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Ferric Heme Superoxide Reductive Transformations to Ferric Heme (Hydro)Peroxide Species: Spectroscopic Characterization and Thermodynamic Implications for H-atom Transfer (HAT)

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Abstract

A new end-on low-spin ferric heme peroxide, [(P^{Im})Fe^{III}—(O₂²⁻)]⁻ (**P^{Im}-P**), and subsequently formed hydroperoxide species, [(P^{Im})Fe^{III}—(OOH)] (**P^{Im}-HP**) are generated utilizing the iron-porphyrinate P^{Im} with its tethered axial base imidazolyl group. Measured thermodynamic parameters, the ferric heme superoxide [(P^{Im})Fe^{III}—(O₂^{•-})] (**P^{Im}-S**) reduction potential (E°) and the **P^{Im}-HP** p*K*_a value, lead to the finding of the OO—H bond dissociation free energy (BDFE) of **P^{Im}-HP** as 69.5 kcal/mol, using a thermodynamic square scheme and Bordwell relationship. The results are validated by the observed oxidizing ability of **P^{Im}-S** via hydrogen atom transfer (HAT) compared to that of the F₈ superoxide complex, [(F₈)Fe^{III}—(O₂^{•-})] (**S**) (F₈ = tetrakis(2,6-difluorophenyl)porphyrinate, without an internally appended axial base imidazolyl), as determined from reactivity comparison of superoxide complexes **P^{Im}-S** and **S** with the hydroxylamine (O—H) substrates TEMPO-H and ABNO-H.

Thermodynamic comparisons for O₂-derived iron-porphyrinate interrelated ferric heme superoxide, peroxide and hydroperoxide analogs in the presence and absence of an appended imidazolyl axial base are presented.

Graphical Abstract

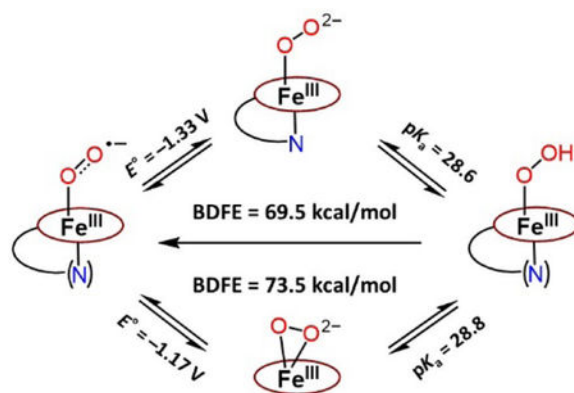
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Conflict of interest

The authors declare no conflict of interest.

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Keywords

end-on peroxide; bond dissociation free energy; BDFE; hydrogen atom transfer; square scheme; thermodynamic parameters

Introduction

Heme-containing enzymes show diverse biological functions such as substrate monooxygenation, oxygen reduction in the respiratory chain, nitric oxide synthesis, oxygen storage and delivery, electron transfer, and H_2O_2 activation and dismutation.^[1] Heme enzyme active sites contain a proximal ligand; cytochrome P450s (CYP450s) as well as NO synthase and chloroperoxidase utilize a cysteinyl residue, whereas other peroxidases include a histidyl proximal ligand and catalases possess a tyrosinate ligand. The identity and nature of the axial ligand contributes to or defines spectroscopic properties, coordination, iron reduction potentials and thus the reactivity. These proximal ligand donors contribute the so-called “push effect”, due to their electron donation.^[1] In particular, the “push effect” of the anionic thiolate ligand in CYP450s leads to a much more negative $\text{Fe}^{\text{III/II}}$ reduction potential^[2] and helps effect the heterolytic O—O bond cleavage of a hydroperoxide species, forming Compound I (Cmpd I, $(\text{P}^+)\text{Fe}^{\text{IV}}=\text{O}$) as the reactive species which hydroxylates substrates.^[1]

CYP450 monooxygenases transfer one atom of O_2 to a substrate, giving hydroxylated, epoxidized, or sulfoxidized products.^[1] The reaction cycle involves formation of heme Fe^{III} —superoxide (the initial $\text{Fe}^{\text{II}}/\text{O}_2$ adduct), with subsequent transformation to Fe^{III} —peroxide and Fe^{III} —hydroperoxide intermediates prior to O—O cleavage to give Cmpd I.^[1,4]

For these three initial Fe^{III} CYP450 intermediates, radiolytic cryoreduction/spectroscopic studies and theoretical calculations lead to the conclusion that all have an end-on binding geometry, with Fe^{III} -coordination only to the O_2 -derived proximal oxygen atom.^[5] Synthetic bioinorganic groups have been interested in the characterization of model compounds for the superoxide, peroxide, or hydroperoxide intermediates, as relevant to O_2 or H_2O_2 activating heme enzymes. There are many known ferric heme superoxide^[6] and hydroperoxide synthetic compounds.^[3,6g,6i,6j,7] Valentine and co-workers^[8] originally synthesized and

characterized a series of ferric heme peroxides possessing a side-on bound O_2^{2-} ligand (η^2 , with both peroxide O-atoms equivalently bound to Fe^{III}). To date, there has been only one report of an end-on (η^1) bound low-spin peroxide synthetic complex, $[(P)Fe^{III}-(\eta^1:O_2^{2-})]^-$ (P possesses an internal imidazolyl axial ligand), that by Naruta and co-workers.^[6g] Recently, we^[3,6d] reported on an end-on ferric heme superoxide, a corresponding hydroperoxide species, and the related ferric heme peroxide, where the O_2 -derived O_2^{2-} ligand is side-on bound (Figure 1, upper right); the chemistry was carried out in tetrahydrofuran (THF), which serves as an axial base ligand. In that work,^[3] thermodynamic parameters, the $[(F_8)Fe^{III}-(O_2^{\bullet-})]$ (*S*) reduction potential and $[(F_8)Fe^{III}-(OOH)]$ (*HP*) pK_a , led to the determination of the ferric heme hydroperoxide BDFE to be 73.5 kcal/mol.

