#### RESEARCH PAPER

**a** OPEN ACCESS **a** Check for updates

Taylor & Francis Taylor & Francis Group

# **Diagnostic and prognostic value of ABC transporter family member ABCG1 gene in clear cell renal cell carcinoma**

Fucheng Meng<sup>a,b[,†](#page-0-1)</sup>, Yafei Xiao<sup>b,†</sup>, Longxiang Xie<sup>b</sup>, Qiao Liu<sup>[c](#page-0-2)</sup>, [a](#page-0-0)nd Keli Qian<sup>a</sup>

<span id="page-0-2"></span><span id="page-0-0"></span><sup>a</sup>Department of Infection Control, The First Affiliated Hospital of Chongqing Medical University, Chongqing, China; <sup>b</sup>Department of Institute of Biomedical Informatics, Cell Signal Transduction Laboratory, Bioinformatics Center, Henan Provincial Engineering Center for Tumor Molecular Medicine, School of Basic Medical Sciences, Henan University, Kaifeng, China; c Department of Pediatric Dentistry, Stomatological Hospital of Chongqing Medical University, Chongqing, China

#### **ABSTRACT**

As the most common histologic subtype of renal cancer, clear cell renal cell carcinoma (ccRCC) poses a serious threat to public health. However, there are no specific molecular-targeted drugs for ccRCC at present. Human ATP-binding cassette (ABC) transporter family plays an important role in homeostasis maintenance. This study aimed to evaluate the potential diagnostic value of *ABC* genes in ccRCC. A total of 952 samples of ccRCC patients (707) and controls (245) from three different datasets were included for analysis. Receiver operating characteristic analysis and t-test were used to analyze the differential expression of *ABC* genes in ccRCC patients and control samples at mRNA level during screening and validations. The Cancer Genome Atlas (TCGA-ccRCC) dataset was utilized to investigate the correlation between ABC genes expression and prognostic value in ccRCC. We then investigated the interactions between *ABCG1* and proteins in the Comparative Toxicogenomics Database (CTD). Finally, we found that ATP-binding cassette transporter G member 1 (ABCG1) was over-expressed in ccRCC patients compared with healthy samples at mRNA level. Cox regression analysis and Kaplan–Meier analysis showed that ccRCC patients with high *ABCG1* expression had better overall survival (OS) than those patients with low expression (hazard ratio (HR) = 0.662, *p* = 0.007). This study demonstrated that *ABCG1* is a potential diagnostic and prognostic biomarker in ccRCC and discussed the molecular mechanisms underlying the relationship between ccRCC and *ABCG1*, which might provide guidance for better management and treatment of ccRCC in the future.

# **Introduction**

<span id="page-0-4"></span><span id="page-0-3"></span>Renal cell carcinoma (RCC) has become one of the most common malignant tumors in urology and accounts for 85% of primary renal cancer. It was estimated that almost 403,262 (2.2%) new cases of kidney cancer and 175,098 (1.8%) deaths worldwide occurred in 2018 [[1](#page-7-0)]. Besides, the global morbidity and mortality of RCC are increasing by approximately 2–3% per decade [[2](#page-7-1)]. Clear cell renal cell carcinoma (ccRCC) is the most common pathological type of RCC in adults. Surgery is recommended as the preferred option in local ccRCC [\[3\]](#page-7-2), with fiveyear survival at more than 90% [\[4\]](#page-7-3). Then, due to the absence of obvious clinical symptoms at the early stage, cancer metastasis has occurred in 25–30% patients at the time of initial diagnosis of ccRCC [[5](#page-7-4)]. Although there has been a significant progress

<span id="page-0-10"></span><span id="page-0-9"></span><span id="page-0-8"></span><span id="page-0-6"></span>in the management of advanced ccRCC, with improved knowledge of disease and the application of targeted drugs. Five-year survival drops to 12% for patients with metastatic ccRCC [\[4\]](#page-7-3). Therefore, identification and validation of biomarkers will be crucial for optimizing the management of ccRCC. In recent years, many molecular biomarkers for ccRCC have been discovered. C1q/tumor necrosis factor (C1QTNF) [\[6\]](#page-7-5) and six-snoRNA (small nucleolar RNA) signature (*SNORA2*, *SNORD12B*, *SNORA59B*, *SNORA70B*, *SNORD93*, and *SNORD116-2*) [[7\]](#page-7-6) could serve as an independent diagnostic and prognostic indicator for ccRCC. Because of the lack of precise and effective molecular targets for the therapy of ccRCC, it is still important to explore new molecular markers or therapeutic targets for the diagnosis and prognosis [\[8\]](#page-7-7).

#### **ARTICLE HISTORY**

Received 17 February 2021 Revised 15 March 2021 Accepted 15 March 2021

#### **KEYWORDS**

ABCG1; ccRCC; diagnosis; prognosis; cholesterol efflux

<span id="page-0-7"></span><span id="page-0-5"></span><span id="page-0-1"></span>CONTACT Qiao Liu 2 500232@hospital.cqmu.edu.cn; Keli Qian 2 qiankeli86@163.com <sup>†</sup>Co-first author

 $\bigoplus$  Supplemental data for this article can be accessed [here](https://doi.org/10.1080/19336950.2021.1909301).

<sup>© 2021</sup> The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

<span id="page-1-1"></span>Human ATP-binding cassette (ABC) transporter family contains 49 members that are divided into eight subfamilies [\[9\]](#page-7-8). ABC transporter family is a widespread membrane-bound protein, which is mainly distributed in liver, intestine, blood–brain barrier, blood–testosterone barrier, placenta, and kidney. ABC protein can transport various endogenous substrates, including inorganic anions, metal ions, peptides, amino acids, sugars, hydrophobic, and metabolites [\[10\]](#page-7-9). Abnormal changes of the ABC genes can lead to multiple diseases, such as cystic fibrosis and disorder of cholesterol metabolism [[11](#page-7-10)]. It has been demonstrated that cholesterol metabolism disorders are related to various cancers, and the cholesterol level in cancer cells elevates obviously compared with normal tissues [[12](#page-7-11),[13](#page-7-12)].

