

MINI-SYMPOSIUM

Genomic mechanisms in Alzheimer's diseaseLars Bertram^{1,2*}  ; Rudolph E. Tanzi³ ¹ Lübeck Interdisciplinary Platform for Genome Analytics (LIGA), Institutes of Neurogenetics and Cardiogenetics, University of Lübeck, Lübeck, Germany.² Centre for Lifespan Changes in Brain and Cognition, Department of Psychology, University of Oslo, Oslo, Norway.³ McCance Center for Brain Health and Genetics and Aging Research Unit, Department of Neurology, Massachusetts General Hospital and Harvard Medical School, Charlestown, MA.**Keywords**

Alzheimer's disease, genomics, GWAS, risk genes.

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Abstract

Alzheimer's disease (AD) is the most common neurodegenerative disease and, owing to its increasing prevalence, represents one of the leading public health problems in aging populations. The molecular causes underlying the onset and progression of AD are manifold and hitherto still incompletely understood. Research over nearly four decades has clearly delineated genetics to play a crucial role in AD susceptibility, likely in concert with non-genetic factors. The field has gained considerable momentum and novel insights over the past 10 years owing to the advent and application of high-throughput genomics technologies in datasets of increasing size. In this contribution to the Mini-Symposium on the Molecular Etiology of Alzheimer's Disease, we review the current status of genomics research in AD. To this end, we scrutinize and discuss the main findings from the two largest and most current genome-wide association studies (GWAS) in the field, that is, the papers published by Jansen *et al* (Nat Genet 51:404–413) and Kunkle *et al* (Nat Genet 51:414–430). Particular focus is laid on genomics findings overlapping across both studies and on the novel insights they provide in terms of improving our understanding of the “genomic mechanisms” underlying this devastating disease.

INTRODUCTION

Alzheimer's disease (AD) is the most common neurodegenerative disorder in humans and is characterized by progressive decline in cognitive functioning ultimately leading to dementia and death. Pathogenetically, AD is triggered by the aberrant deposition of β -amyloid and tau protein, leading to the appearance of senile plaques, formation of neurofibrillary tangles, neuroinflammation, synaptic dysfunction, neuronal loss, and, ultimately, onset of cognitive decline. The molecular events causing neuronal cell death typically precede the onset of cognitive symptoms by a decade or more (23), implying that, once available, therapeutics targeting neuropathology will need to be administered very early, ideally prior to the onset of symptoms. Hence, the identification of genetic risk factors will not only be crucial for furthering our understanding of the molecular mechanisms causing the disease but will also be essential for an “early prediction—early detection—early intervention” approach to preventing the onset of AD-related dementia, and aid in patient stratification schemes in clinical trials.

THE ROLE OF GENETICS IN AD PATHOGENESIS

After the first clinicopathological description of AD by German psychiatrist Alois Alzheimer in 1907 (2) it took

more than half a century to realize that the disease shows familial aggregation consistent with genetic transmission (11). It took another decade until the use of genetic linkage analysis followed by positional cloning led to the discovery of rare mutations in three genes encoding the amyloid-beta precursor protein (*APP*) and presenilins 1 and 2 (*PSEN1/PSEN2*) that cause fully penetrant monogenic forms of AD (22). While a large number of additional mutations in all three genes have been described following the original reports (for an up-to-date summary of monogenic findings in AD see the “AD & FTD Mutation Database” (9)), no additional genetic locus harboring clearly disease-causing mutations has been established since. However, monogenic forms of AD only make up a small fraction of all cases (<5%) while the vast majority of patients suffer from “polygenic AD” (a.k.a. “non-Mendelian AD” or “sporadic AD”). Susceptibility for this latter disease form is determined by the action (and interaction) of numerous independent genomic variants, likely in concert with non-genetic factors, such as environmental exposures (eg, head trauma) and lifestyle choices (eg, alcohol consumption and cigarette smoking). A growing number of studies on a vast array of human disease and non-disease traits has revealed that most—if not all—human phenotypes are driven by complex polygenic backgrounds (8). Thus, with its side-by-side instances of both monogenic and polygenic forms, AD represents a “genetically complex disorder” *par excellence*.

GENOME-WIDE SCREENING IN POLYGENIC TRAITS

DNA sequence variants underlying monogenic diseases, including those causing AD, are exceedingly rare in the population as a whole. This rarity is typically due to the large and highly penetrant molecular effects exerted by such variants resulting in being “selected against” from subsequent generations. DNA sequence variants exerting only moderate molecular effects lead to reduced overall penetrance and are much less subject to selective pressures allowing them to increase in frequency within populations over time. Once the minor allele, that is, the nucleotide with the lowest frequency, is present in 1% or more in the “general” (ie, non-disease) population, a variant is defined as a DNA sequence “polymorphism.” Although the molecular effects and, hence, contribution to disease risk of any such polymorphism may be very small, it is via the *combined* action of many such polymorphisms that a genetically complex disease may eventually develop. Since many, that is, tens to hundreds to thousands, such polymorphisms can contribute to the genetic risk architecture of the same disease, these are said to be of a polygenic background. The identification of common genomic variants relevant for any given disorder is the focus of current gene findings efforts, including those ongoing in AD. The method of choice is genome-wide screening, for example, in the context of genome-wide association studies (GWAS), where the presence of specific alleles or genotypes at polymorphic sites is treated as exposure to predict a certain clinical outcome (here: onset of AD). By design, genome-wide analyses afford a heavy multiple testing burden owing to the very large number of polymorphisms tested (typically several millions) which needs to be accounted for. In European populations it could be shown that approximately 1 million independent DNA sequence variants exist with a minor allele frequency (MAF) >5%. Bonferroni correction for this number of independent tests means that study-wide significance is achieved at $\alpha = 0.05/1\,000\,000 = 5 \times 10^{-8}$, a threshold now widely accepted and used in the field. For more details on this topic as well as a detailed account of the past (and possible future) achievements of GWAS in human genomics research we recommend consulting the review by Visscher *et al* (24).

In AD, about 60 GWAS have been published since 2007 (according to the “NHGRI-EBI Catalog of published genome-wide association studies” [GWAS catalog], URL: <https://www.ebi.ac.uk/gwas/> (6)), although many of these are not independent as the same datasets were utilized in successive meta-analyses. The two most recent and—by sample size—largest AD GWAS were published back-to-back in the March 2019 issue of *Nature Genetics* (14, 16), and represent the core “data” of this review. First, we begin by highlighting the main findings from each of these GWAS with particular emphasis on results overlapping across both studies. This will include a short excursion into rare-variant-based analyses and how data from those studies fit to the results from Jansen *et al* (14) and Kunkle *et al* (16). In

the second part, we will discuss new putative “mechanistic” insights gained from these GWAS findings and their possible implications. We close by providing an outlook on the field for the next 10 years.

