







RESEARCH ARTICLE

Diagnostic accuracy of two commercial SARS-CoV-2 antigen-detecting rapid tests at the point of care in community-based testing centers

Alice Berger¹ , Marie Therese Ngo Nsoga¹ , Francisco Javier Perez-Rodriguez² , Yasmine Abi Aad¹, Pascale Sattounet-Roche², Angèle Gayet-Ageron³ , Cyril Jaksic³, Giulia Torriani⁴, Erik Boehm¹, Ilona Kronig¹, Jilian A. Sacks⁵, Margaretha de Vos⁵ , Frédérique Jacqueroz Bausch⁶, François Chappuis⁶, Adriana Renzoni⁷, Laurent Kaiser^{1,2,7}, Manuel Schibler^{1,2,7}, Isabella Eckerle^{1,2,7} 

1 Division of Infectious Disease, Geneva University Hospitals, Geneva, Switzerland, **2** Geneva Centre for Emerging Viral Diseases, Geneva University Hospitals, Geneva, Switzerland, **3** CRC & Division of Clinical-Epidemiology, Department of Health and Community Medicine, University of Geneva & University Hospitals of Geneva, Geneva, Switzerland, **4** Department of Microbiology and Molecular Medicine, University of Geneva, Geneva, Switzerland, **5** Foundation for Innovative New Diagnostics, Geneva, Switzerland, **6** Department of Primary Care, Geneva University Hospitals, Geneva, Switzerland, **7** Division of Laboratory Medicine, Laboratory of Virology, Geneva University Hospitals, Geneva, Switzerland

 These authors contributed equally to this work.

* Isabella.eckerle@hcuge.ch



 OPEN ACCESS

Citation: Berger A, Nsoga MTN, Perez-Rodriguez FJ, Aad YA, Sattounet-Roche P, Gayet-Ageron A, et al. (2021) Diagnostic accuracy of two commercial SARS-CoV-2 antigen-detecting rapid tests at the point of care in community-based testing centers. PLoS ONE 16(3): e0248921. <https://doi.org/10.1371/journal.pone.0248921>

Editor: Nei-yuan Hsiao, University of Cape Town Faculty of Health Sciences, SOUTH AFRICA

Received: January 11, 2021

Accepted: March 9, 2021

Published: March 31, 2021

Copyright: © 2021 Berger et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Some data cannot be shared publicly because data contain potentially identifying or sensitive patient information. Data is available from the Geneva cantonal Ethics Committee (contact via CCER@etat.ge.ch) for researchers who meet the criteria for access to confidential data.

Funding: The study was supported by the Foundation for Innovative New Diagnostics (<https://www.finddx.org/>), the Fondation privée des

Abstract

Objectives

Determine the diagnostic accuracy of two antigen-detecting rapid diagnostic tests (Ag-RDT) for SARS-CoV-2 at the point of care and define individuals' characteristics providing best performance.

Methods

We performed a prospective, single-center, point of care validation of two Ag-RDT in comparison to RT-PCR on nasopharyngeal swabs.

Results

Between October 9th and 23rd, 2020, 1064 participants were enrolled. The Panbio™ Covid-19 Ag Rapid Test device (Abbott) was validated in 535 participants, with 106 positive Ag-RDT results out of 124 positive RT-PCR individuals, yielding a sensitivity of 85.5% (95% CI: 78.0–91.2). Specificity was 100.0% (95% CI: 99.1–100) in 411 RT-PCR negative individuals. The Standard Q Ag-RDT (SD Biosensor, Roche) was validated in 529 participants, with 170 positive Ag-RDT results out of 191 positive RT-PCR individuals, yielding a sensitivity of 89.0% (95%CI: 83.7–93.1). One false positive result was obtained in 338 RT-PCR negative individuals, yielding a specificity of 99.7% (95%CI: 98.4–100). For individuals presenting

HUG (<https://www.fondationhug.org/>) and Pictet Charitable Foundation (<https://www.group.pictet/responsible-vision/pictet-group-foundation>). M.T. Ngo Nsoga is a beneficiary of the excellence grant from the Swiss confederation (<https://www.sbf.admin.ch/sbf/en/home/education/scholarships-and-grants/swiss-government-excellence-scholarships.html>) and the grant from the humanitarian commission of the university hospitals of Geneva (<https://www.hug.ch/en/external-affairs-directorate>). The Foundation for Innovative New Diagnostics provided rapid tests and elements of the study protocol. The funders had no other roles in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

with fever 1–5 days post symptom onset, combined Ag-RDT sensitivity was above 95%. Lower sensitivity of 88.2% was seen on the same day of symptom development (day 0).

Conclusions

We provide an independent validation of two widely available commercial Ag-RDTs, both meeting WHO criteria of $\geq 80\%$ sensitivity and $\geq 97\%$ specificity. Although less sensitive than RT-PCR, these assays could be beneficial due to their rapid results, ease of use, and independence from existing laboratory structures. Testing criteria focusing on patients with typical symptoms in their early symptomatic period onset could further increase diagnostic value.

Introduction

The Coronavirus Disease 19 (COVID-19) pandemic, caused by SARS-CoV-2, has led to an unprecedented public health crisis. Diagnostic strategies that are low cost, rapid, and easily accessible are critical to control the pandemic. While RT-PCR based assays remain the standard for the detection of emerging respiratory viruses [1, 2], the need for high through-put virus detection has fueled development of Antigen-detecting rapid diagnostic tests (Ag-RDT), that can be performed at the point of care (POC) [3].

Such laboratory-independent tests could allow for control of the pandemic by quickly isolating individuals during their contagious period to prevent further transmission. These tests can affordably help overcome overwhelmed diagnostic laboratories and global PCR-reagent shortages [4, 5]. Given that viral load, as measured by RNA copies, peaks near symptom onset [6–8] and contagiousness begins even earlier [9, 10], RDTs may have the highest sensitivity (SN) in the most contagious individuals. Reported SNs of Ag-RDTs vary widely, and manufacturer reported SNs are often substantially higher than those of independent assessments [11]. Previous studies with NPS rapid antigenic tests (Panbio) for detection of SARS-CoV-2 shows the sensitivity of 95% for Ct ≤ 25 , and 85% for Ct < 30 in symptomatic patients and the sensitivity near of 80% for a period of less than 7 days from symptom onset but the overall sensitivity in this population seems low at 67.3% [12]. A study on two NPS RDT (Panbio and SD Biosensor) versus PCR in symptomatic population found an overall sensitivity of 73.8% and 68.5%; for a CT value < 30 , sensitivities of two RDT are respectively 87% and 81.4% in a population with mostly symptomatic patients [13]. Lambert-Niclot et al. found a sensitivity of 50.0% with RDT NPS (Respi-Strip) versus PCR and for the CT value < 25 CT, sensitivity was 82.2% [14].

