

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect

Science of the Total Environment

journal homepage: www.elsevier.com/locate/scitotenv

Short Communication

Predicted occurrence, ecotoxicological risk and environmentally acquired resistance of antiviral drugs associated with COVID-19 in environmental waters



^a Department of Environmental and Civil Engineering, Toyama Prefectural University, Toyama 939 0398, Japan
 ^b Discipline of Earth Science, Indian Institute of Technology Gandhinagar, Gujarat 382 355, India

HIGHLIGHTS

GRAPHICAL ABSTRACT

- Environmental concentrations of antiviral drugs for COVID-19 were predicted.
 Many drugs and metabolites are not
- readily removed by wastewater treatment.
- Residues of several drugs may pose high ecotoxicological risk in receiving waters.
- Potential of environmental development of antiviral drug resistance is small.
- Proper usage and waste management of antiviral drugs are urgently needed.

ARTICLE INFO

Article history: Received 9 December 2020 Received in revised form 3 February 2021 Accepted 4 February 2021 Available online 15 February 2021

Editor: Damia Barcelo

Keywords: Antiviral drug resistance COVID-19 vaccine Pharmaceuticals QSAR SARS-COV-2



ABSTRACT

Antiviral drugs have been used to treat the ever-growing number of coronavirus disease, 2019 (COVID-19) patients. Consequently, unprecedented amounts of such drug residues discharging into ambient waters raise concerns on the potential ecotoxicological effects to aquatic lives, as well as development of antiviral drug-resistance in wildlife. Here, we estimated the occurrence, fate and ecotoxicological risk of 11 therapeutic agents suggested as drugs for COVID-19 treatment and their 13 metabolites in wastewater and environmental waters, based on drug consumption, physical-chemical property, and ecotoxicological and pharmacological data for the drugs, with the aid of quantitative structure-activity relationship (QSAR) modelling. Our results suggest that the removal efficiencies at conventional wastewater treatment plants will remain low (<20%) for half of the substances, and consequently, high drug residues (e.g. 7402 ng/L ribavirin, 4231 ng/L favipiravir, 730 ng/L lopinavir, 319 ng/L remdesivir; each combined for both unchanged forms and metabolites; and when each drug is administered to 100 patients out of 100,000 populations on a day) can be present in secondary effluents and persist in the environmental waters. Ecotoxicological risk in receiving river waters can be high (risk quotient >1) by a use of favipiravir, lopinavir, umifenovir and ritonavir, and medium (risk quotient >0.1) by a use of chloroquine, hydroxychloroquine, remdesivir, and ribavirin, while the risk will remain low (risk quotient <0.1) for dexamethasone and oseltamivir. The potential of wild animals acquiring antiviral drug resistance was estimated to be low. Our prediction suggests a pressing need for proper usage and waste management of antiviral drugs as well as for improving removal efficiencies of drug residues in wastewater.

© 2021 Elsevier B.V. All rights reserved.

* Corresponding author. *E-mail address:* kuroda@pu-toyama.ac.jp (K. Kuroda).





1. Introduction

Coronavirus disease, 2019 (COVID-19), a highly infectious disease caused by severe acute respiratory syndrome-related coronavirus (SARS-CoV-2), has been declared a pandemic, with 101 million confirmed cases and 2.2 million deaths worldwide as of February 2021 (WHO, 2020). As there are no specific therapeutic drugs recognised for targeting the cure from the SARS-CoV-2, various existing pharmaceuticals have been tested as therapeutic agents to treat COVID-19 patients ("The Race", 2020; Liu et al., 2020a). Studies suggest that remdesivir, an antiviral drug against Ebola, may be effective in shortening the time to recovery in adults hospitalized with COVID-19 (Beigel et al., 2020), and has been approved for COVID-19 treatment in countries such as US and Japan ("Japan Approves Remdesivir", 2020), Recently, low-dose dexamethaxone, a synthetic corticosteroid, has been suggested effective in reducing deaths in COVID-19 patients with ventilation, which is potentially a major breakthrough in COVID-19 treatment ("Dexamethasone Reduces Death", 2020). Several vaccines have been rapidly developed and their mass-delivery and uses have already been started in some countries, but there are many hurdles and uncertainties to overcome, such as logistics and public hesitancy ("Hope of a COVID-19 Vaccine", 2020). Therefore, the unprecedented mass use of these therapeutic drugs is expected to continue worldwide (Kumar et al., 2020a).

After human consumption, pharmaceuticals are excreted from human body and discharged into wastewater as unchanged drugs or metabolites, which are often only partly removed in conventional wastewater treatment plants (WWTPs) (Joss et al., 2005; Nannou et al., 2020). These residues present in receiving environmental waters have posed ecotoxicological concerns (Al Aukidy et al., 2012; Fick et al., 2010; Godoy and Kummrow, 2017; Santos et al., 2010). In particular, during pandemic events, high amounts of antiviral drugs and their metabolites released into environmental waters are likely to pose a high risk to aquatic ecosystems (Jain et al., 2013; Nannou et al., 2020). In addition, such high amounts of antiviral drugs in environmental waters may lead to the development of antiviral drug-resistant viral strains inside the body of specific wild animals, which are natural reservoirs of viruses (Kumar et al., 2020a). That is, when animal reservoirs continuously ingest environmental waters containing elevated levels of antiviral drugs and their metabolites, the viruses inside their bodies may develop resistance through rapid mutations (Jain et al., 2013; Nannou et al., 2020; Singer et al., 2007). We define this type of antiviral drug resistance as environmentally acquired antiviral drug resistance (EDR). EDR has been concerned for influenza A virus during past outbreaks (Fick et al., 2007; Ghosh et al., 2010; Singer et al., 2007). SARS-CoV-2 might similarly develop EDR inside animal hosts such as bats (Zhou et al., 2020), owing to expected mass use of antiviral drugs during the current waves of COVID-19 (Kumar et al., 2020a; Kumar et al., 2020b; Race et al., 2020; Sims and Kasprzyk-Hordern, 2020). To date, however, there has been no quantitative evaluation of EDR of SARS-CoV-2.

The occurrence, fate and ecotoxicity of a diverse range of pharmaceuticals, including antiviral drugs, in WWTPs and in environmental waters during past outbreaks as well as normal times have been summarized (Aymerich et al., 2016; Jain et al., 2013; Kasprzyk-Hordern et al., 2009; Nannou et al., 2020; Ncube et al., 2018; Tran et al., 2018). However, those past studies hardly include most of the therapeutic agents which have been considered for COVID-19 treatment, except for oseltamivir (anti-influenza drug) (Azuma et al., 2012; Fick et al., 2007; Ghosh et al., 2010; Prasse et al., 2010; Singer et al., 2007) and lopinavir/ritonavir (anti-HIV drugs) (Abafe et al., 2018; Wood et al., 2015). The potential occurrence and fate in aquatic environments, the general physical-chemical properties, and ecotoxicological risks of various COVID-19-associated drugs are largely unknown. For screening environmental fate and toxicity of pharmaceuticals, quantitative structure-activity relationship (QSAR) models have been applied to diverse pharmaceuticals (Escher et al., 2011; Kar et al., 2020; Sanderson et al., 2004).

The objective of the study is to provide a model-based evaluation on the occurrence, fate and ecotoxicological effects of a suite of therapeutic agents associated with COVID-19 treatment and their metabolites in wastewater and environmental waters during pandemic events. Predicted environmental concentrations (PECs) were calculated with assumed patient numbers treated with these drugs (100 patients out of 100 k populations are on the course of treatment every day), taking into account drug consumption patterns, excretion from human body, and elimination at WWTPs. QSAR models were used to predict elimination at WWTPs and chronic toxicity to aquatic lives for the substances for which measurement-based data were not available. Furthermore, potential of EDR by animal reservoirs was assessed by in vitro pharmacological data of the drugs against SARS-CoV-2. To our knowledge, this is the first study to estimate ecotoxicological impacts of mass use of multi-antiviral drugs associated with COVID-19 on ambient waters and suggest necessary global precautionary measures.

