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# Diffuse axonal injury has a characteristic multidimensional MRI signature in the human brain

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Axonal injury is a major contributor to the clinical symptomatology in patients with traumatic brain injury. Conventional neuroradiological tools, such as CT and MRI, are insensitive to diffuse axonal injury (DAI) caused by trauma. Diffusion tensor MRI parameters may change in DAI lesions; however, the nature of these changes is inconsistent. Multidimensional MRI is an emerging approach that combines T<sub>1</sub>, T<sub>2</sub>, and diffusion, and replaces voxel-averaged values with distributions, which allows selective isolation of specific potential abnormal components. By performing a combined post-mortem multidimensional MRI and histopathology study, we aimed to investigate T<sub>1</sub>-T<sub>2</sub>-diffusion changes linked to DAI and to define their histopathological correlates. Corpora callosa derived from eight subjects who had sustained traumatic brain injury, and three control brain donors underwent post-mortem ex vivo MRI at 7 T. Multidimensional, diffusion tensor, and quantitative T<sub>1</sub> and T<sub>2</sub> MRI data were acquired and processed. Following MRI acquisition, slices from the same tissue were tested for amyloid precursor protein (APP) immunoreactivity to define DAI severity. A robust image co-registration method was applied to accurately match MRI-derived parameters and histopathology, after which 12 regions of interest per tissue block were selected based on APP density, but blind to MRI. We identified abnormal multidimensional  $T_1$ - $T_2$ , diffusion- $T_2$ , and diffusion- $T_1$  components that are strongly associated with DAI and used them to generate axonal injury images. We found that compared to control white matter, mild and severe DAI lesions contained significantly larger abnormal  $T_1$ - $T_2$  component (P = 0.005 and P < 0.001, respectively), and significantly larger abnormal diffusion-T<sub>2</sub> component (P = 0.005 and P < 0.001, respectively). Furthermore, within patients with traumatic brain injury the multidimensional MRI biomarkers differentiated normal-appearing white matter from mild and severe DAI lesions, with significantly larger abnormal  $T_1$ - $T_2$  and diffusion- $T_2$  components (P = 0.003 and P < 0.001, respectively, for  $T_1$ - $T_2$ ; P = 0.022 and P < 0.001, respectively, for diffusion-T<sub>2</sub>). Conversely, none of the conventional quantitative MRI parameters were able to differentiate lesions and normal-appearing white matter. Lastly, we found that the abnormal  $T_1$ - $T_2$ , diffusion- $T_1$ , and diffusion- $T_2$  components and their axonal damage images were strongly correlated with quantitative APP staining (r = 0.876, P < 0.001; r = 0.727, P < 0.001; and r = 0.743, P < 0.001, respectively), while producing negligible intensities in grey matter and in normal-appearing white matter. These results suggest that multidimensional MRI may provide non-invasive biomarkers for detection of DAI, which is the pathological substrate for neurological disorders ranging from concussion to severe traumatic brain injury.

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**Abbreviations:** DAI = diffuse axonal injury; DTI = diffusion tensor imaging; FA = fractional anisotropy; MD = mean diffusivity; SC = spectral component; TAI = traumatic axonal injury; TBI = traumatic brain injury

## Introduction

Traumatic brain injury (TBI) presents major medical, social and economic challenges world-wide and is associated with high mortality, co-morbidities, and long-term disabilities (Faul and Coronado, 2015; Sharp *et al.*, 2016). Among the various pathological brain lesions produced by impact (e.g. intracerebral bleedings, necrotic-ischaemic lesions, tissue avulsion), diffuse axonal injury (DAI) is caused by mechanical damage to axons multifocally throughout the white matter (Adams *et al.*, 1982).

From severe closed-head injuries to concussions, the degree of injury and the duration of loss of consciousness and coma directly correlate with the extent of axonal pathology (Johnson et al., 2013; Smith and Stewart, 2020). Axons become stiff and brittle following rapid deformation caused by rotation acceleration of the brain, which leads to immediate mechanical damage to the axonal cytoskeleton. As the axonal transport is disrupted, pathological accumulation of different axonal proteins causes axonal swelling, which leads to axonal degeneration (Johnson et al., 2013). Amyloid precursor protein (APP) is one of those accumulated proteins, and it can be detected immunohistochemically within a few hours post injury (Gentleman et al., 1993, 1995; Sherriff et al., 1994). Because of that, and because of its high sensitivity and robustness, APP remains the gold standard for the identification of axonal pathology (Johnson et al., 2013).

Following mechanical impact, DAI is frequently neuroanatomically localized along the different portions of the corpus callosum, particularly at the level of its more anterior portion (Johnson *et al.*, 2013). While most of the brain appears structurally normal and only a minority of axons are affected even in severe TBI, DAI causes collective impairment that can lead to disruption of signal transfer and can have significant implications for cognitive function (McInnes *et al.*, 2017). Although the development of non-invasive biomarkers for DAI has become an important topic within the neuroradiological community, there are few radiologicalpathological studies that correlate histopathological findings, including DAI lesions, and MRI data following acute and chronic TBI exposure.

The microscopic nature of the axonal swelling makes DAI nearly invisible to conventional neuroimaging approaches, such as CT and conventional MRI. Their low spatial resolution, usually above 400 µm, is orders of magnitude larger than the structures of interest. Moreover, mild TBI cases present sparse clusters of affected axons with low density that may reside in different parts of the brain. Showing more promise, diffusion MRI is a technique that provides higher effective spatial sensitivity to changes that are in the order of the tissue microenvironment (below 100 µm) by using water mobility as a probe of tissue microstructure (Le Bihan et al., 1986). The most established diffusion MRI framework is diffusion tensor imaging (DTI) (Basser et al., 1994). Most notably, DTI provides the macroscopic orientationally averaged diffusivity of water in tissue, termed mean diffusivity (MD), and the diffusion anisotropy, termed fractional anisotropy (FA).