The World Health Organization's (WHO) Ag-RDT target product profile aims at SNs $\geq 80\%$ and specificities (SP) of $\geq 97\%$ [5, 15]. Thus, we sought to evaluate the performance of two commercially available Ag-RDTs through a prospective, single-center POC validation in comparison to RT-PCR for detecting SARS-CoV-2 using nasopharyngeal swabs (NPS). We evaluate also the performance of these tests according to the duration of symptoms and viral loads of patients mostly symptomatic.

Methods

Ethics

The study was approved by the Cantonal ethics committee (Nr. 2020–02323). All study participants and/or their legal guardians provided written informed consent.

Setting

The study was performed in two geographically different testing centres run by our institution, the Geneva University Hospital. Both centres are supervised by the same team, and did not differ in their infrastructure, so we analyzed them as a single centre study.

Study design and participants

The primary objective of this prospective study was to assess the diagnostic accuracy (SN and SP) of the Ag-RDTs compared to the reference RT-PCR. Participants were ≥ 16 years old, with suspected SARS-CoV-2 infection according to the local governmental testing criteria. This included suggestive symptoms for COVID-19 and/or recent exposure to a SARS-CoV-2 positive person. Asymptomatic individuals were included if they were notified by the Swiss COVID-19 app or by local health authorities after contact to a confirmed case.

Study procedures

For each participant, two NPS were collected. The first was a standard flocked swab placed in viral transport media (VTM), used routinely for viral genome detection by RT-PCR. The second NPS, provided in the Ag-RDT kit, was obtained from the contralateral nostril and was performed as recommended by the manufacturer. Both swabs were taken by the same trained nurse. All Ag-RDTs were performed immediately at the sample collection site. Adequate personal protective equipment was used while collecting the NPSs and performing the RDTs.

Data collection

Clinical data were collected for each patient upon presentation with a questionnaire including the number of days post symptom onset (DPOS), known contact to a previous SARS-CoV-2 infected person, comorbidities and type of symptoms. The following symptoms were recorded: rhinorrhea (runny nose), odynophagia, myalgia, chills, dry cough, productive cough, red expectoration, fever (anamnestic), anosmia/ageusia (loss of smell or taste), gastrointestinal symptoms, asthenia, dyspnea, thoracic pain and headache.

Comorbidities included in the questionnaire were hypertension, cardiovascular disease, chronic pulmonary disease, diabetes, chronic renal failure, active cancer including lymphoproliferative disease, severe immunosuppression, immunosuppressive therapy, pregnancy, and obesity (BMI >40 kg/m²).

Ag-RDT testing

The two validated Ag-RDTs were Panbio™ COVID-19 Ag Rapid Test Device (Abbott Rapid Diagnostics, US) and Standard™ Q COVID-19 Ag Test (SD Biosensor, distributed by Roche, Switzerland). Both Ag-RDTs were used as recommended by the manufacturers, using only materials provided in the kit. Both assays were manually read, with two individuals reading the results separately. In case of discordant results, the two validators sought a consensus. All visible bands were considered a positive result. All Ag-RDT results were photographically documented.

RT-PCR testing

All participants were tested by a single well, dual target RT-PCR assay for SARS-CoV-2 (cobas® SARS-CoV-2 Test, Cobas 6800, Roche, Switzerland) using NPS in 3mL VTM. For further analysis, only cycle threshold (Ct) values for the E-gene were used. For calculation of viral loads (VL) as SARS-CoV-2 genome copy numbers per mL, a standard curve was obtained

by using a quantified supernatant from a cell culture isolate of SARS-CoV-2. All VLs were calculated from the Ct-values, according to $\log_{10} \text{ SARS-CoV-2 RNA copies/ml} = (\text{Ct}-44.5)/-3.3372$ for Cobas [16, 17].

Statistics

Over a 2-week period, we enrolled all patients who met the SARS-CoV-2 testing criteria; 535 and 529 patients were enrolled during the first and second week, respectively. The target sample size was 530, as it would have sufficient power to generate a 95% confidence interval (CI) with a lower bound above the WHO target of 80%, if the prevalence was 25% and the measured SP was $\geq 87.5\%$.

All continuous variables were presented by their mean \pm standard deviation (SD) and median (interquartile range, IQR), categorical variables were presented by their frequencies and relative proportions. For comparisons of continuous variables, we used a nonparametric Mann-Whitney test due to small sizes; for comparisons of categorical variables, we either performed χ^2 or Fischer's exact tests, depending on applicability.

To enable Ag-RDT result combination, we performed a Bayesian t-test on their sensitivities and specificities. To be able to conduct the t-test, the confidence intervals of both sensitivities and specificities were converted into standard deviation to allow for the t-test to be conducted. The test computes a Bayesian Factor (BF) that allows comparison of the probability of observing our data under H_0 (both tests are equal in term of SN and SP) and H_1 (both tests are different). All analyses were performed using STATA version intercooled 16 (Stata Corp., College Station, TX, USA). Statistical significance was defined as $p < 0.05$ (two-sided).

Results

Between October 9th and October 23rd, 2020, 1064 participants were enrolled and included in the analysis. 535 participants were tested with the Panbio Ag-RDT from October 09th to 16th and 529 participants were tested with the Standard Q Ag-RDT from October 19th to 23rd, 2020.

Characteristics of the study population are shown in [Table 1](#). The mean age of the study participants was 34.9 years (SD ± 10.9) with 53.8% being female. The mean DPOS to testing was 2.7 (SD ± 1 day). Overall, 29.6% of participants were positive by RT-PCR with a mean Ct-value of 22.5 (SD ± 5.1), corresponding to a VL of 1.8E7 SARS-CoV-2 copies/mL. Most patients

Table 1. Characteristics of the study population.

Characteristics	Standard Q	Panbio	Combined	p-value
	(n = 529)	(n = 535)	(n = 1064)	
Mean age (\pm SD, median)	34.9 (± 10.9 , 33)	38.5 (± 13.6 , 36)	36.7 (± 12.5 , 34)	<0.001
Sex distribution, n (%)				0.966
Women	285 (53.9)	287 (53.6)	572 (53.8)	
Men	244 (46.1)	248 (46.4)	492 (46.2)	
Mean DPOS to RT-PCR (\pm SD, median)	2.9 (± 1.5 , 3)	2.6 (± 2.0 , 2)	2.7 (± 1.9 , 2)	0.0125
Result of RT-PCR, n (%)				<0.001
Negative	338 (63.9)	411 (76.8)	749 (70.4)	
Positive	191 (36.1)	124 (23.2)	315 (29.6)	
Mean Ct (\pm SD, median) (n = 315)	22.6 (± 4.9 , 21.8)	22.4 (± 5.4 , 21.0)	22.5 (± 5.1 , 21.5)	0.450

SD, standard deviation; RT-PCR, reverse transcription polymerase chain reaction; DPOS, days post symptom onset; Ct, cycle threshold.

<https://doi.org/10.1371/journal.pone.0248921.t001>

Table 2. Overall SN, SP, positive and negative predictive value of Standard Q and Panbio SARS-CoV-2 Ag-RDT.

