



Article

Synthesis of Densely Immobilized Gold-Assembled Silica Nanostructures

Bomi Seong ¹, Sungje Bock ¹, Eunil Hahm ¹, Kim-Hung Huynh ¹, Jaehi Kim ¹, Sang Hun Lee ², Xuan-Hung Pham ^{1,*} and Bong-Hyun Jun ¹

¹ Department of Bioscience and Biotechnology, Konkuk University, Seoul 143-701, Korea; iambomi33@konkuk.ac.kr (B.S.); bsj4126@konkuk.ac.kr (S.B.); greenice@konkuk.ac.kr (E.H.); huynhkimhung82@gmail.com (K.-H.H.); susia45@gmail.com (J.K.); bjun@konkuk.ac.kr (B.-H.J.)

² Department of Chemical and Biological Engineering, Hanbat National University, Daejeon 34158, Korea; sanghunlee@hanbat.ac.kr

* Correspondence: phamricky@gmail.com; Tel.: +82-2-450-0521

Abstract: In this study, dense gold-assembled SiO₂ nanostructure (SiO₂@Au) was successfully developed using the Au seed-mediated growth. First, SiO₂ (150 nm) was prepared, modified by amino groups, and incubated by gold nanoparticles (ca. 3 nm Au metal nanoparticles (NPs)) to immobilize Au NPs to SiO₂ surface. Then, Au NPs were grown on the prepared SiO₂@Au seed by reducing chloroauric acid (HAuCl₄) by ascorbic acid (AA) in the presence of polyvinylpyrrolidone (PVP). The presence of bigger (ca. 20 nm) Au NPs on the SiO₂ surface was confirmed by transmittance electronic microscopy (TEM) images, color changes to dark blue, and UV-vis spectra broadening in the range of 450 to 750 nm. The SiO₂@Au nanostructure showed several advantages compared to the hydrofluoric acid (HF)-treated SiO₂@Au, such as easy separation, surface modification stability by 11-mercaptoundecanoic acid (R-COOH), 11-mercapto-1-undecanol (R-OH), and 1-undecanethiol (R-CH₃), and a better peroxidase-like catalysis activity for 5,5'-Tetramethylbenzidine (TMB) and hydrogen peroxide (H₂O₂) reaction. The catalytic activity of SiO₂@Au was two times better than that of HF-treated SiO₂@Au. When SiO₂@Au nanostructure was used as a surface enhanced Raman scattering (SERS) substrate, the signal of 4-aminophenol (4-ATP) on the surface of SiO₂@Au was also stronger than that of HF-treated SiO₂@Au. This study provides a potential method for nanoparticle preparation which can be replaced for Au NPs in further research and development.

Keywords: gold nanostructure; dense gold-assembled silica nanostructures; local surface plasmon resonance; peroxidase-like catalysis; surface enhanced Raman scattering



Citation: Seong, B.; Bock, S.; Hahm, E.; Huynh, K.-H.; Kim, J.; Lee, S.H.; Pham, X.-H.; Jun, B.-H. Synthesis of Densely Immobilized Gold-Assembled Silica Nanostructures. *Int. J. Mol. Sci.* **2021**, *22*, 2543. <https://doi.org/10.3390/ijms22052543>

Academic Editor: Ana María Díez-Pascual

Received: 10 February 2021

Accepted: 26 February 2021

Published: 3 March 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Metal nanoparticles (NPs) have attracted much attention due to their quantum size effect resulting in unique physical and chemical properties [1]. Among metal nanostructures, gold (Au) nanostructures are probably the most remarkable metal materials due to their particular features: ease of synthesis manipulation; large surface-to-volume ratio, enabling precise control over the particle's physicochemical properties [2]; strong binding affinity to thiols, disulfides, and amines [3]; unique tunable optical properties due to their size- and shape-dependent biophysical and distinct optoelectronic properties [4–7]; excellent biocompatibility, chemical inertness, and low toxicity [8,9]. As a result, Au nanostructures have been outstanding tools for a variety of potential applications in catalysis [10–15] and biosensing [16–22].

Based on their dimensions, Au nanostructures are classified as one-dimensional (1D) nanostructures (nanorods, nanowires, nanotubes, nanobelts); two-dimensional (2D) nanostructures (nanoplates such as nanoparticles, stars, pentagons, squares/rectangles, dimpled nanoplates, hexagons, truncated triangles); and three-dimensional (3D) nanostructures (nanotadpoles, nanodumbbells, nanopods, nanostars, and nanodendrites) [23]. Even

though spherical or quasi-spherical Au NPs have received the most attention because of the ease of synthesis manipulation, anisotropically Au nanostructures exhibit unique physical and optical properties, such as low percolation threshold and surface plasmon resonance (SPR) [24]. In addition, the optical properties of anisotropic Au nanostructure such as the hollow nanoshell, and nanorods can be tunable with their shape in the visible region and in the near infrared (NIR) region. This property has given rise to the opportunity for using anisotropic Au nanostructures as composite therapeutic agents in clinical medical applications such as diagnostics and therapy [25]. However, the synthesis of higher-dimensional Au nanostructures usually requires specific templates for guiding anisotropic growth [26]. Many templates have been utilized, including biomolecules (DNA), polymers, surfactants, inorganic nanowires, and lithographical patterns [27–31]. However, the synthesis of anisotropic Au nanostructures with precise control of morphology remains a great challenge.

Recently, our group reported Au–Ag alloys assembled SiO₂ (SiO₂@Au–Ag) nanostructure as a strong and reliable surface enhanced Raman scattering (SERS) probe using SiO₂ as a template [32–35]. SiO₂ NPs are inert and their sizes are easily controllable [32]. Furthermore, the density of Au NPs, nanogaps, size and shape of nanostructure can be controlled by SiO₂ template, generating a homogenous SERS substrate [36]. The density of the Au NPs and the gaps between two Au NPs on the surface of SiO₂ can be tunable and provide a stronger localized surface plasmon resonance (LSPR) property. As a result, SiO₂@Au–Ag can be developed as high-strength and reliable SERS substrates for the enzyme-linked immunosorbent assay (ELISA) [37,38], which is an internal SERS [34,35], pesticide detection standard [39], and provides detection of toxic chemical compound in pharmaceuticals [40]. However, the cellular toxicity and easy oxidation of intrinsic Ag of our nanostructure limits their application in vivo [41,42]. Even though some research groups have investigated the attachment of Au NP on the surface of SiO₂, the particle density on the SiO₂ surface was nearly low or non-uniform [43–48]. Therefore, we used the combination of SiO₂ core template and seed-growth methods in this study to develop a new Au-based SiO₂ nanostructure with dense Au density on the surface of SiO₂. This study provides a potential method for preparation of nanoparticles including noble metal that can be replaced for spherical Au NPs, which supports further research and development.

2. Results and Discussion

To prepare SiO₂ nanostructure (SiO₂@Au), silica NPs (~150 nm) were first functionalized by 3-aminopropyltriethoxysilane (APTS) to prepare aminated silica NPs, as shown in Figure 1a. Simultaneously, colloidal Au NPs (3 nm) were synthesized by tetrakis(hydroxymethyl)phosphonium chloride (THPC) and incubated with the aminated silica NPs to prepare Au NPs seed embedded with SiO₂ (SiO₂@Au seed NPs), according to the method reported by Pham et al. [32–35,39]. Subsequently, the Au NPs on the surface of SiO₂@Au seed were grown by reducing a gold precursor (HAuCl₄) in the presence of ascorbic acid (AA) and polyvinylpyrrolidone (PVP) as a stabilizer and structure-directing agent under mild reducing conditions [32]. The gold ions reduced by AA were selectively grown onto SiO₂@Au seed to increase the size of Au on the surface of SiO₂ NPs to generate SiO₂@Au.

