Brain Volume in Pediatric Patients with Sickle Cell Disease: Evidence of Volumetric Growth Delay?

R. Grant Steen, Temitope Emudianughe, Michael Hunte, John Glass, Shengjie Wu, Xiaoping Xiong, and Wilburn E. Reddick

BACKGROUND AND PURPOSE: Despite the large body of data available about somatic growth delay in patients with sickle cell disease (SCD), virtually nothing is known about the effect of the disease on volumetric growth of the brain. This study was designed to test a hypothesis that children with SCD have a disease-related delay in brain volumetric growth compared with healthy children.

METHODS: A cross-sectional study design was used to evaluate 83 children with SCD and 43 age-similar healthy children, including 27 patient siblings. Brain volume was measured by segmenting and classifying MR imaging data, by using at least three separate image sets (T1-, T2-, and proton density-weighted MR images). A linear model was used to compare the various brain volumes with the covariates of group (patient versus control) and age, with age treated as a continuous variable.

RESULTS: With age controlled for, no significant difference was noted in total brain volume between patients and control subjects at age 9.5 years. However, patients showed a deficit specifically in gray matter volume (P=.005), without significant differences in white matter or ventricular volume. The deficit in patient gray matter was greater in central gray matter (P<.005) than in cortical gray matter (P<.02). In healthy control subjects, gray matter volume decreased significantly with age (P<.005), probably due to myelination of white matter tracts. In patients with SCD, gray matter volume did not change with age.

CONCLUSION: Volumetric growth of brain gray matter may be delayed in children with SCD, suggesting that there may be neurodevelopmental consequences of this disease.

Children with sickle cell disease (SCD) tend to have impaired somatic growth, delayed puberty (1–3), and reduced bone mineral density (4). Neonates with SCD are of normal height and weight at birth, but they lag behind their peers by the 6th month of life (5). Patients with hemoglobin SS, the most severe form of SCD, have progressive deficits in weight and height by 2 years of age, relative to children with normal hemoglobin (6). Deficits in body weight and body mass index persist throughout childhood (1,

7–9), with an increasing deficit into adulthood in both sexes (3, 6, 10–13). Reduced body size is often associated with low hematocrit values (13, 14), suggesting that somatic growth may be impaired by chronic anemia (13).

Despite the large body of data available on somatic growth delay in patients with SCD, virtually nothing is known about the effect of the disease on volumetric growth of the brain. Preliminary data suggest that the volume of the caudate head is smaller in patients with SCD (15), but this was an anecdotal observation. Because both somatic growth (1–13) and cognitive ability (16, 17) tend to be impaired in patients with SCD, this suggests a hypothesis of impairment in brain volumetric growth in these patients. To test this hypothesis, we analyzed clinical MR imaging data from children with SCD and compared them to the data from age-similar healthy children. Because SCD has been shown to have a diffuse effect on the brain (15–17), we took the approach of segmenting an anatomically defined subsample of the brain; this approach has been validated in our

Received March 12, 2004; accepted after revision July 6. From the Departments of Radiological Sciences (R.G.S., T.E., M.H., J.G., W.E.R.) and Epidemiology and Biostatistics (S.W., X.X.), St. Jude Children's Research Hospital, Memphis, TN.

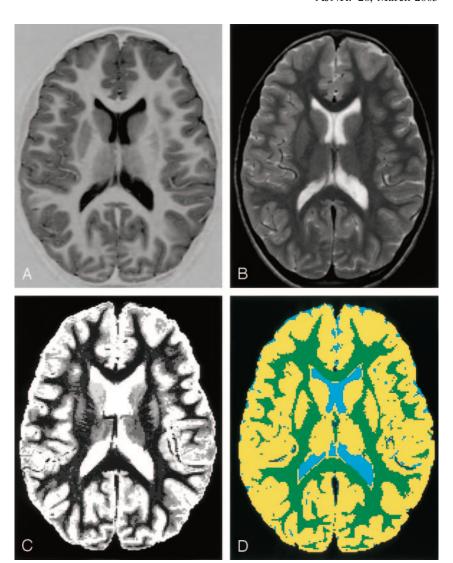
Supported by the National Heart, Lung, and Blood Institute (grant RO1 HL60022 to R.G.S.) and by the American Lebanese Syrian Associated Charities.

Address correspondence to R. Grant Steen, PhD, Department of Psychiatry, University of North Carolina at Chapel Hill, Campus Box 7160, Chapel Hill, NC 27599-7160; e-mail: Grant_Steen@med.unc.edu

© American Society of Neuroradiology

Fig. 1. Images obtained in a 7.8-year-old girl with SCD show how image segmentation was performed.

- A and B, T1-weighted (A) and T2-weighted (B) MR images obtained at the same section level.
- C, Segmented image made by combining T1-weighted, T2- or proton density-weighted, and FLAIR images shows 16 types of tissue as nine different gray scale levels.
- D, Segmented and classified image shows gray matter (yellow), white matter (green), blood vessels and membranes (dark blue), and CSF (light blue). To calculate central gray matter (basal ganglia) volume separate from cortical volume, all tissues external to the basal ganglia were erased. The cortical gray matter volume was then calculated as a function of total gray matter volume.



laboratory as a way to minimize processing time without sacrificing accuracy (18).

