Published in final edited form as:

Nat Rev Dis Primers.; 6(1): 92. doi:10.1038/s41572-020-00224-3.

Head and neck squamous cell carcinoma

Daniel E. Johnson¹, Barbara Burtness², C. René Leemans³, Vivian Wai Yan Lui⁴, Julie E. Bauman⁵, Jennifer R. Grandis^{1,†}

¹Department of Otolaryngology-Head and Neck Surgery, University of California at San Francisco, San Francisco, CA, USA ²Department of Medicine, Yale University School of Medicine and Yale Cancer Center, New Haven, CT, USA ³Department of Otolaryngology-Head and Neck Surgery, Amsterdam University Medical Center, Vrije Universiteit Amsterdam, Cancer Center Amsterdam, Amsterdam, the Netherlands ⁴School of Biomedical Sciences, Faculty of Medicine, The Chinese University of Hong Kong, Hong Kong SAR ⁵Department of Medicine-Hematology/Oncology, University of Arizona, Tucson, AZ, USA

Abstract

Most head and neck cancers are derived from the mucosal epithelium in the oral cavity, pharynx and larynx and are known collectively as head and neck squamous cell carcinoma (HNSCC). Oral cavity and larynx cancers are generally associated with tobacco consumption, alcohol abuse or both, whereas pharynx cancers are increasingly attributed to infection with human papillomavirus (HPV), primarily HPV-16. Thus, HNSCC can be separated into HPV-negative or HPV-positive HNSCC. Despite evidence of histological progression from cellular atypia through various degrees of dysplasia, ultimately leading to invasive HNSCC, most patients are diagnosed with late-stage HNSCC without a clinically evident antecedent premalignant lesion. Traditional staging of HNSCC using the tumour-node-metastasis system has been supplemented by the 2017 AJCC/UICC staging system, which incorporated additional information relevant to HPV-positive disease. The treatment approach is generally multimodal, consisting of surgery followed by chemotherapy plus radiation (chemoradiation or CRT) for oral cavity cancers and primary CRT for pharynx and larynx cancers. The EGFR monoclonal antibody cetuximab is generally used in combination with radiation in HPV-negative HNSCC where co-morbidities prevent the use of cytotoxic chemotherapy. The FDA approved the immune checkpoint inhibitors pembrolizumab and

Author contributions

Introduction (J.R.G.); Epidemiology (V.W.Y.L.); Pathophysiology/mechanisms (D.E.J.); Diagnosis, screening and prevention (J.E.B.); Management (B.A.B.); Quality of life (C.R.L.); Outlook (D.E.J. and J.R.G.).

Competing interests

D.E.J. and J.R.G. are co-inventors of cyclic STAT3 decoy and have financial interests in STAT3 Therapeutics. STAT3 Therapeutics holds an interest in cyclic STAT3 decoy. B.A.B. has received honoraria for consulting from Merck and AstraZeneca. C.R.L. serves on the Advisory Board of Merck & Co. and Rakuten Medical. V.W.Y.L. receives grant support from Lee's Pharmaceutical, Hong Kong Limited, via the University-Industry Collaboration Program (UIM/329; from the Innovation and Technology Fund, Hong Kong government; in 2018–2020), and served as a scientific consultant for Novartis Pharmaceutical (Hong Kong) Limited (Oct 2015-Oct 2016). J.E.B. serves as a scientific consultant to CUE Pharmaceuticals and Astra Zeneca and has research grant support from the IST programs of Aveo and Novartis.

Peer review information

Nature Reviews Disease Primers thanks C. Chung, C. Van Waes, A. Dietz, J. Hess, C. Bradford and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

[†] jennifer.grandis@ucsf.edu.

nivolumab for treatment of recurrent or metastatic HNSCC and pembrolizumab as primary treatment for unresectable disease. Elucidation of the molecular genetic landscape of HNSCC over the past decade has revealed new opportunities for therapeutic intervention. Ongoing efforts aim to integrate our understanding of HNSCC biology and immunobiology to identify predictive biomarkers that will enable delivery of the most effective, least toxic therapies.

Introduction

Head and neck squamous cell carcinomas (HNSCCs) develop from the mucosal epithelium in the oral cavity, pharynx and larynx and are the most common malignancies that arise in the head and neck (Fig. 1). The burden of HNSCC varies across countries/regions and has generally been correlated with exposure to tobacco-derived carcinogens, excessive alcohol consumption, or both. Increasingly, tumours that arise in the oropharynx are linked to prior infection with oncogenic strains of human papillomavirus (HPV), primarily HPV-16, and, to a lesser extent, HPV-18 and other strains ¹⁻³. As the most common oncogenic HPVs, HPV-16 and HPV-18, are covered by FDA-approved HPV vaccines, it is feasible that HPVpositive HNSCC could be prevented by successful vaccination campaigns worldwide. HNSCCs of the oral cavity and larynx are still primarily associated with smoking and are now collectively referred to as HPV-negative HNSCC. No screening strategy has proved to be effective, and careful physical examination remains the primary approach for early detection. Although a proportion of oral premalignant lesions, which present as leukoplakia (white patches) or erythroplakia (red patches), progress to invasive cancer, the majority of patients present with advanced-stage HNSCC without a clinical history of a premalignancy. HNSCC of the oral cavity is generally treated with surgical resection, followed by adjuvant radiation or chemotherapy plus radiation (known as chemoradiation or CRT) depending on the disease stage. CRT has been the primary approach to treat cancers that arise in the pharynx or larynx. HPV-positive HNSCC generally has a more favourable prognosis than HPV-negative HNSCC, and ongoing studies are testing the efficacy of therapeutic dose reduction (of both radiation and chemotherapy) in HPV-positive disease treatment. With the exception of early-stage oral cavity cancers (which are treated with surgery alone) or larynx cancers (which are amenable to surgery or radiation alone), treatment of the majority of HNSCC cases requires multimodality approaches and thus multidisciplinary care. The epidermal growth factor receptor (EGFR; also known as HER1) monoclonal antibody cetuximab is approved by the FDA as a radiation sensitizer, alone or in combination with chemotherapy, for recurrent or metastatic disease⁴. Although inferior to cisplatin as a radiosensitizer in HPV-associated disease, ^{5,6} cetuximab is often used in cisplatin-ineligible patients. The immune checkpoint inhibitors pembrolizumab and nivolumab are approved by the FDA for treatment of cisplatin-refractory recurrent or metastatic HNSCC and pembrolizumab is approved as first-line therapy for patients who present with unresectable or metastatic disease^{7–9}. Detailed molecular characterization as well as immune profiling of HNSCC suggests that incorporation of prognostic and predictive biomarkers into clinical management may overcome obstacles to targeted therapies and enable prolonged survival. In this Primer, we provide an overview of the types of HNSCC and their epidemiology, as well as the pathogenesis of each type and how this influences the management approach.

Epidemiology

Incidence, prevalence and mortality

HNSCC is the sixth most common cancer worldwide, with 890,000 new cases and 450,000 deaths in 2018 (Fig. 2) $^{10-12}$. The incidence of HNSCC continues to rise and is anticipated to rise by 30% (that is, 1.08 million new cases annually) by 2030 (GLOBOCAN; gco.iarc.fr/ today)^{10–12}. The high prevalence of HNSCC in regions such as Southeast Asia and Australia is associated with consumption of specific carcinogen-containing products (described below), whereas increasing rates of oropharyngeal infection with HPV have contributed to the high prevalence of HNSCC in the USA and Western Europe^{13–15}. In general, men are at 2-4-fold higher risk than women for developing HNSCC. The median age of diagnosis for non-virally associated HNSCC is 66 years, whereas the median age of diagnosis for HPVassociated oropharyngeal cancer and Epstein-Barr virus (EBV)-associated nasopharyngeal cancer is ~53 years and ~50 years, respectively ^{16,17}. The survival rate for HNSCC has improved modestly over the past three decades, for example, the 5-year survival rate increased from 55% during the period 1992-1996 to 66% in the period 2002-2006 when analyzed across all age groups and anatomic sites within the SEER registry¹⁸. Subgroup analysis noted improved survival in all age groups except for older patients (>75 years) and all anatomic sites except larynx, where survival was stagnant. Improvement in survival is partially attributable to the emergence of HPV-associated HNSCC, a population with improved prognosis, rather than improvements in multimodality treatment per se; a subsequent SEER analysis incorporating tissue assessment for HPV noted improved survival in HPV-positive but not HPV-negative HNSCC¹⁹. In addition to deaths directly caused by HNSCC, survivors of this cancer have the second highest rate of suicide (63.4 cases per 100,000 individuals) after those with pancreatic cancer (86.4 cases per 100,000 individuals), compared with survivors of other cancers (23.6 cases per 100,000 individuals). Psychological distress and compromised quality of life (QOL) are likely key underlying factors for suicide²⁰.

Risk factors

Epidemiological studies have revealed a diverse range of risk factors for HNSCC, as classified by the International Agency for Research on Cancer (IARC) of the World Health Organization (WHO)²¹. These risk factors include tobacco consumption, alcohol consumption, exposure to environmental pollutants and infection with viral agents, namely, HPV and EBV. Interestingly, several risk factors display geographical or cultural and/or habitual prevalence. Tobacco and alcohol consumption are the high-risk factors that occur most widely geographically. Of note, heavy users of both substances have >35-fold higher risk of developing HNSCC²². Among some Asia-Pacific populations, oral cavity cancer is associated with chewing of areca nut products including 'betel quid', a term that applies to a variety of customized mixtures comprising areca nut (*Areca catechur*, the carcinogen source), betel leaf (the leaf of *Piper betle*), slaked lime and/or tobacco, as well as spices per local custom (IARC Working Group on the Evaluation of Carcinogenic Risk to Humans. Betel-quid and Areca-nut Chewing and Some Areca-nut-derived Nitrosamines. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, No. 85. Lyon (FR): International Agency for Research on Cancer. https://publications.iarc.fr/103 (2004))²¹. The use of areca

nut or betel quid products is linked to particularly high rates of oral cavity cancer in India (first and fourth most common cancer in Indian men and women, respectively), Taiwan and some provinces in mainland China²³. In general, the high male:female ratios for HPV-negative HNSCC incidence reflect the sex-specific patterns of modifiable risk behaviours, including the use of the aforementioned tobacco, smokeless tobacco, areca nut, betel quid and alcohol^{21,24}.

The effect of electronic cigarettes on HNSCC risk remains unknown and will only be evident in the coming decades. Exposure to carcinogenic air pollutants, including organic and inorganic chemicals, as well as particulate matter, is a risk factor for HNSCC, especially in developing countries/regions with worsening air pollution, such as India and China^{25,26}. Other risk factors include ageing, poor oral hygiene and diets lacking in vegetables^{27,28}. In terms of infectious agents, persistent infection with HPV and EBV are known aetiological risk factors for HNSCC arising from the oropharynx and nasopharynx, respectively^{29,30}. The male:female ratio for HPV-positive HNSCC incidence ranges from 3 to 6 (ref.³¹), which is explained by higher rates of persistent oropharyngeal HPV infection among males despite similar prevalence of anogenital HPV infection^{32–34}. HPV infection that leads to HNSCC is mainly transmitted by oral sex, and the incidence of HPV-positive HNSCC continues to rise, especially in populations that are not vaccinated against HPV prior to HPV exposure^{35,36}.

In addition, genetic factors also contribute to HNSCC risk. Individuals with Fanconi anaemia (FA), a rare, inherited genetic disease characterized by impaired DNA repair (owing to mutations in any of the 22 FANC genes), have a 500–700-fold increased risk of developing HNSCC, primarily cancers of the oral cavity³⁷. Although the mechanisms responsible for the unique proclivity of patients with FA to develop HNSCC remain unknown, alterations in FA pathway genes likely have a role. Meta-analyses showed that polymorphisms in genes involved in carcinogen metabolism and in immunity are associated with increased risk, including polymorphisms in cytotoxic T lymphocyte antigen 4 (*CTLA4*; rs231775 and rs4553808), *IL10* (1082A > G), cytochrome P450 1A1 (*CYP1A1*; Ile462Val and glutathione S-transferase µ1 (*GSTM1*; null polymorphism)^{38–41}. Thus, a reduced ability to metabolize carcinogens and weakened immunity may contribute to HNSCC. Decreased use of tobacco products, improved oral health and widespread HPV vaccination should help reduce the global incidence of HNSCC⁴².

Mechanisms/pathophysiology

Formation, progression, cell of origin

HNSCC originates from mucosal epithelial cells that line the oral cavity, pharynx, larynx and sinonasal tract. Histologically, progression to invasive HNSCC follows an ordered series of steps beginning with epithelial cell hyperplasia, followed by dysplasia (mild, moderate and severe), carcinoma *in situ* and, ultimately, invasive carcinoma (Fig. 3). However, of note, most patients diagnosed with HNSCC do not have a history of an antecedent premalignant lesion. Given the heterogeneous nature of HNSCC, the cell of origin depends on anatomical location and aetiological agent (carcinogen versus virus). However, in each case, normal adult stem or progenitor cells are likely candidates for the cell of origin, giving rise, following oncogenic transformation, to cancer stem cells (CSCs) with properties of self-

renewal and pluripotency. HNSCC CSCs with the capacity to generate tumours in transplantation assays constitute only a minor fraction (1-3%) of the cells in primary tumours⁴³ but, despite their inherent resistance to conventional drugs, represent attractive targets for novel targeting agents.

A number of molecular biomarkers of HNSCC CSCs have been proposed, with CD44, CD133 and ALDH1 being the most extensively validated and associated with prognostic significance. CD44 is a cell surface receptor for hyaluronic acid and matrix metalloproteinases (MMPs), which is involved in intercellular interactions and cell migration. HNSCC cells with high levels of CD44 are capable of self-renewal, and CD44 levels in HNSCC tumours are associated with metastasis and poor prognosis^{44,45}. Similarly, increased levels of the membrane-spanning protein CD133 are associated with HNSCC invasiveness and metastasis⁴⁶. ALDH1 is an intracellular enzyme that converts retinol to retinoic acid, plays a part in cellular detoxication and is a marker for both normal stem cells and CSCs. High levels of ALDH1 expression or activity are associated with self-renewal, invasion and metastasis, and may have prognostic significance in HNSCC⁴⁵. In addition, HNSCC cells with CSC properties express elevated levels of the stem cell markers OCT3, OCT4, SOX2 and NANOG, with the levels of these proteins correlating with tumour grade in oral cancers⁴⁷. Immunohistochemical analyses of HNSCC tumours indicate that ~80% of ALDH1-positive cells are in close proximity (100 µm) to a blood vessel, suggesting that the CSCs reside primarily in perivascular niches⁴³.

An important clinical phenomenon to consider when defining the cell of origin in HNSCC is the development of second primary tumours (SPTs). Synchronous and/or metachronous SPTs arise at an extraordinarily high rate after the diagnosis of an initial primary tumour and can be localized at distinct anatomical sites in the head and neck region, oesophagus or lungs^{48,49}. Frequently lethal, SPTs may share some molecular abnormalities with the initial primary tumour or may exhibit marked differences. The concept of 'field cancerization' suggests that carcinogens damage or condemn large anatomical fields⁵⁰. In tobacco-associated HNSCC, the size of the damaged anatomical field may increase with patient age^{51,52}. Considering the concept of field cancerization, the development of SPTs may reflect distinct CSCs arising from independent oncogenic transformations.

Initiating and early events

HPV-negative HNSCC.—Tobacco consumption is the primary risk factor for development of HPV-negative HNSCC. Tobacco consists of over 5,000 different chemicals, of which dozens have been shown to have carcinogenic activity. The chemicals thought to be most responsible for the cancer-causing effects of tobacco are polycyclic aromatic hydrocarbons (PAHs), including benzo(a)pyrene, and nitrosamines, including 4-(methylnitrosamine)-1-(3-pyridyl)-1-butanone (NNK) and N-nitrosonornicotine (NNN)^{53,54}. In smokeless tobacco, nitrosamines are the dominant carcinogen, whereas the carcinogens in areca nut and betel quid are poorly defined⁵⁵. Tobacco-derived carcinogens, including PAHs and nitrosamines, undergo metabolic activation, with detoxication enzymes and pathways promoting excretion (Fig. 4). However, many of the reactive metabolites of these carcinogens can also form covalent DNA adducts, which, if not properly repaired, lead to mutations and other genetic

abnormalities. The propensity for tobacco carcinogens to promote genetic changes and neoplastic transformation likely depends on the balance between metabolic activation versus detoxification and DNA repair (Fig. 4). The use of tobacco products is also associated with inflammation in the exposed tissues. Coincident with inflammation is the local production of cytokines, chemokines and growth factors that can have an important role in promoting proliferation, angiogenesis and, ultimately, carcinogenesis.

Excessive alcohol consumption is another key risk factor for HPV-negative HNSCC and is known to synergize with tobacco use to promote carcinogenesis⁵⁶. Alcohol might serve as a solvent for carcinogens, enhancing the exposure of epithelial cells to these substances⁵⁷. In addition, alcohol is metabolized to acetaldehyde, which is known to form DNA adducts⁵⁸.

HPV-positive HNSCC.—Infection with HPV is an increasingly common risk factor for HNSCC. HPV infection is associated with most oropharyngeal cancers (>70%) and a small minority of cancers at other head and neck anatomical sites^{1,2}. As described below, HPVpositive HNSCC exhibits distinct differences from HPV-negative HNSCC in gene expression, mutational and immune profiles, underscoring the unique biology of this disease. Infection with HPV is an early event in HPV-positive HNSCC, and most of these cancers arise from deep crypts in the palatine and lingual tonsils (Fig. 5). HPV-16 is the primary causative type, although other high-risk HPVs, including HPV-18, HPV-31, HPV-33 and HPV-52, are detected in a small percentage of patients³. The high-risk HPVs, including HPV-16, are small, double-stranded, circular DNA viruses with an ~8 kb genome. In HNSCC tumour specimens, the viral genome is typically found integrated at a single, albeit variant, genomic site⁵⁹. The genome consists of seven early genes (E1–E7) and two late genes (L1 and L2). The L1 and L2 genes encode viral capsid proteins, whereas the E1-E5 genes encode proteins that are primarily involved in replication and transcription of the viral genome. The bicistronic E6 and E7 genes are essential for oncogenic transformation of the host cell. E6 protein forms a complex with the cellular ubiquitination protein E6-AP and the tumour suppressor p53 to promote ubiquitination and proteasomal degradation of p53 (ref. ⁶⁰). In contrast to HPV-negative HNSCC, in which TP53 (encoding p53) is frequently deleted or mutated, this gene is rarely altered in HPV-positive HNSCC, as p53 is eliminated by the action of E6 (ref.⁵⁹). E6 may possess other transforming activities beyond p53 degradation but these functions are less well-characterized^{61–63}. E7 protein binds strongly to the cell cycle regulator retinoblastoma-associated protein (RB1), promoting proteasomal destruction of RB1 and the release of E2F family transcription factors⁶⁴. The liberated E2F proteins drive the cell cycle beyond the restriction point (also known as the G1–S checkpoint) and into S phase. E7 also interacts with and affects the levels and/or cellular activity of a number of other cell cycle regulatory proteins ^{62,65}. The disruption of RB1 function by E7 leads to a feedback upregulation of p16^{INK4A}, and detection of p16^{INK4A} expression is commonly used to classify oropharyngeal tumours as HPV-positive. In addition to E6 and E7, E5 also has a role in oncogenic transformation by helping to drive cell cycle progression^{66,67}.

Genomic alterations and key pathways

There is a tremendous need to identify molecular biomarkers that can be used to predict progression of premalignant HNSCC lesions, prognosticate survival, reveal new targets for intervention and predict response to therapeutic agents. The search for biomarkers has focused on defining the molecular abnormalities that characterize HNSCC. In this section, we highlight findings regarding genetic and epigenetic alterations, as well as dysregulation of cellular signalling pathways, which occur during HNSCC development.

