

Systemic inflammatory response to non-surgical treatment in hypertensive patients with periodontal infection

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Abstract

Hypertension is associated with chronic inflammation in the tissues and organs that are involved in the regulation of arterial pressure, such as kidneys and blood vessels. Periodontal disease affects systemic inflammatory markers, leading to endothelial dysfunction, atherosclerotic plaque instability, dyslipidaemia, and insulin resistance. These conditions can also cause an increase in the blood pressure. Nonsurgical periodontal therapies, such as scaling and root planning, can affect systemic markers of inflammation. We evaluated the effect of scaling and root planning on serum levels of inflammation biomarkers in hypertensive patients. The sample consisted of 19 hypertensive patients with Periodontitis. The patients underwent laboratory tests that included glycaemia, cholesterol, triglycerides and blood count. Blood pressure was measured before periodontal therapy, and the second blood pressure recording was obtained at the re-evaluation appointment. Quantification of peripheral blood cytokines was performed using the Milliplex Inflammation Human Cytokine kit (Interleukin 1- β , Interleukin-4, Interleukin-6, Interleukin-8, Interleukin-10, Interleukin-12 P70, Interleukin-17A, vascular endothelial growth factor and tumor necrosis factor-alpha). All cytokine levels decreased from the initial examination to reassessment. Cytokines that reflected a statistically significant difference included Interleukin-1 β and endothelial vascular growth factor ($P = .04$ and $P = .004$). Hypertensive patients with periodontitis undergoing non-surgical periodontal treatment exhibited a decrease in proinflammatory cytokine levels. Non-surgical periodontal treatment decreases the levels of systemic proinflammatory cytokines in controlled hypertensive patients.

Abbreviations: BOP = bleeding on probing, CAL = clinical attachment level, cHDL = high-density lipoprotein, cLDL = low-density lipoprotein, CRP = C-reactive protein, EDTA = ethylenediaminetetraacetic acid, HUSI = San Ignacio Hospital, IL = interleukin, RT = room temperature, TNF- α = tumor necrosis factor-alpha, VEGF = vascular endothelial growth factor.

Keywords: cytokines, hypertension, mediators of inflammation, periodontal debridement, periodontal disease, periodontal infection

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1. Introduction

Hypertension is associated with chronic inflammation in the tissues and organs that are involved in the regulation of arterial pressure, such as kidneys and blood vessels. Vascular inflammation may contribute to the alteration of functions, such as endothelial resistance and stiffness.^[1–4] Periodontal disease is a multifactor pathology, and its main aetiological factor is the accumulation of bacterial biofilm. This disease affects gingival tissue and subsequently destroys the insertion and support tissues of the teeth.^[5] Periodontal disease severity affects systemic inflammatory markers, such as C-reactive protein (CRP), fibrinogen, tumor necrosis factor-alpha (TNF- α), and Interleukin (IL)-6, leading to endothelial dysfunction, atherosclerotic plaque instability, dyslipidaemia, and insulin resistance. These conditions can also cause an increase in the blood pressure and mortality risk in hypertension patients.^[6–8] The elimination of the infectious and inflammatory load of the periodontal disease may be associated with improved endothelial function and could be accompanied by a decrease in inflammatory markers. Furthermore, periodontal therapy increases nitric oxide production, also leading to improved endothelial function.^[9–11]

Considering this background, the following question arises: What is the effect of non-surgical periodontal therapy (root

scaling and debridement) on serum levels of IL-1 β , IL-4, IL-6, IL-8, IL-10, IL-12 P70, IL-17A, vascular endothelial growth factor (VEGF), and TNF- α in patients with generalized periodontitis stage II, III, and IV and arterial hypertension? Thus, the aim of this study is to analyse whether non-surgical periodontal treatment will improve the systemic inflammatory response. These findings will be important for the interdisciplinary management of blood pressure control and the systemic inflammatory response in hypertensive patients.

2. Methods

2.1. Study design and setting

The present study was observational and evaluates the changes in systemic inflammatory biomarkers before and after non-surgical periodontal therapy. Nineteen patients over 30 years with periodontitis stage II, III, and IV, grade A and B and controlled arterial hypertension were included in this study. Patients diagnosed with hypertension and undergoing pharmacological treatment with a minimum of 6 teeth present in the oral cavity were classified as periodontitis stage II, III, and IV, grade A and B. Patients with diabetes, patients who smoke, patients undergoing antibiotic treatment in the last 3 months and patients who received periodontal treatment in the last 6 months were excluded. Patients with immunological diseases that may alter blood pressure, such as rheumatoid arthritis and systemic lupus erythematosus, were excluded from the study.