2. Materials and methods

We evaluated 11 representative potential therapeutic drugs for COVID-19 treatment (chloroquine, dexamethaxone, favipiravir, hydroxychloroquine, lopinavir, oseltamivir, remdesivir, ribavirin, ritonavir, teicoplanin and umifenovir) and their 13 major metabolites (Table 1), which were selected from literature (Liu et al., 2020a; Wu et al., 2020; Yousefi et al., 2020). The drugs' original purposes are shown in Table 1, and their CAS number and simplified molecular-input line-entry system (SMILES) in Table S1 in the Supplementary data.

2.1. Predicted environmental concentrations

The concentrations of the target substances in raw wastewater, secondary effluent, and river waters were predicted by the following Eqs. (1)-(3), which were adapted from past modelling studies on antivirals and/or down-the-drain chemicals (Singer et al., 2007; Ghosh et al., 2010; Keller et al., 2014):

$$\text{PEC}_{\text{raw}} = \frac{N_{\text{t}}}{100,000} \times \frac{D_{\text{d}} \times f \times 10^{6}}{W_{\text{c}}} \tag{1}$$

$$PEC_{se} = PEC_{raw} \times (1 - R)$$
⁽²⁾

$$PEC_{riv} = \frac{PEC_{se}}{10}$$
(3)

where PEC_{raw} is a predicted concentration in raw wastewater; N_t is a number of patients on the course of treatment with a drug per 100,000 population in a day (assumed as 100); D_d is an average daily drug dose expected for COVID-19 treatment; f is a fraction of excreted substances (to urine and feces) to drug dose; W_c is water consumption per person per day of 200 L, which has been used by European Medicine Agency (EMA) for environmental risk assessment of pharmaceuticals (EMA, 2018); 10⁶ is a conversion factor from mg of substances to ng; PEC_{se} is a predicted concentration in secondary effluent; R is removal efficiency in conventional WWTPs (mentioned below); and PEC_{riv} is a predicted concentration in rivers.

The average daily drug dose D_d ranged from 6 mg/day for dexamethasone to 2473 mg/day for ribavirin. The details of drug dose can be found in Table S1. The fraction of excretion (f), identified based on literature and database search, varied largely, ranging from 0.8% to 83% for unchanged drugs and from 1.5% to 80% for metabolites (Table 1). We assumed dilution of secondary effluent by ten times in the receiving rivers, which represents a minimum dilution in many countries (Keller et al., 2014) and was also used for environmental risk

Table 1

Predicted physical-chemical properties and ecotoxicological characteristics of potential therapeutic agents for COVID-19.

Drugs and metabolites	Original purpose	Average daily dose, D _d (mg/day) ^a	Substances and excreted fraction, f (%) ^b		Antiviral activity	M.W. ^p	LogK _{ow}	PNEC (ng/L)	Removal in WWTP, R	Primary biodegradation
Chloroquine	Malaria	343	Chloroquine (urine and	50% ^c	Active	319.9	4.63 ^q	3700 ^s	63%	weeks to
			feces)							months
-metabolite			N-desethylchloroquine (urine)	10% ^c	Unknown	291.8	3.79	55	22%	days to weeks
Dexamethasone	Corticosteroid	6	Dexamethasone	10% ^d	Active	392.5	1.92 ^q	50 ^t	2.2%	weeks to months
Favipiravir	Influenza	1.600	Favipiravir (urine)	0.8% ^e	Prodrug	157.1	0.72	91	1.9%	days to weeks
-metabolite	minucinbu	1,000	T705M1 (urine)	53.1% ^e	Inactive	173.1	0.99	81	1.9%	days to weeks
Hydroxychloroquine	Malaria	354	Hydroxychloroquine (urine and feces)	47% ^f	Active	335.9	3.03	170	6.0%	weeks to months
Lopinavir	HIV	800	Lopinavir (mostly feces)	22% ^g	Active	628.8	5.94	4.7	92%	days to weeks
-metabolite			M1 (mostly feces)	71% in total ^g	Unknown	642.8	5.54	5.9	89%	days to weeks
-metabolite			M2 (mostly feces)		Unknown	644.8	3.48	30	71%	days to weeks
-metabolites			M3/M4 (mostly feces)		Unknown	644.8	3.46	30	71%	days to weeks
Oseltamivir	Influenza	150	Oseltamivir (urine and feces)	15% ^h	Prodrug	312.4	0.95	4700	1.9%	days to weeks
-metabolite			Oseltamivir carboxylate	80% ^h	Active	284.4	0.18	120000	1.9%	hours to days
Remdesivir	Ebora	110	Remdesivir (urine)	10% ⁱ	Active	602.6	1 74	31	2.1%	days to weeks
-metabolite	Loona	110	GS-451524 (urine)	49% ⁱ	Active	2913	-1.76	240	1.9%	days to weeks
Ribavirin	HCV. RSV	2473	Ribavirin (urine)	17% ^j	Active	244.2	-1.85^{q}	2700	1.9%	hours to days
-metabolite			TCONH ₂ (urine)	44% ^{j, k}	Inactive	112.1	-1.37	830	1.9%	days to weeks
Ritonavir	HIV	200	Ritonavir (mostly feces)	37% ^l	Active	720.9	6.27	2.9	93%	days to weeks
-metabolite			M2 (mostly feces)	60% ^l	Active	736.9	5.17	20	82%	days to weeks
Teicoplanin	Antibiotic	400	Teicoplanin (urine and feces)	83% ^m	Active	1879.7	-1.1 ^r	n.a. ^u	0% ^v	n.a.
Umifenovir	Influenza, SARS	600	Umifenovir (feces)	40% ⁿ	Active	477.4	5.4	9.3	87%	weeks to months
-metabolite			M10 (feces)	3%°	Unknown	556.5	2.91	160	5.0%	weeks to months
-metabolite			M18 (urine)	1.5%	Unknown	653.5	3.34	25000	87%	weeks to months
-metabolite			M20 (urine)	2.1%°	Unknown	669.5	0.76	240000	1.9%	days – weeks

^a Average daily dose (mg) was calculated as the total amount of a drug for expected use for COVID-19 treatment, divided by expected treatment duration (see Table S1).

^b Excretion (%) is the amount, expressed as a fraction of dose, of a parent drug (unchanged drug) or its metabolites which are eliminated from human body via urine and feces. The excretion data were obtained from literature and drug database search.

^c Ducharme and Farinotti (1996).

^d FDA Approved Drug Products: Hemady Dexamethasone Oral Tablets (2019).

^e Ministry of Health, Labour and Welfare (2014).

^f Browning (2014).

^g Health Canada (2019). The fraction of each of four metabolites of lopinavir (M1 to M4) is not available, thus the sum of total metabolite fractions was evaluated.

^h He et al. (1999).

ⁱ FDA: Fact Sheet For Health Care Providers EUA of Remdesivir (2020).

^j FDA Approved Drug Products: Rebetol (ribavirin) oral capsules (2019).

^k Lin et al. (2006).

¹ FDA Approved Drug Products: NORVIR (ritonavir) Capsules, Soft Gelatin for Oral use (2012).

^m Electronic Medicines Compendium: Targocid (teicoplanin) 400 mg powder Monograph (2020).

ⁿ Liu et al. (2009).

^o Deng et al. (2013).

^p Molecular weight.

