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IL-23/IL-17 Axis in Inflammatory Rheumatic Diseases

Hao Li1, **George C. Tsokos**¹

¹Departments of Medicine, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA

Abstract

In inflammatory rheumatic disorders, the immune system attacks and damages the connective tissues and invariably internal organs. During the past decade, remarkable advances having been made towards our understanding on the cellular and molecular mechanisms involved in rheumatic diseases. The discovery of IL-23/IL-17 axis and the delineation of its important role in the inflammation led to the introduction of many needed new therapeutic tools. We will present an overview of the rationale for targeting therapeutically the IL-23/IL-17 axis in rheumatic diseases and the clinical benefit which has been realized so far. Finally, we will discuss the complex interrelationship between IL-23 and IL-17 and the possible uncoupling in certain disease settings.

Keywords

Inflammatory rheumatic disorders; IL-23/IL-17 axis; Biologics

Introduction

Inflammatory rheumatic disorders are a group of diseases with variable phenotypic presentation [1–5]. However, the presence of either systemic inflammation or organ-specific inflammation is a common feature of these diseases [6, 7]. During the past decade, the identification of the proinflammatory function of interleukin-17 (IL-17) [8–10] and the discovery of a novel subset of T helper cells termed Th17 cells [11, 12] which drive inflammation by producing IL-17, the signature cytokine, have led to important insights into chronic inflammation. Interleukin-23 (IL-23), a heterodimeric cytokine comprising two subunits (p19 and p40), controls the production of pro-inflammatory cytokines including IL-17, IL-22, and GM-CSF by promoting the development and expansion of pathogenic Th17 cells [13]. This relationship between IL-23 and Th17s has led to the concept of the IL-23–IL-17 axis as a pivotal pathway driving various autoimmune processes [13–15]

Rheumatic diseases are the most common cause of disability and over 50 million Americans are living with some form of rheumatic diseases [16–18].Until late twentieth century, the main drugs available for the treatment of rheumatic diseases were limited to the use of classical disease-modifying antirheumatic drugs (DMARDs) which were developed without full understanding of involved cellular or molecular mechanisms [19, 20]. The introduction

[✉]Hao Li, hli13@bidmc.harvard.edu.

Genetic and experimental data support the concept that the activation of IL-23/IL-17 axis contributes to the development of a series of inflammatory rheumatic diseases, including psoriasis, psoriatic arthritis (PsA), ankylosing spondylitis (AS), rheumatoid arthritis (RA), and systemic lupus erythematosus (SLE) [22–25]. Since IL-17 production is considered to be under the control of IL-23, it was expected that blockade of either IL-17 or IL-23 should have the same clinical effect [13, 14]. The encouraging clinical results in the treatment of psoriasis and PsA reinforce the blockade of this pathogenic axis [26–30]. However, the negative results in trials of patients with rheumatoid arthritis prompt the question whether synergistic blockade of IL-23/IL-17 axis with other biologics could lead to better therapeutic outcomes [31, 32]. Such a consideration will be discussed herein. Furthermore, the clinical trials in AS indicate that the interaction between IL-23 and IL-17 is more complicated than what we had speculated since blockade of IL-17 but not of IL-23 showed greater therapeutic efficacy in the treatment of patients with AS [33–37]. Therefore, further research is warranted to clarify the common and unique roles for IL-23 and IL-17 in the pathogenesis of inflammatory rheumatic diseases.

who fail to respond; the effort to further understand the involved mechanisms and the

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identification of new targets should not abate.

IL-23 is a heterodimeric cytokine composed of p19 and p40 two subunits and principally produced by dendritic cells (DCs) and macrophages [13, 38–40]. Comparative studies analyzing the susceptibility of mice with deficiency of either IL-12p40, IL-12p35, or IL-23p19 to autoimmunity revealed the crucial roles of IL-23 but not IL-12 in the development of EAE and CIA [41, 42]. Overexpression of IL-23 in mice through transgene or hydrodynamic delivery induces multiple organ inflammation [43–45], a fact which suggests the primary role of IL-23 in driving inflammation in autoimmunity [13, 14, 38].

The interleukin-17 (IL-17) family consists of six members, IL-17A through IL17F [9]. IL-17A, the founding member of this family, exists as either homodimers or heterodimers paired with IL-17F and is mainly produced by T helper 17 (Th17) cells [11, 12, 46], a distinct T cell subset [47]. IL-17 has been implicated in the immunopathology of many autoimmune or chronic inflammatory diseases, by acting on many cell types of nonhematopoietic origin, including fibroblasts, epithelial cells, and synoviocytes [48, 49], which lead to the secretion of a range of pro-inflammatory cytokines (including IL-6, TNFα, and IL-1) and T cell- and neutrophil-attracting chemokines including CCL2, CCL7, CXCL1, and CXCL2 [9, 15, 50, 51]. Additionally, IL-17 works in concert with other pro-inflammatory molecules, particularly TNFα, which in turn amplifies the inflammatory signaling in inflammatory environments [15, 52, 53]. Moreover, IL-17 promotes antibody production during inflammation by acting on IL-17 receptor expressing follicular dendritic cells, T cells and B cells [54–56]. As IL-23 is known to be important in sustaining IL-17 production from Th17 cells [57] and disturbing the Th17/Treg balance [58, 59], the evidence generated in

animal models positioned IL-23/IL-17 axis as a pinnacle therapeutic target for rheumatic diseases characterized by chronic inflammation [13].

Genetic studies in human subjects have linked IL-23 receptor (IL-23R) polymorphisms with susceptibility to autoimmune diseases such as psoriasis, PsA, AS, and multiple sclerosis [22, 24, 25, 60, 61]. Later, more variants in genes encoding critical molecules involved in the IL-23/IL-17 pathway, such as IL-12B, CCR6, STAT3, and TYK2, were identified [24, 62– 64]. The functional impact of some variants has been verified using genetically engineered cells or mice carrying orthologous amino acid substitutions [65–67]. Moreover, the presence of IL-17 and IL-23 in the circulation or locally inflamed tissues has been documented in patients with various inflammatory rheumatoid diseases and elevated levels correlate positively with disease severity, a fact that further supports a pathogenic role for the IL23/ IL-17 axis in patients with inflammatory rheumatoid diseases [13, 37, 49, 59, 68].

Biologics Targeting IL23/IL-17 Axis

Although experimental data from animal models of chronic inflammatory diseases including uveitis, lupus, multiple sclerosis, collagen-induced arthritis, and AS have provided insights, the clinical data generated from trials in humans for this therapeutic concept are still limited. Currently, a series of antibody-based drugs targeting IL-23/IL-17 axis have been developed including those blocking IL-17A (Ixekizumab, Secukinumab, and Netakimab), the IL-17 receptor A subunit (brodalumab and KHK4827), IL-23p19 (Guselkumab, Risankizumab, Tildrakizumab, Mirikizumab, and Brazi-kumab), or IL-23p40 (Ustekinumab) [37, 69, 70] (Fig. 1). Despite the success of these biologics targeting individual cytokines or cytokine receptors (Table 1), bispecific antibodies targeting two cytokines with nonoverlapping proinflammatory roles present an attractive opportunity. COVA 322 and ABT-122, two bispecific antibodies targeting both TNF-α and IL-17A, are currently being tested in clinical studies in patients with rheumatoid arthritis and the results appear encouraging [71, 72] (Fig. 1). Besides the synergism with TNF-α, IL-17A effects could also be potentiated by IL-17F, which displays 50% sequence homology and signals through the same receptor [73]. Related trials with different antibodies synergistically targeting two isoforms of IL-17 (Bimekizumab, ALX-0761 and NI-1401) (Fig. 1) are ongoing [73–75].

Understanding of the IL-23/IL-17 axis-mediated signal cascades advances the search for additional targets for inflammatory rheumatic diseases. Although this axis shares many signaling molecules with other inflammatory pathways like IL-1R and TLR-mediated pathways, there are also distinct molecules which are specific and could represent novel treatment targets [13, 76, 77]. Similarly, deciphering the epigenetics and transcriptional requirements of Th17 cell development also will help identify novel therapeutic candidates [78, 79]. Indeed, small molecules which directly disrupt IL-23 signaling and modulate Th17 linage stability by targeting JAK2 and TYK2-dependent STAT3 activation, displayed impressive efficacy in preclinical disease models [80–83] (Fig. 1). Moreover, functional suppression by targeting downstream signaling molecules (JAK1 and JAK3) of proinflammatory cytokines produced by Th17 (IL-22, IL-21, and GM-CSF) also affords tremendous therapeutical values [81, 82]. Although until now, only limited numbers of JAK inhibitors have been approved for the treatment of rheumatic diseases [84–87] (Table 1), it is

just a matter of time that these new therapeutic reagents will be shown to improve the life quality of patients with various autoimmune diseases, including AS, lupus, psoriasis, and other skin diseases such as atopic dermatitis.

Clinical Trial Data

Psoriasis and Psoriatic Arthritis

Psoriasis is a complex inflammatory skin disease typified by the presence of large, erythematous, scaly plaques [18, 88, 89]. Histology examination frequently reveals the presence of T lymphocyte infiltrates under the cutaneous lesions [90]. Around 30% of patients with psoriasis develop PsA, characterized by inflammation-mediated progressive damage of the peripheral joints, spine, and the entheses [91–95]. Common DMARDs including methotrexate, leflunomide, sulfasalazine, and the more recently included anti-TNF-α biologic agents have significantly slowed down the disease progression and relieved the symptoms [96–99]. However, a considerable proportion of patients do not adequately respond to currently approved therapies; therefore, there is need for the development of novel therapies [96, 100].

Genetic, mechanistic, and clinical data support the concept that activation of IL-23–IL-17 axis contributes to the development of psoriasis and PsA [91, 101–105]. First, genome-wide association studies revealed the association of single-nucleotide polymorphisms (SNPs) mapping to $II12b$, $II23a$, $III7a$, and $III7a$ genes with susceptibility to psoriasis and PsA [60, 106, 107]. A current pathogenic model of psoriasis depicted that the inflammation started with the activation of IL-23-producing dermal dendritic cells [108–110], which in turn activates Th17 cells which release key cytokines including IL-17 and TNF-a [111]. These cytokines act on epidermal keratinocytes and lead to observed tissue lesions [112–116]. Of note, overexpression of IL-17 in mice leads to epidermal hyperplasia and bone destruction which are often observed in humans with psoriasis and PsA [113]. The occurrence of both psoriasis and PsA and the presence of Th17 cells in psoriatic synovium [68, 91, 117] lead to the hypothesis for similar immune responses in synovial tissues although it remains unknown whether there were specific autoantigens shared between the joint and the skin. Furthermore, the assessment of the expression of IL-23, IL-17, and their related receptors in psoriatic skin lesions and inflamed synovium did confirm significantly increased expression of these molecules and their positive correlation with disease severity, which strongly suggests the pivotal role of IL-23/IL-17 axis [91, 103].

The above findings have led to a substantial increase of testing novel IL-17 and IL-23 antagonists in people with psoriasis and PsA. Recently, different highly effective therapies that disrupt interleukin-17 (Secukinumab, Ixekizumab, and Brodalumab) and interleukin-23 (Ustekinumab and Guselkumab) signaling has been approved by FDA for both psoriasis and PsA management [26, 91, 118–124]. Another two agents targeting IL-23 (Risankizumab and Tildrakizumab) have been approved for psoriasis while the clinical trials for PsA are still ongoing [125]. These new biologic therapies have proven to be highly effective and result in significant improvements in approximately 70–90% psoriasis or PsA patients with excellent safety profiles [91, 126]. The unprecedented success of these antagonists validated the essential role of this inflammatory axis in driving chronic inflammation. The reported

common adverse events include headache, upper respiratory tract infection, nasopharyngitis, arthralgias, and infections; however, the safety profiles are still comparable with more classical biologics like etanercept [91, 124, 127]. It should be noted that unlike anti-TNFα reagents which deliver comparable rates of amelioration on both skin and joint pathology, the selective blockade of IL-23/IL-17 axis is more effective in treating psoriatic skin over psoriatic joints. These differential responses bespeak to the complexity of involved processes in the development of psoriatic diseases [91].