2.2. Ethical approval

The Investigation and Ethics Committee from the Odontology Faculty of the Pontificia Universidad Javeriana (CIEFOPUJ)-Bogotá, Colombia approved this study (Reference: 007/2015). The study was explained to the patients, who signed informed consent to participate in the study. We confirm that this study was conducted in accordance with the Helsinki Declaration of 1964, and its subsequent modifications. We confirm that all subjects gave their informed consent to participate in the study.

2.3. Data collection

Periodontal examinations were performed by 2 calibrated evaluators who used a graduated manual North Carolina Probe. Biofilm control was measured using the O'Leary index. The periodontal examination was performed around 6 points in each tooth present in the mouth. The periodontal sulcus or pocket was measured from the gingival margin to the bottom of the pocket or sulcus. It was considered a pathological periodontal pocket when the measurement was 4 mm or greater. Likewise, the gingival margin measure was taken from the cement-enamel junction to the gingival margin. Using this probing depth and gingival margin data, the clinical attachment level was calculated and a periodontal diagnosis was made based on the Workshop of 2017 and Armitage 1999 classification.^[12,13]

After the periodontal examination, laboratory exams were performed in San Ignacio Hospital (HUSI) during the first hour in the morning, and patients were fasting. Exams assessed blood count, glycaemia, triglycerides, High-density lipoprotein cholesterol (cHDL), Low-density lipoprotein cholesterol (cLDL) and serum levels of the inflammatory markers IL-1 β , IL-4, IL-6, IL-8, IL-10, IL-12 P70, IL-17A, VEGF, and TNF- α . After patients had

breakfast and rested for 1 hour, blood pressure was measured by a professional from the Cardiology Department of HUSI.

All patients received oral hygiene instruction and non-surgical periodontal treatment (scaling and root planning) with ultrasonic piezoelectric and Gracey currettes in 1 session. After an average of 4 to 5 weeks after the periodontal treatment, a reevaluation was performed in which the periodontal status of each patient was verified by taking the same reference values of the initial examination appointment. In the same session, blood count, glycaemia, cholesterol, triglycerides, cHDL, cLDL, and systemic inflammatory biomarkers were measured. Furthermore, blood pressure was also measured.

Three blood tubes were obtained as samples, including 2 dry tubes and 1 with an anticoagulant. The resulting sample from the anticoagulant tube with ethylenediaminetetraacetic acid (EDTA) was processed in the clinical laboratory of HUSI to assess blood count and glycaemic values. One of the dry tubes was used for total cholesterol, triglycerides, cHDL, and cLDL analysis. Gel to separate plasma and serum was placed in the other dry tube. The tube was centrifuged at 10,000 rpm for 10 minutes and frozen at -20°C in the HUSI until its use. These samples were used for the cytokines measurements, analysis at the end of the sample collection.

Serum cytokines were quantified using the Luminex system with the Milliplex Inflammation Human Cytokine kit. The test was performed on a plate where reagents and working standards were prepared following the manufacturer's instructions. The pearls were prepared for each cytokine to be evaluated and placed in a single container. Each vessel containing vortex pearls was stirred for 1 minute, and 60 μl of pearl reconstituting agent was placed for each cytokine. The standard, test buffer, and serum matrix were added to the background, standard and control wells, and the serum from the sample was added to the corresponding wells. The bottle with the premixed pearls was placed in the vortex, and 25 μl was added to each well. The bottle was sealed with adhesive paper and wrapped in aluminium foil. The sample was stirred at 700 rpm at 4°C for 16 hours. Then, 2 washes were performed with a magnetic plate, and detection antibodies were added and stirred for 1 hour at room temperature (RT). Then, streptavidin-phycoerythrin was added to each well, and the sample was sealed, covered with aluminium and stirred at RT for 30 minutes. Then, the washing procedure was repeated, and drive fluid was added. The microparticles (pearls) were resuspended in the stirrer for 5 minutes. All samples were assembled in duplicate. Finally, the plate was assessed using Magpix Luminex and analysed by Magpix Luminex 200 software. The equipment gave different values that we used to identify the total count of captured pearls, the average fluorescence intensity and the concentrations of the cytokines of interest. These values were determined according to the standard curve and expressed in pg/ml, and the values were adjusted to the dilution factor. The Luminex kit assay sensitivity has the following ranges in pg/ml of Minimum Detectable Concentration (MinDC) and Minimum Detectable Concentration +2SD, as described below: IFN- γ : 0.8 and 1.1; IL-10: 1.1 and 1.6; IL-12 P70: 0.6 and 1.0; IL-17A: 0.7 and 1.2; IL-1 β : 0.8 and 1.0; IL-4: 4.5 and 7.1; IL-6: 0 and 1.3; IL-8: 0.4 and 0.7; TNF- α : 0.7 and 1.1; VEGF: 26.3 and 47.9.