THE STATUS OF AD GENOMICS RESEARCH

Prior to spring of 2019, it had been over 5 years since the last *bona-fide* GWAS was published for AD (17). That study, conducted by a group of researchers aligned under the auspices of the “International Genomics of Alzheimer's Disease Project” (IGAP), entailed the analysis of ~75 000 individuals allowing the identification of 20 genome-wide significant (ie, P -value $< 5 \times 10^{-8}$) AD risk loci, of which 11 were novel at the time. Since this IGAP-2013 publication, the consortium continued collecting additional independent AD cases and controls culminating in an updated GWAS (16) on approximately 94 000 individuals of European descent (35K AD cases vs. 59K healthy controls; Table 1). The actual genome-wide screening (“discovery phase”) in this IGAP-2019 GWAS was performed in 64 000 individuals followed by validation analyses in another 30 000 individuals and led to the discovery of 25 genome-wide significant loci, five of which were reported as “novel” (16). The second GWAS discussed in this review (14) was published (online) nearly 2 months before the IGAP follow-up study and, overall, utilized nearly seven times as many samples as the IGAP-2019 GWAS (total $n \sim 635\,000$ individuals; Table 1). Partially owing to its much larger sample size, this AD GWAS identified 29 genome-wide significant loci, of which nine were declared “novel” at the time of publication. Notwithstanding the identification of novel genetic risk loci in these studies it is worth noting that the overall number of genome-wide significant findings in AD GWAS continues to be small compared to other GWAS of similar size, for example, Parkinson's disease or schizophrenia (see GWAS catalog for details (6)), possibly indicating that the degree of polygenicity underlying AD may be lower than that of other traits.

Table 1. Summary of key aspects of the two GWAS discussed in this review.

	Jansen <i>et al</i> (2019)	Kunkle <i>et al</i> (2019)
Sample size		
Discovery	455K (72K vs. 383K)	64K (22K vs. 42K)
Follow-up	180K (6.6K vs. 174K)	30K (13K vs. 17K)
Total	636K (79K vs. 557K)	94K (35K vs. 59K)
Results		
# variants analyzed	13 367 300	11 480 633
MAF threshold	No constraint	No constraint
# gw sign. loci	29	15
# gw sign. genes	169	95

Data in “Sample size” are taken directly from the respective GWAS (14, 16), while data in “Results” were derived from the summary statistics re-analyzed for this review.

Of these two most recent AD GWAS, the study published by Jansen *et al* (14) is considered the more “remarkable” by us based on several grounds. First, not only was it submitted and published *before* the IGAP-2019 (16) study, but it was also the first *bona fide* AD GWAS to make all its results (main and supplementary) publicly available as preprint (on bioRxiv at <https://doi.org/10.1101/258533>) prior to peer-review and publication in *Nature Genetics*. Some 6 weeks later, the IGAP-2019 group published their results as preprint as well (<https://doi.org/10.1101/294629>). Second, Jansen *et al* (14) drastically increased the overall sample size (and along with it: statistical power) by nearly an order of magnitude compared to all other previous GWAS in the AD field. This was made possible by utilizing genome-wide data from nearly 48 000 AD (proxy; see below) cases and 330 000 non-AD controls of the UK biobank (UKB) project. UKB is a unique prospective cohort study with deep genetic and phenotypic data collected on ~500 000 individuals from across the United Kingdom (7). At baseline, UKB participants were aged between 49 and 69 years, and therefore mostly too young for having developed polygenic AD, which peaks after the age of 65. To circumvent this problem, and this is the third reason for being a truly remarkable study, Jansen *et al* utilized a method based on “proxy phenotyping” which makes use of parental AD status as recorded in UKB medical records. This approach was recently proposed (18) to be a valid approximation of future AD status in UKB individuals for whom genotype data were available but who had not (yet) developed AD themselves.

MAIN RESULTS FROM THE GWAS BY JANSEN *ET AL* (14)

Some parts of the ensuing results summary were already highlighted in a “News and Views” article which we published earlier this year (5). In their discovery phase, Jansen *et al* (14) combined the data from UKB, IGAP-2013 and two smaller case-control datasets from Europe and the US to arrive at an overall sample size of ~455 000 individuals. As expected owing to the partial sample overlap, the loci reported to show genome-wide significance in (14) included many also highlighted in the IGAP-2013 GWAS (17), but also pinpointed evidence for genome-wide significant (P -value $< 5 \times 10^{-8}$) association at nine additional and novel loci (Table 2), all of which were subsequently tested in an independent replication sample of 180 000 individuals from Iceland (Table 1). At the same time, the data by Jansen *et al* (14) did not confirm several of those reported in the IGAP-2013 study (ie, *MEF2C*, *NME8*, *CELF1*, and *FERMT2*) and renewed association evidence at one locus (ie, *CD33*; originally identified in a GWAS by our group more than 10 years ago (3)) showing clear genome-wide significant (P -value $< 5 \times 10^{-8}$) association with AD risk (Table 2). Also new on the list is *ADAM10*, encoding the key enzyme cleaving APP to preclude A β (the core constituent of β -amyloid and, hence, senile plaques) generation,

which has previously been shown to contain rare variants segregating with AD status in AD families (21), and *APH1B*, whose encoded protein, Aph-1 homolog B, together with the presenilins is a component of the gamma secretase complex, responsible for cleaving APP to produce A β . The other novel loci identified in the GWAS by Jansen *et al* (14) include *ADAMTS4* (located on chromosome 1), *CLNK* (chr. 4), *KAT8* (chr. 16), *ALPK2* (chr. 18), *AC074212.3* (chr. 19), *HESX1* (chr. 3), and *CNTNAP2* (chr. 7). All but the last two loci were pinpointed with common polymorphisms (ie, those with an MAF $> 1\%$; Table 2), while the latter two loci showed their lead signals with rare variants (defined here as MAF $< 1\%$; Table 3).

For a visual summary of the genome-wide association results by Jansen *et al* (14) as a “Manhattan plot” see Figure 1; for a detailed summary of top AD loci see Tables 2 and 3. To facilitate comparison, both sets of results are depicted next to the discovery-phase GWAS findings by Kunkle *et al* (16) in Figure 1. Despite there being a total of 29 loci showing genome-wide significance, it needs to be emphasized that one locus stands out both in terms of statistical support and exerted effect size, that is, locus #26 in (14), located in the *APOE* region on the long arm of chromosome 19. The “standing out” nature of this locus can be grasped from Figure S1 where we plot non-truncated association P -values in the *APOE* region for both GWAS: with P -values at 1×10^{-300} and below, markers in the *APOE* region are hundreds of orders of magnitude more significantly associated with AD risk than any other locus in the genome (the next best association is observed with a variant in the *BINI* locus showing a P -value = 3.38×10^{-44} ; Table 2). Interestingly, of all currently established AD genetic risk factors, *APOE* represents the only locus to emerge from the pre-GWAS “candidate gene era” (4); all other AD loci in Table 2 where established to represent *bona fide* AD genes by genome-wide screening.