<span id="page-1-8"></span><span id="page-1-6"></span><span id="page-1-5"></span><span id="page-1-4"></span><span id="page-1-3"></span><span id="page-1-2"></span>Recently, several studies had explored the role of 10 ABC family members in ccRCC. *ABCA1* [[14](#page-8-0)] and *ABCD1* [[15\]](#page-8-1) were related to the occurrence and development of tumors. *ABCA13* [\[16,](#page-8-2)[17](#page-8-3)], *ABCB1* [\[18\]](#page-8-4), *ABCC1* [[19](#page-8-5)], and *ABCC2* [[20](#page-8-6),[21\]](#page-8-7) were associated with drug resistance and treatment of tumors. *ABCB2* and *ABCB3* were found to be involved in tumor immune evasion [\[21,](#page-8-7)[22](#page-8-8)]. *ABCG2* was correlated with tumor progression, prognosis [\[23\]](#page-8-9) and drug resistance [[18](#page-8-4)], and *ABCB10* was associated with tumor progression as well as prognosis [\[24\]](#page-8-10). In this study, we mainly focused on the expression of ABC genes in multiple datasets to assess their diagnostic and prognostic value in ccRCC.

### **Material and methods**

Gene expression of ABC family members in 952 samples from three independent public datasets (GSE40435 dataset, GSE53757 dataset, and TCGAccRCC dataset) was analyzed by screening and verification. Transcriptional expression of ABC genes from the Oncomine database [\(http://www.](http://www.oncomine.org) [oncomine.org\)](http://www.oncomine.org) was also investigated. Then, a prognostic analysis of the validated gene was conducted on TCGA-ccRCC dataset from UCSC Xena (<https://xenabrowser.net>) [\(Figure 1](#page-1-0)). The procedure was similar to the previous studies [\[25](#page-8-11)].

# <span id="page-1-12"></span>*Screening of ABC genes in the Gene Expression Omnibus (GEO) database – ccRCC vs normal samples*

GSE40435 dataset containing 202 samples (101 ccRCC patients and 101 healthy controls) and the corresponding probe set GPL10558 was obtained from NCBI-GEO database ([https://www.ncbi.nlm.nih.gov/geo/](https://www.ncbi.nlm.nih.gov/geo/query/)  [query/](https://www.ncbi.nlm.nih.gov/geo/query/)). T-test and receiver operating characteristic (ROC) analysis were carried out for analyzing the difference in gene expression on 202 samples.  $p < 0.05$  of t-test and the area under curve (AUC) >0.5 of ROC analysis were considered to be significant statistically.

<span id="page-1-11"></span><span id="page-1-10"></span><span id="page-1-9"></span><span id="page-1-7"></span>

<span id="page-1-0"></span>**Figure 1.** Procedure for the selection of the potential biomarker in ccRCC.

# *Identification of genes that have not been reported in ccRCC*

The genes selected in the screening stage were searched for ccRCC-related research in PubMed (https://www. ncbi.nlm.nih.gov/) on 5 December 2019. Our specific advanced search terms included "Renal clear cell carcinoma" OR "Clear cell renal cell carcinoma" OR "Kidney clear cell carcinoma" OR "KIRC" OR "ccRCC." The genes that had not been studied before were considered new genes and then selected for the following validation.

# *Three rounds of validation*

The first round of validation was performed by investigating the expression of the identified genes in the GSE53757 dataset from the NCBI-GEO database. The ROC analysis and t-test were carried out on 144 samples (72 ccRCC patients and 72 controls). Genes with  $p < 0.05$  and AUC  $> 0.85$  were selected for the following validation. The second round of validation was performed on TCGA-ccRCC dataset. ROC analysis and t-test were performed on 606 samples (72 healthy samples and 534 ccRCC patients). Genes with  $p < 0.05$  and  $AUC > 0.85$  were considered validated. The third round of validation was performed by analyzing transcriptional expression of ABC genes in 20 different tumors in the Oncomine database. The thresholds were as follows: *p* value: 0.05; multiple: 2; genetic rank: top 10%; data type: mRNA. Then, we found out the corresponding study on clear cell carcinoma of the kidney. The genes showing an expression ratio >2 or <0.5 were considered effective to be validated.

#### *Prognostic analysis*

To evaluate the prognostic value of the clinical characteristics in ccRCC patients, we analyzed the relationship between *ABCG1* expression and clinical– pathological parameters including carcinoma in situ, expression, age, gender, survival outcome, overall survival (OS), stage, recurrence, survival after recurrence time (RFS), and smoking history from TCGA-ccRCC dataset. Five hundred and thirty-two patients with clinicopathologic information were equally divided into two groups on the basis of the gene expression. Univariate Cox regression analysis

was carried out to find independent variables. Multivariate Cox regression analysis was performed for the parameters with  $p < 0.2$  in univariate Cox regression analysis to assess the prognostic value.

# *Statistical analysis*

ROC analysis and t-test were carried out with GraphPad 8.0 software during screening and validation. In the prognostic analysis step, univariate and multivariate analyses were performed on SPSS19.0. A chi-square test was performed and the OS curve of validated genes was also constructed using GraphPad 8.0 software.

### **Results**

#### *Screening and validation*

The expression data of 49 *ABC* genes were collected from the GSE40435 dataset and samples were divided into two groups (patient group and control group). ROC analysis and t-test were used to evaluate the ability to discriminate ccRCC patients from control samples. The results showed that 39 genes have statistical significance with AUC  $> 0.5$  and  $p < 0.05$ [\(Table 1\)](#page-3-0). A PubMed search was conducted on 5 December 2019. We found 29 genes that had not been reported to be associated with ccRCC ([Table 1\)](#page-3-0).