Methods

Study Subjects

All patients with SCD and control subjects were enrolled on a protocol approved by the Institutional Review Board at St. Jude Children's Research Hospital. Patients or guardians of all children signed an informed consent form after a brief description of the protocol, whereas adult subjects (older than 18 years) signed their own informed consent form. All children assented to participation.

We imaged a total of 87 patients with SCD between October 1999 and March 2003. Four patients were excluded from further analysis because they had a clinically documented stroke. Among the remaining 83 patients (47 male [57%] and 36 female [43%] patients), their ages ranged from 3.9 to 18.5 years (mean age, 9.4 \pm 3.8 years [SD]). The patients mostly had hemoglobin SS (n = 70), the most serious form of SCD, although there were 11 patients with hemoglobin SC, one with hemoglobin S β^+ , and one whose hemoglobin phenotype was unknown.

We also imaged 43 healthy control subjects during the same period (26 male [61%] and 17 female [39%] subjects whose ages ranged from 4.3 to 18.0 years (mean age, 10.0 ± 3.6 years). The control group included 27 healthy siblings of the patients

with SCD, as well as 16 healthy subjects who were either the healthy siblings of other hospital patients or the children of hospital personnel. Among the control subjects, there were 23 subjects with hemoglobin AA and 20 subjects with hemoglobin AS (sickle cell trait).

MR Imaging

MR imaging evaluations were performed with a 1.5T Vision (Siemens Medical Systems, Iselin, NJ) whole-body imager by using a standard head coil. Weekly quality assurance monitoring of field homogeneity and eddy current compensation was performed, and image quality was monitored daily as a part of the clinical imaging program.

A standard protocol for all subjects included a T1-weighted gradient-echo sequence (Fig 1A) and a T2- or proton density-weighted dual spin-echo sequence (Fig 1B). A fluid-attenuated inversion-recovery (FLAIR) sequence was added in March 2000. All images were acquired in the transverse imaging plane, with the central section of each sequence angled so that the bottom of the section was congruent with the bottom of the genu and splenium on the sagittal scout image. Setting up the acquisition in this way is a highly reproducible way to sample the brain in children (15, 18). Before March 2000, all images were acquired at 5-mm section thickness with a 1-mm gap, to avoid cross talk between section excitations. Beginning in March 2000, all images were acquired as 3-mm sections with a

3-mm gap, then each sequence was repeated with a 3-mm offset, to give 100% coverage of the brain volume. To determine whether the two methods of measuring brain volume gave equivalent results, we undertook a small study comparing the brain volume of five volunteers by using both methods.

Quantitative Analysis of MR Imaging Data

Image registration was performed for each examination (19), to align features in one image set to correspond to features in a related image set, by matching voxels that contain high-intensity gradient magnitudes. A postprocessing algorithm to correct radio-frequency inhomogeneity in the imaging sets was used to ensure uniform signal intensity throughout the brain volume (20). An added advantage of the radio-frequency correction method is the automated removal of extrameningeal tissues, which reduces the amount of postprocessing necessary on the segmented images.

A semi-automated hybrid neural network method (21) was used to segment and classify images, to objectively categorize gray matter and white matter. A sample image (Fig 1C) shows the 16 gray-scale levels in a segmented image. These 16 classifications were mapped in four colors (corresponding to gray matter, white matter, CSF, and vessels or membranes) (Fig 1D), by using a color scheme similar to that used for positron emission tomography. This image was then imported into PhotoShop running on a Macintosh, and the histogram feature was used to measure the number of pixels of each tissue type. To separately determine the number of pixels in central gray matter (basal ganglia) and cortical gray matter, cortical gray matter pixels were erased in the PhotoShop image, and the histogram feature was used to count the number of central gray matter pixels. The number of cortical gray matter pixels was then calculated by subtracting central gray matter pixels from total gray matter pixels. Previous work (18, 20-22) has established that these methods are valid and robustly reliable.

The volume of brain parenchyma was calculated by multiplying the number of pixels of each tissue type by the known pixel volume. Pixel volumes were summed across the full volume of interest, and the following equation was used to calculate brain volume: Brain Volume (mL) = (Pixel Count • $[FOV/256]^2$ • Section Thickness)/1000, where FOV was either 210 or 230 mm and section thickness was either 3 or 5 mm. In the earlier examinations in which a 5-mm section thickness was used, this section was interpolated to an effective section thickness of 6 mm. In the later examinations in which a 3-mm section thickness was used, two adjacent sections were summed to obtain an effective section thickness of 6 mm.

Volume calculations are thus based on a 6-mm slab through the basal ganglia (Fig 1), regardless of which section thickness was used during imaging. Volume estimation from such an anatomically defined subset of a full clinical examination significantly correlated with the whole cerebrum ($r \approx 0.94$), with an average error of 2% in total gray matter volume and 6% in total white matter volume (18).

Effects of Blood Phenotype on Brain Volume

Hemoglobin phenotype (AA, AS, SS) data were obtained concurrent with imaging. For this secondary analysis, we excluded 13 patients with a hemoglobin phenotype other than SS. Our hypothesis was that there would be a relationship between hemoglobin phenotype and brain volume, such that patients with hemoglobin SS would have a smaller brain volume than that of children with hemoglobin AA or AS.