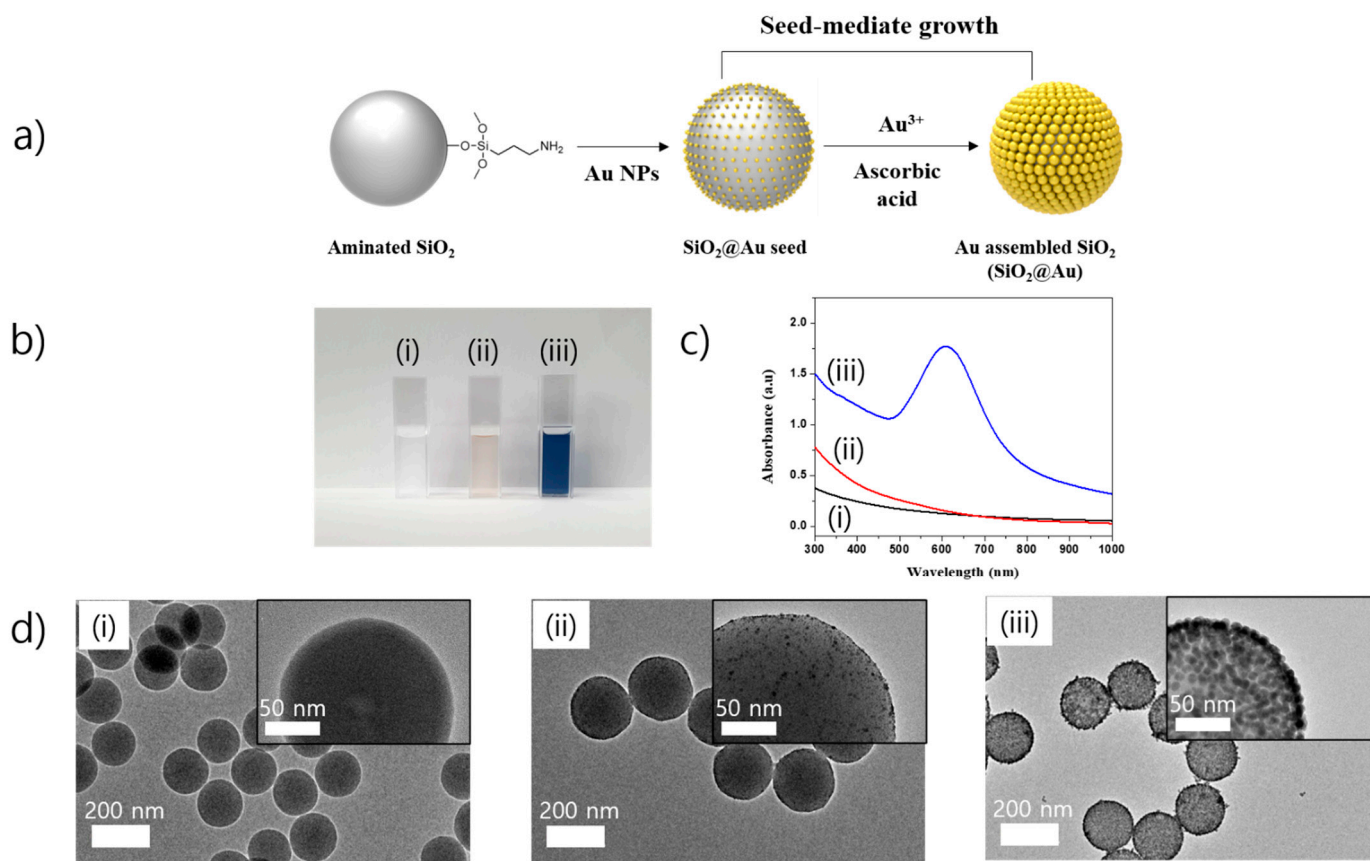


Figure 1. (a) Illustration of preparation of gold embedded silica nanostructure by using the combination of seed-growth method and reduction of chloroauric acid by ascorbic acid and polyvinylpyrrolidone. (b) Optical images, (c) UV-Vis spectra, and (d) Transmittance electronic microscopy (TEM) images of SiO₂-based nanostructures (50 µg/mL): (i) SiO₂ metal nanoparticles (NPs); (ii) SiO₂ nanostructure (SiO₂@Au) seed and (iii) SiO₂@Au nanostructure.

2.1. Synthesis of SiO₂@Au Nanostructure

We investigated the characteristics of SiO₂@Au synthesized by reduction of HAuCl₄ and AA in the presence of PVP. Figure 1b shows the optical images of nanostructures in our research. SiO₂ NPs with ca. 150 nm in diameter are shown in a transparent solution in Figure 1b(i). When Au NPs were coated and grown on the surface of aminated SiO₂, the color of the solution changed to light brown, as shown in Figure 1b(ii). Then, 2, 5, and 10 mg of SiO₂@NH₂ were incubated in 10 mL Au NPs suspension overnight (Figure S1a). The surface of SiO₂ NPs was decorated with many small Au NPs to generate the SiO₂@Au seed (Figure 1b(ii) and Figure S1a). The Au NPs density on the SiO₂ surface became inverted with the amount of SiO₂ used. Then, 1 mg of SiO₂@Au seed was used for a growth of Au NPs on the SiO₂ in Figure S1b–d. The sizes of Au NPs seed on the SiO₂@Au surface increased with the Au³⁺ concentration in the range of 100 to 300 µM at all 2, 5, and 10 mg SiO₂@NH₂ (Figure S1b–d). The ultraviolet-visible (UV-Vis) spectra of obtained SiO₂@Au nanostructures synthesized at different concentrations of Au³⁺ were investigated in Figure S1e. The optical property of SiO₂@Au was slightly tunable by adjusting the Au³⁺ concentration in the range of 100 to 300 µM. At 100 µM Au³⁺, the maximum absorbance peaks of obtained SiO₂@Au synthesized by 2, 5, and 10 mg SiO₂@NH₂ were observed at 523, 522, and 521 nm, respectively. Whereas these peaks were red-shifted to 547, 543, and 541 nm at 200 µM Au³⁺ and 571, 561, and 572 nm at 300 µM Au³⁺, respectively. These results were consistent with the Mie's theory which states that an increase of particle size leads the electric surface charge density and shifts the plasmon absorption band to longer wavelengths [23]. However, it is difficult to obtain the different desired SiO₂@Au nanostructures

as expected when using 1 mg SiO₂@Au seed. The final size of Au NPs is dependent on the number of seeds and total concentration of Au³⁺ in the growth solution [49]. In our study, the Au³⁺ concentration in the growth solution was fixed at 100, 200, and 300 μM. Therefore, the final size of Au NPs on the surface of SiO₂ depended on the number of SiO₂@Au seeds (or SiO₂@Au seed amount). Indeed, 1 mg of SiO₂@Au seed was used for Au NPs growth process in our study, which resulted in Au NPs on the SiO₂ surface not being able to grow larger as seen in the TEM images and UV-Vis spectra (Figure S1). To obtain a monodisperse SiO₂@Au nanostructure, the SiO₂@Au seed was therefore decreased to 0.2 mg for the growth of Au NPs on the SiO₂ NPs in the next study. When 0.2 mg SiO₂@Au seed was used as seed, the color of the obtained SiO₂@Au suspension obviously changed from light brown to dark blue when the Au³⁺ concentration increased to 150 μM, as shown in Figure 1b(iii). According to the literature, Au NPs exhibit color and LSPR bands in the visible light that are dependent on size and shape [49–52]. Any changes in the absorbance or wavelength of the LSPR band provide a difference in size, shape, and aggregation state. This result indicated that the growth of Au NPs on the SiO₂ surface was well performed and the absorbance, transmission, and reflection of light passed through the SiO₂@Au suspension were also different at various synthesis conditions of SiO₂@Au nanostructures. The UV-Vis spectra of the SiO₂@Au nanostructures synthesized at 150 μM Au³⁺ were investigated in Figure 1c to show additional characteristics of SiO₂@Au nanostructures. The suspension of SiO₂ and SiO₂@Au seed did not show any absorbance in the range of 300–1000 nm because of its small Au NPs on the SiO₂ surface [37,53]. Whereas the suspension of SiO₂@Au synthesized at 150 μM Au³⁺ showed a broadband in the range of 450 to 750 nm with the maximum peak at 615 nm. Furthermore, the absorbance intensity of SiO₂@Au nanostructure solution was sharply increased. Consistent with the UV-Vis spectra, the transmittance electronic microscopy (TEM) images of SiO₂@Au nanostructures also showed that the size of Au NPs on the surface of SiO₂@Au increased dramatically comparing to the SiO₂@Au seed at 150 μM Au³⁺ (Figure 1d), which led to the sharp broadening of LSPR bands of UV-Vis spectrum. While the average size of SiO₂@Au seed was 200 ± 9 nm (*n* = 170, Figure S2a), the average size of SiO₂@Au was 211 ± 7 nm (*n* = 172, Figure S2b). Consequently, the red-shifted and broadened absorbance properties of SiO₂@Au nanostructures indicated the growth of Au NPs to bigger size on the SiO₂ surface [23,49]. Therefore, the SiO₂@Au nanostructure synthesized at 150 μM Au³⁺ was used for further study.