Since its first use to investigate DAI following TBI (Arfanakis et al., 2002), DTI has been established as a sensitive tool for depicting axonal integrity in regions with TBIassociated lesions (Bazarian et al., 2007; Budde and Frank, 2010; Haacke et al., 2010; Shenton et al., 2012; Hulkower et al., 2013; Skinner et al., 2017). In particular, FA had stood out as being a sensitive biomarker for TBI. Unfortunately, despite DTI's high sensitivity, conflicting findings of either decreases (Arfanakis et al., 2002; Inglese et al., 2005; Mac Donald et al., 2007; Miles et al., 2008; Kinnunen et al., 2011; Matsushita et al., 2011) or increases (Bazarian et al., 2007, 2012; Mayer et al., 2010, 2012; Henry et al., 2011; Borich et al., 2013) of the FA in TBI groups compared with control subjects have been reported. Importantly, these studies reported contradictory results with respect to FA regardless of the time from injury.

Predominantly, an additional and more fundamental obstacle for using conventional MRI in detecting tissue alterations following mild TBI is that it provides information averaged over the entire voxel. Voxel-averaged scalar images can only provide macroscopic information with respect to the voxel size (typically on the order of 1 mm<sup>3</sup>), thus limited by MRI's relatively low image resolution. In a mammalian brain, an individual voxel contains multiple chemical and physical microenvironments such as axons, neurons, glia, myelin, blood vessels, and CSF. It should be expected then, that DAI would affect a very small volume fraction of any given voxel and would most likely remain undetectable using voxel-averaged MRI methods. As a result, the inability to separate normal and pathological tissue within a voxel is the most likely cause of the non-specificity of conventional MRI methods, including DTI, in detecting and diagnosing DAI lesions.

Following recent technological breakthroughs (Benjamini and Basser, 2016; Kim et al., 2017; Topgaard, 2017; Hutter et al., 2018), multidimensional correlation MRI is currently emerging as an attractive MRI modality to study heterogeneous biological media (Benjamini, 2020; Benjamini and Basser, 2020). Overcoming the aforementioned voxel-averaging (i.e. partial volume) limitation, this approach jointly encodes several magnetic resonance contrasts (such as  $T_1$ ,  $T_2$ , and diffusion) and yields a multidimensional distribution of those parameters within each voxel, thus accomplishing two main goals: (i) providing a distribution instead of a mean, thus allowing the identification of multiple components within a voxel (Does and Gore, 2002; Ronen et al., 2006); and (ii) adding more magnetic resonance dimensions to help separate different biological components (Celik et al., 2013; Benjamini and Basser, 2017; Slator et al., 2019; Benjamini et al., 2020; Kim et al., 2020; Pas et al., 2020).

To provide a basis for future clinical translation, we aimed here to define the ways in which DAI pathology affects MRI biomarkers, and to establish whether multidimensional MRI can be used to obtain axonal injury images. To this end, we performed comprehensive APP histopathological assessments on the same specimens that were previously scanned using multidimensional and conventional MRI. We compared sections from control corpus callosum, normal-appearing corpus callosum in TBI cases, and DAI lesions in the corpus callosum, to enable the identification of subvoxel tissue components that are linked to axonal injury. Because our study focuses on TBI patients, we henceforth use the term traumatic axonal injury (TAI) to refer to DAI caused by a trauma (Sharp and Ham, 2011; Hill et al., 2016). Our hope is that non-invasive imaging and prediction of pathological APP accumulation would provide an important biomarker for DAI and means to detect it clinically.

## Materials and methods

#### **Donor specimens**

We evaluated autopsy-derived brain autopsy specimens from two different human brain collections. Formalin-fixed portions of approximately  $20 \times 20 \times 10 \text{ mm}^3$  of the corpus callosum were obtained from four civilian subjects enrolled in the Transforming Research and Clinical Knowledge in Traumatic Brain Injury study (TRACK-TBI; https://tracktbi.ucsf.edu/trans forming-research-and-clinical-knowledge-tbi) (Cases 1, 2, 4, and 5), and seven military subjects from the Center for Neuroscience and Regenerative Medicine Brain Tissue Repository (https:// www.researchbraininjury.org, Uniformed Services University of the Health Sciences, Bethesda, MD; Cases 3 and 6-11). For each case, a next-of-kin or legal representative provided a written consent for donation of the brain for use in research. The brain tissues used have undergone procedures for donation of the tissue, its storage, and use of available clinical information that have been approved by the USU Institutional Review Board (IRB) prior to the initiation of the study. All experiments were performed in accordance with current federal, state, Department of Defense, and NIH guidelines and regulations for post-mortem analysis. A detailed description of demographics for the subjects from whom brain tissue samples were obtained is listed in Table 1 and Supplementary Table 1.

#### **MRI** acquisition

Prior to MRI scanning, each formalin-fixed brain specimen was transferred to a phosphate-buffered saline (PBS) filled container for 12 days to ensure that any residual fixative was removed from the tissue. The specimen was then placed in a 25 mm tube, and immersed in perfluoropolyether (Fomblin LC/8, Solvay Solexis), a proton-free fluid void of a proton-MRI signal. Specimens were imaged using a 7 T Bruker vertical bore MRI scanner equipped with a microimaging probe and a 25 mm quadrupole RF coil.

Multidimensional data were acquired using a 3D inversion recovery diffusion-weighted sequence with a repetition time of 1000 ms, and an isotropic voxel dimension of 300  $\mu$ m. To encode the multidimensional magnetic resonance space spanned by T<sub>1</sub> and T<sub>2</sub> (i.e. T<sub>1</sub>-T<sub>2</sub>), by T<sub>1</sub> and diffusion (i.e. MD-T<sub>1</sub>), and by T<sub>2</sub> and diffusion (i.e. MD-T<sub>2</sub>), 56, 223, and 223 images were acquired, respectively, according to a previously published sampling scheme (Pas *et al.*, 2020), which resulted in respective acquisition times of 4.5, 17.8, and 17.8 h. Additional parameters of the MRI pulse sequence can be found in the Supplementary material.

A standard DTI imaging protocol was applied with the same imaging parameters as the multidimensional data and using 21 diffusion gradient directions and four b-values ranging from 0 to 1400 s/mm<sup>2</sup>, which resulted in an acquisition time of 4.5 h.

Lastly, a high-resolution MRI scan with an isotropic voxel dimension of 100  $\mu$ m was acquired using a fast low angle shot (FLASH) sequence (Matthaei *et al.*, 1985) with a flip angle of 49.6° to serve as a high resolution reference image and facilitate co-registration of histopathological and magnetic resonance images. The acquisition time was approximately 2 h.