Statistical Analysis

A general linear model was used to estimate mean brain volume as a linear function of age at MR imaging, with separate analyses for volume of all tissues, white matter, gray matter, and CSF. In this model, brain volume was the response

TABLE 1: Validation of the methods for measuring brain volume performed in five volunteers

BRAIN VOLUME IN SICKLE CELL DISEASE

Structure	Volume (mL) Measured by 5-mm Thick Sections	Volume (mL) Measured by 3-mm Thin Sections
White matter	33.86 ± 3.33	34.00 ± 3.02
Gray matter	58.28 ± 3.77	58.44 ± 3.60
CSF	9.68 ± 3.26	9.54 ± 3.18

Note.—Data are mean \pm SD. The mean difference between the two methods was 0.053 mL. A paired t test comparing 15 measurements made on thick sections to the same measurements made on thin sections was not significant (t statistic = 0.5524 with 14 degrees of freedom; P = 589)

variable, whereas the subject group (patient versus control) and the age at MR imaging were covariates. We also used gender as a covariate, but preliminary analysis suggested that gender did not have a significant effect on brain volume in these children. Since boys and girls were equally represented in the two subject groups, we did not include gender in the final analysis. Brain volume was fitted as a function of the following equation: Volume = (Intercept for Group) + (Age • Group), where group is the subject group (patients with SCD versus control subjects). We compared baseline brain volume (intercept) and the rate of change of brain volume (slope) between the two groups, and we also compared the mean volume between groups at various ages. All analyses were performed using the SAS/STAT statistical package (23).

Results

Method Validation

A comparison of brain volume measured in five subjects by the two different methods (3-mm thin section versus 5-mm thick section) showed no significant differences that could be attributed to the method itself (Table 1). This suggests that comparing results from the two methods is legitimate.

Descriptive Comparison of Brain Volume

Patients and healthy control subjects did not differ significantly in age, suggesting that age-related differences between patients and control subjects should be negligible (Table 2). Total brain volume was significantly less (P < .04) in patients than in healthy control subjects. There was 4% less gray matter volume in patients (P < .01), with a greater deficit in central gray matter (basal ganglia) volume (6%, P < .01) than in cortical gray matter volume (3%, P < .03). No significant difference was noted in either white matter volume or ventricular volume between patients with SCD and control subjects (Table 2).

Modeling Brain Volume Differences

A scatterplot of total brain volume (Fig 2) suggests that age-related trends in growth may differ between patients and control subjects. Even though patient and control ages were not significantly different, patients were about 8 months younger than control subjects (Table 2). Thus, it is possible that the age

458 STEEN AJNR: 26, March 2005

TABLE 2: Comparison of descriptive data in patients with SCD and control subjects without controlling for age

Comparison	Patients $(n = 83)^*$	Controls $(n = 43)^*$	% Difference	P value [†]
Subject age (y)	9.36 ± 3.83	10.02 ± 3.61	-6.40	NS
Proportion male (%)	56.6	58.1	-2.60	NS
Volume (mL)				
Total brain	94.72 ± 8.11	97.24 ± 6.10	-2.60	.04
White matter	27.16 ± 4.55	27.52 ± 4.59	-1.30	NS
Gray matter	61.30 ± 6.10	63.99 ± 5.18	-4.20	.01
Central gray matter	10.60 ± 1.46	11.32 ± 1.51	-6.40	.01
Cortical gray matter	50.96 ± 4.37	52.58 ± 4.59	-3.10	.03
CSF	6.26 ± 2.55	5.73 ± 1.90	9.20	NS

Note.—NS indicates not significant. Age did not differ significantly between patients and controls, so this approach may be valid.

[†] Statistical testing was performed with a two-sample t test.

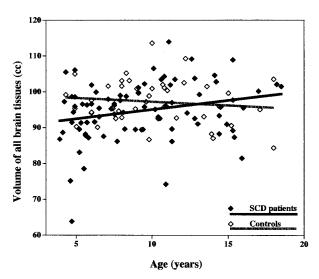


Fig 2. Scatterplot shows volume of all tissues in patients with SCD and in healthy control children.

difference between patients and control subjects is clinically significant, even though it is not statistically significant.

Therefore, we controlled for differences in age between patients and control subjects by modeling brain volume as a function of age at 9.5 years, the grand mean age, or the average age across all subjects (Table 3). This approach should maximize sensitivity to volume differences between patients and control subjects, since the grand mean age is the age at which results are statistically the most robust. When age was controlled for in this way, there was no longer a difference in total volume of all tissues between patients and control subjects. Yet the difference in gray matter volume between patients and control subjects became more robustly significant (P = .005). The deficit in patient gray matter was larger and more significant in central gray matter (P < .005) than in cortical gray matter (P < .02). No significant differences were noted between patients and control subjects in white matter or ventricular volume (Table 3).

A least-squares approach was then used to compare brain volume in patients and control subjects at age 5 years and at age 11.5 years (Table 3). At age 5

years, patients showed volume deficits in total brain volume (P = .01), in total gray matter volume (P = .0004), in central gray matter volume (P = .001), and in cortical gray matter volume (P = .002). There was an excess of CSF volume in patients (P < .007). By age 11.5 years, all of these differences had become nonsignificant.