Herein, the newly generated end-on low-spin peroxide, $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im}-P**), and its subsequent conversion to hydroperoxide species, $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im}-HP**) are described. The **P^{Im}** porphyrinate possesses an appended axial base strongly donating imidazolyl group (Figure 1), in partial analogy to the strongly donating cysteinate base in CYP450s; Fe^{III} is bound to only one oxygen atom of the peroxide moiety. Also, a reduction potential for the $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im}-S**)/ $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im}-P**) couple and a pK_a for the $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im}-HP**) acid-base pair are established here. These thermodynamic parameters and use of the Bordwell relationship square scheme (Figure 1) allow the determination of OO—H bond dissociation free energy (BDFE) for **P^{Im}-HP**. We compare and contrast our thermodynamic findings and HAT reactivity with substrates to those obtained with a previously studied system employing the F_8 porphyrinate (Figure 1).

Results and Discussion

The ferric heme superoxide $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im}-S**) generated by bubbling O_2 to a solution of $[(P^{Im})Fe^{II}]$ (Figure 1), was previously characterized (ν_{O-O} , 1180 ($^{18}O_2$, -56) cm^{-1} ; ν_{Fe-O} , 575 ($^{18}O_2$, -23) cm^{-1}).^[6h,9] Addition of cobaltocene ($CoCp_2$) to **P^{Im}-S** at -80 °C in THF results in the formation of the ferric heme peroxide complex $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im}-P**); UV-vis spectral shifts occurring are 423 to 424 nm (Soret) and 532 to 535, 567 nm (Figure 2A). These product electronic absorption spectra are very similar to those of Naruta's end-on low-spin ferric heme peroxide, as mentioned the first of its kind.^[6g] The UV-vis features of **P^{Im}-P** and Naruta's analog exhibit higher energy Soret λ_{max} values than the side-on bound ferric heme peroxides (also lacking a trans axial base ligand), as in $[(F_8)Fe^{III}-(\eta^2:O_2^{2-})]^-$ (Figure 1, upper right).^[3,6d,6f,8] The low-spin end-on formulation is confirmed by the EPR spectra which display signals at $g = 2.25$, 2.14, and 1.95 (Figure 2A), matching Naruta's analog,^[6g] as well as for those end-on ferric peroxide obtained by cryoreduction methods in heme enzymes.^[5] Such EPR signatures also are distinct from side-on heme peroxide intermediates which are intermediate spin species, $g = 4.2$ ($S = 3/2$).^[3,6d,6f,8]

Further, $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im}-P**) was characterized using rR spectroscopy. Two isotopic sensitive bands were observed, at 810 ($^{18}O_2 = -34$) cm^{-1} and 578 ($^{18}O_2 = -26$) cm^{-1} corresponding to the ν_{O-O} and ν_{Fe-O} stretches, respectively (Figure 2B). This latter stretching frequency value matches the expected value ($\nu_{Fe-O,calc} (^{16}O_2/^{18}O_2) = -26$ cm^{-1}) using the harmonic oscillator approximation. The magnitude of the isotope shift for ν_{O-O} is

smaller than the calculated value ($\nu_{\text{O-O,calc}}(^{16}\text{O}_2/^{18}\text{O}_2) = -46 \text{ cm}^{-1}$); however, the same trend is found for Naruta's complex $[(\text{P})\text{Fe}^{\text{III}}(\eta^1\text{:O}_2^{2-})]^-$ ($\nu_{\text{O-O}}, 808$ ($^{18}\text{O}_2, -37$) cm^{-1} ; $\nu_{\text{Fe-O}}, 585$ ($^{18}\text{O}_2, -25$) cm^{-1}).^[6g] It is notable that $\nu_{\text{Fe-O}}$ in $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\eta^1\text{:O}_2^{2-})]^-$ (**P^{Im}-P**) is much larger than that found for the side-on peroxide species which has no internally appended axial base imidazolyl, $[(\text{F}_8)\text{Fe}^{\text{III}}(\eta^2\text{:O}_2^{2-})]^-$ (Figure 1, upper right; $\nu_{\text{O-O}}, 806$ ($^{18}\text{O}_2, -46$) cm^{-1} ; $\nu_{\text{Fe-O}}, 466$ ($^{18}\text{O}_2, -19$) cm^{-1}).^[3] The increased Fe—O stretching frequency is likely a result of both the monodentate coordination and the σ -symmetry O_2 donation into the fully unoccupied low-spin Fe d_{z^2} orbital which is stronger than the π -symmetry donation into the half-occupied Fe d_{xz}/d_{yz} orbital of the side-on peroxide complex. These effects would strengthen the Fe—O bond of **P^{Im}-P**, and favor the end-on binding mode, giving a low-spin six-coordinate species $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\eta^1\text{:O}_2^{2-})]^-$ (**P^{Im}-P**).