2.2. Characteristics of SiO₂@Au Nanostructure

As shown in the previous study, the presence of SiO₂ template in SiO₂-based nanostructure facilitates the control of density, size, shape, and nanogap of metal NPs on the SiO₂ surface [36]. It is believed that SiO₂ template also facilitates the separation of SiO₂-based nanostructures in reaction compared to those without SiO₂ template. In our study, SiO₂@Au nanostructure was first obtained from the growth reaction at 150 μM Au³⁺, then was incubated in 24% hydrofluoric acid (HF) to etch the SiO₂ core template as seen in Figure 2a. Figure 2b shows an obvious color change from blue to pink when the SiO₂ template was etched by HF. The TEM images of SiO₂@Au nanostructure with HF treatment were collected. Figure 1d(iii) shows the morphology of SiO₂@Au without HF treatment with the average size of 211 ± 7 nm (Figure S2b). Many Au NPs presented on the surface of SiO₂ template. However, the nanostructure with HF treatment showed many small Au NPs with different shape (Figure 2c(ii)). The average size of Au NPs was 7.7 ± 1.5 nm (*n* = 172, Figure S2c). This meant that SiO₂ core was completely etched and disappeared in SiO₂@Au nanostructure and remained as small Au NPs. The optical properties of SiO₂@Au with and without HF treatment (Au) were significantly different, as shown in Figure 2d. Both the maximum peak position and absorbance intensity of SiO₂@Au with HF treatment (Au) decreased dramatically. The maximum peak position of SiO₂@Au at 615 nm was blue-shifted to 546 nm when SiO₂ core was etched. The difference in the LSPR wavelength and absorbance intensity of Au (SiO₂@Au with HF treatment) was due to the difference in aggregation state of nanostructure that was caused by the disappearance of SiO₂ core

template [49]. To confirm the aggregation state of SiO₂@Au with and without SiO₂ core, the nanostructures were redispersed in 1 M NaCl. The maximum LSPR peak of SiO₂@Au in NaCl showed a 10 nm red-shift compared to it in distilled water. Whereas SiO₂@Au treated with HF was 22 nm red-shifted in 1 M NaCl (Figure S3). This indicates that SiO₂@Au treated with HF was more easily aggregated than SiO₂@Au without HF treatment.

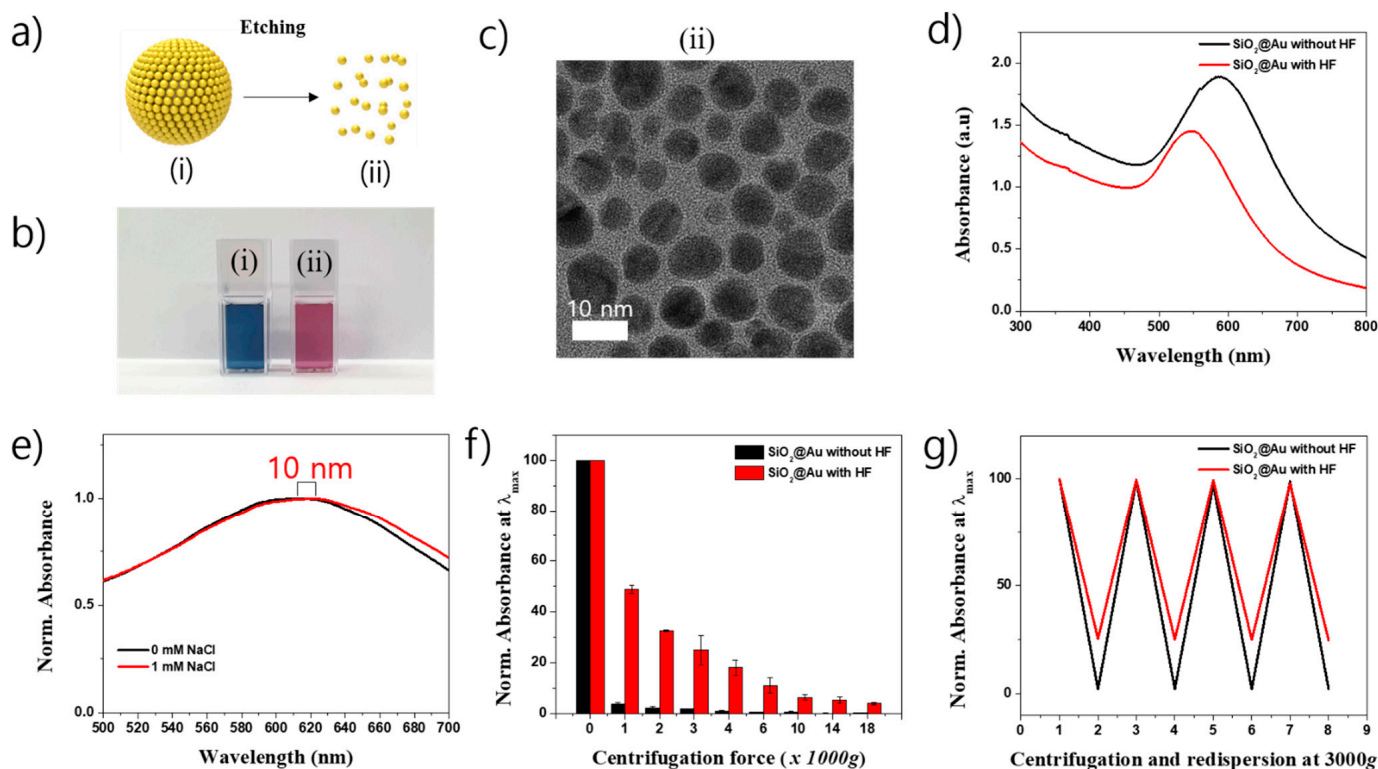


Figure 2. (a) Etching, (b) optical images, (c) TEM images, and (d) UV-Vis spectra of SiO₂@Au nanostructures (50 μ g/mL): (i) SiO₂@Au without hydrofluoric acid (HF) treatment and (ii) SiO₂@Au treated with HF. (e) Red-shifting of UV-Vis spectra, (f) centrifugation speed, and (g) centrifugation (2,4,6,8 in x-axis) and redispersion (3,5,7 in x-axis) of SiO₂@Au synthesized at 150 M Au³⁺ with and without HF treatment.