HNSCC is characterized by genetic instability, with frequent loss or gain of chromosomal regions⁵⁹. The availability of a model of ordered histological progression of HNSCC has enabled assignment of some chromosomal abnormalities to specific stages of progression^{57,68} (Fig. 3). Loss of 9p21 occurs during progression of normal head and neck epithelial mucosa to hyperplasia. The 9p21 region includes the tumour suppressor genes (TSGs) *CDKN2A* (encoding the CDK4 and CDK6 inhibitor p16^{INK4A}) and *ARF* (encoding p14, a stabilizer of p53). Progression from hyperplasia to dysplasia is marked by loss of 3p21 and 17p13, the site of *TP53*. The transition from dysplasia to carcinoma *in situ* involves loss of 11q13, 13q21 and 14q32, whereas loss of 6p, 8, 4q27, and 10q23 is observed in the progression to invasive carcinoma. Collectively, these studies of chromosomal abnormalities reveal that multiple genetic alterations may be required for full transformation to invasive HNSCC⁶⁸. Furthermore, whether progression of HNSCC is strictly dependent on the temporal sequence of these alterations or, instead, on their collective accumulation, remains unresolved.

The Cancer Genome Atlas (TCGA) contains a wealth of data on copy number alterations (CNAs), mutational profiles, mRNA expression and microRNA (miRNA) expression from over 520 human HNSCC tumours⁵⁹. A detailed analysis of 279 of these tumours, consisting of 243 HPV-negative and 36 HPV-positive tumours, revealed a high degree of genomic instability; tumours had an average of 141 CNAs and 62 chromosomal structural abnormalities (for example, fusions)⁵⁹. HNSCC tumours exhibited frequent mutation of CDKN2A (22% of tumours) and TP53 (72% of tumours). Coupled with the aforementioned highly frequent chromosomal loss of CDKN2A and TP53 in HNSCC tumours, these TSGs are regarded as the most frequently altered genes in HNSCC. However, these alterations are largely restricted to HPV-negative tumours, owing to the action of HPV E6 and E7 proteins in eliminating p53 and RB1 (p16^{INK4A} inhibits phosphorylation of RB1). Mutational profiling revealed that HNSCC-associated mutations are statistically enriched in 11 genes⁵⁹. Interestingly, this list of frequently mutated genes is dominated by known and potential tumour suppressors, including TP53, CDKN2A, FAT1, NOTCH1, KMT2D, NSD1 and TGFBR2. PIK3CA, encoding the catalytic subunit of phosphoinositide 3-kinase (PI3K), is the only oncogene that is found to be frequently mutated (\sim 14%) in HNSCC^{59,69–72}. Mutations in RAS genes are infrequent, with HRAS mutations being the most common (~4% of tumours). Thus, in contrast to many other solid tumour malignancies that are frequently driven by mutations in RAS or other oncogenes, HNSCC might be more frequently driven by loss of tumour suppressors. Comparison of HPV-positive and HPVnegative tumours failed to detect previously reported differences in overall mutation rates⁷¹. However, HPV-positive tumours are uniquely enriched for frequent loss of TRAF3 and

amplification of *E2F1*, whereas HPV-negative tumours are enriched for *CDKN2A* and *TP53* alterations, frequent focal deletions in other TSGs (such as *NSD1*, *FAT1*, *NOTCH1* and *SMAD4*) and frequent focal amplification of the genes encoding the receptor tyrosine kinases EGFR, HER2 (also known as ERBB2) and FGFR1⁵⁹. Mutations in the genes encoding NRF2 (*NFE2L2*) and KEAP1 (*KEAP1*), key regulators of oxidative stress, are also common and occur exclusively in HPV-negative HNSCC⁵⁹.

Two additional members of the *TP53* gene family, *TP63* and *TP73*, are frequently altered in HNSCC. *TP63* encodes two major isoforms, Np63 and TAp63 (containing a truncated or complete transactivation (TA) domain, respectively), and is overexpressed in a majority of HNSCC tumours⁷³. Np63 promotes HNSCC tumour growth by multiple mechanisms, including suppression of apoptosis and p16^{INK4A} expression and induction of mitogenic signalling^{73–76}. By contrast, TAp73, a major isoform encoded by *TP73*, exhibits tumour suppressor activity and the function of TAp73 is commonly abrogated in HNSCC. For example, stimulation of HNSCC cells with TNF results in the induction of c-REL oncoprotein that binds to Np63, displacing TAp73 from Np63–TAp73 complexes and inactivating TAp73^{77,78}. Phosphorylation of Tap73 by casein kinase 2 or Polo-like kinase 2 also leads to TAp73 inactivation and results in induction of NANOG, SOX2 and OCT4, promoting the stem cell-like properties of HNSCC tumour cells^{79,80}.

In addition to genetic alterations, epigenetic changes also have a role in driving HNSCC oncogenesis. Although HNSCC tumours are characterized by global hypomethylation of DNA, hypermethylation and resultant downregulated expression of key TSGs, including *CDKN2A*, *RARB* (encoding RARβ), *DCC* and *MGMT*, occurs frequently^{81–83}. MGMT is notable, as this protein is involved in the repair of DNA damage from tobacco carcinogens, whereas RARβ mediates epithelial cell differentiation.

Numerous studies have demonstrated aberrant expression of signalling proteins and/or activation of signalling pathways in HNSCC tumours. EGFR is overexpressed in 80–90% of HNSCC tumours and is associated with poor overall survival (OS) and progression-free survival (PFS)^{84,85}. Molecular targeting of EGFR with monoclonal antibodies (such as cetuximab) is an FDA-approved strategy for inhibiting EGFR signalling in HNSCC. Overexpression of other receptor tyrosine kinases, including HER2 and MET, also occurs and may contribute to HNSCC resistance to EGFR-targeting agents^{73,86,87}. Overexpression of the cytokine IL-6 and its receptor is also implicated in poor prognosis of HNSCC^{88,89}. Amplification of *CCND1*, resulting in overexpression of cyclin D1, is associated with the progression of dysplastic lesions to carcinoma *in situ* and with poor clinical prognosis^{57,90} (Fig. 3). Aberrant expression of MMPs and HIF1α and their effect on HNSCC progression are discussed later.

Among signalling pathways that commonly drive tumour development, the PI3K–AKT–mTOR pathway is the most frequently altered oncogenic pathway in HNSCC^{59,91}. Components of this pathway are genetically altered in most HNSCC tumours, with frequent alterations in *PIK3CA* occurring by both mutation (14%) and gene amplification (16%). Loss of function of phosphatase and tensin homologue (PTEN), a negative regulator of PI3K–AKT signalling, results from both genetic and epigenetic alterations and occurs in

~30% of HNSCC tumours 92 (Fig. 3). STAT3 signalling is hyperactivated in HNSCC and correlates with poor prognosis, although STAT3 is rarely mutated 93,94 . Mutations in the genes encoding the protein tyrosine phosphatase receptors (PTPRs) PTPRT and PTPRD, which occur frequently in HNSCC, have been demonstrated as one cause of STAT3 hyperactivation in head and neck cancers 95,96 . STAT3 signalling drives the expression of genes that promote cellular proliferation and survival as well as genes encoding growth factors and cytokines that promote immunosuppression (such as VEGF, IL-6, IL-10 and TGF β) 94 . Oncogenic signalling by the WNT- β -catenin pathway also contributes to HNSCC 97 . Last, the RAS-MAPK pathway, which contributes to the growth and survival of HNSCC tumour cells, has been reported by one group to be infrequently mutated in HNSCC, whereas another group reported mutations of this pathway in 18% of HNSCC tumours 59 .

Tumour microenvironment

The tumour microenvironment (TME) in HNSCC is a complex and heterogeneous mix of tumour cells and stromal cells, which include endothelial cells, cancer-associated fibroblasts (CAFs) and immune cells. Both tumour cells and CAFs produce growth factors, such as VEGF, which recruit endothelial cells, stimulating neovascularization and supply of oxygen and nutrients to the tumour. In turn, endothelial cells secrete factors that support the survival and self-renewal of CSCs⁴³. CAFs have a key role in HNSCC progression and are distinguished from normal fibroblasts by a persistent state of activation and expression of α -smooth muscle actin (α SMA)⁹⁸. CAFs secrete a broad range of growth factors (such as EGF, VEGF and HGF), cytokines (such as IL-6) and chemokines that promote tumour cell growth, angiogenesis and recruitment of immunosuppressive immune cells^{99,100}. In addition, CAFs are the primary source in the TME of MMPs^{99,100}, which are involved in degradation and remodelling of the extracellular matrix (ECM) and the release and activation of matrix-embedded growth factors (such as FGFs, VEGF and TGF β) that further stimulate tumour cell proliferation, angiogenesis and immunosuppression. Elevated β -SMA levels in HNSCC tumours correlate with poor prognosis¹⁰¹.

The TME of HNSCC tumours also includes innervating neurons derived from the peripheral nervous system. In addition, HNSCC tumours contain newly formed adrenergic neurons whose presence acts to stimulate tumour growth¹⁰². Loss of *TP53* seems to have a role in reprogramming sensory neurons in the HNSCC TME to an adrenergic, tumour promoting phenotype¹⁰².

Immune evasion.—The immune component of the HNSCC TME consists of tumour-infiltrating lymphocytes (TILs; including T cells, B cells and natural killer (NK) cells) and myeloid-lineage cells (including macrophages, neutrophils, dendritic cells and myeloid-derived suppressor cells (MDSCs)). In general, HNSCC tumours are highly infiltrated by immune cells, although the extent and composition of the immune cell infiltrate varies according to anatomical subsite and aetiological agent (smoking versus HPV)^{103,104}. Furthermore, distinct immune phenotypes have been identified for HNSCC tumours, and several molecular signatures, incorporating diverse markers, have been identified as classifiers^{105–107}. These signatures might prove useful for predicting response to different

therapies, particularly checkpoint inhibition. High levels of TILs generally correspond to better outcomes in HNSCC but this is dependent on the balance of cells with anti-tumour activity (effector T (T_{eff}) cells) versus those with immunosuppressive activity (regulatory T (T_{reg}) cells) cells in the TIL population 108,109 . Numerous studies have reported that the TME of most HNSCC tumours is highly immunosuppressive 104,110 . Anti-tumour immunity in the TME is mediated largely by T_{eff} cells and NK cells, whereas immune suppression and tumour cell growth is mediated by T_{reg} cells, MDSCs and M2 macrophages. Elevated levels of CD8+ T_{eff} cells and NK cells in the TME are associated with improved survival 100 . By contrast, elevated levels of T_{reg} cells, MDSCs, neutrophils or M2 macrophages are associated with advanced stage HNSCC or poor prognosis 100 .

The immune cell abundance and composition of HPV-positive tumours is notably different from HPV-negative tumours ^{103,104}. HPV-positive tumours typically have a greater abundance of TILs than HPV-negative tumours. Importantly, patients with HPV-positive tumours containing high levels of TILs have excellent outcomes, whereas patients with HPV-positive tumours containing low levels of TILs exhibit survival outcomes similar to those of patients with HPV-negative HNSCC¹¹¹.

HNSCC tumours evade immune surveillance by a number of different mechanisms. The milieu of the HNSCC TME is rich in immunosuppressive growth factors and cytokines that promote recruitment or activity of MDSCs, T_{reg} cells and M2 macrophages while inhibiting the anti-tumour effects of $T_{\rm eff}$ cells and NK cells, of which IL-6, IL-10, VEGF and TGF β are particularly important⁹⁹. Factors in the HNSCC TME, including IL-10 and TGFβ, also promote macrophage polarization to the immunosuppressive M2 phenotype¹¹². Genetic and epigenetic alterations result in decreased tumour cell levels of human leukocyte antigen (HLA) and defects in antigen processing, leading to decreased recognition and cytolysis of tumour cells^{113,114}. In addition, HNSCC tumours, particularly advanced stage cancers, demonstrate upregulation of programmed cell death 1 ligand 1 (PDL1), which attenuates the cytolytic activity of T cells^{7,8}. Similarly, MDSCs and T_{reg} cells recruited to the HNSCC TME express PDL1 and another immunosuppressive molecule, cytotoxic T lymphocyte antigen 4 (CTLA4), respectively. In the specific case of HPV-positive HNSCC, the viral E5, E6 and E7 proteins promote immune evasion by effects on cellular gene and protein expression in tumor cells^{115,116}. The frequent loss in HPV-positive HNSCC of TRAF3, which encodes a protein that functions in antiviral immunity, also likely contributes to immune evasion in HPV-positive disease^{117,118}.

Hypoxia.—HNSCC tumours are also characterized by hypoxia, with roughly similar levels of hypoxia in HPV-positive and HPV-negative tumours of the same stage¹¹⁹. High levels of hypoxia in tumours portend poor prognosis and resistance to radiation therapy^{120–124}. Hypoxia induces the expression of HIF1α, a subunit of the transcription factor HIF1, which drives expression of a range of genes encoding proteins that promote angiogenesis (VEGF) and degradation of ECM (MMPs). HIF1 also upregulates tumour cell expression of glucose transporters (for example, GLUT1) and enzymes that contribute to metabolic reprogramming of the tumour from oxidative phosphorylation to glycolysis (known as the Warburg effect)⁷³. In tumour-infiltrating immune cells, hypoxia and HIF1 induce expression

of pro-inflammatory and immune-modulatory cytokines and chemokines (such as IL-1 β and TNF)⁷³.

The oral microbiome.—An emerging field of study is the role of the microbiota in the HNSCC TME, particularly in oral cancer. Poor oral health is associated with oral cancer, as well as other cancers, and tobacco, alcohol and HPV are all known to modulate the composition of the oral cavity microbiota¹²⁵. Specific bacterial and fungal signatures that are enriched in oral cancer have been reported and continue to be refined^{126,127}.

Metastasis

Multiple pathways and processes contribute to the invasion and metastasis of HNSCC tumour cells. MMPs produced by both tumour and stromal cells in the TME have an integral role in degrading and remodelling the ECM, thereby promoting tumour cell invasion. High levels of MMP2, MMP9 and MMP13 in HNSCC tumours are associated with invasion, metastasis and poor prognosis ^{128–130}. Interestingly, the HNSCC CSC marker CD44 serves as a cell surface receptor that binds to and promotes the activity of MMP9^{44,131}. CD44 and MMP9 co-localize to the invasive front of HNSCC tumours and their expression levels correlate with metastasis ¹³².

The epithelial—mesenchymal transition (EMT; the conversion of tumour cells from an epithelial to a mesenchymal phenotype) has a key role in HNSCC metastasis. Cells that undergo EMT exhibit downregulation of E-cadherin, upregulation of vimentin, reduction of cell adhesion and enhanced migration and invasiveness. EMT-associated changes in E-cadherin and vimentin levels are associated with increased metastasis of HNSCC tumours 100,133. The transcription factors TWIST, SNAIL and SLUG mediate downregulation of E-cadherin during EMT, with co-expression of TWIST, SNAIL and HIF1α correlating with high rates of HNSCC metastasis 134,135. Hypoxic conditions in the TME can drive EMT in tumour cells, as HIF1α induces the expression of vimentin, TWIST and SNAIL 100; this mechanism may explain the association of elevated levels of hypoxia with HNSCC metastasis 120,121,134. The process of EMT is also closely linked with the acquisition of stem cell properties, including expression of HNSCC CSC markers 135.

Tumour cell expression of CD44, CD133 or ALDH1 is closely associated with metastasis in HNSCC 45. Similarly, OCT3, OCT4 and NANOG promote invasiveness of HNSCC cells and may serve as a predictor of metastasis 45.

Of note, EMT is not a fixed and irreversible process¹³⁶ but instead can be context-dependent, occurring in distinct cellular populations at particular sites within the tumour^{136,137}. Reversal of EMT may be necessary for the establishment of macrometastatic tumour sites, underscoring the plasticity of the process¹³⁶. Furthermore, tumour cells may exhibit only partial EMT (p-EMT), a partial acquisition of EMT markers or properties. Single-cell transcriptomic analyses of primary and metastatic HNSCC tumours revealed that cells with p-EMT are localized to the leading edge of tumours and that p-EMT can serve as an independent predictor of metastasis, tumour grade and detrimental pathologic features¹³⁷.

The capacity of HNSCC tumour cells to metastasize also requires an ability to detach from the basement membrane and associated ECM components. Typically, when normal epithelial

cells detach they lose access to key survival factors and undergo a form of programmed cell death termed anoikis ¹³⁸. Metastasis is dependent on suppression of, or acquired resistance to, anoikis. Growth factors and cytokines in the TME, particularly IL-6, EGF and HGF, activate tumour cell signalling pathways, including the RAS–MAPK, PI3K–AKT–mTOR and STAT3 pathways, which suppresse anoikis ^{100,139,140}. Of note, anoikis-suppressive factors in the TME are derived from CAFs, endothelial cells, infiltrating immune cells and the tumour cells themselves ¹⁰⁰, underscoring the complex cross-talk between cell types that contributes to metastasis in HNSCC.

Diagnosis, screening and prevention

Clinical presentation

HNSCC is a cancer of adults, with a median age at diagnosis of 66 years for HPV-negative HNSCC, 53 years for HPV-positive HNSCC and 50 years for EBV-positive HNSCC^{16,17,141}. Irrespective of environmental or viral aetiology, males have a substantially higher risk than females for all forms of HNSCC. The classical presenting symptoms of HNSCC depend on both the anatomical site of the primary tumour and the aetiology of the tumour (either environmental carcinogens, HPV or EBV).

Oral cavity tumours.—Cancers of the oral cavity, including the mobile tongue, floor of mouth, buccal mucosa, alveolar ridges, retromolar trigone and hard palate, classically present with a non-healing mouth sore or ulcer. Oral cavity tumours are often diagnosed at an early stage due to the patient's self-identification of the mass lesion and symptoms that interfere with the fundamental functions of eating and speaking, such as pain with chewing or dysarthria (difficulty speaking). Clinical suspicion should be heightened by the coexistence of risk factors for environmental carcinogenesis, including use of smokeless or combustible tobacco, areca nut and betel quid, and alcohol, and poor dentition.

p16^{INK4A}-negative oropharyngeal and hypopharyngeal tumours.—Primary tumours of the oropharynx, including the base of the tongue, the palatine tonsils, the soft palate and the hypopharynx, typically become symptomatic at a later stage, owing to their hidden anatomical location. When present, symptoms such as dysphagia (difficulty eating), odynophagia (pain when swallowing) or otalgia (ear pain) often herald a more advanced tumour. A personal history of combustible tobacco and alcohol use should raise the index of suspicion when such non-specific pharyngeal symptoms are present.

Laryngeal tumours.—Patients with cancers of the larynx frequently present with voice changes or florid hoarseness, resulting in diagnosis at an early stage. If tumours are neglected, patients can present with dyspnea (difficulty breathing) and, ultimately, airway obstruction, prompting tracheostomy. The behavioural risk factors of combustible tobacco and alcohol use are multiplicative at the larynx and other HPV-negative sites ¹⁴².

p16^{INK4A}-positive oropharyngeal tumours.—Patients with HPV-related cancers of the oropharynx most commonly present with a new, painless level II (lymph nodes at the upper jugular level) neck mass and an asymptomatic primary tumour. Because the risk factors for HPV-negative HNSCC are often absent, keen awareness of the 21st century epidemic of

HPV-related HNSCC in North America and Western Europe is crucial for appropriate suspicion and early diagnosis. The dominant risk factors for HPV-related HNSCC include male sex and lifetime number of sexual partners³⁴. However, the average latency period of 10–30 years from initial oral infection to ultimate diagnosis of p16^{INK4A}-positive oropharyngeal cancer means that risk factors might be remote¹⁴³. Thus, in all adults, a new neck mass is considered malignant until demonstrated otherwise. In a systematic review of minimally invasive transoral surgery to diagnostically assess an occult primary HNSCC tumour presenting as a cervical neck mass, the majority of tumours were p16^{INK4A}-positive¹⁴⁴. Diagnostic transoral robotic surgery, including palatine and lingual tonsillectomies, identified ~70% of primary HNSCC tumours, which were most commonly localized in the base of tongue¹⁴⁵.