The JAK/STAT pathway is well linked to the IL-23/−17 axis [13, 128], for example, IL-23 acts through JAK2-TYK2/STAT3-STAT4 pathway and IL-22 [13, 38, 129], an important cytokine produced by Th17 cells, acts through the JAK1/TYK2/STAT1-STAT3 pathway [129]. The first generation of JAK inhibitors includes Tofacitinib, Baricitinib, Ruxolitinib, and Oclacitinib [128]. Although most of them have demonstrated efficacy in the treatment of plaque psoriasis, the long-term safety data require further evaluation considering their limited selectivity [130, 131]. Until now, the risk for serious adverse events appears comparable with that of approved biologic agents [130, 131]. Following the approval of Tofacitinib, the most extensively studied JAK inhibitor, for the treatment of patients with active psoriatic arthritis [132, 133], the development of the second generation of JAK inhibitors with improved selectivity including Peficitinib, Filgotinib, Upadacitinib, and Lestaurtinib has been undertaken seeking better efficacy and safety [81, 134]. In addition, a selective inhibitor of TYK2 (BMS-98616) also showed positive results in a phase II study of people with psoriasis [135].

Recent emerging data suggest that the simultaneous inhibition of two inflammatory cytokines with nonoverlapping functions may provide better efficacy [136]. Specifically, TNF-α and IL-17A, two mediators in the psoriatic pathogenic process, act synergistically on keratinocytes for the induction of key genes involved in inflammation and pathogenesis [15]. Along this line, a few bispecific agents have been designed and clinical trials are ongoing although the safety profile remains a concern particularly the risk of increased rates of infections [137].

Rheumatoid Arthritis

In 1999, Miossec and co-workers demonstrated the presence of IL-17A in synovial tissues from patients with rheumatoid arthritis which first indicated the potential inflammatory role of IL-17A in rheumatoid arthritis [15, 138, 139]. This concept was rapidly supported by numerous studies using various murine models [140]. Subsequently, mice lacking IL-23p19 but not IL-12p35 were shown to be resistant to collagen-induced arthritis due to the absence of IL-17-producing CD4 T cells [42]. After the establishment of Th17 lineage in 2005 [11, 12], several studies documented the increased presence of IL-23, IL-17, Th17 cells, and other IL-23R-expressing cells in the circulation or inflamed joints of people with RA patients which correlated positively with disease activity or joint damage [13, 141–143]. In addition, data from different groups affirmed that IL-17A enhanced osteoclastogenesis and angiogenesis by acting locally on synoviocytes and osteoblasts which lead to synovitis and joint destruction [48]. Of note, IL-23/IL-17 axis has also been reported essential in the control of antibody glycosylation profiles which determine autoantibody activities [144]

(Fig. 2). Together, all evidence has emphasized that IL-23/IL-17 axis should be considered for the development of targeted therapies to treat people with RA [145, 146].

Several clinical trials which aimed to evaluate the therapeutic values of blocking IL-23 or IL-17 in RA have now been completed [140]. However, only poor or moderate efficacy was observed. For example, clinical trials with Secukinumab in biological-naive RA patients or patients with inadequate response to methotrexate or anti-TNFð showed that while there was clinical efficacy of IL-17A blockade, the effect, as judged by ACR20 response, was relatively modest [140]. Studies have also investigated IL-17RA blockade using Brodalumab, and similarly, the trial did not meet the set efficacy endpoints [147]. Consistently, treatment with Ustekinumab or Guselkumab did not significantly reduce the signs and symptoms in patients with RA as well [31].

These results bring out the question why blockade of IL-23/IL-17 axis did not deliver therapeutic value in RA. There are different possibilities. First, RA is a heterogeneous disease and IL-23/IL-17 axis may not be the dominant player for every patient, which is supported by the observation that not all patients display high IL-17A levels or Th17 cell frequencies [148]. It has been shown that IL-17A expression in the joint correlates with serum CRP levels, and interestingly, one Secukinumab trial reported better responses in patients with elevated CRP levels. It would be intriguing to identify the patient subgroups which may respond better to IL-17 or IL-23 blocking biologics. Of note, IL-23 is not required for early Th17 development although it is strictly necessary for late stage IL-17A production [57]. Beside IL-17A, Th17s produce a variety of other inflammatory factors including IL-17F, IL-21, IL-22, and GM-CSF [14, 149, 150], which provide additional explanation for the poor efficacy of current therapies targeting IL-23 or IL-17 alone in RA [151]. Notably, a recent phase II study with Bimekizumab (dual blockade of IL-17A and IL-17F) in RA patients with an inadequate response to anti-TNF showed great therapeutic efficacy [152]. In addition, the synergistic effects of IL-17 with other inflammatory cytokines need attention, for example, IL-17A potentiates the effects of TNFα [9, 15, 153]. More recently, simultaneous blockade of IL-17A and TNF is under investigation using ABT-122, a bispecific antibody which targets both TNF and IL-17A [154].

In contrast to antibody-based therapies, indirect blocking of cytokine production by targeting JAKs offers additional approaches for the management of patients with RA [155]. There are three JAK inhibitors currently approved for the treatment of people with active RA (Tofacitinib, Baricitinib, and Upadacitinib) after demonstrating great efficacy in extensive clinical trials [85]. All agents efficiently inhibit structural damage progression [156]. In clinical practice, JAK inhibitors are increasingly being used after classical DMARD failure [85, 156, 157].

Axial Spondyloarthritis

Axial spondyloarthritis (AS) is a chronic inflammatory disease that affects primarily the spine and the sacroiliac joints [5, 158]. Genetic and epigenetic fine mapping studies have provided evidence for a close relationship between AS and PsA. For example, an association with the IL23R locus exists in PsA and AS [22]. In addition, the integrated genomics

In addition to the genetic evidence, experimental data have also linked the IL-23/IL-17 axis to AS [159]. First, intracellular misfolded HLA-B27, a molecule strongly associated with AS, stimulates the IL-23–IL-17 pathway in vitro [160]. Second, overexpression of IL-23 in mice induces enthesitis and peripheral arthritis which resembles human AS [44]. In this model, IL-23 responding T cells reside in entheseal sites, produce IL-17, and initiate local tissue damage [161]. Moreover, the effects of these two cytokines on osteoclasts and on bone resorption suggest that they might have a catabolic effect on the bone [45, 113, 159, 162]. Of note, in contrast to IL-17, IL-23 did not have any effect on the proliferation and differentiation of osteoblasts, which may explain the discrepancy between individual IL-17 and IL-23 blockade in AS trials [159].

Antibodies targeting IL-17 including Secukinumab, Ixekizumab, Bimekizumab, and Netakimab did show superiority in improving AS over placebo in clinical trials [35, 36, 163, 164]. Surprisingly, in contrast to the positive results of IL-17 inhibitors, the clinical trial results of two IL-23 blocking antibodies Ustekinumab and Risankizumab were clearly negative [34, 165]. Although it cannot be completely excluded that higher doses of Ustekinumab or Risankizumab may be more efficient in reaching the inflamed tissues and provide therapeutical values, the data from psoriasis and PsA clinical trials with different doses did not support this speculation [37]. The data achieved from other inflammatory rheumatic diseases shows a tight connection between IL-23 and IL-17 [13, 38] and the results of either IL-17 or IL-23 inhibition are somewhat similar [14, 68, 91]. Therefore, it is surprising to see the discrepancy of IL-17 and IL-23 blockade in AS clinical trials. A number of considerations have been verbalized to explain this divergence. First, there is a variety of immune cells other than Th17s which can produce IL-17, and especially IL-17F independent of IL-23 [166, 167]. Second, there are many types of cells including B cells and epithelial cells, which can respond to IL-23 without producing IL-17 [144, 168]. Last but not least, the temporal difference between IL-17-mediated and IL-23-mediated process may lead to the observed discrepancy, for example, IL-23 may contribute more to disease initiation but not to progression in patients with AS [159]. However, further studies are needed to explore the details of the underlining molecular and cellular mechanisms.

Notably, two JAK inhibitors, Tofacitinib (JAK1 and JAK3) and Filgotinib (a selective inhibitor for JAK1), significantly reduced the signs and symptoms in clinical trials in AS although the exact mechanisms whereby they regulate AS needs to be further explored [169, 170].

Systemic Lupus Erythematosus

Systemic lupus erythematosus (SLE) is a clinically heterogeneous autoimmune disease characterized by a loss of self-tolerance with the development of autoantibodies against a multitude of self-antigens [1, 171]. There is sufficient evidence that IL-23 and IL-17 are involved in the pathogenesis of SLE [59, 70, 172–174]. Increased frequency of IL-17 producing CD4 T cells and double negative (DN) TCRðβ⁻CD4⁻CD8⁻ T cells were observed in peripheral blood of SLE patients and these cells infiltrated inflamed tissues including the

skin and the kidney [59, 175, 176]. Accordingly, the circulating IL-23 and IL-17 levels are elevated in SLE patients and correlate positively with disease severity [59, 70, 177–180].

Data from animal models have provided insights on the cellular and molecular mechanisms which underlie SLE development. Il23 deficiency in lupus-prone mice significantly ameliorates disease followed by a dramatic reduction of IL-17-producing T cells including Th17s and DN T cells [181, 182]. Consistently, IL-23 overexpression in vivo exaggerates disease progression by promoting expansion of Th17s and DN T cells [183] (Fig. 2). However, the results from IL-17 signaling deficiency in different murine lupus models are paradoxical. In BXD2 mice, IL-23-dependent Th17 cells promote autoantibody production by producing IL-17 which in turn regulates B cell migration inside germinal center area for more rounds of somatic hypermutation [55, 184] (Fig. 2). This concept was subsequently validated by in vivo adoptive transfer of Th17 cells [54]. However, III 7a deficiency in MRL.lpr mice has minimal effects on the course of nephritis [185]. The distinct cellular and molecular mechanisms for disease progression in different murine models may explain the reported discrepancy [186]. Together, the experimental evidence leads to the speculation that IL-23 and IL-17 producing T cells somewhat orchestrate the dysregulated immune responses in lupus.

In theory, all antibodies targeting IL-23/IL-17 axis which have been approved for the management of other inflammatory rheumatic diseases could be tested in patients with SLE. It is disappointing that the phase 3 clinical trial of Ustekinumab in SLE was discontinued due to lack of expected efficacy following planned futility analysis although the phase 2 results were promising [187]. In addition, Barcitinib, an inhibitor for both JAK1 and JAK2, improved significantly the symptoms in patients with active SLE in a phase 2 trial [188, 189] while the phase 3 trial is ongoing.

Currently, SLE treatment faces big challenges since the results of most trials have been disappointing [190]. Considering the high clinical heterogeneity of SLE [191–193], it becomes self-evident that there is need to administer biologics targeting processes which are responsible for the expression of the disease in defined subsets of patients. The need for personalized treatment in patients with SLE cannot be overemphasized. Along this line, three different trials will be conducted in people with discoid lupus erythematosus (Secukinumab, [NCT03866317\)](https://clinicaltrials.gov/ct2/show/NCT03866317) and patients with active lupus nephritis (Secukinumab, [NCT04181762](https://clinicaltrials.gov/ct2/show/NCT04181762); Guselkumab, [NCT04376827\)](https://clinicaltrials.gov/ct2/show/NCT04376827) (Table 2).

Conclusion

After the establishment of pathogenic role of IL-23/IL-17 axis in different autoimmune and inflammatory diseases [9, 38], diligent work by many groups of researchers, clinicians, and industry colleagues has opened a new era for treatment opportunities for people who suffer from inflammatory rheumatic diseases. Although encouraging clinical results emerge, the overall outcome remains complex [14, 37]. The agents targeting IL-23/IL-17 generally work impressively well in psoriasis and even surpass the effect of anti-TNF therapy but they display only moderate therapeutic efficacy for people with PsA [68, 91]. Moreover, most of these IL-23/IL17 directed biologics have delivered negative results in the management of

RA despite the fact that strong experimental evidence had demonstrated the involvement of this axis in the pathogenesis of RA [15, 48, 50, 140]. Last but not least, even though IL-17 inhibitors have been proven quite effective, IL-23 blocking agents have not shown therapeutic value in the treatment of patients with AS [37]. Despite the advances made in the above discussed diseases, little is known about the role of this axis in other rheumatic diseases including Sjogren's syndrome, vasculitis, and gout.