## Histology and immunohistochemistry

After MRI scanning, each corpus callosum tissue block was transferred for histopathological processing. Tissue blocks from each brain specimen were processed using an automated tissue processor (ASP 6025, Leica Biosystems). After tissue processing, each tissue block was embedded in paraffin and cut in a series of 5-µm thick consecutive sections. The first two sections were

Table | Main demographic and histopathological findings in patients with history of TBI and healthy controls

Case	Age	Sex	Cause of death	PMI, h	Survival time	Relevant medical history	TBI clinical outcome	DAI grade <sup>b</sup>	Microsco confirme CC, APP	pically d TAI in -positive
									Extent	Severity, left/right
I	70	Male	MVA resulting in SAH and SDH	<12	3 days	Alcohol abuse, depression, diabetes	Fatal	II	Bilateral	+ + +/+
2	60	Male	MVA resulting in IPH	<12	26 days	NA	Fatal	II	Bilateral	+/+
3	33	Male	MVA resulting in IPH	30	NA	Cholecystectomy, anxiety	Fatal	1	Bilateral	+/+
4	49	Male	Fall resulting in IPH and SAH	<12	16 h	NA	Fatal	III	Bilateral	+/+
5	23	Female	MVA resulting in SDH	<12	5 days	NA	Fatal	II	Unilateral (right)	0/+
6	36	Male	Suicide resulting in IPH	<12	> I year	Blast-induced TBI, anxiety, paranoia, depression, psychosis	Non-fatal <sup>a</sup>	I	Unilateral (right)	NA/+ +
7	58	Male	Hepatic cirrhosis with ascites	<12	> I year	Impact-induced TBI, alcohol abuse, drug abuse	Non-fatal	II	Unilateral (left)	+ /0
8	32	Male	Hypoxemia and cere- bral anoxia	12	> I year	Impact-induced TBI, severe headache, hypertension.	Non-fatal	I	Unilateral (right)	0/+
9	52	Male	Sudden cardio-re- spiratory arrest	19	NA	NA	Control	NA	Absent	0/NA
10	44	Male	Sudden cardio-re- spiratory arrest	22	NA	NA	Control	NA	Absent	0/0
11	54	Male	Sudden cardio-re- spiratory arrest	29	NA	NA	Control	NA	Absent	0/0

+/++/+ + = semi-quantitative severity assessment based on APP staining; IPH = intraparenchymal hemorrhage; MVA = motor vehicle accident; NA = not available; PMI = post-mortem interval; SAH = subarachnoid haemorrhage; SDH = subdural haemorrhage.

<sup>a</sup>Non-fatal refers to the documented blast-induced TBI and not the immediate cause of death.

<sup>b</sup>Based on Adams et al. (1989).

selected for haematoxylin and eosin, and Luxol fast blue (LFB) stains, while the remaining sections were selected for immunohistochemistry procedures using a Leica Bond III automated immunostainer with a diaminobenzidine chromogen detection system (DS9800, Leica Biosystems). Immunohistochemistry was performed for anti-APP for the detection of TAI, anti-ionized calcium-binding adapter molecule 1 (Iba-1) to assess microglia reactivity, glial fibrillary acidic protein (GFAP), to evaluate presence of astrogliosis and myelin basic protein (MBP) to evaluate possible myelin loss. Two sections per antibody were stained at 300-µm apart from each other, in accordance with the MRI slice thickness. Staining procedures details can be found in the Supplementary material.

All stained sections were digitally scanned using an Aperio scanner system (Aperio AT2 - High Volume, Digital whole slide scanning scanner, Leica Biosystems, Inc.) and stored in Biolucida, a hub for 2D and 3D image data (MBF Bioscience) for further assessment and analyses. A Zeiss Imager A2 (ImagerA2 microscope, Zeiss) bright-field microscope with  $\times 40$  and  $\times 63$  magnification lenses was used to identify and photograph histological and pathological details, as needed.

### Grading of the injury

It is well recognized that both clinical and pathological aspects of TBI (i.e. closed head injury) are heterogeneous (Thompson *et al.*, 2020). We therefore used three separate classifications

and gradings to evaluate the cases examined in this study. First, the clinical outcomes in each case where a distinction was made based on whether the TBI was fatal or non-fatal. Second, the DAI grading was based on the distribution of axonal injury in the brain as proposed by Adams *et al.* (1989). Lastly, we used a semi-quantitative grading based on APP immunostaining, i.e. APP-positive DAI lesions (Blumbergs *et al.*, 1995). The extent of the DAI lesions was classified as unilateral or bilateral according to their presence in one or both hemispheres and their level of severity was semi-quantitatively assessed based on APP estimated loads as occupying <50% (+), ~50% (++) or >50% (+++) of the microscopically-examined cross-sectional section surface.

### Quantification of axonal damage

Images of APP stained sections were digitized using an Aperio whole slide scanning scanner system (Leica Biosystems) at  $\times 20$  magnification. The following steps, all implemented using MATLAB (The Mathworks, Natick, MA), were taken to allow for a quantitative analysis of the APP images. First, the images were transformed into a common, normalized space to enable improved quantitative analysis (Macenko *et al.*, 2009). Then, the normalized images were deconvolved to unmix the primary (APP) and secondary (haematoxylin and eosin) stains, and background to three separate channels (Ruifrok and Johnston, 2001). Once an APP-only image has been obtained, a final



Figure 1 Histological findings from three representative cases. Case I (A–F), Case 2 (H–M), and Case 4 (O–T). Micrographs from approximately the same regions of (A, H and O) Iba1 (microglia), (B, I and P) GFAP (astrocytes), (C, J and Q) Luxol fast blue (myelin), (D, K and R) MBP (myelin), (E, L and S) haematoxylin and eosin. [F(i), M(i) and T(i)] Original APP images, [F(ii), M(ii) and T(ii)], which are deconvolved to obtain the APP density, in F(iii), M(iii) and T(iii). Scale bar = 200  $\mu$ m in all micrographs and 5 mm in the whole-mount corpus collosum APP images in G, N and U. Case I survived 3 days post TBI and presented reactive microglia (A), astrogliosis (B), no apparent myelin loss (C and D), and TAI APP lesions (F). Case 2 survived 26 days post TBI and presented reactive microglia (H), very sparse astrogliosis (I), no apparent myelin loss (J and K), and TAI APP lesions (M). Case 4 survived 16 h post TBI and did not present any glial reactivity (O and P), no apparent myelin loss (Q and R), but did present TAI APP lesions (T).

thresholding step was taken to exclude non-specific staining and to allow for a subsequent per cent area calculation. These steps are illustrated in Fig. 1F, M and T.