We modeled the effects of SCD and subject age on brain volume by using a univariate approach (Table 4). For healthy control subjects, total brain volume was Volume = $99.34 + (-0.21 \cdot Age)$, whereas for patients with SCD, total brain volume was Volume = $89.93 + (0.51 \cdot Age)$.

No significant interaction was noted between age and total brain volume in control subjects, suggesting that volumetric growth of the brain is complete in children by about age 5 years (Table 4). However, among control subjects, a significant age-related *decrease* was noted in volume of gray matter (P = .005), central gray matter (P < .04), and cortical gray matter (P < .002), whereas volumes of total brain volume and white matter in the control subjects did not change as a function of age (Table 4).

Conversely, among patients, a significant age-related increase was noted in total brain volume (P < .02) and volume of white matter (P = .0007), suggesting that the brain is still growing and myelinating in patients of the age range studied. No significant change was noted in volume of gray matter, central gray matter, or cortical gray matter (Table 4). Thus, there are significant trend differences between patients and control subjects (Fig 3). The finding that brain volume trends are in opposite directions in patients and control subjects could account for the fact that differences between patients and control subjects disappear by age 11.5 years (Table 3).

Effects of Hemoglobin Phenotype on Brain Volume

Patients with hemoglobin SS were significantly different from control children with hemoglobin AA in many ways (Table 5). At age 5 years, patients had smaller brain volume (P = .02), smaller gray matter volume (P = .0002), smaller central gray matter vol-

^{*} Data are mean ± SD.

TABLE 3: Least-squares means of brain volumes between the 83 patients with SCD and 43 control subjects as a function of age

Volume and Group	5.0 Years Old	P Value	9.5 Years Old	P Value	11.50 Years	P Value
Total brain						
Control	98.29 ± 1.94	.0127	97.35 ± 1.13	NS	96.93 ± 1.21	NS
SCD	92.49 ± 1.22		94.79 ± 0.80		95.82 ± 0.92	
White matter						
Control	26.63 ± 1.16	NS	27.43 ± 0.68	NS	27.79 ± 0.72	NS
SCD	25.25 ± 0.72		27.22 ± 0.48		28.10 ± 0.55	
Gray matter						
Control	67.47 ± 1.50	.0004	64.35 ± 0.87	.0052	62.96 ± 0.94	NS
SCD	61.07 ± 0.94		61.30 ± 0.62		61.41 ± 0.71	
Central gray matter						
Control	11.98 ± 0.38	.0014	11.39 ± 0.22	.0045	11.13 ± 0.24	NS
SCD	10.51 ± 0.24		10.60 ± 0.16		10.64 ± 0.18	
Cortical gray matter						
Control	55.54 ± 1.14	.0021	52.89 ± 0.67	.0195	51.71 ± 0.72	NS
SCD	51.29 ± 0.72		50.95 ± 0.48		50.80 ± 0.54	
CSF						
Control	4.18 ± 0.61	.0067	5.56 ± 0.36	NS	6.18 ± 0.38	NS
SCD	6.17 ± 0.38		6.27 ± 0.26		6.31 ± 0.29	

Note.—NS indicates not significant. Data are mean \pm standard error.

TABLE 4: Age-related change in brain volume in 83 patients with SCD and 43 control subjects

		Standard	
Effect	Estimate*	Error	P Value [†]
Total brain volume			
Intercept for controls	99.34	3.36	<.0001
Intercept for SCD	89.93	2.12	<.0001
Age \times controls	-0.21	0.32	NS
Age × SCD patients	0.51	0.21	.0162
White matter			
Intercept for controls	25.74	2.00	<.0001
Intercept for SCD	23.06	1.27	<.0001
$Age \times controls$	0.18	0.19	NS
Age × SCD patients	0.44	0.13	.0007
Gray matter			
Intercept for controls	70.95	2.59	<.0001
Intercept for SCD	60.81	1.63	<.0001
Age \times controls	-0.69	0.24	.0050
Age × SCD patients	0.05	0.16	NS
Central gray matter			
Intercept for controls	12.64	0.66	<.0001
Intercept for SCD	10.41	0.42	<.0001
$Age \times controls$	-0.13	0.06	.0360
Age × SCD patients	0.02	0.04	NS
Cortical gray matter			
Intercept for controls	58.49	1.98	<.0001
Intercept for SCD	51.66	1.25	<.0001
$Age \times controls$	-0.59	0.19	.0019
Age × SCD patients	-0.07	0.12	NS
CSF			
Intercept for controls	2.65	1.05	.0130
Intercept for SCD	6.05	0.66	<.0001
$Age \times controls$	0.31	0.10	.0024
Age × SCD patients	0.02	0.07	NS

^{*} Estimates of fitted model between groups.

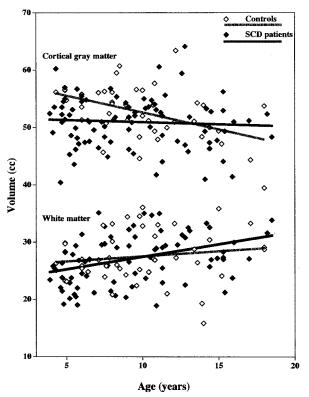


Fig 3. Scatterpolots show cortical gray matter volume (top) and white matter volume (bottom) in patients with SCD and in healthy control children.

ume (P < .008), smaller cortical gray matter volume (P = .0005), and larger CSF volume (P = .01). By age 9.5 years, some of these differences had disappeared, but patients still had smaller gray matter volume (P < .002), smaller central gray matter volume (P = .03), and smaller cortical gray matter volume (P < .002). By age 11.5 years, all of these differences had disappeared.

