



OPEN

Eosinopenia and neutrophil-to-lymphocyte count ratio as prognostic factors in exacerbation of COPD

Tomasz Karauda¹, Kamil Kornicki², Amer Jarri³, Adam Antczak²,
Joanna Miłkowska-Dymanowska¹, Wojciech J. Piotrowski¹, Sebastian Majewski¹,
Paweł Górski¹ & Adam Jerzy Białas³✉

Exacerbations of Chronic Obstructive Pulmonary Disease (AECOPDs) are one of the most important clinical aspects of the disease, and when requiring hospital admission, they significantly contribute to mortality among COPD patients. Our aim was to assess the role of eosinopenia and neutrophil-to-lymphocyte count (NLR) as markers of in-hospital mortality and length of hospitalization (LoH) among patients with ECOPD requiring hospitalization. We included 275 patients. Eosinopenia was associated with in-hospital deaths only when coexisted with lymphocytopenia, with the specificity of 84.4% (95% CI 79.6–88.6%) and the sensitivity of 100% (95% CI 35.9–100%). Also, survivors presented longer LoH ($P < 0.0001$). $NLR \geq 13.2$ predicted in-hospital death with the sensitivity of 100% (95% CI 35.9–100%) and specificity of 92.6% (95% CI 88.8–95.4%), however, comparison of LoH among survivors did not reach statistical significance ($P = 0.05$). Additionally, when we assessed the presence of coexistence of eosinopenia and lymphocytopenia first, and then apply NLR, sensitivity and specificity in prediction of in-hospital death was 100% (95% CI 35.9–100) and 93.7% (95% CI 90.1–96.3), respectively. Moreover, among survivors, the occurrence of such pattern was associated with significantly longer LoH: 11 (7–14) vs 7 (5–10) days ($P = 0.01$). The best profile of sensitivity and specificity in the prediction of in-hospital mortality in ECOPD can be obtained by combined analysis of coexistence of eosinopenia and lymphocytopenia with elevated NLR. The occurrence of a such pattern is also associated with significantly longer LoH among survivors.

Exacerbations of chronic obstructive pulmonary disease (AECOPDs) are one of the most important clinical aspects to the disease. The clinical manifestations of COPD exacerbations are highly variable and reflect broad heterogeneity in the pathobiology of the disease. Regardless of its variability, exacerbations are clearly associated with negative impact on the health status, increasing rates of hospitalization, readmission, and disease progression^{1–3}. AECOPDs requiring hospital admission are the leading cause of hospitalization and significantly contribute to mortality among COPD patients⁴. Therefore, research on prognostic factors in this group is of special interest.

With a great deal of reasonable criticism, eosinopenia was already proposed as a marker of infection, in differentiating infectious from non-infectious causes of elevated C-reactive protein (CRP) and identifying sepsis or bacteremia⁵. Eosinopenia was also proposed to be a predictor of a short or long-term survival in some diseases, including AECOPD^{6–10}. Thus, we reanalyzed the problem of eosinopenia in AECOPD patients. Additionally, we assessed another previously discussed, and logically linked with eosinopenia, prognostic factor—neutrophil-to-lymphocyte count (NLR). This parameter has also been studied as a marker of sepsis and predictor of bacteremia⁵. The roles of both these parameters in AECOPD still remain ambiguous and controversial. The aim of this study was to assess the role of eosinopenia and NLR as markers of in-hospital mortality and length of hospitalization. The relationships between both parameters and their possible pathophysiological background were analyzed as well.

¹Department of Pneumology and Allergy, Medical University of Lodz, Lodz, Poland. ²Department of General and Oncological Pulmonology, Medical University of Lodz, Lodz, Poland. ³Department of Pathobiology of Respiratory Diseases, Medical University of Lodz, 22nd Kopcińskiego Street, 90-153 Lodz, Poland. ✉email: adam.bialas@umed.lodz.pl

Patients and methods

This study is a retrospective analysis of the data collected in the digital base of the Barlicki Memorial Teaching Hospital of the Medical University of Lodz. The protocol of the study was approved by the institutional ethics committee. Also, due to retrospective character of the study, in which consent would be impossible to obtain, concordantly with the Declaration of Helsinki, the Committee waived the need of such consent.

All methods in the study were performed in accordance with the relevant guidelines and regulations.

Enrollment of our study included only adult Caucasians with the exacerbation of COPD, diagnosed concordantly with current GOLD recommendations³. We included only exacerbations that required hospital admission, both of infectious and non-infectious character. Full blood count and white blood cell differentiation were examined in these patients, which was performed on the hospital admission. Venous blood was collected by venipuncture into tubes with ethylenediaminetetraacetic acid as an anticoagulant. The samples were examined with the automated hematology analyzer. Cell counting was performed using electrical impedance method. Arterial blood gases parameters were analyzed using automated analyzer.

The exclusion criteria were any other known chronic lung disease, any hematological disorder—active, or in the past medical history, any active malignancy, exacerbation of respiratory symptoms associated with other acute causes. We also excluded patients with clinically and radiologically proven pneumonia and those, who received any dose of systemic corticosteroids prior to the admission. Additionally, we excluded all patients who were transferred to the hospital by an ambulance, because retrospective character of the study would preclude reliable verification that these patients did not receive corticosteroids prior to the hospital admission.

AECOPD was defined as an event characterized by a rapid decline in the patient's respiratory symptoms that is beyond normal day-to-day variations and lead to changes in medication. In-hospital mortality was defined as any AECOPD-related death after the hospital admission.

NLR was calculated as a ratio of absolute counts of peripheral blood neutrophils to lymphocytes.

Length of hospitalization was assessed among survivors to avoid false shortage of hospitalization time in case of early in-hospital death.