Characteristics	Standard Q	Panbio	Combined
SN, % (95%CI)	89.0 (83.7–93.1)	85.5 (78.0–91.2)	87.6 (83.5–91.0)
SP, % (95%CI)	99.7 (98.4–100)	100 (99.1–100)	99.9 (99.3–100)
Positive predictive value, % (95%CI)	99.4 (96.8–100)	100 (96.6–100)	99.6 (98.0–100)
Negative predictive value, % (95%CI)	94.1 (91.2–96.3)	95.8 (93.4–97.5)	95 (93.3–96.5)

Positivity rate at the time of study for Standard Q was 36.1% and at the time of study for Panbio was 23.2%.

<https://doi.org/10.1371/journal.pone.0248921.t002>

(97.8%) were symptomatic upon presentation at the testing centre, with only 3 reporting no symptoms. Symptoms information was missing for 4 patients. The study population tested with the Standard Q was younger than that tested with the Panbio assay (34.9 ± 10.9 vs 38.5 ± 13.6 years, respectively, $p < 0.001$) and DPOS differed slightly (2.9 ± 1.5 vs 2.6 ± 2.0 days, respectively, $p = 0.0125$). Ct-values did not differ significantly between the two groups ($p = 0.450$): 22.6 (SD ± 4.9) in the Standard Q group vs. 22.4 (SD ± 5.4) in the Panbio group, corresponding to $1.7E7$ and $1.9E7$ SARS-CoV-2 RNA copies/mL, respectively. The RT-PCR positivity rate was 23.2% and 36.1% for the population tested with the Panbio and the Standard Q, respectively, corresponding to an increase in the overall PCR positivity rate and reflecting the rapidly increasing local incidence during the time of this study.

The overall test performance for the Standard Q was 89.0% SN (95%CI: 83.7–93.1) and 99.7% SP (95%CI: 98.4–100%). The overall test performance for the Panbio assay was 85.5% SN (95%CI: 78.0–91.2) and 100% SP (95%CI: 99.1–100) (Table 2). The absolute numbers of positive and negative detections per method (by RT-PCR and Ag-RDT) and number of true positive, false negatives, false positives and true negatives can be found in Table 3.

Ct-values of samples with positive Ag-RDT results ranged from 14.2–34.0 and 14.4–34.2 for Panbio and Standard Q ($p = 0.1766$), respectively, while Ct-values of samples of samples that tested falsely negative by Ag-RDT ranged from 16.0–39.7 and 19.8–37.4 ($p = 0.7998$), respectively. Median Ct-values of Ag-RDT positive samples (Panbio: 20.4, IQR: 18.1–23.8; Standard Q: 21.2, IQR 18.6–24) were lower than those of Ag-RDT negative samples (Panbio: 30.5, IQR: 27–35.9; Standard Q: 30.4, IQR: 25.7–33.9) (Fig 1). Furthermore, we evaluated overall Ag-RDT results in relation to Ct-values/viral load as well DPOS (Fig 2). False-negative results occurred in both assay across all DPOS.

We compared SN and SP between the two Ag-RDTs and concluded that we could accept, with high probabilities, (respectively likelihood ratio of $BF_{01} = 10.2$ and 11.9) the hypothesis of equivalent SN and SP. Based on this, a combined SN of 87.6% (95%CI: 83.5–91.0) and a combined SP of 99.9% (95%CI: 99.3–100) for both Ag-RDTs were calculated with a positive predictive value of 99.6% (95%CI: 98.0–100) and a negative predictive value of 95.0% (95%CI: 93.3–96.5). In order to identify subpopulations in which maximal SN could be reached with these tests, we analyzed SN by DPOS, Ct-values as determined by RT-PCR, type of symptoms, comorbidities, and previous contact with a confirmed SARS-CoV-2 infection.

Table 3. RT-PCR and Ag-RDT results.

	Total	Ag-RDT +/ RT-PCR + (TP)	Ag-RDT –/RT-PCR + (FN)	Ag-RDT +/ Rt-PCR – (FP)	Ag-RDT –/RT-PCR – (TN)
Standard Q	529	170	21	1	337
Panbio	535	106	18	0	411

+, positive; -, negative; TP, true positive; FN, false negative; FP, false positive.

<https://doi.org/10.1371/journal.pone.0248921.t003>

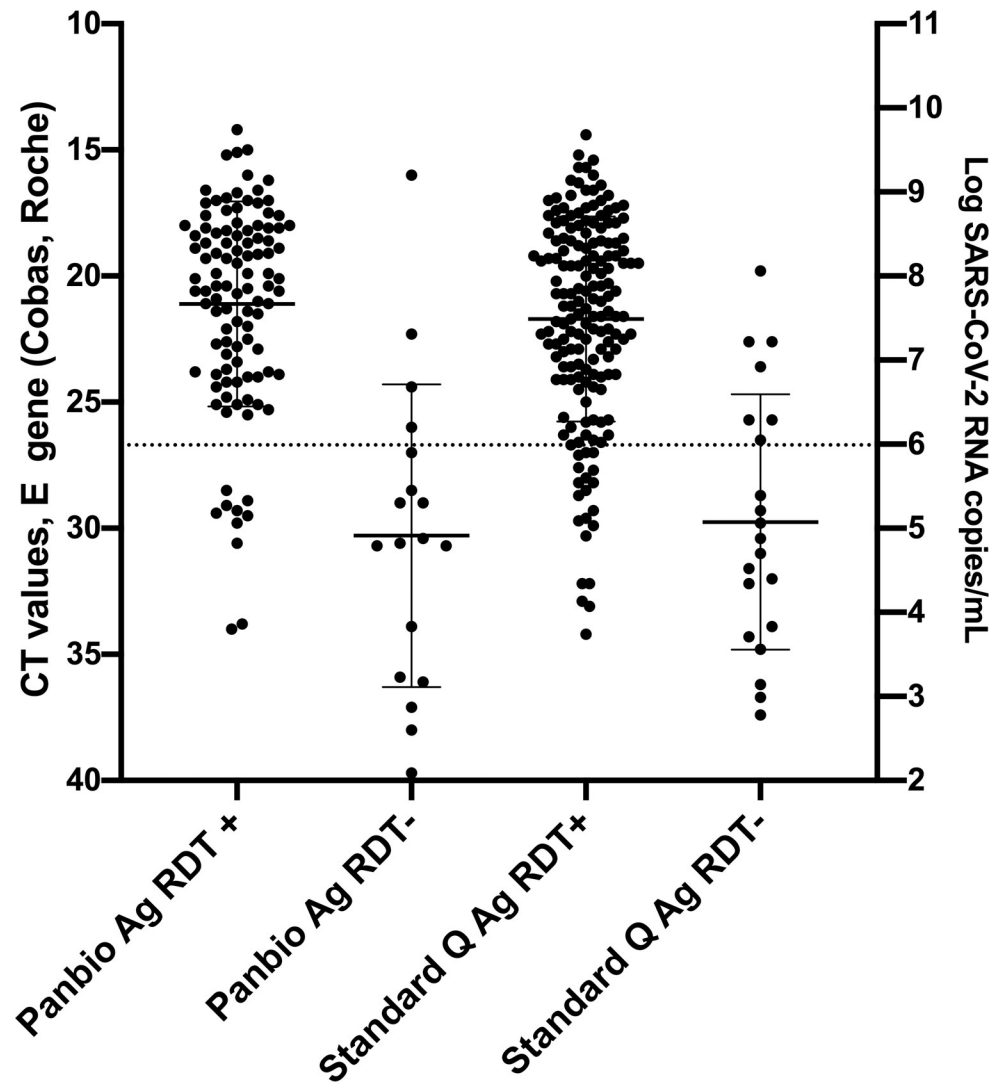


Fig 1. Ct values, viral load and Ag-RDT results for RT-PCR-positive individuals tested with Standard Q (n = 191) and Panbio (n = 124). Horizontal bars represent median and standard deviation. Dotted line: Ct value of 26.7 or $1E6$ SARS-CoV-2 RNA copy numbers/mL.