MAIN RESULTS FROM THE GWAS BY KUNKLE *ET AL* (16) (IGAP-2019)

Owing to its smaller sample size, the IGAP-2019 GWAS identified fewer loci at genome-wide significance than the study by Jansen *et al* (14). Overall, there were 25 loci highlighted by Kunkle *et al* (16) as AD risk factors, five of which were deemed “novel” by the authors, namely *ADAM10* (on chromosome 15), *IQCK* and *WWOX* (both chr. 16), *ACE* (chr. 17), and *ADAMTS1* (chr. 21; Table 2). Technically, *ADAM10* was first published as a genome-wide significant AD locus 2 months prior in the GWAS by Jansen *et al* (14) to which it is, hence, credited for the purpose of this review. The other four novel IGAP-2019 AD GWAS loci either show no or only very modest evidence of association with AD risk in the larger datasets by Jansen *et al* (14), which is why these are interpreted only as “potential” or “possibly false positive” findings by us (Table 2). Conversely, Kunkle *et al* (16) also did not replicate some of the novel signals in the Jansen GWAS (namely: *ADAMTS4* and *ALPK2*), although this may simply be attributed to the

Table 2. Summary of current common variant (MAF \geq 0.01) AD genomics findings from the GWAS by Jansen *et al.* (14) and Kunkle *et al.* (16)

Chr	Pos	LeadSNP	P-value Jansen		MAF [†]	P-value Kunkle		AD effect	Nearest gene	AD pathway	Potential link to AD pathogenesis
			A1 vs. A2	et al		A vs. G	et al				
1	161155392	rs4575098	A vs. G	2.05E-10	0.240	2.34E-02*	Risk	ADAMTS4	None	Neuroprotection: Extracellular Matrix Protease	
1	207786828	rs2093760	A vs. G	1.10E-18	0.225	1.66E-15*	Risk	CR1	Immune	Innate Immunity;	
2	127891427	rs4663105	A vs. C	3.38E-44	0.412	2.16E-26*	Risk	BIN1	Lipid	Neuroinflammation	
2	233981912	rs10933431	G vs. C	8.92E-10	0.240	3.42E-09**	Protection	INPP5D	None	Cellular Protein Trafficking	
4	11026028	rs6448453	A vs. G	1.93E-09	0.228	4.90E-05*	Risk	CLNK	None	Autophagy; Viral Infection	
6	32583357	rs9469112	T vs. A	8.41E-11	0.153	2.32E-07**	Protection	HLA-DRB1	Immune	Innate Immunity;	
6	47432637	rs9381563	C vs. T	2.52E-10	0.344	3.57E-10**	Risk	CD2AP	None	Neuroinflammation	
7	99971834	rs4727449rs1859788	A vs. G	2.22E-15	0.323	1.22E-09**	Protection	ZCWPW1	None	Adaptive Immunity	
7	143108158	rs7810606	T vs. C	3.59E-11	0.425	1.13E-06**	Protection	EPHA1	None	Blood Brain Barrier; A β	
8	27464929	rs28834970/rs4236673	A vs. G	2.61E-19	0.390	5.60E-23**	Protection	CLU/PTK2B	Immune; lipid; tau	Transcytosis	
10	11717397	rs11257238	C vs. T	1.26E-08	0.382	2.61E-07**	Risk	ECHDC3	None	Innate Immunity;	
11 [†]	47380340	rs3740688	G vs. T	4.50E-05	0.458	5.46E-13**	Protection	SPI1/CELF1	Immune	Neuroinflammation	
11	59958380	rs2081545	A vs. C	1.55E-15	0.342	5.35E-17**	Protection	MS4A6A	Immune	Signal Transduction	
11	85776544	rs867611	G vs. A	2.19E-18	0.342	3.41E-19**	Protection	PICALM	APP	A β clearance/Signal Transduction	
11	121435587	rs11218343	C vs. T	1.09E-11	0.035	2.88E-12**	Protection	SORL1	Lipid; APP	Lipid Metabolism	
14 [†]	53391680	rs17125924	G vs. A	5.26E-06	0.099	1.42E-09**	Protection	FERMT2	n.a.	Potential false-positive result (not replicated in UKB)	
14	92938855	rs12590654	A vs. G	1.65E-10	0.347	8.79E-09*	Protection	SLC24A4	None	Innate Immunity;	
15	59022615	rs442495	C vs. T	1.31E-09	0.334	2.51E-7**	Protection	ADAM10	Immune	Neuroinflammation	
15	63569902	rs117618017	T vs. C	3.35E-08	0.132	2.38E-04*	Risk	APH1B	None	Blood Brain Barrier; A β	
16*	19808163	rs7185636	C vs. T	1.40E-01	0.156	2.4E-08***	Protection	IQCK	n.a.	Transcytosis	
16	31133100	rs59735493	A vs. G	3.98E-08	0.324	7.42E-03*	Protection	KAT8	None	Cellular Protein Trafficking	
16*	79355857	rs62039712	G vs. A	7.66E-01	0.094	3.70E08*	Risk	WVVOX	n.a.	Potential false-positive result (not replicated in UKB)	
17	5138980	rs113260531	A vs. G	9.16E-10	0.118	3.70E-04**	Risk	SCIMP	None	Calcium Homeostasis	
17	47450775	rs28394864	A vs. G	1.87E-08	0.471	4.85E-03*	Risk	ABI3	None	Sheddase; APP Processing	
17	61538148	rs138190086	A vs. G	2.65E-04	0.017	5.30E-09***	Risk	ACE	Immune	γ -secretase; APP Processing	
18	56189459	rs76726049	C vs. T	3.30E-08	0.011	1.76E-01*	Risk	ALPK2	None	Likely false-positive result (not replicated in Jansen)	
19	1039323	rs111278892	G vs. C	7.93E-11	0.165	1.10E-07*	Risk	ABCA7	Lipid; APP	Innate Immunity;	
19	45411941	rs429358	C vs. T	<1E-900	0.155	1.17E-881*	Risk	APOE	Lipid; APP; tau	Neuroinflammation	

Table 2. (Continued)