To establish blood pressure, measurements were obtained from both arms with an interval of at least 2 minutes. The patient had to meet the following requirements for adequate intake, sit with back supported and arms at heart level, blood pressure should be

Table 1**Clinical parameters pre-treatment and post-treatment.**

Clinical parameters	Pre-treatment		Post-treatment		P value
	Mean	SE	Mean	SE	
Teeth with periodontitis	12.89	1.13	8.84	1.11	.00*
Probing Depth (mm)	3.17	0.12	2.89	0.10	.00*
CAL (mm)	3.38	0.27	3.15	0.29	.01*
BOP (%)	58.43	4.38	39.75	3.20	.0001**
BIOFILM (%)	41.36	2.23	28.80	2.08	.0004**

BOP=bleeding on probing, CAL=clinical attachment level, SE=standard error.

* Paired *t* test $\leq .05$.

** Test of proportions ≤ 0.05 .

taken after 5 minutes of rest, the patient should not have smoked or consumed caffeine 30 minutes before the blood pressure reading, do not speak during the measurement and, support the discreetly strapped arm with the palm facing up, preferably on the right arm or the dominant arm. An electronic blood pressure monitor was used, the same instrument was used in all patients.

2.4. Statistic analysis

Demographic characteristics and the results of the periodontal evaluation and the systemic biomarkers of inflammation are reported as the means, medians, ranges, standard deviations and 95% confidence intervals. Comparisons between groups were performed using Student's *t* test or chi-square as appropriate. Here, $P < .05$ (2-tailed) was considered significant.

3. Results

The study included a total of 22 patients, of which 3 were excluded due to incomplete follow-up. Of the final sample, 52.6% were women (10 patients), and 47.3% were men (9 patients), with an average age of 57.6 years (SD 2.2)

Of the 19 patients included in the study, periodontal status was assessed according to the 2017 Workshop classification at the start of the study and at the time of re-evaluation. In total, 2 patients were diagnosed with Stage III Grade A Periodontitis, 10 with Stage III Grade Periodontitis B, 2 with Stage IV Grade A Periodontitis and 5 with Stage IV Grade B Periodontitis. Regarding the 1999 Armitage classification, at the beginning of treatment, 17 patients presented with generalized severe chronic periodontitis and 2 with generalized moderate chronic periodontitis. At the time of reevaluation, 4 presented with generalized moderate chronic periodontitis, 8 generalized severe

chronic periodontitis, 2 localized moderate chronic periodontitis and 5 presented localized severe chronic periodontitis.

A total of 464 teeth were evaluated, of which 265 had periodontal pockets at the initial evaluation. The pre-treatment range was 4 to 14 mm. At the month of the re-evaluation, the number of teeth for which 2 exodonces were performed in the hygienic phase was 462, and there were 175 teeth with periodontal pockets. The number of periodontal pockets in the re-evaluation ranged from 4 to 12 mm. When evaluating the measured clinical parameters at the initial evaluation the average number of teeth with periodontitis was 12.89, the average sulcus or pocket depth was 3.17 mm, the clinical attachment level was 3.38 mm, the bleeding on probing percentage was 58.43% and the plaque index was 41.36%, the following changes were observed at the time of the re-evaluation: the average number of teeth with periodontitis was 4.05, the average sulcus or pocket depth was 0.28 mm, the clinical attachment level was 0.23 mm, the bleeding on probing percentage was 18.67% and the plaque index was 12.55%. These findings indicate reductions in all the parameters evaluated (Table 1).