Continuous data was presented as the mean with SD or median with interquartile range (IQR), depending on the distribution of data. In comparing multiple groups, one-way ANOVA with pairwise comparisons using *t* tests with pooled SD or Kruskal–Wallis rank sum test with pairwise comparisons using Mann–Whitney *U* test was used according to tests assumptions. Bonferroni method was used for *P* value adjustment. Receiver operating characteristics (ROC) with area under the ROC curve (AUROC) analyses were performed to measure the accuracy of absolute eosinophil count and NLR in prediction of in-hospital mortality and to identify its cut-off values for further analysis. Variables were compared using the unpaired Student's *t*-test, Welch *t*-test or the Wilcoxon rank sum test with continuity correction, depending on data normality and homogeneity of variance. Categorical data were presented as absolute value and percentage. Such data were compared using Chi-square test or Fisher's exact test according to test assumptions. Correlation analysis was performed using Kendall's rank correlation tau. Statistical analysis was performed using R software¹¹.

Results

General characteristics. We included 275 patients. Five patients (1.82%) died in hospital. Baseline population data are presented in Table 1. Majority of patients suffered from infectious AECOPD (*n* = 146; 53.09%). This group did not differ significantly in eosinophil count from non-infectious AECOPD [0.07 (0.02–0.18) vs 0.1 (0.03–0.21); *P* = 0.09]. However, in infectious AECOPD we observed significantly higher NLR values [4.3 (1.89–9.27) vs 2.6 (1–4.93); *P* = 0.0003].

The ROC curves for absolute counts of eosinophils and NLR in peripheral blood in prediction of in-hospital death are presented in Fig. 1. The AUROC for eosinophils was 0.88 (95% CI 0.76–0.99) and for NLR was 0.96 (95% CI 0.93–0.99).

We also analyzed AUROC for the components of NLR, however its values were lower—0.92 for lymphocyte count, identifying $1 \times 10^3/\mu\text{l}$ as a cut-off value and 0.77 for neutrophil count with cut off value of $7.3 \times 10^3/\mu\text{l}$.

Eosinopenia. In ROC analysis, an absolute number of eosinophils $\leq 0.04 \times 10^3/\mu\text{l}$ was identified as an optimal cut-off point for eosinopenia. Using this cut-off value, eosinopenia was diagnosed in 99 (36%) patients. All five patients who died presented on hospital admission with eosinopenia (*P* = 0.006 for the Fisher's Exact Test). This value was characterized by sensitivity of 100% (95% CI 35.9–100%), however specificity of 65.2% (95% CI 59.2–70.9%). The diagnostic accuracy was 0.66 (95% CI 0.6–0.71).

Comparison of study parameters upon stratification by presence of EP is presented in Table 2.

We found a significant negative correlation between eosinophil count and length of hospitalization ($\tau = -0.11$; *P* = 0.008), CRP ($\tau = -0.11$; *P* = 0.02); WBC ($\tau = -0.15$; *P* = 0.0004) and BE ($\tau = -0.1$; *P* = 0.03).

NLR. Elevated NLR was identified when the value was ≥ 13.2 . Such values were observed among 25 (9.09%) patients, including all five patients who died during hospitalization. This cut-off point was characterized by sensitivity of 100% (95% CI 35.9–100%) and specificity estimated for 92.6% (95% CI 88.8–95.4%). The diagnostic accuracy was 0.93 (95% CI 0.9–0.96). Comparison of study parameters upon stratification by NLR value is presented in Table 3.

Kendall's test did not detect significant correlation between SaO_2 and NLR ($\tau = -0.06$; *P* = 0.13).

In Table 2 we can see that patients who presented with eosinopenia had significantly higher NLR values: 8.44 (4.37–12.72) vs 2.26 (0.14–3.69); *P* < 0.0001. We observed parallel situation when we analyzed results from Table 3 – patients with high NLR values had significantly lower eosinophil count: $0.01 (0-0.02) \times 10^3/\mu\text{l}$ vs $0.1 (0.03-0.21) \times 10^3/\mu\text{l}$; *P* < 0.0001.

Parameter	Total n = 275
Age (years), mean (SD)	69.42 (9.59)
Male, n (%)	152 (55.27)
Active smoker, n (%)	102 (37.09)
Character of ECOPD	
Infectious, n (%)	146 (53.09)
CRP (mg/l), median (IQR)	10.88 (3.4–25.26)
WBC ($\times 10^3/\mu\text{l}$), median (IQR)	9.49 (7.77–12.24)
Eosinophils ($\times 10^3/\mu\text{l}$), median (IQR)	0.09 (0.02–0.19)
Lymphocytes ($\times 10^3/\mu\text{l}$), median (IQR)	1.6 (1–2.15)
Neutrophils ($\times 10^3/\mu\text{l}$), median (IQR)	6.7 (4.85–9.9)
NLR, median (IQR)	3.19 (1.48–6.96)
Hemoglobin (g/dl), median (IQR)	14 (12.8–15.15)
pH, median (IQR)	7.42 (7.39–7.45)
pCO ₂ (mmHg), median (IQR)	38.7 (35.25–45.28)
pO ₂ (mmHg), median (IQR)	63 (55.4–72)
SaO ₂ (%), median (IQR)	92.4 (89–94.88)
HCO ₃ ⁻ (mEq/l), median (IQR)	25.3 (23.45–28.1)
BE (mEq/l), median (IQR)	1 (–0.63 to 2.93)
Comorbidities	
Ischemic heart disease, n (%)	86 (31.27)
AF ^a , n (%)	31 (11.27)
Arrhythmia other than AF ^b , n (%)	8 (2.91)
Congestive heart failure, n (%)	116 (42.18)
Arterial hypertension, n (%)	179 (65.09)
Type 2 diabetes, n (%)	66 (24)
Others, n (%)	152 (55.27)

Table 1. Baseline study data. *AF* atrial fibrillation, *CRP* c-reactive protein, *IQR* interquartile range, *WBC* white blood count. ^aFirst detected in the admission, paroxysmal, persistent or permanent. ^bIn the admission or in the past medical history.