 $^{^{\}dagger}$ The P value for the "Intercept for controls" and the "Intercept for SCD" tests whether the respective intercept is significantly different from zero. The P value for "Age \times controls" and "Age \times SCD patients" tests whether there is a significant effect of age in controls and patients, respectively, by testing whether the slope is significantly different from zero slope.

460 STEEN AJNR: 26, March 2005

TABLE 5: Least-squares means of brain volumes as a function of genotype in 70 patients with hemoglobin SS, 20 control subjects with hemoglobin AS, and 23 control subjects with hemoglobin AA

Volume and			
Genotype	5.0 Years Old	9.5 Years Old	11.5 Years Old
Total brain		Means ± SE	
AA	98.79 ± 2.50		96.95 ± 1.66
AS	97.37 ± 3.28	97.12 ± 1.70	97.01 ± 1.87
SS	92.18 ± 1.34		95.66 ± 1.00
		P Value	
AA vs SS	0.0217	NS	NS
AS vs SS	NS	NS	NS
AA vs AS	NS	NS	NS
White matter		Means \pm SE	
AA	25.61 ± 1.47	26.53 ± 0.92	26.94 ± 0.98
AS	27.90 ± 1.93	28.48 ± 0.99	28.73 ± 1.10
SS	25.00 ± 0.79	26.96 ± 0.53	27.83 ± 0.59
		P Value	
AA vs SS	NS	NS	NS
AS vs SS	NS	NS	NS
AA vs AS	NS	NS	NS
Gray matter		Means \pm SE	
AA	68.95 ± 1.95	65.55 ± 1.19	64.03 ± 1.26
AS	65.52 ± 2.49	62.95 ± 1.29	61.82 ± 1.40
SS	60.70 ± 1.02	61.17 ± 0.68	61.37 ± 0.77
		P Value	
AA vs SS	0.0002	0.0018	NS
AS vs SS	NS	NS	NS
AA vs AS	NS	NS	NS
Central gray matter		Means ± SE	
AA	11.99 ± 0.49	11.37 ± 0.31	11.09 ± 0.32
AS	11.94 ± 0.64	11.41 ± 0.33	11.17 ± 0.37
SS	10.48 ± 0.26	10.61 ± 0.18	10.66 ± 0.20
		P Value	
AA vs SS	0.0078	0.0340	NS
AS vs SS	0.0374	0.0343	NS
AA vs AS	NS	NS	NS
Cortical gray matter		Means \pm SE	
AA	56.96 ± 1.45	54.18 ± 0.91	52.94 ± 0.96
AS	53.80 ± 1.90	51.40 ± 0.98	50.33 ± 1.08
SS	51.07 ± 0.78	50.86 ± 0.52	50.77 ± 0.58
		P Value	
AA vs SS	0.0005	0.0019	NS
AS vs SS	NS	NS	NS
AA vs AS	NS	0.0397	NS
CSF		Means \pm SE	
AA	4.22 ± 0.77	5.44 ± 0.48	5.98 ± 0.51
AS	3.95 ± 1.00	5.70 ± 0.52	6.47 ± 0.58
SS	6.48 ± 0.41	6.46 ± 0.28	6.45 ± 0.31
		P Value	
AA vs SS	0.0111	NS	NS
AS vs SS	0.0221	NS	NS
AA vs AS	NS	NS	NS

Note.—NS indicates not significant; SE, standard error.

We also assessed whether brain volumes differed between control children with hemoglobin AS (sickle cell trait) and patients with hemoglobin SS (Table 5). This comparison is particularly important because AS control subjects were all siblings of the patients. Therefore, this comparison should control for variables such as family or genetic influences and for socioeconomic factors that can potentially impact brain volume growth. Despite the reduced sample size of control subjects available for comparison, pa-

tients had a smaller central gray matter volume at age 5 years (P < .04) and at age 9.5 years (P = .03), and a larger CSF volume at age 5.0 years (P = .02).

Finally, we assessed whether sickle cell trait itself had an effect on brain volume, by separately modeling control children with sickle trait (hemoglobin AS) and control children with hemoglobin AA (Table 5). Children with sickle trait typically do not have severe anemia (24) but they had a significantly smaller cortical gray matter volume at age 9.5 years than that of children with hemoglobin AA (P < .04), despite the reduced sample size. No other significant differences were noted between children with hemoglobin AA and those with AS.

Discussion

We report that gray matter volume is significantly smaller in patients with SCD than in healthy children (Table 2), especially when age effects are modeled (Tables 3 and 4). Volumetric deficits in patients are larger and more significant in central gray matter than in cortical gray matter (Table 3), and gray matter deficits appear to be a function of different patterns of volumetric growth in patients and healthy children (Table 4). Specifically, patient total brain volume significantly increases at an age when control brain volume has already stabilized (Fig 2, Table 4). The changes that we describe could be the result of normal developmental processes that are occurring at a slower-than-normal rate (Table 4), as a function of hemoglobin type (Table 5). Alternatively, patients with SCD may be showing subtle neurodevelopmental effects of their disease.