^q Experimentally determined (US EPA ECOTOX knowledgebase, 2020, https://cfpub.epa.gov/ecotox/)

^r Experimentally determined (Rowland, 1990)

^s Based on Zurita et al., 2005; eEC₅₀ for *D. magna* at a 72h exposure.

^t Based on DellaGreca et al., 2004; chronic toxicity for *C. dubia* at a 7d exposure.

^u Not available.

^v The removal efficiency of teicoplanin in WWTP was not predictable by EPISuite, thus a removal efficiency of 0% was assumed.

assessment by EMA (2018). To give a conservative estimation, no instream attenuation was assumed.

2.2. Physical-chemical properties

Removal efficiency in conventional WWTPs (employing activated sludge process as secondary treatment) was obtained as 'total removal at STPs' predicted by STPWIN program in EPI Suite™ (EPA, 2021). LogK_{ow} was searched for experimentally derived octanol-water distribution coefficient (K_{ow}), but it was available only for chloroquine, dexamethasone, ribavirin and teicoplanin (Table 1). For the other drugs and metabolites, LogK_{ow} was estimated by Kowwin v.1.68 in EPI Suite™. Considering its importance in determining environmental fate, LogK_{ow} was also calculated

by SPARC program (Hilal et al., 2003) for comparison. Kowwin modelling is based on a database of substances with known K_{ow} , whereas SPARC program calculates strictly from molecular structure (Hilal et al., 2003). If there is more than an order of magnitude difference in K_{ow} between the two programs, the ALOGPS 2.1 program by VCC labs (VCC Laboratory, 2009) was tested, and the K_{ow} from either Kowwin or SPARC closer to the value predicted by AGLOPS program was used. The LogK_{ow} values evaluated by Kowwin, SPARC and ALOGPS 2.1 are shown in Table S2.

2.3. Ecotoxicity

Chronic toxicity of the target substances was evaluated, either using experimentally derived ecotoxicity (when available) or otherwise using predicted ecotoxicity by ECOSAR, a computerized structure activity relationship for aquatic toxicity (EPA, 2020). Experimentally derived ecotoxicity data was searched by US EPA Ecotox knowledgebase (https://cfpub.epa.gov/ecotox/) and Google Scholar, and was obtained for only chloroquine (Zurita et al., 2005) and dexamethasone (DellaGreca et al., 2004). As ecotoxicity of chloroquine was obtained for only acute toxicity, the median effective concentration (EC₅₀; hereafter, denoted as *eEC*₅₀ to differentiate from viral inhibitory concentration) was converted to chronic toxicity by acute-to-chronic ratio of 10 (Mayo-Bean et al., 2017). For the remaining substances, chronic ecotoxicity was predicted by ECOSAR, and the smallest values of chronic ecotoxicity for three model organisms (daphnia, algae and fish) were taken for a conservative estimate. For each substance, the predicted no-effect concentration (PNEC) was estimated as the chronic toxicity value divided by UF, a standard uncertainty factor, as shown in Eq. (4); the UF value of 1000 was conventionally adopted to consider the intra- and interspecies variability in the sensitivity (Hernando et al., 2006):

$$PNEC = \frac{eEC_{50}}{1000} \tag{4}$$

In addition, the mode of action in aquatic organisms was predicted by VEGA (2019) for each substance.

Risk quotient (RQ) was calculated for each substance as the ratio between PEC_{riv} and PNEC, as shown in Eq. (5):

$$RQ = \frac{PEC_{riv}}{PNEC}$$
(5)

The risk is classified into three levels: RQ 0.01–0.1, low risk; RQ 0.1–1, medium risk; and RQ >1, high risk (Hernando et al., 2006).

2.4. Environmentally acquired antiviral drug resistance

The drug concentration which inhibits *in vitro* viral growth by 50% (the half maximal inhibitory concentration; IC_{50}) is a measure of susceptibility of viruses to antiviral agents (Pillay and Zambon, 1998),

|--|

Summary of determined vIC₅₀ and vEC₅₀ of antiviral drugs against SARS-CoV-2.

and it can also be expressed as half maximal effective concentration (EC₅₀); here, we denote IC_{50} and EC_{50} of antiviral agents as vIC_{50} and vEC_{50} to differentiate from ecotoxicological median effective concentration eEC₅₀.

The likelihood of developing antiviral resistance by a virus is the largest when the drug concentrations are close to vIC_{50} (Pillay and Zambon, 1998). Thus, we evaluated the potential of EDR by animal reservoirs exposed to environmental waters, by defining *EDR potential* (EDRP) as the minimum values between the ratio of PEC_{riv} to vIC_{50} values of an antiviral drug and its reciprocal (Eq. (6)):

$$EDRP = Min\left(\frac{PEC_{riv}}{vIC_{50}}, \frac{vIC_{50}}{PEC_{riv}}\right)$$
(6)

By definition, EDRP of 1 is the maximal value. The vIC₅₀ and vEC₅₀ values of the target pharmaceuticals determined *in vitro* against SARS-CoV-2 were summarized from literature (Table 2). Note that, the determined vIC₅₀/vEC₅₀ values varied by an order of magnitude, depending on experimental conditions (e.g., multiplicity of infection: MOI (Liu et al., 2020b), time after infection of test cells (Gonçalves et al., 2020)). To make a conservative estimate, the lowest vIC₅₀/vEC₅₀ values were used for EDRP calculation. Metabolites are also evaluated for EDRP, assuming the same vIC₅₀ values as those of the parent substances.

3. Results and discussion

3.1. Physical-chemical properties and environmental fate

The predicted physical-chemical properties of the target substances are summarized in Table 1. Approximately half of the parent drugs (6/11, 54%) and the metabolites (6/12, 50%) were found to be hydrophilic (LogK_{ow} <3). These hydrophilic substances mostly have low molecular weight (mw <400), but a few substances had high molecular weight (e.g., remdesivir, mw 602.6; teicoplanin, mw 1709.4; umifenovir M10, mw 556.5; and umifenovir M20, mw 669.5) but low LogK_{ow} values (1.74, -1.10, 2.91 and 0.76, respectively). The predicted removal in conventional WWTPs was low for the half of the

Antiviral drugs	vIC ₅₀ /vEC ₅₀ (µM)	vIC ₅₀ /vEC ₅₀ used for EDRP calculation				
		μΜ	Converted to µg/L			
Chloroquine	1.03 (Holwerda et al., 2020)	1.03	329			
	1.13 (Wang et al., 2020a)					
	1.31 (Ohashi et al., 2020)					
	5.47 (Yao et al., 2020)					
	2.71–7.36 (Liu et al., 2020b)					
	7.28, 12.0 (Jeon et al., 2020)					
	9.27 (Xiong et al., 2020)					
Favipiravir	62 (Wang et al., 2020a)	62	9740			
	>500 (Jeon et al., 2020)					
Hydroxychloroquine	0.72 (Yao et al., 2020)	0.72	242			
	4.51–12.96 (Liu et al., 2020b)					
	9.21–11.17 (Weston et al., 2020)					
Lopinavir	1.73 (Ohashi et al., 2020)	1.73	1088			
	4.9–5.2 (Gonçalves et al., 2020)					
	5.73 (Yamamoto et al., 2020)					
	9.12, 15.27 (Jeon et al., 2020)					
Oseltamivir	>100 (Tan and Jin, 2020)	100	31,200			
	>100 (Wang et al., 2020b)					
Remdesivir	0.77 (Wang et al., 2020a)	0.77	464			
	1.842 (Holwerda et al., 2020)					
	2.5 (Liu et al., 2020c)					
	8.24, 11.41 (Jeon et al., 2020)					
Ribavirin	109.5 (Wang et al., 2020a)	109.5	26,740			
Ritonavir	8.63 (Yamamoto et al., 2020)	8.63	6222			
Teicoplanin	1.66 (Zhang et al., 2020)	1.66	3120			
Umifenovir	4.11 (Wang et al., 2020b)	4.11	1962			
	30 (Lu, 2020)					



Fig. 1. Comparison between LogK_{ow} values and removal efficiencies at WWTPs, both predicted by EPI Suite™. Note that removal efficiency of teicoplanin was not predictable by EPI Suite™, thus was assumed as 0.

substances (removal efficiency <20% for 12 substances), whereas high removal efficiency (>80%) was predicted for only six substances (chloroquine, lopinavir, ritonavir, umifenovir, and two metabolites). The predicted high removal efficiency would be largely associated with adsorptive behavior of the substances; predicted LogK_{ow} values and predicted removal efficiencies were expressed in a sigmoid-like growth curve (Fig. 1). In addition, biodegradability at WWTPs predicted by STPWIN ('Biodegradation in STP') was only less than 0.77%. 'Primary biodegradation', which indicates the time required for the transformation of a substance to an initial metabolite (EPA, 2021), was 'days to weeks' and 'weeks to months' for most of the target substances.