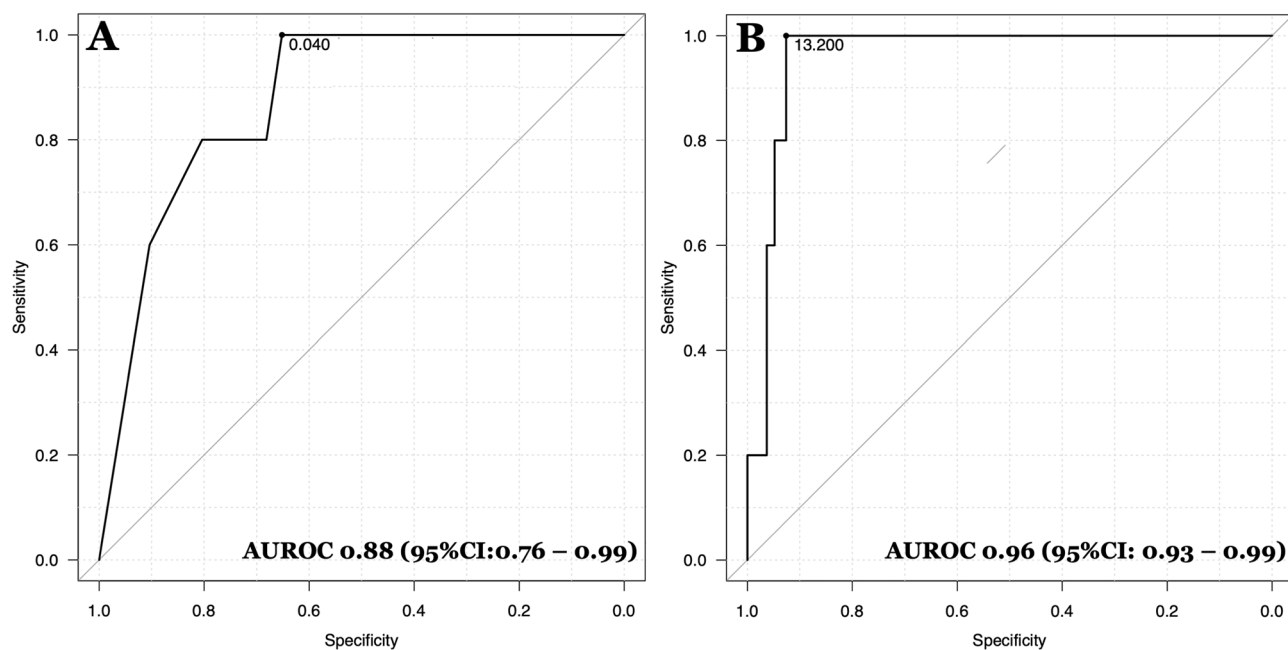


Figure 1. The ROC curve for absolute eosinophil count (A) and NLR (B) as predictors of in-hospital mortality. Cut-off points were marked with specificity and sensitivity values in brackets.

Parameter	Eosinopenia n = 99 (36%)	Normal eosinophil count n = 176 (64%)	P-value
Age (years), mean (SD)	70.24 (9.48)	68.96 (9.64)	0.29
Male, n (%)	51 (51.51)	101 (57.39)	0.42
Active smoker, n (%)	58 (58.59)	44 (25)	<0.0001
Length of hospitalization ^c (days), median (IQR)	8 (6–12.75)	7 (5–10)	0.004
Character of ECOPD			
Infectious, n (%)	60 (60.61)	86 (48.86)	0.06
CRP (mg/l), median (IQR)	15.53 (6.79–30.29)	10.14 (2.79–23.41)	0.07
WBC ($\times 10^3/\mu\text{l}$), median (IQR)	11.4 (8.17–13.2)	8.9 (7.67–11.57)	0.0007
Lymphocytes ($\times 10^3/\mu\text{l}$), median (IQR)	1.1 (0.7–1.8)	1.8 (1.2–2.4)	<0.0001
Neutrophils ($\times 10^3/\mu\text{l}$), median (IQR)	9.2 (6.1–11.55)	6.1 (4.6–7.95)	<0.0001
NLR, median (IQR)	8.44 (4.37–12.72)	2.26 (0.14–3.69)	<0.0001
Hemoglobin (g/dl), median (IQR)	13.9 (12.6–15.15)	14.2 (13–15.13)	0.29
pH, median (IQR)	7.42 (7.39–7.46)	7.42 (7.39–7.45)	0.65
pCO ₂ (mmHg), median (IQR)	38.9 (35.2–47.65)	38.7 (35.3–44.5)	0.86
pO ₂ (mmHg), median (IQR)	63.9 (54.5–73.5)	63 (55.65–71)	0.96
SaO ₂ (%), median (IQR)	92.7 (88.05–95.3)	92.4 (89.65–94.8)	0.86
HCO ₃ ⁻ (mEq/l), median (IQR)	26 (23.93–28.68)	24.95 (23.2–27.98)	0.1
BE (mEq/l), median (IQR)	1.5 (-0.08 to 3.73)	0.75 (-0.7 to 2.48)	0.04
Comorbidities			
Ischemic heart disease, n (%)	37 (37.37)	49 (27.84)	0.1
AF ^a , n (%)	20 (20.2)	11 (6.25)	0.0004
Arrhythmia other than AF ^b , n (%)	2 (2.02)	6 (3.41)	0.72
Congestive heart failure, n (%)	45 (45.45)	71 (40.34)	0.41
Arterial hypertension, n (%)	57 (57.58)	122 (69.32)	0.05
Type 2 diabetes, n (%)	26 (26.26)	40 (22.73)	0.51
Others, n (%)	83 (83.84)	69 (39.2)	0.0003