<https://doi.org/10.1371/journal.pone.0248921.g001>

Combined SN varied according to Ct-values: it was highest in samples with low Ct-values, with a SN of 98.4% (95% CI: 94.2–99.8) for $Ct \leq 20$ ($\leq 1.0E8$ SARS-CoV-2 copies/mL), decreased slightly to 95.5% (95% CI: 89.9–98.5) for $20 < Ct \leq 25$ ($< 1.0E8$ SARS-CoV-2 copies/mL $\leq 3.2E6$), dropped further to 89.9% (95% CI: 86.0–93.0) for $Ct \leq 35$ ($\leq 3.2E3$ SARS-CoV-2 copies/mL) and was lowest (only 40.9% (95% CI: 20.7–63.6)) for $30 < Ct \leq 35$ ($< 1.0E5$ SARS-CoV-2 copies/mL $\leq 3E3$) (Fig 3). The SN for all samples with a Ct value ≤ 26.7 ($\leq 1E6$ SARS-CoV-2 copies/mL), an assumed cut-off for presence of infectious virus, was 95.7% (95% CI: 92.4–97.8).

SN increased with DPOS, from 88.2% at 0 DPOS (95% CI: 63.6–98.5) to 94.3% (95% CI: 84.3–98.8, $p = 0.030$) at 1 DPOS, and remained high until 5 DPOS. The highest SN was seen between 1 DPOS and 4–5 DPOS, ranging from 94.3% (95% CI: 84.3–98.8) to 94.8% (95% CI: 85.6–98.9), with a decline after 5 DPOS (Fig 4A).

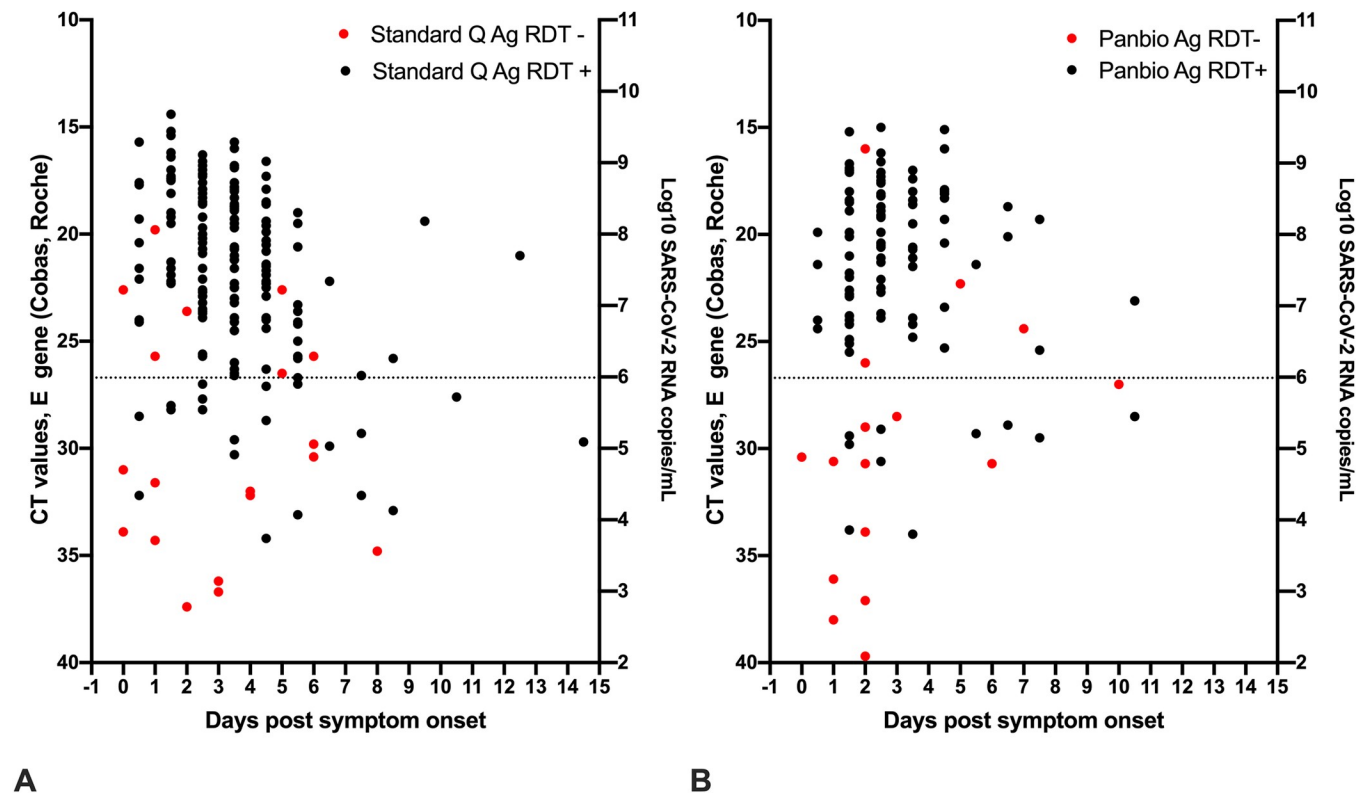


Fig 2. Ct values, viral load, days post symptom onset and Ag-RDT results for 190 patients tested with Standard Q (A) and 116 patients tested with Panbio (B) for which information on day of symptom onset was available. Dotted line: Ct value of 26.7 or 1E6 SARS-CoV-2 RNA copy numbers/mL.

<https://doi.org/10.1371/journal.pone.0248921.g002>

Additionally, we analyzed SN according to specific symptoms, differentiating between typical COVID-19 symptoms (fever/chills, cough and anosmia/ageusia) and more non-specific symptoms of respiratory infection (all other symptoms reported). The highest SN of 93.8% (87.0–97.7) was observed for patients presenting with fever/chills and cough at the time of testing, followed by patients presenting with anosmia/ageusia or cough and fever/chills with a SN of 93.7% (95%CI: 87.4–97.4), but only 73.8% (95%CI: 58.0–86.1) in patients presenting with non-specific signs (Fig 4B). No difference in SN was seen between patients with (89.3%, 95% CI: 71.8–97.7, $n = 28$) or without (87.5%, 95%CI: 83.1–91.1, $n = 287$) comorbidities ($p = 0.999$). Typical symptoms were more frequent in patients with comorbidities (100%, 15/15 patients) than in patients without comorbidities (86.5%, 96/138 patients) ($p = 0.012$), however sample size was small. No difference was seen in patients with or without contact with a recently positive case ($p = 0.065$). We further analysed by DPOS, and found that the highest SN was seen in patients with fever/chills and presenting between 1 and 5 DPOS, at 95.7% SN (95%CI: 91.0–98.4).