# *First round of validation*

The 29 genes obtained from the above steps were then validated in the GSE53757 dataset. The 10 genes showing  $AUC > 0.85$  and  $p < 0.05$  were allowed to enter the second round of validation, namely *ABCA3*, *ABCA8*, *ABCA9*, *ABCA12*, *ABCC3*, *ABCC6*, *ABCC8*, *ABCD3*, *ABCF1*, and *ABCG1* (Table S1). Notably, *ABCA12* showed the most significant difference in expression in ccRCC patients vs healthy samples, and the *ABCG1* gene with the highest AUC value.

# *Second round of validation*

Ten genes selected from the first-round validation were analyzed on the TCGA-ccRCC dataset for the second round of validation. Five genes with  $AUC > 0.85$  and  $p < 0.05$  were

<span id="page-3-0"></span>



<sup>a</sup>Novel genes were marked in bold.

statistically significant, namely, *ABAC12*, *ABCC3*, *ABCD3*, *ABCF1*, and *ABCG1* [\(Table](#page-3-1) [2\)](#page-3-1). ROC analysis was carried out to assess the diagnostic value of these five genes. The AUC values of the five genes indicated that they could identify ccRCC patients from normal samples effectively and independently [\(Figure 2\)](#page-4-0).

<span id="page-3-1"></span>**Table 2.** T-test and ROC analysis of ABC transporter family members based on the TCGA-ccRCC dataset.

No.	Gene	<b>Expression</b> (ccRCC)	<b>Expression</b> (normal)	ccRCC/ normal	AUC	p
	ARCA12	6.50	3.42	1.90	0.88	< 0.0001
$\mathcal{P}$	ABCC3	11.56	9.14	1.26	0.93	< 0.0001
3	ABCD3	10.43	11.49	0.91	0.95	< 0.0001
4	ARCF1	10.24	10.67	0.96	0.87	< 0.0001
5	ARCG1	10.44	9.09	1.15	በ 91	< 0.0001

#### *Third round of validation*

Transcriptional expression of the above five genes was verified in the Oncomine database. The expression of *ABCC3*, *ABCF1*, and *ABCG1* in 20 types of cancers is shown in Figure S1. *ABCA12* and *ABCD3* had no available data. There are eight, five, and eight datasets of *ABCC3*, *ABCF1*, and *ABCG1* for renal cancer, respectively. However, there is no clear cell renal cell carcinoma vs normal in the five datasets of *ABCF1.*  The result showed that *ABCG1* was highly overexpressed in all datasets (Table S2, Table S3).

#### *Prognostic analysis*

To explore the prognostic value of *ABCG1* expression in the TCGA-ccRCC dataset, Cox regression



<span id="page-4-0"></span>**Figure 2.** ROC analysis of the expression data for diagnostic assessment of five genes according to TCGA database. AUC statistics are used to evaluate the capacity to discriminate ccRCC samples from normal controls with specificity and sensitivity.

analysis and Kaplan–Meier analysis were performed. The relationship between *ABCG1* expression and clinical characteristics in ccRCC is shown in [Table 3.](#page-4-1) We found significant differences in living status between the high-expression group and the low-expression group ( $p = 0.01$ ), but there was no statistical difference in gender, clinical stage, smoking history, and recurrence history

between the two groups. Univariate Cox regression analysis showed that age, clinical stage, and *ABCG1* expression were associated with OS [\(Table 4\)](#page-5-0). Meanwhile, multivariate Cox regression analysis demonstrated that *ABCG1* expression might be an independent prognostic factor for ccRCC patients. Kaplan–Meier analysis also showed that ccRCC patients with high *ABCG1* 

		ABCG1 mRNA expression				
Parameters	Group	High $(n = 266)$	Low $(n = 266)$	χ2	р	
Age (Mean $+$ SD)		$60.80 \pm 12.77$	$60.36 \pm 12.03$			
Gender	Female	82	104	4.001	0.056	
	Male	184	162			
Clinical stage	I/II	164	188	4.872	0.034	
	<b>III/IV</b>	101	77			
	Discrepancy					
Recurrence status	No	68	53	0.325	0.655	
	Yes	15	9			
	Null	183	204			
Smoking history		29	16	2.147	0.143	
	2/3/4/5	20	21			
	Null	229	217			
Living status	Living	193	164	7.161	0.010	
	Dead	73	102			

<span id="page-4-1"></span>**Table 3.** Chi-square test of clinicopathologic parameters and *ABCG1* mRNA expression in the TCGA-ccRCC cohort.

	Univariate analysis			Multivariate analysis			
Parameters	<b>HR</b>	95%CI		<b>HR</b>	95%CI	р	
Age							
$≥60$ vs $<60$	1.808	1.321-2.478	0.000	1.563	1.140-2.144	0.006	
Gender							
Female vs male	1.060	$0.779 - 1.442$	0.710		$\overline{\phantom{a}}$	-	
Clinical stage							
$III/IV$ vs $I/II$	3.838	2.799-5.292	0.000	3.675	2.676-5.046	0.000	
Smoking history							
$2/3/4/5$ vs 1	0.778	$0.254 - 2.383$	0.661		۰	-	
ABCG1 expression							
High vs low	0.665	$0.492 - 0.898$	0.008	0.662	$0.490 - 0.895$	0.007	

<span id="page-5-0"></span>**Table 4.** Univariate and multivariate Cox regression analysis of clinical pathologic features according to the TCGA-ccRCC dataset.

expression was significantly associated with better OS than those patients with low *ABCG1* expression ( $p = 0.0067$ , hazard ratio (HR) = 0.6621) [\(Figure 3](#page-5-1)).