The limitations of this study arise mostly from the cross-sectional study design. All the data we report are cross-sectional, yet we are using these data to infer patterns of growth over time (Fig 3). This can be risky if there are significant differences in the composition of the patient and control groups, or if volumetric growth of brain tissues is not linear. We are reasonably confident that the composition of the patient and control groups is comparable (Table 2), both in terms of gender and in terms of age distribution. However, we cannot be sure that brain volumetric growth is linear, even though we have chosen to fit our data with a linear model. We elected to use a simple linear fit since such a model is more easily disproven than either a model with more terms or a model with quadratic functions (25). However, if more data were available, it might become preferable to use a more sophisticated model. A nonlinear model might be especially important if there are subtle interactions between age and disease. Because patients with SCD tend to be at greatest risk of stroke in the age range of 4–9 years (26), it is reasonable to propose that there is an increase in the rate of brain volumetric growth during this period (16). A linear model would not be an adequate description of such a growth spurt, particularly if healthy children do not have a similarly timed spurt (27).

Another potential limitation of this study is that we measured brain volume in a single slab, rather than in

the entire brain volume. We believe that this approach is justified in these patients by the finding that SCD has a diffuse effect on the entire brain (15–17). There is compelling evidence that every gray matter tissue is impacted by SCD, both in terms of the risk of focal injury (15) and in terms of diffuse effects on gray matter T1 (16, 25). Furthermore, the approach of evaluating a subsample of the brain has been validated in our laboratory as a way to minimize image-processing time without sacrificing accuracy (18).

A third potential limitation of our study is that we did not incorporate sex into the final analysis. A preliminary analysis of our data found significant effects of age on brain volume, but interactions with covariates (group x gender) and (group x age x gender) were not statistically significant. Therefore, we elected to leave gender out of the final analysis, since the proportion of boys was nearly equivalent in the patient and control groups. We are aware that cerebral volume is roughly 9% larger in male than in female subjects (28, 29), but earlier studies have shown that there are no sex-related differences in patterns of gray matter or white matter growth, after correcting for brain volume (29). Similarly, we elected to leave race out of the final analysis, since brain volumetric growth in black and white infants is equivalent, and brain growth rate in young children is related neither to race nor to gender (30).

The pattern of volumetric growth in healthy children has been moderately well characterized. Intracranial volume increases by only ~300 mL from age 3 months to 10 years (31), and there is little change in total cerebral volume after the age of 5 years (32, 33). Cortical gray matter volume peaks at age 4 years and decreases thereafter, whereas cortical white matter volume increases until at least age 20 years (31), due to the ongoing process of brain myelination. At age 9 years, white matter volume is only 85% of adult volume, whereas gray matter volume is actually larger than the adult volume, since white matter myelination is not complete until well past age 9 years (34). In general, white matter volume increases at a faster rate than gray matter volume throughout early childhood, as myelination proceeds (34, 35). Cross-sectional data in healthy children had suggested a plateau in gray matter volume by about age 13 years (36), but recent longitudinal data suggest that gray matter volume actually peaks at age 12 years and begins to decline thereafter (37). There is no significant difference in rate of volume loss as a function of sex (37).

Our data suggest that there is a decrease in cortical gray matter volume in healthy children between the ages of 5 and 11.5 years (Tables 3 and 4), which is consistent with the literature (31). Many studies have shown that gray matter volume decreases with age in adults (31, 38–41), although fewer studies have documented loss of gray matter volume in adolescents (31, 42–45) or children (29, 31, 43, 45). Our study is actually more unusual in reporting that patients with SCD do *not* show an age-related decrement in gray matter.

Our results suggest that brain volume in young patients is smaller than normal because brain growth pro-

cesses are delayed or impaired in children with SCD (Tables 3 and 4). In particular, we note that total brain volume is still increasing significantly in patients at an age when brain volumetric growth has already ceased in control subjects (Table 4). Furthermore, patients had 9% less gray matter than control subjects at age 5.0 years (P = .0004) and 5% less gray matter than control subjects at age 9.5 years (P = .0052) (Table 3). The volumetric deficit in gray matter of patients cannot be accounted for by encephalomalacia, since this should result in a progressive decrease in the volume of white matter over time, whereas a significant increase in white matter was actually seen in patients (Table 4). Although our prior work shows that patients with SCD can have an age-related increase in encephalomalacia (46), we excluded patients with clinical stroke from this analysis. Furthermore, if encephalomalacia had caused a significant gray matter volume loss in patients, one would expect the CSF volume to increase with age, but no significant age-related increase in patient CSF volume was seen (Table 4). We noted a significant age-related increase in CSF volume in the control subjects, which may result from the significant age-related decrease in volume of central and cortical gray matter in control subjects (Table 4). Thus, the simplest explanation for the gray matter deficit in patients at age 9.5 years (Table 3) is that central gray matter does not reach a maximal volume as early in patients as it does in control subjects.