EBV-positive nasopharyngeal tumours.—The most common presenting symptoms of nasopharyngeal carcinoma include a cervical neck mass, epistaxis (nosebleeds) and unilateral nasal obstruction ¹⁴⁶. Owing to the proximity of the nasopharynx to the base of skull, more advanced tumours can present with conductive hearing loss or cranial nerve palsies. In east and southeast Asia, where EBV-positive nasopharyngeal carcinoma is endemic, the presentation of a new neck mass in an adult requires a complete head and neck evaluation with close attention to the nasopharyx.

Diagnosis

Histopathology.—The diagnosis of HNSCC must be established by biopsy of the primary tumour and/or neck mass¹⁴⁷. The biopsy method depends upon the location of the lesion. Primary tumours are typically approached with cup forceps, incisional biopsy or excisional biopsy, whereas the suspicious cervical neck mass should undergo fine needle aspiration (FNA). Excisional biopsy of a neck mass is not recommended unless the FNA biopsy sample has been persistently non-diagnostic, a primary site has not been identified on cross-sectional imaging or panendoscopy and/or lymphoma is suspected due to concurrent noncervical lymphadenopathy.

As HNSCC is derived from the stratified epithelium of the upper aerodigestive mucosa, the histopathological spectrum is characterized by the extent of cellular atypia and squamous differentiation (Fig. 6). A well-differentiated tumour closely resembles the stratified epithelium, with mature-appearing cells organizing into layers with irregular keratinization, most classically manifesting as a 'keratin pearl' (Fig. 6a). A poorly differentiated tumour is characterized by immature cells with nuclear pleomorphism and atypical mitoses, with minimal to no organized stratification or keratinization (Fig. 6b). Of note, the observed extent of squamous differentiation is closely associated with aetiology: HPV-negative HNSCCs are more often moderately or well differentiated, with preservation of stratification and keratinization, whereas HPV-positive HNSCCs are poorly differentiated or even display basaloid morphology on histopathological analysis⁵⁷.

The diagnosis of HNSCC can usually be made on the basis of routine histopathology with haemotoxylin and eosin staining. However, in the case of poorly differentiated or basaloid tumours, immunohistochemistry may be necessary to confirm an epithelial origin. HNSCC

tumours routinely stain with pancytokeratin antibodies and markers of squamous differentiation, including antibodies against cytokeratin 5, cytokeratin 6 and p63 (ref. ¹⁴⁸). HPV testing is mandatory for all oropharyngeal or unknown primary tumours, as it is a major determinant of modern staging and prognosis. Multiple techniques are available for determination of HPV status, including detection of E6 and E7 mRNA (by reverse transcription PCR), HPV DNA (by PCR or in situ hybridization) or the cell cycle protein p16^{INK4A} (by immunohistochemistry) ¹⁴³. Consensus recommendations from the College of American Pathologists include testing all newly diagnosed oropharyngeal cancers for HPV by immunohistochemistry for p16^{INK4A}, with a diagnostic threshold of diffuse nuclear and cytoplasmic staining in >70% of tumour cells ¹⁴⁹ (Fig. 6c,d). Of note, the level of p16^{INK4A} is not a direct measurement of HPV infection; p16^{INK4A} is upregulated on the degradation of RB, and as such, is a functional surrogate for levels of the oncoprotein E7.

Staging.—Stage and HPV status are now recognized as the major determinants of HNSCC prognosis in North America and Western Europe, although HPV-positive HNSCC was not identified as a disease with a distinct aetiology, molecular characteristics and prognosis until 2010 (ref. 150). Until 2017, the staging of HNSCC involved classification of disease within each classic anatomical subsite (oral cavity, oropharynx, hypopharynx or larynx) according to the tumour-node-metastasis (TNM) system, independent of HPV status ¹⁵¹. In 2017, the Union for International Cancer Control (UICC) and American Joint Commission on Cancer (AJCC) released the 8th edition of the Cancer Staging Manual ¹⁵². This version contained three important changes to improve hazard discrimination and outcome prediction across HNSCC, including the addition of depth of invasion to tumour staging in oral cavity cancers, the addition of extracapsular nodal extension to nodal staging in non-viral HNSCC and codification of a novel staging system for HPV-positive HNSCC¹⁵². The new staging system for p16^{INK4A}-positive oropharyngeal cancer improved prognostic discrimination compared with UICC/AJCC version 7, when applied to retrospective cohorts 153-155. The revision of nodal staging, in particular the classification of any number of ipsilateral nodes <6 cm (UICC/AJCC version 7 N1-N2b) into N1 disease, and the influence of nodal stage on overall stage group, resulted in a large stage migration from stages III-IVa in version 7 to stages I-II in version 8. Although hazard discrimination has likely improved for HPVpositive HNSCC, prospective outcomes data are needed to validate the version 8 staging system. Of note, version 8 does not account for tobacco use, which may be an important modifier of prognosis in p16^{INK4A}-positive oropharyngeal cancer ¹⁵⁰. Unlike HPV-positive HNSCC, current staging of nasopharyngeal carcinoma in the UICC/AJCC 8th edition is strictly based on anatomy, without incorporation of viral or environmental aetiology. Nomograms incorporating baseline plasma EBV DNA into the staging system significantly improve hazard discrimination for EBV-positive nasopharyngeal carcinoma, which is likely to be incorporated in subsequent versions of the Cancer Staging Manual 156,157.

Following histopathological confirmation of HNSCC, the staging evaluation includes the following procedures in all patients irrespective of HPV status: complete head and neck examination with direct inspection of the oral cavity and fibre-optic nasopharyngolaryngoscopy as indicated; cross-sectional imaging of the head and neck by CT or MRI to establish the extent of locoregional disease; and chest CT to rule out distant

metastatic disease. Where available, PET–CT is preferred for distant metastatic staging in patients with locally advanced tumours or nodal disease¹⁵⁸. In patients with tobacco-related HNSCC, the risk for a second tobacco-related primary tumour of the upper aerodigestive tract also warrants staging with panendoscopy under anaesthesia, including direct laryngoscopy, oesophagoscopy and bronchoscopy¹⁵⁹. As patients with HPV-positive or EBV-positive HNSCC rarely manifest SPTs, panendoscopy can be omitted in most cases¹⁴³.

Primary prevention

'Primary prevention' involves interventions to reduce the incidence of disease in the first place, by decreasing exposures, altering modifiable behaviours or increasing resistance in healthy people who are at risk. When considered as a target for primary prevention, most cases of HNSCC would indeed be preventable with successful global elimination of tobacco use and implementation of HPV vaccination.

Tobacco and areca nut use.—Worldwide, the majority of HNSCC cases are caused by individual use of tobacco, a known carcinogen. Tobacco use, whether smokeless (chew, snuff) or combustible (cigarettes, cigars or pipes), and chewing of areca nut products including betel quid, are modifiable risk factors. Evidence-based interventions for tobacco cessation at the individual level include programmes of behavioural and psychological support that identify tobacco users, provide personalized advice to quit, assess readiness to quit, provide assistance with a quit attempt and arrange follow up. Quit attempts are more likely to be successful with the use of at least one of seven US FDA-approved first-line pharmacotherapies, including 5 methods of nicotine replacement therapy (NRT; the nicotine patch, gum, lozenge, nasal spray or oral inhaler) and 2 non-nicotine oral medications, bupropion and varenicline 160. Evidence-based interventions at the national policy level have been ranked according to cost-effectiveness by the WHO, which classifies the following four strategies as 'best buys': increased excise taxes on tobacco products; large graphic warnings on cigarette packages; comprehensive bans on tobacco advertising; and enforcement of a national, comprehensive, ban on smoking in public spaces¹⁶¹. Unlike tobacco use, for which the WHO has published evidence-based policies for cessation, no global policy exists for the control of areca nut or betel quid use¹⁶². As with tobacco, the areca nut is addictive, although the biological basis for addiction is less well understood. Research into both the mechanisms and social determinants of dependence is required in order to advance prevention and cessation programmes.

HPV vaccination.—Three prophylactic HPV vaccinations are currently approved by the FDA and each is recommended for a specific sex and age group: the bivalent vaccine (HPV-16 and HPV-18) for females 9–26 years of age, the quadrivalent vaccine (HPV-16, HPV-18, HPV-6 and HPV-11) for females and males 9–26 years of age, and the nonavalent vaccine (HPV-16, HPV-18, HPV-6, HPV-11, HPV-31, HPV-33, HPV-45, HPV-52 and HPV-58) for females and males 9–45 years of age. Whereas the indications for prophylactic HPV vaccination were established from the results of randomized, placebo-controlled trials evaluating the burden of anogenital HPV infections, the effectiveness against oral HPV infection has been analyzed retrospectively in three populations. In Costa Rica, a phase III trial of the bivalent vaccine demonstrated 93% efficacy against oral infection with HPV-16

or HPV–18163. Young adults 18–30 years of age with a self-reported history of HPV vaccination demonstrated a considerably lower rate of vaccine-type-specific oral HPV infection, in repeated cross-sectional data analysis of the National Health and Nutrition Examination Survey (NHANES) 2009–2014¹⁶⁴. Finally, a cross-sectional study of NHANES participants 18–33 years of age estimated the individual and population level efficacy of the quadrivalent HPV vaccine. Efficacy against HPV vaccine-type-specific infection was 88% for oral HPV; however, the population-level effect on the burden of oral HPV-16, HPV-18, HPV-6 and HPV-11 infections was only 17% overall, 25% in women and 7% in men, owing to low vaccination rates ¹⁶⁵.

The US Centers for Disease Control and Prevention (CDC) Advisory Committee on Immunization Practices recommends HPV vaccination for all children and adults 9–26 years of age ¹⁶⁶. More recently, the FDA approved HPV vaccination for adults 27–45 years of age who were not adequately vaccinated earlier ¹⁶⁷. Currently, two doses of the vaccine are recommended for children 15 years of age or younger, whereas three doses are recommended for those older than 15 years of age. Although widespread vaccination has the potential to eliminate HPV-positive HNSCC, the low vaccination rates and exponential increase in incidence observed among males in the USA indicate that herd immunity is at least a generation away³⁴.

Secondary prevention

Secondary prevention refers to the early detection of latent, asymptomatic disease and subsequent interventions to halt disease progression to a harmful state. In cancer, secondary prevention typically involves screening, such as mammography to find and treat early stage breast cancers or Papanicolaou smears to identify and ablate precancerous HPV lesions of the cervix. Although recognized oral premalignant lesions (OPLs), including leukoplakia, erythroplakia and dysplastic leukoplakia, are associated with increased risk for HPVnegative HNSCC, most OPLs themselves do not transform into invasive cancer^{168,169}. Rather, HPV-negative HNSCC arises from the grossly normal upper aerodigestive mucosa subject to 'epithelial field cancerization', where chronic exposure to carcinogens has resulted in molecular changes preconditioning towards malignant progression⁵⁰. As such, patients with a first HPV-negative HNSCC are at high risk for a SPT of the upper aerodigestive tract, including in the head and neck, oesophagus or lung¹⁵⁹. Due to the alarming rate of SPT formation, HNSCC has been a fertile field for the evaluation and eventual failure of multiple chemoprevention agents. For example, in the 1980s, high-dose isotretinoin, a synthetic vitamin A analogue, was found to reverse OPLs and prevent formation of SPTs in two landmark randomized trials; however, ultimately high-dose isotretinoin was too toxic for chronic daily use and has been abandoned 170,171. More recently, the EPOC trial investigated erlotinib, a small-molecule inhibitor of EGFR, in patients with OPLs who had loss of heterozygosity (LOH) at 9p and 3p. Although erlotinib was found to be ineffective, LOH was prospectively confirmed as a molecular biomarker of oral cancer risk¹⁷².

An effective and tolerable chemoprevention agent against HPV-negative HNSCC remains an elusive but noble ambition. Modern trials of chemoprevention of tobacco-related cancers

now focus on the cytoprotective, anti-inflammatory or immunomodulatory potential of repurposed pharmaceutical and nutraceutical compounds that are known to be well-tolerated during prolonged exposure. Ongoing secondary prevention trials include the evaluation of aspirin and other non-steroidal anti-inflammatory drugs to reduce cyclooxygenase 2 (COX2) signalling ¹⁷³; the study of metformin, an anti-hyperglycaemic biguanide that might block mTOR signalling in transforming epithelial cells ¹⁷⁴; and trials of 'green chemoprevention' agents derived from cruciferous vegetables rich in isothiocyanates, which are thought to induce carcinogen detoxication and, possibly, innate immunity ^{175,176}.

Screening

No validated tool exists for screening for HPV-positive HNSCC. HPV vaccinations are effective only for primary prevention, as their primary mechanism involves induction of anti-L1 capsid antibodies, which block the first infection step of viral entry. Furthermore, unlike cervical cancer, HPV-positive HNSCC lacks a cytological or gross precursor lesion. Although epidemiology studies consistently demonstrate an association between oral HPV DNA detection or anti-HPV-16 E6 seropositivity and risk for oropharyngeal cancer, the rarity of HPV-positive HNSCC reduces the clinical utility of such measures ¹⁴³.

Of note, a randomized, prospective screening trial in 20,174 asymptomatic males was conducted in Hong Kong and demonstrated that plasma EBV DNA detection had 97.1% sensitivity and 98.6% specificity for the identification of nasopharyngeal carcinoma⁴². Furthermore, nasopharyngeal carcinoma was detected at a significantly earlier stage in the clinical trial population than in a historical cohort, with 71% versus 20% stage I–II disease, respectively.

Management

The treatment approach to every individual patient is guided by anatomical subsite, stage, disease characteristics, functional considerations and patient wishes. Although natural history is different for HPV-related than for tobacco-related HNSCC, and intensive therapy is more difficult for elderly patients, these considerations do not lead to absolute differences in how patients should be managed.

Surgery, radiation and chemotherapy

The principal modalities of curative therapy for locally or locoregionally confined HNSCC are resection, radiation and systemic therapy. Treatment planning should aim for the most highly curative approach, while optimizing preservation of function. For patients with a small primary cancer with no clinical nodal involvement, or involvement of only a single node, cure rates of over 80% can be achieved with single modality intervention (resection or radiation)¹⁷⁷. Surgery is commonly elected for oral cavity cancers, whereas radiation might be more commonly employed for pharyngeal and laryngeal cancers. For laryngeal cancers, a moderately hypo-fractionated radiation schedule results in better locoregional control and survival than standard radiation therapy ^{178,179}. Advances in minimally invasive resection, including transoral robotic or laser resection and larynx-preserving partial laryngectomy, as well as improved reconstructive techniques, have extended the indications for primary

surgical management in the hands of high-volume head and neck surgical oncologists ^{180,181}. Occult metastases in the draining cervical lymph nodes might be present even with small, invasive primary tumours, and use of elective neck dissection improves survival ¹⁸². In the event of treatment failure after single modality radiation or surgery, salvage with the alternate modality offers a high chance of cure ^{181,183}. For tumours with more advanced tumour or nodal stage, post-operative radiation or chemoradiation, guided by pathological risk factors, reduces the risk of recurrence and improves survival ^{184,185}.

Pathological features indicative of increased risk of occurrence include extra-nodal extension, close or involved surgical margins, or perineural invasion; when these are present, administration of high-dose cisplatin chemotherapy concurrent with radiation further improves disease-free survival and impacts survival in the highest risk groups 184,185. The use of tri-modality therapy (that is, the addition of CRT following surgery) is known to increase the late toxicities of radiation, including chronic dysphagia and aspiration, and might increase the risk of non-cancer mortality in survivors ¹⁸⁶. For this reason, it is considered important to accurately predict the extent of disease, including the presence of extra-nodal extension, prior to initiation of definitive therapy, to avoid the need to add CRT after what was initially projected to be single modality surgical therapy, as currently happens in 65% of node-positive HPV-associated oropharyneal cancer that are managed primarily with resection ¹⁸⁷. Deep-learning neural networks, other radiomics approaches and functional imaging with PET offer promise in improving pre-operative diagnostic accuracy 188,189. Furthermore, there is a >50% likelihood of distant metastases in patients with HPV-positive oropharyngeal cancer and >5 nodes involved — such patients are also poor candidates for initial surgical management, given the potential positive impact of chemotherapy on micro-metastases¹⁷⁷.

Definitive treatment with CRT is recommended as non-surgical treatment for patients with advanced T stage (T3), more than one involved node, a single bulky node and for function preservation. The standard chemotherapy regimen is single agent cisplatin given at 100 mg/m² every 3 weeks¹⁹⁰, with a 3-year survival of 37% in a predominantly HPV-negative population; however, in patients who are poor candidates for high-dose cisplatin owing to pre-existing hearing loss or renal injury, weekly administration of 40-mg/m² cisplatin might lead to a comparable outcome ¹⁹¹. Although patients with T4 laryngeal cancer have previously been excluded from trials of larynx-preservation strategies because of the concern that even complete response would not lead to restoration of larynx function ¹⁸³, in patients who respond to a single cycle of induction chemotherapy and retain good laryngeal function, larynx preservation may be considered 192. The EGFR-directed antibody cetuximab is also an effective radiation sensitizer⁴; however, survival is reduced when cetuximab is substituted for cisplatin in HPV-positive oropharyngeal cancer^{5,6}. EBV-related nasopharyngeal cancer is treatment-responsive but associated with increased risk of distant metastases. Additional chemotherapy exposure after the completion of CRT, or as induction therapy, leads to an ~6% improvement in survival and should be considered the standard of care for locally advanced disease 193,194. Trials that attempt to tailor adjuvant therapy in patients with persistent circulating EBV DNA have to date not demonstrated the utility of this approach¹⁹⁵. Combined modality therapy is associated with substantial acute and late toxicities, including chronic dysphagia with a risk of aspiration pneumonia, and possibly

increased non-cancer mortality^{186,196}; therefore, expert multidisciplinary care, aggressive symptom management and swallowing rehabilitation are crucial to functional recovery.

The use of multidrug chemotherapy before definitive chemoradiation in tumours outside the nasopharynx has been studied extensively and does not improve overall survival compared with high-dose cisplatin and radiation ^{197,198}. However, objective response proportion is high ¹⁹⁹, and induction chemotherapy can be used to reduce tumour volume in selected situations, such as impending airway obstruction or when a patient is not able to lie flat for radiation simulation but is not a candidate for immediate surgery. Induction chemotherapy may also be used to control disease when curative surgery cannot immediately be accessed, as has been the case in areas with surging incidence of COVID-19²⁰⁰.

The recognition that patients with HPV-associated oropharyngeal cancer, small tumours and light smoking history enjoy exceptionally high survival rates 150,201 , but are treated with aggressive multimodality regimens that were evaluated predominantly in the more treatment-resistant HPV-negative population, raises the question of whether the toxicities and risks of such full-dose therapy are always warranted. A number of single-arm trials seem to demonstrate that for non-smokers with non-bulky tumours, response to induction chemotherapy predicts radiation sensitivity to the extent that radiation dose can safely be reduced from 70 Gy to 54 Gy 202,203 ; however, randomized trials will be required to confirm that this is an appropriate standard of care.