Chr	Pos	LeadSNP	A1 vs. A2	MAF [†]	P-value Jansen <i>et al.</i>	P-value Kunkle <i>et al.</i>	AD effect	Nearest gene	AD pathway	Potential link to AD pathogenesis
19	46241841	rs76320948	T vs. C	0.059	4.64E-08	1.22E-04*	Risk	AC074212.3	None	?
19	51727962	rs3865444	A vs. C	0.336	6.34E-09	5.27E-06**	Protection	CD33	None	Innate Immunity; Neuroinflammation
20	54998544	rs6014724	G vs. A	0.089	6.56E-10	3.65E-07*	Protection	CASS4	None	Signal Transduction
21*	28156856	rs2830500	A vs. C	0.336	1.65E-02	2.60E-08***	Protection	ADAMTS1	n.a.	Likely false-positive result (not replicated in Jansen)

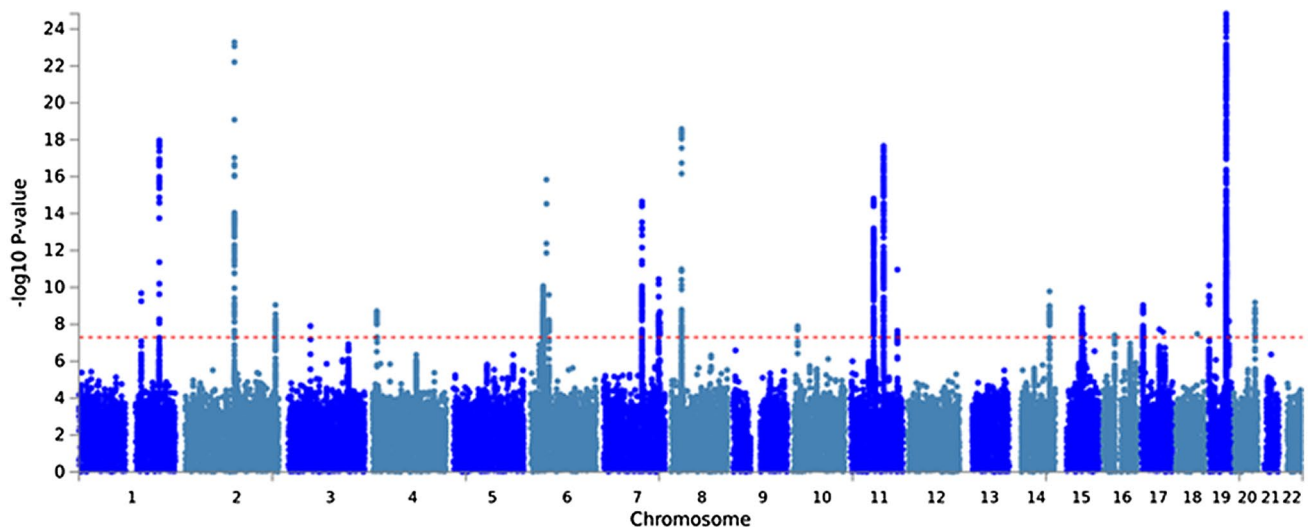
"Chr," "Pos," "A1 vs. A2," "AD effect" taken from Jansen *et al.* "Nearest gene" determined from UCSC genome browser (GRCh37, hg19). Blue highlight indicates "novel" finding in GWAS by Jansen *et al.*, grey highlight indicates "novel" findings in GWAS by Kunkle *et al.*, no highlight indicates previously described GWAS findings. AD pathway as determined in either or both GWAS: "immune" = "immune system response"; "lipid" = "lipid metabolism"; "APP" = "APP metabolism"; "tau" = "tau protein binding" (see text for more details).
 "Potential link to AD pathogenesis" = based on the authors' review (and interpretation) of the literature.
 *Stage 1 result from Kunkle *et al.*
 **Stage 2 result from Kunkle *et al.*
 ***Stage 3 result from Kunkle *et al.*
[†]SPI1/CELF and FERMT2 represent results featured in Kunkle *et al.* that poorly replicate in the UKB portion of Jansen *et al.*, these are interpreted as possibly false-positives here.
[‡]MAF = minor allele frequency; from European controls (non-Finnish) as provided on GnomAD [v.2.1.1.; <https://gnomad.broadinstitute.org/>].
[§]IQCK, VWOX, and ADAMTS1 represent results featured in Kunkle *et al.* that are not replicated at P-value < 0.01 in Jansen *et al.*; these are interpreted as possibly false-positives here.

Table 3. Summary of current rare variant (MAF ≤ 0.01) AD genomics findings from the GWAS by Jansen *et al.* (14) and Kunkle *et al.* (16)

Chr	Pos	LeadSNP	A1 vs. A2	MAF [†]	P-value Jansen	P-value Kunkle	AD effect	Nearest gene	AD pathway	Potential link to AD pathogenesis
3	57226150	rs184384746	T vs. C	0.002	1.24E-08	n.a.	Risk	HESX1	None	Homoebox Gene; Development
6	41129252	rs75932628	T vs. C	0.002	2.95E-15	2.95E-12*	Risk	TREM2	Immune system response	Innate Immunity; neuroinflammation
7	145950029	rs114360492	T vs. C	0.0003	2.10E-09	n.a.	RISK	CNTNAP2	None	Neuronal Development
16 [†]	81942028	rs72824905	G vs. C	0.01	2.11E-03	792E-03*	Protection	PLCG2	None	Microglial activation; neuroinflammation
17 [†]	47297297	rs616338	T vs. C	0.01	7.81E-07	n.a.	Risk	ABI3	None	Microglial activation; neuroinflammation

"Chr," "Pos," "A1 vs. A2," "AD effect" taken from Jansen *et al.* "Nearest gene" determined from UCSC genome browser (GRCh37, hg19). Blue highlight indicates "novel" AD locus in GWAS by Jansen *et al.*, grey highlight indicates "novel" AD locus in GWAS by Kunkle *et al.*, no highlight indicates previously described GWAS findings.
 AD pathway as determined in either or both GWAS: "immune" = "immune system response"; "lipid" = "lipid metabolism"; "APP" = "APP metabolism"; "tau" = "tau protein binding" (see text for more details).
 "Potential link to AD pathogenesis" = based on the authors' review (and interpretation) of the literature.
 *Stage 1 result from Kunkle *et al.*
 **Stage 2 result from Kunkle *et al.*
 ***Stage 3 result from Kunkle *et al.*
[†]Variants in PLCG2, and ABI3 were highlighted in a rare variant GWAS by the IGAP group (Sims *et al.*, 2018) (20) published prior to the Kunkle *et al.*, 2019 (16) study and are listed here for reasons of completeness.
[‡]MAF = minor allele frequency; from European controls (non-Finnish) as provided on GnomAD [v.2.1.1.; <https://gnomad.broadinstitute.org/>].