From each tissue section, based on APP staining, corpus callosum TAI lesions were identified by an experienced neuropathologist (D.I.) as white matter areas with swollen axonal varicosities, axonal bulbs, or distorted axons. Accordingly, regions of interest of normal-appearing white matter and TAI lesions were manually defined. Regions of interest of normal white matter from control cases were defined as well. Additionally, grey matter regions of interest from adjacent cingulate cortex were defined in all sections. Twelve regions of interest, covering together on average 81 mm<sup>2</sup> of tissue, were identified per tissue section. Four cases and their regions of interest are shown in Supplementary Fig. 5. After extracting the regions of interest, APP density was expressed as the percentage of total area within the region of interest in the binary deconvolved APP image. In total, 132 regions of interest from 11 cases were included in this study.

#### **MRI** analysis

#### **Diffusion tensor MRI processing**

Diffusion tensor imaging parameters (Basser *et al.*, 1994) were calculated using in-house MATLAB (The Mathworks, Natick, MA) code based on previous work (Barmpoutis and Vemuri, 2010).

#### Voxel-averaged T<sub>1</sub> and T<sub>2</sub> map processing

Conventional quantitative relaxation maps were first computed by fitting the signal decay to monoexponential functions. The  $T_1$  value was computed by fitting a subset of the multidimensional data that included 20 images with inversion times in the range of 12 ms and 980 ms. The  $T_2$  value was computed by fitting a subset of the multidimensional data that included 20 images with echo times in the range of 10.5 ms and 125 ms.

#### **Multidimensional MRI processing**

Here we implemented a marginally-constrained,  $l_2$ -regularized, non-negative least square optimization to compute the multidimensional distribution in each voxel, as previously described (Benjamini and Basser, 2017; Pas *et al.*, 2020). It is a well-tested approach that had been proved robust and reliable (Provencher, 1982; Kroeker and Henkelman, 1986; Song *et al.*, 2002; Hürlimann *et al.*, 2003; Mitchell *et al.*, 2012; Celik *et al.*, 2013; Benjamini and Basser, 2018), which in this study had resulted in three types of distributions in each voxel: T<sub>1</sub>-T<sub>2</sub>, MD-T<sub>2</sub>, and MD-T<sub>1</sub>.

If one considers the multidimensional distributions as spectra, it is possible to use them to generate images of specific spectral components by means of integration over a predefined parameter range (e.g. the rectangle  $T_1$ - $T_2$  range highlighted in Fig. 2). The integral value is a number between 0 and 1, representing a certain spectral component (SC) in a given multidimensional distribution, which can be computed in each voxel to generate an image of that specific SC (Labadie *et al.*, 1994).

While the  $T_1$ - $T_2$ -MD spectra contain rich information from multiple tissue components such as myelin (Laule *et al.*, 2008), we focused here on discovering spectral substrates of axonal injury. Two factors guided us in the search: (i) we know the injury is primarily a white matter injury; and (ii) we know the  $T_1$ - $T_2$ -

MD range of what is deemed as 'normal' white matter from previous multidimensional MRI studies (and current results) (Benjamini and Basser, 2017; Pas et al., 2020). Our driving hypothesis was therefore that microscopic white matter injury would affect the normal white matter spectral component, and that the injury-associated spectral information would be found in the vicinity of the normal white matter component. Based on this notion, we have identified a  $T_1$ - $T_2$ -MD range,  $T_1 = [91.03,$ 339.32] ms,  $T_2 = [6.70, 34.85]$  ms, and MD = [0.004, 0.146] $\mu$ m<sup>2</sup>/ms, in which we observed the presence of injury-associated spectral information. We developed a spectral thresholding algorithm to account for variability between different corpus callosum samples, and to extracts the injury information in a more accurate and unsupervised way. We used an adaptive threshold in the spectral domain to detect the edge that distinguishes between the normal white matter and the axonal injury spectral components. Additional information regarding multidimensional MRI processing can be found in the Supplementary material and in Supplementary Fig. 1.

Repeatability and reproducibility of the  $T_1$ - $T_2$ , MD- $T_1$ , and MD- $T_2$  TAI SC images were evaluated by repeating the multidimensional MRI acquisitions for Cases 1 and 2. For each of these subjects, the duplicate datasets were processed using the same pipeline and then compared. Bland-Altman plots were used to quantify the agreement between all the non-zero voxels in the injury SC images from the first and second acquisitions (Bland and Altman, 1986). The standard deviation of the measurement error (in units × 100 SC) in Case 1 was 5.3 for  $T_1$ - $T_2$  injury SC, 5.5 for MD- $T_1$  injury SC, and 5.3 for MD- $T_2$  injury SC (Supplementary Fig. 2), and in Case 2 it was 4.8 for  $T_1$ - $T_2$  injury SC, 4.3 for MD- $T_1$ injury SC, and 6.5 for MD- $T_2$  injury SC (Supplementary Fig. 3).

#### Histology MRI co-registration

The high-resolution magnetic resonance images were used as anatomical references to which the histological images were registered to. Areas in the histological images that grossly diverged from the wet tissue state (i.e. the magnetic resonance images) due to deformation were manually removed, while maintaining the image aspect ratio. Following convergence of 2D affine coregistration of histology and magnetic resonance images (Image Processing Toolbox, MATLAB, The Mathworks, Natick, MA), we performed a 2D diffeomorphic registration refinement between the APP image slices and MRI volumes. This was done to recover true in-plane tissue shape and bridge over residual differences between the modalities. The diffeomorphic registration procedure in this study was performed using an efficient implementation of the greedy diffeomorphic algorithm (Joshi et al., 2004), provided as an open-source software package (https://github.com/pyushkevich/greedy). The greedy software was initialized and used as previously described (Adler et al., 2018). The transformed histology images were overlaid on magnetic resonance images to assess the quality of the coregistration.