Table 2. Study parameters upon stratification by eosinophil count. *AF* atrial fibrillation, *CRP* c-reactive protein, *IQR* interquartile range, *WBC* white blood count. ^aFirst detected in the admission, paroxysmal, persistent or permanent. ^bIn the admission or in the past medical history. ^cAmong survivors.

Kendall's rank correlation tau revealed significant correlation between eosinophil and lymphocyte count: $\tau = 0.28$ ($P < 0.0001$) and negative correlation with NLR: $\tau = -0.38$ ($P < 0.0001$).

Coexistence of eosinopenia with lymphocytopenia. The coexistence of eosinopenia with lymphocytopenia (defined as lymphocyte count $\leq 1 \times 10^3/\mu\text{l}$) was observed in 47 (17.09%) patients and only such pattern was associated with in-hospital deaths in the analyzed group. There were no deaths in patients who presented with eosinopenia without lymphocytopenia ($P = 0.0001$ for the Fisher's Exact Test). The coexistence of eosinopenia with lymphocytopenia was characterized by higher specificity than previously analyzed eosinopenia [84.4% (95% CI 79.6–88.6%) vs 65.2% (95% CI 59.2–70.9%) and the same sensitivity of 100% (95% CI 35.9–100%)]. The diagnostic accuracy was 0.84 (95% CI 0.8–0.89).

Taking into account above findings, we reanalyzed the eosinopenia group in the context of previously analyzed study parameters. Results of additional analyses are presented in Table 4, which illustrates study parameters upon stratification by coexistence of eosinopenia and lymphocytopenia. Kruskal Wallis rank sum test, after pairwise comparisons, showed that only coexistence of eosinopenia with lymphocytopenia was associated with longer hospitalization ($P = 0.006$ vs normal eosinophils and lymphocytes; $P = 0.02$ for eosinopenia alone, and $P = 0.00004$ vs lymphocytopenia alone). Length of hospitalization in isolated eosinopenia or lymphocytopenia subgroups was not significantly different than in patients with normal counts of these cells ($P = 1.0$ and $P = 0.12$ respectively)—Fig. 2.

When we first assess the presence of coexistence of eosinopenia and lymphocytopenia, and then apply NLR, we can achieve the best sensitivity and specificity compared to the situation when these parameters are used individually: 100% (95% CI 35.9–100) and 93.7% (95% CI 90.1–96.3), respectively. The diagnostic accuracy for such approach was 0.94 (95% CI 0.90–0.96).

Also, among survivors, occurrence of such pattern was associated with significantly longer hospital stay: 11 (7–14) vs 7 (5–10) days, $P = 0.01$.

Discussion

Our aim was to assess the role of eosinopenia, lymphocytopenia and NLR as markers of in-hospital mortality and length of hospitalization (LoH) among patients with AECOPD requiring hospitalization.

Parameter	NLR \geq 13.2 n = 25 (9.09%)	NLR < 13.2 n = 250 (90.91%)	P-value
Age (years), mean (SD)	70.24 (10.83)	69.34 (9.47)	0.66
Male, n (%)	14 (56)	138 (55.2)	0.94
Active smoker, n (%)	13 (52)	89 (35.6)	0.11
Length of hospitalization ^c (days), median (IQR)	10 (7–12.5)	7 (5–10)	0.05
Character of ECOPD			
Infectious, n (%)	17 (68)	129 (51.6)	0.12
CRP (mg/l), median (IQR)	17.34 (8.92–22.6)	10.73 (3.37–25.31)	0.5
WBC ($\times 10^3/\mu\text{l}$), median (IQR)	11.9 (10.86–16)	9.13 (7.68–12)	< 0.0001
Eosinophils ($\times 10^3/\mu\text{l}$), median (IQR)	0.01 (0–0.02)	0.1 (0.03–0.21)	< 0.0001
Hemoglobin (g/dl), median (IQR)	14.1 (13–14.9)	14 (12.8–15.18)	0.83
pH, median (IQR)	7.42 (7.37–7.43)	7.42 (7.39–7.45)	0.12
pCO ₂ (mmHg), median (IQR)	42.3 (35.8–52.4)	38.7 (35–44.5)	0.2
pO ₂ (mmHg), median (IQR)	59.3 (54–65.1)	63.3 (55.4–73.1)	0.08
SaO ₂ (%), median (IQR)	90 (86–92.7)	92.7 (89.4–95.1)	0.008
HCO ₃ ⁻ (mEq/l), median (IQR)	26.75 (24.03–29.55)	25.2 (23.35–28.1)	0.31
BE (mEq/l), median (IQR)	1.6 (-1.18 to 4.5)	1 (-0.6 to 2.9)	0.78
Comorbidities			
Ischemic heart disease, n (%)	9 (36)	77 (30.8)	0.59
AF ^a , n (%)	6 (24)	25 (10)	0.05
Arrhythmia other than AF ^b , n (%)	0 (0)	8 (3.2)	1.0
Congestive heart failure, n (%)	14 (56)	102 (40.8)	0.14
Arterial hypertension, n (%)	13 (52)	166 (66.4)	0.15
Type 2 diabetes, n (%)	4 (16)	62 (24.8)	0.33
Others, n (%)	15 (60)	137 (54.8)	0.62

Table 3. Study parameters upon stratification by NLR value. *AF* atrial fibrillation, *CRP* c-reactive protein, *IQR* interquartile range, *WBC* white blood count. ^aFirst detected in the admission, paroxysmal, persistent or permanent. ^bIn the admission or in the past medical history. ^cAmong survivors.