A Jansen et al. (2019)



B Kunkle et al. (2019)

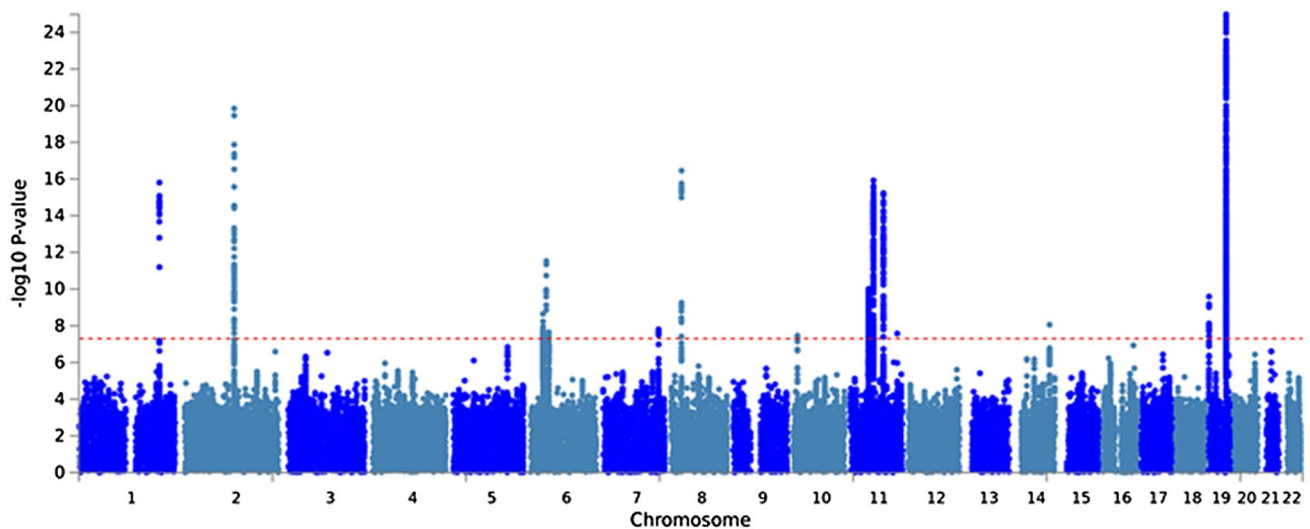


Figure 1. Manhattan plots of “discovery phase” GWAS findings from the two GWAS discussed in this review. A. Jansen *et al* (14); B. Kunkle *et al* (16). Results are based on data distributed by the two respective studies (see “Data Availability Statement” section for more details). *P*-values are truncated at $1E-25$ for didactic reasons. For Manhattan plots of non-truncated results see Figure S1.

much smaller sample size (and, hence, reduction in power) of the IGAP-2019 study. While these could therefore also represent false-positive findings and more data are needed to more conclusively address this possibility, we consider the likelihood of such an outcome as relatively low, and therefore count these latter two signals as *genuine* AD genetic loci for the purpose of this review.

For a visual summary of the genome-wide association results by Kunkle *et al* (16) as a “Manhattan plot” see Figure 1 (and Figure S1); for a detailed summary of top AD loci see Tables 2 and 3. To facilitate comparison, the results of the IGAP-2019 GWAS are depicted next to the discovery-phase GWAS findings by Jansen *et al* (14) in

Figure 1. As for the results by Jansen *et al* (14), markers in the *APOE* region outshine all other genome-wide significant AD loci by a large margin, again emphasizing the predominant role of this locus in AD pathogenesis (Figure S1B).

OVERLAPPING FINDINGS ACROSS GWAS BY JANSEN ET AL AND KUNKLE ET AL

Before engaging in considerations on the potential “mechanistic” implications of the current AD genomics findings,

one needs to derive a subset of results that stands a large likelihood of surviving the test of time, that is, likely representing *genuine* AD loci. As outlined above, Jansen *et al* (14) reported to have identified nine novel loci at genome-wide significance. Two of these, that is, *HESX1* and *CNTNAP2*, were elicited by rare-variants (MAF < 1%; Table 3) for which IGAP-2019 did not compute or make available summary association statistics at the time of writing. Based on the strength of the reported association evidence, we still consider them as *genuine* AD risk loci here. Furthermore, five of the remaining seven novel AD loci elicited by common variants (MAF > 1%) highlighted by Jansen *et al* (14) showed the same direction of effect and association *P*-values < 0.01 in IGAP-2019, namely *CLNK*, *ADAM10*, *APH1B*, *KAT8*, and *AC074212.3* (Table 2). Therefore, for the purpose of this review they are also considered as *genuine* AD loci. Interestingly, the reverse is not true for most loci: of all 4 novel IGAP-2019 loci, only *ACE* on chromosome 17 replicates in Jansen *et al* (14) and, thus, appears to represent a *genuine* AD finding. Finally, there remain two loci with somewhat unclear association evidence, that is, *SPII/CELF1* (on chromosome 11) and *FERMT2* (chr. 14). Both of these were originally identified in the IGAP-2013 study (17) and continue to show genome-wide significant association with AD risk in IGAP-2019 (Table 2). This is not surprising given that 80% of individuals in IGAP-2019 were also included in IGAP-2013. However, Jansen *et al* (14) did not consider these loci further due to the “lower association signals in the UKB data set” (ie, *P*-values of 0.02 and 0.004 for *SPII/CELF1* and *FERMT2*, respectively). Given the extremely modest association evidence for these two loci in UKB, we tend to agree with the notion of Jansen *et al* (14), and will also not consider these two loci further in the remainder of this review.

In summary, taking all the available evidence together, most genome-wide association findings show good correspondence across both studies. Overall, there emerge 32 apparently *genuine* AD risk loci at the day of writing of this review (August 2019), 27 (out of 32) from common variant (Table 2) and 5 (out of 5) from rare-variant based-results (Table 3). Note that the latter results also include two loci, that is, *PLCG2* and *ABI3*, which recently emerged from an “exome chip” GWAS (20) on what appears to be largely the same dataset also utilized for the IGAP-2019 GWAS. Variants in both loci showed *P*-values < 0.01 in the Jansen *et al* (14) results (Table 3), thus, fulfilling our criteria of “replication.”

INSIGHTS INTO GENOMIC MECHANISMS OF AD FROM GWAS

The *identification* of complex trait genetic loci represents only the first step in what inevitably proves to be a “long and winding road” to advancing our understanding of the pathogenic mechanisms underlying the trait in question. This process amounts to “making sense” of GWAS findings and putting them into the (correct) context with other molecular data to understand why and when a specific

disease has developed in cases, but not in controls. A next and probably more important aspect is to utilize these genetic insights to predict an individual's risk for the disease in question *prior* to the actual onset of first symptoms, with the aim to offer specific genetic counseling and/or therapeutic options (if these exist) similar to what is now possible for many monogenic disease mutations. In the beginning of this review, we summarized this procedure as the development of “early prediction—early detection—early intervention” procedures in AD. Notwithstanding its clinical importance, the translation of polygenic genomics findings into medical genetics practice is not discussed further in this review. Instead, we focus on the interpretation of the GWAS findings from a “disease mechanism” viewpoint.