Measurement-based removal efficiencies of the target substances during activated sludge treatment processes were largely unavailable, except for oseltamivir, oseltamivir carboxylate (the active metabolite of oseltamivir), lopinavir and ritonavir. Regarding oseltamivir and oseltamivir carboxylate, the predicted removal efficiencies at conventional WWTPs (2% for both) were in accordance with their low removal efficiencies determined during activated sludge treatment (none for oseltamivir carboxylate: Fick et al., 2007; 10% for both substances: Azuma et al., 2012; none for oseltamivir and 59% for oseltamivir carboxylate: Prasse et al., 2010). Regarding lopinavir and ritonavir, high removal efficiencies (92% and 93%, respectively) were predicted. In comparison, their measurement-based removal efficiencies in two municipal WWTPs in South Africa (employing activated sludge with nutrient removal, anaerobic digestion, followed by maturation ponds) somewhat differed (43% to 71% and -192% to -58%, respectively (Abafe et al., 2018)); however, this results could be treated with care, because the wastewater samples were taken by grab sampling. Favipiravir was predicted to be persistent during activated sludge process (2% removal); this is in accordance with its persistence against biodegradation in a batch-scale experiment (Azuma et al., 2017). Note that, favipiravir was easily degraded by sunlight in the latter study, indicating a rapid decrease in environmental waters.

3.2. Predicted occurrence in wastewater and environmental waters

The large concentrations in secondary effluents were predicted for $TCONH_2$ (5339 ng/L), the major active metabolite of ribavirin, followed by T705M1 (4168 ng/L; the major inactive metabolite of favipiravir) and ribavirin (2063 ng/L), as shown in Table 3. On the contrary, low PECs in secondary effluents were predicted for dexamethasone (2.9 ng/L), ritonavir (26 ng/L) and remdesivir (54 ng/L), because of low dose (dexamethasone), high removal at activated sludge process (ritonavir) and high rate of transformation to metabolites (for remdesivir, with

265 ng/L of the active metabolite, GS-451524). For all substances, concentrations in the river waters were lower by a factor of 10, because of assumed dilution.

As for oseltamivir, the PEC in secondary effluents in this study (118 ng/L and 589 ng/L) was similar with the maximum concentrations in treated wastewater (293 ng/L to 672 ng/L) determined during pandemic events in Japan (Azuma et al., 2017; Azuma et al., 2012; Ghosh et al., 2010), but lower than the predicted concentrations of oseltamivir carboxylate in UK and US rivers of 31.8 µg/L during the peak of influenza outbreaks (Singer et al., 2007). Favipiravir has been rarely detected in wastewater effluents after activated sludge process and in river waters in Japan during the past influenza season, presumably because of low usage of favipiravir to influenza patients in Japan and low excretion unchanged (0.8%) (Azuma et al., 2017). Lopinavir was abundant in wastewater in South Africa (1200-2500 in influents, 130-3800 ng/L in effluents)(Abafe et al., 2018; Wood et al., 2015). Concentrations of ribavirin were below limit of quantification in raw wastewater and treated wastewater in Germany and China (Peng et al., 2014; Prasse et al., 2010). Ritonavir has been determined in wastewater in South Africa (mean 1600-3200 ng/L) (Abafe et al., 2018), treated hospital effluent in Switzerland (max 108 ng/L)(Kovalova et al., 2012), and surface water in France (max. 12 ± 5 ng/L)(Aminot et al., 2015). The predicted concentrations of lopinavir and ritonavir in raw wastewater were several times lower than the abovementioned high concentrations of lopinavir and ritonavir in wastewater in South Africa, probably owing to the daily usage for HIV treatment in South Africa (Abafe et al., 2018). The occurrence of the other substances has not been determined in environmental waters.

Clearly, the PECs of the target substances greatly depend on N_t, the number of treated patients per 100 k population in a day. Since expected treatment duration is 5-10 days for most therapeutics in this study, the assumed N_t value of 100 would imply that there are additional 10–20 patients treated by a drug per 100 k population each day. As of February 2021, multiple countries have reported more than 10 new daily confirmed cases per 100 k population; for example, Gibraltar (364), Belgium (143), Portugal (126), Switzerland (98), France (78), the US (75), Spain (74), Germany (69), Qatar (64), Italy (64), Chile (46), South Africa (32), Brazil (27), and Russia (20) (7-day moving average; WHO, 2020). These numbers would have been much larger at regional/county level; in the US, 7-day moving average of daily new cases per 100 k population has been up to more than 500 in 88 counties (1.63 million population in total), and up to more than 1000 in 34 counties (0.74 million population in total) as of Feb 1, 2020 (USAFacts US Coronavirus Cases and Deaths, 2021). Considering these numbers and the ratio of severely or critically ill patients of COVID-19 (19%) (Wu and McGoogan, 2020), the predicted number of patients given in this study can be a likely scenario in many parts of the world, and the number can be even larger in areas with high infection rates.

3.3. High predicted ecotoxicological risk for favipiravir, lopinavir and umifenovir

Ritonavir showed the highest chronic toxicity to aquatic organisms (PNEC 2.9 ng/L; Table 3), followed by lopinavir (4.7 ng/L), lopinavir M1 (5.9 ng/L), umifenovir (9.3 ng/L) and ritonavir M2 (20 ng/L). Ritonavir has been widely concerned for its high ecotoxicological risk because of its exceptionally high hydrophobicity (Escher et al., 2011). PNEC of the other substances were predicted at more than 30 ng/L, up to 120,000 ng/L.

Consequently, high ecotoxicological risk was suggested in the receiving river waters for favipiravir-T705M1 (RQ 5.2), metabolites of lopinavir (0.78–3.9), umifenovir (1.7) and lopinavir (1.5) (Table 3). Medium risk was predicted for seven substances: ritonavir (0.89) and its metabolite M2 (0.53), TCONH₂, the major metabolite of ribavirin (0.64), hydroxychloroquine (0.47), *N*-desethylchloroquine, the major metabolite of chloroquine (0.25), remdesivir (0.17), and GS-451524,

Table 3

Predicted Environmental Concentration (PEC), Predicted No Effect Concentration (PNEC), ecological Risk Quotients (RQ) and Environmentally acquired antiviral Drug Resistance Potential (EDRP) of potential therapeutic agents for COVID-19 in wastewater and environmental waters.