The average blood pressure was 135.15/86.1 mmHg at the beginning of the study and 133.26/83.15 mmHg at the end of treatment, and the difference was not statistically significant (Table 2). Clinical laboratory serum tests revealed an increase of 0.26 mg/dl in glycaemia at the time of reassessment, triglycerides, cHDL and cLDL were reduced by 10.6, 16.2, 38.6, and 8.3 mg/dl, respectively. Statistically significant *P* values for the level of triglycerides and cHDL (Table 3).

Patient cytokine levels decreased from the initial examination to re-evaluation. The following initial values were observed: 0.53 pg/ml IL-1 β , 2.98 pg/ml IL-4, 0.31 pg/ml IL-6, 9.17 pg/ml IL-8, 1.15 pg/ml IL-10, 1.82 pg/ml IL-12 P70, 3.8 pg/ml IL-17A, 15.84 pg/ml TNF- α , and 152.10 pg/ml VEGF. The following values were obtained after periodontal treatment: 0.30 pg/ml IL-1 β ,

Table 2**Blood pressure pre-treatment and post-treatment.**

Blood pressure (mmHg)	Pre-treatment		Post-treatment		Mean difference	CI 95%	P value*
	Mean	SE	Mean	SE			
Systolic	135.15	3.69	133.26	2.47	1.89	−5.89	.61
Diastolic	86.10	1.80	83.15	1.14	2.94	−0.45	.08

CI 95%=95% confidence interval, SE=standard error

* Paired *t* test $P \leq .05$.

Table 3**Laboratory tests pre-treatment and post-treatment.****Laboratory tests results (mg/dl)**

	Pre-treatment		Post-treatment		Mean difference	CI 95%	P value**
	Mean	SE	Mean	SE			
Glycemia	100.78	2.23	101.05	2.19	-0.26	-5.30	.91
Cholesterol	201.05	8.28	190.45	8.34	10.6	-0.51	.06
Triglycerides	176.38	18.08	160.14	17.62	16.24	0.33	.04
HDL*	83.71	10.53	45.05	2.70	38.65	17.27	.001
LDL†	124.23	6.88	115.89	4.87	8.33	-2.87	.13

CI 95%=95% confidence interval, SE=standard error.

* High-density lipoprotein

† Low-density lipoprotein.

** Paired *t* test $P \leq .05$.

1.23 pg/ml IL-4, 0.24 pg/ml IL-6, 7.79 pg/ml IL-8, 0.86 pg/ml IL-10, 0.94 pg/ml IL-12 P70, 3.19 pg/ml IL-17A, 14.29 pg/ml TNF- α , and 101.55 pg/ml VEGF. IL-1 β and VEGF ($P=.04$ and $P=.004$) exhibited statistically significant differences when evaluated with the Wilcoxon test (Table 4; Fig. 1).

A cut-off point for each cytokine was established based on reference values reported in the literature. Of the articles evaluated, there is no definitive cut-off point for cytokines since they can be detected by different techniques such as ELISPOT, Flow Cytometry, ELISA, and Luminex. This cut-off point is described in each study in a particular way, for example, it is obtained from the mean plus two (2) standard deviations, and a range is determined. Each study establishes it independently depending on the technique used, the selected groups, and the biological sample analyzed. On the other hand, each laboratory obtains its cut-off point or reference value. We obtained these values from the Mayo Clinic (Bogotá), a recognized clinic in Colombia.^[14-17] IL-1 β , IL-6, IL-8, and IL-10 initial and final treatment values were normal based on cut-off points. At the initial evaluation, only 2 patients presented normal TNF- α values, and 17 presented increased values. However, in the final evaluation, 3 patients presented normal values, and 16 values increased based on the cut-off point ($P=.018^*$). (Table 5).

4. Discussion

Scientific literature has described the relationship between hypertension and periodontal disease due to the inflammatory

response that underlies these 2 pathologies. Periodontal therapy could decrease the levels of inflammation biomarkers and positively impact the development of hypertension.