The separation and redispersion of SiO₂@Au nanostructures was also observed in Figure 2f,g and Figure S3b–d. The SiO₂@Au with and without HF treatment were centrifuged at various centrifugation speeds in the range of 1000 to 18,000 \times g. The SiO₂@Au was almost settled down to a bottle of microtube at a centrifugal speed of 3000 \times g and left a supernatant which showed a low absorbance. Meanwhile, the absorbance intensity of the supernatant of SiO₂@Au treated with HF decreased gradually to 50% at 1000 \times g, 25% at 3000 \times g and 6.3% at 10,000 \times g. From these results, we concluded that SiO₂@Au was easily centrifuged and separated from the reaction solution. In the next experiment, we carried out centrifuge and successively redispersed the SiO₂@Au at the centrifugation speed of 3000 \times g. As seen in Figure 2g and Figure S3c, both SiO₂@Au with and without HF treatment was easily redispersed in phosphate buffer saline containing 0.1% Tween 20 (PBST). Therefore, we concluded that the SiO₂@Au nanostructures were easily separated and redispersed in PBST at mild conditions. The use of SiO₂ as the core template opens up a new opportunity for easy preparation and separation of the nanostructure.

2.3. Surface Modification of SiO₂@Au Nanostructure

According to the literature, the electric field of the incident light induces polarization and excites the free conduction electron on the surface of the nanostructure to generate the LSPR spectrum. Therefore, any variation in the LSPR absorbance intensity or wavelength represents the difference in particle size, shape, aggregation state, as well as free

electron cloud on the surface of the nanostructure [49,52]. We used the UV-Vis spectrum to observe the optical properties of the SiO₂@Au nanostructures modified by three kinds of ligand (11-mercaptoundecanoic acid (R-COOH), 11-mercapto-1-undecanol (R-OH), and 1-undecanethiol (R-CH₃)). Similarly, SiO₂@Au which removed the SiO₂ core by HF treatment was also used as a control sample to compare its optical properties to SiO₂@Au nanostructures. The results are shown in Figure S4 and Figure 3. When R-COOH, R-CH₃ and R-OH ligands were modified on the SiO₂@Au surface, the absorbance intensities of all prepared SiO₂@Au nanostructures obviously decreased (Figure S4). In addition, the extinction peak positions of SiO₂@Au nanostructures modified by ligands were all blue-shifts (Figure 3a(i)). Maximum peak of SiO₂@Au nanostructures modified by R-COOH ligand dramatically decreased 50 nm from 615 nm to 557 nm. Similarly, the SiO₂@Au nanostructures modified by R-OH ligand also showed a 75 nm blue-shift to 540 nm and those modified by R-CH₃ showed a 42 nm blue-shift to 573 nm, respectively (Figure 3b(i)). Meanwhile, the HF-treated SiO₂@Au modified by R-COOH, R-OH, and R-CH₃ ligands exhibited different behaviors (Figure 3a(ii)). Even though the extinction peak positions of the HF-treated SiO₂@Au modified by R-COOH and R-OH also showed decreases in LSPR wavelength like those of SiO₂@Au, the blue-shift differences in wavelength were small, just 15 and 18 nm for R-COOH and R-OH ligands, respectively (Figure 3b(ii)). Blue-shift decreases in wavelength of SiO₂@Au nanostructures modified by R-COOH and R-OH are due to hydrophilic properties of OH and COOH groups of the modified nanostructure regardless of HF treatment. Interestingly, the HF-treated SiO₂@Au modified by R-CH₃ showed an opposite behavior compared to that of SiO₂@Au nanostructure. While the SiO₂@Au nanostructure modified by R-CH₃ showed a blue-shift from 615 nm to 573 nm, the HF-treated SiO₂@Au nanostructure modified by R-CH₃ showed a red-shift from 546 to 561 nm. This indicates the presence of aggregation of the HF-treated SiO₂@Au nanostructure modified by R-CH₃. In contrast, presence of R-CH₃ groups on the surface of SiO₂@Au nanostructures showed little effect on the optical property of SiO₂@Au nanostructure because of the big size of SiO₂@Au (~211 nm) compared to HF-treated SiO₂@Au (~8 nm). This is because of the intrinsic hydrophobic property of CH₃ group strongly present on the surface of small Au, as shown in Figure 2c (~8 nm). The result was confirmed by replacing R-CH₃ group by 4-aminophenol (4-ATP) with a benzene ring. Figure S5a,b show that the extinction bands of both SiO₂@Au with and without HF treatment were red-shifted because of the intrinsic hydrophobic aromatic ring of 4-ATP. From these results, we once again concluded that the SiO₂@Au nanostructure possessed a stable surface for ligand modification compared to that of Au NPs without the SiO₂ core.

2.4. SERS and Peroxidase-Like Activity of SiO₂@Au Nanostructure

The application of the SiO₂@Au nanostructure was briefly performed in our study. In SERS measurement, we used 4-ATP as a Raman reporter to observe the SERS properties of SiO₂@Au with and without HF treatment using a 780 nm diode pump solid-state laser. Figure 4a shows the SERS spectra of SiO₂@Au and HF-treated SiO₂@Au before and after incubating in 1 mM 4-ATP solution. Although several Raman signals were shown in 4-ATP untreated condition, after incubating in 1 mM 4-ATP solution, typical bands of 4-ATP were clearly obtained for both SiO₂@Au and HF-treated SiO₂@Au due to the electromagnetic enhancement of the decorated Au NPs. Dominant and distinct bands were seen at 1081 cm⁻¹ which were assigned to C-H in-plane bending vibration. The band at 1592 cm⁻¹ and 391 cm⁻¹ were attributed to C-C ring and C-S stretching, respectively [32]. Interestingly, the SERS bands at 391, 1081, and 1592 cm⁻¹ of SiO₂@Au were stronger than those of HF-treated SiO₂@Au due to the hot spot generated by Au NPs on the surface of SiO₂@Au. These results demonstrate the importance of pre-seeding Au NPs to ensure the growth of Au NPs onto the SiO₂ surface to generate various hot spots, enhance the electromagnetic field on or near the surface of SiO₂@Au, and amplify the SERS signal of SiO₂@Au compared to HF-treated SiO₂@Au without the SiO₂ core template.

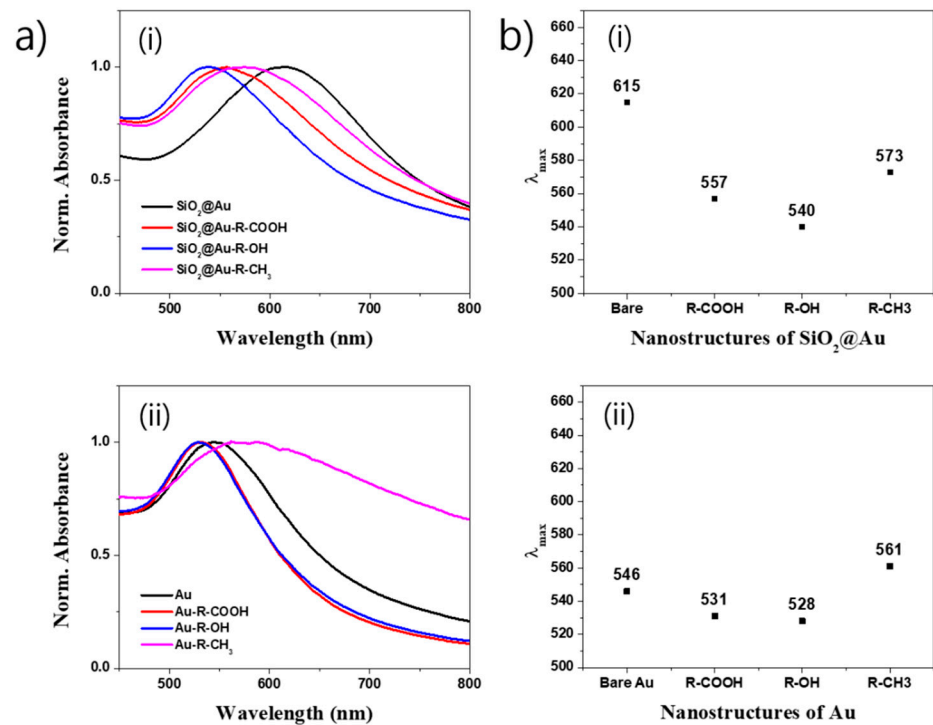


Figure 3. Effect of surface modification of 1-undecanethiol (R-CH₃), 11-mercaptoundecanoic acid (R-COOH), and 11-mercapto-1-undecanol (R-OH) ligands on (a) the UV-Vis spectra and (b) plot of extinction maximum band position of SiO₂@Au nanostructures synthesized at 150 M Au³⁺.