Immunotherapy

Among patients with recurrent or metastatic HNSCC, some may be cured by salvage resection, re-irradiation (particularly for nasopharyngeal cancer)²⁰⁴ or metastatectomy (particularly for HPV-positive cancer)²⁰⁵ (Fig. 7). The remaining patients are considered for systemic therapy. First-line treatment should include the immune checkpoint inhibitor pembrolizumab, an IgG4 humanized antibody against programmed cell death 1 (PD1), for patients with PDL1-expressing tumours or microsatellite instability, unless there is a contraindication to immunotherapy because of an underlying autoimmune disorder. A phase III trial compared pembrolizumab monotherapy or the combination of pembrolizumab with a platinating agent and 5-fluorouracil to the same chemotherapy with cetuximab⁹. Chemotherapy plus pembrolizumab improved overall survival compared with chemotherapy plus cetuximab (13 months versus 10.7 months, respectively; HR 0.77, p=-0.0034), with comparable response and toxicity results. Pembrolizumab alone was non-inferior to combination chemotherapy plus cetuximab among all patients with HNSCC (median overall survival of 11.6 months versus 10.7 months, respectively; HR 0.85). Among patients with expression of the biomarker PDL1, defined as a combined positive score (CPS) 20 or 1 (when CPS reflects all PDL1-expressing tumour cells, macrophages and immune cells as a proportion of the total number of tumour cells counted), survival was superior for pembrolizumab monotherapy versus chemotherapy plus cetuximab 14.9 versus 12.3 months, respectively). However, response proportion was higher for the chemotherapy combination; thus, many practitioners will favour pembrolizumab and a chemotherapy combination for patients with higher disease burden and those who are more symptomatic, while reserving pembrolizumab monotherapy for those with lower tumour volume and the highest PDL1

expression. A minority of patients with HNSCC who are treated with immune checkpoint inhibition may experience accelerated disease progression, often termed hyperprogression²⁰⁶. Hyperprogression is most likely for HPV-negative disease, bulky local or regional recurrence, and when immunotherapy is used without chemotherapy. Although hyperprogression is associated with worse outcome, objective and even durable responses to chemotherapy can be observed in this setting, and vigilance and rapid alteration in therapy are important to maximize disease control²⁰⁷. For patients who experience immune-related adverse events, such as pneumonitis, colitis or other organ injury, treatment interruption and systemic corticosteroids are indicated²⁰⁸.

Patients who are not candidates for first-line immunotherapy should receive cetuximab plus combination chemotherapy with a platinating agent and either 5-fluorouracil²⁰⁹ or paclitaxel. In the second-line, where the prognosis for survival is worse, PD1 inhibition may be reconsidered for some patients despite a history of autoimmune disease, as immune checkpoint inhibitors are also active in cisplatin-refractory patients. Two randomized trials compared PD1 inhibition with either nivolumab⁷ or pembrolizumab²¹⁰ to single-agent standard-of-care chemotherapy in patients who were cisplatin-refractory because they had progressed on cisplatin-based therapy for recurrent and/or metastatic disease, or because cancer had recurred within 6 months of concurrent cisplatin and radiation treatment. In each case, PD1 inhibition improved survival compared with chemotherapy; a small proportion of patients experience very durable benefit and some may benefit from continued therapy after progression^{211,212}. For patients with metastatic and/or recurrent HNSCC, many clinical trials are currently available that are exploring novel immunotherapies, as well as the very promising combinations of immune checkpoint inhibition with multi-targeted kinase inhibitors and other anti-angiogenic agents²¹³.

Throughout treatment, attention to symptom management, functional rehabilitation and appropriate incorporation of palliative care services are key to maintaining performance status and quality of life (QOL), and supporting patients at the end of life.

Quality of life

Patient health and supportive care

Given the complex nature of everyday functions within the head and neck area, the inherent consequences of HNSCC and its treatment and the increasing choices of treatments have a large effect on the health-related QOL (HRQOL) of patients with HNSCC.

The wide array and combinations of treatments all have their specific sequelae, including physical, emotional, functional, social, and occupational dysfunction, as well as a profound effect on the families of patients with HNSCC²¹⁴. Furthermore, HRQOL is significantly associated with survival^{215,216}. For example, a clinically meaningful association exists between HRQOL scores measured at diagnosis and overall survival of patients after treatment (HR = 0.90, CI 0.86–0.94, p<0.001)²¹⁷. Depending on the primary tumour site, patients with HNSCC might be confronted with specific symptoms, such as oral dysfunction and swallowing and speech problems, during treatment, which often improve 6 months after treatment²¹⁸. However, long-term reduction in QOL in HNSCC survivors (at 10-year follow

up) is common²¹⁹. On average, overall HRQOL deteriorated by 11% when compared with pre-treatment, and by 15% when compared with years 1 and 2 post-treatment²¹⁹.

To provide individualized (supportive) care, monitoring HRQOL in a structured manner in clinical research and practice is important. HRQOL is typically assessed using patient-reported outcome measures (PROMs)²²⁰. Widely used PROMs among patients with HNSCC include the European Organisation for Research and Treatment of Cancer (EORTC) Quality of Life Questionnaires (QLQ-C30 and QLQ-HN35 or QLQ-HN43) ²²¹ and the Functional Assessment of Cancer Therapy - Head & Neck Cancer (FACT-H&N)²²².

Survivorship

With the increase in incidence and improved survival rates, more people have to cope with living beyond a diagnosis of HNSCC and its treatment (see Box 1). From a more individualized perspective, this post-diagnosis survival is defined as "living with, through and beyond a cancer diagnosis" ^{223,224}. For HNSCC specifically, patients are confronted with numerous, profound disabilities owing to the anatomical complexity of the head and neck region, which can also affect patients' families²²⁵. Swallowing and speech impairments occur in ~50% of HNSCC survivors following radiotherapy treatment and are often present long-term²²⁶. For example, a majority (68%) of HNSCC survivors reported voice problems even 10 years after radiotherapy²²⁷. Furthermore, the two-year prevalence of dysphagia is 45% among HNSCC survivors (all therapies) and is 4–8 times more likely to occur in these individuals than in those who never had cancer²²⁸.

Moving forward

An integrated approach to survivorship care in HNSCC survivors, including (early) rehabilitation, psychosocial care, lifestyle interventions and self-management, will likely be implemented in the next 5–10 years²²⁹. In addition, peer support can be important to address the general and specific survivorship needs of individual patients²³⁰.

Substantial efforts are being made to improve HRQOL for patients with HNSCC by developing e-health applications, including self-monitoring of HRQOL and self-help interventions²³¹. These innovations carry the promise of sustainably improving supportive care and empowerment of HNSCC survivors²³². Measurement precision of HRQOL by PROMs can benefit from other data sources as biomarkers and from objective measures (collected using, for example, wearable devices, such as a smart watch, that record and analyse physical activity)²³³. Furthermore, emerging data-driven analytics can be used to study and improve forecasts of HRQOL²³³.

One of the current major challenges is to make collected HRQOL data universally available to patients and clinicians in a meaningful way that will facilitate the design of treatment regimens and post-therapy decisions and indicate appropriate supportive care for the individual patient.

Outlook

Advances in biotechnology, drug development, robotic surgery, radiotherapy approaches and molecular characterization of human cancers (including HNSCC) in the 21st century were expected to lead to improved outcomes for patients with HNSCC. However, despite these advances, outcomes have remained mostly unchanged for the past few decades, especially for HPV-negative HNSCC, and short-term and long-term treatment-associated morbidities are still substantial. Most patients still present with advanced-stage disease and are treated with platinum-based chemotherapy regimens that were approved by the FDA in 1978. Patients who survive their first HNSCC (especially HPV-negative HNSCC) remain at risk for development of a SPT of the upper aerodigestive tract for the rest of their lives. The requirement for multidisciplinary care (including, amongst others, head and neck surgery, radiation oncology, medical oncology, head and neck pathology, speech language pathology, nutrition, prosthodontics and oral medicine) coupled with the rarity of HNSCC in most countries, underscores our recommendation that all patients with HNSCC should be treated in high-volume centres and by experienced multi-disciplinary teams. An analysis conducted in the USA employing the National Cancer Database showed that patients undergoing curative radiotherapy at high-volume facilities and academic centres showed increased survival compared with patients treated at low-volume centres²³⁴. Even when patients with HNSCC are treated in a multi-institutional chemoradiation clinical trial, the overall survival of those patients treated at centres that enroll more patients in trials in the USA was significantly greater than that of patients treated at institutions that enroll fewer patients²³⁵. In a pooled analysis of six randomized, controlled trials in Italy, survival was improved when patients with HNSCC were treated at high-case-volume centres compared with lowcase-volume centers²³⁶. Despite improvements in treatment options, particularly the incorporation of immune checkpoint inhibitors into the standard of care, we also recommend that all patients with advanced, incurable HNSCC be offered the opportunity to participate in clinical trials that are based on a strong biological rationale. Table 1 lists trials of HNSCC treatments in the National Clinical Trials Network (NCTN) that are currently open for patient enrolment. Fortunately, numerous clinical trials are ongoing and precision medicine approaches are emerging.

HPV-positive HNSCC

The awareness that HPV contributes to the majority of oropharyngeal HNSCC cases has markedly affected clinical practice. HPV vaccination has been shown to prevent infection with HNSCC-causing HPV types, including HPV-16 and HPV-18, which are responsible for the majority of HPV-positive HNSCC. In the USA, HPV vaccination is generally administered at the discretion of the paediatrician and the parents, with only Virginia, Rhode Island and the District of Columbia requiring the HPV vaccine for public school attendance. The World Health Organization recommends HPV vaccination as a part of routine vaccinations in all countries. Barriers to widespread vaccination of all girls and boys prior to HPV infection include cost, general vaccine skepticism and concerns that administration of a vaccine against a sexually transmitted virus will encourage sexual activity. Stated simply, HPV vaccination of all children and young adults prior to HPV infection is likely to dramatically reduce, if not eliminate completely, HPV-positive HNSCC. Whereas the

incidence of smoking-related HNSCC continues to decline worldwide, that of HPV-positive HNSCC is on the rise²³⁷. The number of men with HPV-positive HNSCC has now surpassed the number of women with HPV-associated cervical cancer in the USA²³⁸, suggesting that without widespread HPV vaccination, HPV-positive HNSCC could reach epidemic proportions. We encourage clinicians who treat patients with HNSCC to partner with paediatricians, public health officials and politicians to increase HPV vaccination rates worldwide.

As the pathogenesis of HPV-positive HNSCC seems to be distinct from that of HPVnegative HNSCC, a tantalizing possibility is that HPV-positive tumours have unique vulnerabilities that could be exploited for therapeutic purposes. Non-smokers who develop HPV-positive HNSCC generally survive their head and neck cancer, and clinical strategies to date have focused on identifying treatments that are associated with reduced morbidity. The known toxicities of the EGFR inhibitor cetuximab are notably less than those of platinum chemotherapy. In an effort to reduce treatment morbidity and maintain the high survival rate of HPV-positive HNSCC, phase III clinical trials have compared cetuximab plus radiation to cisplatin plus radiation. These studies demonstrated that cetuximab is inferior to cisplatin in HPV-positive disease, thus precluding this FDA-approved molecular targeting agent as a treatment option for HPV-positive HNSCC^{5,6}. Studies leading to the incorporation of immune checkpoint inhibitors, including pembrolizumab and nivolumab, into standard of care for HNSCC have included both HPV-positive and HPV-negative patients. FDAapproval of these agents for HNSCC treatment confirmed the power of harnessing the immune system for HNSCC therapy. However, given the improved outcome for most patients with HPV-positive HNSCC, they are less likely to receive these immune checkpoint inhibitors as part of standard of care compared with individuals with HPV-negative HNSCC. Ongoing and future research aimed at elucidating the immune profile of HPV-positive HNSCC is expected to help define therapeutic targets in this population. Mitigating the toxicities associated with standard of care, including platinum chemotherapy and radiation, for these patients remains an important objective.

Molecular characterization of HNSCC

Substantial progress has been made over the past decade in the molecular characterization of both HPV-positive and HPV-negative HNSCC, culminating in The Cancer Genome Atlas (TCGA)^{59,71,72}. These collective efforts have confirmed the preponderance of *TP53* mutations, identified previously unknown *NOTCH1* mutations and provided a platform to test precision medicine approaches in HNSCC. Success stories include the recognition that *HRAS* mutations may serve as predictive biomarkers for treatment with farnesyl transferase inhibitors, such as tipifarnib²³⁹, and evidence that oncogenic *PIK3CA* mutations are common and could be used to identify patients with HNSCC who are most likely to benefit from PI3K pathway inhibitors^{69,240}. Although some centres are routinely evaluating the mutational profile of HNSCC tumours, the tumours in the majority of patients with HNSCC remain uncharacterized, even when these patients present with recurrent or metastatic disease. Furthermore, the role of tumour mutational burden as a predictive biomarker for immunotherapy remains incompletely understood and efforts are underway to establish guidelines that are designed to harmonize tumour mutation burden evaluation and

reporting²⁴¹. Prospective, targeted sequencing, supported by efforts such as the American Association for Cancer Research Project Genie, aim to provide a publicly accessible international cancer registry of real-world data assembled through data sharing²⁴². Importantly, single-cell transcriptomic analysis represents a powerful new tool for defining diverse intratumoral expression programmes and differences among tumours from different patients, as has been demonstrated in HNSCC^{137,243}. In addition, increased use of relevant preclinical models, including patient-derived xenografts^{244,245} and organoids²⁴⁶, should facilitate the translation of laboratory discoveries to patients with HNSCC.

Immunotherapy

The importance of the immune system in the development and treatment of HNSCC has long been recognized, leading to early efforts to treat this malignancy using strategies to activate endogenous immunity. The FDA approval of immune checkpoint inhibitors in 2016 for recurrent or metastatic disease, and for front-line treatment of inoperable or metastatic cancer in 2019, definitively altered the landscape of HNSCC therapy and clinical trial opportunities. For patients who benefit from these agents, responses seem to be durable 7–9. However, to date, only a subset of patients with HNSCC are expected to respond to immune checkpoint inhibitors, and reliable predictive biomarkers are needed. Evidence exists that PDL1 expression levels in the tumour may indicate a likelihood of clinical benefit, which led to the 2019 approval of pembrolizumab as first-line therapy in patients with HNSCC whose tumours had a CPS 1)⁹.

In addition to FDA-approved and emerging agents that target PD1 or PDL1, the anti-CTLA4 monoclonal antibody ipilimumab is also in active clinical development (>20 trials are in progress) for HNSCC treatment. In fact, the non-redundant functions of PD1 and CTLA4 in regulating the adaptive immune system serve as the basis for combining PD1 and CTLA4 inhibitors for cancer therapy, including HNSCC, in ongoing clinical trials. Whereas PD1 and PDL1 inhibitors are generally well tolerated, CTLA4 inhibitors are associated with more toxicity²⁴⁷, and it will be important to balance efficacy and toxicity when combining these agents. In addition, although adaptive immune cell therapies (eg. CAR T cells) and therapeutic vaccines are not FDA approved, they are under active investigation in head and neck cancer and represent a promising new avenue for therapy.

The era of immune-oncology has highlighted the limitations of traditional preclinical HNSCC models, including cell lines and xenografts derived from cell lines and patient tumours. Even though patient-derived xenografts are a more accurate model of the patient tumour than cell lines, the xenograft stroma largely consists of mouse cells and these models are generally grown in immunocompromised mice. The advent of immunocompetent HNSCC models offers an opportunity to evaluate immunotherapies in more realistic animal models of HNSCC prior to initiating clinical testing. For example, a report of a syngeneic carcinogen-induced HNSCC tumour in immunocompetent mice demonstrates the ability of these models to assess responses to anti-CTLA4 antibodies²⁴⁸. Given the limitations of preclinical models to study immune perturbations, the development of novel clinical trial platforms in conjunction with immune profiling approaches offer opportunities to test the effect of novel immunotherapies in the clinical setting with minimal risk²⁴⁹.

Acknowledgements

This work was supported by the following: NIH R35CA231998 (J.R.G.), U54CA209891 (J.R.G.), R01DE023685 (J.R.G. and D.E.J.), R01DE028289 (D.E.J. and J.R.G.), NET-QUBIC-Dutch Cancer Society grant VU 2013-5930 (C.R.L.), General Research Fund #17121616 and #14168517 (V.W.Y.L.), Research Impact Fund R4015-19F and R4017-18 (V.W.Y.L.), the Health and Medical Research Fund by the Food and Health Bureau, the Government of the Hong Kong Special Administrative Region #15160691 (V.W.Y.L.), University-Industry Collaboration Program UIM/329 by the Innovation and Technology Fund, Hong Kong government, Hong Kong SAR (V.W.Y.L.), the Hong Kong Cancer Fund, Hong Kong SAR (V.W.Y.L.), NIH/NCI UG1CA242596 (J.E.B.), and NIH/NCI P30CA023074 (J.E.B.).

References

- 1. Stein AP et al. Prevalence of Human Papillomavirus in Oropharyngeal Cancer: A Systematic Review. Cancer J 21, 138–146, doi:10.1097/PPO.0000000000000115 (2015). [PubMed: 26049691]
- Isayeva T, Li Y, Maswahu D & Brandwein-Gensler M Human papillomavirus in non-oropharyngeal head and neck cancers: a systematic literature review. Head Neck Pathol 6 Suppl 1, S104–120, doi:10.1007/s12105-012-0368-1 (2012). [PubMed: 22782230]
- Michaud DS et al. High-risk HPV types and head and neck cancer. Int J Cancer 135, 1653–1661, doi:10.1002/ijc.28811 (2014). [PubMed: 24615247]
- Bonner JA et al. Radiotherapy plus cetuximab for squamous-cell carcinoma of the head and neck. N Engl J Med 354, 567–578 (2006). [PubMed: 16467544]
- Gillison ML et al. Radiotherapy plus cetuximab or cisplatin in human papillomavirus-positive oropharyngeal cancer (NRG Oncology RTOG 1016): a randomised, multicentre, non-inferiority trial. Lancet 393, 40–50, doi:10.1016/S0140-6736(18)32779-X (2019). [PubMed: 30449625]
- 6. Mehanna H et al. Radiotherapy plus cisplatin or cetuximab in low-risk human papillomavirus-positive oropharyngeal cancer (De-ESCALaTE HPV): an open-label randomised controlled phase 3 trial. Lancet 393, 51–60, doi:10.1016/S0140-6736(18)32752-1 (2019). [PubMed: 30449623]
- 7. Ferris RL et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. N Engl J Med 375, 1856–1867, doi:10.1056/NEJMoa1602252 (2016). [PubMed: 27718784]
- Seiwert TY et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-012): an open-label, multicentre, phase 1b trial. Lancet Oncol 17, 956–965, doi:10.1016/S1470-2045(16)30066-3 (2016). [PubMed: 27247226]
- Burtness B et al. Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-048): a randomised, open-label, phase 3 study. Lancet 394, 1915–1928, doi:10.1016/ S0140-6736(19)32591-7 (2019). [PubMed: 31679945]
- Ferlay J et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. Int J Cancer 144, 1941–1953, doi:10.1002/ijc.31937 (2019). [PubMed: 30350310]
- Bray F et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 68, 394–424, doi:10.3322/ caac.21492 (2018). [PubMed: 30207593]
- 12. Ferlay J, Ervik M, Lam F, Colombet M, Mery L, Pineros M, Znaor A, Soerjomataram I, Bray F Global Cancer Observatory: Cancer Today. Lyon, France: International Agency for Research on Cancer, https://gco.iarc.fr/today (2018).
- 13. Hashibe M et al. Alcohol drinking in never users of tobacco, cigarette smoking in never drinkers, and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. J Natl Cancer Inst 99, 777–789, doi:99/10/777 [pii] 10.1093/jnci/djk179 (2007). [PubMed: 17505073]
- 14. Mehanna H et al. Prevalence of human papillomavirus in oropharyngeal and nonoropharyngeal head and neck cancer--systematic review and meta-analysis of trends by time and region. Head Neck 35, 747–755, doi:10.1002/hed.22015 (2013). [PubMed: 22267298]