In the genomics community, the set of analyses focusing on the interpretation (in terms of putative disease mechanisms and/or translational potential) of specific GWAS findings is often described as “post-GWAS analyses.” In general terms, these approaches entail the utilization of different computational tools and analyses with the aim to integrate high-resolution data from other genomics domains (eg, transcriptomics or epigenomics), as well as other “-omics” data in general (eg, proteomics and metabolomics). The main aims of these efforts can perhaps be summarized (and simplified) as follows: First, delineate the functionally relevant genes located within the identified AD GWAS loci. This is important because many AD loci actually contain several plausible gene candidates in the immediate vicinity of associated lead variants. Second, characterize the functionally relevant molecular genetic mechanisms *within* delineated gene candidates, for example, assess whether the predominant effect on pathogenesis is the result of an exonic variant changing the function of the encoded protein or that of a regulatory variant changing the gene's expression. Third, if a non-exonic variant is the likely culprit, assess whether it affects the expression profile of implicated genes in the tissue of interest, for example, in the case of AD the brain, for example, hippocampus or specific cortical areas. Fourth, delineate the overarching mechanistic “themes,” for example, specific pathways or gene/protein networks, with the aim to assess how the genomics findings may fit with evidence from other molecular domains, for example, neuropathological data. Almost always, these and other analyses are applied in parallel and in combination to arrive at the most likely solution. The hope behind these efforts is to arrive at a tractable mechanistic hypothesis which can then be tested by dedicated laboratory experiments to prove or disprove causality of a specific disease-associated sequence variant.

Owing to the complexity of the matter, we can here only provide some first and early insights into potential mechanisms offered by the newest AD genomics findings. For the sake of simplicity, we only touch on the four main “mechanistic domains” outlined in the previous paragraph and summarize selected results of post-GWAS analyses presented in the two primary GWAS publications that form the basis of this review. We refer to the other papers in this Mini-Symposium for insights into additional potential mechanisms underlying AD pathogenesis.

DELINEATING FUNCTIONALLY RELEVANT GENES WITHIN THE AD ASSOCIATED GENETIC LOCI

In the context of GWAS, the term “genetic locus” describes a specific stretch of genomic DNA collectively showing association evidence with the outcome trait in question, here the onset of AD. The physical dimensions, that is, length, of each locus can vary quite substantially between chromosomal regions and mainly depend on the extent of correlation between DNA sequence variants located within the locus, a situation referred to as “linkage disequilibrium”. A genomic region not much affected by chromosomal recombination at meiosis will be larger (and possibly contain more genes) than recombination “hotspots” which are smaller and typically contain fewer genes. In the GWAS by Jansen *et al*, the average AD locus encompassed ~138 000 base pairs (bp) in length (range 1–823 000 bp; calculated based on information from table S2 of (14)). Using three different mapping strategies the authors of that study concluded that up to 192 genes may be linked to the 29 AD loci identified by genome-wide screening. Using equivalent mapping strategies, the authors of the IGAP-2019 study delineated up to 400 gene candidates underlying the GWAS signals in their analyses (16). In theory, all of these could be involved in AD pathogenesis and hence represent culprits for eliciting the observed GWAS signal within each locus. In some instances, it is even possible that several DNA sequence variants within the same locus are independently associated with disease risk and these do not necessarily have to be located in the same gene. The *APOE* locus on chromosome 19q13.32 shall serve as a illustrative example of the problem faced when attempting to translate “GWAS loci” to “disease genes” (Figure S2): here, the entire AD-associated GWAS locus extends over ~823 000 bp. Near the main association signal, it contains several highly correlated variants mapping into different genes located in close proximity. Most of the other non-*APOE* AD GWAS loci highlighted in the GWAS by Jansen *et al* (14) and Kunkle *et al* (16) contain several, albeit less than in the *APOE* region, such gene candidates. Knowledge of the other “mechanistic domains” (see below) can help to further pinpoint the pathophysiological culprits, that is, genes eliciting the observed GWAS signals.

CHARACTERIZATION OF THE UNDERLYING MOLECULAR GENETIC MECHANISMS

An integral part of mapping relevant disease genes within GWAS loci is characterizing the assumed molecular genetic mechanisms of the associated DNA sequence variants. For instance, an exonic loss-of-function variant, for example, introducing a premature stop-codon severely affecting the function of the encoded protein, may elicit stronger and more significant disease-effects than an intronic variant without any obvious molecular consequences. However, exonic variants (and others near the coding sequence) are quite rare and rarely serve as plausible mechanistic explanation

for the disease association. Figure 2 depicts the distribution of variant locations for all genome-wide significant loci in the Jansen *et al* (14) GWAS (Figure S3 for both GWAS, side-by-side). It shows that the two most frequent disease-associated variant categories are “intergenic” (ie, outside the coding sequence, between genes) and “intronic.” While these categories may at first appear less compelling than “exonic” or “splicing” or “UTR” it is becoming increasingly clear from other work that much of the intergenic space is actively involved in the regulation of gene expression (15) and may, therefore, be of relevance in the onset and progression of disease. Systematic functional mapping of variants in the 29 loci highlighted in the Jansen *et al* (14) GWAS revealed that in addition to *APOE* a total of only four loci (ie, *CR1* [chr. 1], *PILRA* [chr. 7], *APH1B* [chr. 15], and *CD33* [chr. 19]) contained non-synonymous exonic variants deemed to be “credible causal” for the observed association signals (see table S9 in (14)); conversely, the association signals in the other loci are, therefore, likely elicited by non-exonic variants. Interestingly, within the *APOE* locus on chromosome 19 this approach identified a total of 16 non-synonymous exonic variants across nine different genes: the one deemed as “credible causal” using the same framework was the well-known epsilon4-allele (at variant rs429358) in the *APOE* gene itself.

CHARACTERIZATION OF POTENTIAL VARIANT EFFECTS ON GENE EXPRESSION

Non-exonic (and even some exonic) variants are hypothesized to exert their molecular effects by influencing the regulation of gene expression (10). To this end, the recent advent of high-dimensional reference datasets for tissue-specific gene expression profiles based on whole transcriptome RNA sequencing data, has vastly facilitated the interpretation of putative regulatory effects of DNA sequence variants (26).