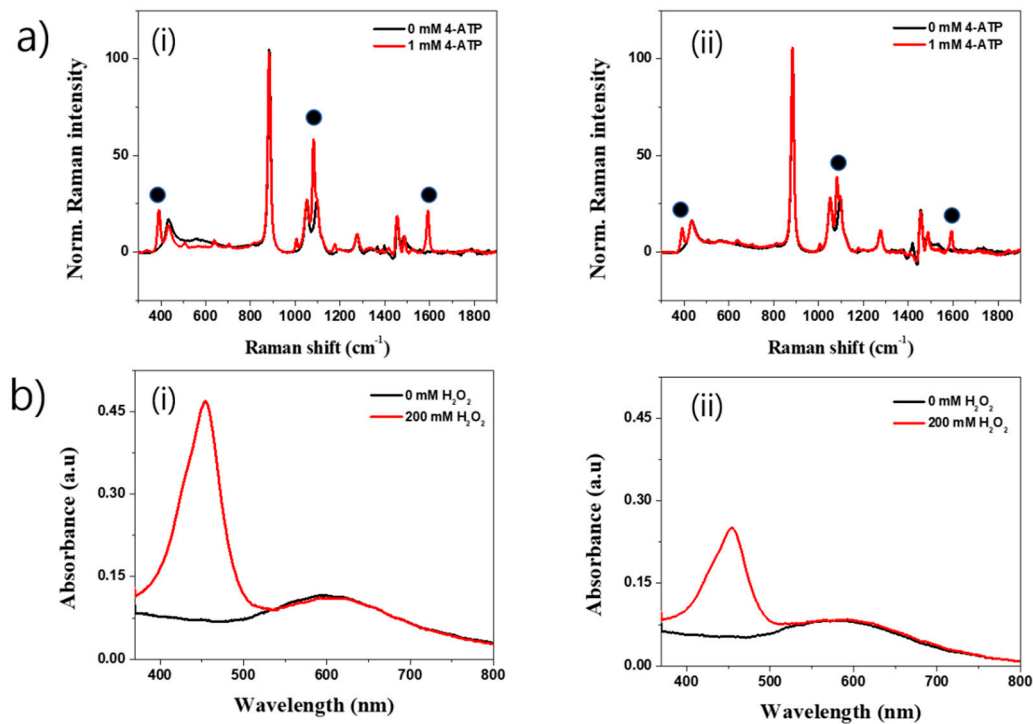


Figure 4. (a) Surface enhanced Raman scattering of SiO₂@Au nanostructures suspended in ethanol solution and (b) peroxidase-like catalytic activity of SiO₂@Au nanostructures synthesized at 150 M Au³⁺ (i) without and (ii) with HF treatment.

In addition, Au NPs-based nanostructures showed peroxidase-like catalytic activities in previous reports [54–56]. In this study, we used the SiO₂@Au nanostructure to catalyze for 5,5'-Tetramethylbenzidine (TMB) and hydrogen peroxide (H₂O₂) reaction. Figure 4b shows the UV-Vis spectra of the SiO₂@Au with and without HF treatment. In the presence of H₂O₂ and under the catalysis of nanostructures, TMB was oxidized to TMB⁺, followed by conversion to TMB²⁺ in sulfuric acid (H₂SO₄) condition as mentioned in Figure S4c. The presence of TMB²⁺ in reaction will show a clear absorbance peak at ~450 nm. Indeed, TMB was successfully converted to TMB²⁺ under the catalysis of the SiO₂@Au by the presence of a clear and strong absorbance band at 453 nm (Figure 4b(i)). Similarly, the HF-treated SiO₂@Au nanostructures could also convert TMB to TMB²⁺, as shown in in Figure 4b(ii). However, the absorbance intensity at 453 nm of the SiO₂@Au was two times higher than that of the SiO₂@Au with HF treatment. This is perhaps because of the easy aggregation of the HF-treated SiO₂@Au nanostructure or because the surface of Au NPs was varied in the HF treatment as mentioned in a previous report [54].

3. Materials and Methods

3.1. Chemicals and Reagents

Tetraethylorthosilicate (TEOS), 3-aminopropyltriethoxysilane (APTS), tetrakis (hydroxymethyl)phosphonium chloride (THPC), polyvinylpyrrolidone (PVP), phosphate buffer saline (PBS), phosphate buffer saline containing 0.1% Tween 20 (PBST), ascorbic acid (AA), chloroauric acid (HAuCl₄), hydrofluoric acid (HF), 4-aminothiophenol (4-ATP), 4-mercaptobenzoic acid (4-MBA), 1-undecanethiol (R-CH₃), 11-mercaptoundecanoic acid (R-COOH), 11-mercapto-1-undecanol (R-OH), hydrogen peroxide (H₂O₂), and 5,5'-Tetramethylbenzidine (TMB) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Ethanol (EtOH), sulfuric acid (H₂SO₄), buffer pH 4.0, and aqueous ammonium hydroxide (NH₄OH, 27%) were purchased from Daejung (Sihung, Gyeonggi-do, South Korea); and ultrapure water (18.2 MΩ cm) was produced by a Millipore water purification system (EXL water purification; Vivagen Co., Seongnam, Gyeonggi-do, Korea).

3.2. Preparation of SiO₂@Au NPs

The SiO₂@Au seed was prepared as in the previous report [32,33]. The SiO₂@Au NPs were prepared by incubating Au NPs suspension (10 mL) with aminated silica NPs (2 mg) overnight. The colloids were carefully centrifuged and washed thoroughly using EtOH. The NPs were then redispersed in 2.0 mL of 1 mg/mL PVP of obtained 1 mg/mL SiO₂@Au seed.

The SiO₂@Au NPs were carefully prepared via the reduction and deposition of Au using AA onto SiO₂@Au seed in PVP. Moreover, 200 μL of SiO₂@Au (1.0 mg/mL) was briefly dispersed in 9.8 mL of water that contained 10 mg of PVP under stirring. Thereafter, 20 μL of HAuCl₄ (10 mM) was added to the suspension, followed by the addition of 40 μL of AA (10 mM). The suspension was incubated for 15 min to completely reduce the Au³⁺ ions to Au⁰. By repeating the reduction steps, the final Au³⁺ concentration was controlled to be 150 μM. The SiO₂@Au NPs were obtained by the centrifugation of the suspension at 8500 rpm for 15 min, then the NPs were washed thoroughly using EtOH to remove any excess reagent. The SiO₂@Au NPs were then redispersed in 1 mL of absolute EtOH to obtain a 200 μg/mL SiO₂@Au NP suspension.

3.3. Etching of Silica Core of SiO₂@Au NPs

In order to etch the silica core of SiO₂@Au, 500 μL of SiO₂@Au (100 μg/mL) in PBS solution which contained 0.1% Tween 20 (PBST), was added in 500 μL of 48% HF, followed by incubation for 24 h and centrifugation for 15 min at 17,000 rpm to obtain the Au NPs suspension. The prepared Au NPs were washed thoroughly using PBST to remove any excess reagent. The Au NPs were then redispersed in 1 mL of PBST, to obtain a 50 μg/mL Au NPs suspension.

3.4. Separation and Dispersion of SiO₂@Au

First, 50 µg/mL SiO₂@Au suspension was prepared in PBST and centrifuged for 10 min at various centrifugation speeds in the range of 1000 to 18,000 × *g*. The supernatant was collected and the UV-Vis spectroscopy measured in the range of 300–800 nm. The pellet was redispersed by sonication. The centrifugation and redispersion were repeated until finishing. Similarly, the separation and redispersion of 50 µg/mL Au suspension was carried out as SiO₂@Au suspension.