Given the evidence that IL-23 and IL-17 have complex roles in the development of inflammation in diverse patients with rheumatic diseases [13], the discrepancy of therapeutic values by targeting IL-23/IL-17 axis in different rheumatic diseases [49, 140, 159], the inconsistent clinical outcomes by targeting IL-17 vs targeting IL-23 in the same disease setting, and the diverse response to IL-23/IL-17 axis targeted therapies in patients with the same rheumatic disease (responders vs non-responders) [15, 37, 49, 50], research should be directed towards the identification of subsets of patients with any of these diseases who have better chance of responding to a targeted therapy. The sharing of genetic susceptibility and cellular/molecular pathogenic processes by various rheumatic diseases demands an advanced reclassification of diseases apart from the established clinical nosology. It is obvious that the cellular and molecular processes that we have identified as contributors of the expression of rheumatic diseases are interconnected logically and represent in some ways a "house of cards" whereby removal of any of them results in clinical benefits in certain patients. The efforts to develop biologics to tackle two molecules using a hybrid biologic [194] or administer two biologics simultaneously may represent a promising path to take. Exciting prospects are offered but the use of small molecule inhibitors of intracellular kinases essential for the signal transduction downstream of the engagement of cytokines with their cognate receptors. The opportunity to use these drugs orally represents great advantage to physicians and patients alike. Yet, a simple inspection of any of the diagrams which depict various cytokine-kinase signal cascades reveals the fact that many of them are used interchangeably [195–197]. This is confounded further by the fact that small drug inhibitors are almost never specific. Also, the same kinases are present and are of functional importance in non-immune cells and therefore the short- and long-term side effects should be considered and hopefully can be lessened by carefully titrating the administered dose.

References

- 1. Tsokos GC (2011) Systemic lupus erythematosus. N Engl J Med 365(22):2110–2121. 10.1056/ NEJMra1100359 [PubMed: 22129255]
- 2. Smolen JS, Aletaha D, McInnes IB (2016) Rheumatoid arthritis. Lancet 388(10055):2023–2038. 10.1016/S0140-6736(16)30173-8 [PubMed: 27156434]
- 3. McGonagle D, McDermott MF (2006) A proposed classification of the immunological diseases. PLoS Med 3(8):e297. 10.1371/journal.pmed.0030297 [PubMed: 16942393]
- 4. Nestle FO, Kaplan DH, Barker J (2009) Psoriasis. N Engl J Med 361(5):496–509. 10.1056/ NEJMra0804595 [PubMed: 19641206]
- 5. Sieper J, Poddubnyy D (2017) Axial spondyloarthritis. Lancet 390(10089):73–84. 10.1016/ S0140-6736(16)31591-4 [PubMed: 28110981]
- 6. Lewis JE, Fu SM, Gaskin F (2013) Autoimmunity, end organ damage, and the origin of autoantibodies and autoreactive T cells in systemic lupus erythematosus. Discov Med 15(81):85–92 [PubMed: 23449110]

- 7. Tsokos GC (2020) Autoimmunity and organ damage in systemic lupus erythematosus. Nat Immunol 21(6):605–614. 10.1038/s41590-020-0677-6 [PubMed: 32367037]
- 8. Chabaud M, Fossiez F, Taupin JL, Miossec P (1998) Enhancing effect of IL-17 on IL-1-induced IL-6 and leukemia inhibitory factor production by rheumatoid arthritis synoviocytes and its regulation by Th2 cytokines. J Immunol 161(1):409–414 [PubMed: 9647250]
- 9. Miossec P, Kolls JK (2012) Targeting IL-17 and TH17 cells in chronic inflammation. Nat Rev Drug Discov 11(10):763–776. 10.1038/nrd3794 [PubMed: 23023676]
- 10. Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, Armitage RJ (1995) Human IL-17: a novel cytokine derived from T cells. J Immunol 155(12):5483–5486 [PubMed: 7499828]
- 11. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, Weaver CT (2005) Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. Nat Immunol 6(11):1123–1132. 10.1038/ni1254 [PubMed: 16200070]
- 12. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, Wang Y, Hood L, Zhu Z, Tian Q, Dong C (2005) A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. Nat Immunol 6(11):1133–1141. 10.1038/ni1261 [PubMed: 16200068]
- 13. Gaffen SL, Jain R, Garg AV, Cua DJ (2014) The IL-23-IL-17 immune axis: from mechanisms to therapeutic testing. Nat Rev Immunol 14(9):585–600. 10.1038/nri3707 [PubMed: 25145755]
- 14. Iwakura Y, Ishigame H (2006) The IL-23/IL-17 axis in inflammation. J Clin Invest 116(5):1218– 1222. 10.1172/JCI28508 [PubMed: 16670765]
- 15. Beringer A, Noack M, Miossec P (2016) IL-17 in chronic inflammation: From discovery to targeting. Trends Mol Med 22(3):230–241. 10.1016/j.molmed.2016.01.001 [PubMed: 26837266]
- 16. Theis KA, Roblin DW, Helmick CG, Luo R (2018) Prevalence and causes of work disability among working-age US adults 2011–2013 NHIS. Disabil Health J 11(1):108–115. 10.1016/ j.dhjo.2017.04.010 [PubMed: 28476583]
- 17. Hootman JM, Helmick CG, Barbour KE, Theis KA, Boring MA (2016) Updated projected prevalence of self-reported doctor-diagnosed arthritis and arthritis-attributable activity limitation among US adults, 2015–2040. Arthritis Rheumatol 68(7):1582–1587. 10.1002/art.39692 [PubMed: 27015600]
- 18. Rachakonda TD, Schupp CW, Armstrong AW (2014) Psoriasis prevalence among adults in the United States. J Am Acad Dermatol 70(3):512–516. 10.1016/j.jaad.2013.11.013 [PubMed: 24388724]
- 19. Saad J, Mathew D (2020) Nonsteroidal Anti-Inflammatory Drugs (NSAID) Toxicity. In: StatPearls. Treasure Island (FL)
- 20. Sostres C, Gargallo CJ, Lanas A (2013) Nonsteroidal anti-inflammatory drugs and upper and lower gastrointestinal mucosal damage. Arthritis Res Ther 15(Suppl 3):S3. 10.1186/ar4175
- 21. Shirota Y, Illei GG, Nikolov NP (2008) Biologic treatments for systemic rheumatic diseases. Oral Dis 14(3):206–216. 10.1111/j.1601-0825.2008.01440.x [PubMed: 18282173]
- 22. Wellcome Trust Case Control C, Australo-Anglo-American Spondylitis C, Burton PR, Clayton DG, Cardon LR, Craddock N, Deloukas P, Duncanson A, Kwiatkowski DP, McCarthy MI, Ouwehand WH, Samani NJ, Todd JA, Donnelly P, Barrett JC, Davison D, Easton D, Evans DM, Leung HT, Marchini JL, Morris AP, Spencer CC, Tobin MD, Attwood AP, Boorman JP, Cant B, Everson U, Hussey JM, Jolley JD, Knight AS, Koch K, Meech E, Nutland S, Prowse CV, Stevens HE, Taylor NC, Walters GR, Walker NM, Watkins NA, Winzer T, Jones RW, McArdle WL, Ring SM, Strachan DP, Pembrey M, Breen G, St Clair D, Caesar S, Gordon-Smith K, Jones L, Fraser C, Green EK, Grozeva D, Hamshere ML, Holmans PA, Jones IR, Kirov G, Moskivina V, Nikolov I, O'Donovan MC, Owen MJ, Collier DA, Elkin A, Farmer A, Williamson R, McGuffin P, Young AH, Ferrier IN, Ball SG, Balmforth AJ, Barrett JH, Bishop TD, Iles MM, Maqbool A, Yuldasheva N, Hall AS, Braund PS, Dixon RJ, Mangino M, Stevens S, Thompson JR, Bredin F, Tremelling M, Parkes M, Drummond H, Lees CW, Nimmo ER, Satsangi J, Fisher SA, Forbes A, Lewis CM, Onnie CM, Prescott NJ, Sanderson J, Matthew CG, Barbour J, Mohiuddin MK, Todhunter CE, Mansfield JC, Ahmad T, Cummings FR, Jewell DP, Webster J, Brown MJ, Lathrop MG, Connell J, Dominiczak A, Marcano CA, Burke B, Dobson R, Gungadoo J, Lee KL, Munroe PB, Newhouse SJ, Onipinla A, Wallace C, Xue M, Caulfield M, Farrall M, Barton A, Biologics in RAG, Genomics Study Syndicate Steering C, Bruce IN, Donovan H, Eyre S, Gilbert PD, Hilder SL, Hinks AM, John SL, Potter C, Silman AJ, Symmons DP, Thomson W, Worthington J, Dunger DB,

Widmer B, Frayling TM, Freathy RM, Lango H, Perry JR, Shields BM, Weedon MN, Hattersley AT, Hitman GA, Walker M, Elliott KS, Groves CJ, Lindgren CM, Rayner NW, Timpson NJ, Zeggini E, Newport M, Sirugo G, Lyons E, Vannberg F, Hill AV, Bradbury LA, Farrar C, Pointon JJ, Wordsworth P, Brown MA, Franklyn JA, Heward JM, Simmonds MJ, Gough SC, Seal S, Breast Cancer Susceptibility C, Stratton MR, Rahman N, Ban M, Goris A, Sawcer SJ, Compston A, Conway D, Jallow M, Newport M, Sirugo G, Rockett KA, Bumpstead SJ, Chaney A, Downes K, Ghori MJ, Gwilliam R, Hunt SE, Inouye M, Keniry A, King E, McGinnis R, Potter S, Ravindrarajah R, Whittaker P, Widden C, Withers D, Cardin NJ, Davison D, Ferreira T, Pereira-Gale J, Hallgrimsdo'ttir IB, Howie BN, Su Z, Teo YY, Vukcevic D, Bentley D, Brown MA, Compston A, Farrall M, Hall AS, Hattersley AT, Hill AV, Parkes M, Pembrey M, Stratton MR, Mitchell SL, Newby PR, Brand OJ, Carr-Smith J, Pearce SH, McGinnis R, Keniry A, Deloukas P, Reveille JD, Zhou X, Sims AM, Dowling A, Taylor J, Doan T, Davis JC, Savage L, Ward MM, Learch TL, Weisman MH, Brown M (2007) Association scan of 14,500 nonsynonymous SNPs in four diseases identifies autoimmunity variants. Nat Genet 39(11):1329–1337. 10.1038/ng.2007.17 [PubMed: 17952073]

