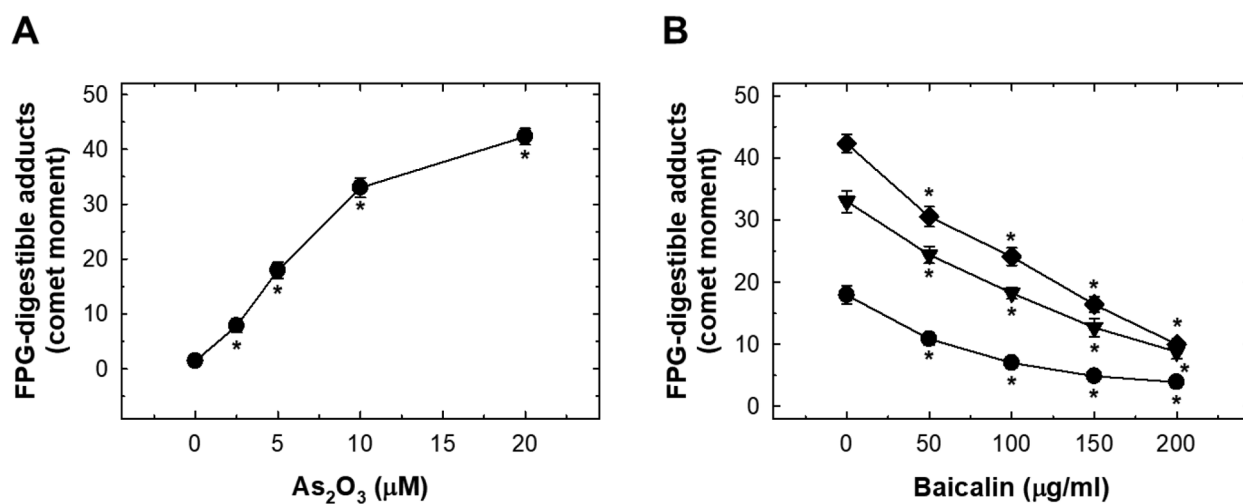


Figure 5. The effects of As₂O₃ and baicalin on formamidopyrimidine-DNA glycosylase (FPG)-digestible adducts in human umbilical vein endothelial cells (HUVECs). A: HUVECs were treated with 0-20 μM As₂O₃ for 24 h and FPG-digestible adducts produced were detected with comet assay. The data are presented as comet moments. B: The effect on FPG-digestible adducts in HUVECs after pretreatment with baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▼) or 0 μM (●) As₂O₃ for 24 h. Data are the mean±SD for three independent experiments. *Significantly different at $p < 0.05$ compared with untreated cells.

it is worth determining the practical cardio-protective effects of Chinese traditional medicine, such as baicalin. Arsenic exposure induced HUVEC dysfunction through pro-apoptotic and oxidative stress, which might be responsible for its adverse effects on blood vessels. The protective role of Chinese traditional medicine may be of great help in prevention of cardiovascular disorders. In this study, the

effects of baicalin on As₂O₃-induced endothelial cell apoptosis, oxidative stress and oxidative DNA adduct production were revealed for the first time.

Increased endothelial cell apoptosis may contribute to the loss of endothelial integrity and lead to vascular disorders (34, 35). Our study confirmed a significant increase in HUVEC apoptosis when cells were treated with As₂O₃,