#### Statistical analysis

Demographic variables and post-mortem interval (i.e. time between death and start of autopsy) were compared between controls, non-fatal and fatal TBI patients (per clinical outcome)



**Figure 2 Multidimensional spectra of the TAI lesions in representative cases with increasing degree of severity (left to right).** (A–E)  $T_1$ - $T_2$ , (F–J) MD- $T_2$ , and (K–O) MD- $T_1$ . The preselected  $T_1$ - $T_2$ -MD range,  $T_1$  = [91.03, 339.32] ms,  $T_2$  = [6.70, 34.85] ms, and MD = [0.004, 0.146]  $\mu$ m<sup>2</sup>/ms is highlighted as pink rectangles. Below each distribution, a magnification of the highlighted spectral region of interest is shown. The progressive shift towards shorter  $T_1$ , shorter  $T_2$ , and slower diffusivity as the severity of the injury increases is evident.

using Mann-Whitney U-test or unpaired two-tailed Student's *t*-test.

Linear mixed-effects models were used to compare MRIderived and histopathological measures. We fitted a linear mixed-effects model using three predictors: region of interest (control corpus callosum, normal-appearing white matter in mild TAI corpus callosum, normal-appearing white matter in severe TAI corpus callosum, mild TAI lesions, and severe TAI lesions), clinical outcome (control, fatal and non-fatal TBI), and subject (i.e. case number). To allow unbiased statistical comparisons, the voxel-averaged MRI parameters were adjusted across all tissue samples by dividing them by the mean of each parameter in each specific sample to correct for possible between-sample differences arising from post-mortem effects. We fitted the model for each of the measured quantities (magnetic resonance variables and APP density), with fixed-effects for region of interest, and potentially correlated random-effects for intercept and region of interest grouped by the clinical outcome, and for intercept and region of interest grouped by the case number. The random-effects were selected to account for MRI differences that might exist due to clinical outcome, patient-specific variations, and possible within-subject correlation among measures. False discovery rate (FDR) correction was carried out to take the overall number of pairwise contrasts into account.

The potential correlation between all conventional voxel-averaged and multidimensional MRI measures and APP density in the different tissue types was investigated using linear regression, with Pearson's correlation coefficient, r, reported.

A *P*-value of 0.05 was considered statistically significant. MATLAB was used for the computation.

#### **Data availability**

The datasets generated and analysed during the current study are available from the corresponding author on reasonable request.

## Results

## Traumatic axonal injury pathology in the corpus callosum

Table 1 and Supplementary Table 1 summarize the main demographic data for each examined brain and histopathological findings observed in each studied corpus callosum. No significant differences were found for age and post-mortem interval between fatal and non-fatal TBI, and controls. No significant differences were found for APP-based TAI grade, APP-based TAI extent and APP-based severity between the fatal and non-fatal TBI cases (Supplementary Table 1).

The post-injury survival time in all fatal TBI cases was sufficiently long to allow for pathological APP accumulation (Sherriff *et al.*, 1994). Except for Case 6, subjects who died of causes unrelated to TBI had documented head trauma. The subject described in Case 6 sustained a fatal head trauma that resulted in immediate death, which would likely have prevented significant pathological accumulation of APP. Considering that APP is the histopathological marker we evaluate in this study, and because of their documented blast-induced TBI, we classified Case 6 as non-fatal TBI. It is noteworthy that classification as fatal or non-fatal TBI is important for understanding the clinical context; however, it does not affect the MRI findings and determinations in this study, which are made regardless of the cause of death.

Figure 1 shows simultaneous views from three representative TBI cases immunostained for markers of microglia (Iba-1), astrocytes (GFAP), myelin (MBP), and TAI lesions (APP), histology stains for myelin (LFB) and general histological assessment (haematoxylin and eosin), and the deconvolved APP images along with the obtained APP densities.

Levels of myelin density as assessed by both MBP immunohistochemistry and LFB stain did not show any marked loss across all considered regions (e.g. Fig. 1C, D, J, K, Q and R). By contrast, microglia activation and reactivity varied across all cases (e.g. Fig. 1A, H and O). Likewise, astroglial reactivity was variable as well (e.g. Fig. 1B, I and P). Astroglial and microglial cells reactivities depend on various biological timing factors, most notably, time interval differences between the TBI exposure and autopsy (Ramlackhansingh *et al.*, 2011), which, in our study, varied greatly between the fatal and non-fatal TBI groups.

Additionally, apart from Case 1, haematoxylin and eosinbased assessment of each corpus callosum did not reveal evidence of ischaemic-necrotic lesions, presence of vascular lesions, microhaemorrhages, or tissue rarefaction.

One hundred and thirty-two tissue regions of interest were selected based on the APP density images and blind to MRI, consisting of four APP-positive regions from each TAI case (total of 32), four normal-appearing white matter regions from each TAI case (total of 32), eight white matter regions from the control cases (total of 24), and four cortical grey matter regions from all cases (total of 44). In total, 132 regions of interest from 11 cases were included in this study.

## Transverse and longitudinal relaxations shorten with traumatic axonal injury severity

We observed a consistent and gradual change in the way in which  $T_1$ ,  $T_2$ , and MD were distributed as the APP-based severity of the TAI was increasing. Figure 2 shows five cases with increased injury severity and their respective  $T_1$ - $T_2$ (Fig. 2A–E), MD- $T_2$  (Fig. 2F–J), and MD- $T_1$  (Fig. 2K–O) distributions averaged from 20 voxels within control white matter and TAI lesions. The shift in the distributions can be illustrated by examining the most affected  $T_1$ - $T_2$ -MD ranges, which are highlighted as pink rectangles. A magnification of these ranges in the spectra are shown below each distribution. Qualitatively, the magnified regions show a gradual shortening of  $T_1$  and  $T_2$  as the severity of the injury is increased (Fig. 2, left to right). A shift towards lower MD values is also noticeable, although not as consistent as changes in the relaxation distributions.

## Multidimensional and voxelaveraged MRIs of traumatic axonal injury

Based on the control and TAI cases we examined here, we have identified a  $T_1$ - $T_2$ -MD range, ( $T_1 = [91.03, 339.32]$  ms,  $T_2 = [6.70, 34.85]$  ms, and MD =  $[0.004, 0.146] \mu m^2/$  ms) in which we assessed most of the spectral information regarding the injury resides. Integration according to the spectral signature within this range, as detailed in the 'Materials and methods' section, was performed, resulting in MD- $T_1$ , MD- $T_2$ , and  $T_1$ - $T_2$  injury SC images.