3.5. Surface Modification of SiO₂@Au

SiO₂@Au was incubated with 1 mM ligands, such as 1-undecanethiol (R-CH₃), 11-mercaptoundecanoic acid (R-COOH), and 11-mercapto-1-undecanol (R-OH) for 6 h at room temperature to modify the surface of Au NPs by CH₃, COOH, and OH groups, respectively. Briefly, 50 µg SiO₂@Au suspension was prepared in 500 µL EtOH. Then, 2 mM ligand solutions (500 µL) were added into SiO₂@Au suspension and incubated for 6 h under stirring. The nanostructure was collected at 17,000 rpm for 15 min and the obtained pellet was redispersed in 1000 µL PBST to obtain an SiO₂@Au-ligand suspension. Similarly, the surface modification of Au NPs was carried out as SiO₂@Au NPs.

3.6. Peroxidase-like Catalytic Activity of SiO₂@Au

In order to obtain the peroxidase-like catalysis of nanostructures, TMB was dissolved in EtOH to obtain a stock solution of 10 mM TMB. Similarly, a stock solution of 2 M H₂O₂ was freshly prepared. Thereafter, 100 µL of 10 mM TMB, 700 µL of buffer pH 4.0, 100 µL of 2 M H₂O₂, and 100 µL of nanostructure (200 µg/mL) were incubated for 30 min at 25 °C in vortex mixer. The mixture was added to 500 µL of 1 M H₂SO₄ to stop the reaction and incubated for 10 min for color development. The suspension was measured by UV-Vis spectroscopy at 453 nm.

3.7. Instrument

Transmission electron microscope images of the sample were captured by using Libra 120 field-emission transmission electron microscope (Carl Zeiss, Oberkochen, Baden-Württemberg, Germany) with a maximum accelerated voltage of 120 kV and JEM-F200 multi-purpose electron microscope (JEOL, Akishima, Tokyo, Japan) with a maximum accelerated voltage of 200 kV. Optical properties of the sample were observed by an Optizen POP UV/Vis spectrometer (Mecasys, Seoul, Korea). Centrifugation of the sample was performed by using a Microcentrifuge 1730R (LaboGene, Lyngen, Denmark). The Raman signal of nanostructure suspension in the capillary tube was recorded by a micro-Raman system (LabRam 300, JY-Horiba, Tokyo, Japan) equipped with an optical microscope (BX41, Olympus, Tokyo, Japan). The SERS signals were collected with a back-scattering geometry using a ×10 objective lens (0.90 NA, Olympus) and with a spectrometer using a thermoelectrically cooled CCD detector. A 780 nm diode-pumped solid-state laser (CL780-100-S, CrystaLaser, Reno, NV, USA) was used as a photoexcitation source with 50 mW of laser power shot at the sample.

4. Conclusions

Dense Au NPs on the surface of SiO₂@Au was successfully developed using the combination of Au seed-mediated growth in this study. The growth of Au NPs on the surface of SiO₂ was confirmed by the color changes from light brown to dark blue, TEM images, and broad bands in the UV-vis spectra in the range of 450 nm to 750 nm with the maximum peak at 615 nm, which indicated bigger size of Au NPs on SiO₂ surface when the Au³⁺ concentration increased to 150 µM. In addition, the prepared SiO₂@Au nanostructure showed several advantages compared to the SiO₂@Au treated by HF solution, such as easy separation from the solution at 3000 × *g* for 10 min and stability for surface modification by R-COOH, R-OH, and R-CH₃. Furthermore, the SiO₂@Au nanostructure was used as a SERS substrate for 4-ATP detection and a peroxidase-like catalyst material for TMB and

H₂O₂ reactions. As a result, the SiO₂@Au showed a stronger SERS signal of SiO₂@Au compared to HF-treated SiO₂@Au. Similarly, the catalytic activity of SiO₂@Au was two times better than that of the SiO₂@Au with HF treatment. This study provides a potential method for nanoparticle preparation that can be replaced for Au NPs, which supports further research and development.

Supplementary Materials: The following are available online at <https://www.mdpi.com/1422-0067/22/5/2543/s1>, Figure S1: TEM images and UV-Vis spectra of SiO₂@Au nanostructures synthesized at (i) 2 mg (ii) 5 mg, and (iii) 10 mg SiO₂@NH₂ in (a) 0, (b) 100, (c) 200, and (d) 300 mM Au³⁺ concentration; Figure S2: Histogram of nanoparticle size: (a) SiO₂@Au seed ($n = 170$), (b) SiO₂@Au synthesized at 150 μM Au³⁺ ($n = 172$), and HF treated-SiO₂@Au ($n = 172$). Figure S3. (a) Red-shifting of UV-Vis spectra of SiO₂@Au treated HF, (b) effect of centrifugation speed, (c) UV-Vis spectra of SiO₂@Au and HF-treated SiO₂@Au, (d) optical absorbance plot of SiO₂@Au nanostructures synthesized at 150 mM Au³⁺ with and without HF treatment after centrifugation (2, 4, 6, 8 in x-axis) and redispersion (3, 5, 7 in x-axis); Figure S4. Effect of surface modification of (a) 1-undecanethiol (R-CH₃), 11-mercaptoundecanoic acid (R-COOH), and 11-mercapto-1-undecanol (R-OH) ligands and (b) 4-ATP on the UV-Vis spectra of (i) SiO₂@Au nanostructures and HF-treated SiO₂@Au synthesized at 150 mM Au³⁺; Figure S5. Normalized absorbance spectra of (a) SiO₂@Au nanostructures and (b) HF-treated SiO₂@Au synthesized at 150 mM Au³⁺. (c) Mechanism of peroxidase-like catalytic activity of SiO₂@Au nanostructures.

Author Contributions: Conceptualization, X.-H.P. and B.-H.J.; methodology, B.S., S.B. and X.-H.P.; investigation, B.S.; formal analysis, K.-H.H. and E.H.; software, S.H.L.; writing—original draft preparation, B.S., X.-H.P.; writing—review and editing, J.K. and B.-H.J.; supervisor, B.-H.J. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the KU Research Professor Program of Konkuk University and funded by the Ministry of Science and ITC (NRF-2019R1G1A1006488) and by Ministry of Education (NRF-2018R1D1A1B07045708).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article or supplementary material.

Acknowledgments: The authors are grateful for the financial support from the NRF of Korea. Further, the author gives thanks for the financial support by Konkuk University.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Zhao, P.; Li, N.; Astruc, D. State of the art in gold nanoparticle synthesis. *Coord. Chem. Rev.* **2013**, *257*, 638–665. [[CrossRef](#)]
2. Zhang, Y.; Chu, W.; Foroushani, A.D.; Wang, H.; Li, D.; Liu, J.; Barrow, C.J.; Wang, X.; Yang, W. New gold nanostructures for sensor applications: A review. *Materials* **2014**, *7*, 5169–5201. [[CrossRef](#)]
3. Zhou, Y.; Wang, C.Y.; Zhu, Y.R.; Chen, Z.Y. A novel ultraviolet irradiation technique for shape-controlled synthesis of gold nanoparticles at room temperature. *Chem. Mater.* **1999**, *11*, 2310–2312. [[CrossRef](#)]
4. Zhang, Y.; Qian, J.; Wang, D.; Wang, Y.; He, S. Multifunctional gold nanorods with ultrahigh stability and tunability for in vivo fluorescence imaging, SERS detection, and photodynamic therapy. *Angew. Chem. Int. Ed.* **2013**, *52*, 1148–1151. [[CrossRef](#)] [[PubMed](#)]
5. Zhang, Z.; Wang, J.; Nie, X.; Wen, T.; Ji, Y.; Wu, X.; Zhao, Y.; Chen, C. Near infrared laser-induced targeted cancer therapy using thermoresponsive polymer encapsulated gold nanorods. *J. Am. Chem. Soc.* **2014**, *136*, 7317–7326. [[CrossRef](#)]
6. Jain, P.K.; Huang, X.; El-Sayed, I.H.; El-Sayed, M.A. Noble Metals on the Nanoscale: Optical and Photothermal Properties and Some Applications in Imaging, Sensing, Biology, and Medicine. *Acc. Chem. Res.* **2008**, *41*, 1578–1586. [[CrossRef](#)]
7. Yu, Y.Y.; Chang, S.-S.; Lee, C.-L.; Wang, C.R.C. Gold Nanorods: Electrochemical Synthesis and Optical Properties. *J. Phys. Chem. B* **1997**, *101*, 6661–6664. [[CrossRef](#)]
8. Yeh, Y.C.; Creran, B.; Rotello, V.M. Gold nanoparticles: Preparation, properties, and applications in bionanotechnology. *Nanoscale* **2012**, *4*, 1871–1880. [[CrossRef](#)] [[PubMed](#)]
9. Elahi, N.; Kamali, M.; Baghersad, M.H. Recent biomedical applications of gold nanoparticles: A review. *Talanta* **2018**, *184*, 537–556. [[CrossRef](#)]