Such approach seems to be justified by pathophysiological background. Namely, we would analyze eosinopenia in the context of considering AECOPD as a state of an acute stress. Undoubtedly, COPD is associated with both physical and emotional stress. Extremal, but pictorial evidence for possible levels of stress which can be released in this condition may be presented in context of relatively frequent association between COPD and Tako-Tsubo cardiomyopathy, which is linked with a rapid elevation of circulating catecholamine, triggered by emotional and/or physical stress, as a key mechanism¹². Also, Zurfluh et al. reported a time-dependent effect with higher levels pointing towards higher mortality at short term associated with activation of stress hormones (particularly cortisol and cortisone)¹³.

We should also remember that acute stress, and associated role of adrenal glucocorticoids in this process, is not the only cause of eosinopenia in peripheral blood. There are other contributing mechanisms including migration of eosinophils to the site of inflammation, rapid peripheral sequestration, suppression of egress of mature eosinophils from the bone marrow, and suppression of eosinophils production^{14,15}.

Eosinopenia in AECOPD was concluded to have some clinical implications in already published studies. Namely, Holland et al.⁶ reported that patients with eosinopenia had a longer hospital stay and were more likely to die in hospital. Additionally, Rahimi-Rad et al. found an association between eosinopenia and an unfavorable prognosis within 30-day after discharge⁷. Both Holland et al. and Rahimi-Rad et al. used cut-off value of 40 cells/mm³ to diagnose eosinopenia. In the past, we also reported that eosinopenia would be a prognostic factor of in-hospital mortality in ECOPD¹⁶, however, in the present study, we used the analysis which was not applied by above mentioned authors, therefore ROC curve assessment. From one hand, AUROC analysis may still justify such statement, but from the other hand, relatively low specificity 65.2% of this marker decreased our enthusiasm significantly.

Also, for further evaluation of the clinical implications of eosinopenia, discussion of its eventual consequences should be taken into account. First, we should start with the physiological role of these cells in the immune response. Among others, eosinophils promote humoral immunity by priming of B cells and have a role in the maintenance of type-2 immunity¹⁷, and regulation of T-helper-1 and Th2 immunity¹⁸. However, paradoxical to their physiological effects, evidence suggests that a reduction of eosinophils appears to have no negative effect on normal health. This statement is based not only on animal studies, but also on humans who received monoclonal antibodies reducing eosinophil count¹⁹. Therefore, even if we conclude that we indirectly assessed the exacerbation of stress or other contributing mechanisms, the occurrence of eosinopenia itself probably has no further significant clinical importance.

Parameter	Normal eosinophil and lymphocyte count n = 90 (32.73%)	Lymphocytopenia without eosinopenia n = 86 (31.27%)	Eosinopenia without lymphocytopenia n = 52 (18.91%)	Eosinopenia with lymphocytopenia n = 47 (17.09%)	P-value
Age (years), mean (SD)	68.92 (10.11)	69 (9.19)	69.33 (8.85)	71.26 (10.12)	0.55
Male, n (%)	40 (44.44)	61 (70.93)	26 (50)	25 (53.19)	0.004
Active smoker, n (%)	25 (27.78)	19 (22.09)	34 (65.38)	24 (51.06)	<0.0001
Length of hospitalization ^c (days), median (IQR)	8 (6–11)	6.5 (5–9)	7 (6–10)	10.5 (7–14)	<0.0001
Character of EOCOPD					
Infective, n (%)	39 (43.33)	47 (54.65)	29 (55.77)	31 (65.96)	0.08
CRP (mg/l), median (IQR)	9.62 (2.78–19.74)	10.71 (3.29–29.96)	18.62 (7.57–29.3)	13.52 (6.61–29)	0.22
WBC ($\times 10^3/\mu\text{l}$), median (IQR)	9.72 (8.29–11.97)	8.34 (6.9–9.83)	11.6 (8.58–13.2)	10.86 (8.04–13.54)	<0.0001
Eosinophils ($\times 10^3/\mu\text{l}$), median (IQR)	0.18 (0.1–0.28)	0.14 (0.08–0.27)	0.02 (0.01–0.03)	0.01 (0–0.02)	<0.0001
Lymphocytes ($\times 10^3/\mu\text{l}$), median (IQR)	2.4 (2–2.8)	1.2 (0.9–1.5)	1.75 (1.4–2.23)	0.7 (0.6–0.9)	<0.0001
Neutrophils ($\times 10^3/\mu\text{l}$), median (IQR)	6 (4.7–8.18)	6.2 (4.5–7.5)	8.05 (6.65–11.4)	10 (6.95–12.45)	<0.0001
NLR, median (IQR)	2.15 (1.03–3.09)	2.62 (0.09–5.68)	4.65 (2.34–7.09)	12.78 (9.88–18.04)	<0.0001
Hemoglobin (g/dl), median (IQR)	14.5 (13.3–15.2)	13.7 (12.83–15)	13.85 (12.5–15.13)	14 (12.6–15.25)	0.33
pH, median (IQR)	7.43 (7.39–7.46)	7.42 (7.39–7.44)	7.44 (7.4–7.46)	7.42 (7.37–7.43)	0.06
pCO ₂ (mmHg), median (IQR)	37.9 (35–42.6)	39.75 (36.53–46.3)	37.8 (34.55–41.13)	41.3 (35.75–55.35)	0.06
pO ₂ (mmHg), median (IQR)	64.9 (57–75.2)	61 (55.1–68.13)	64 (56.43–74.63)	60.2 (52.45–72.5)	0.33
SaO ₂ (%), median (IQR)	92.9 (90.1–95.3)	91.8 (88.73–93.8)	93.85 (90.3–95.85)	91.6 (86.6–94.8)	0.07
HCO ₃ ⁻ (mEq/l), median (IQR)	24.8 (22.9–27.18)	25.1 (23.2–28.1)	25.3 (23.6–27.7)	26.9 (24.3–29.8)	0.08
BE (mEq/l), median (IQR)	0.75 (–0.78 to 2.2)	0.75 (–0.7 to 2.83)	1.2 (–0.1 to 3)	1.7 (0–4.3)	0.18
Comorbidities, n (%):					
Ischemic heart disease	23 (25.56)	26 (30.23)	19 (36.54)	18 (38.3)	0.37
AF ^a	4 (4.44)	7 (8.14)	12 (23.08)	8 (17.02)	0.003
Arrhythmia other than AF ^b	3 (3.33)	3 (3.49)	1 (1.92)	1 (2.13)	1.0
Congestive heart failure	38 (42.22)	33 (38.37)	19 (36.54)	26 (55.32)	0.22
Arterial hypertension	62 (68.89)	60 (69.77)	34 (65.38)	23 (48.94)	0.08
Type 2 diabetes	22 (24.44)	18 (20.93)	20 (38.46)	6 (12.77)	0.02
Others	36 (40)	47 (54.65)	42 (80.77)	27 (57.45)	<0.001