Substances	PEC (ng/L)			PNEC	RQ			EDRP (x 10^{-3})			
	Raw wastewater	Secondary effluent	River	(ng/L)	Raw wastewater	Secondary effluent	River	Raw wastewater	Secondary effluent	River	
Chloroquine	857	320	32	3700	0.23	0.086	0.0086	2.6	0.97	0.097	
N-desethylchloroquine	171	135	13	55	3.1	2.5	0.25	0.57	0.45	0.045	
Dexamethasone	3.0	2.9	0.29	50	0.060	0.058	0.0060	n.a. ^c	n.a.	n.a.	
Favipiravir	64	63	6.3	91	0.71	0.69	0.069	0.0066	0.0064	0.00064	
T705M1	4248	4168	417	81	53	52	5.2	0.40	0.39	0.039	
Hydroxychloroquine	833	783	78.3	170	5.0	4.7	0.47	3.4	3.2	0.32	
Lopinavir	880	71	7.1	4.7	190	15	1.5	0.81	0.066	0.0066	
Lopinavir-M1	2840 ^a	659 ^a	66 ^a	5.9	96-480 ^b	7.8–39 ^b	0.78-3.9 ^b	2.6 ^d	0.59 ^d	0.059 ^d	
Lopinavir-M2				30							
Lopinavir-M3/M4				30							
Oseltamivir	113	110	11	4700	0.024	0.023	0.0023	0.0035	0.0035	0.00035	
Oseltamivir	600	589	59	120000	0.0049	0.0048	0.00048	0.021	0.021	0.0021	
carboxylate											
Remdesivir	55	54	5.4	31	1.8	1.7	0.17	0.12	0.12	0.012	
GS-451524	270	265	26	240	1.1	1.1	0.11	1.2	1.2	0.12	
Ribavirin	2102	2063	206	2700	0.77	0.75	0.075	0.079	0.077	0.0077	
TCONH ₂	5440	5339	534	830	6.5	6.4	0.6	0.44	0.44	0.044	
Ritonavir	373	26	2.6	2.9	128	8.9	0.89	0.060	0.0042	0.00042	
Ritonavir-M2	604	106	11	20	30	5.3	0.53	0.095	0.017	0.0017	
Teicoplanin	1654	1654	165	n.a.	n.a.	n.a.	n.a.	0.53	0.53	0.053	
Umifenovir	1200	157	16	9.3	130	17	1.7	0.61	0.080	0.0080	
Umifenovir-M10	90	86	8.6	160	0.58	0.55	0.055	0.039	0.037	0.0037	
Umifenovir-M18	45	6	0.6	25000	0.0018	0.00023	0.000023	0.017	0.0022	0.00022	
Umifenovir-M20	63	62	6.2	240000	0.00027	0.00026	0.000026	0.023	0.022	0.0022	

^a The fraction of each of the four metabolites of lopinavir (M1 to M4) was not available, thus the sum of the four metabolites is shown.

^b The fraction of each of the four metabolites of lopinavir (M1 to M4) was not available, thus the range of RQ is shown, using the maximum and the minimum PNEC to the sum of four metabolites.

^c n.a.; not available. EDRP of dexamethasone was not available because it is a corticosteroid, not an antiviral agent.

^d The fraction of each of the four metabolites of lopinavir (M1 to M4) was not available, thus the EDRP was evaluated with the sum of the four metabolites.

the major metabolite of remdesivir (0.11). Low risk was predicted for the remaining nine substances: chloroquine, dexamethasone, favipiravir, oseltamivir, oseltamivir carboxylate, ribavirin and metabolites of umifenovir. The predicted high ecological risk posed by lopinavir and ritonavir in this study is in accordance with a previous study, in which the two antiretroviral drugs dominated RO in hospital effluents (Escher et al., 2011). Our predictions, however, suggests an appreciably high ecotoxicological risk posed by the metabolites of remdesivir and favipiravir, which have been recommended for the treatment of COVID-19 in several countries. Prediction by VEGA shows that narcosis is the major mode of action in aquatic organisms for all the substances, except for remdesivir, ritonavir and ritonavir-M2 (acetylcholinesterase inhibition), and ribavirin (reactivity). Apart from these antiviral drugs, high ecotoxicity by narcosis has also been suggested for some pharmaceuticals derived from domestic and hospital effluents (e.g. diclofenac) in receiving waters (Al Aukidy et al., 2012; Escher et al., 2011); thus, the ecotoxicity of these pharmaceuticals can be added up in the receiving waters and lead to even higher ecotoxicity. Overall, significant ecological adverse effects can be posed in river waters receiving the discharge of antiviral drugs and metabolites.

3.4. Environmentally acquired antiviral drug resistance potential (EDRP) of SARS-CoV-2 in the natural reservoirs

For all the antiviral drugs in this study, the risk of EDRP against SARS-CoV-2 appears to be insignificant, because there are at least three orders of magnitude difference between PEC and vLC₅₀/vEC₅₀ for all substances (Tables 3). In river waters, largest EDRP was found for hydroxychloroquine (0.00032), followed by GS-451524 (the major active metabolite of remdesivir; 0.00012), and chloroquine (0.00097). The small EDRP in the present study was primarily due to the large vLC₅₀/vEC₅₀ values of the therapeutic drugs in this study against SARS-CoV-2 (0.72 to >100 μ M; 242 to >31,200 μ g/L). While in the case of

influenza, vLC₅₀ of oseltamivir carboxylate, the active form of oseltamivir, is much smaller (e.g., 0.28–0.81 nM; 80–230 ng/L) (Gubareva et al., 2001; Monto et al., 2006). Hence, environmental concentrations of oseltamivir carboxylate can be comparable to vLC₅₀ during influenza outbreak, suggesting a significant risk of EDR in the body of water fowls (the natural reservoir of influenza virus) in wastewater-impacted water bodies (Azuma et al., 2012; Fick et al., 2007; Ghosh et al., 2010; Jain et al., 2013; Nannou et al., 2020; Singer et al., 2007).

Regardless of the small EDRP as above, we must note that numerous populations of wild or domestic animals potentially possess SARS-CoV-2; coronaviruses are known to circulate in mammals, and various animals can be direct or intermediate host for SARS-CoV-2. In the case of SARS-CoV-2, bats have been suggested as animal reservoirs as they carry a coronavirus named RaTG13, which is genetically 96.2% identical to SARS-CoV-2 (Zhou et al., 2020). Pangolins also have coronaviruses similar to SARS-CoV-2 (Lam et al., 2020), but they are unlikely the reservoir and they likely acquired these coronaviruses after spillover from the natural hosts (Hu et al., 2021). Besides bats and pangolins, some of domestic or cultured animals such as cats, ferrets and minks are susceptible to SARS-CoV-2, and infections between individuals have been observed for cats and minks (Oreshkova et al., 2020; Shi et al., 2020). In the past coronavirus-outbreaks, palm civets were found to be the intermediate host animals for SARS-CoV, and dromedary camels for MERS-CoV (Hu et al., 2021).

Therefore, regardless of the small EDRP as above, it is recommended that residues of antiviral drugs in wastewater must be reduced. In domestic/cultured settings, wastewater-impacted waters should not be given to animals which are susceptible to SARS-CoV-2. Similarly to antiviral drugs, excessive usage of therapeutic or non-therapeutic antimicrobials has been a matter of concern over disruption of natural biological systems as well as development of antimicrobial resistance in the aquatic systems (Usman et al., 2020).