### **Discussion**

<span id="page-5-5"></span><span id="page-5-4"></span><span id="page-5-2"></span>Many studies have demonstrated that ABC family genes play important roles in the maintaining cellular environment [[26,](#page-8-12)[27\]](#page-8-13), cholesterol metabolism [[9,](#page-7-8)[10,](#page-7-9)[28–31\]](#page-8-14), disease occurrence [[-](#page-8-15) [32–34\]](#page-8-15), and tumor resistance [[35,](#page-8-16)[36\]](#page-8-17). ABC transporter genes could promote drug efflux and enhance chemical resistance of cancer cells [[37](#page-8-18)]. Mutations in the *ABC* genes could affect

<span id="page-5-9"></span><span id="page-5-8"></span><span id="page-5-7"></span><span id="page-5-3"></span>the phenotypes of cancer cells such as proliferation, differentiation, migration, and invasion [[38](#page-8-19)]. *ABCG1* is involved in lipid balance and cholesterol efflux from macrophages [[10](#page-7-9)]. *ABCG1* is also able to transport sterols, which can regulate the expression of macrophage inflammatory cytokines, chemokines, and lymphocyte proliferation response [[28](#page-8-14)]. *ABCG1* was found to be a potential biomarker for lung cancer [\[39](#page-8-20)[,40\]](#page-8-21), head and neck squamous cell carcinoma [[41](#page-8-22)], and prostate cancer [\[42](#page-8-23)]. However, it has not been studied in ccRCC. After screening, identification, and three rounds of verification, *ABCG1* was selected from 49 ABC transporter genes. We first showed that *ABCG1* has the

<span id="page-5-6"></span>

<span id="page-5-1"></span>**Figure 3.** Kaplan–Meier survival curve of *ABCG1* mRNA expression in ccRCC patients. Survival curves suggested that patients with decreased *ABCG1* mRNA level significantly correlated with poorer OS (*p* = 0.0067).

diagnostic and prognostic value for ccRCC patients.

<span id="page-6-4"></span><span id="page-6-3"></span><span id="page-6-2"></span><span id="page-6-1"></span>Metabolic change is the main feature of tumors [[43](#page-9-0)], and ccRCC is also considered as metabolic disease [\[44](#page-9-1)], which is characterized by the accumulation of cholesterol, cholesterol esters, other neutral lipids, and glycogen [\[45\]](#page-9-2). The total cholesterol content in the ccRCC tissues is eight times higher than that of the normal kidney, and the esterified cholesterol content is 35 times higher than that of the normal kidney [\[46\]](#page-9-3). The abnormalities in cholesterol metabolism in ccRCC cells may affect the physiological and biochemical functions of cells and produce pathological changes. Many studies have shown that serum cholesterol levels are associated with ccRCC invasion and prognosis [\[47–49\]](#page-9-4). Patients with low preoperative cholesterol levels have lower OS than patients with high cholesterol levels [\[47](#page-9-4)], and cholesterol metabolism may be involved in ccRCC metastasis [\[50\]](#page-9-5). The function of the ABCG1 gene is mainly related to cell cholesterol outflow [\[10\]](#page-7-9), which indicates that *ABCG1*  may play an important role in the tumorigenesis and progress of ccRCC.

<span id="page-6-5"></span>Our study showed that *ABCG1* was overexpressed among patients with ccRCC compared with normal people [\(Table 1–2,](#page-3-0) Table S1, [Figure](#page-5-1)  [3\)](#page-5-1). It may be hypothesized that when normal cells mutate into cancer cells, the energy demand increases, which activates a certain cholesterol transport mechanism and begins to take cholesterol from the outside. The decrease in serum cholesterol levels is correlated with the uptake of low-density lipoprotein in serum by tumor cells [[49](#page-9-6)]. Yang et al. found that the accumulation of cholesterol is one of the characteristics of ccRCC [[50](#page-9-5)]. As serum cholesterol decreased, *ABCG1*  began to be highly expressed and promoted the efflux of cellular cholesterol to maintain serum cholesterol level.

<span id="page-6-7"></span><span id="page-6-6"></span>Moreover, we found that ccRCC patients with high expression level had longer OS time than those with low expression level. It may be that the high expression of *ABCG1* can inhibit the growth of cancer cells and affect the survival of cancer cells by reducing the cholesterol content in cancer cells. Wu et al. found that liver X receptor 623 (LXR623) downregulates low-density lipoprotein receptor (LDLR) expression while upregulating *ABCA1*, leading to a decrease in intracellular cholesterol content and the occurrence of

apoptosis [[14](#page-8-0)]. They speculated that LXR623 could kill tumor cells by promoting cholesterol outflow [\[14\]](#page-8-0). *ABCG1* and *ABCA1* have many similarities: both of them are regulated by LXR623 [[51–54\]](#page-9-7) and they have 101 common interacting chemicals (Figure S2), promoting the outflow of cholesterol from macrophages [[51–54\]](#page-9-7) and regulating the expression of macrophage inflammatory cytokines [\[28](#page-8-14)], etc. Hence, it may be possible to kill or suppress tumor cells by upregulating *ABCG1* with LXR623.