Table 4. Study parameters upon stratification by coexistence of eosinopenia and lymphocytopenia. *AF* atrial fibrillation, *CRP* c-reactive protein, *IQR* interquartile range, *WBC* white blood count. ^aFirst detected in the admission, paroxysmal, persistent or permanent. ^bIn the admission or in the past medical history. ^cAmong survivors.

Taking into account all above mentioned, and the fact that glucocorticoids can reduce eosinophils, raise neutrophils and reduce lymphocytes^{20,21} we decided to evaluate eosinopenia in a broader context, by adding the assessment of lymphocytopenia and NLR into analysis. The relation between NLR and in-hospital mortality also was previously reported in patients with AECOPD²². Yao et al. presented ROC curve analysis for using NLR to predict in-hospital mortality—AUROC was 0.803, and an optimal cut-off value was 6.24. Authors reported the sensitivity of 81.08%, and specificity of 69.17%. We observed both better sensitivity and specificity for NLR, however we obtained higher cut-off value in ROC curve analysis. In our study, NLR presented much better accuracy parameters and higher value of AUROC than obtained for eosinopenia. Moreover, we have observed that these parameters are not fully independent and that there is a significant relationship between the values of eosinophil count and NLR. That is why we started to analyze these associations in the context of individual components of NLR, observing intriguing interplay between eosinophils and lymphocytes. Interestingly, patients who presented with the coexistence of eosinopenia and lymphocytopenia had the lowest counts of both eosinophils and lymphocytes. This is justified because these patients probably had the highest level of stress, which can suppress both types of cells. However, this relationship itself seems to also have interesting clinical implications.

In turn, eosinophils and neutrophils are granulocytes that originate from the same myeloblast progenitor in the bone marrow, and upon differentiating, each of these granulocytes leaves the marrow and migrates to the inflamed tissues to enact their effector functions. From the other hand, lymphocytes, which originate from the different progenitor, secrete cytokines that coordinate eosinophil and neutrophil responses. From one hand, type 2 CD4⁺ T cells and type 2 innate lymphoid cells produce IL-5 that prompts eosinophil production^{23,24}, while from the other hand, Th17 cells, CD8⁺ T cells, $\gamma\delta$ T cells, and type 3 innate lymphoid cells secrete IL-17A, leading to neutrophil maturation^{25,26}. Infection by different types of pathogens may lead to different effects—either eosinophil, or neutrophil accumulation²⁷. Wiesner et al. reported that Rag2/IL-2R $\gamma^{-/-}$ mice that lack lymphocytes, and observed that without lymphocytes, infected mice had significantly impaired eosinophilia compared with similarly infected wild-type mice, yet neutrophil accumulation remained unimpaired. Authors also concluded