- 23. Mells GF, Hirschfield GM (2015) Making the most of new genetic risk factors genetic and epigenetic fine mapping of causal autoimmune disease variants. Clin Res Hepatol Gastroenterol 39(4):408–411. 10.1016/j.clinre.2015.05.002 [PubMed: 26160476]
- 24. Farh KK, Marson A, Zhu J, Kleinewietfeld M, Housley WJ, Beik S, Shoresh N, Whitton H, Ryan RJ, Shishkin AA, Hatan M, Carrasco-Alfonso MJ, Mayer D, Luckey CJ, Patsopoulos NA, De Jager PL, Kuchroo VK, Epstein CB, Daly MJ, Hafler DA, Bernstein BE (2015) Genetic and epigenetic fine mapping of causal autoimmune disease variants. Nature 518(7539):337–343. 10.1038/nature13835 [PubMed: 25363779]
- 25. Sarin R, Wu X, Abraham C (2011) Inflammatory disease protective R381Q IL23 receptor polymorphism results in decreased primary CD4+ and CD8+ human T-cell functional responses. Proc Natl Acad Sci U S A 108(23):9560–9565. 10.1073/pnas.1017854108 [PubMed: 21606346]
- 26. Mease PJ, McInnes IB, Kirkham B, Kavanaugh A, Rahman P, van der Heijde D, Landewe R, Nash P, Pricop L, Yuan J, Richards HB, Mpofu S, Group FS (2015) Secukinumab inhibition of interleukin-17A in patients with psoriatic arthritis. N Engl J Med 373(14):1329–1339. 10.1056/ NEJMoa1412679 [PubMed: 26422723]
- 27. Mease PJ, van der Heijde D, Ritchlin CT, Okada M, Cuchacovich RS, Shuler CL, Lin CY, Braun DK, Lee CH, Gladman DD, Group S-PS (2017) Ixekizumab, an interleukin-17A specific monoclonal antibody, for the treatment of biologic-naive patients with active psoriatic arthritis: results from the 24-week randomised, double-blind, placebo-controlled and active (adalimumab) controlled period of the phase III trial SPIRIT-P1. Ann Rheum Dis 76(1):79–87. 10.1136/ annrheumdis-2016-209709 [PubMed: 27553214]
- 28. Deodhar A, Gottlieb AB, Boehncke WH, Dong B, Wang Y, Zhuang Y, Barchuk W, Xu XL, Hsia EC, Group CS (2018) Efficacy and safety of guselkumab in patients with active psoriatic arthritis: a randomised, double-blind, placebo-controlled, phase 2 study. Lancet 391(10136):2213–2224. 10.1016/S0140-6736(18)30952-8 [PubMed: 29893222]
- 29. Gordon KB, Duffin KC, Bissonnette R, Prinz JC, Wasfi Y, Li S, Shen YK, Szapary P, Randazzo B, Reich K (2015) A phase 2 trial of guselkumab versus adalimumab for plaque psoriasis. N Engl J Med 373(2):136–144. 10.1056/NEJMoa1501646 [PubMed: 26154787]
- 30. Gordon KB, Strober B, Lebwohl M, Augustin M, Blauvelt A, Poulin Y, Papp KA, Sofen H, Puig L, Foley P, Ohtsuki M, Flack M, Geng Z, Gu Y, Valdes JM, Thompson EHZ, Bachelez H (2018) Efficacy and safety of risankizumab in moderate-to-severe plaque psoriasis (UltIMMa-1 and UltIMMa-2): results from two double-blind, randomised, placebo-controlled and ustekinumabcontrolled phase 3 trials. Lancet 392(10148):650–661. 10.1016/S0140-6736(18)31713-6 [PubMed: 30097359]
- 31. Smolen JS, Agarwal SK, Ilivanova E, Xu XL, Miao Y, Zhuang Y, Nnane I, Radziszewski W, Greenspan A, Beutler A, Baker D (2017) A randomised phase II study evaluating the efficacy and safety of subcutaneously administered ustekinumab and guselkumab in patients with active rheumatoid arthritis despite treatment with methotrexate. Ann Rheum Dis 76(5):831–839. 10.1136/annrheumdis-2016-209831 [PubMed: 28087506]
- 32. Blanco FJ, Moricke R, Dokoupilova E, Codding C, Neal J, Andersson M, Rohrer S, Richards H (2017) Secukinumab in active rheumatoid arthritis: a phase III randomized, double-blind, active

comparator- and placebo-controlled study. Arthritis Rheumatol 69(6):1144–1153. 10.1002/ art.40070 [PubMed: 28217871]

- 33. Deodhar A, Gensler LS, Sieper J, Clark M, Calderon C, Wang Y, Zhou Y, Leu JH, Campbell K, Sweet K, Harrison DD, Hsia EC, van der Heijde D (2019) Three multicenter, randomized, doubleblind, placebo-controlled studies evaluating the efficacy and safety of ustekinumab in axial spondyloarthritis. Arthritis Rheumatol 71(2):258–270. 10.1002/art.40728 [PubMed: 30225992]
- 34. Baeten D, Ostergaard M, Wei JC, Sieper J, Jarvinen P, Tam LS, Salvarani C, Kim TH, Solinger A, Datsenko Y, Pamulapati C, Visvanathan S, Hall DB, Aslanyan S, Scholl P, Padula SJ (2018) Risankizumab, an IL-23 inhibitor, for ankylosing spondylitis: results of a randomised, doubleblind, placebo-controlled, proof-of-concept, dose-finding phase 2 study. Ann Rheum Dis 77(9):1295–1302. 10.1136/annrheumdis-2018-213328 [PubMed: 29945918]
- 35. Baeten D, Sieper J, Braun J, Baraliakos X, Dougados M, Emery P, Deodhar A, Porter B, Martin R, Andersson M, Mpofu S, Richards HB, Group MS (2015) Secukinumab, an Interleukin-17A Inhibitor. Ankylosing Spondylitis N Engl J Med 373(26):2534–2548. 10.1056/NEJMoa1505066 [PubMed: 26699169]
- 36. van der Heijde D, Cheng-Chung Wei J, Dougados M, Mease P, Deodhar A, Maksymowych WP, Van den Bosch F, Sieper J, Tomita T, Landewe R, Zhao F, Krishnan E, Adams DH, Pangallo B, Carliergroup C-Vs, H (2018) Ixekizumab, an interleukin-17A antagonist in the treatment of ankylosing spondylitis or radiographic axial spondyloarthritis in patients previously untreated with biological disease-modifying anti-rheumatic drugs (COAST-V): 16 week results of a phase 3 randomised, double-blind, active-controlled and placebo-controlled trial. Lancet 392(10163):2441–2451. 10.1016/S0140-6736(18)31946-9 [PubMed: 30360964]
- 37. Sieper J, Poddubnyy D, Miossec P (2019) The IL-23-IL-17 pathway as a therapeutic target in axial spondyloarthritis. Nat Rev Rheumatol 15(12):747–757. 10.1038/s41584-019-0294-7 [PubMed: 31551538]
- 38. Teng MW, Bowman EP, McElwee JJ, Smyth MJ, Casanova JL, Cooper AM, Cua DJ (2015) IL-12 and IL-23 cytokines: from discovery to targeted therapies for immune-mediated inflammatory diseases. Nat Med 21(7):719–729. 10.1038/nm.3895 [PubMed: 26121196]
- 39. Sheibanie AF, Tadmori I, Jing H, Vassiliou E, Ganea D (2004) Prostaglandin E2 induces IL-23 production in bone marrow-derived dendritic cells. FASEB J 18(11):1318–1320. 10.1096/ fj.03-1367fje [PubMed: 15180965]
- 40. Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, Vega F, Yu N, Wang J, Singh K, Zonin F, Vaisberg E, Churakova T, Liu M, Gorman D, Wagner J, Zurawski S, Liu Y, Abrams JS, Moore KW, Rennick D, de Waal-Malefyt R, Hannum C, Bazan JF, Kastelein RA (2000) Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immunity 13(5):715–725. 10.1016/s1074-7613(00)00070-4 [PubMed: 11114383]
- 41. Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, Lucian L, To W, Kwan S, Churakova T, Zurawski S, Wiekowski M, Lira SA, Gorman D, Kastelein RA, Sedgwick JD (2003) Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 421(6924):744–748. 10.1038/nature01355 [PubMed: 12610626]
- 42. Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, Sedgwick JD, Cua DJ (2003) Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. J Exp Med 198(12):1951–1957. 10.1084/jem.20030896 [PubMed: 14662908]
- 43. Wiekowski MT, Leach MW, Evans EW, Sullivan L, Chen SC, Vassileva G, Bazan JF, Gorman DM, Kastelein RA, Narula S, Lira SA (2001) Ubiquitous transgenic expression of the IL-23 subunit p19 induces multiorgan inflammation, runting, infertility, and premature death. J Immunol 166(12):7563–7570. 10.4049/jimmunol.166.12.7563 [PubMed: 11390512]
- 44. Sherlock JP, Joyce-Shaikh B, Turner SP, Chao CC, Sathe M, Grein J, Gorman DM, Bowman EP, McClanahan TK, Yearley JH, Eberl G, Buckley CD, Kastelein RA, Pierce RH, Laface DM, Cua DJ (2012) IL-23 induces spondyloarthropathy by acting on ROR-gammat+ CD3+CD4-CD8 entheseal resident T cells. Nat Med 18(7):1069–1076. 10.1038/nm.2817 [PubMed: 22772566]
- 45. Adamopoulos IE, Tessmer M, Chao CC, Adda S, Gorman D, Petro M, Chou CC, Pierce RH, Yao W, Lane NE, Laface D, Bowman EP (2011) IL-23 is critical for induction of arthritis, osteoclast

formation, and maintenance of bone mass. J Immunol 187(2):951–959. 10.4049/ jimmunol.1003986 [PubMed: 21670317]

- 46. Gaffen SL (2009) Structure and signalling in the IL-17 receptor family. Nat Rev Immunol 9(8):556–567. 10.1038/nri2586 [PubMed: 19575028]
- 47. Wynn TA (2005) T(H)-17: a giant step from T(H)1 and T(H)2. Nat Immunol 6(11):1069–1070. 10.1038/ni1105-1069 [PubMed: 16239919]
- 48. Hot A, Miossec P (2011) Effects of interleukin (IL)-17A and IL-17F in human rheumatoid arthritis synoviocytes. Ann Rheum Dis 70(5):727–732. 10.1136/ard.2010.143768 [PubMed: 21345813]
- 49. Lubberts E (2015) The IL-23-IL-17 axis in inflammatory arthritis. Nat Rev Rheumatol 11(10):562. 10.1038/nrrheum.2015.128
- 50. Beringer A, Miossec P (2019) Systemic effects of IL-17 in inflammatory arthritis. Nat Rev Rheumatol 15(8):491–501. 10.1038/s41584-019-0243-5 [PubMed: 31227819]
- 51. Robert M, Miossec P (2018) IL-17 in rheumatoid arthritis and precision medicine: from synovitis expression to circulating bioactive levels. Front Med (Lausanne) 5:364. 10.3389/fmed.2018.00364 [PubMed: 30693283]
- 52. Adamopoulos IE, Chao CC, Geissler R, Laface D, Blumenschein W, Iwakura Y, McClanahan T, Bowman EP (2010) Interleukin-17A upregulates receptor activator of NF-kappaB on osteoclast precursors. Arthritis Res Ther 12(1):R29. 10.1186/ar2936 [PubMed: 20167120]
- 53. Yago T, Nanke Y, Ichikawa N, Kobashigawa T, Mogi M, Kamatani N, Kotake S (2009) IL-17 induces osteoclastogenesis from human monocytes alone in the absence of osteoblasts, which is potently inhibited by anti-TNF-alpha antibody: A novel mechanism of osteoclastogenesis by IL-17. J Cell Biochem 108(4):947–955. 10.1002/jcb.22326 [PubMed: 19728295]
- 54. Mitsdoerffer M, Lee Y, Jager A, Kim HJ, Korn T, Kolls JK, Cantor H, Bettelli E, Kuchroo VK (2010) Proinflammatory T helper type 17 cells are effective B-cell helpers. Proc Natl Acad Sci U S A 107(32):14292–14297. 10.1073/pnas.1009234107 [PubMed: 20660725]
- 55. Hsu HC, Yang P, Wang J, Wu Q, Myers R, Chen J, Yi J, Guentert T, Tousson A, Stanus AL, Le TV, Lorenz RG, Xu H, Kolls JK, Carter RH, Chaplin DD, Williams RW, Mountz JD (2008) Interleukin 17-producing T helper cells and interleukin 17 orchestrate autoreactive germinal center development in autoimmune BXD2 mice. Nat Immunol 9(2):166–175. 10.1038/ni1552 [PubMed: 18157131]
- 56. Majumder S, Amatya N, Revu S, Jawale CV, Wu D, Rittenhouse N, Menk A, Kupul S, Du F, Raphael I, Bhattacharjee A, Sie-benlist U, Hand TW, Delgoffe GM, Poholek AC, Gaffen SL, Biswas PS, McGeachy MJ (2019) IL-17 metabolically reprograms activated fibroblastic reticular cells for proliferation and survival. Nat Immunol 20(5):534–545. 10.1038/s41590-019-0367-4 [PubMed: 30962593]
- 57. McGeachy MJ, Chen Y, Tato CM, Laurence A, Joyce-Shaikh B, Blumenschein WM, McClanahan TK, O'Shea JJ, Cua DJ (2009) The interleukin 23 receptor is essential for the terminal differentiation of interleukin 17-producing effector T helper cells in vivo. Nat Immunol 10(3):314– 324. 10.1038/ni.1698 [PubMed: 19182808]
- 58. Li H, Hsu HC, Wu Q, Yang P, Li J, Luo B, Oukka M, Steele CH 3rd, Cua DJ, Grizzle WE, Mountz JD (2014) IL-23 promotes TCR-mediated negative selection of thymocytes through the upregulation of IL-23 receptor and RORgammat. Nat Commun 5:4259. 10.1038/ncomms5259 [PubMed: 25001511]
- 59. Koga T, Ichinose K, Kawakami A, Tsokos GC (2019) The role of IL-17 in systemic lupus erythematosus and its potential as a therapeutic target. Expert Rev Clin Immunol 15(6):629–637. 10.1080/1744666X.2019.1593141 [PubMed: 30874446]
- 60. Huffmeier U, Lascorz J, Bohm B, Lohmann J, Wendler J, Mossner R, Reich K, Traupe H, Kurrat W, Burkhardt H, Reis A (2009) Genetic variants of the IL-23R pathway: association with psoriatic arthritis and psoriasis vulgaris, but no specific risk factor for arthritis. J Invest Dermatol 129(2):355–358. 10.1038/jid.2008.233 [PubMed: 18800148]
- 61. Di Meglio P, Di Cesare A, Laggner U, Chu CC, Napolitano L, Villanova F, Tosi I, Capon F, Trembath RC, Peris K, Nestle FO (2011) The IL23R R381Q gene variant protects against immune-mediated diseases by impairing IL-23-induced Th17 effector response in humans. PLoS ONE 6(2):e17160. 10.1371/journal.pone.0017160 [PubMed: 21364948]