Discussion

This study provides an independent, POC validation of two commercial Ag-RDTs relative to RT-PCR and according to demographic and clinical information. This combined validation of two similar assays provides performance data in a real-life high incidence test setting.

Both RDTs performed well with an overall SN of 87.6% (95%CI: 83.5–91) and a very high SP of 99.9% (95%CI: 99.3–100) during a time of very high SARS-CoV-2 weekly incidence (375/100,000 to 824/100,000 inhabitants) and a SARS-CoV-2 RT-PCR positivity rate >20%.

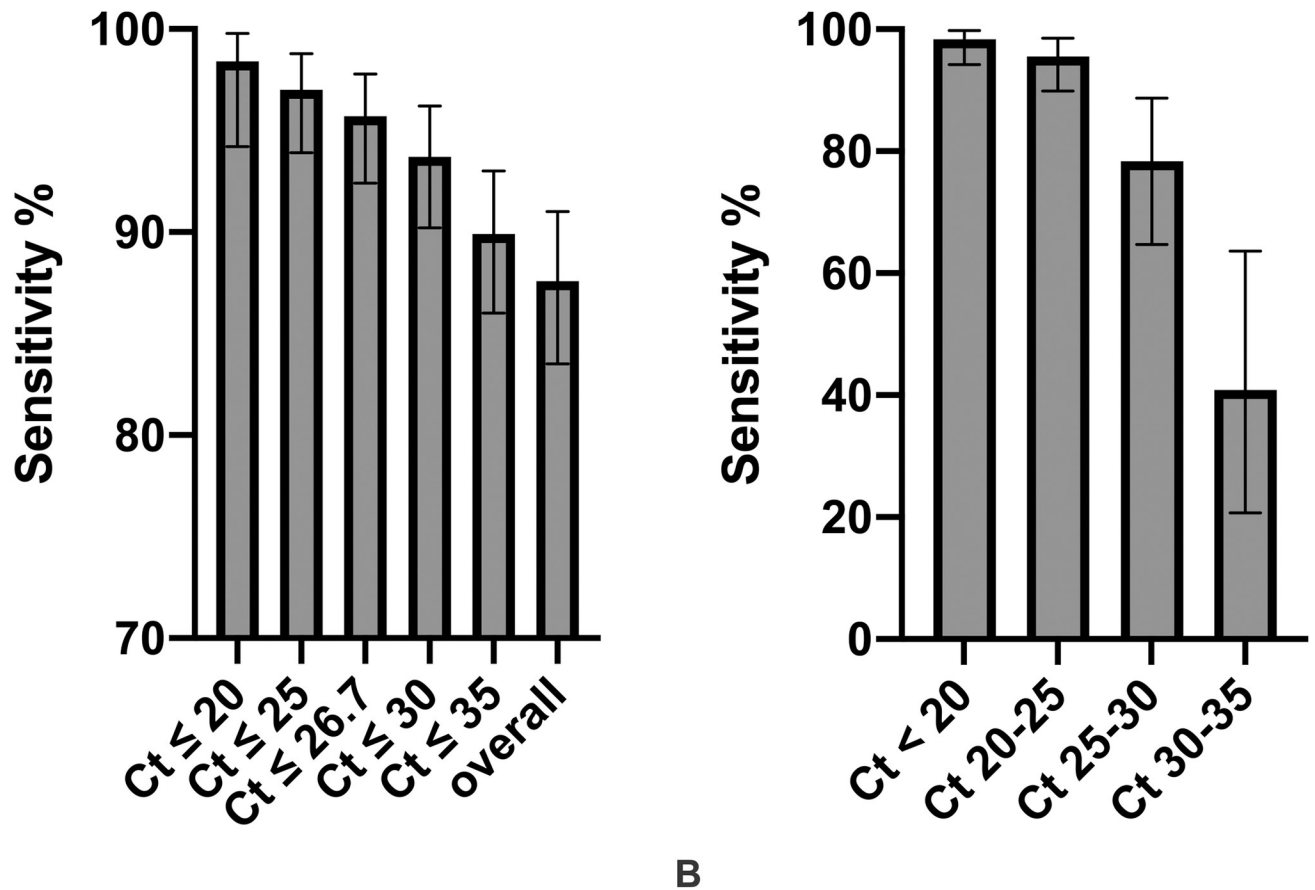


Fig 3. A. Combined SN of the two Ag-RDTs according to Ct-values of the RT-PCR. B. Combined SN of the two Ag-RDTs according to subgroups of Ct-values of the RT-PCR. Ct values correspond to the following SARS-CoV-2 RNA copy numbers/mL: Ct 20: 1.0E8; Ct 25: 3.2E6; Ct 26.7: 1E6; Ct 30: 1.0E5, Ct 35: 3.2E3.

<https://doi.org/10.1371/journal.pone.0248921.g003>

SN was higher in sub-populations with earlier DPOS numbers and characteristic COVID-19 symptoms. Importantly at 0 DPOS, the SN is lower than during the subsequent days, and the sensitivity drops rapidly when the Ct-values increase above a threshold of 30, mostly after 6–7 days.

The highest VL and thus transmission probability occurs within the first week of symptom onset, with VLs peaking around the time of symptom onset [6]. Culturable virus has been predominantly found in the first week after symptom onset, down to a VL around 1E6 copies/ml [17–20], which has been set as a detection cut-off for Ag-RDTs by the WHO [15]. The SN of the Ag-RDTs validated here, for VLs compatible with contagiousness, was 95.7%. Correspondingly, the highest Ag-RDT SN was also observed at early DPOS numbers and in patients with low Ct-values, again suggesting reliable identification of contagious individuals.

Our findings at the POC are in line with other validations performed in different countries and prevalences, although study designs and specimens used varied considerably between studies. Standard Q was reported to have SNs between 70.6–88.7%, while SP remained high throughout these studies between 97.6–100% [21–27]. A clinical study performed similarly to ours in a much lower-incidence setting (<1% RT-PCR positivity rate), found a SN/SP of 76.6%/100%, using a mixture of NPS and combined oro- and naso-pharyngeal swabs from a total of 2417 participants with 47 RT-PCR positive samples yielding 36 Ag-RDT [21].