When excess triflic acid (HOTf) is added to $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{2-})]^-$ (**P^{Im}-P**), Fe—O bond cleavage occurs, releasing H_2O_2 (89 % yield; horseradish peroxidase assay, see Figure S1), corroborating the complex's peroxide formulation. **P^{Im}-P** is singly protonated by adding 1 equiv of 2,6-lutidinium triflate $[(\text{LutH}^+)](\text{OTf})$ at -80 °C in THF, giving $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{OOH})]$ (**P^{Im}-HP**) with absorptions at 423 and 533 nm (Figure 3A). EPR spectra of **P^{Im}-HP** confirm an end-on low-spin ferric heme hydroperoxide structure ($g = 2.25, 2.14, \text{ and } 1.95$) as shown in Figure 3B. These values correspond closely to those already known for synthetically derived end-on low-spin hydroperoxide complexes^[3,6f,6g,6i,6j,7a,7b] as well as for ferric heme hydroperoxides generated within hemoglobin and myoglobin.^[10a,b] A high yield of H_2O_2 (90.1 %) is also obtained when **P^{Im}-HP** is acidified using HOTf (see Figure S1). Further, we performed rR spectroscopy; while only noisy spectra could be obtained (Figure S2), $\nu_{\text{O-O}}$ (808 ($^{18}\text{O}_2, -48$) cm^{-1}) and $\nu_{\text{Fe-O}}$ (579 ($^{18}\text{O}_2, -36$) cm^{-1}) resonances could be well discerned. These parameters, for **P^{Im}-HP**, are quite similar to those for the other synthetic or protein low-spin ferric heme hydroperoxide complexes (vide supra).^[3,6g,7c,10]

We find $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{\bullet-})]$ (**P^{Im}-S**) and $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{2-})]^-$ (**P^{Im}-P**) are interconvertible (Figure 1) using redox reagents. Addition of a ferrocenium reagent (Fc^+ ; $E_{1/2} = 0 \text{ V vs Fc}^{+/0}$) to a solution of **P^{Im}-P** leads to the reappearance of the optical bands (423 and 533 nm) associated with **P^{Im}-S** (Figure S3). This result is supported by the diamagnetic behaviour (i.e., EPR silent; 10 K, THF) of the resulting **P^{Im}-S** solution. The further addition of CoCp_2 leads to re-reduction giving the peroxide species **P^{Im}-P** (Figure S4).

Interestingly, titration of $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{\bullet-})]$ (**P^{Im}-S**) with aliquots of CoCp_2 ($E_{1/2} = -1.33 \text{ V vs Fc}^{+/0}$ in THF)^[11] at -80 °C in THF gives equilibrium mixtures; UV-vis monitoring (at either 423 or 567 nm) (Figure 4A, Table S1, Figure S5), allowed determination of K_{eq} for each aliquot added. From these results and utilizing the Nernst equation, the reduction potential for the couple, $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{\bullet-})]$ (**P^{Im}-S**)/ $[(\text{P}^{\text{Im}})\text{Fe}^{\text{III}}(\text{O}_2^{2-})]^-$ (**P^{Im}-P**) is calculated to be $-1.33 \text{ V} \pm 0.01 \text{ V vs Fc}^{+/0}$. This value is more negative by $\sim 160 \text{ mV}$ than that reported for $[(\text{F}_8)\text{Fe}^{\text{III}}(\text{O}_2^{\bullet-})]$ ($-1.17 \text{ V vs Fc}^{+/0}$) (Figure S6) which has no internally appended axial base in the porphyrinate complex.^[3] Consistent with this finding is the observation that in order to fully reduce $[(\text{F}_8)\text{Fe}^{\text{III}}(\text{O}_2^{\bullet-})]$ to its peroxide analogue, one equiv CoCp_2 was used, while multiple equiv of CoCp_2 are required to fully reduce the

presently described superoxide complex **P^{Im}-S**. These findings suggest a “push effect” due to the axial imidazolyl ligand which is covalently appended to the heme of P^{Im} iron-porphyrinate complexes.

Dey and co-workers^[2] reported the reduction potentials of iron-porphyrinate complexes (Fe^{III/II}) which have thiolate and imidazolyl linkers bound to the iron center of porphyrinate complexes. A thiolate ligated porphyrin complex shows 500 mV more negative Fe^{III/II} reduction potential compared to an imidazolyl ligated porphyrinate analogue; the thiolate is a much better donor than imidazolyl. Never-the-less, in our case the greater charge donation by an axial imidazolyl ligand in P^{Im} reduces the reduction potential of [(P^{Im})Fe^{III}—(O₂^{•-})] (**P^{Im}-S**) in comparison with that of [(F₈)Fe^{III}—(O₂^{•-})].