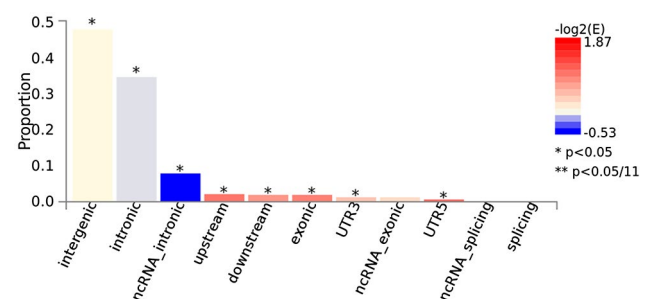


Figure 2. Functional consequences of SNPs on genes for the GWAS by Jansen *et al* (14). The histogram displays the proportion of SNPs (all SNPs in LD with independent significant SNPs) which have corresponding functional annotation assigned by ANNOVAR. Bars are colored by \log_2 (enrichment) relative to all SNPs in the selected reference panel. Plots are made with FUMA (25) using data described in “Data Availability Statement” section. For equivalent results of the GWAS by Kunkle *et al* (16) see Figure S3.

Such variants are often called “expression quantitative trait loci” (eQTL), meaning that they elicit quantitative (and statistically significant) effects on the expression of genes. In essence, these studies are nothing but GWAS using quantitative gene expression data as outcome variables. The field was revolutionized by the launch of the “GTEx” (Genotype-Tissue Expression) project, whose overarching aim is to characterize variation in gene expression across individuals and diverse tissues in humans (1). Owing to the availability of gene expression data in 13 different brain regions, data accumulated by the GTEx consortium are highly relevant to the neuroscience community (26). Importantly for the efforts described here, all GTEx data and results are made freely available via a dedicated website (www.gtexportal.org) and numerous variant-mapping algorithms have integrated these data into their “variant effect prediction.” Among these is the “Functional Mapping and Annotation” (FUMA) of GWAS findings tool (25), developed by the same group that also spearheaded the Jansen *et al* (14) AD GWAS.

There are multiple different ways of connecting DNA sequence variant to gene expression data. One way routinely applied by FUMA are “gene property analyses for tissue specificity.” Essentially, these analyses probe for tissue specificity of the phenotype by testing for potential relationships between tissue specific gene expression profiles (based on GTEx data) and disease-gene associations (based on genome-wide gene-based association statistics of the underlying single-variant results). Applying these analyses to the discovery phase gene-based GWAS results from the Jansen *et al* (14) study revealed study-wide significant

association between AD risk genes and tissue-specific gene expression in spleen, whole blood, and lung (Figures 3 and S4A). Interestingly, no significant association was observed between AD risk genes and gene expression in the various different brain regions analyzed by GTEx. A very similar pattern was observed when analyzing the discovery-phase AD GWAS results from the Kunkle *et al* study (16) which also showed significant tissue-specific expression in non-brain regions such as whole blood, liver, and spleen (Figure S4B). These and other related sets of gene expression results converged on the notion that AD risk genes overlap more with immune-system related tissues than with brain- or neuron-specific datasets. For instance, the brain cell types with the highest expression of AD risk genes were microglia, a cell type involved in the brain’s immune system response, rather than neuronal cell types (see figure 4 of (14)).

DELINEATING OVERARCHING MECHANISTIC “THEMES” BY MEANS OF PATHWAY ANALYSES

The final set of “functional characterization” analyses discussed in this section relate to various types of “pathway analyses” to elucidate common mechanistic themes combining genetics with protein function. There are a multitude of different approaches to link disease genes to pathways, but probably the most widely used test in this context is to probe for an enrichment of associated genes in certain predefined gene sets (eg, converging on

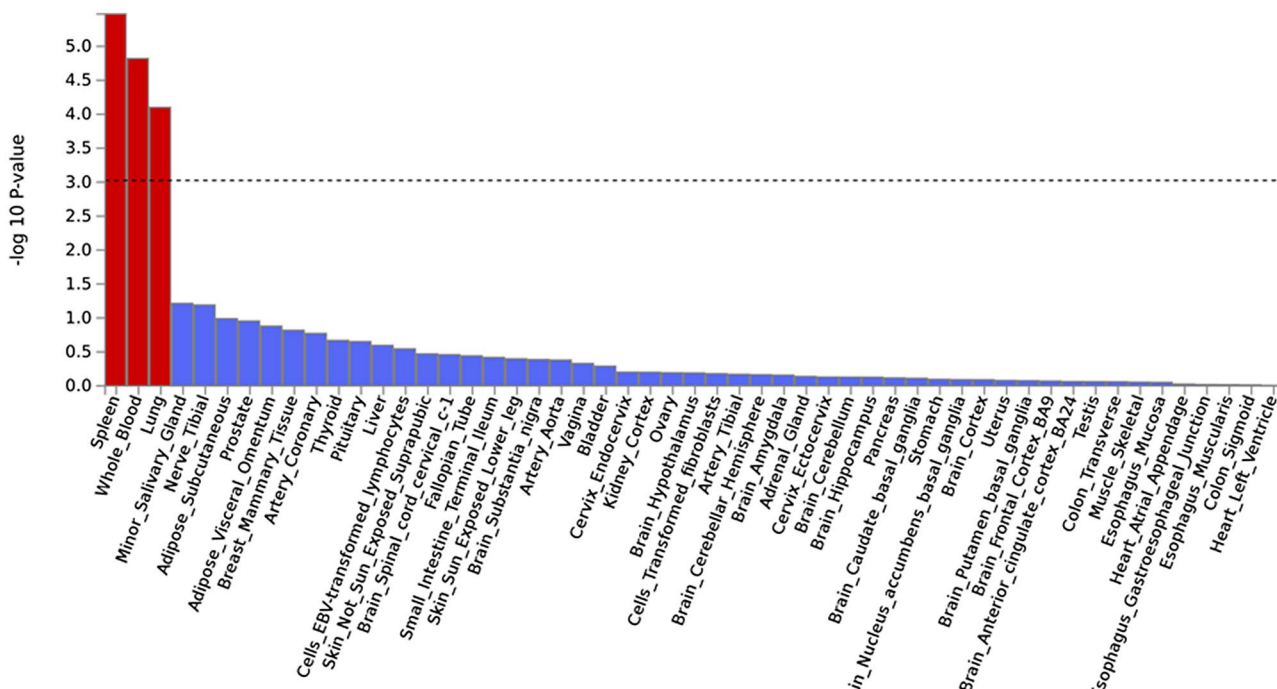


Figure 3. Results of tissue expression analysis for the GWAS by Jansen *et al* (14). Results are based on MAGMA gene-property analyses as implemented in FUMA (25) using GTEx (1) v6 data on 53 tissue types. Input GWAS data as described in “Data Availability Statement” section. Red bars represent study-wide significant results correcting for the 53 tissue types.

certain biological mechanisms defined by gene ontology [GO] classification). Applying such GO-based enrichment analyses for the two GWAS discussed in this review highlighted two biological mechanisms in both studies, that is, processes involved in “lipid metabolism” and “APP metabolism/ A β formation.” In addition, the GWAS by Kunkle *et al* (16) also identified “immune system response” and “tau protein binding” as significantly enriched among their set of AD-associated GWAS genes. While the connection to lipid metabolism and immune system response pathways had already been described in similar enrichment analyses of the IGAP-2013 data (13), the connection to APP metabolism represents a new outcome of the refined AD association results of the two largest and most recent GWAS. As such it provides further support for the notion that the “amyloid hypothesis” of AD—which posits that A β mismetabolism is the primary driver of AD-related pathogenesis (for reviews see refs (22) and (19))—may also be at play in late-onset, polygenic AD, in addition to its well-known role in monogenic AD.