10. Dimitratos, N.; Lopez-Sanchez, J.A.; Hutchings, G.J. Selective liquid phase oxidation with supported metal nanoparticles. *Chem. Sci.* **2012**, *3*, 20–44. [[CrossRef](#)]
11. Min, B.K.; Friend, C.M. Heterogeneous gold-based catalysis for green chemistry: Low-temperature CO oxidation and propene oxidation. *Chem. Rev.* **2007**, *107*, 2709–2724. [[CrossRef](#)]
12. Bond, G.C.; Thompson, D.T. Catalysis by Gold. *Catal. Rev.* **1999**, *41*, 319–388. [[CrossRef](#)]
13. Chen, M.; Goodman, D.W. Catalytically active gold: From nanoparticles to ultrathin films. *Acc. Chem. Res.* **2006**, *39*, 739–746. [[CrossRef](#)] [[PubMed](#)]
14. Hashmi, A.S.K.; Hutchings, G.J. Gold Catalysis. *Angew. Chem. Int. Ed.* **2006**, *45*, 7896–7936. [[CrossRef](#)] [[PubMed](#)]
15. Haruta, M.; Daté, M. Advances in the catalysis of Au nanoparticles. *Appl. Catal. A Gen.* **2001**, *222*, 427–437. [[CrossRef](#)]
16. Jain, P.K.; El-Sayed, I.H.; El-Sayed, M.A. Au nanoparticles target cancer. *Nano Today* **2007**, *2*, 18–29. [[CrossRef](#)]
17. Sperling, R.A.; Gil, P.R.; Zhang, F.; Zanella, M.; Parak, W.J. Biological applications of gold nanoparticles. *Chem. Soc. Rev.* **2008**, *37*, 1896–1908. [[CrossRef](#)]
18. Schroeder, A.; Heller, D.A.; Winslow, M.M.; Dahlman, J.E.; Pratt, G.W.; Langer, R.; Jacks, T.; Anderson, D.G. Treating metastatic cancer with nanotechnology. *Nat. Rev. Cancer* **2012**, *12*, 39–50. [[CrossRef](#)]
19. Saha, K.; Agasti, S.S.; Kim, C.; Li, X.; Rotello, V.M. Gold nanoparticles in chemical and biological sensing. *Chem. Rev.* **2012**, *112*, 2739–2779. [[CrossRef](#)]
20. Bardhan, R.; Lal, S.; Joshi, A.; Halas, N.J. Theranostic nanoshells: From probe design to imaging and treatment of cancer. *Acc. Chem. Res.* **2011**, *44*, 936–946. [[CrossRef](#)]
21. Zhang, X. Gold Nanoparticles: Recent Advances in the Biomedical Applications. *Cell Biochem. Biophys.* **2015**, *72*, 771–775. [[CrossRef](#)]
22. Yuan, F.; Chen, H.; Xu, J.; Zhang, Y.; Wu, Y.; Wang, L. Aptamer-based luminescence energy transfer from near-infrared-to-near-infrared upconverting nanoparticles to gold nanorods and its application for the detection of thrombin. *Chem. Eur. J.* **2014**, *20*, 2888–2894. [[CrossRef](#)]
23. Ortiz-Castillo, J.E.; Gallo-Villanueva, R.C.; Madou, M.J.; Perez-Gonzalez, V.H. Anisotropic gold nanoparticles: A survey of recent synthetic methodologies. *Coord. Chem. Rev.* **2020**, *425*, 213489. [[CrossRef](#)]
24. Zeng, S.; Yong, K.-T.; Roy, I.; Dinh, X.-Q.; Yu, X.; Luan, F. A Review on Functionalized Gold Nanoparticles for Biosensing Applications. *Plasmonics* **2011**, *6*, 491–506. [[CrossRef](#)]
25. Zhang, W.; Wang, F.; Wang, Y.; Wang, J.; Yu, Y.; Guo, S.; Chen, R.; Zhou, D. pH and near-infrared light dual-stimuli responsive drug delivery using DNA-conjugated gold nanorods for effective treatment of multidrug resistant cancer cells. *J. Control. Release* **2016**, *232*, 9–19. [[CrossRef](#)]
26. Wang, J.; Dong, X.; Xu, R.; Li, S.; Chen, P.; Chan-Park, M.B. Template-free synthesis of large anisotropic gold nanostructures on reduced graphene oxide. *Nanoscale* **2012**, *4*, 3055–3059. [[CrossRef](#)]
27. Kim, H.J.; Roh, Y.; Hong, B. Selective Alignment of Gold Nanowires Synthesized With DNA as Template by Surface-Patterning Technique. *IEEE Trans. Nanotechnol.* **2010**, *9*, 254–257. [[CrossRef](#)]
28. Park, S.; Moon, S.C.; Chen, D.; Farris, R.J.; Russell, T.P. Preparation of 1 inch gold nanowires from PS-b-P4VP block copolymers. *J. Mater. Chem.* **2010**, *20*, 1198–1202. [[CrossRef](#)]
29. Murphy, C.J.; Sau, T.K.; Gole, A.; Orendorff, C.J. Surfactant-Directed Synthesis and Optical Properties of One-Dimensional Plasmonic Metallic Nanostructures. *MRS Bull.* **2005**, *30*, 349–355. [[CrossRef](#)]
30. Wong, T.C.; Li, C.P.; Zhang, R.Q.; Lee, S.T. Gold nanowires from silicon nanowire templates. *Appl. Phys. Lett.* **2004**, *84*, 407–409. [[CrossRef](#)]
31. Xiang, C.; Kung, S.-C.; Taggart, D.K.; Yang, F.; Thompson, M.A.; Güell, A.G.; Yang, Y.; Penner, R.M. Lithographically Patterned Nanowire Electrodeposition: A Method for Patterning Electrically Continuous Metal Nanowires on Dielectrics. *ACS Nano* **2008**, *2*, 1939–1949. [[CrossRef](#)] [[PubMed](#)]
32. Pham, X.-H.; Lee, M.; Shim, S.; Jeong, S.; Kim, H.-M.; Hahm, E.; Lee, S.H.; Lee, Y.-S.; Jeong, D.H.; Jun, B.-H. Highly sensitive and reliable SERS probes based on nanogap control of a Au-Ag alloy on silica nanoparticles. *RSC Adv.* **2017**, *7*, 7015–7021. [[CrossRef](#)]
33. Shim, S.; Pham, X.-H.; Cha, M.G.; Lee, Y.-S.; Jeong, D.H.; Jun, B.-H. Size effect of gold on Ag-coated Au nanoparticle-embedded silica nanospheres. *RSC Adv.* **2016**, *6*, 48644–48650. [[CrossRef](#)]
34. Pham, X.-H.; Hahm, E.; Kang, E.; Ha, Y.N.; Lee, S.H.; Rho, W.-Y.; Lee, Y.-S.; Jeong, D.H.; Jun, B.-H. Gold-silver bimetallic nanoparticles with a Raman labeling chemical assembled on silica nanoparticles as an internal-standard-containing nanoprobe. *J. Alloys Compd.* **2019**, *779*, 360–366. [[CrossRef](#)]
35. Pham, X.-H.; Hahm, E.; Kang, E.; Son, B.S.; Ha, Y.; Kim, H.-M.; Jeong, D.H.; Jun, B.-H. Control of Silver Coating on Raman Label Incorporated Gold Nanoparticles Assembled Silica Nanoparticles. *Int. J. Mol. Sci.* **2019**, *20*, 1258. [[CrossRef](#)] [[PubMed](#)]
36. Bong-Hyun, J.; Gunsung, K.; Sinyoung, J.; Suk, N.M.; Xuan-Hung, P.; Homan, K.; Myung-Haing, C.; Jong-Ho, K.; Yoon-Sik, L.; Hong, J.D. Silica Core-based Surface-enhanced Raman Scattering (SERS) Tag: Advances in Multifunctional SERS Nanoprobes for Bioimaging and Targeting of Biomarkers. *Bull. Korean Chem. Soc.* **2015**, *36*, 963–978. [[CrossRef](#)]
37. Pham, X.-H.; Hahm, E.; Kim, T.H.; Kim, H.-M.; Lee, S.H.; Lee, Y.-S.; Jeong, D.H.; Jun, B.-H. Enzyme-catalyzed Ag Growth on Au Nanoparticle-assembled Structure for Highly Sensitive Colorimetric Immunoassay. *Sci. Rep.* **2018**, *8*, 1–7. [[CrossRef](#)]
38. Pham, X.-H.; Hahm, E.; Kim, T.H.; Kim, H.-M.; Lee, S.H.; Lee, S.C.; Kang, H.; Lee, H.-Y.; Jeong, D.H.; Choi, H.S.; et al. Enzyme-amplified SERS immunoassay with Ag-Au bimetallic SERS hot spots. *Nano Res.* **2020**, *13*, 3338–3346. [[CrossRef](#)]