To complete the thermodynamic square scheme analysis (Figure 1),^[12] the p*K*_a value of ferric heme hydroperoxide [(P^{Im})Fe^{III}—(OOH)] (**P^{Im}-HP**) was evaluated at -80 °C in THF. Full deprotonation of **P^{Im}-HP** was accomplished by spectral titration using the derivatized phosphazene base EtP₂(dma) (see Table S2 for the chemical structure; conjugate base p*K*_a = 28.1; THF solvent, RT).^[13] The concentrations of each species (**P^{Im}-P**, **P^{Im}-HP**, EtP₂(dma), and EtP₂(dma)H⁺) were evaluated by examination of the observed absorbance at either 423 or 567 nm in the UV-vis spectra (Figure 4B, Table S2, Figure S7). Such titration experiments lead to the establishment of *K*_{eq} constants for the protonation of **P^{Im}-P** to give **P^{Im}-HP**, allowing the calculation of the p*K*_a of the ferric heme hydroperoxide to be 28.6 ± 0.5 (-80 °C, THF). The reversibility of acid-base reaction was further demonstrated by adding [(LutH⁺)(OTf)] to the solution generated in this titration, which gives back **P^{Im}-HP**, (Figure S8).

This allows the completion of the thermodynamic square scheme for electron and proton transfer reactions (Figure 1) using the presently determined thermodynamic parameters, *E*^o (-1.33 V vs. Fc⁺⁰, vide supra) and p*K*_a (28.6, vide supra), and determination of the BDFE of the OO—H in the ferric heme hydroperoxide complex [(P^{Im})Fe^{III}—(OOH)] (**P^{Im}-HP**). According to eq. 1 (Bordwell relationship, C_G stands for the H⁺/H[•] standard reduction potential in a particular solvent),^[12] where C_G = 61 kcal/mol in THF, the **P^{Im}-HP** BDFE is calculated to be 69.5 kcal/mol (eq. 2).

$$\text{BDFE} = 1.37(\text{p}K_a) + 23.06E^o + C_{G, \text{ solv}} \quad (1)$$

$$\text{BDFE}_{\text{OO-H}} = 1.37(28.6) + 23.06(-1.33) + 61 = \mathbf{69.5 \text{ kcal / mol}} \quad (2)$$

Our own previous report is the only other experimental evaluation of the thermodynamics and use of the square scheme as applied to ferric heme superoxide, peroxide (Figure 1, upper right) and hydroperoxide relatives, i.e., for the F₈ heme containing superoxide, (side-on) peroxide and hydroperoxide, [(F₈)Fe^{III}—(OOH)] BDFE_{OO-H} = 73.5 kcal/mol.^[3] Our OO—H BDFE value presented here with P^{Im} is shown to be ~ 4 kcal/mol less, and the difference resides primarily in the variation in the superoxide/peroxide redox couple (Figure 5).

The pK_a values for $[(F_8)Fe^{III}-(OOH)]$ vs $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im}-HP**) are essentially identical,^[14] but the E° reduction potential is measurably more negative for the (**P^{Im}-S**) / (**P^{Im}-P**) couple (Figure 5). This makes chemical sense as the P^{Im} system possesses the strong imidazolyl axial base ligand donor, that not present in the F₈ heme system. Also, the dissimilarity of BDFE values ($[(P^{Im})Fe^{III}-(OOH)]$ BDFE_{OO-H} = 69.5 kcal/mol and $[(F_8)Fe^{III}-(OOH)]$ BDFE_{OO-H} = 73.5 kcal/mol) is supported by their hydrogen atom transfer (HAT) reactivities toward H atom donors (vide infra). As we noted previously^[3] our BDFE results are also roughly in accord with computationally derived ferric heme hydroperoxides (those modeling proteins), where a proximal ligand is either an imidazolyl or —SH group, BDEs (64–66 kcal/mol).^[15]

HAT reactivity (shown in the diagonal of Figure 1) studies were performed with $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im}-S**) toward external substrates to support our experimentally determined BDFE value. Addition of an excess of substrates such as 9,10-dihydroanthracene (76 kcal/mol in DMSO)^[12] and xanthene (72.2 kcal/mol in THF)^[13] to the solutions of **P^{Im}-S** at –80 °C in THF led to no reaction. These observations are consistent with our BDFE determination of 69.5 kcal/mol for $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im}-HP**). In contrast, TEMPO-H (66.5 (THF), 72.6 (aq. buffer), and 69.6 (benzene) kcal/mol)^[13,16] does react with complex **P^{Im}-S**. The final spectra (UV-vis and EPR; Figure 6A, Figure S9 and S10) reveal a high yield of TEMPO•, while **P^{Im}-HP** seems to only form in small amounts (~20% yield), probably due to side-reactions. The results support the conclusion that an initial H-atom abstraction from TEMPO-H to give **P^{Im}-HP** occurred. Based on these reactivity studies, the BDFE of **P^{Im}-HP** can be bracketed to be, 66.5 < BDFE_{OO-H} < 72 kcal/mol, which is consistent with our experimentally determined BDFE value, 69.5 kcal/mol.