As for all other “functional mapping” domains highlighted in this section, there is a vast array of additional types of analyses allowing to integrate the evidence from genetics studies (such as GWAS) to those from other “-omics” domains with the aim to derive common pathways relevant for pathogenesis and/ or therapeutic interventions. A substantial number of additional results from these analyses are highlighted in the primary publications by Jansen *et al* (14) and Kunkle *et al* (16). To learn more about these findings and their implications we encourage the interested reader to scrutinize the very detailed Supporting Information provided with both publications on the *Nature Genetics* website.

CONCLUDING REMARKS AND OUTLOOK

At the day of writing (August of 2019), AD genomics can be summarized succinctly with the results from two GWAS investigating a large number of independent case control datasets of European descent using state-of-the-art methodology. From these studies, a total of 32 independent genomic loci showing compelling association with AD emerge in analyses of both common (Table 2) and rare (Table 3) genomic variants. While the exact nature of the underlying genes biologically responsible for the observed genetic associations remains elusive for most of the implied loci, a number of overlapping mechanistic themes emerge. First, both GWAS converge on the notion that most AD associated DNA variants are located in non-coding portions of the genome, especially in regions with effects on gene transcription. This is in line with GWAS results from other complex phenotypes (8, 24) and has important bearings on the design of future genomic studies: if most of the functionally relevant variation occurs outside genes, technologies focusing on coding regions only (such as exon variant genotyping or whole-exome sequencing) will likely not be suitable to decipher the genetic basis of AD and other conditions. Instead, more emphasis should be laid

on the regions *between* genes (eg, using whole-genome sequencing) and their functional implications and interactions (eg, using epigenomic and transcriptomic profiling, covered elsewhere in this Mini-Symposium). Second, and in line with previous work in the AD field, the computational modeling performed by both GWAS emphasizes lipid metabolism and immune system response as crucial components in the pathogenesis of AD. For the latter, this is corroborated by gene-set enrichment results showing expression in immune system-related tissues (ie, whole blood, spleen, and liver) and, perhaps more importantly, in a key population of immune cells in the brain (microglia). Third, while no direct association signals were observed in the genes causing early-onset monogenic AD (ie, *APP*, *PSEN1*, and *PSEN2*) the GWAS variants by both teams show a highly significant enrichment in other genes involved in the regulation of “APP metabolism/ A β formation.” This is the first time that APP metabolism emerged as a main functional category in genetic analyses of polygenic AD, providing further support for the amyloid hypothesis in this form of the disease. Last but not least, the results of both GWAS were published in “pre-print” form prior to entering the peer-review process (<https://www.biorxiv.org/>). The authors should be commended for this decision, as it has effectively allowed the community to work with their findings for almost a year before formal publication.

Despite their seminal scopes and unique analytic angles extending our understanding of genomic mechanisms underlying AD pathogenesis, the current GWAS results still leave some important questions unanswered. For instance, the new data in both studies did not markedly increase the proportion of phenotypic variance explained by genetics, a situation often described as the “missing heritability problem” in complex traits (27). Hence, if the phenotypic variance cannot be sufficiently explained by “simple” DNA variants of the type typically assayed by GWAS (eg, single base-changes and small insertion-deletions), the elusive heritability must be “hidden” elsewhere, for instance in other types of genomic variants (necessitating other genotyping/sequencing methods) and/or in genetic interactions among loci (necessitating novel analytic approaches highlighted elsewhere in this Mini-Symposium). Third, both GWAS discussed here (as most GWAS published for AD to date), focus on datasets of European descent. The reason for this apparent selectivity is often simply convenience: European-descent populations are typically those with the most readily available phenotype and genotype data. It will be exciting to observe how the utilization of datasets from other descent-groups will (re-) shape our knowledge and understanding of genomics mechanisms underlying AD (12). Finally, delineating the precise molecular mechanisms linking “genomic dysfunction” to “cognitive dysfunction,” for example, via “immune system dysfunction,” are still, works-in-progress and will require the development and application of novel methods effectively linking readouts from “-omics”-based studies to cellular function *in vivo* to establish causality of the observed statistical associations.

ACKNOWLEDGEMENTS

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DATA AVAILABILITY STATEMENT

All figures shown in this review were based on the re-analysis and plotting of summary statistics made available alongside the original GWAS using FUMA (25). Specifically, for the Jansen *et al* (14) study we utilized the discovery phase association results available at https://ctg.cncr.nl/software/summary_statistics. For the Kunkle *et al* (16) study, we used the Stage 1 summary data available at <https://www.niagads.org/igap-rv-summary-stats-kunkle-p-value-data>.

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SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article at the publisher's web site:

Figure S1. Manhattan plots of “discovery phase” GWAS findings from the two GWAS discussed in this review. **A.** Jansen

et al (14); **B. Kunkle *et al*** (16). Equivalent to Figure 1 except that *P*-values were not truncated.

Figure S2. Regional association plot for the *APOE* region on chromosome 19q13.32 in the GWAS by Jansen *et al* (14). Plots were constructed with FUMA (25) on data described in “Data Availability Statement” section. Color code for SNPs: Each SNP is color-coded based on the highest r^2 to one of the independent significant SNPs, if that is greater or equal to $r^2 = 0.6$. Other SNPs (ie, $r^2 < 0.6$) are colored in grey. The top lead SNPs in genomic risk loci, lead SNPs and ind. sig. SNPs are

circled in black and colored in dark-purple, purple and red, respectively. Color code for genes: Red: Mapped genes. Blue: Non-mapped protein-coding genes. Dark grey: Non-mapped non-coding genes.

Figure S3. Functional consequences of SNPs on genes for the GWAS by Jansen *et al* 14 and Kunkle *et al* (16). See legend to Figure 2 for more details.

Figure S4. Results of tissue expression analysis for the GWAS by Jansen *et al* 14 and Kunkle *et al* (16). See legend to Figure 3 for more details.