Figure 3 shows conventional and multidimensional magnetic resonance and histological images of three representative cases: control, non-fatal and fatal TBI (Cases 10, 6, and 2, respectively). Histological images of the control case show negative APP staining (Fig. 3A), compared with positive APP staining in the injured samples (Fig. 3D and G). Conventional, voxel-averaged relaxation and diffusion images of the control, non-fatal and fatal cases, are shown in Fig. 3B, E and H, respectively. Lastly, the multidimensional injury images of the control, non-fatal and fatal cases, are shown in Fig. 3C, F and I, respectively. Note that to facilitate visualization, the injury SC images were thresholded at 10% of the maximal intensity and overlaid on grayscale proton density images. More multidimensional injury images are shown in Supplementary Fig. 4.

# Histopathological, multidimensional and conventional MRI findings

We grouped the 132 tissue regions into controls, normalappearing white matter in TBI patients, and TAI lesions in TBI patients. We further partitioned the TBI patients into mild and severe TAI according to the APP staining-based severity in Table 1 to assess whether the examined MRI metrics are able to identify TAI with different degrees of APPbased severity (Cases 1–4 and 6 were grouped as severe TAI and Cases 5, 7 and 8 as mild TAI). We also applied a commonly used strategy (Holleran *et al.*, 2017; Cooper *et al.*, 2019) to correct for possible between-subject differences arising from post-mortem effects; we adjusted each voxelaveraged MRI parameter by dividing them by the mean for that parameter across all white matter (in control subjects) and normal-appearing white matter (in subjects with TBI) voxels in each brain sample.

Compared to controls' white matter, mild TAI lesions showed increased  $T_1$ - $T_2$  injury SC (FDR-corrected *P*-value,  $P_{\rm FDR} = 0.005$ ), increased MD- $T_2$  injury SC ( $P_{\rm FDR} = 0.005$ ), and increased per cent area APP ( $P_{\rm FDR} < 0.001$ ) (Fig. 4 and Supplementary Table 2).

Compared to controls white matter, severe TAI lesions showed increased  $T_1$ - $T_2$  injury SC ( $P_{FDR} < 0.001$ ), increased MD- $T_1$  injury SC ( $P_{FDR} = 0.023$ ), increased MD- $T_2$  injury SC ( $P_{FDR} < 0.001$ ), and increased per cent area APP ( $P_{FDR} < 0.001$ ) (Fig. 4 and Supplementary Table 2).

Compared to normal-appearing white matter tissue regions in TBI patients with mild TAI, mild TAI lesions showed increased  $T_1$ - $T_2$  injury SC ( $P_{FDR} = 0.003$ ), increased MD- $T_2$  injury SC ( $P_{FDR} = 0.022$ ), and increased per cent area APP ( $P_{FDR} < 0.001$ ) (Table 2 and Fig. 4).

Compared to normal-appearing white matter tissue regions in TBI patients with severe TAI, severe TAI lesions showed increased  $T_1$ - $T_2$  injury SC ( $P_{FDR} < 0.001$ ), increased MD- $T_1$  injury SC ( $P_{FDR} = 0.036$ ), increased MD- $T_2$  injury SC ( $P_{FDR} < 0.001$ ), and increased per cent area APP ( $P_{FDR} < 0.001$ ) (Table 2 and Fig. 4).

The MD-T<sub>1</sub> and MD-T<sub>2</sub> injury biomarkers were the only ones that were able to distinguish between mild TAI and severe TAI lesions (P = 0.008 and P = 0.015, respectively); however, they did not survived correction for multiple comparisons.

None of the other MRI metrics showed significant between-group differences (Table 2, Fig. 4 and Supplementary Table 2).

## Correlation between MRI measures and APP density

To determine whether MRI could potentially detect TAI in TBI patients, we performed radiological–pathological correlation analyses with histological APP density and all the investigated MRI parameters.

Figure 5 summarizes the association between the investigated magnetic resonance metrics—conventional voxel-averaged and multidimensional images—and the pathological findings in normal white matter, cortical grey matter, and TAI tissue blocks.

To assess the relationship of the MRI parameters with the degree of injury, all tissue regions of interest were grouped together and correlated with the APP density (Fig. 5, solid lines). We found that APP density was strongly and significantly correlated with the  $T_1$ - $T_2$  injury SC (r = 0.876, P < 0.001), the MD- $T_1$  injury SC (r = 0.727, P < 0.001), and the MD- $T_2$  injury SC (r = 0.743, P < 0.001). Importantly, these results indicated that higher intensity of the multidimensional injury biomarkers is associated with increased DAI severity, regardless of the tissue type studied. From the conventional voxel-averaged images, we found that APP density had very weak yet significant correlations with adjusted FA (r = 0.166, P = 0.032), with adjusted  $T_1$  (r = -0.241, P = 0.003) and with adjusted  $T_2$  (r = -0.287, P < 0.001).

To assess the capability of the MRI parameters to differentiate between normal and injured white matter with respect to APP density, normal white matter and TAI regions of interest were grouped together (Fig. 5, dashed lines). We found that the correlations of multidimensional injury biomarker images with APP density have barely changed by excluding the cortical grey matter regions of interest. Conversely, this grouping resulted in insignificant voxelaveraged MRI metrics correlations with respect to APP density, showing that none of these variables can be used to distinguish lesion and non-lesion white matter tissue.