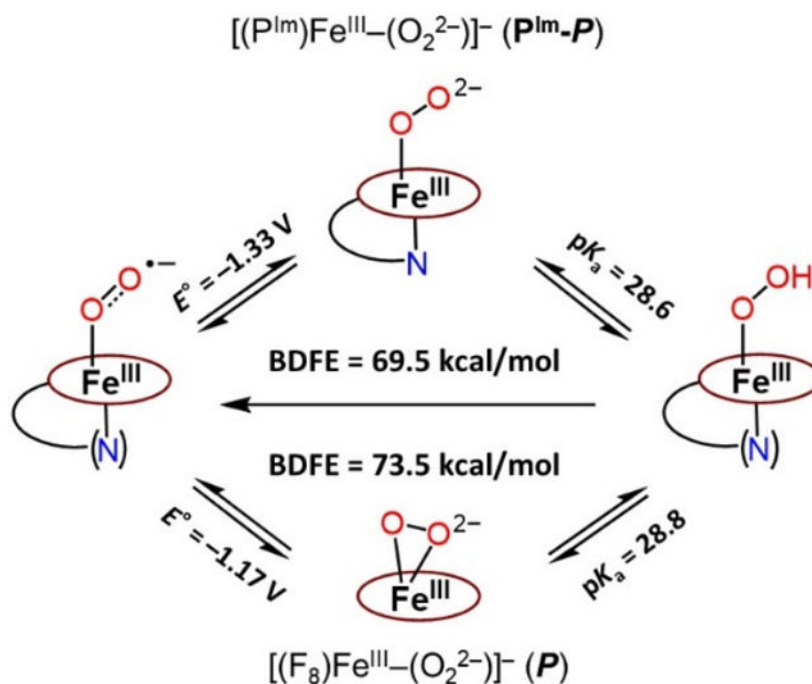
As we previously showed that $[(F_8)Fe^{III}-(O_2^{\bullet-})]$ (**S**) reacts with TEMPO-H to cleanly give the TEMPO• and corresponding hydroperoxide complex $[(F_8)Fe^{III}-(OOH)]$ (**HP**), we now see that **S** and $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im}-S**) reactions toward TEMPO-H are comparable. Thus, we sought to find a different substrate that might differentiate between the oxidizing capabilities of the two superoxide complexes, which is what we have found here based on our thermodynamic parameter analysis. The superoxide complex **S** should be a somewhat stronger oxidant for HAT chemistry as based on our finding of the ~ 4 kcal/mol difference in **HP** versus **P^{Im}-HP** complexes' BDFEs (vide supra).

Thus, we turned to the use of the hydroxylamine substrate ABNO-H (9-azabicyclo[3.3.1]nonane-*N*-hydroxide, BDFE = 76.0 (aq. buffer) and 76.2 (benzene) kcal/mol)^[16] whose BDFE value is in the range 3.4-to-6.6 kcal/mol higher than that of TEMPO-H. As summarized in Figure 6, there was no reaction between $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im}-S**) and ABNO-H (Figure 6A and Figure S11), while spectroscopic monitoring (UV-vis and EPR) of the corresponding $[(F_8)Fe^{III}-(O_2^{\bullet-})]$ (**S**) plus ABNO-H reaction (Figure 6B and Figure S12) revealed both the generation of the organic radical product (ABNO•) along with the formation of the low-spin end-on hydroperoxide species, **HP** (See Figure S12 for further details). The HAT reaction of ABNO-H with **P^{Im}-S** is thermodynamically uphill. In contrast, the oxidizing ability of **S** via HAT from ABNO-H is favored. ABNO-H is a borderline substrate which enables us to discern the clear difference in the reactivities between **P^{Im}-S**

and *S*; the results are fully consistent with our findings of heme-hydroperoxide OO—H BDFE's via thermodynamic analyses, here and previously.^[3]

Conclusion

In conclusion, using the P^{Im} iron-porphyrinate which includes an imidazolyl moiety appended to the periphery of a fluorinated tetraphenylporphyrin, we here generated a new (only the second one known) end-on low-spin ferric heme peroxide $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im} -P**) and also a hydroperoxide $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im} -HP**) species which are relevant to the catalytic cycle of CYP450s and many other heme enzymes which activate O_2 or H_2O_2 . The end-on peroxide binding mode with the P^{Im} heme contrasts with that of the F_8 heme (without the appended axial imidazolyl ligand) which forms a side-on peroxide species. The reversibility of the $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im} -S**)/ $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im} -P**) redox process and the $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im} -P**)/ $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im} -HP**) acid-base equilibrium allowed for the calculation of reduction potential and pK_a , respectively. With the measured thermodynamic parameters, the ferric heme OO—H BDFE was determined to be 69.5 kcal/mol, contrasting with the value of 73.5 kcal/mol found for $[(F_8)Fe^{III}-(OOH)]$ (**HP**).



This difference is a result of the existence of the appended axial imidazolyl ligand in the P^{Im} system. Its “push effect” appears to contribute significantly to the lower reduction potential found for (**P^{Im} -S**)/(**P^{Im} -P**) compared to that measured for the (*S*)/(*P*) couple with F_8 -heme. Confirmation of our thermodynamically derived parameters comes from comparison of the $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ (**P^{Im} -S**) versus $[(F_8)Fe^{III}-(O_2^{\bullet-})]$ (*S*) reactivity results employing the hydroxylamine substrates TEMPO-H and/or ABNO-H. Future studies will explore further