15. Jiang H et al. Can public health policies on alcohol and tobacco reduce a cancer epidemic? Australia's experience. BMC Med 17, 213, doi:10.1186/s12916-019-1453-z (2019). [PubMed: 31771596]

- Windon MJ et al. Increasing prevalence of human papillomavirus-positive oropharyngeal cancers among older adults. Cancer 124, 2993–2999, doi:10.1002/cncr.31385 (2018). [PubMed: 29710393]
- Fung SY, Lam JW & Chan KC Clinical utility of circulating Epstein-Barr virus DNA analysis for the management of nasopharyngeal carcinoma. Chin Clin Oncol 5, 18, doi:10.21037/ cco.2016.03.07 (2016). [PubMed: 27121878]
- Pulte D & Brenner H Changes in survival in head and neck cancers in the late 20th and early 21st century: a period analysis. Oncologist 15, 994–1001, doi:10.1634/theoncologist.2009-0289 (2010). [PubMed: 20798198]
- Chaturvedi AK et al. Human papillomavirus and rising oropharyngeal cancer incidence in the United States. J Clin Oncol 29, 4294–4301, doi:10.1200/JCO.2011.36.4596 (2011). [PubMed: 21969503]
- 20. Osazuwa-Peters N et al. Suicide risk among cancer survivors: Head and neck versus other cancers. Cancer 124, 4072–4079, doi:10.1002/cncr.31675 (2018). [PubMed: 30335190]
- Cancer, I. A. f. R. o. List of Classifications by cancer sites with sufficient or limited evidence in humans, Volumes 1 to 127. IARC Monographs On The Identification Of Carcinogenic Hazards To Humans, https://monographs.iarc.fr/wp-content/uploads/2019/07/ Classification_by_cancer_site_127.pdf> (
- 22. Blot WJ et al. Smoking and drinking in relation to oral and pharyngeal cancer. Cancer Res 48, 3282–3287 (1988). [PubMed: 3365707]
- Zhang LW et al. Incidence and mortality trends in oral and oropharyngeal cancers in China, 2005–2013. Cancer Epidemiol 57, 120–126, doi:10.1016/j.canep.2018.10.014 (2018). [PubMed: 30396144]
- 24. Organization, W. H. WHO global report on trends in prevalence of tobacco smoking 2015. Geneva. WHO Press (2015).
- Wong IC, Ng YK & Lui VW Cancers of the lung, head and neck on the rise: perspectives on the genotoxicity of air pollution. Chin J Cancer 33, 476–480, doi:10.5732/cjc.014.10093 (2014).
 [PubMed: 25011457]
- 26. Mishra A & Meherotra R Head and neck cancer: global burden and regional trends in India. Asian Pac J Cancer Prev 15, 537–550, doi:10.7314/apjcp.2014.15.2.537 (2014). [PubMed: 24568456]
- 27. Guha N et al. Oral health and risk of squamous cell carcinoma of the head and neck and esophagus: results of two multicentric case-control studies. Am J Epidemiol 166, 1159–1173, doi:10.1093/aje/kwm193 (2007). [PubMed: 17761691]
- 28. Freedman ND et al. Fruit and vegetable intake and head and neck cancer risk in a large United States prospective cohort study. Int J Cancer 122, 2330–2336, doi:10.1002/ijc.23319 (2008). [PubMed: 18092323]
- 29. Hennessey PT, Westra WH & Califano JA Human papillomavirus and head and neck squamous cell carcinoma: recent evidence and clinical implications. J Dent Res 88, 300–306, doi:10.1177/0022034509333371 (2009). [PubMed: 19407148]
- 30. Tsang CM, Lui VWY, Bruce JP, Pugh TJ & Lo KW Translational genomics of nasopharyngeal cancer. Semin Cancer Biol 61, 84–100, doi:10.1016/j.semcancer.2019.09.006 (2020). [PubMed: 31521748]
- 31. Viens LJ et al. Human Papillomavirus-Associated Cancers United States, 2008–2012. MMWR Morb Mortal Wkly Rep 65, 661–666, doi:10.15585/mmwr.mm6526a1 (2016). [PubMed: 27387669]
- 32. Gillison ML et al. Prevalence of oral HPV infection in the United States, 2009–2010. JAMA 307, 693–703, doi:10.1001/jama.2012.101 (2012). [PubMed: 22282321]
- 33. Chaturvedi AK et al. NHANES 2009–2012 Findings: Association of Sexual Behaviors with Higher Prevalence of Oral Oncogenic Human Papillomavirus Infections in U.S. Men. Cancer Res 75, 2468–2477, doi:10.1158/0008-5472.CAN-14-2843 (2015). [PubMed: 25873485]

34. Gillison ML, Chaturvedi AK, Anderson WF & Fakhry C Epidemiology of Human Papillomavirus-Positive Head and Neck Squamous Cell Carcinoma. J Clin Oncol 33, 3235–3242, doi:10.1200/JCO.2015.61.6995 (2015). [PubMed: 26351338]

- 35. Gillison ML, Chaturvedi AK & Lowy DR HPV prophylactic vaccines and the potential prevention of noncervical cancers in both men and women. Cancer 113, 3036–3046, doi:10.1002/cncr.23764 (2008). [PubMed: 18980286]
- 36. Rowhani-Rahbar A et al. Antibody responses in oral fluid after administration of prophylactic human papillomavirus vaccines. J Infect Dis 200, 1452–1455, doi:10.1086/606026 (2009). [PubMed: 19698077]
- 37. Velleuer E & Dietrich R Fanconi anemia: young patients at high risk for squamous cell carcinoma. Mol Cell Pediatr 1, 9, doi:10.1186/s40348-014-0009-8 (2014). [PubMed: 26567103]
- 38. Fang M, Huang W, Mo D, Zhao W & Huang R Association of Five Snps in Cytotoxic T-Lymphocyte Antigen 4 and Cancer Susceptibility: Evidence from 67 Studies. Cell Physiol Biochem 47, 414–427, doi:10.1159/000489953 (2018). [PubMed: 29794444]
- 39. Niu YM et al. Increased risks between Interleukin-10 gene polymorphisms and haplotype and head and neck cancer: a meta-analysis. Sci Rep 5, 17149, doi:10.1038/srep17149 (2015). [PubMed: 26612133]
- 40. Wang Y, Yang H, Duan G & Wang H The association of the CYP1A1 Ile462Val polymorphism with head and neck cancer risk: evidence based on a cumulative meta-analysis. Onco Targets Ther 9, 2927–2934, doi:10.2147/OTT.S106264 (2016). [PubMed: 27274286]
- 41. Cadoni G et al. A review of genetic epidemiology of head and neck cancer related to polymorphisms in metabolic genes, cell cycle control and alcohol metabolism. Acta Otorhinolaryngol Ital 32, 1–11 (2012). [PubMed: 22500060]
- 42. Chan KCA et al. Analysis of Plasma Epstein-Barr Virus DNA to Screen for Nasopharyngeal Cancer. N Engl J Med 377, 513–522, doi:10.1056/NEJMoa1701717 (2017). [PubMed: 28792880]
- 43. Krishnamurthy S et al. Endothelial cell-initiated signaling promotes the survival and self-renewal of cancer stem cells. Cancer Res 70, 9969–9978, doi:10.1158/0008-5472.CAN-10-1712 (2010). [PubMed: 21098716]
- 44. Faber A et al. CD44 as a stem cell marker in head and neck squamous cell carcinoma. Oncol Rep 26, 321–326, doi:10.3892/or.2011.1322 (2011). [PubMed: 21617876]
- 45. Yu SS & Cirillo N The molecular markers of cancer stem cells in head and neck tumors. J Cell Physiol 235, 65–73, doi:10.1002/jcp.28963 (2020). [PubMed: 31206697]
- 46. Zhang Q et al. A subpopulation of CD133(+) cancer stem-like cells characterized in human oral squamous cell carcinoma confer resistance to chemotherapy. Cancer Lett 289, 151–160, doi:10.1016/j.canlet.2009.08.010 (2010). [PubMed: 19748175]
- 47. Chiou SH et al. Positive correlations of Oct-4 and Nanog in oral cancer stem-like cells and high-grade oral squamous cell carcinoma. Clin Cancer Res 14, 4085–4095, doi:10.1158/1078-0432.CCR-07-4404 (2008). [PubMed: 18593985]
- 48. Leon X et al. Second, third, and fourth head and neck tumors. A progressive decrease in survival. Head Neck 34, 1716–1719, doi:10.1002/hed.21977 (2012). [PubMed: 22307753]
- 49. Leon X et al. Risk of onset of second neoplasms and successive neoplasms in patients with a head and neck index tumour. Acta Otorrinolaringol Esp 71, 9–15, doi:10.1016/j.otorri.2018.11.003 (2020). [PubMed: 30987756]
- 50. Slaughter DP, Southwick HW & Smejkal W Field cancerization in oral stratified squamous epithelium; clinical implications of multicentric origin. Cancer 6, 963–968 (1953). [PubMed: 13094644]
- Ryser MD, Lee WT, Ready NE, Leder KZ & Foo J Quantifying the Dynamics of Field Cancerization in Tobacco-Related Head and Neck Cancer: A Multiscale Modeling Approach. Cancer Res 76, 7078–7088, doi:10.1158/0008-5472.CAN-16-1054 (2016). [PubMed: 27913438]
- 52. Sheth SH, Johnson DE, Kensler TW & Bauman JE Chemoprevention targets for tobacco-related head and neck cancer: past lessons and future directions. Oral Oncol 51, 557–564, doi:10.1016/j.oraloncology.2015.02.101 (2015). [PubMed: 25868717]
- 53. Hecht SS Tobacco smoke carcinogens and lung cancer. J Natl Cancer Inst 91, 1194–1210, doi:10.1093/jnci/91.14.1194 (1999). [PubMed: 10413421]

54. Hoffmann D & Hoffmann I The changing cigarette, 1950–1995. J Toxicol Environ Health 50, 307–364, doi:10.1080/009841097160393 (1997). [PubMed: 9120872]

- 55. Warnakulasuriya S & Straif K Carcinogenicity of smokeless tobacco: Evidence from studies in humans & experimental animals. Indian J Med Res 148, 681–686, doi:10.4103/ijmr.IJMR_149_18 (2018). [PubMed: 30778001]
- 56. Talamini R et al. Combined effect of tobacco and alcohol on laryngeal cancer risk: a case-control study. Cancer Causes Control 13, 957–964, doi:10.1023/a:1021944123914 (2002). [PubMed: 12588092]
- 57. Pai SI & Westra WH Molecular pathology of head and neck cancer: implications for diagnosis, prognosis, and treatment. Annu Rev Pathol 4, 49–70, doi:10.1146/annurev.pathol.4.110807.092158 (2009). [PubMed: 18729723]
- 58. Brooks PJ & Theruvathu JA DNA adducts from acetaldehyde: implications for alcohol-related carcinogenesis. Alcohol 35, 187–193, doi:10.1016/j.alcohol.2005.03.009 (2005). [PubMed: 16054980]
- 59. Cancer Genome Atlas N Comprehensive genomic characterization of head and neck squamous cell carcinomas. Nature 517, 576–582, doi:10.1038/nature14129 (2015). [PubMed: 25631445]
- 60. Scheffner M, Huibregtse JM, Vierstra RD & Howley PM The HPV-16 E6 and E6-AP complex functions as a ubiquitin-protein ligase in the ubiquitination of p53. Cell 75, 495–505 (1993). [PubMed: 8221889]
- 61. White EA et al. Comprehensive analysis of host cellular interactions with human papillomavirus E6 proteins identifies new E6 binding partners and reflects viral diversity. J Virol 86, 13174–13186, doi:10.1128/JVI.02172-12 (2012). [PubMed: 23015706]
- 62. Tomaic V Functional Roles of E6 and E7 Oncoproteins in HPV-Induced Malignancies at Diverse Anatomical Sites. Cancers (Basel) 8, doi:10.3390/cancers8100095 (2016).
- 63. Eckhardt M et al. Multiple Routes to Oncogenesis Are Promoted by the Human Papillomavirus-Host Protein Network. Cancer Discov 8, 1474–1489, doi:10.1158/2159-8290.CD-17-1018 (2018). [PubMed: 30209081]
- 64. Dyson N, Howley PM, Munger K & Harlow E The human papilloma virus-16 E7 oncoprotein is able to bind to the retinoblastoma gene product. Science 243, 934–937, doi:10.1126/science.2537532 (1989). [PubMed: 2537532]
- 65. White EA et al. Systematic identification of interactions between host cell proteins and E7 oncoproteins from diverse human papillomaviruses. Proc Natl Acad Sci U S A 109, E260–267, doi:10.1073/pnas.1116776109 (2012). [PubMed: 22232672]
- 66. Venuti A et al. Papillomavirus E5: the smallest oncoprotein with many functions. Mol Cancer 10, 140, doi:10.1186/1476-4598-10-140 (2011). [PubMed: 22078316]
- 67. Estevao D, Costa NR, Gil da Costa RM & Medeiros R Hallmarks of HPV carcinogenesis: The role of E6, E7 and E5 oncoproteins in cellular malignancy. Biochim Biophys Acta Gene Regul Mech 1862, 153–162, doi:10.1016/j.bbagrm.2019.01.001 (2019). [PubMed: 30707946]
- 68. Califano J et al. Genetic progression model for head and neck cancer: implications for field cancerization. Cancer Res 56, 2488–2492 (1996). [PubMed: 8653682]
- 69. Lui VW et al. Frequent mutation of the PI3K pathway in head and neck cancer defines predictive biomarkers. Cancer Discov 3, 761–769, doi:10.1158/2159-8290.CD-13-0103 (2013). [PubMed: 23619167]
- 70. Nichols AC et al. High frequency of activating PIK3CA mutations in human papillomavirus-positive oropharyngeal cancer. JAMA Otolaryngol Head Neck Surg 139, 617–622, doi:10.1001/jamaoto.2013.3210 (2013). [PubMed: 23787421]
- Agrawal N et al. Exome sequencing of head and neck squamous cell carcinoma reveals inactivating mutations in NOTCH1. Science 333, 1154–1157, doi:10.1126/science.1206923 (2011). [PubMed: 21798897]
- 72. Stransky N et al. The mutational landscape of head and neck squamous cell carcinoma. Science 333, 1157–1160, doi:10.1126/science.1208130 (2011). [PubMed: 21798893]
- 73. Alsahafi E et al. Clinical update on head and neck cancer: molecular biology and ongoing challenges. Cell Death Dis 10, 540, doi:10.1038/s41419-019-1769-9 (2019). [PubMed: 31308358]

74. Rocco JW & Ellisen LW p63 and p73: life and death in squamous cell carcinoma. Cell Cycle 5, 936–940, doi:10.4161/cc.5.9.2716 (2006). [PubMed: 16687923]

- 75. Rocco JW, Leong CO, Kuperwasser N, DeYoung MP & Ellisen LW p63 mediates survival in squamous cell carcinoma by suppression of p73-dependent apoptosis. Cancer Cell 9, 45–56 (2006). [PubMed: 16413471]
- 76. Yang X et al. DeltaNp63 versatilely regulates a Broad NF-kappaB gene program and promotes squamous epithelial proliferation, migration, and inflammation. Cancer Res 71, 3688–3700, doi:10.1158/0008-5472.CAN-10-3445 (2011). [PubMed: 21576089]
- 77. Lu H et al. TNF-alpha promotes c-REL/DeltaNp63alpha interaction and TAp73 dissociation from key genes that mediate growth arrest and apoptosis in head and neck cancer. Cancer Res 71, 6867–6877, doi:10.1158/0008-5472.CAN-11-2460 (2011). [PubMed: 21933882]
- 78. Si H et al. TNF-alpha modulates genome-wide redistribution of DeltaNp63alpha/TAp73 and NF-kappaB cREL interactive binding on TP53 and AP-1 motifs to promote an oncogenic gene program in squamous cancer. Oncogene 35, 5781–5794, doi:10.1038/onc.2016.112 (2016). [PubMed: 27132513]
- 79. Lu H et al. CK2 phosphorylates and inhibits TAp73 tumor suppressor function to promote expression of cancer stem cell genes and phenotype in head and neck cancer. Neoplasia 16, 789–800, doi:10.1016/j.neo.2014.08.014 (2014). [PubMed: 25379016]
- 80. Hu Z et al. Polo-like kinase 2 acting as a promoter in human tumor cells with an abundance of TAp73. Onco Targets Ther 8, 3475–3488, doi:10.2147/OTT.S90302 (2015). [PubMed: 26640387]
- 81. Foy JP et al. New DNA methylation markers and global DNA hypomethylation are associated with oral cancer development. Cancer Prev Res (Phila) 8, 1027–1035, doi:10.1158/1940-6207.CAPR-14-0179 (2015). [PubMed: 26342026]
- 82. Viswanathan M, Tsuchida N & Shanmugam G Promoter hypermethylation profile of tumorassociated genes p16, p15, hMLH1, MGMT and E-cadherin in oral squamous cell carcinoma. Int J Cancer 105, 41–46, doi:10.1002/ijc.11028 (2003). [PubMed: 12672028]
- 83. Ha PK & Califano JA Promoter methylation and inactivation of tumour-suppressor genes in oral squamous-cell carcinoma. Lancet Oncol 7, 77–82, doi:10.1016/S1470-2045(05)70540-4 (2006). [PubMed: 16389187]
- 84. Rubin Grandis J et al. Levels of TGF-alpha and EGFR protein in head and neck squamous cell carcinoma and patient survival. J Natl Cancer Inst 90, 824–832 (1998). [PubMed: 9625170]
- 85. Zhu X et al. Prognostic role of epidermal growth factor receptor in head and neck cancer: a meta-analysis. J Surg Oncol 108, 387–397, doi:10.1002/jso.23406 (2013). [PubMed: 24038070]
- 86. Madoz-Gurpide J et al. Activation of MET pathway predicts poor outcome to cetuximab in patients with recurrent or metastatic head and neck cancer. J Transl Med 13, 282, doi:10.1186/s12967-015-0633-7 (2015). [PubMed: 26319934]
- 87. Arnold L, Enders J & Thomas SM Activated HGF-c-Met Axis in Head and Neck Cancer. Cancers (Basel) 9, doi:10.3390/cancers9120169 (2017).
- 88. Karakasheva TA et al. IL-6 Mediates Cross-Talk between Tumor Cells and Activated Fibroblasts in the Tumor Microenvironment. Cancer Res 78, 4957–4970, doi:10.1158/0008-5472.CAN-17-2268 (2018). [PubMed: 29976575]
- 89. Tsai MS, Chen WC, Lu CH & Chen MF The prognosis of head and neck squamous cell carcinoma related to immunosuppressive tumor microenvironment regulated by IL-6 signaling. Oral Oncol 91, 47–55, doi:10.1016/j.oraloncology.2019.02.027 (2019). [PubMed: 30926062]
- 90. Marsit CJ, Black CC, Posner MR & Kelsey KT A genotype-phenotype examination of cyclin D1 on risk and outcome of squamous cell carcinoma of the head and neck. Clin Cancer Res 14, 2371–2377, doi:10.1158/1078-0432.CCR-07-4368 (2008). [PubMed: 18413827]
- 91. Wang Z, Valera JC, Zhao X, Chen Q & Gutkind JS mTOR co-targeting strategies for head and neck cancer therapy. Cancer Metastasis Rev 36, 491–502, doi:10.1007/s10555-017-9688-7 (2017). [PubMed: 28822012]
- 92. Squarize CH et al. PTEN deficiency contributes to the development and progression of head and neck cancer. Neoplasia 15, 461–471, doi:10.1593/neo.121024 (2013). [PubMed: 23633918]