<span id="page-6-12"></span><span id="page-6-11"></span><span id="page-6-10"></span><span id="page-6-9"></span><span id="page-6-8"></span>In addition, *ABCG1* could affect tumor growth by regulating macrophages. Macrophages participate in the formation of tumor microenvironment [\[55,](#page-9-8)[56\]](#page-9-9), tumor growth, and metastasis [\[57–60\]](#page-9-10), apoptosis [\[61\]](#page-9-11), and play an important role in tumor immunity [\[62\]](#page-9-12). There are two types of it: M1 cells can produce a large number of inflammatory cytokines, which can activate the immune response and play an anti-tumor role; M2 cells promote angiogenesis, remodeling, and tumor growth [[63\]](#page-9-13). In most tumor models, most of the macrophages in the tumor are shown as tumor promoting M2 phenotype [\[62\]](#page-9-12). Researches have shown that the deficiency of *ABCG1* increases the signaling of Toll-like receptors in macrophages, leading to an enhanced inflammatory response of macrophages to LPS or other TLR ligands [\[64–67\]](#page-9-14), and also reduces the number and proportion of M2 phenotype [\[29,](#page-8-24)[68\]](#page-9-15). This means that the upregulation of *ABCG1* leads to an increase in M2 macrophages, which is conducive to tumor growth. However, this is inconsistent with the result that the high *ABCG1* expression group can have a longer survival time, so more work is needed to explain the problem.

<span id="page-6-13"></span><span id="page-6-0"></span>Furthermore, we identified 35 potential active drugs from 165 interplay substances with *ABCG1* in the Comparative Toxicogenomics Database (CTD) [\(http://](http://ctdbase.org) [ctdbase.org](http://ctdbase.org)), of which 18 drugs can upregulate *ABCG1*  expression at mRNA level, while 17 drugs can downregulate *ABCG1* expression (Table S4). In addition, six *ABCG1* target genes were identified from 60 *ABCG1*  interacting genes: *ACSL4* [\[69](#page-9-16)], *AP1S2* [\[69\]](#page-9-16), *ELAVL1*  [\[70\]](#page-10-0), *HNRNPL* [\[71\]](#page-10-1), *SGO1* [\[69\]](#page-9-16), and *UBC* [\[72](#page-10-2)] (Table S5).

<span id="page-6-14"></span>There are still several limitations in this study. In the screening and the first rounds of validation analysis, *ABCA12* and *ABCD3* were significantly different between ccRCC and normal tissues. Unfortunately, there was no data available for *ABCA12* and *ABCD3*  in the Oncomine database. But it could not exclude that *ABCA12* may be associated with the diagnosis and prognosis in ccRCC. Protein expression of *ABC*  genes was analyzed before prognostic analysis in the HPA database [\(https://www.proteinatlas.org/\)](https://www.proteinatlas.org/). Grayscale conversion analysis was carried out on a total of 57 histological sections (11 normal tissue sections and 46 pathological sections of ccRCC patients) using ImageJ (Table S6). The t-test was then performed and a scatter plot was constructed according to the area percentage of histological sections (Figure S3). As shown in Figure S3, *ABCC3* was not statistically significant in t-test, and in the protein expression level of *ABCG1* was reduced in ccRCC patients, while positively expressed in normal tissue sections. It is not consistent with the results of mRNA level, which may be related to the type and specificity of antibody and the small sample size. Additional works and experiments need to be performed to validate them.

### **Conclusion**

Excluding the ABC family members that have been studied, through multiple rounds of validation, a novel diagnostic and prognostic biomarker of ccRCC – *ABCG1* – was found. According to the high expression of *ABCG1* in ccRCC and its correlation with better prognosis, it may be helpful for the diagnosis and provides new ideas for the development of molecular-targeted drugs for ccRCC.

### **Acknowledgments**

This study was supported by Program for Innovative Talents of Science and Technology in Henan Province (No. 202102310205), China Postdoctoral Science Foundation (No. 2017M62237), and Henan Postdoctoral Foundation (No. 001702052).

#### **Disclosure statement**

No potential conflict of interest was reported by the authors.

#### **Funding**

This work was supported by the Program for Innovative Talents of Science and Technology in Henan Province [202102310205]; Henan Postdoctoral Foundation

[001702052]; China Postdoctoral Science Foundation [2017M62237].

#### **References**

- <span id="page-7-0"></span>[1] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. [2018](#page-0-3) Nov;68(6):394–424.
- <span id="page-7-1"></span>[2] Gupta K, Miller JD, Li JZ, et al. Epidemiologic and socioeconomic burden of metastatic renal cell carcinoma (mRCC): a literature review. Cancer Treat Rev. [2008](#page-0-4) May;34(3):193–205.
- <span id="page-7-2"></span>[3] Escudier B, Porta C, Schmidinger M, et al. Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. [2019](#page-0-5)  May 1;30(5):706–720.
- <span id="page-7-3"></span>[4] Atkins MB, Tannir NM. Current and emerging therapies for first-line treatment of metastatic clear cell renal cell carcinoma. Cancer Treat Rev. [2018](#page-0-6)  Nov;70:127–137.
- <span id="page-7-4"></span>[5] Motzer RJ, Bander NH, Nanus DM. Renal-cell carcinoma. N Engl J Med. [1996](#page-0-7) Sep 19;335 (12):865–875.
- <span id="page-7-5"></span>[6] Lin W, Chen X, Chen T, et al. C1QTNF6 as a novel diagnostic and prognostic biomarker for clear cell renal cell carcinoma. DNA Cell Biol. [2020](#page-0-8) Apr 13;39 (6):1000–1011.
- <span id="page-7-6"></span>[7] Zhao Y, Yan Y, Ma R, et al. Expression signature of six-snoRNA serves as novel non-invasive biomarker for diagnosis and prognosis prediction of renal clear cell carcinoma. J Cell Mol Med. [2020](#page-0-9) Feb;24 (3):2215–2228.
- <span id="page-7-7"></span>[8] Xu D, Xu Y, Lv Y, et al. Identification of four pathological stage-relevant genes in association with progression and prognosis in clear cell renal cell carcinoma by integrated bioinformatics analysis. Biomed Res Int. [2020](#page-0-10);2020:2137319.
- <span id="page-7-8"></span>[9] Dean M, Hamon Y, Chimini G. The human ATP-binding cassette (ABC) transporter superfamily. J Lipid Res. [2001](#page-1-1) Jul;42(7):1007–1017.
- <span id="page-7-9"></span>[10] Vasiliou V, Vasiliou K, Nebert DW. Human ATP-binding cassette (ABC) transporter family. Hum Genomics. [2009](#page-1-2) Apr 3;3(3):281–290.
- <span id="page-7-10"></span>[11] Lam F, Hussain S, Sinha J. An unusual cause of a limp in a child: developmental coxa vara. Emerg Med J. [2001](#page-1-3)  Jul;18(4):314.
- <span id="page-7-11"></span>[12] Rudling M, Collins VP. Low density lipoprotein receptor and 3-hydroxy-3-methylglutaryl coenzyme A reductase mRNA levels are coordinately reduced in human renal cell carcinoma. Biochim Biophys Acta. [1996](#page-1-4) Jan 5;1299(1):75–79.
- <span id="page-7-12"></span>[13] Dessi S, Batetta B, Pulisci D, et al. Cholesterol content in tumor tissues is inversely associated with high-density lipoprotein cholesterol in serum in