- 62. Uddin M, Codner D, Hasan SM, Scherer SW, O'Rielly DD, Rahman P (2015) Integrated genomics identifies convergence of ankylosing spondylitis with global immune mediated disease pathways. Sci Rep 5:10314. 10.1038/srep10314 [PubMed: 25980808]
- 63. Shao M, Xu S, Yang H, Xu W, Deng J, Chen Y, Gao X, Guan S, Xu S, Shuai Z, Pan F (2020) Association between IL-17A and IL-17F gene polymorphism and susceptibility in inflammatory arthritis: a meta-analysis. Clin Immunol 213:108374. 10.1016/j.clim.2020.108374 [PubMed: 32146336]
- 64. Song K, Liu L, Zhang X, Chen X (2020) An update on genetic susceptibility in lupus nephritis. Clin Immunol 210:108272. 10.1016/j.clim.2019.108272 [PubMed: 31683055]
- 65. Spach KM, Noubade R, McElvany B, Hickey WF, Blankenhorn EP, Teuscher C (2009) A single nucleotide polymorphism in Tyk2 controls susceptibility to experimental allergic encephalomyelitis. J Immunol 182(12):7776–7783. 10.4049/jimmunol.0900142 [PubMed: 19494301]
- 66. Gorman JA, Hundhausen C, Kinsman M, Arkatkar T, Allenspach EJ, Clough C, West SE, Thomas K, Eken A, Khim S, Hale M, Oukka M, Jackson SW, Cerosaletti K, Buckner JH, Rawlings DJ (2019) The TYK2-P1104A autoimmune protective variant limits coordinate signals required to generate specialized T cell subsets. Front Immunol 10:44. 10.3389/fimmu.2019.00044 [PubMed: 30740104]
- 67. Shaw MH, Boyartchuk V, Wong S, Karaghiosoff M, Ragimbeau J, Pellegrini S, Muller M, Dietrich WF, Yap GS (2003) A natural mutation in the Tyk2 pseudokinase domain underlies altered susceptibility of B10.Q/J mice to infection and autoimmunity. Proc Natl Acad Sci U S A 100(20):11594–11599. 10.1073/pnas.1930781100 [PubMed: 14500783]
- 68. Suzuki E, Mellins ED, Gershwin ME, Nestle FO, Adamopoulos IE (2014) The IL-23/IL-17 axis in psoriatic arthritis. Autoimmun Rev 13(4–5):496–502. 10.1016/j.autrev.2014.01.050 [PubMed: 24424175]
- 69. Jeon C, Sekhon S, Yan D, Afifi L, Nakamura M, Bhutani T (2017) Monoclonal antibodies inhibiting IL-12, −23, and −17 for the treatment of psoriasis. Hum Vaccin Immunother 13(10):2247–2259. 10.1080/21645515.2017.1356498 [PubMed: 28825875]
- 70. Robert M, Miossec P (2020) Interleukin-17 and lupus: enough to be a target? For which patients? Lupus 29(1):6–14. 10.1177/0961203319891243 [PubMed: 31791181]
- 71. Silacci M, Lembke W, Woods R, Attinger-Toller I, Baenziger-Tobler N, Batey S, Santimaria R, von der Bey U, Koenig-Friedrich S, Zha W, Schlereth B, Locher M, Bertschinger J, Grabulovski D (2016) Discovery and characterization of COVA322, a clinical-stage bispecific TNF/IL-17A inhibitor for the treatment of inflammatory diseases. MAbs 8(1):141–149. 10.1080/19420862.2015.1093266 [PubMed: 26390837]
- 72. Mease PJ, Genovese MC, Weinblatt ME, Peloso PM, Chen K, Othman AA, Li Y, Mansikka HT, Khatri A, Wishart N, Liu J (2018) Phase II study of ABT-122, a tumor necrosis factor- and interleukin-17A-targeted dual variable domain immunoglobulin, in patients with psoriatic arthritis with an inadequate response to methotrexate. Arthritis Rheumatol 70(11):1778–1789. 10.1002/ art.40579 [PubMed: 29855175]
- 73. Ritchlin CT, Kavanaugh A, Merola JF, Schett G, Scher JU, Warren RB, Gottlieb AB, Assudani D, Bedford-Rice K, Coarse J, Ink B, McInnes IB (2020) Bimekizumab in patients with active psoriatic arthritis: results from a 48-week, randomised, double-blind, placebo-controlled, doseranging phase 2b trial. Lancet 395(10222):427–440. 10.1016/S0140-6736(19)33161-7 [PubMed: 32035552]
- 74. Svecova D, Lubell MW, Casset-Semanaz F, Mackenzie H, Gren-ningloh R, Krueger JG (2019) A randomized, double-blind, placebo-controlled phase 1 study of multiple ascending doses of subcutaneous M1095, an anti-interleukin 17A/F nanobody, in moderate-to-severe psoriasis. J Am Acad Dermatol 81(1):196–203. 10.1016/j.jaad.2019.03.056 [PubMed: 30926369]
- 75. Chiricozzi A, De Simone C, Fossati B, Peris K (2019) Emerging treatment options for the treatment of moderate to severe plaque psoriasis and psoriatic arthritis: evaluating bimekizumab and its therapeutic potential. Psoriasis (Auckl) 9:29–35. 10.2147/PTT.S179283 [PubMed: 31214486]

- 76. Weaver CT, Hatton RD, Mangan PR, Harrington LE (2007) IL-17 family cytokines and the expanding diversity of effector T cell lineages. Annu Rev Immunol 25:821–852. 10.1146/ annurev.immunol.25.022106.141557 [PubMed: 17201677]
- 77. Vignali DA, Kuchroo VK (2012) IL-12 family cytokines: immunological playmakers. Nat Immunol 13(8):722–728. 10.1038/ni.2366 [PubMed: 22814351]
- 78. Zhou L, Littman DR (2009) Transcriptional regulatory networks in Th17 cell differentiation. Curr Opin Immunol 21(2):146–152. 10.1016/j.coi.2009.03.001 [PubMed: 19328669]
- 79. Ivanov II, Zhou L, Littman DR (2007) Transcriptional regulation of Th17 cell differentiation. Semin Immunol 19(6):409–417. 10.1016/j.smim.2007.10.011 [PubMed: 18053739]
- 80. Huh JR, Littman DR (2012) Small molecule inhibitors of RORgammat: targeting Th17 cells and other applications. Eur J Immunol 42(9):2232–2237. 10.1002/eji.201242740 [PubMed: 22949321]
- 81. Schwartz DM, Kanno Y, Villarino A, Ward M, Gadina M, O'Shea JJ (2017a) JAK inhibition as a therapeutic strategy for immune and inflammatory diseases. Nat Rev Drug Discov 16(12):843– 862. 10.1038/nrd.2017.201 [PubMed: 29104284]
- 82. O'Shea JJ, Kontzias A, Yamaoka K, Tanaka Y, Laurence A (2013) Janus kinase inhibitors in autoimmune diseases. Ann Rheum Dis 72(Suppl 2):ii111–ii115. 10.1136/ annrheumdis-2012-202576 [PubMed: 23532440]
- 83. Leonard WJ, O'Shea JJ (1998) Jaks and STATs: biological implications. Annu Rev Immunol 16:293–322. 10.1146/annurev.immunol.16.1.293 [PubMed: 9597132]
- 84. Mogul A, Corsi K, McAuliffe L (2019) Baricitinib: the second FDA-approved JAK inhibitor for the treatment of rheumatoid arthritis. Ann Pharmacother 53(9):947–953. 10.1177/1060028019839650 [PubMed: 30907116]
- 85. Taylor PC (2019) Clinical efficacy of launched JAK inhibitors in rheumatoid arthritis. Rheumatology (Oxford) 58(Suppl 1):i17–i26. 10.1093/rheumatology/key225 [PubMed: 30806707]
- 86. Fleischmann R (2017) A review of tofacitinib efficacy in rheumatoid arthritis patients who have had an inadequate response or intolerance to methotrexate. Expert Opin Pharmacother 18(14):1525–1533. 10.1080/14656566.2017.1370453 [PubMed: 28829236]
- 87. Wang F, Sun L, Wang S, Davis JM 3rd, Matteson EL, Murad MH, Luo F, Vassallo R (2020) Efficacy and safety of tofacitinib, baricitinib, and upadacitinib for rheumatoid arthritis: a systematic review and meta-analysis. Mayo Clin Proc 95(7):1404–1419. 10.1016/ j.mayocp.2020.01.039 [PubMed: 32499126]
- 88. Takeshita J, Grewal S, Langan SM, Mehta NN, Ogdie A, Van Voorhees AS, Gelfand JM (2017) Psoriasis and comorbid diseases: implications for management. J Am Acad Dermatol 76(3):393– 403. 10.1016/j.jaad.2016.07.065 [PubMed: 28212760]
- 89. Harden JL, Krueger JG, Bowcock AM (2015) The immunogenetics of psoriasis: a comprehensive review. J Autoimmun 64:66–73. 10.1016/j.jaut.2015.07.008 [PubMed: 26215033]
- 90. Hawkes JE, Yan BY, Chan TC, Krueger JG (2018) Discovery of the IL-23/IL-17 signaling pathway and the treatment of psoriasis. J Immunol 201(6):1605–1613. 10.4049/jimmunol.1800013 [PubMed: 30181299]
- 91. Boutet MA, Nerviani A, Gallo Afflitto G, Pitzalis C (2018) Role of the IL-23/IL-17 Axis in Psoriasis and Psoriatic Arthritis: The Clinical Importance of Its Divergence in Skin and Joints. Int J Mol Sci 19 (2). doi:10.3390/ijms19020530
- 92. Parisi R, Symmons DP, Griffiths CE, Ashcroft DM, Identification of P, Associated ComorbidiTy project t, M (2013) Global epidemiology of psoriasis: a systematic review of incidence and prevalence. J Invest Dermatol 133(2):377–385. 10.1038/jid.2012.339 [PubMed: 23014338]
- 93. Ogdie A, Weiss P (2015) The epidemiology of psoriatic arthritis. Rheum Dis Clin North Am 41(4):545–568. 10.1016/j.rdc.2015.07.001 [PubMed: 26476218]
- 94. Rouzaud M, Sevrain M, Villani AP, Barnetche T, Paul C, Richard MA, Jullien D, Misery L, Le Maitre M, Aractingi S, Aubin F, Joly P, Cantagrel A, Ortonne JP, Beylot-Barry M (2014) Is there a psoriasis skin phenotype associated with psoriatic arthritis? Systematic literature review. J Eur Acad Dermatol Venereol 28(Suppl 5):17–26. 10.1111/jdv.12562 [PubMed: 24985559]
- 95. Rida MA, Chandran V (2020) Challenges in the clinical diagnosis of psoriatic arthritis. Clin Immunol 214:108390. 10.1016/j.clim.2020.108390 [PubMed: 32200113]