39. Pham, X.-H.; Hahm, E.; Huynh, K.-H.; Son, B.S.; Kim, H.-M.; Jeong, D.H.; Jun, B.-H. 4-Mercaptobenzoic Acid Labeled Gold-Silver-Alloy-Embedded Silica Nanoparticles as an Internal Standard Containing Nanostructures for Sensitive Quantitative Thiram Detection. *Int. J. Mol. Sci.* **2019**, *20*, 4841. [[CrossRef](#)]
40. Pham, X.-H.; Hahm, E.; Huynh, K.-H.; Kim, H.-M.; Son, B.S.; Jeong, D.H.; Jun, B.-H. Sensitive and selective detection of 4-aminophenol in the presence of acetaminophen using gold-silver core-shell nanoparticles embedded in silica nanostructures. *J. Ind. Eng. Chem.* **2020**, *83*, 208–213. [[CrossRef](#)]
41. Stensberg, M.C.; Wei, Q.; McLamore, E.S.; Porterfield, D.M.; Wei, A.; Sepúlveda, M.S. Toxicological studies on silver nanoparticles: Challenges and opportunities in assessment, monitoring and imaging. *Nanomedicine* **2011**, *6*, 879–898. [[CrossRef](#)]
42. Ferdous, Z.; Nemmar, A. Health Impact of Silver Nanoparticles: A Review of the Biodistribution and Toxicity Following Various Routes of Exposure. *Int. J. Mol. Sci.* **2020**, *21*, 2375. [[CrossRef](#)]
43. Westcott, S.L.; Oldenburg, S.J.; Lee, T.R.; Halas, N.J. Formation and Adsorption of Clusters of Gold Nanoparticles onto Functionalized Silica Nanoparticle Surfaces. *Langmuir* **1998**, *14*, 5396–5401. [[CrossRef](#)]
44. Sadtler, B.; Wei, A. Spherical ensembles of gold nanoparticles on silica: Electrostatic and size effects. *Chem. Commun.* **2002**, *15*, 1604–1605. [[CrossRef](#)]
45. Ryan, D.; Nagle, L.; Rensmo, H.; Fitzmaurice, D. Programmed Assembly of Binary Nanostructures in Solution. *J. Phys. Chem. B* **2002**, *106*, 5371–5377. [[CrossRef](#)]
46. Hiramatsu, H.; Osterloh, F.E. pH-Controlled Assembly and Disassembly of Electrostatically Linked CdSe–SiO₂ and Au–SiO₂ Nanoparticle Clusters. *Langmuir* **2003**, *19*, 7003–7011. [[CrossRef](#)]
47. Lu, L.; Zhang, H.; Sun, G.; Xi, S.; Wang, H.; Li, X.; Wang, X.; Zhao, B. Aggregation-Based Fabrication and Assembly of Roughened Composite Metallic Nanoshells: Application in Surface-Enhanced Raman Scattering. *Langmuir* **2003**, *19*, 9490–9493. [[CrossRef](#)]
48. Xue, J.; Wang, C.; Ma, Z. A facile method to prepare a series of SiO₂@Au core/shell structured nanoparticles. *Mater. Chem. Phys.* **2007**, *105*, 419–425. [[CrossRef](#)]
49. Leng, W.; Pati, P.; Vikesland, P.J. Room temperature seed mediated growth of gold nanoparticles: Mechanistic investigations and life cycle assesment. *Environ. Sci. Nano* **2015**, *2*, 440–453. [[CrossRef](#)]
50. Liz-Marzán, L.M. Tailoring Surface Plasmons through the Morphology and Assembly of Metal Nanoparticles. *Langmuir* **2006**, *22*, 32–41. [[CrossRef](#)]
51. Murphy, C.J.; Sau, T.K.; Gole, A.M.; Orendorff, C.J.; Gao, J.; Gou, L.; Hunyadi, S.E.; Li, T. Anisotropic Metal Nanoparticles: Synthesis, Assembly, and Optical Applications. *J. Phys. Chem. B* **2005**, *109*, 13857–13870. [[CrossRef](#)] [[PubMed](#)]
52. Kelly, K.L.; Coronado, E.; Zhao, L.L.; Schatz, G.C. The Optical Properties of Metal Nanoparticles: The Influence of Size, Shape, and Dielectric Environment. *J. Phys. Chem. B* **2003**, *107*, 668–677. [[CrossRef](#)]
53. Duff, D.G.; Baiker, A.; Edwards, P.P. A new hydrosol of gold clusters. 1. Formation and particle size variation. *Langmuir* **1993**, *9*, 2301–2309. [[CrossRef](#)]
54. Liu, C.-P.; Chen, K.-C.; Su, C.-F.; Yu, P.-Y.; Lee, P.-W. Revealing the Active Site of Gold Nanoparticles for the Peroxidase-Like Activity: The Determination of Surface Accessibility. *Catalysts* **2019**, *9*, 517. [[CrossRef](#)]
55. Li, R.S.; Liu, H.; Chen, B.B.; Zhang, H.Z.; Huang, C.Z.; Wang, J. Stable gold nanoparticles as a novel peroxidase mimic for colorimetric detection of cysteine. *Anal. Methods* **2016**, *8*, 2494–2501. [[CrossRef](#)]
56. Chang, C.-C.; Hsu, T.-L.; Chen, C.-P.; Chen, C.-Y. Enhancement of the Peroxidase-Like Activity of Iodine-Capped Gold Nanoparticles for the Colorimetric Detection of Biothiols. *Biosensors* **2020**, *10*, 113. [[CrossRef](#)] [[PubMed](#)]