93. Geiger JL, Grandis JR & Bauman JE The STAT3 pathway as a therapeutic target in head and neck cancer: Barriers and innovations. Oral Oncol 56, 84–92, doi:10.1016/j.oraloncology.2015.11.022 (2016). [PubMed: 26733183]

- 94. Johnson DE, O'Keefe RA & Grandis JR Targeting the IL-6/JAK/STAT3 signalling axis in cancer. Nat Rev Clin Oncol 15, 234–248, doi:10.1038/nrclinonc.2018.8 (2018). [PubMed: 29405201]
- 95. Lui VW et al. Frequent mutation of receptor protein tyrosine phosphatases provides a mechanism for STAT3 hyperactivation in head and neck cancer. Proc Natl Acad Sci U S A 111, 1114–1119, doi:10.1073/pnas.1319551111 (2014). [PubMed: 24395800]
- 96. Peyser ND et al. Loss-of-Function PTPRD Mutations Lead to Increased STAT3 Activation and Sensitivity to STAT3 Inhibition in Head and Neck Cancer. PLoS One 10, e0135750, doi:10.1371/journal.pone.0135750 (2015). [PubMed: 26267899]
- 97. Alamoud KA & Kukuruzinska MA Emerging Insights into Wnt/beta-catenin Signaling in Head and Neck Cancer. J Dent Res 97, 665–673, doi:10.1177/0022034518771923 (2018). [PubMed: 29771197]
- 98. Augsten M Cancer-associated fibroblasts as another polarized cell type of the tumor microenvironment. Front Oncol 4, 62, doi:10.3389/fonc.2014.00062 (2014). [PubMed: 24734219]
- 99. Canning M et al. Heterogeneity of the Head and Neck Squamous Cell Carcinoma Immune Landscape and Its Impact on Immunotherapy. Front Cell Dev Biol 7, 52, doi:10.3389/ fcell.2019.00052 (2019). [PubMed: 31024913]
- 100. Peltanova B, Raudenska M & Masarik M Effect of tumor microenvironment on pathogenesis of the head and neck squamous cell carcinoma: a systematic review. Mol Cancer 18, 63, doi:10.1186/s12943-019-0983-5 (2019). [PubMed: 30927923]
- 101. Marsh D et al. Stromal features are predictive of disease mortality in oral cancer patients. J Pathol 223, 470–481, doi:10.1002/path.2830 (2011). [PubMed: 21294121]
- 102. Amit M et al. Loss of p53 drives neuron reprogramming in head and neck cancer. Nature 578, 449–454, doi:10.1038/s41586-020-1996-3 (2020). [PubMed: 32051587]
- 103. Partlova S et al. Distinct patterns of intratumoral immune cell infiltrates in patients with HPV-associated compared to non-virally induced head and neck squamous cell carcinoma. Oncoimmunology 4, e965570, doi:10.4161/21624011.2014.965570 (2015). [PubMed: 25949860]
- 104. Mandal R et al. The head and neck cancer immune landscape and its immunotherapeutic implications. JCI Insight 1, e89829, doi:10.1172/jci.insight.89829 (2016). [PubMed: 27777979]
- 105. Chen YP et al. Identification and validation of novel microenvironment-based immune molecular subgroups of head and neck squamous cell carcinoma: implications for immunotherapy. Ann Oncol 30, 68–75, doi:10.1093/annonc/mdy470 (2019). [PubMed: 30407504]
- 106. Brooks JM et al. Development and Validation of a Combined Hypoxia and Immune Prognostic Classifier for Head and Neck Cancer. Clin Cancer Res 25, 5315–5328, doi:10.1158/1078-0432.CCR-18-3314 (2019). [PubMed: 31182433]
- 107. Feng B et al. Integrative Analysis of Multi-omics Data Identified EGFR and PTGS2 as Key Nodes in a Gene Regulatory Network Related to Immune Phenotypes in Head and Neck Cancer. Clin Cancer Res 26, 3616–3628, doi:10.1158/1078-0432.CCR-19-3997 (2020). [PubMed: 32161122]
- 108. Nguyen N et al. Tumor infiltrating lymphocytes and survival in patients with head and neck squamous cell carcinoma. Head Neck 38, 1074–1084, doi:10.1002/hed.24406 (2016). [PubMed: 26879675]
- 109. Fang J et al. Prognostic significance of tumor infiltrating immune cells in oral squamous cell carcinoma. BMC Cancer 17, 375, doi:10.1186/s12885-017-3317-2 (2017). [PubMed: 28549420]
- 110. Whiteside TL Immunobiology of head and neck cancer. Cancer Metastasis Rev 24, 95–105, doi:10.1007/s10555-005-5050-6 (2005). [PubMed: 15785875]
- 111. Ward MJ et al. Tumour-infiltrating lymphocytes predict for outcome in HPV-positive oropharyngeal cancer. Br J Cancer 110, 489–500, doi:10.1038/bjc.2013.639 (2014). [PubMed: 24169344]
- 112. Costa NL et al. Tumor-associated macrophages and the profile of inflammatory cytokines in oral squamous cell carcinoma. Oral Oncol 49, 216–223, doi:10.1016/j.oraloncology.2012.09.012 (2013). [PubMed: 23089461]

113. Ferris RL, Hunt JL & Ferrone S Human leukocyte antigen (HLA) class I defects in head and neck cancer: molecular mechanisms and clinical significance. Immunol Res 33, 113–133, doi:10.1385/IR:33:2:113 (2005). [PubMed: 16234579]

- 114. Ferris RL, Whiteside TL & Ferrone S Immune escape associated with functional defects in antigen-processing machinery in head and neck cancer. Clin Cancer Res 12, 3890–3895, doi:10.1158/1078-0432.CCR-05-2750 (2006). [PubMed: 16818683]
- 115. Grabowska AK & Riemer AB The invisible enemy how human papillomaviruses avoid recognition and clearance by the host immune system. Open Virol J 6, 249–256, doi:10.2174/1874357901206010249 (2012). [PubMed: 23341860]
- 116. DiMaio D & Petti LM The E5 proteins. Virology 445, 99–114, doi:10.1016/j.virol.2013.05.006 (2013). [PubMed: 23731971]
- 117. Karim R et al. Human papillomavirus (HPV) upregulates the cellular deubiquitinase UCHL1 to suppress the keratinocyte's innate immune response. PLoS Pathog 9, e1003384, doi:10.1371/journal.ppat.1003384 (2013). [PubMed: 23717208]
- 118. Gu Z, Shi W, Zhang L, Hu Z & Xu C USP19 suppresses cellular type I interferon signaling by targeting TRAF3 for deubiquitination. Future Microbiol 12, 767–779, doi:10.2217/fmb-2017-0006 (2017). [PubMed: 28391724]
- 119. Sorensen BS et al. Radiosensitivity and effect of hypoxia in HPV positive head and neck cancer cells. Radiother Oncol 108, 500–505, doi:10.1016/j.radonc.2013.06.011 (2013). [PubMed: 23953409]
- 120. Brizel DM, Sibley GS, Prosnitz LR, Scher RL & Dewhirst MW Tumor hypoxia adversely affects the prognosis of carcinoma of the head and neck. Int J Radiat Oncol Biol Phys 38, 285–289, doi:10.1016/s0360-3016(97)00101-6 (1997). [PubMed: 9226314]
- 121. Swartz JE et al. Poor prognosis in human papillomavirus-positive oropharyngeal squamous cell carcinomas that overexpress hypoxia inducible factor-1alpha. Head Neck 38, 1338–1346, doi:10.1002/hed.24445 (2016). [PubMed: 27027530]
- 122. Gottgens EL, Ostheimer C, Span PN, Bussink J & Hammond EM HPV, hypoxia and radiation response in head and neck cancer. Br J Radiol, 20180047, doi:10.1259/bjr.20180047 (2018).
- 123. Nordsmark M et al. Prognostic value of tumor oxygenation in 397 head and neck tumors after primary radiation therapy. An international multi-center study. Radiother Oncol 77, 18–24, doi:10.1016/j.radonc.2005.06.038 (2005). [PubMed: 16098619]
- 124. Linge A et al. Low Cancer Stem Cell Marker Expression and Low Hypoxia Identify Good Prognosis Subgroups in HPV(–) HNSCC after Postoperative Radiochemotherapy: A Multicenter Study of the DKTK-ROG. Clin Cancer Res 22, 2639–2649, doi:10.1158/1078-0432.CCR-15-1990 (2016). [PubMed: 26755529]
- 125. Bornigen D et al. Alterations in oral bacterial communities are associated with risk factors for oral and oropharyngeal cancer. Sci Rep 7, 17686, doi:10.1038/s41598-017-17795-z (2017). [PubMed: 29247187]
- 126. Mager DL et al. The salivary microbiota as a diagnostic indicator of oral cancer: a descriptive, non-randomized study of cancer-free and oral squamous cell carcinoma subjects. J Transl Med 3, 27, doi:10.1186/1479-5876-3-27 (2005). [PubMed: 15987522]
- 127. Banerjee S et al. Microbial Signatures Associated with Oropharyngeal and Oral Squamous Cell Carcinomas. Sci Rep 7, 4036, doi:10.1038/s41598-017-03466-6 (2017). [PubMed: 28642609]
- 128. Luukkaa M et al. Association between high collagenase-3 expression levels and poor prognosis in patients with head and neck cancer. Head Neck 28, 225–234, doi:10.1002/hed.20322 (2006). [PubMed: 16302191]
- 129. Patel BP, Shah SV, Shukla SN, Shah PM & Patel PS Clinical significance of MMP-2 and MMP-9 in patients with oral cancer. Head Neck 29, 564–572, doi:10.1002/hed.20561 (2007). [PubMed: 17252594]
- 130. Viros D et al. Prognostic role of MMP-9 expression in head and neck carcinoma patients treated with radiotherapy or chemoradiotherapy. Oral Oncol 49, 322–325, doi:10.1016/j.oraloncology.2012.10.005 (2013). [PubMed: 23122824]

131. Samanna V, Ma T, Mak TW, Rogers M & Chellaiah MA Actin polymerization modulates CD44 surface expression, MMP-9 activation, and osteoclast function. J Cell Physiol 213, 710–720, doi:10.1002/jcp.21137 (2007). [PubMed: 17508356]

- 132. Sterz CM et al. A basal-cell-like compartment in head and neck squamous cell carcinomas represents the invasive front of the tumor and is expressing MMP-9. Oral Oncol 46, 116–122, doi:10.1016/j.oraloncology.2009.11.011 (2010). [PubMed: 20036607]
- 133. Nijkamp MM et al. Expression of E-cadherin and vimentin correlates with metastasis formation in head and neck squamous cell carcinoma patients. Radiother Oncol 99, 344–348, doi:10.1016/j.radonc.2011.05.066 (2011). [PubMed: 21684617]
- 134. Yang MH et al. Direct regulation of TWIST by HIF-1alpha promotes metastasis. Nat Cell Biol 10, 295–305, doi:10.1038/ncb1691 (2008). [PubMed: 18297062]
- 135. Zhang Z, Filho MS & Nor JE The biology of head and neck cancer stem cells. Oral Oncol 48, 1–9, doi:10.1016/j.oraloncology.2011.10.004 (2012). [PubMed: 22070916]
- 136. Williams ED, Gao D, Redfern A & Thompson EW Controversies around epithelial-mesenchymal plasticity in cancer metastasis. Nat Rev Cancer 19, 716–732, doi:10.1038/s41568-019-0213-x (2019). [PubMed: 31666716]
- 137. Puram SV et al. Single-Cell Transcriptomic Analysis of Primary and Metastatic Tumor Ecosystems in Head and Neck Cancer. Cell 171, 1611–1624 e1624, doi:10.1016/j.cell.2017.10.044 (2017). [PubMed: 29198524]
- 138. Nirmala JG & Lopus M Cell death mechanisms in eukaryotes. Cell Biol Toxicol 36, 145–164, doi:10.1007/s10565-019-09496-2 (2020). [PubMed: 31820165]
- 139. Zeng Q et al. Hepatocyte growth factor inhibits anoikis in head and neck squamous cell carcinoma cells by activation of ERK and Akt signaling independent of NFkappa B. J Biol Chem 277, 25203–25208, doi:10.1074/jbc.M201598200 (2002). [PubMed: 11994287]
- 140. Neiva KG et al. Cross talk initiated by endothelial cells enhances migration and inhibits anoikis of squamous cell carcinoma cells through STAT3/Akt/ERK signaling. Neoplasia 11, 583–593, doi:10.1593/neo.09266 (2009). [PubMed: 19484147]
- 141. Huang SH et al. Refining American Joint Committee on Cancer/Union for International Cancer Control TNM stage and prognostic groups for human papillomavirus-related oropharyngeal carcinomas. J Clin Oncol 33, 836–845, doi:10.1200/JCO.2014.58.6412 (2015). [PubMed: 25667292]
- 142. Hashibe M et al. Interaction between tobacco and alcohol use and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. Cancer Epidemiol Biomarkers Prev 18, 541–550, doi:1055–9965.EPI-08–0347 [pii] 10.1158/1055-9965.EPI-08-0347 (2009). [PubMed: 19190158]
- 143. Taberna M et al. Human papillomavirus-related oropharyngeal cancer. Ann Oncol 28, 2386–2398, doi:10.1093/annonc/mdx304 (2017). [PubMed: 28633362]
- 144. Fu TS, Foreman A, Goldstein DP & de Almeida JR The role of transoral robotic surgery, transoral laser microsurgery, and lingual tonsillectomy in the identification of head and neck squamous cell carcinoma of unknown primary origin: a systematic review. J Otolaryngol Head Neck Surg 45, 28, doi:10.1186/s40463-016-0142-6 (2016). [PubMed: 27142355]
- 145. Meccariello G et al. The emerging role of trans-oral robotic surgery for the detection of the primary tumour site in patients with head-neck unknown primary cancers: A meta-analysis. Auris Nasus Larynx 46, 663–671, doi:10.1016/j.anl.2019.04.007 (2019). [PubMed: 31064689]
- 146. Chen YP et al. Nasopharyngeal carcinoma. Lancet 394, 64–80, doi:10.1016/S0140-6736(19)30956-0 (2019). [PubMed: 31178151]
- 147. Pynnonen MA et al. Clinical Practice Guideline: Evaluation of the Neck Mass in Adults. Otolaryngol Head Neck Surg 157, S1–S30, doi:10.1177/0194599817722550 (2017). [PubMed: 28891406]
- 148. Yi CH, Jim Zhai Q & Wang BY Updates on Immunohistochemical and Molecular Markers in Selected Head and Neck Diagnostic Problems. Arch Pathol Lab Med 141, 1214–1235, doi:10.5858/arpa.2016-0245-RA (2017). [PubMed: 28854343]

149. Lewis JS Jr. et al. Human Papillomavirus Testing in Head and Neck Carcinomas: Guideline From the College of American Pathologists. Arch Pathol Lab Med 142, 559–597, doi:10.5858/arpa.2017-0286-CP (2018). [PubMed: 29251996]

- 150. Ang KK et al. Human papillomavirus and survival of patients with oropharyngeal cancer. N Engl J Med 363, 24–35, doi:10.1056/NEJMoa0912217 (2010). [PubMed: 20530316]
- 151. Edge SB, B. D, Compton CC, Fritz AG, Greene FL, Trotti A, editors. . AJCC cancer staging manual (7th ed). . (New York, NY: Springer).
- 152. Amin MB, et al. (eds). AJCC Cancer Staging Manual, 8th edn (American Joint Committee on Cancer and Springer International Publishing, 2017).
- 153. Lydiatt W, O'Sullivan B & Patel S Major Changes in Head and Neck Staging for 2018. Am Soc Clin Oncol Educ Book 38, 505–514, doi:10.1200/EDBK_199697 (2018). [PubMed: 30231400]
- 154. van Gysen K et al. Validation of the 8(th) edition UICC/AJCC TNM staging system for HPV associated oropharyngeal cancer patients managed with contemporary chemoradiotherapy. BMC Cancer 19, 674, doi:10.1186/s12885-019-5894-8 (2019). [PubMed: 31288767]
- 155. Wurdemann N et al. Prognostic Impact of AJCC/UICC 8th Edition New Staging Rules in Oropharyngeal Squamous Cell Carcinoma. Front Oncol 7, 129, doi:10.3389/fonc.2017.00129 (2017). [PubMed: 28713770]
- 156. Lee VH et al. The addition of pretreatment plasma Epstein-Barr virus DNA into the eighth edition of nasopharyngeal cancer TNM stage classification. Int J Cancer 144, 1713–1722, doi:10.1002/ijc.31856 (2019). [PubMed: 30192385]
- 157. Guo R et al. Proposed modifications and incorporation of plasma Epstein-Barr virus DNA improve the TNM staging system for Epstein-Barr virus-related nasopharyngeal carcinoma. Cancer 125, 79–89, doi:10.1002/cncr.31741 (2019). [PubMed: 30351466]
- 158. Pfister DG et al. Head and Neck Cancers, Version 2.2020, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw. 18, 873–898 (2020). [PubMed: 32634781]
- 159. Coca-Pelaz A et al. The risk of second primary tumors in head and neck cancer: A systematic review. Head Neck 42, 456–466, doi:10.1002/hed.26016 (2020). [PubMed: 31750595]
- 160. U.S. Department of Health and Human Services. Smoking Cessation: A report of the Surgeon General. Atlanta, GA: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health, https://www.hhs.gov/sites/default/files/2020-cessation-sgr-full-report.pdf (2020).
- 161. World Health Organization. Tackling NCDs: "Best buys" and other recommended interventions for the prevention and control of noncommunicable diseases. Geneva: World Health Organization; https://apps.who.int/iris/handle/10665/259232. License: CC BY-NC-SA 3.0 IGO (2017).
- 162. Mehrtash H et al. Defining a global research and policy agenda for betel quid and areca nut. Lancet Oncol 18, e767–e775, doi:10.1016/S1470-2045(17)30460-6 (2017). [PubMed: 29208442]
- 163. Herrero R et al. Reduced prevalence of oral human papillomavirus (HPV) 4 years after bivalent HPV vaccination in a randomized clinical trial in Costa Rica. PLoS One 8, e68329, doi:10.1371/journal.pone.0068329 (2013). [PubMed: 23873171]
- 164. Hirth JM, Chang M, Resto VA & Group HPVS Prevalence of oral human papillomavirus by vaccination status among young adults (18–30years old). Vaccine 35, 3446–3451, doi:10.1016/j.vaccine.2017.05.025 (2017). [PubMed: 28526331]
- 165. Chaturvedi AK et al. Effect of Prophylactic Human Papillomavirus (HPV) Vaccination on Oral HPV Infections Among Young Adults in the United States. J Clin Oncol 36, 262–267, doi:10.1200/JCO.2017.75.0141 (2018). [PubMed: 29182497]
- 166. Meites E, Kempe A & Markowitz LE Use of a 2-Dose Schedule for Human Papillomavirus Vaccination - Updated Recommendations of the Advisory Committee on Immunization Practices. MMWR Morb Mortal Wkly Rep 65, 1405–1408, doi:10.15585/mmwr.mm6549a5 (2016). [PubMed: 27977643]
- 167. Meites E et al. Human Papillomavirus Vaccination for Adults: Updated Recommendations of the Advisory Committee on Immunization Practices. MMWR Morb Mortal Wkly Rep 68, 698–702, doi:10.15585/mmwr.mm6832a3 (2019). [PubMed: 31415491]

168. Napier SS & Speight PM Natural history of potentially malignant oral lesions and conditions: an overview of the literature. J Oral Pathol Med 37, 1–10, doi:10.1111/j.1600-0714.2007.00579.x (2008). [PubMed: 18154571]