similar thermodynamic relationships to determine how ligand effects may control such properties and chemical reactivity.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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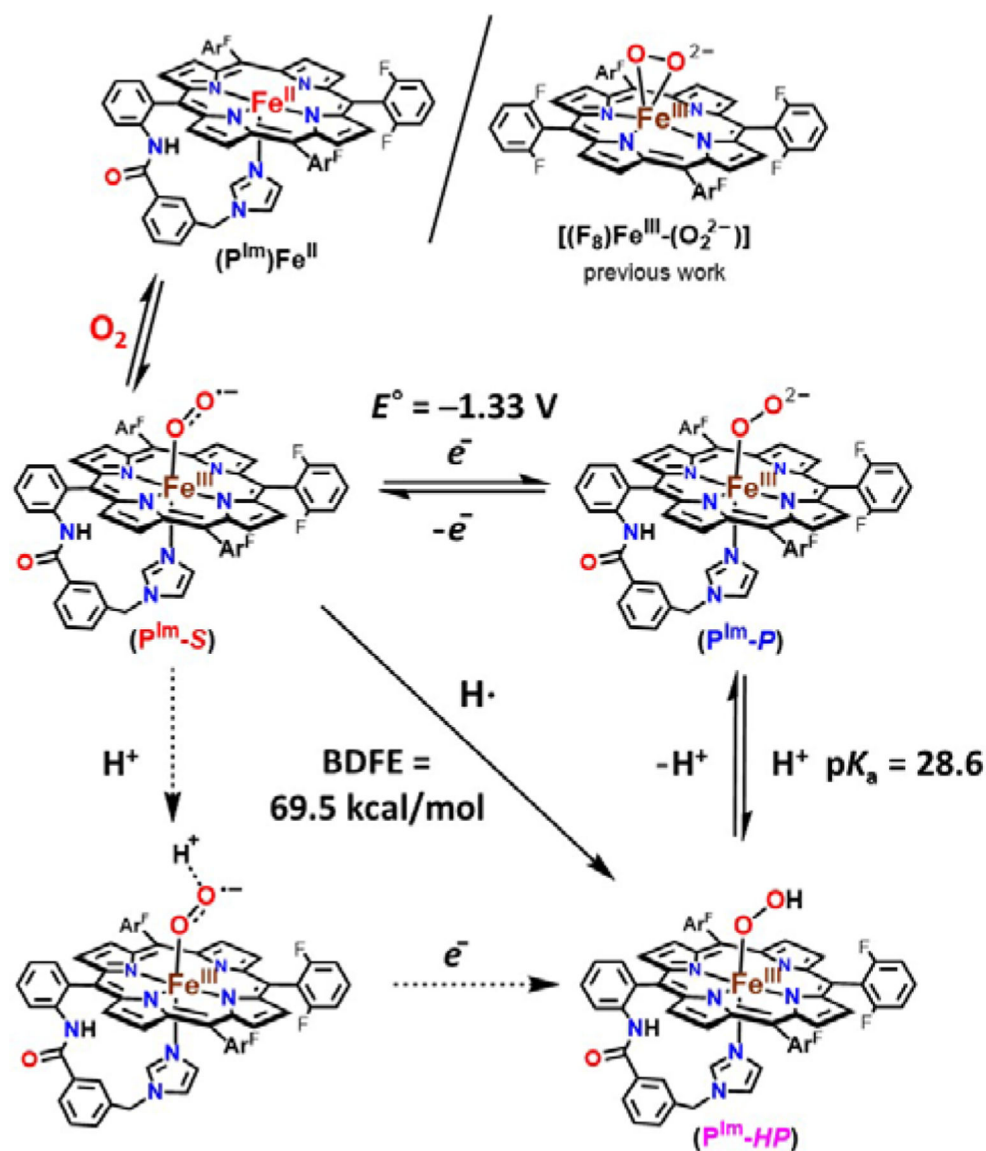


Figure 1. Stepwise generation of superoxide, peroxide, and hydroperoxide heme analogues and relevant thermodynamic square scheme (E° as determined vs $Fc^{+/0}$ and pK_a) and $OO-H$ BDFE of ferric heme hydroperoxide. Ar^F = 2,6-difluorophenyl group. Upper right: the previously studied ferric heme peroxide $[(F_8)Fe^{III}-(\eta^2-O_2^{2-})]^-$.^[3]