- 96. Kim WB, Jerome D, Yeung J (2017) Diagnosis and management of psoriasis. Can Fam Physician 63(4):278–285 [PubMed: 28404701]
- 97. Ritchlin CT, Kavanaugh A, Gladman DD, Mease PJ, Helliwell P, Boehncke WH, de Vlam K, Fiorentino D, Fitzgerald O, Gottlieb AB, McHugh NJ, Nash P, Qureshi AA, Soriano ER, Taylor WJ, Group for R, Assessment of P, Psoriatic A (2009) Treatment recommendations for psoriatic arthritis. Ann Rheum Dis 68(9):1387–1394. 10.1136/ard.2008.094946 [PubMed: 18952643]
- 98. Toussi A, Maverakis N, Le ST, Sarkar S, Raychaudhuri SK, Raychaudhuri SP (2020) Updated therapies for the management of psoriatic arthritis. Clin Immunol 220:108536. 10.1016/ j.clim.2020.108536 [PubMed: 32681979]
- 99. Bell S, Nahle Z, Adamopoulos IE (2020) Psoriatic arthritis; overcoming the challenges by creating opportunities. Clin Immunol 218:108519. 10.1016/j.clim.2020.108519 [PubMed: 32621978]
- 100. Ritchlin CT, Colbert RA, Gladman DD (2017) Psoriatic Arthritis. N Engl J Med 376(10):957– 970. 10.1056/NEJMra1505557 [PubMed: 28273019]
- 101. Eder L, Chandran V, Gladman DD (2015) What have we learned about genetic susceptibility in psoriasis and psoriatic arthritis? Curr Opin Rheumatol 27(1):91–98. 10.1097/ BOR.0000000000000136 [PubMed: 25415529]
- 102. Bowes J, Ashcroft J, Dand N, Jalali-Najafabadi F, Bellou E, Ho P, Marzo-Ortega H, Helliwell PS, Feletar M, Ryan AW, Kane DJ, Korendowych E, Simpson MA, Packham J, McManus R, Brown MA, Smith CH, Barker JN, McHugh N, FitzGerald O, Warren RB, Barton A (2017) Crossphenotype association mapping of the MHC identifies genetic variants that differentiate psoriatic arthritis from psoriasis. Ann Rheum Dis 76(10):1774–1779. 10.1136/annrheumdis-2017-211414 [PubMed: 28821532]
- 103. Di Cesare A, Di Meglio P, Nestle FO (2009) The IL-23/Th17 axis in the immunopathogenesis of psoriasis. J Invest Dermatol 129(6):1339–1350. 10.1038/jid.2009.59 [PubMed: 19322214]
- 104. Prinz I, Sandrock I, Mrowietz U (2020) Interleukin-17 cytokines: Effectors and targets in psoriasis-A breakthrough in understanding and treatment. J Exp Med 217 (1). doi:10.1084/ jem.20191397
- 105. Hile G, Kahlenberg JM, Gudjonsson JE (2020) Recent genetic advances in innate immunity of psoriatic arthritis. Clin Immunol 214:108405. 10.1016/j.clim.2020.108405 [PubMed: 32247832]
- 106. Filer C, Ho P, Smith RL, Griffiths C, Young HS, Worthington J, Bruce IN, Barton A (2008) Investigation of association of the IL12B and IL23R genes with psoriatic arthritis. Arthritis Rheum 58(12):3705–3709. 10.1002/art.24128 [PubMed: 19035472]
- 107. Valdimarsson H (2007) The genetic basis of psoriasis. Clin Dermatol 25(6):563–567. 10.1016/ j.clindermatol.2007.08.010 [PubMed: 18021893]
- 108. Tomfohrde J, Silverman A, Barnes R, Fernandez-Vina MA, Young M, Lory D, Morris L, Wuepper KD, Stastny P, Menter A et al. (1994) Gene for familial psoriasis susceptibility mapped to the distal end of human chromosome 17q. Science 264(5162):1141–1145. 10.1126/ science.8178173 [PubMed: 8178173]
- 109. Wang A, Bai Y (2020) Dendritic cells: The driver of psoriasis. J Dermatol 47(2):104–113. 10.1111/1346-8138.15184 [PubMed: 31833093]
- 110. Chen L, Deshpande M, Grisotto M, Smaldini P, Garcia R, He Z, Gulko PS, Lira SA, Furtado GC (2020) Skin expression of IL-23 drives the development of psoriasis and psoriatic arthritis in mice. Sci Rep 10(1):8259. 10.1038/s41598-020-65269-6 [PubMed: 32427877]
- 111. Miossec P (2003) Interleukin-17 in rheumatoid arthritis: If T cells were to contribute to inflammation and destruction through synergy. Arthritis Rheum 48(3):594–601. 10.1002/ art.10816 [PubMed: 12632409]
- 112. Wang CQF, Akalu YT, Suarez-Farinas M, Gonzalez J, Mitsui H, Lowes MA, Orlow SJ, Manga P, Krueger JG (2013) IL-17 and TNF synergistically modulate cytokine expression while suppressing melanogenesis: Potential relevance to psoriasis. J Invest Dermatol 133(12):2741– 2752. 10.1038/jid.2013.237 [PubMed: 23732752]
- 113. Adamopoulos IE, Suzuki E, Chao CC, Gorman D, Adda S, Maverakis E, Zarbalis K, Geissler R, Asio A, Blumenschein WM, McClanahan T, De Waal MR, Gershwin ME, Bowman EP (2015) IL-17A gene transfer induces bone loss and epidermal hyperplasia associated with psoriatic

arthritis. Ann Rheum Dis 74(6):1284–1292. 10.1136/annrheumdis-2013-204782 [PubMed: 24567524]

- 114. Harper EG, Guo C, Rizzo H, Lillis JV, Kurtz SE, Skorcheva I, Purdy D, Fitch E, Iordanov M, Blauvelt A (2009) Th17 cytokines stimulate CCL20 expression in keratinocytes in vitro and in vivo: implications for psoriasis pathogenesis. J Invest Dermatol 129(9):2175–2183. 10.1038/ jid.2009.65 [PubMed: 19295614]
- 115. Chiricozzi A, Guttman-Yassky E, Suarez-Farinas M, Nograles KE, Tian S, Cardinale I, Chimenti S, Krueger JG (2011) Integrative responses to IL-17 and TNF-alpha in human keratinocytes account for key inflammatory pathogenic circuits in psoriasis. J Invest Dermatol 131(3):677–687. 10.1038/jid.2010.340 [PubMed: 21085185]
- 116. Tomalin LE, Russell CB, Garcet S, Ewald DA, Klekotka P, Nirula A, Norsgaard H, Suarez-Farinas M, Krueger JG (2020) Short-term transcriptional response to IL-17 receptor-A antagonism in the treatment of psoriasis. J Allergy Clin Immunol 145(3):922–932. 10.1016/ j.jaci.2019.10.041 [PubMed: 31883845]
- 117. McGonagle DG, McInnes IB, Kirkham BW, Sherlock J, Moots R (2019) The role of IL-17A in axial spondyloarthritis and psoriatic arthritis: recent advances and controversies. Ann Rheum Dis 78(9):1167–1178. 10.1136/annrheumdis-2019-215356 [PubMed: 31278139]
- 118. Krueger GG, Langley RG, Leonardi C, Yeilding N, Guzzo C, Wang Y, Dooley LT, Lebwohl M, Group CPS (2007) A human interleukin-12/23 monoclonal antibody for the treatment of psoriasis. N Engl J Med 356(6):580–592. 10.1056/NEJMoa062382 [PubMed: 17287478]
- 119. Coates LC, Kavanaugh A, Mease PJ, Soriano ER, Laura Acosta-Felquer M, Armstrong AW, Bautista-Molano W, Boehncke WH, Campbell W, Cauli A, Espinoza LR, FitzGerald O, Gladman DD, Gottlieb A, Helliwell PS, Husni ME, Love TJ, Lubrano E, McHugh N, Nash P, Ogdie A, Orbai AM, Parkinson A, O'Sullivan D, Rosen CF, Schwartzman S, Siegel EL, Toloza S, Tuong W, Ritchlin CT (2016) Group for research and assessment of psoriasis and psoriatic arthritis 2015 treatment recommendations for psoriatic arthritis. Arthritis Rheumatol 68(5):1060–1071. 10.1002/art.39573 [PubMed: 26749174]
- 120. Markham A (2017) Guselkumab: first global approval. Drugs 77(13):1487–1492. 10.1007/ s40265-017-0800-7 [PubMed: 28819723]
- 121. Group U-S, Group U-S, Group U-S, Gordon KB, Blauvelt A, Papp KA, Langley RG, Luger T, Ohtsuki M, Reich K, Amato D, Ball SG, Braun DK, Cameron GS, Erickson J, Konrad RJ, Muram TM, Nickoloff BJ, Osuntokun OO, Secrest RJ, Zhao F, Mallbris L, Leonardi CL (2016) Phase 3 trials of ixekizumab in moderate-to-severe plaque psoriasis. N Engl J Med 375(4):345– 356. 10.1056/NEJMoa1512711 [PubMed: 27299809]
- 122. Lebwohl M, Strober B, Menter A, Gordon K, Weglowska J, Puig L, Papp K, Spelman L, Toth D, Kerdel F, Armstrong AW, Stingl G, Kimball AB, Bachelez H, Wu JJ, Crowley J, Langley RG, Blicharski T, Paul C, Lacour JP, Tyring S, Kircik L, Chimenti S, Callis Duffin K, Bagel J, Koo J, Aras G, Li J, Song W, Milmont CE, Shi Y, Erondu N, Klekotka P, Kotzin B, Nirula A (2015) Phase 3 studies comparing brodalumab with ustekinumab in psoriasis. N Engl J Med 373(14):1318–1328. 10.1056/NEJMoa1503824 [PubMed: 26422722]
- 123. Mease PJ, Genovese MC, Greenwald MW, Ritchlin CT, Beaulieu AD, Deodhar A, Newmark R, Feng J, Erondu N, Nirula A (2014) Brodalumab, an anti-IL17RA monoclonal antibody, in psoriatic arthritis. N Engl J Med 370(24):2295–2306. 10.1056/NEJMoa1315231 [PubMed: 24918373]
- 124. Armstrong AW, Read C (2020) Pathophysiology, clinical presentation, and treatment of psoriasis: a review. JAMA 323(19):1945–1960. 10.1001/jama.2020.4006 [PubMed: 32427307]
- 125. Papp KA, Blauvelt A, Bukhalo M, Gooderham M, Krueger JG, Lacour JP, Menter A, Philipp S, Sofen H, Tyring S, Berner BR, Visvanathan S, Pamulapati C, Bennett N, Flack M, Scholl P, Padula SJ (2017) Risankizumab versus ustekinumab for moderate-to-severe plaque psoriasis. N Engl J Med 376(16):1551–1560. 10.1056/NEJMoa1607017 [PubMed: 28423301]
- 126. Li X, Andersen KM, Chang HY, Curtis JR, Alexander GC (2020) Comparative risk of serious infections among real-world users of biologics for psoriasis or psoriatic arthritis. Ann Rheum Dis 79(2):285–291. 10.1136/annrheumdis-2019-216102 [PubMed: 31672774]
- 127. Gossec L, Baraliakos X, Kerschbaumer A, de Wit M, McInnes I, Dougados M, Primdahl J, McGonagle DG, Aletaha D, Balanescu A, Balint PV, Bertheussen H, Boehncke WH, Burmester

GR, Canete JD, Damjanov NS, Kragstrup TW, Kvien TK, Landewe RBM, Lories RJU, Marzo-Ortega H, Poddubnyy D, Rodrigues Manica SA, Schett G, Veale DJ, Van den Bosch FE, van der Heijde D, Smolen JS (2020) EULAR recommendations for the management of psoriatic arthritis with pharmacological therapies: 2019 update. Ann Rheum Dis 79(6):700–712. 10.1136/ annrheumdis-2020-217159 [PubMed: 32434812]