- 169. Lee JJ et al. Predicting cancer development in oral leukoplakia: ten years of translational research. Clin Cancer Res 6, 1702–1710 (2000). [PubMed: 10815888]
- 170. Hong WK et al. 13-cis-retinoic acid in the treatment of oral leukoplakia. N Engl J Med 315, 1501–1505, doi:10.1056/NEJM198612113152401 (1986). [PubMed: 3537787]
- 171. Hong WK et al. Prevention of second primary tumors with isotretinoin in squamous-cell carcinoma of the head and neck. N Engl J Med 323, 795–801, doi:10.1056/NEJM199009203231205 (1990). [PubMed: 2202902]
- 172. Rosin MP et al. Use of allelic loss to predict malignant risk for low-grade oral epithelial dysplasia. Clin Cancer Res 6, 357–362 (2000). [PubMed: 10690511]
- 173. Garland LL et al. Effect of Intermittent Versus Continuous Low-Dose Aspirin on Nasal Epithelium Gene Expression in Current Smokers: A Randomized, Double-Blinded Trial. Cancer Prev Res (Phila) 12, 809–820, doi:10.1158/1940-6207.CAPR-19-0036 (2019). [PubMed: 31451521]
- 174. Vitale-Cross L et al. Metformin prevents the development of oral squamous cell carcinomas from carcinogen-induced premalignant lesions. Cancer Prev Res (Phila) 5, 562–573, doi:10.1158/1940-6207.CAPR-11-0502 (2012). [PubMed: 22467081]
- 175. Bauman JE et al. Prevention of Carcinogen-Induced Oral Cancer by Sulforaphane. Cancer Prev Res (Phila) 9, 547–557, doi:10.1158/1940-6207.CAPR-15-0290 (2016). [PubMed: 27339168]
- 176. Hu L et al. Gene targets of sulforaphane in head and neck squamous cell carcinoma. Mol Med Rep 20, 5335–5344, doi:10.3892/mmr.2019.10766 (2019). [PubMed: 31661135]
- 177. Lee NCJ et al. Patterns of failure in high-metastatic node number human papillomavirus-positive oropharyngeal carcinoma. Oral Oncol 85, 35–39, doi:10.1016/j.oraloncology.2018.08.001 (2018). [PubMed: 30220317]
- 178. Lyhne NM et al. The DAHANCA 6 randomized trial: Effect of 6 vs 5 weekly fractions of radiotherapy in patients with glottic squamous cell carcinoma. Radiother Oncol 117, 91–98, doi:10.1016/j.radonc.2015.07.004 (2015). [PubMed: 26255764]
- 179. Bledsoe TJ et al. Hypofractionated Radiotherapy for Patients with Early-Stage Glottic Cancer: Patterns of Care and Survival. J Natl Cancer Inst 109, doi:10.1093/jnci/djx042 (2017).
- 180. Weinstein GS et al. Transoral robotic surgery: a multicenter study to assess feasibility, safety, and surgical margins. Laryngoscope 122, 1701–1707, doi:10.1002/lary.23294 (2012). [PubMed: 22752997]
- 181. Forastiere AA et al. Use of Larynx-Preservation Strategies in the Treatment of Laryngeal Cancer: American Society of Clinical Oncology Clinical Practice Guideline Update. J Clin Oncol 36, 1143–1169, doi:10.1200/JCO.2017.75.7385 (2018). [PubMed: 29172863]
- 182. D'Cruz AK et al. Elective versus Therapeutic Neck Dissection in Node-Negative Oral Cancer. N Engl J Med 373, 521–529, doi:10.1056/NEJMoa1506007 (2015). [PubMed: 26027881]
- 183. Forastiere AA et al. Concurrent chemotherapy and radiotherapy for organ preservation in advanced laryngeal cancer. N Engl J Med 349, 2091–2098, doi:10.1056/NEJMoa031317 (2003). [PubMed: 14645636]
- 184. Cooper JS et al. Postoperative concurrent radiotherapy and chemotherapy for high-risk squamous-cell carcinoma of the head and neck. N Engl J Med 350, 1937–1944, doi:10.1056/NEJMoa032646 (2004). [PubMed: 15128893]
- 185. Bernier J et al. Postoperative irradiation with or without concomitant chemotherapy for locally advanced head and neck cancer. N Engl J Med 350, 1945–1952, doi:10.1056/NEJMoa032641 (2004). [PubMed: 15128894]
- 186. Forastiere AA et al. Long-term results of RTOG 91–11: a comparison of three nonsurgical treatment strategies to preserve the larynx in patients with locally advanced larynx cancer. J Clin Oncol 31, 845–852, doi:10.1200/JCO.2012.43.6097 (2013). [PubMed: 23182993]
- 187. Kelly JR et al. Upfront surgery versus definitive chemoradiotherapy in patients with human Papillomavirus-associated oropharyngeal squamous cell cancer. Oral Oncol 79, 64–70, doi:10.1016/j.oraloncology.2018.02.017 (2018). [PubMed: 29598952]

188. Kann BH et al. Pretreatment Identification of Head and Neck Cancer Nodal Metastasis and Extranodal Extension Using Deep Learning Neural Networks. Sci Rep 8, 14036, doi:10.1038/s41598-018-32441-y (2018). [PubMed: 30232350]

- 189. Lowe VJ et al. Multicenter Trial of [(18)F]fluorodeoxyglucose Positron Emission Tomography/ Computed Tomography Staging of Head and Neck Cancer and Negative Predictive Value and Surgical Impact in the N0 Neck: Results From ACRIN 6685. J Clin Oncol 37, 1704–1712, doi:10.1200/JCO.18.01182 (2019). [PubMed: 30768363]
- 190. Adelstein DJ et al. An intergroup phase III comparison of standard radiation therapy and two schedules of concurrent chemoradiotherapy in patients with unresectable squamous cell head and neck cancer. J Clin Oncol 21, 92–98, doi:10.1200/JCO.2003.01.008 (2003). [PubMed: 12506176]
- 191. Bauml JM et al. Cisplatin Every 3 Weeks Versus Weekly With Definitive Concurrent Radiotherapy for Squamous Cell Carcinoma of the Head and Neck. J Natl Cancer Inst 111, 490–497, doi:10.1093/jnci/djy133 (2019). [PubMed: 30239887]
- 192. Worden FP et al. Chemoselection as a strategy for organ preservation in patients with T4 laryngeal squamous cell carcinoma with cartilage invasion. Laryngoscope 119, 1510–1517, doi:10.1002/lary.20294 (2009). [PubMed: 19504552]
- 193. Blanchard P et al. Chemotherapy and radiotherapy in nasopharyngeal carcinoma: an update of the MAC-NPC meta-analysis. Lancet Oncol 16, 645–655, doi:10.1016/S1470-2045(15)70126-9 (2015). [PubMed: 25957714]
- 194. Zhang Y et al. Gemcitabine and Cisplatin Induction Chemotherapy in Nasopharyngeal Carcinoma. N Engl J Med 381, 1124–1135, doi:10.1056/NEJMoa1905287 (2019). [PubMed: 31150573]
- 195. Chan ATC et al. Analysis of Plasma Epstein-Barr Virus DNA in Nasopharyngeal Cancer After Chemoradiation to Identify High-Risk Patients for Adjuvant Chemotherapy: A Randomized Controlled Trial. J Clin Oncol, JCO2018777847, doi:10.1200/JCO.2018.77.7847 (2018).
- 196. Trotti A et al. TAME: development of a new method for summarising adverse events of cancer treatment by the Radiation Therapy Oncology Group. Lancet Oncol 8, 613–624, doi:10.1016/S1470-2045(07)70144-4 (2007). [PubMed: 17543584]
- 197. Haddad R et al. Induction chemotherapy followed by concurrent chemoradiotherapy (sequential chemoradiotherapy) versus concurrent chemoradiotherapy alone in locally advanced head and neck cancer (PARADIGM): a randomised phase 3 trial. Lancet Oncol 14, 257–264, doi:10.1016/S1470-2045(13)70011-1 (2013). [PubMed: 23414589]
- 198. Cohen EE et al. Phase III randomized trial of induction chemotherapy in patients with N2 or N3 locally advanced head and neck cancer. J Clin Oncol 32, 2735–2743, doi:10.1200/ JCO.2013.54.6309 (2014). [PubMed: 25049329]
- 199. Posner MR et al. Cisplatin and fluorouracil alone or with docetaxel in head and neck cancer. N Engl J Med 357, 1705–1715, doi:10.1056/NEJMoa070956 (2007). [PubMed: 17960013]
- 200. Kowalski LP et al. COVID-19 pandemic: effects and evidence-based recommendations for otolaryngology and head and neck surgery practice. Head Neck, doi:10.1002/hed.26164 (2020).
- 201. Fakhry C et al. Improved survival of patients with human papillomavirus-positive head and neck squamous cell carcinoma in a prospective clinical trial. J Natl Cancer Inst 100, 261–269, doi:djn011 [pii] 10.1093/jnci/djn011 (2008). [PubMed: 18270337]
- 202. Marur S et al. E1308: Phase II Trial of Induction Chemotherapy Followed by Reduced-Dose Radiation and Weekly Cetuximab in Patients With HPV-Associated Resectable Squamous Cell Carcinoma of the Oropharynx- ECOG-ACRIN Cancer Research Group. J Clin Oncol 35, 490– 497, doi:10.1200/JCO.2016.68.3300 (2017). [PubMed: 28029303]
- 203. Chen AM et al. Reduced-dose radiotherapy for human papillomavirus-associated squamous-cell carcinoma of the oropharynx: a single-arm, phase 2 study. Lancet Oncol 18, 803–811, doi:10.1016/S1470-2045(17)30246-2 (2017). [PubMed: 28434660]
- 204. Lee AW et al. Retrospective analysis of patients with nasopharyngeal carcinoma treated during 1976–1985: survival after local recurrence. Int J Radiat Oncol Biol Phys 26, 773–782, doi:10.1016/0360-3016(93)90491-d (1993). [PubMed: 8344845]

205. Fakhry C et al. Human papillomavirus and overall survival after progression of oropharyngeal squamous cell carcinoma. J Clin Oncol 32, 3365–3373, doi:10.1200/JCO.2014.55.1937 (2014). [PubMed: 24958820]

- 206. Saada-Bouzid E et al. Hyperprogression during anti-PD-1/PD-L1 therapy in patients with recurrent and/or metastatic head and neck squamous cell carcinoma. Ann Oncol 28, 1605–1611, doi:10.1093/annonc/mdx178 (2017). [PubMed: 28419181]
- 207. Saleh K et al. Response to salvage chemotherapy after progression on immune checkpoint inhibitors in patients with recurrent and/or metastatic squamous cell carcinoma of the head and neck. Eur J Cancer 121, 123–129, doi:10.1016/j.ejca.2019.08.026 (2019). [PubMed: 31574417]
- 208. Puzanov I et al. Managing toxicities associated with immune checkpoint inhibitors: consensus recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. J Immunother Cancer 5, 95, doi:10.1186/s40425-017-0300-z (2017). [PubMed: 29162153]
- 209. Vermorken JB et al. Platinum-based chemotherapy plus cetuximab in head and neck cancer. N Engl J Med 359, 1116–1127, doi:359/11/1116 [pii] 10.1056/NEJMoa0802656 (2008). [PubMed: 18784101]
- 210. Cohen EEW et al. Pembrolizumab versus methotrexate, docetaxel, or cetuximab for recurrent or metastatic head-and-neck squamous cell carcinoma (KEYNOTE-040): a randomised, open-label, phase 3 study. Lancet 393, 156–167, doi:10.1016/S0140-6736(18)31999-8 (2019). [PubMed: 30509740]
- 211. Mehra R et al. Efficacy and safety of pembrolizumab in recurrent/metastatic head and neck squamous cell carcinoma: pooled analyses after long-term follow-up in KEYNOTE-012. Br J Cancer 119, 153–159, doi:10.1038/s41416-018-0131-9 (2018). [PubMed: 29955135]
- 212. Haddad R et al. Nivolumab treatment beyond RECIST-defined progression in recurrent or metastatic squamous cell carcinoma of the head and neck in CheckMate 141: A subgroup analysis of a randomized phase 3 clinical trial. Cancer 125, 3208–3218, doi:10.1002/cncr.32190 (2019). [PubMed: 31246283]
- 213. Taylor MH et al. Phase IB/II Trial of Lenvatinib Plus Pembrolizumab in Patients With Advanced Renal Cell Carcinoma, Endometrial Cancer, and Other Selected Advanced Solid Tumors. J Clin Oncol 38, 1154–1163, doi:10.1200/JCO.19.01598 (2020). [PubMed: 31961766]
- 214. Verdonck-de Leeuw IM et al. The course of health-related quality of life in head and neck cancer patients treated with chemoradiation: a prospective cohort study. Radiother Oncol 110, 422–428, doi:10.1016/j.radonc.2014.01.002 (2014). [PubMed: 24582144]
- 215. Rettig EM et al. Health-related quality of life before and after head and neck squamous cell carcinoma: Analysis of the Surveillance, Epidemiology, and End Results-Medicare Health Outcomes Survey linkage. Cancer 122, 1861–1870, doi:10.1002/cncr.30005 (2016). [PubMed: 27182694]
- 216. Quinten C et al. Baseline quality of life as a prognostic indicator of survival: a meta-analysis of individual patient data from EORTC clinical trials. Lancet Oncol 10, 865–871, doi:10.1016/S1470-2045(09)70200-1 (2009). [PubMed: 19695956]
- 217. Rogers SN et al. Quality of life, cognitive, physical and emotional function at diagnosis predicts head and neck cancer survival: analysis of cases from the Head and Neck 5000 study. Eur Arch Otorhinolaryngol 277, 1515–1523, doi:10.1007/s00405-020-05850-x (2020). [PubMed: 32062743]
- 218. Hammerlid E, Silander E, Hornestam L & Sullivan M Health-related quality of life three years after diagnosis of head and neck cancer--a longitudinal study. Head Neck 23, 113–125, doi:10.1002/1097-0347(200102)23:2<113::aid-hed1006>3.0.co;2-w (2001). [PubMed: 11303628]
- 219. Mehanna HM & Morton RP Deterioration in quality-of-life of late (10-year) survivors of head and neck cancer. Clin Otolaryngol 31, 204–211, doi:10.1111/j.1749-4486.2006.01188.x (2006). [PubMed: 16759240]
- 220. Verdonck-de Leeuw IM, van Nieuwenhuizen A & Leemans CR The value of quality-of-life questionnaires in head and neck cancer. Curr Opin Otolaryngol Head Neck Surg 20, 142–147, doi:10.1097/MOO.0b013e32834f5fd7 (2012). [PubMed: 22249168]

221. Singer S et al. International validation of the revised European Organisation for Research and Treatment of Cancer Head and Neck Cancer Module, the EORTC QLQ-HN43: Phase IV. Head Neck 41, 1725–1737, doi:10.1002/hed.25609 (2019). [PubMed: 30636188]

- 222. List MA et al. The Performance Status Scale for Head and Neck Cancer Patients and the Functional Assessment of Cancer Therapy-Head and Neck Scale. A study of utility and validity. Cancer 77, 2294–2301, doi:10.1002/(SICI)1097-0142(19960601)77:11<2294::AID-CNCR17>3.0.CO;2-S (1996). [PubMed: 8635098]
- 223. Nekhlyudov L et al. Head and Neck Cancer Survivorship Care Guideline: American Society of Clinical Oncology Clinical Practice Guideline Endorsement of the American Cancer Society Guideline. J Clin Oncol 35, 1606–1621, doi:10.1200/JCO.2016.71.8478 (2017). [PubMed: 28240970]
- 224. Ringash J et al. Head and Neck Cancer Survivorship: Learning the Needs, Meeting the Needs. Semin Radiat Oncol 28, 64–74, doi:10.1016/j.semradonc.2017.08.008 (2018). [PubMed: 29173757]
- 225. Bonomo P et al. Quality Assessment in Supportive Care in Head and Neck Cancer. Front Oncol 9, 926, doi:10.3389/fonc.2019.00926 (2019). [PubMed: 31620372]
- 226. Rinkel RN et al. Prevalence of swallowing and speech problems in daily life after chemoradiation for head and neck cancer based on cut-off scores of the patient-reported outcome measures SWAL-QOL and SHI. Eur Arch Otorhinolaryngol 273, 1849–1855, doi:10.1007/s00405-015-3680-z (2016). [PubMed: 26071622]
- 227. Kraaijenga SA et al. Assessment of voice, speech, and related quality of life in advanced head and neck cancer patients 10-years+ after chemoradiotherapy. Oral Oncol 55, 24–30, doi:10.1016/j.oraloncology.2016.02.001 (2016). [PubMed: 26874554]
- 228. Hutcheson KA et al. Two-year prevalence of dysphagia and related outcomes in head and neck cancer survivors: An updated SEER-Medicare analysis. Head Neck 41, 479–487, doi:10.1002/hed.25412 (2019). [PubMed: 30536748]
- 229. Bjordal K & Bottomley A Making Advances in Quality of Life Studies in Head and Neck Cancer. Int J Radiat Oncol Biol Phys 97, 659–661, doi:10.1016/j.ijrobp.2016.11.051 (2017). [PubMed: 28244403]
- 230. Dawson C et al. Patient advocacy in head and neck cancer: Realities, challenges and the role of the multi-disciplinary team. Clin Otolaryngol 45, 437–444, doi:10.1111/coa.13508 (2020). [PubMed: 31971339]
- 231. van der Hout A et al. Role of eHealth application Oncokompas in supporting self-management of symptoms and health-related quality of life in cancer survivors: a randomised, controlled trial. Lancet Oncol 21, 80–94, doi:10.1016/S1470-2045(19)30675-8 (2020). [PubMed: 31838009]
- 232. Parke SC et al. Identifying Gaps in Research on Rehabilitation for Patients With Head and Neck Cancer: A Scoping Review. Arch Phys Med Rehabil 100, 2381–2388, doi:10.1016/j.apmr.2019.03.022 (2019). [PubMed: 31082380]
- 233. Kampshoff CS, Verdonck-de Leeuw IM, van Oijen MG, Sprangers MA & Buffart LM Ecological momentary assessments among patients with cancer: A scoping review. Eur J Cancer Care (Engl) 28, e13095, doi:10.1111/ecc.13095 (2019). [PubMed: 31090160]
- 234. David JM et al. Treatment at high-volume facilities and academic centers is independently associated with improved survival in patients with locally advanced head and neck cancer. Cancer 123, 3933–3942, doi:10.1002/cncr.30843 (2017). [PubMed: 28640546]
- 235. Wuthrick EJ et al. Institutional clinical trial accrual volume and survival of patients with head and neck cancer. J Clin Oncol 33, 156–164, doi:10.1200/JCO.2014.56.5218 (2015). [PubMed: 25488965]
- 236. Bossi P et al. Impact of treatment expertise on the outcome of patients with head and neck cancer treated within 6 randomized trials. Head Neck 40, 2648–2656, doi:10.1002/hed.25389 (2018). [PubMed: 30447127]
- 237. Ellington TD et al. Trends in Incidence of Cancers of the Oral Cavity and Pharynx United States 2007–2016. MMWR Morb Mortal Wkly Rep 69, 433–438, doi:10.15585/mmwr.mm6915a1 (2020). [PubMed: 32298244]

238. Sonawane K et al. Oral Human Papillomavirus Infection: Differences in Prevalence Between Sexes and Concordance With Genital Human Papillomavirus Infection, NHANES 2011 to 2014. Ann Intern Med 167, 714–724, doi:10.7326/M17-1363 (2017). [PubMed: 29049523]

- Tipifarnib Targets HRAS-Mutant Cancers. Cancer Discov 9, 1637–1638, doi:10.1158/2159-8290.CD-NB2019-129 (2019).
- 240. Elkabets M et al. AXL mediates resistance to PI3Kalpha inhibition by activating the EGFR/PKC/mTOR axis in head and neck and esophageal squamous cell carcinomas. Cancer Cell 27, 533–546, doi:10.1016/j.ccell.2015.03.010 (2015). [PubMed: 25873175]
- 241. Merino DM et al. Establishing guidelines to harmonize tumor mutational burden (TMB): in silico assessment of variation in TMB quantification across diagnostic platforms: phase I of the Friends of Cancer Research TMB Harmonization Project. J Immunother Cancer 8, doi:10.1136/jitc-2019-000147 (2020).
- 242. Castellanos E & Baxi SS Letting the GENIE Out of Its Bottle: Examining the Potential of Real-World Clinicogenomic Data. Cancer Discov 10, 490–491, doi:10.1158/2159-8290.CD-20-0039 (2020). [PubMed: 32238395]
- 243. Cillo AR et al. Immune Landscape of Viral- and Carcinogen-Driven Head and Neck Cancer. Immunity 52, 183–199 e189, doi:10.1016/j.immuni.2019.11.014 (2020). [PubMed: 31924475]
- 244. Li H et al. Proteomic Characterization of Head and Neck Cancer Patient-Derived Xenografts. Mol Cancer Res 14, 278–286, doi:10.1158/1541-7786.MCR-15-0354 (2016). [PubMed: 26685214]
- 245. Karamboulas C et al. Patient-Derived Xenografts for Prognostication and Personalized Treatment for Head and Neck Squamous Cell Carcinoma. Cell Rep 25, 1318–1331 e1314, doi:10.1016/j.celrep.2018.10.004 (2018). [PubMed: 30380421]
- 246. Driehuis E et al. Oral Mucosal Organoids as a Potential Platform for Personalized Cancer Therapy. Cancer Discov 9, 852–871, doi:10.1158/2159-8290.CD-18-1522 (2019). [PubMed: 31053628]
- 247. Wang H et al. Immune Checkpoint Inhibitor Toxicity in Head and Neck Cancer: From Identification to Management. Front Pharmacol 10, 1254, doi:10.3389/fphar.2019.01254 (2019). [PubMed: 31708780]
- 248. Wang Z et al. Syngeneic animal models of tobacco-associated oral cancer reveal the activity of in situ anti-CTLA-4. Nat Commun 10, 5546, doi:10.1038/s41467-019-13471-0 (2019). [PubMed: 31804466]
- 249. Barry KC et al. A natural killer-dendritic cell axis defines checkpoint therapy-responsive tumor microenvironments. Nat Med 24, 1178–1191, doi:10.1038/s41591-018-0085-8 (2018). [PubMed: 29942093]
- 250. Shah FD et al. A review on salivary genomics and proteomics biomarkers in oral cancer. Indian J Clin Biochem 26, 326–334, doi:10.1007/s12291-011-0149-8 (2011). [PubMed: 23024467]

Box 1.