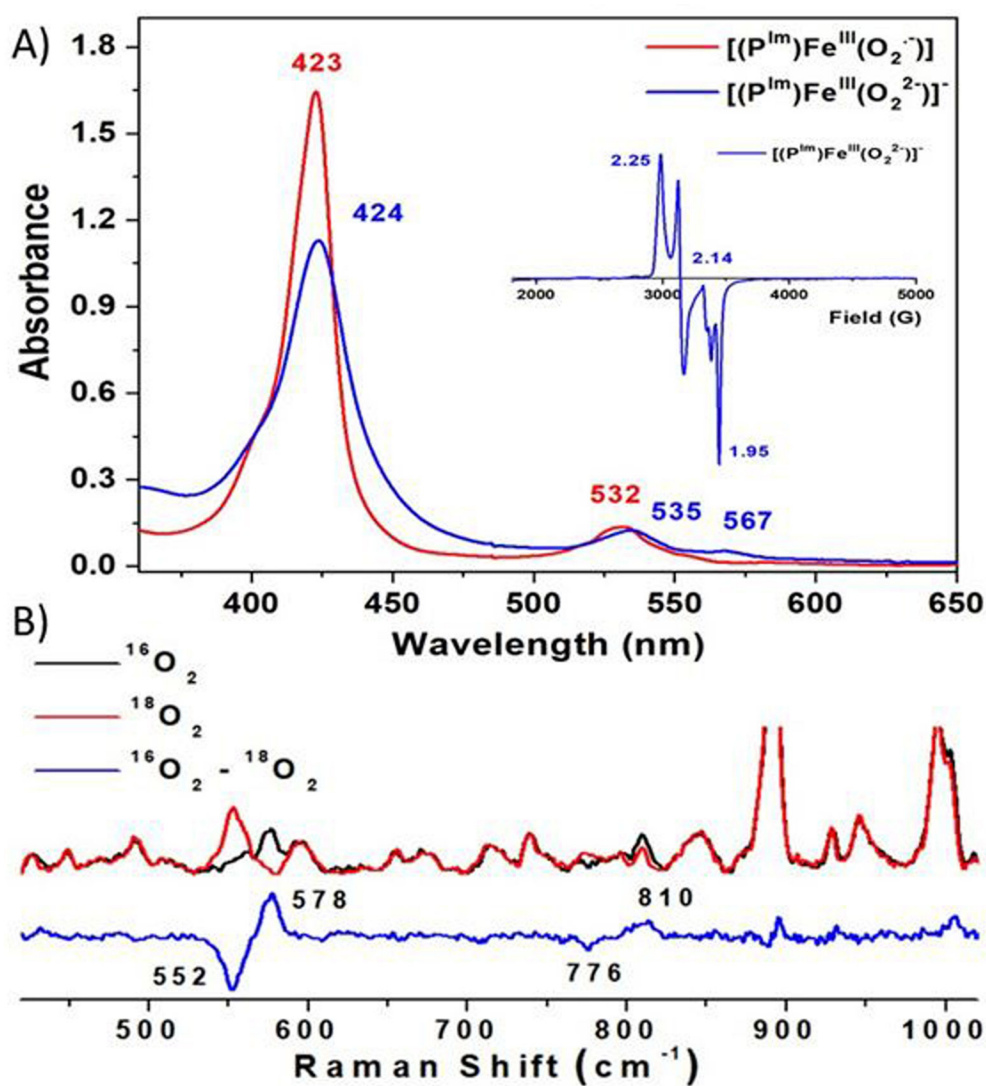


Figure 2. (A) Electronic absorption spectra of $[(P^{Im})Fe^{III}(O_2^{\bullet-})]$ (P^{Im-S}) (red) to $[(P^{Im})Fe^{III}(O_2^{2-})]^-$ (P^{Im-P}) (blue) at -80 °C in THF. Inset: Frozen THF solution EPR (10 K) spectrum of P^{Im-P} . The small feature ($g = 1.99$) corresponds to an excess of cobaltocene.^[6d] (B) Resonance Raman spectra of $[(P^{Im})Fe^{III}(O_2^{2-})]^-$ (P^{Im-P}) in frozen THF obtained at 77 K with 413 nm excitation. The $^{16}O_2$ — $^{18}O_2$ difference spectrum is shown in blue.

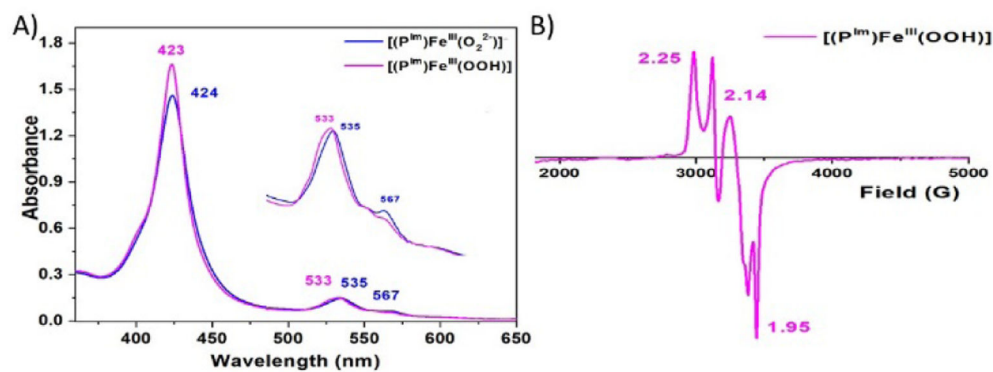


Figure 3.

(A) UV-vis spectra illustrating the conversion of $[(P^{Im})Fe^{III}-(O_2^{2-})]^-$ (**P^{Im}-P**) to $[(P^{Im})Fe^{III}-(OOH)]$ (**P^{Im}-HP**) by addition of $[(LutH^+)](OTf)$ at $-80\text{ }^\circ\text{C}$ in THF. (B) Low-spin EPR spectrum (10 K) of **P^{Im}-HP**. Note that there is $g = 1.99$ signal from excess of cobaltocene used for generation of the peroxide complex. An extra signal at $g > 2.00$ (compared to the EPR spectrum shown in Figure 2) is attributed to an unknown impurity.