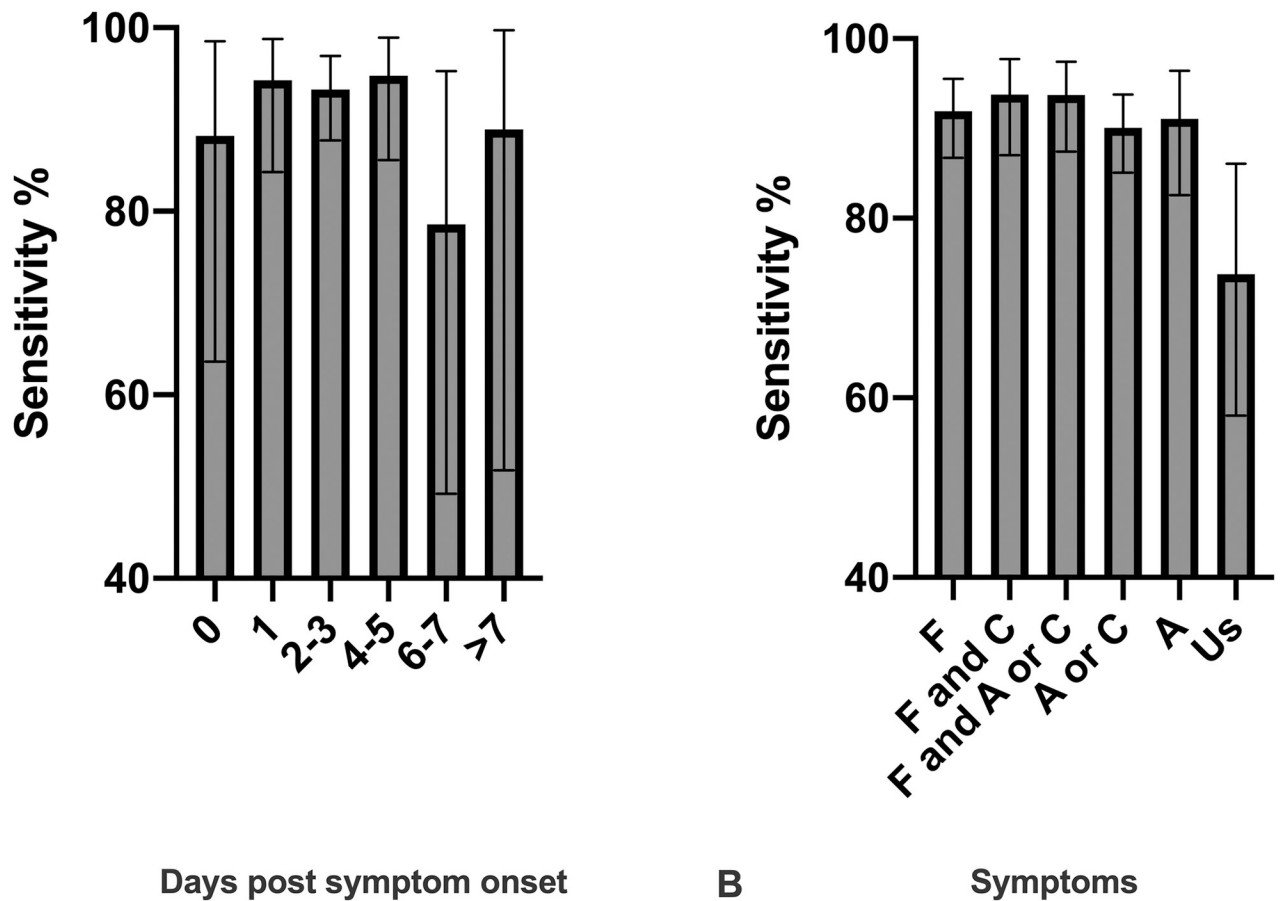


Fig 4. A. Combined SN of the two Ag-RDTs according to days post symptom onset. Number of patients per category: Day 0, n = 17; day 1, n = 53, day 2–3, n = 135; day 4–5, n = 58; day 6–7, n = 14; > 7 days, n = 9. **B.** Combined SN of the two Ag-RDTs according to symptoms F, fever/chills; C, cough, A, anosmia/ageusia (loss of smell or taste), Us, unspecific symptoms (all other symptoms excluding fever/chills, cough and anosmia/ageusia). Number of patients per category: F, n = 172; F and C, n = 97; F and A or C, n = 111; A or C, n = 202; A, n = 79; Us, n = 42.

<https://doi.org/10.1371/journal.pone.0248921.g004>

For Panbio, other studies have reported SNs ranging from 73.3–91.7% with SP in the range of 94.9–100% [28–31]. Notably, the highest reported SN of 91.7%/98.9% comes from a study, not done at a POC, using frozen NPS specimens [28]. Although the use of frozen samples is possible for RDTs [22, 28], it is not their intended use and does not represent their use in the field. It is also unknown if a freeze-thaw cycle can affect the accessibility of viral antigens.

While significant variation in Ag-RDT SN is observed across studies, there is remarkable similarity within certain Ct-value ranges—although caution must be exercised comparing Ct-values of different assays. For Ct-values of <25, the Standard Q test was reported to have a SN of 100% [21], while the Panbio was reported to have a SN of 97.1% [30] or 98.2% [28], which is in agreement with our results of 97%. In contrast to most other validations, we did observe false-negative Ag-RDT results in patients with high VLs across a range of DPOS. These patients are likely contagious and able to transmit SARS-CoV-2, with VLs associated with culturable virus.

We did not find any validation of Ag-RDTs that has analyzed SN based on type of symptoms, which could be an additional factor for testing algorithms. This finding also highlights that Ag-RDTs are a valid diagnostic tool in symptomatic individuals, while the benefit of their use in asymptomatic individuals or in patients with atypical or minor symptoms remains to be

investigated. In a situation with sufficient RT-PCR capacity, testing algorithms focusing on a subgroup of symptomatic patients, while testing others with more sensitive RT-PCR might be an option to make the best use of Ag-RDTs. In our data set, the best SN is found in symptomatic individuals with symptoms suggestive of COVID-19, between 1 and 5 DPOS.

Our study has several strengths. The Ag-RDTs were performed at the POC in parallel to RT-PCR, and is one of the largest in terms of RT-PCR positive individuals. The test population represents a population screened for public health intervention, and not for diagnostic purposes in a hospital setting: mainly young unhospitalized adults without comorbidities, who mostly had typical and mild COVID-19 symptoms. This currently describes the majority of SARS-CoV-2 infected individuals, and an important group for limiting community transmission.

Although Ag-RDTs are less sensitive than RT-PCR and, we saw false-negative Ag-RDT results in patients with high VL, the public health benefit of quickly identifying a large proportion of infected individuals would still outweigh the disadvantages of occasional missed diagnoses [32].

Furthermore, our validation showed very high SP, with only one false-positive Ag-RDT result in the whole study. Interestingly, the patient with this putative false-positive tested positive 3 days later for SARS-CoV-2 by RT-PCR. This study was conducted in a high prevalence setting, thus extrapolating the findings of our study to low prevalence settings must be done with caution.

In conclusion, we show good diagnostic accuracy of both Ag-RDTs, especially for rule-in purposes of infected individuals and in patients with certain criteria. The SN for identification of SARS-CoV-2 infections, rapidity of results, and the laboratory-independence make these Ag-RDTs promising tools for SARS-CoV-2 infection control.