Beyond the periodontal diagnosis, this study found that when performing basic periodontal treatment, average periodontal clinical parameters, such as the level of clinical insertion (from 3.38 to 3.15), the rate of bleeding (from 58.43% to 39.75%) and the biofilm index (from 41.36% to 28.80%), improved with statistically significant differences for all the analysed clinical parameters. These findings were expected and are consistent with that reported by Aristizábal et al.^[18] Al Bush et al published the effects of surgical and non-surgical periodontal treatment on vascular markers in hypertensive patients. Both the surgical group and the non-surgical group presented statistically significant decreases in periodontal clinical parameters. These results are consistent with our study; however, in this investigation, non-surgical periodontal therapy was performed in all the patients in the study.^[19]

Regarding blood pressure before and after treatment, a slight decrease in systolic and diastolic blood pressure was evidenced from an average of 135.15/86.1 to an average of 133.26/83.15 mmHg, respectively. This finding is consistent with the study by Pietropaoli D. and colleagues in 2014, demonstrating that the mean systolic blood pressure was approximately 2.3 to 3 mmHg increased in the presence of periodontitis among hypertensive adults undergoing treatment ($P<.0001$). These results further indicate that good periodontal health is associated with a better profile of systolic blood pressure during antihypertensive

Table 4**Cytokines levels pre-treatment and post-treatment.****Cytokines levels (pg/ml)**

	Pre-treatment		Post-treatment		Mean difference	CI 95%	P value*
	Mean	SE	Mean	SE			
IL-1 β	0.53	0.14	0.30	0.07	0.22	-0.09	.04
IL-4	2.98	1.62	1.23	1.03	1.75	-0.78	.27
IL-6	0.31	0.18	0.24	0.13	0.07	-0.05	.30
IL-8	9.17	1.05	7.79	0.62	1.37	-0.74	.22
IL-10	1.15	0.43	0.86	0.26	0.29	-0.50	.42
IL-12 P70	1.82	1.08	0.94	0.48	0.87	-0.49	.36
IL-17A	3.8	1.08	3.19	0.69	0.60	-0.47	.74
TNF- α	15.84	1.06	14.29	1.05	1.54	-0.38	.18
VEGF	152.10	28.70	101.55	18.36	50.55	16.56	.004

CI 95%=95% confidence interval, IL=interleukin, SE=standard error, TNF- α =tumor necrosis factor-alpha, VEGF=vascular endothelial growth factor.

* Wilcoxon signed-rank test $P \leq .05$.

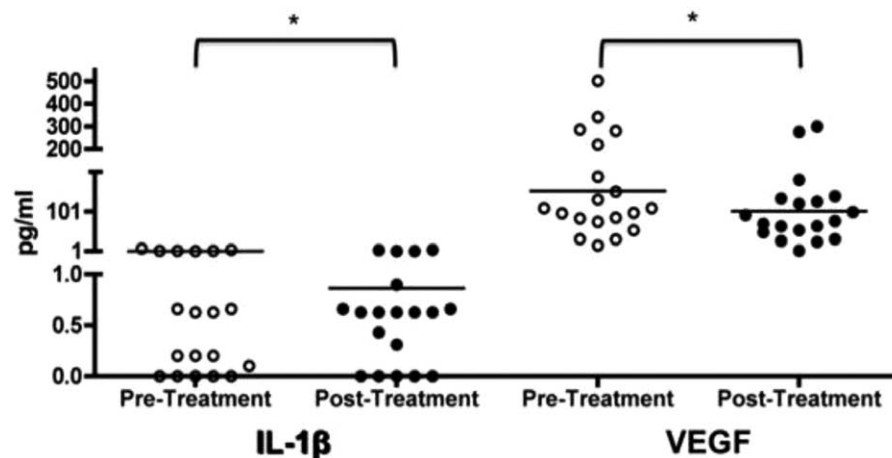


Figure 1. Patient cytokine levels decreased from the initial examination to re-evaluation. Interleukin 1 β (IL-1 β) and vascular endothelial growth factor (VEGF) expression in initial examination (pre-treatment) and re-evaluation (post-treatment) in 19 patients with different periodontitis status and diagnosed with hypertension. The concentrations obtained were expressed in pg/ml and comparisons between groups were performed using Wilcoxon test.

therapy.^[20] The slight change (which was not significant for systolic and diastolic pressure) in blood pressure before and after treatment can be attributed to the sample of patients who were all on medical antihypertensive treatment in the present study. However, we consider that even a difference of 1.89 (systolic pressure) and 2.94 (diastolic pressure) mmHg represents an improvement in the systemic condition of hypertensive patients. Evidence of the change in blood pressure after periodontal treatment in the literature is controversial. In 2013, D’Aiuto et al reported limited evidence on the effects of periodontal therapy on changes in blood pressure. These findings support our results where a slight improvement was also evident although the patients were hypertensive controlled.^[21]