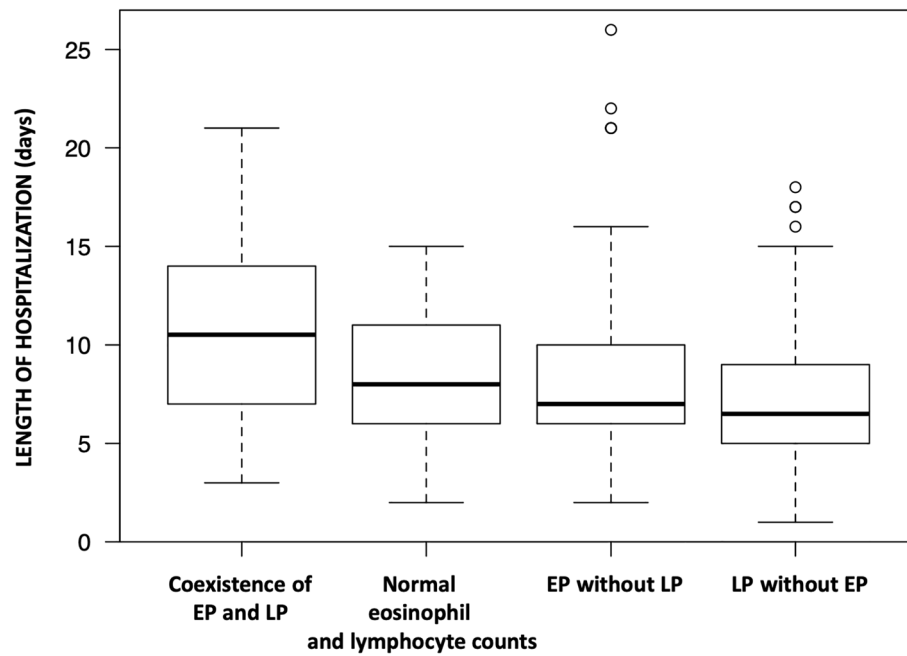


Figure 2. Boxplot for the comparison of length of hospitalization upon stratification by the presence of eosinopenia and lymphocytopenia coexistence. *EP* eosinopenia, *LP* lymphocytopenia.

that singular elimination of Th cells (while leaving all other lymphocyte subsets intact, including ILCs and NK cells) may result in a loss of eosinophils that can be replaced by IL-17A-dependent neutrophilia²⁷.

Focusing on COPD population, Freeman et al. observed decreased concentration of CD4⁺ and CD8⁺ T cells in peripheral blood during AECOPD. From the other hand, Ross et al. observed that exacerbated airway neutrophilia in cigarette smoke-exposed mice infected with nontypeable *Haemophilus influenzae*, large subgroup of bacterial AECOPD, was associated with an induction of IL-17A²⁸. There is also evidence for a decrease in both total CD4⁺ and CD8⁺ T cell counts after infusion of cortisol. The effect started 90 min after infusion²⁹.

How would these issues contribute to an increase in mortality? Namely, CD4⁺ T cells are major players involved in responses to infectious diseases, enabling B cells to differentiate into plasma cells, helping CD8⁺ T cells develop into cytotoxic cells, as well as are required for long-term CD8 memory generation. CD4⁺ T cells also mediate activation of macrophages, playing a critical role in the viral and bacterial control^{30–35}. Therefore, decrease in this population of cells would result in catastrophic effects for homeostasis of the immune response.

The above-mentioned evidence may partially and indirectly justify our observations and allow to draw a hypothetical pathway that lymphocytopenia would be a primary pathology, which leads to eosinopenia and neutrophilia. Playing together with stress response, which potentially aggravates such pattern, these factors may contribute to a significantly increased risk of in-hospital death. However, for confirmation of these hypotheses, there is a need for a prospective study which directly will analyze all above mentioned relationships: stress response, lymphocytes subpopulations assessment, eosinophil and neutrophil counts with broad profile of cytokines, including IL-17A.

There are some major limitations of our study, which need to be considered. As a retrospective study, it is burdened by all limitations associated with this type of data collection, including an absence of data regarding potential confounding factors. As the most important we indicate above mentioned lack of measurement of stress hormones and inflammatory cytokines. It should be also worth to diagnose pathogens responsible for infectious AECOPDs. Taking into account the above-mentioned issues, the results of our study should be considered as hypothesis generating and should be furtherly confirmed in a sufficiently powered prospective analysis, designed taking into account above mentioned limitations.

Conclusions

All analyzed parameters in our study have certain value as prognostic factors of in-hospital mortality in AECOPD. However, the best profile of sensitivity and specificity can be obtained by combined analysis of coexistence of eosinopenia and lymphocytopenia with elevated NLR. Occurrence of such pattern is also associated with significantly longer time of hospitalization among survivors.