- 128. Schwartz DM, Kanno Y, Villarino A, Ward M, Gadina M, O'Shea JJ (2017b) JAK inhibition as a therapeutic strategy for immune and inflammatory diseases. Nat Rev Drug Discov 17(1):78. 10.1038/nrd.2017.267
- 129. Sabat R, Ouyang W, Wolk K (2014) Therapeutic opportunities of the IL-22-IL-22R1 system. Nat Rev Drug Discov 13(1):21–38. 10.1038/nrd4176 [PubMed: 24378801]
- 130. Kvist-Hansen A, Hansen PR, Skov L (2020) Systemic treatment of psoriasis with JAK inhibitors: A review. Dermatol Ther (Hei-delb) 10(1):29–42. 10.1007/s13555-019-00347-w
- 131. Colbert RA, Ward MM (2017) JAK inhibitors taking on psoriatic arthritis. N Engl J Med 377(16):1582–1584. 10.1056/NEJMe1709907 [PubMed: 29045217]
- 132. Gladman D, Rigby W, Azevedo VF, Behrens F, Blanco R, Kaszuba A, Kudlacz E, Wang C, Menon S, Hendrikx T, Kanik KS (2017) Tofacitinib for psoriatic arthritis in patients with an inadequate response to TNF inhibitors. N Engl J Med 377(16):1525–1536. 10.1056/ NEJMoa1615977 [PubMed: 29045207]
- 133. Paik J, Deeks ED (2019) Tofacitinib: a review in psoriatic arthritis. Drugs 79(6):655–663. 10.1007/s40265-019-01091-3 [PubMed: 30895473]
- 134. O'Shea JJ, Gadina M (2019) Selective Janus kinase inhibitors come of age. Nat Rev Rheumatol 15(2):74–75. 10.1038/s41584-018-0155-9 [PubMed: 30622297]
- 135. Papp K, Gordon K, Thaci D, Morita A, Gooderham M, Foley P, Girgis IG, Kundu S, Banerjee S (2018) Phase 2 trial of selective tyrosine kinase 2 inhibition in psoriasis. N Engl J Med 379(14):1313–1321. 10.1056/NEJMoa1806382 [PubMed: 30205746]
- 136. Schett G, Elewaut D, McInnes IB, Dayer JM, Neurath MF (2013) How cytokine networks fuel inflammation: toward a cytokine-based disease taxonomy. Nat Med 19(7):822–824. 10.1038/ nm.3260 [PubMed: 23836224]
- 137. Zhao Q (2020) Bispecific antibodies for autoimmune and inflammatory diseases: clinical progress to date. BioDrugs 34(2):111–119. 10.1007/s40259-019-00400-2 [PubMed: 31916225]
- 138. Chabaud M, Durand JM, Buchs N, Fossiez F, Page G, Frappart L, Miossec P (1999) Human interleukin-17: A T cell-derived proinflammatory cytokine produced by the rheumatoid synovium. Arthritis Rheum 42(5):963–970. 10.1002/1529-0131(199905)42:5<963::AID-ANR15>3.0.CO;2-E [PubMed: 10323452]
- 139. Smolen JS (2020) Insights into the treatment of rheumatoid arthritis: a paradigm in medicine. J Autoimmun 110:102425. 10.1016/j.jaut.2020.102425 [PubMed: 32143989]
- 140. Taams LS (2020) Interleukin-17 in rheumatoid arthritis: Trials and tribulations. J Exp Med 217 (3). doi:10.1084/jem.20192048
- 141. Du J, Wang X, Tan G, Liang Z, Zhang Z, Yu H (2020) The association between genetic polymorphisms of interleukin 23 receptor gene and the risk of rheumatoid arthritis: An updated meta-analysis. Clin Immunol 210:108250. 10.1016/j.clim.2019.108250 [PubMed: 31430553]
- 142. Manolova I, Ivanova M, Vasilev G, Stoilov R, Miteva L, Stanilova S (2020) Impact of IL12B polymorphisms on genetic susceptibility and IL-12p40 and IL-23 serum levels in rheumatoid arthritis. Immunol Invest 49(1–2):1–14. 10.1080/08820139.2019.1622561 [PubMed: 31161840]
- 143. Mo WX, Yin SS, Chen H, Zhou C, Zhou JX, Zhao LD, Fei YY, Yang HX, Guo JB, Mao YJ, Huang LF, Zheng WJ, Zhang W, Zhang JM, He W, Zhang X (2017) Chemotaxis of Vdelta2 T cells to the joints contributes to the pathogenesis of rheumatoid arthritis. Ann Rheum Dis 76(12):2075–2084. 10.1136/annrheumdis-2016-211069 [PubMed: 28866647]
- 144. Pfeifle R, Rothe T, Ipseiz N, Scherer HU, Culemann S, Harre U, Ackermann JA, Seefried M, Kleyer A, Uderhardt S, Haugg B, Hueber AJ, Daum P, Heidkamp GF, Ge C, Bohm S, Lux A, Schuh W, Magorivska I, Nandakumar KS, Lonnblom E, Becker C, Dudziak D, Wuhrer M, Rombouts Y, Koeleman CA, Toes R, Winkler TH, Holmdahl R, Herrmann M, Bluml S, Nimmerjahn F, Schett G, Kronke G (2017) Regulation of autoantibody activity by the IL-23-

TH17 axis determines the onset of autoimmune disease. Nat Immunol 18(1):104–113. 10.1038/ ni.3579 [PubMed: 27820809]

- 145. Fragoulis GE, Siebert S, McInnes IB (2016) Therapeutic targeting of IL-17 and IL-23 cytokines in immune-mediated diseases. Annu Rev Med 67:337–353. 10.1146/ annurevmed-051914-021944 [PubMed: 26565676]
- 146. Mankia K, Di Matteo A, Emery P (2020) Prevention and cure: The major unmet needs in the management of rheumatoid arthritis. J Autoimmun 110:102399. 10.1016/j.jaut.2019.102399 [PubMed: 31899021]
- 147. Pavelka K, Chon Y, Newmark R, Lin SL, Baumgartner S, Erondu N (2015) A study to evaluate the safety, tolerability, and efficacy of brodalumab in subjects with rheumatoid arthritis and an inadequate response to methotrexate. J Rheumatol 42(6):912–919. 10.3899/jrheum.141271 [PubMed: 25877498]
- 148. van Baarsen LG, Lebre MC, van der Coelen D, Aarrass S, Tang MW, Ramwadhdoebe TH, Gerlag DM, Tak PP (2014) Heterogeneous expression pattern of interleukin 17A (IL-17A), IL-17F and their receptors in synovium of rheumatoid arthritis, psoriatic arthritis and osteoarthritis: Possible explanation for nonresponse to anti-IL-17 therapy? Arthritis Res Ther 16(4):426. 10.1186/ s13075-014-0426-z [PubMed: 25146432]
- 149. Pastor-Fernandez G, Mariblanca IR, Navarro MN (2020) Decoding IL-23 Signaling Cascade for New Therapeutic Opportunities. Cells 9 (9). doi:10.3390/cells9092044
- 150. Long D, Chen Y, Wu H, Zhao M, Lu Q (2019) Clinical significance and immunobiology of IL-21 in autoimmunity. J Autoimmun 99:1–14. 10.1016/j.jaut.2019.01.013 [PubMed: 30773373]
- 151. Burns LA, Maroof A, Marshall D, Steel KJA, Lalnunhlimi S, Cole S, Catrina A, Kirkham B, Taams LS (2020) Presence, function, and regulation of IL-17F-expressing human CD4(+) T cells. Eur J Immunol 50(4):568–580. 10.1002/eji.201948138 [PubMed: 31850514]
- 152. Yamamoto K, Takeuchi T, Yamanaka H, Ishiguro N, Tanaka Y, Eguchi K, Watanabe A, Origasa H, Shoji T, Sakamaki Y, van der Heijde D, Miyasaka N, Koike T (2014) Efficacy and safety of certolizumab pegol plus methotrexate in Japanese rheumatoid arthritis patients with an inadequate response to methotrexate: The J-RAPID randomized, placebo-controlled trial. Mod Rheumatol 24(5):715–724. 10.3109/14397595.2013.864224 [PubMed: 24313916]
- 153. Yue C, You X, Zhao L, Wang H, Tang F, Zhang F, Zhang X, He W (2010) The effects of adalimumab and methotrexate treatment on peripheral Th17 cells and IL-17/IL-6 secretion in rheumatoid arthritis patients. Rheumatol Int 30(12):1553–1557. 10.1007/s00296-009-1179-x [PubMed: 19847432]
- 154. Genovese MC, Weinblatt ME, Aelion JA, Mansikka HT, Peloso PM, Chen K, Li Y, Othman AA, Khatri A, Khan NS, Padley RJ (2018) ABT-122, a bispecific dual variable domain immunoglobulin targeting tumor necrosis factor and interleukin-17A, in patients with rheumatoid arthritis with an inadequate response to methotrexate: a randomized. Double-Blind Study Arthritis Rheumatol 70(11):1710–1720. 10.1002/art.40580 [PubMed: 29855172]
- 155. Winthrop KL (2017) The emerging safety profile of JAK inhibitors in rheumatic disease. Nat Rev Rheumatol 13(4):234–243. 10.1038/nrrheum.2017.23 [PubMed: 28250461]
- 156. Kerschbaumer A, Sepriano A, Smolen JS, van der Heijde D, Dougados M, van Vollenhoven R, McInnes IB, Bijlsma JWJ, Burmester GR, de Wit M, Falzon L, Landewe R (2020) Efficacy of pharmacological treatment in rheumatoid arthritis: a systematic literature research informing the 2019 update of the EULAR recommendations for management of rheumatoid arthritis. Ann Rheum Dis 79(6):744–759. 10.1136/annrheumdis-2019-216656 [PubMed: 32033937]
- 157. Harigai M (2019) Growing evidence of the safety of JAK inhibitors in patients with rheumatoid arthritis. Rheumatology (Oxford) 58(Suppl 1):i34–i42. 10.1093/rheumatology/key287 [PubMed: 30806708]
- 158. Dougados M (2020) Treat to target in axial spondyloarthritis: from its concept to its implementation. J Autoimmun 110:102398. 10.1016/j.jaut.2019.102398 [PubMed: 31926832]
- 159. Gravallese EM, Schett G (2018) Effects of the IL-23-IL-17 pathway on bone in spondyloarthritis. Nat Rev Rheumatol 14(11):631–640. 10.1038/s41584-018-0091-8 [PubMed: 30266977]
- 160. DeLay ML, Turner MJ, Klenk EI, Smith JA, Sowders DP, Colbert RA (2009) HLA-B27 misfolding and the unfolded protein response augment interleukin-23 production and are

associated with Th17 activation in transgenic rats. Arthritis Rheum 60(9):2633–2643. 10.1002/ art.24763 [PubMed: 19714651]