Patient experience

A 58-year old male patient who is a board member of a large international company, underwent platinum-based chemoradiotherapy (70 Gy) for a T1N1 HPV- positive oropharyngeal carcinoma from the left tonsil one year ago, and describes his experience:

"The reaction from my team at work was superb. We agreed I would focus 100% on myself, and within a week I was entirely out of the loop. It was sad as well, to find out how quickly you become dispensable. These feelings stayed with me for months, even after I returned to work 6 months later. Disconcerting was the discovery that my treatment for a stage 1 cancer would be basically the same as for advanced cancer.

The first round of chemo went surprisingly well. The radiotherapy sessions were well organized and pleasantly on time. The second chemo session hammered me, and the radiotherapy sessions started taking their toll. My appetite went down as my mouth went dry and my sense of taste slipped away. By the time of the third chemo session I absolutely knew and felt that I was in a war of attrition. Somewhere along the line I developed a persistent diarrhea that added to the feeling of being drained. Entering my last week of radiotherapy, I was dehydrated, underfed, and weak to the point of having to lie down while waiting for the radiotherapy sessions.

Thankfully some of my worst fears did not materialize, such as being put on a feeding tube or getting radiotherapy burns that would have required additional interventions. During the treatment my throat did swell and became constrained, but in my view this was not the major cause of the relatively mild swallowing or speech problems I experienced. The main factors were having a dry mouth and being foggy-brained. The fogginess took about two months to clear, about as long as it took me to stop carrying a water bottle everywhere all the time. I had anticipated this would take longer and I think it's a benefit of having been irradiated on one side of the throat only.

There are five remaining physical symptoms from my treatment: mouth dryness (back at 85% of where I was), loss of taste, alopecia spots in my beard and frequent tinnitus in the ears, and lower energy levels. At work and at home and socially things are basically back to normal as well. Generally, I take things easier and I don't stress about the discomforts; it's way better than where I was and could have ended up.

Whatever happens, it's clear that the fear of falling badly sick again and spending days in hospital is something that consciously or unconsciously besets me and probably all surviving patients with cancer."

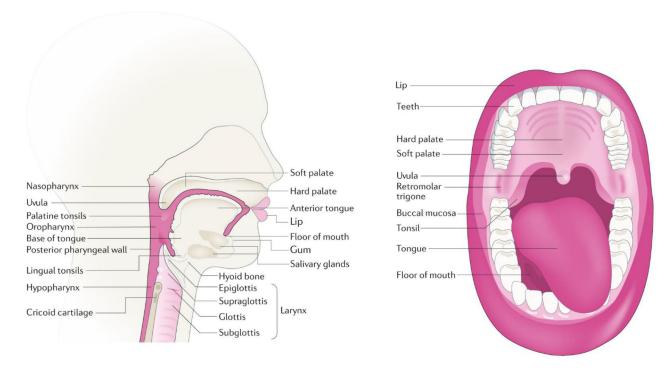
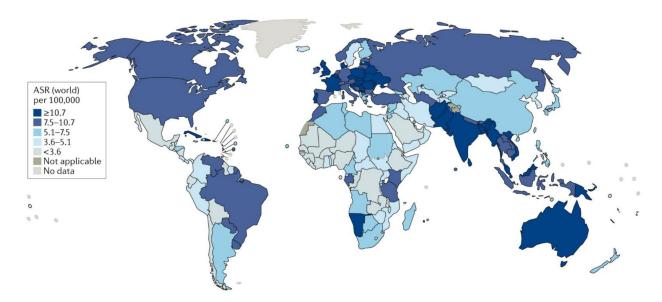


Figure 1. Anatomical sites of HNSCC development.

Head and neck squamous cell carcinoma (HNSCC) arises from the mucosal epithelium of the oral cavity (lips, buccal mucosa, hard palate, anterior tongue, floor of mouth and retromolar trigone), nasopharynx, oropharynx (palantine tonsils, lingual tonsils, base of tongue, soft palate, uvula and posterior pharyngeal wall), hypopharynx (the bottom part of the throat, extending from the hyoid bone to the cricoid cartilage) and larynx. Human papilloma virus-associated HNSCCs arise primarily from the palantine and lingual tonsils of the oropharynx, whereas tobacco-associated HNSCCs arise primarily in the oral cavity, hypopharynx and larynx.



 $\label{lem:continuous} \textbf{Figure 2. Global incidence of head and neck squamous cell carcinoma.}$

The estimated age-standardized rate (ASR) of HNSCC incidence worldwide is shown for men and women combined ^{10–12}. Data from GLOBOCAN, 2018 ¹⁰. Map was generated using the GLOBOCAN website mapping tool (https://gco.iarc.fr/today/online-analysis-map) by selecting the 'hypopharynx', 'larynx', 'lip, oral cavity', 'nasopharynx' and 'oropharynx' cancer sites.

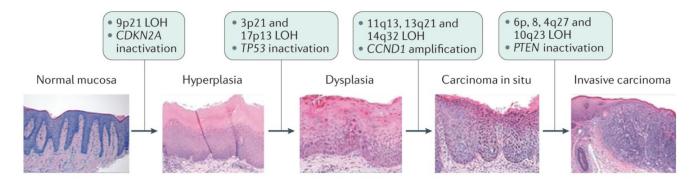


Figure 3. Progression of HNSCC and key genetic events.

The mucosal epithelium lining the oral cavity, pharynx, larynx and sinonasal tract is the site of origin for head and neck squamous cell carcinoma (HNSCC). In a model of ordered histological progression of HNSCC⁶⁸, mucosal epithelial cell hyperplasia is followed by dysplasia, and carcinoma in situ precedes the development of invasive carcinoma. Specific genetic events have been found to be enriched at each stage of progression and are indicated. Of note, unlike in most cancers in which oncogenic mutations typically drive tumorigenesis, HNSCC formation usually involves the inactivation of tumour suppressor genes, such as CDKN2A and TP53 (encoding p16INK4A and p53, respectively) in early stages and PTEN (encoding phosphatase and tensin homologue (PTEN)) at later stages. LOH, loss of heterozygosity. Histopathology images of hyperplasia, dysplasia, carcinoma in situ and invasive carcinoma are reprinted from ref. 250²⁵⁰, Springer Nature Limited. Histopathology image of normal mucosa courtesy of R. Jordan, University of California, San Francisco.

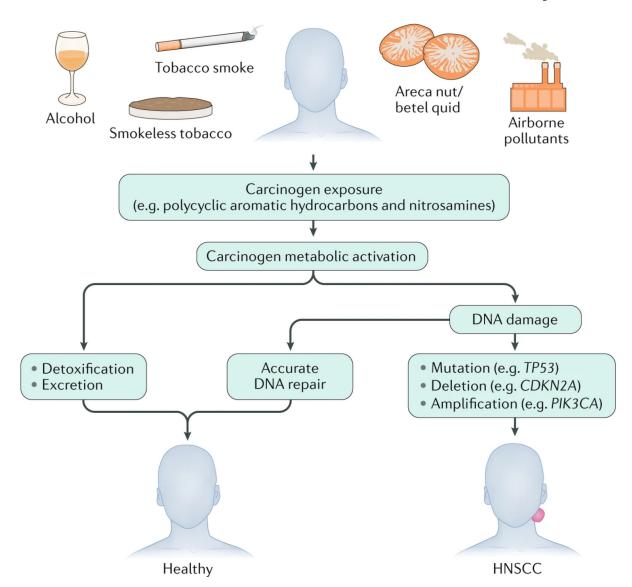
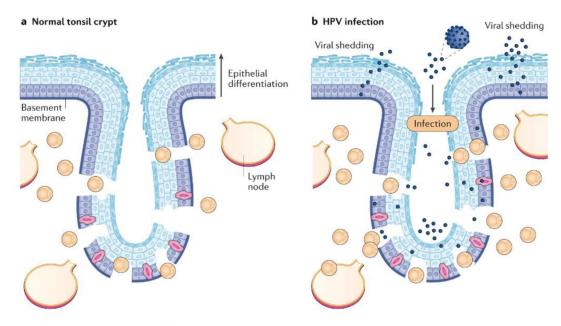


Figure 4. Development of carcinogen-associated, HPV-negative HNSCC.

Consumption of tobacco products or betel quid (the leaf of *Piper betle*) and areca nut (*Areca catechu*), exposure to environmental pollutants or excessive alcohol consumption are primary factors in the development of human papilloma virus (HPV)-negative head and neck squamous cell carcinoma (HNSCC). Tobacco and tobacco smoke, in particular, are rich in polycyclic aromatic hydrocarbons and nitrosamines, which are known human carcinogens and are associated with a strongly increased risk of HNSCC. Metabolic activation of carcinogens results in the formation of reactive metabolites, which, if not detoxified and excreted, can damage DNA, typically by generating bulky DNA adducts. If the DNA damage is faithfully and accurately repaired, there may be no lasting consequences. However, if the damaged DNA is not promptly repaired, or is repaired errantly by lower fidelity repair mechanisms, then permanent damage in the form of mutations, deletions and amplifications can occur. The accumulation of alterations in key tumour suppressor genes (such as *TP53* and *CDKN2A*, which encode p53 and p16^{INK4A}, respectively) or signalling

pathways (such as PI3K–AKT–mTOR and RAS–MAPK pathway genes) is associated with the onset, progression and poor prognosis of HPV-negative HNSCC.



c HPV-mediated malignant transformation

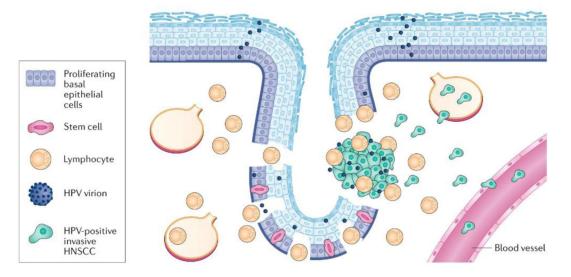


Figure 5. HPV infection of the tonsil crypt and development of HPV-positive HNSCC.

a | Throughout the tonsil epithelium, proliferating basal epithelial cells constitute the cell layer adjacent to the basement membrane. Differentiation of basal epithelial cells leads to their detachment from the basement membrane and upward migration, with progressively increasing differentiation leading to the sloughing off of terminally differentiated, non-proliferating cells. The palatine and lingual tonsils are also characterized by numerous tissue invaginations, commonly termed crypts, which are particularly enriched in stem cells at their base. A unique, reticulated squamous epithelium lines the crypt structure and gaps or fissures in the basement membrane and basal layer also occur. The presence of these fissures allows lymphocytes to enter the crypts and directly interact with foreign, external antigens. **b** | During infection with human papilloma virus (HPV), the reticulated and disrupted nature of

the crypt squamous epithelium allows viral access to stem cells, proliferating basal cells and the basement membrane. Infiltrating immune cells also make contact with the viral particles. In the case of a productive infection, distinct viral genes and proteins are induced and/or activated during the different stages of epithelial cell differentiation, culminating in the production and shedding of new viral particles. \mathbf{c} | Stem cells or proliferating basal cells represent probable cells of origin for HPV-positive head and neck squamous cell carcinoma (HNSCC). Stable integration of the viral genome into the host genome, and the concerted action of HPV E6 and E7 proteins on cellular p53 and RB levels, respectively, acts to promote cellular transformation. The accumulation of additional genetic alterations is needed to induce full transformation, including the acquisition of invasive and metastatic phenotypes.

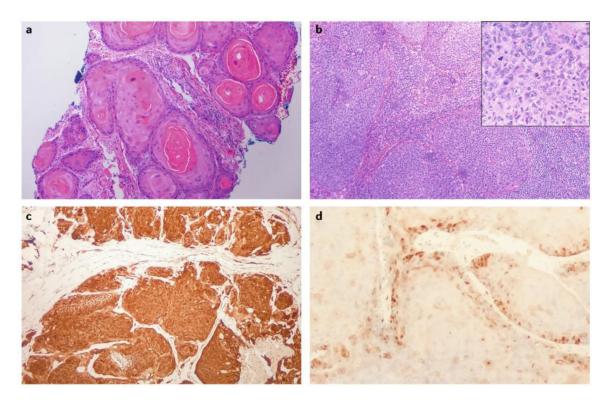


Figure 6. Histopathology of HNSCC.

 ${\bf a}$ | Well-differentiated squamous cell carcinoma of the oral tongue, demonstrating mature cells with semi-organized keratinization and featuring a 'keratin pearl' (10x). ${\bf b}$ | A poorly differentiated squamous cell carcinoma of the base of tongue (10x). Inset shows immature cells with nuclear polymorphism and atypical mitoses without apparent stratification or keratinization (40x). ${\bf c}$ | A p16 INK4A -positive oropharyngeal squamous cell carcinoma characterized by diffuse nuclear and cytoplasmic staining for the cell-cycle protein p16 INK4A by immunohistochemistry, indicative of HPV-positive disease (10x). ${\bf d}$ | A p16 INK4A -negative oropharyngeal squamous cell carcinoma demonstrates minimal staining by the same anti-p16 INK4A - antibody, a staining intensity that is indicative of HPV-negative disease (40x). In parts ${\bf a}$ and ${\bf b}$, staining is with haematoxylin and eosin. HNSCC, head and neck squamous cell carcinoma.

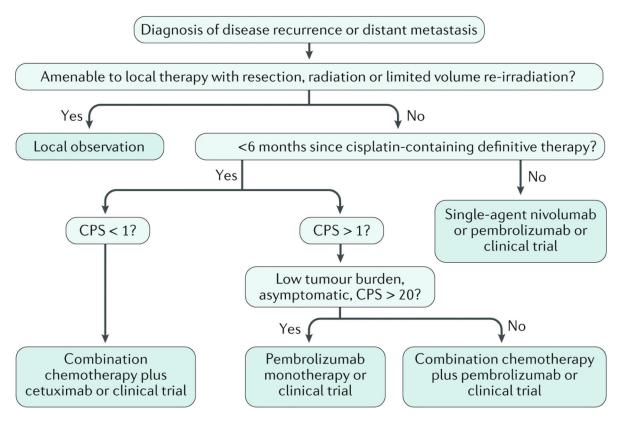


Figure 7. Algorithm for treatment-decision making for recurrent and/or metastatic HNSCC. After a diagnosis of recurrent disease or distant metastasis, patients with disease that is amenable to local therapy receive resection, radiation or limited volume irradiation and are then subject to observation. However, in patients who are not amenable to these therapies, systemic therapy is indicated. Pembrolizumab is chosen for those with expression of programmed cell death 1 ligand 1 (PDL1) expression (composite positive score (CPS)>1; CPS is: (the number of PDL1-staining tumor and immune cells/total number of viable tumour cells counted)X100), given alone for those with high expression and asymptomatic disease (CPS>20), or given with platinum-based chemotherapy for those with lower PDL1 expression (CPS 1–19) or higher symptom burden. If PDL1 expression is absent (CPS<1), platinum doublet chemotherapy with cetuximab is appropriate first-line therapy. Patients who relapse after platinum-based therapy are candidates for nivolumab or pembrolizumab.

Table 1.

Current NCTN head and neck cancer trials

Protocol number	Phase	Protocol title
EA3132	II	Randomized trial of radiotherapy with or without cisplatin for surgically resected squamous cell carcinoma of the head and neck (SCCHN) with <i>TP53</i> sequencing
EA3161	II/III	Randomized study of maintenance nivolumab versus observation in patients with locally advanced, intermediate risk HPV-positive OPCA
EA3163	II	Randomized trial of neo-adjuvant chemotherapy followed by surgery and post-operative radiation versus surgery and post-operative radiation for organ preservation of T3 and T4a nasal and paranasal sinus squamous cell carcinoma (NPNSCC)
NRG-HN001	II/III	Randomized studies of individualized treatment for nasopharyngeal carcinoma based on biomarker Epstein Barr Virus (EBV) deoxyribonucleic acid (DNA)
NRG-HN004	II/III	Randomized of radiotherapy with concurrent MEDI4736 (durvalumab) versus radiotherapy with concurrent cetuximab in patients with stage III-IVB head and neck cancer with a contraindication to cisplatin
NRG-HN005	II/III	Randomized trial of de-intensified radiation therapy for patients with early-stage, p16-positive, non-smoking associated oropharyngeal cancer
RTOG-1008	II	Randomized study of adjuvant concurrent radiation and chemotherapy versus radiation alone in resected high- risk malignant salivary gland tumours
RTOG-1216	II/III	Randomized trial of surgery and postoperative radiation delivered with concurrent cisplatin versus docetaxel versus docetaxel and cetuximab for high-risk squamous cell cancer of the head and neck
EAY131	II	Molecular analysis for therapy choice (MATCH)
S1609	II	Dual anti-CTLA4 and anti-PD1 blockade in rare tumours (DART)

^aTrials that are open for patient enrolment as of 15 April 2020. CTLA4, cytotoxic T lymphocyte antigen 4; HNSCC, head and neck squamous cell carcinoma; NCTN, National Clinical Trials Network; OPCA, oral and pharyngeal cancer; PD1, programmed cell death 1.