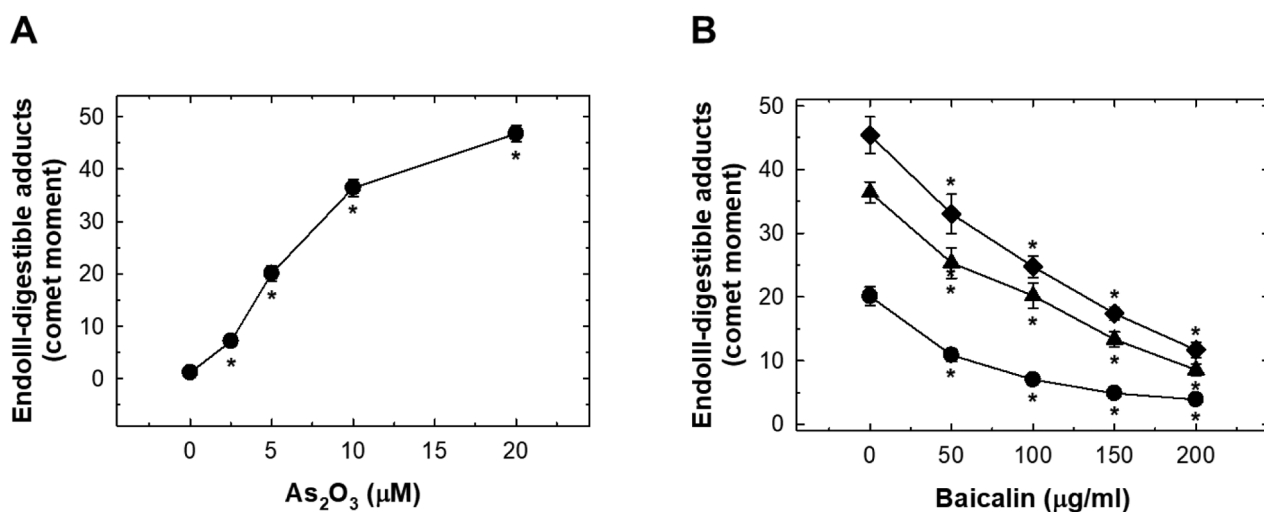


Figure 6. The effects of As₂O₃ and baicalin on endonuclease III-digestible adducts in human umbilical vein endothelial cells (HUVECs). A: HUVECs were treated with 0-20 μM As₂O₃ for 24 h and FPG-digestible adducts produced were detected with comet assay. The data are presented as comet moments. B: The effect on endonuclease III-digestible adducts in HUVECs after pretreatment with baicalin for 1 h, and co-treatment of baicalin with 20 μM (◆), 10 μM (▼) or 0 μM (●) As₂O₃ for 24 h. Data are the mean±SD for three independent experiments. *Significantly different at p<0.05 compared with untreated cells.

while pre-treatment with baicalin reduced As₂O₃-induced HUVEC apoptosis (Figure 2B). The degree of As₂O₃-induced apoptosis was not as strong as in Yu *et al.*'s finding, reporting that 5 μM As₂O₃ was capable suppressed cell viability by about 40% of (36). Our results are consistent with Ma *et al.*'s finding that 5 μM As₂O₃ induced apoptosis in less than 20% of HUVECs (37). The As₂O₃ induced generation of ROS was rapidly detected, and sustained for as long as 12 h (Figure 3A). Pre-treatment with baicalin did not alter the level of ROS itself but did suppress ROS production induced by As₂O₃. A number of risk factors for cardiovascular disorders were reported to be associated with increased vascular ROS (38, 39). Our studies also showed that As₂O₃ activated NADPH oxidase, which was reversed by pre-treatment with baicalin. Activation of NADPH oxidase may be the major mechanism related to As₂O₃-mediated intracellular generation of ROS.

One of the highlights of this was the detection of FPG-digestible adducts and endonuclease III-digestible adducts. FPG specifically cleaves oxidized purines, including 8-oxoguanine, 5-hydroxycytosine, 5-hydroxyuracil, 2,6-diamino-4-hydroxy-5-*N*-methylformamidopyrimidine, and 4,6-diamino-5-formamidopyrimidine (40), while endonuclease III specifically cleaves oxidized pyrimidines, including thymine glycol, 5,6-dihydrothymine, 5-hydroxydihydrothymine, 5-hydroxycytosine, 5-hydroxyuracil, and uracil glycol (41). We also demonstrated that baicalin pre-treatment reduced the formation of As₂O₃-induced oxidative adducts, although the detailed mechanisms of this need further investigation.

In conclusion, at a non-toxic dose, baicalin is of reducing the As₂O₃-induced apoptosis, oxidative stress and DNA adducts. The underlying mechanisms may provide novel insights into potential clinical cardioprotective effects.

Conflicts of Interest

The Authors declare no conflicts of interest in regard to this study.

Authors' Contributions

C.L.T. and C.W.T. conceived and designed the experiments. W.S.C. and C.W.T. performed the experiments. J.C.L. and T.C.H. analyzed the data. C.L.T. and J.C.L. contributed reagents/materials/analysis tools. C.W.T. and D.T.B. wrote the article.

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