- 161. Wendling D, Prati C, Chouk M, Verhoeven F (2020) Effects of anti-IL-23 and anti-IL-17: the hidden side of spondyloarthritis polymorphism? Joint Bone Spine 87(1):5–7. 10.1016/ j.jbspin.2019.06.012 [PubMed: 31324528]
- 162. Chen S, Blijdorp I, van Mens L, Bowcutt R, Latuhihin T, van de Sande M, Shaw S, Yeremenko N, Baeten D (2020) IL-17A and IL-17F expression and functional responses in rheumatoid arthritis and peripheral spondyloarthritis. J Rheumatol. 10.3899/jrheum.190571
- 163. Erdes S, Nasonov E, Kunder E, Pristrom A, Soroka N, Shesternya P, Dubinina T, Smakotina S, Raskina T, Krechikova D, Povarova T, Plaksina T, Gordeev I, Mazurov V, Reshetko O, Zonova E, Eremeeva A, Chernyaeva E, Makulova T, Ivanov R (2020) Primary efficacy of netakimab, a novel interleukin-17 inhibitor, in the treatment of active ankylosing spondylitis in adults. Clin Exp Rheumatol 38(1):27–34.
- 164. van der Heijde D, Gensler LS, Deodhar A, Baraliakos X, Poddubnyy D, Kivitz A, Farmer MK, Baeten D, Goldammer N, Coarse J, Oortgiesen M, Dougados M (2020) Dual neutralisation of interleukin-17A and interleukin-17F with bimekizumab in patients with active ankylosing spondylitis: results from a 48-week phase IIb, randomised, double-blind, placebo-controlled, dose-ranging study. Ann Rheum Dis 79(5):595–604. 10.1136/annrheumdis-2020-216980 [PubMed: 32253184]
- 165. Mease P (2019) Ustekinumab fails to show efficacy in a phase III axial spondyloarthritis program: The importance of negative results. Arthritis Rheumatol 71(2):179–181. 10.1002/art.40759 [PubMed: 30600936]
- 166. Baeten DL, Kuchroo VK (2013) How cytokine networks fuel inflammation: interleukin-17 and a tale of two autoimmune diseases. Nat Med 19(7):824–825. 10.1038/nm.3268 [PubMed: 23836225]
- 167. Buonocore S, Ahern PP, Uhlig HH, Ivanov II, Littman DR, Maloy KJ, Powrie F (2010) Innate lymphoid cells drive interleukin-23-dependent innate intestinal pathology. Nature 464(7293):1371–1375. 10.1038/nature08949 [PubMed: 20393462]
- 168. Aden K, Rehman A, Falk-Paulsen M, Secher T, Kuiper J, Tran F, Pfeuffer S, Sheibani-Tezerji R, Breuer A, Luzius A, Jentzsch M, Hasler R, Billmann-Born S, Will O, Lipinski S, Bharti R, Adolph T, Iovanna JL, Kempster SL, Blumberg RS, Schreiber S, Becher B, Chamaillard M, Kaser A, Rosenstiel P (2016) Epithelial IL-23R signaling licenses protective IL-22 responses in intestinal inflammation. Cell Rep 16(8):2208–2218. 10.1016/j.celrep.2016.07.054 [PubMed: 27524624]
- 169. van der Heijde D, Deodhar A, Wei JC, Drescher E, Fleishaker D, Hendrikx T, Li D, Menon S, Kanik KS (2017) Tofacitinib in patients with ankylosing spondylitis: a phase II, 16-week, randomised, placebo-controlled, dose-ranging study. Ann Rheum Dis 76(8):1340–1347. 10.1136/ annrheumdis-2016-210322 [PubMed: 28130206]
- 170. van der Heijde D, Baraliakos X, Gensler LS, Maksymowych WP, Tseluyko V, Nadashkevich O, Abi-Saab W, Tasset C, Meuleners L, Besuyen R, Hendrikx T, Mozaffarian N, Liu K, Greer JM, Deodhar A, Landewe R (2018) Efficacy and safety of filgotinib, a selective Janus kinase 1 inhibitor, in patients with active ankylosing spondylitis (TORTUGA): Results from a randomised, placebo-controlled, phase 2 trial. Lancet 392(10162):2378–2387. 10.1016/ S0140-6736(18)32463-2 [PubMed: 30360970]
- 171. Aringer M (2020) Inflammatory markers in systemic lupus erythematosus. J Autoimmun 110:102374. 10.1016/j.jaut.2019.102374 [PubMed: 31812331]
- 172. Frangou E, Georgakis S, Bertsias G (2020) Update on the cellular and molecular aspects of lupus nephritis. Clin Immunol 216:108445. 10.1016/j.clim.2020.108445 [PubMed: 32344016]
- 173. Martin JC, Baeten DL, Josien R (2014) Emerging role of IL-17 and Th17 cells in systemic lupus erythematosus. Clin Immunol 154(1):1–12. 10.1016/j.clim.2014.05.004 [PubMed: 24858580]
- 174. Yang HX, Zhang W, Zhao LD, Li Y, Zhang FC, Tang FL, He W, Zhang X (2009) Are CD4+CD25-Foxp3+ cells in untreated new-onset lupus patients regulatory T cells? Arthritis Res Ther 11(5):R153. 10.1186/ar2829 [PubMed: 19821980]
- 175. Crispin JC, Oukka M, Bayliss G, Cohen RA, Van Beek CA, Stillman IE, Kyttaris VC, Juang YT, Tsokos GC (2008) Expanded double negative T cells in patients with systemic lupus

erythematosus produce IL-17 and infiltrate the kidneys. J Immunol 181(12):8761–8766. 10.4049/ jimmunol.181.12.8761 [PubMed: 19050297]

- 176. Koga T, Ichinose K, Tsokos GC (2017) T cells and IL-17 in lupus nephritis. Clin Immunol 185:95–99. 10.1016/j.clim.2016.04.010 [PubMed: 27109641]
- 177. Dai H, He F, Tsokos GC, Kyttaris VC(2017) IL-23 limits the production of IL-2 and promotes autoimmunity in lupus. J Immunol 199(3):903–910. 10.4049/jimmunol.1700418 [PubMed: 28646040]
- 178. Lopez P, Rodriguez-Carrio J, Martinez-Zapico A, Perez-Alvarez AI, Benavente L, Caminal-Montero L, Suarez A (2020) IgM anti-phosphorylcholine antibodies associate with senescent and IL-17+ T cells in SLE patients with a pro-inflammatory lipid profile. Rheumatology (Oxford) 59(2):407–417. 10.1093/rheumatology/kez264 [PubMed: 31302689]
- 179. Vukelic M, Laloo A, Kyttaris VC (2020) Interleukin 23 is elevated in the serum of patients with SLE. Lupus:961203320952841. 10.1177/0961203320952841
- 180. Li M, Yang C, Wang Y, Song W, Jia L, Peng X, Zhao R (2020) The Expression of P2X7 receptor on Th1, Th17, and regulatory T cells in patients with systemic lupus erythematosus or rheumatoid arthritis and its correlations with active disease. J Immunol 205(7):1752–1762. 10.4049/jimmunol.2000222 [PubMed: 32868411]
- 181. Kyttaris VC, Zhang Z, Kuchroo VK, Oukka M, Tsokos GC (2010) Cutting edge: IL-23 receptor deficiency prevents the development of lupus nephritis in C57BL/6-lpr/lpr mice. J Immunol 184(9):4605–4609. 10.4049/jimmunol.0903595 [PubMed: 20308633]
- 182. Sharabi A, Tsokos GC (2020) T cell metabolism: New insights in systemic lupus erythematosus pathogenesis and therapy. Nat Rev Rheumatol 16(2):100–112. 10.1038/s41584-019-0356-x [PubMed: 31949287]
- 183. Li H, Adamopoulos IE, Moulton VR, Stillman IE, Herbert Z, Moon JJ, Sharabi A, Krishfield S, Tsokos MG, Tsokos GC (2020) Systemic lupus erythematosus favors the generation of IL-17 producing double negative T cells. Nat Commun 11(1):2859. 10.1038/s41467-020-16636-4 [PubMed: 32503973]
- 184. Hong H, Gao M, Wu Q, Yang P, Liu S, Li H, Burrows PD, Cua D, Chen JY, Hsu HC, Mountz JD (2020) IL-23 promotes a coordinated B cell germinal center program for class-switch recombination to IgG2b in BXD2 mice. J Immunol 205(2):346–358. 10.4049/jimmunol.2000280 [PubMed: 32554431]
- 185. Schmidt T, Paust HJ, Krebs CF, Turner JE, Kaffke A, Bennstein SB, Koyro T, Peters A, Velden J, Hunemorder S, Haag F, Steinmetz OM, Mittrucker HW, Stahl RA, Panzer U (2015) Function of the Th17/interleukin-17A immune response in murine lupus nephritis. Arthritis Rheumatol 67(2):475–487. 10.1002/art.38955 [PubMed: 25385550]
- 186. Perry D, Sang A, Yin Y, Zheng YY, Morel L (2011) Murine models of systemic lupus erythematosus. J Biomed Biotechnol 2011:271694. 10.1155/2011/271694 [PubMed: 21403825]
- 187. van Vollenhoven RF, Hahn BH, Tsokos GC, Wagner CL, Lipsky P, Touma Z, Werth VP, Gordon RM, Zhou B, Hsu B, Chevrier M, Triebel M, Jordan JL, Rose S (2018) Efficacy and safety of ustekinumab, an IL-12 and IL-23 inhibitor, in patients with active systemic lupus erythematosus: results of a multi-centre, double-blind, phase 2, randomised, controlled study. Lancet 392(10155):1330–1339. 10.1016/S0140-6736(18)32167-6 [PubMed: 30249507]
- 188. Wallace DJ, Furie RA, Tanaka Y, Kalunian KC, Mosca M, Petri MA, Dorner T, Cardiel MH, Bruce IN, Gomez E, Carmack T, DeLozier AM, Janes JM, Linnik MD, de Bono S, Silk ME, Hoff-man RW (2018) Baricitinib for systemic lupus erythematosus: A double-blind, randomised, placebo-controlled, phase 2 trial. Lancet 392(10143):222–231. 10.1016/S0140-6736(18)31363-1 [PubMed: 30043749]
- 189. Alunno A, Padjen I, Fanouriakis A, Boumpas DT (2019) Pathogenic and Therapeutic Relevance of JAK/STAT Signaling in Systemic Lupus Erythematosus: Integration of Distinct Inflammatory Pathways and the Prospect of Their Inhibition with an Oral Agent. Cells 8 (8). 10.3390/ cells8080898
- 190. Murphy G, Isenberg DA (2019) New therapies for systemic lupus erythematosus past imperfect, future tense. Nat Rev Rheumatol 15(7):403–412. 10.1038/s41584-019-0235-5 [PubMed: 31165780]

- 191. Wenzel J (2019) Cutaneous lupus erythematosus: New insights into pathogenesis and therapeutic strategies. Nat Rev Rheumatol 15(9):519–532. 10.1038/s41584-019-0272-0 [PubMed: 31399711]
- 192. Schwartz N, Stock AD, Putterman C (2019) Neuropsychiatric lupus: new mechanistic insights and future treatment directions. Nat Rev Rheumatol 15(3):137–152. 10.1038/s41584-018-0156-8 [PubMed: 30659245]
- 193. Li Q, Wu H, Liao W, Zhao M, Chan V, Li L, Zheng M, Chen G, Zhang J, Lau CS, Lu Q (2018) A comprehensive review of immune-mediated dermatopathology in systemic lupus erythematosus. J Autoimmun 93:1–15. 10.1016/j.jaut.2018.07.007 [PubMed: 30017673]
- 194. Sedykh SE, Prinz VV, Buneva VN, Nevinsky GA (2018) Bispecific antibodies: Design, therapy, perspectives. Drug Des Devel Ther 12:195–208. 10.2147/DDDT.S151282
- 195. Leonard WJ, Lin JX, O'Shea JJ (2019) The gammac family of cytokines: Basic biology to therapeutic ramifications. Immunity 50(4):832–850. 10.1016/j.immuni.2019.03.028 [PubMed: 30995502]
- 196. Schwartz DM, Bonelli M, Gadina M, O'Shea JJ (2016) Type I/II cytokines, JAKs, and new strategies for treating autoimmune diseases. Nat Rev Rheumatol 12(1):25–36. 10.1038/ nrrheum.2015.167 [PubMed: 26633291]
- 197. Stark GR, Cheon H, Wang Y (2018) Responses to Cytokines and Interferons that Depend upon JAKs and STATs. Cold Spring Harb Perspect Biol 10 (1). 10.1101/cshperspect.a028555

Fig. 1.

Biologics targeting IL-23/IL-17 axis in rheumatic diseases. Specific blocking antibodies and small molecules targeting IL-23/IL-17 axis which are currently used in clinical practice or still under development are depicted (MQ: macrophages; DC: dendritic cells)

Fig. 2.

IL-23/IL-17 axis contributes to autoantibody-driven diseases. B cells migrate between the dark and light zones of the germinal centers (GCs), a process mediated by CXCL12/CXCR4 and CXCL13/CXCR5, to undergo somatic hypermutation, clonal expansion, and affinitybased selection. Exposure to IL-17 reduces their migration which results in more rounds of somatic hypermutation and the production of IgG with increased affinity. Plasma cells emigrating from GCs produce autoantibodies. IL-23-activated T_H17 cells suppress St6gal1 expression in antibody-producing cells via IL-21 and IL-22, which results in changes in the glycosylation profile as well as increased inflammatory activity of IgG (cDC: classical dendritic cells; FDC: follicular dendritic cells)

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Table 1

Biologics and small molecule drugs targeting the IL-23/IL-17 axis approved by FDA to treat patients with rheumatic diseases Biologics and small molecule drugs targeting the IL-23/IL-17 axis approved by FDA to treat patients with rheumatic diseases

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Biologics and small molecule drugs in clinical trials targeting the IL-23/IL-17 axis in SLE patients Biologics and small molecule drugs in clinical trials targeting the IL-23/IL-17 axis in SLE patients