patients with gastrointestinal cancer. Cancer. [1994](#page-1-4) Jan 15;73(2):253–258.

- <span id="page-8-0"></span>[14] Wu G, Wang Q, Xu Y, et al. Targeting the transcription factor receptor LXR to treat clear cell renal cell carcinoma: agonist or inverse agonist? Cell Death Dis. [2019](#page-1-5) May 28;10(6):416.
- <span id="page-8-1"></span>[15] Hour TC, Kuo YZ, Liu GY, et al. Downregulation of ABCD1 in human renal cell carcinoma. Int J Biol Markers. [2009](#page-1-5) Jul-Sep;24(3):171–178.
- <span id="page-8-2"></span>[16] Yun EJ, Zhou J, Lin CJ, et al. The network of DAB2IP-miR-138 in regulating drug resistance of renal cell carcinoma associated with stem-like phenotypes. Oncotarget. [2017](#page-1-6) Sep 15;8(40):66975–66986.
- <span id="page-8-3"></span>[17] Arai E, Sakamoto H, Ichikawa H, et al. Multilayeromics analysis of renal cell carcinoma, including the whole exome, methylome and transcriptome. Int J Cancer. [2014](#page-1-6) Sep 15;135(6):1330–1342.
- <span id="page-8-4"></span>[18] Reustle A, Fisel P, Renner O, et al. Characterization of the breast cancer resistance protein (BCRP/ABCG2) in clear cell renal cell carcinoma. Int J Cancer. [2018](#page-1-7) Dec 15;143(12):3181–3193.
- <span id="page-8-5"></span>[19] Li S, Yang J, Wang J, et al. Down-regulation of miR-210-3p encourages chemotherapy resistance of renal cell carcinoma via modulating ABCC1. Cell Biosci. [2018;](#page-1-6)8(1):9.
- <span id="page-8-6"></span>[20] Haenisch S, Zimmermann U, Dazert E, et al. Influence of polymorphisms of ABCB1 and ABCC2 on mRNA and protein expression in normal and cancerous kidney cortex. Pharmacogenomics J. [2007](#page-1-8) Feb 7;7  $(1):56-65.$
- <span id="page-8-7"></span>[21] Saleeb RM, Farag M, Lichner Z, et al. Modulating ATP binding cassette transporters in papillary renal cell carcinoma type 2 enhances its response to targeted molecular therapy. Mol Oncol. [2018](#page-1-9) Oct;12(10):1673–1688.
- <span id="page-8-8"></span>[22] Seliger B, Atkins D, Bock M, et al. Characterization of human lymphocyte antigen class I antigen-processing machinery defects in renal cell carcinoma lesions with special emphasis on transporter-associated with antigen-processing down-regulation. Clin Cancer Res. [2003](#page-1-9) May;9(5):1721–1727.
- <span id="page-8-9"></span>[23] Kuo CY, Yoo TJ. In vitro inhibition of tritiated thymidine uptake in Morris hepatoma cells by normal rat liver extract: a possible liver chalone. J Natl Cancer Inst. [1977](#page-1-10) Dec;59 (6):1691–1695.
- <span id="page-8-10"></span>[24] Huang Y, Zhang Y, Jia L, et al. Circular RNA ABCB10 promotes tumor progression and correlates with pejorative prognosis in clear cell renal cell carcinoma. Int J Biol Markers. [2019](#page-1-11) Jun;34(2):176–183.
- <span id="page-8-11"></span>[25] D'Arcangelo D, Giampietri C, Muscio M, et al. WIPI1, BAG1, and PEX3 autophagy-related genes are relevant melanoma markers. Oxid Med Cell Longev. [2018](#page-1-12);2018.
- <span id="page-8-12"></span>[26] Chai AB, Ammit AJ, Gelissen IC. Examining the role of ABC lipid transporters in pulmonary lipid homeostasis and inflammation. Respir Res. [2017](#page-5-2) Feb 28;18(1):41.
- <span id="page-8-13"></span>[27] Zhu L, Tang F, Lei Z, et al. Antiapoptotic properties of MALT1 protease are associated with redox homeostasis

in ABC-DLBCL cells. Mol Carcinog. [2019](#page-5-2) Sep 26;58 (12):2340–2352. .