Acknowledgments

We thank all nurses and staff at the testing Centre Sectors of our institution as well as the patients for their willingness to participate in the study. We thank Catia Machado-Delgado for excellent technical assistance and Stéphanie Baggio for help with data analysis.

Author Contributions

Conceptualization: Erik Boehm, Ilona Kronig, Jilian A. Sacks, Margaretha de Vos, François Chappuis, Adriana Renzoni, Laurent Kaiser, Manuel Schibler, Isabella Eckerle.

Data curation: Francisco Javier Perez-Rodriguez, Frédérique Jacqueroiz Bausch.

Formal analysis: Alice Berger, Marie Therese Ngo Nsoga, Francisco Javier Perez-Rodriguez, Pascale Sattonnet-Roche, Angèle Gayet-Ageron, Cyril Jaksic, Frédérique Jacqueroiz Bausch, Manuel Schibler, Isabella Eckerle.

Investigation: Alice Berger, Marie Therese Ngo Nsoga, Yasmine Abi Aad, Pascale Sattonnet-Roche, Giulia Torriani, Manuel Schibler.

Methodology: Jilian A. Sacks, Margaretha de Vos, Laurent Kaiser, Manuel Schibler, Isabella Eckerle.

Project administration: Laurent Kaiser.

Resources: Marie Therese Ngo Nsoga, Jilian A. Sacks, Margaretha de Vos.

Validation: Alice Berger, Marie Therese Ngo Nsoga, Angèle Gayet-Ageron, Manuel Schibler, Isabella Eckerle.

Visualization: Francisco Javier Perez-Rodriguez.

Writing – original draft: Erik Boehm, Laurent Kaiser, Manuel Schibler, Isabella Eckerle.

Writing – review & editing: Alice Berger, Marie Therese Ngo Nsoga, Francisco Javier Perez-Rodriguez, Yasmine Abi Aad, Pascale Sattonnet-Roche, Angèle Gayet-Ageron, Cyril Jaksic, Giulia Torriani, Erik Boehm, Ilona Kronig, Jilian A. Sacks, Margaretha de Vos, Frédérique Jacquerioz Bausch, François Chappuis, Adriana Renzoni, Laurent Kaiser, Isabella Eckerle.

References

1. Corman VM, Eckerle I, Bleicker T, Zaki A, Landt O, Eschbach-Bludau M, et al. Detection of a novel human coronavirus by real-time reverse-transcription polymerase chain reaction. *Eurosurveillance*. 2012 Sep 27; 17(39):20285. <https://doi.org/10.2807/ese.17.39.20285-en> PMID: 23041020
2. Drosten C, Günther S, Preiser W, van der Werf S, Brodt H-R, Becker S, et al. Identification of a Novel Coronavirus in Patients with Severe Acute Respiratory Syndrome. *New England Journal of Medicine*. 2003 May 15; 348(20):1967–76. <https://doi.org/10.1056/NEJMoa030747> PMID: 12690091
3. Guglielmi G. The explosion of new coronavirus tests that could help to end the pandemic. *Nature*. 2020 Jul; 583(7817):506–9. <https://doi.org/10.1038/d41586-020-02140-8> PMID: 32681157
4. Mina MJ, Parker R, Larremore DB. Rethinking Covid-19 Test Sensitivity—A Strategy for Containment. *N Engl J Med*. 2020 Sep 30; Epub ahead of print. <https://doi.org/10.1056/NEJMp2025631> PMID: 32997903
5. Antigen-detection in the diagnosis of SARS-CoV-2 infection using rapid immunoassays [Internet]. 2020 [cited 2020 Oct 8]. Available from: <https://www.who.int/publications-detail-redirect/antigen-detection-in-the-diagnosis-of-sars-cov-2infection-using-rapid-immunoassays>
6. He X, Lau EHY, Wu P, Deng X, Wang J, Hao X, et al. Temporal dynamics in viral shedding and transmissibility of COVID-19. *Nature Medicine*. 2020 May; 26(5):672–5. <https://doi.org/10.1038/s41591-020-0869-5> PMID: 32296168
7. Zou L, Ruan F, Huang M, Liang L, Huang H, Hong Z, et al. SARS-CoV-2 Viral Load in Upper Respiratory Specimens of Infected Patients. *New England Journal of Medicine*. 2020 Mar 19; 382(12):1177–9. <https://doi.org/10.1056/NEJMc2001737> PMID: 32074444
8. Walsh KA, Jordan K, Clyne B, Rohde D, Drummond L, Byrne P, et al. SARS-CoV-2 detection, viral load and infectivity over the course of an infection. *J Infect*. 2020 Jun 29; 81(3):357–71. <https://doi.org/10.1016/j.jinf.2020.06.067> PMID: 32615199
9. Lauer SA, Grantz KH, Bi Q, Jones FK, Zheng Q, Meredith HR, et al. The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. *Ann Intern Med*. 2020 Mar 10; 172(9):577–82. <https://doi.org/10.7326/M20-0504> PMID: 32150748
10. Byrne AW, McEvoy D, Collins AB, Hunt K, Casey M, Barber A, et al. Inferred duration of infectious period of SARS-CoV-2: rapid scoping review and analysis of available evidence for asymptomatic and symptomatic COVID-19 cases. *BMJ Open*. 2020 Aug 5; 10(8):e039856. <https://doi.org/10.1136/bmjopen-2020-039856> PMID: 32759252
11. Walle IV, Leitmeyer K, Broberg EK, Group TEC-19 microbiological laboratories. Meta-analysis of the clinical performance of commercial SARS-CoV-2 nucleic acid, antigen and antibody tests up to 22 August 2020. *medRxiv*. 2020 Sep 18;2020.09.16.20195917.
12. Masiá M, Fernández-González M, Sánchez M, Carvajal M, García JA, Gonzalo N, et al. Nasopharyngeal Panbio COVID-19 antigen performed at point-of-care has a high sensitivity in symptomatic and asymptomatic patients with higher risk for transmission and older age. *Open Forum Infect Dis* [Internet]. [cited 2021 Feb 24]; Available from: <https://academic.oup.com/ofid/advance-article/doi/10.1093/ofid/ofab059/6126749>
13. Drevinek P, Hurych J, Kepka Z, Briksi A, Kulich M, Zajac M, et al. The sensitivity of SARS-CoV-2 antigen tests in the view of large-scale testing. *medRxiv*. 2020 Nov 24;2020.11.23.20237198.
14. Lambert-Niclot S, Cuffel A, Pape SL, Vauloup-Fellous C, Morand-Joubert L, Roque-Afonso A-M, et al. Evaluation of a Rapid Diagnostic Assay for Detection of SARS-CoV-2 Antigen in Nasopharyngeal Swabs. *Journal of Clinical Microbiology* [Internet]. 2020 Jul 23 [cited 2020 Oct 8]; 58(8). Available from: <https://jcm.asm.org/content/58/8/e00977-20> <https://doi.org/10.1128/JCM.00977-20> PMID: 32404480
15. COVID-19 Target product profiles for priority diagnostics to support response to the COVID-19 pandemic v.1.0 [Internet]. 2020 [cited 2020 Oct 8]. Available from: <https://www.who.int/publications/m/item/covid-19-target-product-profiles-for-priority-diagnostics-to-support-response-to-the-covid-19-pandemic-v.0.1>
16. Baggio S, L'Huillier AG, Yerly S, Bellon M, Wagner N, Rohr M, et al. SARS-CoV-2 viral load in the upper respiratory tract of children and adults with early acute COVID-19. *Clin Infect Dis* [Internet]. 2020 Aug 6