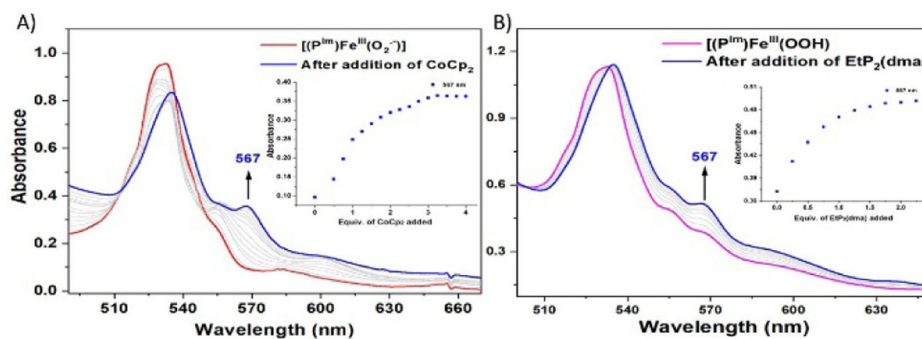


Figure 4. UV-vis spectroscopic monitoring of the incremental addition of (A) CoCp₂ to a solution of [(P^{Im})Fe^{III}—(O₂^{•-})] (**P^{Im}-S**) (red). Inset: monitoring of the absorbance at 567 nm (blue). (B) EtP₂(dma) addition to a solution of [(P^{Im})Fe^{III}—(OOH)] (**P^{Im}-HP**) (pink). Inset: monitoring of the absorbance at 567 nm (blue).

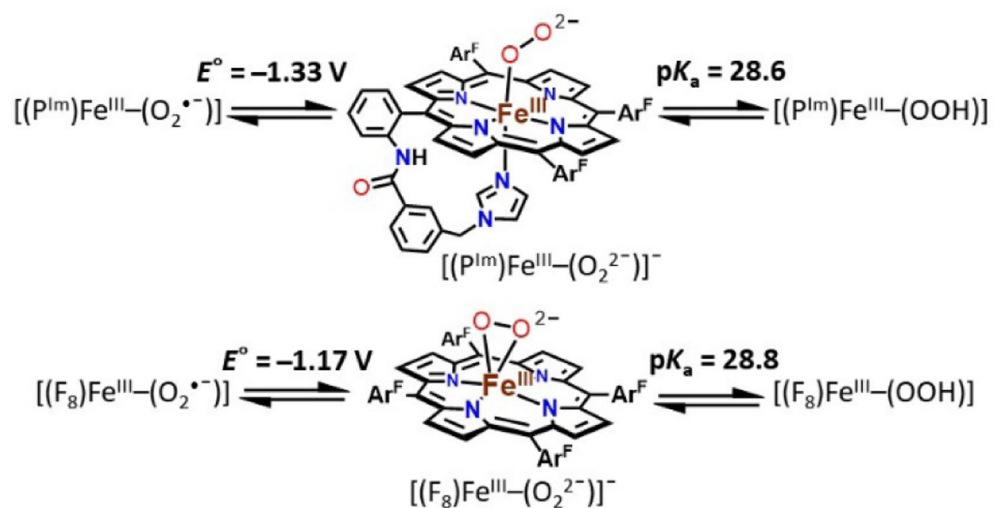


Figure 5. Comparison of P^{Im} and F₈ iron-porphyrinate complex for thermodynamic parameters and the geometry of peroxide species.

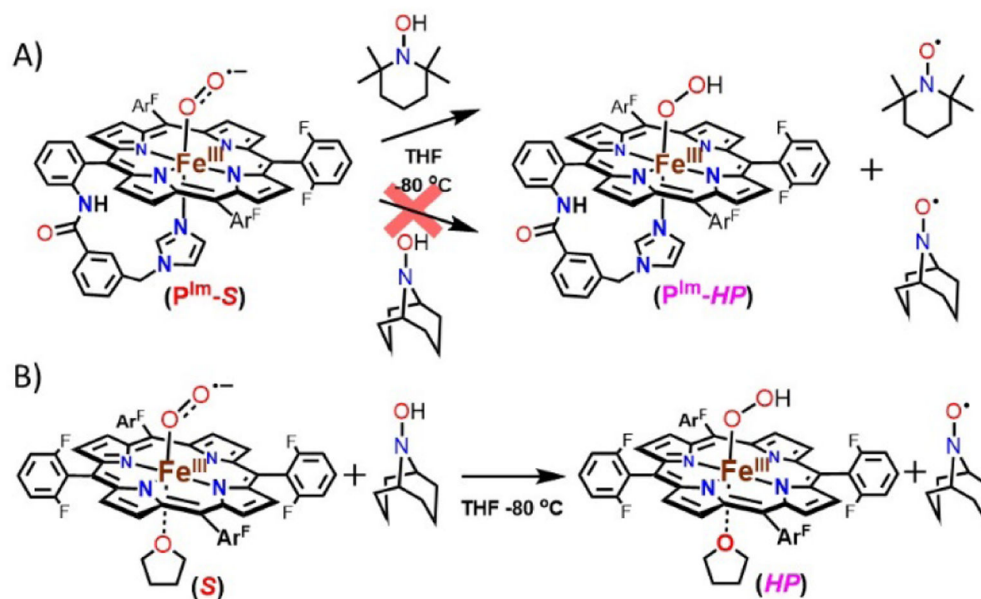


Figure 6.

(A) Hydrogen atom transfer (HAT) reaction by $[(P^{Im})Fe^{III}-(O_2^{\bullet-})]$ ($P^{Im}-S$) from TEMPO-H and ABNO-H at $-80\text{ }^\circ\text{C}$ in THF. (B) HAT from $[(F_8)Fe^{III}-(O_2^{\bullet-})]$ (S) toward ABNO-H at $-80\text{ }^\circ\text{C}$ in THF.