An alternative possibility is that central gray matter atrophy occurred at a very young age in our patients (< 5 years). In support of this possibility, we note that CSF volume was larger in patients than in control subjects at age 5 years (Table 3). Furthermore, control subjects showed a significant loss of central gray matter (Table 4), but still had more central gray matter than patients at age 5 and 9.5 years (Table 3). It will be critical in future work to characterize the pattern of volumetric brain growth in very young patients (< 5 years old), to determine whether atrophy or growth delay is a better explanation for the volume deficit in central gray matter noted at age 5 years (Table 3).

Conclusion

We describe volumetric deficits specific to gray matter, in patients with SCD, whereas there is no evidence of a volumetric deficit in white matter. These findings appear to be consistent with measurements that show reduction of brain tissue T1 specifically in gray matter, with no T1 abnormality in the white matter of patients with SCD (15, 16, 25, 46). Our results thus suggest that gray matter is selectively vulnerable to injury in children with SCD, perhaps because of the severe anemia that can be present in these patients (16, 17). Our results show that children with SCD can experience neurodevelopmental consequences of their disease.

Acknowledgments

We thank Mary Freeman, Crystal Manchester, Carolyn Phillips, and Mark Summers, who performed 462 STEEN AJNR: 26, March 2005

all of the MR imaging, and the patients and families of St. Jude who dedicated their time and effort.

In Memoriam

This article is dedicated to the memory of our good friend and valued colleague, Dr. Temitope Emudianughe-Okponobi, who died recently from complications of cancer. She was a wonderful person, a fine physician, and a clear-thinking scientist. She fought against great odds, and she will be deeply missed.

References

- Platt OS, Rosenstock W, Espeland MA. Influence of sickle hemoglobinopathies on growth and development. N Engl J Med 1984;311:7–12
- Barden EM, Zemel BS, Kawchak DA, Goran MI, Ohene-Frempong K, Stallings VA. Total and resting energy expenditure in children with sickle cell disease. J Pediatr 2000;136:73–79
- Barden EM, Kawchak DA, Ohene-Frempong K, Stallings VA, Zemel BS. Body composition in children with sickle cell disease. Am J Clin Nutr 2002;76:218–225
- Soliman AT, Bererhi H, Darwish A, Alzalabani MM, Wali Y, Ansari B. Decreased bone mineral density in prepubertal children with sickle cell disease: correlation with growth parameters, degree of siderosis, and secretion of growth factors. J Trop Pediatr 1998;44:194–198
- VanderJagt DJ, Kanellis GJ, Isichei C, Patuszyn A, Glew RH. Serum and urinary amino acid levels in sickle cell disease. J Trop Pediatr 1997;43:220–225
- Stevens MC, Maude GH, Cupidore L, Jackson H, Hayes RJ, Serjeant GR. Prepubertal growth and skeletal maturation in children with sickle cell diease. *Pediatrics* 1986;78:124–132
- Stevens MC, Hayes RJ, Serjeant GR. Body shape in young children with homozygous sickle cell disease. Pediatrics 1983;71:610–614
- Oyedeji GA, Olamijulo SK, Asinaike AI, Esimai VC, Odunusi EO, Aladekomo TA. Anthro-pometric measurement in children aged 0-6 years in a Nigerian village. East Afr Med J 1995;72:523-526
- Patey RA, Sylvester KP, Rafferty GF, Dick M, Greenough A. The importance of using ethnically appropriate reference ranges for growth assessment in sickle cell disease. Arch Dis Child 2002;87:352–353
- Phebus CK, Gloninger MF, Maciak BJ. Growth patterns by age and sex in children with sickle cell diease. J Pediatr 1984;105:28–33
- Zago MA, Kerbauy J, Souza HM, et al. Growth and sexual maturation of Brazilian patients with sickle cell diseases. Trop Geogr Med 1992;44:317–321
- Modebe O, Ifenu SA. Growth retardation in homozygous sickle cell disease: role of calorie intake and possible gender-related differences. Am J Hematol 1993;44:149–154
- Singhal A, Morris J, Thomas P, Dover G, Higgs D, Serjeant G. Factors affecting prepubertal growth in homozygous sickle cell disease. Arch Dis Child 1996;74:502–506
- Silva CM, Viana MB. Growth deficits in children with sickle cell disease. Arch Med Res 2002;33:308–312
- Steen RG, Langston JW, Ogg RJ, Xiong X, Ye Z, Wang WC. Diffuse T1 reduction in gray matter of sickle cell disease patients: evidence of selective vulnerability to damage? Mag Reson Imag 1999;17:503-515
- Steen RG, Xiong X, Mulhern RK, Langston JW, Wang WC. Subtle brain abnormalities in children with sickle cell disease: relationship to blood hematocrit. Ann Neurol 1999;45:279–286
- Steen RG, Miles M, Helton K, et al. Cognitive impairment in children with hemoglobin SS sickle cell disease: relationship to MR imaging findings and hematocrit. Am J Neuroradiol 2003;24:382–389
- Glass JO, Ji Q, Glas LS, Reddick WE. Prediction of total cerebral tissue volumes in normal-appearing brain from sub-sampled segmentation volumes. Mag Reson Imaging 2003;21:977–982
- Ostuni JL, Levin RL, Frank JA, DeCarli C. Correspondence of closest gradient voxels: a robust registration algorithm. J Mag Reson Imaging 1997;7:410–415
- Ji Q, Reddick WE, Glass JO, Krynetskiy E. Quantitative study of the renormalization transformation method to correct intensity inhomogeneity in MR images. Presented at the SPIE International