3.5. Uncertainties

We acknowledge following uncertainties in our predictions. First, usage of each drug would differ depending on regulatory status, drug characteristics (e.g., dosage form, utility, adverse reactions) and patients' health conditions. There are also cases where practitioners and general public are using more than one drug for their own precautions from COVID-19, perhaps owing to a lack of reliable guidelines on specific drug usage for COVID-19. Such practice would result in even higher amount of drugs and their metabolites releasing into the environment, further exacerbating the ecotoxicological impacts. Second, the QSAR models used in this study are supposed to be only for screening analysis of chemicals (EPA, 2021). Therefore, further studies are required for precise evaluations of chemical properties, environmental behavior and ecotoxicity of the substances. Third, the prediction we provided is for a given snapshot concentrations of drug residues, which needs to be substantiated through time-course analysis of drug concentrations in ambient waters during pandemic events, as was done for oseltamivir during an influenza outbreak (Singer et al., 2007). In terms of spatial distribution of drug residues in environmental waters, specific facilities (i.e. hospitals or quarantined hotels and residences) can be important point sources, to which a large proportion of symptomatic patients are often transferred. The impact of such specific medical facilities on environmental pharmaceutical discharge can be particularly significant in small catchments (Kuroda et al., 2016).

4. Conclusions

In the fight against COVID-19, medication is obviously essential in saving human lives and speeding up the recovery. Meanwhile, the potential negative environmental impacts of increased drug usage should not be overlooked. Our study suggests the following:

- 1. Conventional WWTPs are not capable of efficiently eliminating (removal efficiency <20%) dexamethasone, favipiravir, hydroxychloroquine, oseltamivir, remdesivir, ribavirin and their metabolites from raw wastewater. Therefore, effluents from conventional WWTPs may contain high concentrations of these drugs and their metabolites (up to 7402 ng/L, combined for ribavirin and its metabolite TCONH₂), potentially posing high risk to aquatic lives.
- 2. High risk quotients in effluent-receiving rivers are predicted for T705M1, a metabolite of favipiravir (RQ 5.2), metabolites of lopinavir (0.78–3.9), umifenovir (1.7) and lopinavir (1.5). Use of chloroquine, hydroxychloroquine, remdesivir, ribavirin and ritonavir also implies medium ecotoxicological risk (RQ >0.1) by the parent compounds or their metabolites in rivers.
- 3. EDR is less concerning for SARS-CoV-2, because PECs of the antiviral drugs in rivers are more than a thousand times smaller than reported vEC_{50}/vLC_{50} values of antiviral drugs against SARS-CoV-2. Nevertheless, efforts to reduce environmental discharge of antiviral drugs and their metabolites are important in terms of EDR prevention, as there are numerous populations of SARS-CoV-2-susceptible animals.

In order to address these issues, proper usage and management of antiviral drugs, and proper management of unused pharmaceuticals must be shared and implemented. Direct disposal of drugs into wastewater must be avoided, and using wastewater-impacted waters for animals must be refrained. In the long term, upgrading WWTPs with advanced treatments, such as ozonation, must be facilitated to efficiently remove diverse pharmaceuticals. On-site treatment of hospital effluents can also be effective in reducing environmental discharge of pharmaceuticals. Proper collection and treatment of wastewater in developing communities are a challenge, and thus additional investment is necessary. In order to facilitate such discussions, measurementbased evaluation of occurrence, fate, and ecotoxicological risk of various therapeutics associated with COVID-19 in WWTPs and environmental waters is urgently warranted in many parts of the world.

CRediT authorship contribution statement

Keisuke Kuroda: Conceptualization, Visualization, Investigation, Data curation, Writing – original draft. **Cong Li:** Formal analysis. **Kiran Dhangar:** Investigation. **Manish Kumar:** Visualization, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This study was financially supported by Kurita Water and Environment Foundation (KWEF) research grant (No. 198078).

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi. org/10.1016/j.scitotenv.2021.145740.

References

- Abafe, O.A., Späth, J., Fick, J., Jansson, S., Buckley, C., Stark, A., et al., 2018. LC-MS/MS determination of antiretroviral drugs in influents and effluents from wastewater treatment plants in KwaZulu-Natal, South Africa. Chemosphere 200, 660–670.
- Al Aukidy, M., Verlicchi, P., Jelic, A., Petrovic, M., Barcelò, D., 2012. Monitoring release of pharmaceutical compounds: occurrence and environmental risk assessment of two WWTP effluents and their receiving bodies in the Po Valley, Italy. Sci. Total Environ. 438, 15–25.
- Aminot, Y., Litrico, X., Chambolle, M., Arnaud, C., Pardon, P., Budzindki, H., 2015. Development and application of a multi-residue method for the determination of 53 pharmaceuticals in water, sediment, and suspended solids using liquid chromatographytandem mass spectrometry. Anal. Bioanal. Chem. 407, 8585–8604.
- Andy Cowper: Hope of a covid-19 vaccine is good news, but comes with many uncertainties. The BMJopinion, 2020.
- Aymerich, I., Acuña, V., Barceló, D., García, M.J., Petrovic, M., Poch, M., et al., 2016. Attenuation of pharmaceuticals and their transformation products in a wastewater treatment plant and its receiving river ecosystem. Water Res. 100, 126–136.
- Azuma, T., Nakada, N., Yamashita, N., Tanaka, H., 2012. Synchronous dynamics of observed and predicted values of anti-influenza drugs in environmental waters during a seasonal influenza outbreak. Environmental Science & Technology 46, 12873–12881.
- Azuma, T., Ishida, M., Hisamatsu, K., Yunoki, A., Otomo, K., Kunitou, M., et al., 2017. Fate of new three anti-influenza drugs and one prodrug in the water environment. Chemosphere 169, 550–557.
- Beigel, J.H., Tomashek, K.M., Dodd, L.E., Mehta, A.K., Zingman, B.S., Kalil, A.C., et al., 2020. Remdesivir for the treatment of Covid-19 – preliminary report. N. Engl. J. Med. 383, 1813–1826.
- Browning, D.J., 2014. Hydroxychloroquine and Chloroquine Retinopathy. Springer US, New York.
- DellaGreca, M., Fiorentino, A., Isidori, M., Lavorgna, M., Previtera, L., Rubino, M., et al., 2004. Toxicity of prednisolone, dexamethasone and their photochemical derivatives on aquatic organisms. Chemosphere 54, 629–637.
- Deng, P., Zhong, D., Yu, K., Zhang, Y., Wang, T., Chen, X., 2013. Pharmacokinetics, metabolism, and excretion of the antiviral drug arbidol in humans. Antimicrob. Agents Chemother. 57 (4), 1743–1755.
- Dexamethasone reduces death in hospitalised patients with severe respiratory complications of COVID-19. University of Oxford, 2020.
- Ducharme, J., Farinotti, R., 1996. Clinical pharmacokinetics and metabolism of chloroquine. Focus on recent advancements. Clin. Pharmacokinet. 31, 257–274.
- Electronic Medicines Compendium: Targocid (teicoplanin) 400 mg powder Monograph, 2020. https://www.medicines.org.uk/emc/medicine/27321.
- Escher, B.I., Baumgartner, R., Koller, M., Treyer, K., Lienert, J., McArdell, C.S., 2011. Environmental toxicology and risk assessment of pharmaceuticals from hospital wastewater. Water Res. 45, 75–92.
- European Medicine Agency (EMA) Guideline on the environmental risk assessment of medicinal products for human use, 2018.
- FDA Approved Drug Products: Hemady Dexamethasone Oral Tablets, 2019. https://www. accessdata.fda.gov/drugsatfda_docs/label/2019/211379s000lbl.pdf.
- FDA Approved Drug Products: NORVIR (ritonavir) Capsules, Soft Gelatin for Oral use, 2012. https://www.accessdata.fda.gov/drugsatfda_docs/label/2012/020945s033lbl. pdf.
- FDA Approved Drug Products: Rebetol (ribavirin) oral capsules, 2019. https://www. accessdata.fda.gov/drugsatfda_docs/label/2019/020903s055,021546s011lbl.pdf.
- Fick, J., Lindberg, R.H., Tysklind, M., Haemig, P.D., Waldenström, J., Wallensten, A., et al., 2007. Antiviral oseltamivir is not removed or degraded in normal sewage water

treatment: implications for development of resistance by influenza a virus. PLoS One 2, e986.