- <span id="page-8-14"></span>[28] Yvan-Charvet L, Wang N, Tall AR. Role of HDL, ABCA1, and ABCG1 transporters in cholesterol efflux and immune responses. Arterioscler Thromb Vasc Biol. [2010](#page-5-3) Feb;30(2):139–143.
- <span id="page-8-24"></span>[29] Sag D, Cekic C, Wu R, et al. The cholesterol transporter ABCG1 links cholesterol homeostasis and tumour immunity. Nat Commun. [2015](#page-6-0) Feb 27;6(1):6354.
- [30] Kober AC, Manavalan APC, Tam-Amersdorfer C, et al. Implications of cerebrovascular ATP-binding cassette transporter G1 (ABCG1) and apolipoprotein M in cholesterol transport at the blood-brain barrier. Biochim Biophys Acta Mol Cell Biol Lipids. 2017 Jun;1862 (6):573–588.
- [31] Vaughan AM, Oram JF. ABCA1 and ABCG1 or ABCG4 act sequentially to remove cellular cholesterol and generate cholesterol-rich HDL. J Lipid Res. 2006 Nov;47(11):2433–2443.
- <span id="page-8-15"></span>[32] Pereira CD, Martins F, Wiltfang J, et al. ABC transporters are key players in Alzheimer's disease. J Alzheimers Dis. [2018](#page-5-4);61(2):463–485.
- [33] Schumacher T, Benndorf RA, Transport ABC. Proteins in cardiovascular disease—a brief summary. Molecules. 2017 Apr 6;22(4):589.
- [34] Tarling EJ, De Aguiar Vallim TQ, Edwards PA. Role of ABC transporters in lipid transport and human disease. Trends Endocrinol Metab. 2013 Jul;24(7):342–350.
- <span id="page-8-16"></span>[35] Harandi NM, Stavness I, Woo J. Subject-specific biomechanical modelling of the oropharynx: towards speech production. Comput Methods Biomech Biomed Eng Imaging Vis. [2017](#page-5-5);5(6):416–426.
- <span id="page-8-17"></span>[36] Liu X. ABC family transporters. Adv Exp Med Biol. [2019](#page-5-5);1141:13–100.
- <span id="page-8-18"></span>[37] Briz O, Perez-Silva L, Al-Abdulla R, et al. What "The Cancer Genome Atlas" database tells us about the role of ATP-binding cassette (ABC) proteins in chemoresistance to anticancer drugs. Expert Opin Drug Metab Toxicol. [2019](#page-5-6) Jul;15(7):577–593.
- <span id="page-8-19"></span>[38] Fletcher JI, Williams RT, Henderson MJ, et al. ABC transporters as mediators of drug resistance and contributors to cancer cell biology. Drug Resist Updat. [2016](#page-5-7) May;26:1–9.
- <span id="page-8-20"></span>[39] Tian C, Huang D, Yu Y, et al. ABCG1 as a potential oncogene in lung cancer. Exp Ther Med. [2017](#page-5-8) Jun;13 (6):3189–3194.
- <span id="page-8-21"></span>[40] Wang Y, Liu H, Ready NE, et al. Genetic variants in ABCG1 are associated with survival of nonsmall-cell lung cancer patients. Int J Cancer. [2016](#page-5-8) Jun 1;138 (11):2592–2601.
- <span id="page-8-22"></span>[41] Gonzalez HE, Gujrati M, Frederick M, et al. Identification of 9 genes differentially expressed in head and neck squamous cell carcinoma. Arch Otolaryngol Head Neck Surg. [2003](#page-5-9) Jul;129(7):754–759.
- <span id="page-8-23"></span>[42] Demidenko R, Razanauskas D, Daniunaite K, et al. Frequent down-regulation of ABC transporter genes

in prostate cancer. BMC Cancer. [2015](#page-5-9) Oct 12;15 (1):683.

- <span id="page-9-0"></span>[43] Jiang L, Zhao L, Bi J, et al. Glycolysis gene expression profilings screen for prognostic risk signature of hepatocellular carcinoma. Aging (Albany NY). [2019](#page-6-1) Dec 2;11(23):10861–10882.
- <span id="page-9-1"></span>[44] Massari F, Ciccarese C, Santoni M, et al. Metabolic alterations in renal cell carcinoma. Cancer Treat Rev. [2015](#page-6-2) Nov;41(9):767–776.
- <span id="page-9-2"></span>[45] Drabkin HA, Gemmill RM. Cholesterol and the development of clear-cell renal carcinoma. Curr Opin Pharmacol. [2012](#page-6-3) Dec;12(6):742–750.
- <span id="page-9-3"></span>[46] Gebhard RL, Clayman RV, Prigge WF, et al. Abnormal cholesterol metabolism in renal clear cell carcinoma. J Lipid Res. [1987](#page-6-4) Oct;28(10):1177–1184.
- <span id="page-9-4"></span>[47] Kang HW, Seo SP, Kim WT, et al. Low preoperative serum cholesterol level is associated with aggressive pathologic features and poor cancer-specific survival in patients with surgically treated renal cell carcinoma. Int J Clin Oncol. [2018](#page-6-5) Feb;23(1):142–150.
- [48] Lee H, Kim YJ, Hwang EC, et al. Preoperative cholesterol level as a new independent predictive factor of survival in patients with metastatic renal cell carcinoma treated with cyto-reductive nephrectomy. BMC Cancer. 2017 May 25;17(1):364.
- <span id="page-9-6"></span>[49] Peng D, He ZS, Li XS, et al. A novelpredictor of survival with renal cell carcinoma after nephrectomy. J Endourol. [2017](#page-6-6) Apr;31(4):397–404.
- <span id="page-9-5"></span>[50] Yang H, Li W, Lv Y, et al. Exploring the mechanism of clear cell renal cell carcinoma metastasis and key genes based on multi-tool joint analysis. Gene. [2019](#page-6-7) Dec 15;720:144103.
- <span id="page-9-7"></span>[51] Frambach S, De Haas R, Smeitink JAM, et al. Brothers in arms: ABCA1- and ABCG1-mediated cholesterol efflux as promising targets in cardiovascular disease treatment. Pharmacol Rev. [2020](#page-6-8) Jan;72(1):152–190.
- [52] Ren K, Li H, Zhou HF, et al. Mangiferin promotes macrophage cholesterol efflux and protects against atherosclerosis by augmenting the expression of ABCA1 and ABCG1. Aging (Albany NY). 2019 Dec 2;11(23):10992–11009.
- [53] Huesca-Gomez C, Torres-Paz YE, Martinez-Alvarado R, et al. Association between the transporters ABCA1/ G1 and the expression of miR-33a/144 and the carotid intima media thickness in patients with arterial hypertension. Mol Biol Rep. 2020 Feb;47(2):1321–1329.
- [54] Koga M, Kanaoka Y, Okamoto M, et al. Varenicline aggravates atherosclerotic plaque formation in nicotine-pretreated ApoE knockout mice due to enhanced oxLDL uptake by macrophages through downregulation of ABCA1 and ABCG1 expression. J Pharmacol Sci. 2020 Jan;142(1):9–15.
- <span id="page-9-8"></span>[55] Lewis CE, Pollard JW. Distinct role of macrophages in different tumor microenvironments. Cancer Res. [2006](#page-6-9)  Jan 15;66(2):605–612.
- <span id="page-9-9"></span>[56] Movahedi K, Laoui D, Gysemans C, et al. Different tumor microenvironments contain functionally distinct