In their systematic review, Muñoz-Aguilera et al found a prevalence of hypertension in patients with periodontal disease versus those without disease. It was concluded that periodontitis could be associated with an increased risk of hypertension in a linear way. The findings highlight the potential to improve cardiovascular disease outcomes by improving oral health in the general population.^[22]

When the systemic inflammatory response was analysed through cytokine levels, a decrease after periodontal treatment

was evident in all the interleukins analysed. Statistically significant differences were noted for IL-1 β and VEGF cytokines ($P = .04$ and $P = .004$), reflecting the decrease in the local bacterial load. In this sense, the literature has also analysed systemic inflammatory cytokines after periodontal treatment. Montenegro et al evaluated patients with stable coronary disease and periodontitis. They found that non-surgical periodontal therapy leads to lower levels of CRP, IL-6, and IL-8 in cardiovascular patients.^[23]

In 2019, a systematic review by D’Isidoro et al analysed the influence of non-surgical periodontal treatment on inflammation biomarkers, reporting that periodontal treatment contributed to the resolution of oral inflammation and could positively influence the levels of biomarkers of systemic inflammation.^[24] Similarly, Roca-Millan et al also found a decrease in TNF- α but not IL-1 α after non-surgical instrumentation and oral hygiene instruction.^[25]

Higashi et al, Vidal et al and Piconi et al, analysed the effect of periodontal therapy on plasma levels of biomarkers of inflammation. Levels of biomarkers of inflammation decreased after periodontal treatment.^[26–28] These results are consistent with those found in our study given that cytokine levels decreased

Table 5

Cut-off point of cytokines levels pre-treatment and post-treatment.

Cut-off point of cytokines levels

	Cut-off point (pg/ml)	Pre-treatment	Post-treatment	P value
IL-10	≤9.1	19	19	Not calculable
	>9.1	0	0	
IL-1 β	≤5	19	19	Not calculable
	>5	0	0	
IL-6	≤3.4	18	19	Not calculable
	>3.4	1	0	
IL-8	≤50	19	19	Not calculable
	>50	0	0	
TNF- α	≤8.1	2	3	.018*
	>8,1	17	16	

* Fisher’s exact test ≤.05.

after periodontal treatment. The novelty of this study is that we analysed a greater number of cytokines, 1 month after the treatment and the patients were hypertensive controlled.

Another relevant finding in our study was the statistically significant decrease in VEGF from 152.10 to 101.55 with a *P* value of .004. The increase in VEGF is related to arterial hypertension. In 2017, Touyz et al analysed recent advances in hypertension and cardiovascular toxicity upon inhibition of VEGF. They emphasized that this biomarker is associated with a higher incidence of cardiovascular pathologies, including hypertension, ischaemic heart disease, heart failure, QT prolongation, and thromboembolism. The magnitude of hypertension induced by this biomarker is significant, and almost all trials report an increase in blood pressure (BP) greater than 150/100 mmHg. The development of hypertension depends on the dose of VEGF.^[29]

We consider that the significant reduction of VEGF in addition to IL-1 β is an important predictor in the improvement of systemic inflammation in patients with arterial hypertension. Together with the improvement of other risk factors, these effects contribute to improved disease control.

In addition, the importance of periodontal therapy must be taken into account, since an improvement in local periodontal inflammation can decrease systemic interleukin levels and help control one of the risk factors of arterial hypertension, namely, systemic inflammation. Therefore, interdisciplinary management together with periodontal treatment can improve the systemic condition of hypertensive patients by controlling risk factors, such as periodontal infection.

5. Conclusions

According to the study performed here and taking into account its limitations, it can be concluded that non-surgical periodontal treatment decreases the levels of systemic proinflammatory cytokines in hypertensive patients. Periodontal treatment improved blood pressure levels and clinical parameters, such as probing depth, clinical insertion level, bleeding and biofilm rate.

These findings are relevant for the interdisciplinary management of hypertensive patients in the control of risk factors for their disease. Today more than ever, more studies of the levels of inflammatory biomarkers after periodontal treatment are needed with a longer follow-up time; however, the results obtained as predictive parameters in subsequent studies should be taken into account and could have a positive impact on public health.

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