- Fick, J., Lindberg, R.H., Tysklind, M., Larsson, D.G.J., 2010. Predicted critical environmental concentrations for 500 pharmaceuticals. Regul. Toxicol. Pharmacol. 58, 516–523.
- Ghosh, G.C., Nakada, N., Yamashita, N., Tanaka, H., 2010. Oseltamivir carboxylate, the active metabolite of oseltamivir phosphate (Tamiflu), detected in sewage discharge and river water in Japan. Environ. Health Perspect. 118, 103–107.
- Godoy, A.A., Kummrow, F., 2017. What do we know about the ecotoxicology of pharmaceutical and personal care product mixtures? A critical review. Crit. Rev. Environ. Sci. Technol. 47, 1453–1496.
- Gonçalves A, Bertrand J, Ke R, Comets E, de Lamballerie X, Malvy D, et al. Timing of antiviral treatment initiation is critical to reduce SARS-CoV-2 viral load. CPT: Pharmacometrics & Systems Pharmacology 2020; Sep; 9(9): 509–514.
- Gubareva, L.V., Webster, R.G., Hayden, F.G., 2001. Comparison of the activities of Zanamivir, Oseltamivir, and RWJ-270201 against clinical isolates of influenza virus and neuraminidase inhibitor-resistant variants. Antimicrob. Agents Chemother. 45, 3403–3408.
- He, G., Massarella, J., Ward, P., 1999. Clinical pharmacokinetics of the Prodrug Oseltamivir and its active metabolite Ro 64-0802. Clin. Pharmacokinet. 37 (6), 471–484.
- Health Canada (2019) Product Monograph: Kaletra (lopinavir/ritonavir) for oral use. https://pdf.hres.ca/dpd_pm/00053305.PDF.
- Hernando, M.D., Mezcua, M., Fernández-Alba, A.R., Barceló, D., 2006. Environmental risk assessment of pharmaceutical residues in wastewater effluents, surface waters and sediments. Talanta 69, 334–342.
- Hilal SH, Karickhoff SW, Carreira L. PREDICTION OF CHEMICAL REACTIVITY PARAMETERS AND PHYSICAL PROPERTIES OF ORGANIC COMPOUNDS FROM MOLECULAR STRUC-TURE USING SPARC, 2003.
- Holwerda M, V'kovski P, Wider M, Thiel V, Dijkman R. Identification of five antiviral compounds from the Pandemic Response Box targeting SARS-CoV-2. bioRxiv 2020: 2020.05.17.100404.
- Hu, B., Guo, H., Zhou, P., Shi, Z.-L., 2021. Characteristics of SARS-CoV-2 and COVID-19. Nat. Rev. Microbiol. 19, 141–154.
- Jain S, Kumar P, Vyas RK, Pandit P, Dalai AK. Occurrence and Removal of Antiviral Drugs in Environment: A Review. Water, Air, & Soil Pollution 2013; 224: 1410.
- Japan Approves Remdesivir For Use On Severe COVID-19 Patients. Forbes, 2020.
- Jeon, S., Ko, M., Lee, J., Choi, I., Byun, S.Y., Park, S., et al., 2020. Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs. Antimicrob. Agents Chemother. 64, e00819–e00820.
- Joss, A., Keller, E., Alder, A.C., Göbel, A., McArdell, C.S., Ternes, T., et al., 2005. Removal of pharmaceuticals and fragrances in biological wastewater treatment. Water Res. 39, 3139–3152.
- Kar, S., Sanderson, H., Roy, K., Benfenati, E., Leszczynski, J., 2020. Ecotoxicological assessment of pharmaceuticals and personal care products using predictive toxicology approaches. Green Chem. 22, 1458–1516.
- Kasprzyk-Hordern, B., Dinsdale, R.M., Guwy, A.J., 2009. The removal of pharmaceuticals, personal care products, endocrine disruptors and illicit drugs during wastewater treatment and its impact on the quality of receiving waters. Water Res. 43, 363–380.
- Keller, V.D.J., Williams, R.J., Lofthouse, C., Johnson, A.C., 2014. Worldwide estimation of river concentrations of any chemical originating from sewage-treatment plants using dilution factors. Environ. Toxicol. Chem. 33, 447–452.
- Kovalova, L., Siegrist, H., Singer, H., Wittmer, A., McArdell, C.S., 2012. Hospital wastewater treatment by membrane bioreactor: performance and efficiency for organic micropollutant elimination. Environmental Science & Technology 46, 1536–1545.
- Kumar, M., Kuroda, K., Dhangar, K., Mazumder, P., Sonne, C., 2020a. Rinklebe J, et al. Potential Emergence of Antiviral-Resistant Pandemic Viruses via Environmental Drug Exposure of Animal Reservoirs, Environmental Science & Technology.
- Kumar, M., Mazumder, P., Mohapatra, S., Kumar Thakur, A., Dhangar, K., Taki, K., et al., 2020b. A chronicle of SARS-CoV-2: seasonality, environmental fate, transport, inactivation, and antiviral drug resistance. J. Hazard. Mater. 124043.
- Kuroda, K., Itten, R., Kovalova, L., Ort, C., Weissbrodt, D.G., McArdell, C.S., 2016. Hospitaluse pharmaceuticals in Swiss Waters modeled at high spatial resolution. Environmental Science & Technology 50, 4742–4751.
- Lam, T.T.-Y., Jia, N., Zhang, Y.-W., Shum, M.H.-H., Jiang, J.-F., Zhu, H.-C., et al., 2020. Identifying SARS-CoV-2-related coronaviruses in Malayan pangolins. Nature 583, 282–285.
- Lin, C.C., Xu, C., Zhu, N., Yeh, L.T., 2006. Absorption, metabolism, and excretion of [14C] Viramidine in humans. Antimicrob. Agents Chemother. 50 (7), 2368–2373.
- Liu, M.Y., Wang, S., Yao, W.F., Wu, H.Z., Meng, S.N., Wei, M.J., 2009. Pharmacokinetic properties and bioequivalence of two formulations of arbidol: an open-label, single-dose, randomized-sequence, two-period crossover study in healthy chinese male volunteers. Clin. Ther. 31 (4), 784–792.
- Liu, C., Zhou, Q., Li, Y., Garner, L.V., Watkins, S.P., Carter, L.J., et al., 2020a. Research and Development on therapeutic agents and vaccines for COVID-19 and related human coronavirus diseases. ACS Central Science 6, 315–331.
- Liu, J., Cao, R., Xu, M., Wang, X., Zhang, H., Hu, H., et al., 2020b. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. Cell Discovery 6, 16.
- Liu S, Lien CZ, Selvaraj P, Wang TT. Evaluation of 19 antiviral drugs against SARS-CoV-2 Infection. bioRxiv 2020c: 2020.04.29.067983.
- Lu, H., 2020. Drug treatment options for the 2019-new coronavirus (2019-nCoV). BioScience Trends 14, 69–71.
- Mayo-Bean, K., Moran-Bruce, K., Meylan, W., Ranslow, P., Lock, M., Nabholz, J.V., et al., 2017. Methodology Document for the Ecological Structure-Activity Relationship Model (ECOSAR) Class Program.
- Ministry of Health, Labour and Welfare, 2014. Report on the deliberation results. March 4, 2014 (Avigan Tablet 200 mg). https://www.pmda.go.jp/files/000210319.pdf.