Received: 22 June 2020; Accepted: 16 February 2021

Published online: 26 February 2021

References

1. Wedzicha, J. A. & Seemungal, T. A. R. COPD exacerbations: Defining their cause and prevention. *Lancet Lond. Engl.* **370**, 786–796 (2007).
2. Seemungal, T. A. *et al.* Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *Am. J. Respir. Crit. Care Med.* **157**, 1418–1422 (1998).
3. Global Initiative for Chronic Obstructive Pulmonary Disease. Global Strategy for the diagnosis, management, and prevention of Chronic Obstructive Pulmonary Disease. https://goldcopd.org/wp-content/uploads/2019/12/GOLD-2020-FINAL-ver1.2-03Dec19_WMV.pdf. Accessed 31 October 2020.
4. Lima, F. V., Yen, T. Y. M. & Patel, J. K. Trends in in-hospital outcomes among adults hospitalized with exacerbation of chronic obstructive pulmonary disease. *COPD* **12**, 636–642 (2015).
5. Karakostas, S., Kalemaki, D., Tzagkarakis, E. & Lydakis, C. Pitfalls in studies of eosinopenia and neutrophil-to-lymphocyte count ratio. *Infect. Dis. Lond. Engl.* **50**, 163–174 (2018).
6. Holland, M., Alkhalil, M., Chandromouli, S., Janjua, A. & Babores, M. Eosinopenia as a marker of mortality and length of stay in patients admitted with exacerbations of chronic obstructive pulmonary disease. *Respirol. Carlton Vic.* **15**, 165–167 (2010).
7. Rahimi-Rad, M. H., Asgari, B., Hosseinzadeh, N. & Eishi, A. Eosinopenia as a marker of outcome in acute exacerbations of chronic obstructive pulmonary disease. *Maedica* **10**, 10–13 (2015).
8. Bolayir, A. *et al.* The effect of eosinopenia on mortality in patients with intracerebral hemorrhage. *J. Stroke Cerebrovasc. Dis. Off. J. Natl. Stroke Assoc.* **26**, 2248–2255 (2017).
9. Korkmaz, Ö. *et al.* Is preoperative eosinopenia an independent predictor of early mortality for coronary artery bypass surgery?. *Heart Surg. Forum* **19**, E088–093 (2016).
10. Yip, B. & Ho, K. M. Eosinopenia as a predictor of unexpected re-admission and mortality after intensive care unit discharge. *Anaesth. Intensive Care* **41**, 231–241 (2013).
11. R Core Team. (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.
12. Manfredini, R. *et al.* Heart and lung, a dangerous liaison-Tako-tsubo cardiomyopathy and respiratory diseases: A systematic review. *World J. Cardiol.* **6**, 338–344 (2014).
13. Zurfluh, S. *et al.* Association of adrenal hormone metabolites and mortality over a 6-year follow-up in COPD patients with acute exacerbation. *Clin. Chem. Lab. Med.* **56**, 669–680 (2018).
14. Abidi, K. *et al.* Eosinopenia is a reliable marker of sepsis on admission to medical intensive care units. *Crit. Care Lond. Engl.* **12**, R59 (2008).
15. Abidi, K. *et al.* Eosinopenia, an early marker of increased mortality in critically ill medical patients. *Intensive Care Med.* **37**, 1136–1142 (2011).
16. Bialas, A. *et al.* Eosinopenia as a prognostic factor in patients with acute exacerbation of chronic obstructive pulmonary disease. *Eur. Respir. J.* **50**, PA2110 (2017).
17. Spencer, L. A. & Weller, P. F. Eosinophils and Th2 immunity: Contemporary insights. *Immunol. Cell Biol.* **88**, 250–256 (2010).
18. Stehle, C., Saikali, P. & Romagnani, C. Putting the brakes on ILC2 cells. *Nat. Immunol.* **17**, 43–44 (2016).
19. Gleich, G. J., Klion, A. D., Lee, J. J. & Weller, P. F. The consequences of not having eosinophils. *Allergy* **68**, 829–835 (2013).
20. Shoenfeld, Y., Gurewich, Y., Gallant, L. A. & Pinkhas, J. Prednisone-induced leukocytosis. Influence of dosage, method and duration of administration on the degree of leukocytosis. *Am. J. Med.* **71**, 773–778 (1981).
21. Altman, L. C., Hill, J. S., Hairfield, W. M. & Mullarkey, M. F. Effects of corticosteroids on eosinophil chemotaxis and adherence. *J. Clin. Invest.* **67**, 28–36 (1981).
22. Yao, C., Liu, X. & Tang, Z. Prognostic role of neutrophil-lymphocyte ratio and platelet-lymphocyte ratio for hospital mortality in patients with AECOPD. *Int. J. Chron. Obstruct. Pulmon. Dis.* **12**, 2285–2290 (2017).
23. Licona-Limón, P., Kim, L. K., Palm, N. W. & Flavell, R. A. TH2, allergy and group 2 innate lymphoid cells. *Nat. Immunol.* **14**, 536–542 (2013).
24. Nussbaum, J. C. *et al.* Type 2 innate lymphoid cells control eosinophil homeostasis. *Nature* **502**, 245–248 (2013).
25. Eyerich, S., Eyerich, K., Cavani, A. & Schmidt-Weber, C. IL-17 and IL-22: Siblings, not twins. *Trends Immunol.* **31**, 354–361 (2010).
26. Zheng, Y. *et al.* Interleukin-22, a T(H)17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis. *Nature* **445**, 648–651 (2007).
27. Wiesner, D. L., Smith, K. D., Kashem, S. W., Bohjanen, P. R. & Nielsen, K. Different lymphocyte populations direct dichotomous eosinophil or neutrophil responses to pulmonary cryptococcus infection. *J. Immunol. Baltim. Md* **1950**(198), 1627–1637 (2017).
28. Roos, A. B. *et al.* IL-17A and the promotion of neutrophilia in acute exacerbation of chronic obstructive pulmonary disease. *Am. J. Respir. Crit. Care Med.* **192**, 428–437 (2015).
29. Dimitrov, S. *et al.* Cortisol and epinephrine control opposing circadian rhythms in T cell subsets. *Blood* **113**, 5134–5143 (2009).
30. North, R. J. & Jung, Y.-J. Immunity to tuberculosis. *Annu. Rev. Immunol.* **22**, 599–623 (2004).
31. Román, E. *et al.* CD4 effector T cell subsets in the response to influenza: Heterogeneity, migration, and function. *J. Exp. Med.* **196**, 957–968 (2002).
32. Brown, D. M., Román, E. & Swain, S. L. CD4 T cell responses to influenza infection. *Semin. Immunol.* **16**, 171–177 (2004).
33. Sun, J. C., Williams, M. A. & Bevan, M. J. CD4+ T cells are required for the maintenance, not programming, of memory CD8+ T cells after acute infection. *Nat. Immunol.* **5**, 927–933 (2004).
34. Shedlock, D. J. & Shen, H. Requirement for CD4 T cell help in generating functional CD8 T cell memory. *Science* **300**, 337–339 (2003).
35. Schoenberger, S. P., Toes, R. E., van der Voort, E. I., Offringa, R. & Melief, C. J. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. *Nature* **393**, 480–483 (1998).

Author contributions

A.J.B. conceived the idea of the study. T.K. and A.J.B. contributed to the design of the research. All authors were involved in data collection, however with a predominance of T.K.. All authors were involved in data analysis. A.J.B. and T.K. performed statistical analysis. All authors were involved in analysis of the results. All authors edited and approved the final version of the manuscript.

Funding

The costs of this study were defrayed from regular finances of the Department of Pneumology and Allergy of Medical University of Lodz, Poland (503/1-151-03).

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to A.J.B.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021