**Figure 3 Multidimensional and voxel-averaged MRIs of TAI. (A–C)** Control brain (Case 10). (**A**) APP histological images co-registered to MRIs. Deconvolved histological image: red = APP stain. Negligible APP density was detected. (**B**) Conventional MRI maps of T<sub>1</sub>, T<sub>2</sub>, FA and MD do not show visible abnormalities. (**C**) Multidimensional injury maps overlaid onto proton density images show absent of significant injury. (**D–F**) Non-fatal TBI brain (Case 6). (**D**) APP histological images co-registered to MRIs show visible TAI lesions in the corpus callosum. (**E**) Conventional MRI maps of T<sub>1</sub>, T<sub>2</sub>, FA and MD do not show visible abnormalities in the corpus callosum. (**F**) Multidimensional injury maps overlaid onto proton density images show significant injury in white matter (in particular, the T<sub>1</sub>-T<sub>2</sub> injury SC). (**G–I**) Fatal TBI brain (Case 2). (**G**) APP histological images co-registered to MRIs in regions of white matter/grey matter interface (white arrows). (**H**) Conventional MRI maps of T<sub>1</sub>, T<sub>2</sub>, FA and MD do not show visible abnormalities in white matter/grey matter interface. (**I**) Multidimensional injury maps overlaid onto proton density images show substantial injury along the white matter/grey matter interface.



Figure 4 Between-group comparisons of voxel-averaged and multidimensional biomarkers and histopathological measures. Box plots showing between-group differences among control corpus callosum, normal-appearing white matter in mild TAI corpus callosum, mild TAI lesions, normal-appearing white matter in severe TAI corpus callosum, and severe TAI lesions. (A)  $T_1$ - $T_2$  injury SC; (B) MD- $T_1$  injury SC; (C) MD- $T_2$  injury SC; (D) adjusted voxel-averaged FA; (E) adjusted voxel-averaged  $T_1$ ; and (F) adjusted voxel-averaged  $T_2$ . All parameters are averaged across their corresponding spatial regions of interest.

To assess the capability of the MRI parameters to differentiate between grey matter and injured white matter with respect to APP density, grey matter and TAI regions of interest were grouped together (Fig. 5, dotted lines). Here too, we found that the correlations of multidimensional injury biomarker images with APP density have barely changed by excluding the normal white matter regions of interest. All of the voxel-averaged MRI parameters were significantly correlated with APP density (adjusted FA: r = 0.379, P < 0.001; adjusted MD: r = -0.356, P < 0.001; adjusted T<sub>1</sub>: r = -0.375, P < 0.001; adjusted T<sub>2</sub>: r = -0.468, P < 0.001), showing that they are able to distinguish between grey matter and axonal injury. However, given their inability to differentiate normal white matter and axonal injury, these findings merely show that voxel-averaged parameters can distinguish grey matter and white matter.

## Discussion

This study crucially identified potential imaging biomarkers of DAI pathology from the joint analysis of multidimensional MRI and histopathological data in the corpus callosum. The ability to selectively extract a specific  $T_1$ - $T_2$ -MD spectral range, or 'spectral signature', and provide its corresponding image, allows multidimensional MRI to achieve good separation between subjects and microscopic lesion and non-lesion regions. We found that the multidimensional MRI axonal injury images are significantly and strongly correlated with histological evidence of DAI. Furthermore, these non-invasive DAI biomarkers produce negligible intensities in grey matter and in normal-appearing white matter.

Multidimensional MRI allows us to capture the strong association of T<sub>1</sub> and T<sub>2</sub> shortening and the severity of the microscopic axonal injury. Proteins, which have very short T<sub>2</sub> values, interact with free and partially bound water, reducing both T<sub>1</sub> and T<sub>2</sub> (Edzes and Samulski, 1977; Zhang et al., 2007; Filo et al., 2019). Axonal injury causes microtubule disruption that induces long-term pathological co-accumulation of APP and other intra-axonal proteins (Chen et al., 2004), and to the formation of axonal varicosities (Fig. 6). While non-invasive and direct detection of proteins may not be feasible with adequate spatial resolution, we suggest that these protein aggregates could act as endogenous relaxation enhancers of the surrounding water, which can be captured by observed shortened  $T_1$ - $T_2$  (Fig. 6). However, quantitative biophysical modelling and further detailed experiments will be required to confirm or refute this hypothesis.