- Symposium on Medical Imaging, Image Processing Conference, San Diego, CA; February 23–28, 2002
- Reddick WE, Glass JO, Cook EN, Elkin TD, Deaton RJ. Automated segmentation and classification of multispectral magnetic resonance images of brain using artifical neural networks. *IEEE Trans Med Imaging* 1997;16:911–918
- Reddick WE, Glass JO, Langston JW, Helton KJ. Quantitative MRI assessment of leuko-encephalopathy. Mag Reson Med 2002:47:912–920
- 23. SAS/STAT User's Guide, Version 8. Cary, NC: SAS Institute, Inc.; 1999
- Oski FA. Differential diagnosis of anemia. In: Oski FA, Nathan DG, eds. Hematology of Infancy and Childhood. Vol. 1. Philadelphia: W. B. Saunders Co.; 1987:265–273
- Steen RG, Schroeder J. Age-related changes in the human brain: proton T1 in healthy children and in children with sickle cell disease. Mag Reson Imaging 2003;21:9–15
- Ohene-Frempong K, Weiner SJ, Sleeper LA, et al. Cerebrovascular accidents in sickle cell disease: rates and risk factors. Blood 1998;91:288–294
- 27. Mann MD. The growth of the brain and skull in children. $\it Brain~Res~1984; 315:169-178$
- Giedd JN, Rumsey JM, Castellanos FX, et al. A quantitative MRI study of the corpus callosum in children and adolescents. Develop Brain Res 1996;91:274–280
- Rajapakse JC, Giedd JN, DeCarli C, et al. A technique for singlechannel MR brain tissue segmentation: application to a pediatric sample. Mag Reson Imaging 1996;14:1053–1065
- Ho KC, Roessmann U, Hause L, Monroe G. Correlation of perinatal brain growth with age, body size, sex, and race. J Neuropathol Exp Neurol 1986;45:179–188
- Pfefferbaum A, Mathalon DH, Sullivan EV, Rawles JM, Zipursky RB, Lim KO. A quantitative magnetic resonance imaging study of changes in brain morphology from infancy to late adulthood. Arch Neurol 1994;51:874–887
- 32. Reiss AL, Abrams MT, Singer HS, Ross JL, Denckla MB. Brain development, gender and IQ in children: a volumetric imaging study. Brain 1996;119:1763–1774
- Giedd J. Brain development, IX: Human brain growth. Am J Psychiatry 1999;156:4
- 34. Caviness VS, Kennedy DN, Richelme C, Rademacher J, Filipek PA. The human brain age 7-11 years: a volumetric analysis based on magnetic resonance images. Cereb Cortex 1996;6:726-736
- Matsuzawa J, Matsui M, Konishi T, et al. Age-related volumetric changes of brain gray and white matter in healthy infants and children. Cereb Cortex 2001;11:335–342
- 36. James ACD, Crow TJ, Renowden S, Wardell AMJ, Smith DM, Anslow P. Is the course of brain development in schizophrenia delayed? Evidence from onsets in adolescence. Schizophren Res 1999;40:1–10
- Giedd JN, Blumenthal J, Jeffries NO, et al. Brain development during childhood and adolescence: a longitudinal MRI study. Nat Neurosci 1999;2:861–863
- Jernigan TL, Trauner DA, Hesselink JR, Tallal PA. Maturation of human cerebrum observed in vivo during adolescence. Brain 1991:114:2037–2049
- Lim KO, Zipursky RB, Watts MC, Pfefferbaum A. Decreased gray matter in normal aging: an in vivo magnetic resonance study. J Gerontol 1992;47:B26–B30
- Gunning-Dixon FM, Head D, McQuani J, Acker JD, Raz N. Differential aging of the human striatum: a prospective MR imaging study. Am J Neuroradiol 1998;19:1501–1507
- Ge Y, Grossman RI, Babb JS, Rabin ML, Mannon LJ, Kolson DL. Age-related total gray matter and white matter changes in normal adult brain. I: Volumetric MR imaging analysis. Am J Neuroradiol 2002;23:1327–1333
- Jernigan TL, Tallal P. Late childhood changes in brain morphology observable with MRI. Dev Med Child Neurol 1990;32:379–385
- Lange N, Giedd JN, Castellanos FX, Vaituzis AC, Rapoport JL.
 Variability of human brain structure size: ages 4–20 years. Psychiat Res 1997;74:1–12
- Rapoport JL, Castellanos FX, Gogate N, Janson K, Kohler S, Nelson P. Imaging normal and abnormal brain development: new perspectives for child psychiatry. Aust N Z J Psychiat 2001;35:272–281
- Castellanos FX, Lee PP, Sharp W, et al. Developmental trajectories of brain volume abnormalities in children and adolescents with attention-deficit hyperactivity disorder. JAMA 2002;288:1740–1748
- Steen RG, Xiong X, Langston JW, Helton KJ. Brain injury in children with sickle cell disease: prevalence and etiology. Ann Neurol 2003;54:564–572