- Monto, A.S., McKimm-Breschkin, J.L., Macken, C., Hampson, A.W., Hay, A., Klimov, A., et al., 2006. Detection of influenza viruses resistant to neuraminidase inhibitors in global surveillance during the first 3 years of their use. Antimicrob. Agents Chemother. 50, 2395–2402.
- Nannou, C., Ofrydopoulou, A., Evgenidou, E., Heath, D., Heath, E., Lambropoulou, D., 2020. Antiviral drugs in aquatic environment and wastewater treatment plants: a review on occurrence, fate, removal and ecotoxicity. Sci. Total Environ. 699, 134322.
- Ncube, S., Madikizela, L.M., Chimuka, L., Nindi, M.M., 2018. Environmental fate and ecotoxicological effects of antiretrovirals: a current global status and future perspectives. Water Res. 145, 231–247.
- Ohashi H, Watashi K, Saso W, Shionoya K, Iwanami S, Hirokawa T, et al. Multidrug treatment with nelfinavir and cepharanthine against COVID-19. bioRxiv 2020: 2020.04.14.039925.
- Oreshkova N, Molenaar RJ, Vreman S, Harders F, Oude Munnink BB, Hakze-van der Honing RW, et al. SARS-CoV-2 infection in farmed minks, the Netherlands, April and May 2020. Eurosurveillance 2020; 25: 2001005.
- Peng, X., Wang, C., Zhang, K., Wang, Z., Huang, Q., Yu, Y., et al., 2014. Profile and behavior of antiviral drugs in aquatic environments of the Pearl River Delta, China. Sci. Total Environ. 466–467, 755–761.
- Pillay, D., Zambon, M., 1998. Antiviral drug resistance. BMJ 317, 660-662.
- Prasse, C., Schlüsener, M.P., Schulz, R., Ternes, T.A., 2010. Antiviral drugs in wastewater and surface waters: a new pharmaceutical class of environmental relevance? Environmental Science & Technology 44, 1728–1735.
- Race, M., Ferraro, A., Galdiero, E., Guida, M., Núñez-Delgado, A., Pirozzi, F., et al., 2020. Current emerging SARS-CoV-2 pandemic: potential direct/indirect negative impacts of virus persistence and related therapeutic drugs on the aquatic compartments. Environ. Res. 188, 109808.
- Rowland, M., 1990. Clinical Pharmacokinetics of Teicoplanin. Clinical Pharmacokinetics 18, 184–209.
- Sanderson, H., Johnson, D.J., Reitsma, T., Brain, R.A., Wilson, C.J., Solomon, K.R., 2004. Ranking and prioritization of environmental risks of pharmaceuticals in surface waters. Regul. Toxicol. Pharmacol. 39, 158–183.
- Santos, L.H.M.L.M., Araújo, A.N., Fachini, A., Pena, A., Delerue-Matos, C., Montenegro, M.C.B.S.M., 2010. Ecotoxicological aspects related to the presence of pharmaceuticals in the aquatic environment. J. Hazard. Mater. 175, 45–95.
- Shi J, Wen Z, Zhong G, Yang H, Wang C, Huang B, et al. Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS-coronavirus 2. Science (New York, N.Y.) 2020; 368: 1016–1020.
- Sims, N., Kasprzyk-Hordern, B., 2020. Future perspectives of wastewater-based epidemiology: monitoring infectious disease spread and resistance to the community level. Environ. Int. 139, 105689.
- Singer, A.C., Nunn, M.A., Gould, E.A., Johnson, A.C., 2007. Potential risks associated with the proposed widespread use of Tamiflu. Environ. Health Perspect. 115, 102–106.
- Tan Q, Jin Y. Ostavimir is ineffective against COVID-19: in silico assessment, in vitro and retrospective study. medRxiv 2020: 2020.05.15.20102392.
- The race against COVID-19. Nature Nanotechnology. 15, 2020, pp. 239-240.
- Tran, N.H., Reinhard, M., Gin, K.Y.-H., 2018. Occurrence and fate of emerging contaminants in municipal wastewater treatment plants from different geographical regions-a review. Water Res. 133, 182–207.
- US EPA ECOlogical Structure-Activity Relationship Model (ECOSAR) Version 2.0. United States Environmental Protection Agency, Washington, DC, USA., 2020.
- US EPA ECOTOX knowledgebase, 2020. https://cfpub.epa.gov/ecotox/.
- US EPA Estimation Programs Interface SuiteTM for Microsoft® Windows, 4.11. United States Environmental Protection Agency, Washington, DC, USA., 2021.
- USAFacts US Coronavirus Cases and Deaths, 2021. https://usafacts.org/visualizations/coronavirus-covid-19-spread-map/.
- Usman, M., Farooq, M., Hanna, K., 2020. Environmental side effects of the injudicious use of antimicrobials in the era of COVID-19. Sci. Total Environ. 745, 141053. VCC Laboratory. ALOGPS 2.1, 2009.
- VEGA QSAR, 2019. Istituto di Ricerche Farmacologiche Mario Negri IRCCS. Laboratory of Environmental Chemistry and Toxicology.
- Wang, M., Cao, R., Zhang, L., Yang, X., Liu, J., Xu, M., et al., 2020a. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. Cell Res. 30, 269–271.
- Wang, X., Cao, R., Zhang, H., Liu, J., Xu, M., Hu, H., et al., 2020b. The anti-influenza virus drug, arbidol is an efficient inhibitor of SARS-CoV-2 in vitro. Cell Discovery 6, 28.
- Weston S, Coleman CM, Haupt R, Logue J, Matthews K, Frieman MB. Broad anticoronaviral activity of FDA approved drugs against SARS-CoV-2 in vitro and SARS-
- CoV in vivo. bioRxiv 2020: 2020.03.25.008482.
- WHO (2020) WHO Coronavirus Disease (COVID-19) Dashboard.
- Wood, T.P., Duvenage, C.S.J., Rohwer, E., 2015. The occurrence of anti-retroviral compounds used for HIV treatment in South African surface water. Environ. Pollut. 199, 235–243.
- Wu, Z., McGoogan, J.M., 2020. Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China: summary of a report of 72 314 cases from the Chinese Center for Disease Control and Prevention. JAMA 323, 1239–1242.
- Wu, R., Wang, L., Kuo, H.-C.D., Shannar, A., Peter, R., Chou, P.J., et al., 2020. An update on current therapeutic drugs treating COVID-19. Current Pharmacology Reports 6, 56–70.
- Xiong H-L, Cao J-L, Shen C-G, Ma J, Qiao X-Y, Shi T-S, et al. Several FDA-approved drugs effectively inhibit SARS-CoV-2 infection in vitro. bioRxiv 2020: 2020.06.05.135996.
- Yamamoto N, Matsuyama S, Hoshino T, Yamamoto N. Nelfinavir inhibits replication of severe acute respiratory syndrome coronavirus 2 in vitro. bioRxiv 2020: 2020.04.06.026476.
- Yao, X., Ye, F., Zhang, M., Cui, C., Huang, B., Niu, P., et al., 2020. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of

severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Clin. Infect. Dis. 71 (15), 732–739.

- Yousefi, B., Valizadeh, S., Ghaffari, H., Vahedi, A., Karbalaei, M., Eslami, M., 2020. A global treatments for coronaviruses including COVID-19. J. Cell. Physiol. 235 (12), 9133–9142.
- Zhang J, Ma X, Yu F, Liu J, Zou F, Pan T, et al. Teicoplanin potently blocks the cell entry of 2019-nCoV. bioRxiv 2020: 2020.02.05.935387.
- Zhou, P., Yang, X.-L., Wang, X.-G., Hu, B., Zhang, L., Zhang, W., et al., 2020. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 579, 270–273.
- Zurita, J.L. Jos, Á., Ad, Peso, Salguero, M., López-Artíguez, M., Repetto, G., 2005. Ecotoxicological evaluation of the antimalarial drug chloroquine. Aquat. Toxicol. 75, 97–107.