subsets of macrophages derived from Ly6C(high) monocytes. Cancer Res. [2010](#page-6-9) Jul 15;70(14):5728–5739.

- <span id="page-9-10"></span>[57] Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. Nat Rev Cancer. [2004](#page-6-10) Jan 4;4(1):71–78.
- [58] Lin EY, Li JF, Gnatovskiy L, et al. Macrophages regulate the angiogenic switch in a mouse model of breast cancer. Cancer Res. 2006 Dec 1;66(23):11238–11246.
- [59] Sica A, Larghi P, Mancino A, et al. Macrophage polarization in tumour progression. Semin Cancer Biol. 2008 Oct;18(5):349–355.
- [60] Hiraoka K, Zenmyo M, Watari K, et al. Inhibition of bone and muscle metastases of lung cancer cells by a decrease in the number of monocytes/macrophages. Cancer Sci. 2008 Aug;99(8):1595–1602.
- <span id="page-9-11"></span>[61] Li Y, Schwabe RF, DeVries-Seimon T, et al. Free cholesterol-loaded macrophages are an abundant source of tumor necrosis factor-alpha and interleukin-6: model of NF-kappaB- and map kinase-dependent inflammation in advanced atherosclerosis. J Biol Chem. [2005](#page-6-10) Jun 10;280 (23):21763–21772.
- <span id="page-9-12"></span>[62] Solinas G, Germano G, Mantovani A, et al. Tumorassociated macrophages (TAM) as major players of the cancer-related inflammation. J Leukoc Biol. [2009](#page-6-11)  Nov;86(5):1065–1073.
- <span id="page-9-13"></span>[63] Wei H, Tarling EJ, McMillen TS, et al. ABCG1 regulates mouse adipose tissue macrophage cholesterol levels and ratio of M1 to M2 cells in obesity and caloric restriction. J Lipid Res. [2015](#page-6-12) Dec;56(12):2337–2347.
- <span id="page-9-14"></span>[64] Buczek M, Escudier B, Bartnik E, et al. Resistance to tyrosine kinase inhibitors in clear cell renal cell carcinoma: from the patient's bed to molecular mechanisms. Biochim Biophys Acta. [2014](#page-6-13) Jan;1845(1):31–41.
- [65] Van Der Mijn JC, Mier JW, Broxterman HJ, et al. Predictive biomarkers in renal cell cancer: insights in drug resistance mechanisms. Drug Resist Updat. 2014 Oct-Dec;17(4–6):77–88.
- [66] Randall JM, Millard F, Kurzrock R. Molecular aberrations, targeted therapy, and renal cell carcinoma: current state-of-the-art. Cancer Metastasis Rev. 2014 Dec;33(4):1109–1124.
- [67] Garcia-Donas J, Rodriguez-Moreno JF, Romero-Laorden N, et al. Renal carcinoma pharmacogenomics and predictors of response: steps toward treatment individualization. Urol Oncol. 2015 Apr;33(4):179–186.
- <span id="page-9-15"></span>[68] Yvan-Charvet L, Welch C, Pagler TA, et al. Increased inflammatory gene expression in ABC transporter-deficient macrophages: free cholesterol accumulation, increased signaling via toll-like receptors, and neutrophil infiltration of atherosclerotic lesions. Circulation. [2008](#page-6-0) Oct 28;118(18):1837–1847.
- <span id="page-9-16"></span>[69] Hein MY, Hubner NC, Poser I, et al. A human interactome in three quantitative dimensions organized by stoichiometries and abundances. Cell. [2015](#page-6-14) Oct 22;163 (3):712–723.
- <span id="page-10-0"></span>[70] Abdelmohsen K, Srikantan S, Yang X, et al. Ubiquitinmediated proteolysis of HuR by heat shock. Embo J. [2009](#page-6-14)  May 6;28(9):1271–1282.
- <span id="page-10-1"></span>[71] Fei T, Chen Y, Xiao T, et al. Genome-wide CRISPR screen identifies HNRNPL as a prostate cancer

dependency regulating RNA splicing. Proc Natl Acad Sci U S A. [2017](#page-6-14) Jun 27;114(26):E5207–E5215.

<span id="page-10-2"></span>[72] Na CH, Jones DR, Yang Y, et al. Synaptic protein ubiquitination in rat brain revealed by antibody-based ubiquitome analysis. J Proteome Res. [2012](#page-6-14) Sep 7;11(9):4722–4732.