[cited 2021 Feb 12]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7454380/> <https://doi.org/10.1093/cid/ciaa1157> PMID: 32761228

17. L'Huillier AG, Torriani G, Pigny F, Kaiser L, Eckerle I. Culture-Competent SARS-CoV-2 in Nasopharynx of Symptomatic Neonates, Children, and Adolescents. *Emerg Infect Dis*. 2020 Oct; 26(10):2494–7. <https://doi.org/10.3201/eid2610.202403> PMID: 32603290
18. Wölfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Müller MA, et al. Virological assessment of hospitalized patients with COVID-2019. *Nature*. 2020 May; 581(7809):465–9. <https://doi.org/10.1038/s41586-020-2196-x> PMID: 32235945
19. Kampen JJA van, Vijver DAMC van de, Fraaij PLA, Haagmans BL, Lamers MM, Okba N, et al. Shedding of infectious virus in hospitalized patients with coronavirus disease-2019 (COVID-19): duration and key determinants. *medRxiv*. 2020 Jun 9;2020.06.08.20125310.
20. Vetter P, Eberhardt CS, Meyer B, Murillo PAM, Torriani G, Pigny F, et al. Daily Viral Kinetics and Innate and Adaptive Immune Response Assessment in COVID-19: a Case Series. *mSphere* [Internet]. 2020 Dec 23 [cited 2021 Feb 12]; 5(6). Available from: <https://msphere.asm.org/content/5/6/e00827-20> <https://doi.org/10.1128/mSphere.00827-20> PMID: 33177214
21. Krueger LJ, Gaeddert M, Koepfel L, Bruemmer L, Gottschalk C, Miranda IB, et al. Evaluation of the accuracy, ease of use and limit of detection of novel, rapid, antigen-detecting point-of-care diagnostics for SARS-CoV-2. *medRxiv*. 2020 Oct 4;2020.10.01.20203836.
22. Cerutti F, Burdino E, Milia MG, Alice T, Gregori G, Bruzzone B, et al. Urgent need of rapid tests for SARS CoV-2 antigen detection: Evaluation of the SD-Biosensor antigen test for SARS-CoV-2. *J Clin Virol*. 2020 Nov; 132:104654. <https://doi.org/10.1016/j.jcv.2020.104654> PMID: 33053494
23. Gupta A, Khurana S, Das R, Srigan D, Singh A, Mittal A, et al. Rapid chromatographic immunoassay-based evaluation of COVID-19: A cross-sectional, diagnostic test accuracy study & its implications for COVID-19 management in India. *The Indian journal of medical research*. 2020 Oct 31; Online ahead of print. https://doi.org/10.4103/ijmr.IJMR_3305_20 PMID: 33146157
24. FIND. FIND Evaluation of SD Biosensor, Inc. STANDARD™ Q COVID-19 Ag FIA External Report. 2020. FIND; 2020.
25. Chaimayo C, Kaewnaphan B, Tanlieng N, Athipanyasilp N, Sirijatuphat R, Chayakulkeeree M, et al. Rapid SARS-CoV-2 antigen detection assay in comparison with real-time RT-PCR assay for laboratory diagnosis of COVID-19 in Thailand. *Virology Journal*. 2020 Nov 13; 17(1):177. <https://doi.org/10.1186/s12985-020-01452-5> PMID: 33187528
26. Nalumansi A, Lutalo T, Kayiwa J, Watera C, Balinandi S, Kiconco J, et al. Field Evaluation of the Performance of a SARS-CoV-2 Antigen Rapid Diagnostic Test in Uganda using Nasopharyngeal Samples. *International Journal of Infectious Diseases*. 2020 Oct 30; Epub ahead of print. <https://doi.org/10.1016/j.ijid.2020.10.073> PMID: 33130198
27. Gremmels H, Winkel BMF, Schuurman R, Rosingh A, Rigter NAM, Rodriguez O, et al. Real-life validation of the Panbio™ COVID-19 antigen rapid test (Abbott) in community-dwelling subjects with symptoms of potential SARS-CoV-2 infection. *EClinicalMedicine* [Internet]. 2021 Jan 1 [cited 2021 Feb 24]; 31. Available from: [https://www.thelancet.com/journals/eclinm/article/PIIS2589-5370\(20\)30421-1/abstract](https://www.thelancet.com/journals/eclinm/article/PIIS2589-5370(20)30421-1/abstract) <https://doi.org/10.1016/j.eclinm.2020.100677> PMID: 33521610
28. Alemany A, Baró B, Ouchi D, Rodó P, Ubals M, Corbacho-Monné M, et al. Analytical and clinical performance of the panbio COVID-19 antigen-detecting rapid diagnostic test. *J Infect* [Internet]. 2021 Jan 7 [cited 2021 Feb 24]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7788317/> <https://doi.org/10.1016/j.jinf.2020.12.033> PMID: 33421447
29. Albert E, Torres I, Bueno F, Huntley D, Molla E, Fernández-Fuentes MÁ, et al. Field evaluation of a rapid antigen test (Panbio™ COVID-19 Ag Rapid Test Device) for COVID-19 diagnosis in primary healthcare centres. *Clin Microbiol Infect* [Internet]. 2020 Nov 13 [cited 2021 Feb 24]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7662075/> <https://doi.org/10.1016/j.cmi.2020.11.004> PMID: 33189872
30. Linares M, Pérez-Tanoira R, Carrero A, Romanyk J, Pérez-García F, Gómez-Herruz P, et al. Panbio antigen rapid test is reliable to diagnose SARS-CoV-2 infection in the first 7 days after the onset of symptoms. *Journal of Clinical Virology*. 2020 Oct 16; 104659. <https://doi.org/10.1016/j.jcv.2020.104659> PMID: 33160179
31. Fenollar F, Bouam A, Ballouche M, Fuster L, Prudent E, Colson P, et al. Evaluation of the Panbio Covid-19 rapid antigen detection test device for the screening of patients with Covid-19. *J Clin Microbiol*. 2020 Nov 2; JCM.02589-20.
32. Larremore DB, Wilder B, Lester E, Shehata S, Burke JM, Hay JA, et al. Test sensitivity is secondary to frequency and turnaround time for COVID-19 surveillance. *medRxiv* [Internet]. 2020 Sep 8 [cited 2020 Nov 18]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7325181/> <https://doi.org/10.1101/2020.06.22.20136309> PMID: 32607516