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	ariable N/ in i	AWM mTAI = 12	m <b>TAI</b> lesions n = 12	NAWM in sTAI n = 20	sTAI lesions n = 20	mTAI lesions versus NAWM in mTAI		sTAI lesions versus NAWM in sTAI		sTAI lesions versus mTAI lesions	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Ξ.	ean (SE)	Mean (SE)	Mean (SE)	Mean (SE)	Estimated mean difference (95% CI)	P/P <sub>FDR</sub>	Estimated mean difference (95% Cl)	P/P <sub>FDR</sub>	Estimated mean difference (95% Cl)	P/P <sub>FDR</sub>
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	I-T <sub>2</sub> SC, × 100 1.0	09 (0.39)	4.09 (0.97)	0.67 (0.29)	4.70 (1.03)	3.00 (0.95, 5.05)	< 0.001/0.003	4.03 (1.93, 6.13)	< 0.00   / < 0.00	0.74 (1.93, 3.41)	0.667/0.943
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	<sup>1</sup> D-T <sub>1</sub> SC, × 100 0.5	56 (0.29)	0.41 (0.20)	0.46 (0.19)	2.20 (0.64)	-0.15 (-0.84, 0.54)	0.707/0.793	1.74 (0.43, 3.05)	0.018/0.036	1.57 (0.24, 2.90)	090.0/800.060
Adjusted FA 1.00 (0.07) 0.94 (0.23) 1.00 (0.05) 0.96 (0.15) -0.06 (-0.54, 0.41) 0.7793/0.793 -0.04 (-0.36, 0.28) 0.790/0.790 0   Adjusted MD 1.00 (0.06) 1.04 (0.06) 1.00 (0.05) 1.13 (0.11) 0.04 (-0.13, 0.21) 0.6575/0.793 0.13 (-0.09, 0.36) 0.261/0.418 -0   Adjusted T1 1.00 (0.03) 0.97 (0.03) 1.00 (0.02) 0.95 (0.06) -0.03 (-0.11, 0.05) 0.564/0.793 0.13 (-0.09) 0.523/0.597 -0   Adjusted T2 1.00 (0.04) 0.91 (0.04) 1.00 (0.02) 0.94 (0.08) -0.03 (-0.11, 0.05) 0.126/0.793 0.13 (-0.17, 0.09) 0.523/0.597 -0   Adjusted T2 1.00 (0.04) 0.91 (0.04) 1.00 (0.03) 0.94 (0.08) -0.09 (-0.20, 0.02) 0.126/0.251 -0.06 (-0.23, 0.11) 0.497/0.597 0   % area APP 0.40 (1.99) 20.67 (3.71) 0.97 (1.5.4) 26.65 (6.35) 20.27 (12.02, 28.52) <0.001 / < 0.001	<sup>1</sup> D-T <sub>2</sub> SC, × 100 0.2	22 (0.24)	1.14 (0.24)	0.19 (0.19)	2.84 (0.65)	0.92 (0.25, 1.58)	0.008/0.022	2.65 (1.32, 3.98)	< 0.00   / < 0.00	1.72 (0.36, 3.08)	0.015/0.060
Adjusted MD 1.00 (0.06) 1.04 (0.06) 1.00 (0.05) 1.13 (0.11) 0.04 (-0.13, 0.21) 0.675/0.793 0.13 (-0.09, 0.36) 0.261/0.418 -0   Adjusted T1 1.00 (0.03) 0.97 (0.03) 1.00 (0.02) 0.95 (0.06) -0.03 (-0.11, 0.05) 0.504/0.793 -0.04 (-0.17, 0.09) 0.523/0.597 -0   Adjusted T2 1.00 (0.04) 0.91 (0.03) 0.94 (0.08) -0.09 (-0.20, 0.02) 0.94 (0.08) -0.09 (-0.20, 0.02) 0.497/0.597 -0   Adjusted T2 1.00 (0.04) 0.91 (0.03) 0.94 (0.08) -0.09 (-0.20, 0.02) 0.126/0.251 -0.06 (-0.23, 0.11) 0.497/0.597 0   % area APP 0.40 (1.99) 20.67 (3.71) 0.97 (1.54) 26.65 (6.35) 20.27 (12.02, 28.52) <0.001 / < 0.001	djusted FA I.C	00 (0.07)	0.94 (0.23)	1.00 (0.05)	0.96 (0.15)	-0.06 (-0.54, 0.41)	0.793/0.793	-0.04 (-0.36, 0.28)	0.790/0.790	0.02 (-0.53, 0.57)	0.943/0.943
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% area APP 0.40 (1.99) 20.67 (3.71) 0.97 (1.54) 26.65 (6.35) 20.27 (12.02, 28.52) < 0.001 / < 0.001 25.68 (12.87, 38.49) < 0.001 / < 0.001 5	djusted T <sub>2</sub> 1.0	00 (0.04)	0.91 (0.04)	1.00 (0.03)	0.94 (0.08)	-0.09 (-0.20, 0.02)	0.126/0.251	-0.06 (-0.23, 0.11)	0.497/0.597	0.03 (-0.15, 0.21)	0.730/0.943
	area APP 0.4	40 (1.99)	20.67 (3.71)	0.97 (1.54)	26.65 (6.35)	20.27 (12.02, 28.52)	< 0.001/< 0.001	25.68 (12.87, 38.49)	< 0.00   / < 0.00	5.96 (–8.45, 20.37)	0.419/0.903

among mild TAL severe TAL NAWM in mild and severe TAL tissue regions Table 2 Comparisons of MRI parameters and pathological measures

Robust quantitative radiological-pathological correlations allowed us to visualize, within each TBI patient, regions with substantial TAI lesions and regions that were not affected by trauma. We showed that voxel-averaged MRI approaches were unable to detect any differences between controls, normal-appearing white matter in TBI subjects, and TAI lesions (Fig. 4, Table 2 and Supplementary Table 2). The main reason for this insensitivity is that using these approaches one cannot selectively extract the injury-affected spectral range per subject because these images contain only scalar averages, not distributions. Conversely, multidimensional MRI parameters, primarily T1-T2 and MD-T2, were able to distinguish between lesions and normal-appearing white matter regions within TBI patients (Table 2 and Supplementary Table 2). Replacing the conventionally obtained voxel-averaged scalar value with a distribution increases the sensitivity to detect subtle changes, that would

Consistent with previous literature (Johnson *et al.*, 2013), we found that the corpus callosum was affected by DAI following trauma. Apart from Case 1, none of the corpus callosa we evaluated showed macroscopic damage, cytotoxic oedema, or large (>1 mm) lesions. No myelin loss was observed in all the examined cases, and glial cells reactivity varied greatly between cases, most likely due to variability in the survival time. These histopathological findings suggest that the damage from TAI in the cases we examined was microscopically limited, and in most cases did not result in observable macroscopic loss.

have been otherwise averaged-out.

Remarkably, there have been only a few quantitative MRI studies of TBI that combined relaxometry and diffusion with histological correlation. Our findings of decreased  $T_1$  and  $T_2$ are in contrast with previous animal models studies that reported an increase in T<sub>2</sub> following TBI. The increase in T<sub>2</sub> in those studies was associated with oedema (Ducreux et al., 2005; Obenaus et al., 2007; Immonen et al., 2009; Li et al., 2016), necrosis (Mac Donald et al., 2007), and haemorrhage (Long et al., 2015). In animal models of TBI, where moderate to severe injury is prevalent, macroscopic damage that causes oedema and haemorrhage may dominate the MRI signal. Most of the cases in our study had a survival time longer than 14 days, which is typically the time it takes oedema to resolve. Of the three acute cases with survival time under 7 days (Cases 1, 4, and 5), oedema and loss of tissue were observed only in Case 1 (care was taken such that none of the analysed tissue blocks were selected from that area).

Although DTI metrics are considered to be sensitive to microstructural alterations due to TBI (Mac Donald *et al.*, 2007; Holleran *et al.*, 2017), it is becoming increasingly evident that they are inconsistent in their observed response (Budde and Annese, 2013; Kamnaksh *et al.*, 2015; Dennis *et al.*, 2018). Reflecting this heterogeneity, voxel-averaged FA and MD in our study presented a large variability in all groups that had led to insignificant changes due to injury.

Common to all *ex vivo* human MRI studies, our data include the effects of post-mortem degeneration, fixation and



**Figure 5 APP density.** APP density (% area) from 132 tissue regions, consisting of 4 APP-positive regions from each TAI case (total of 32, blue dots), four to six normal-appearing white matter regions from all cases (total of 56, red dots), and four cortical grey matter regions from all cases (total of 44, yellow dots), and the corresponding magnetic resonance parameter correlations. Individual data-points represent the mean region of interest value from each post-mortem tissue sample. Scatterplots of the mean (with 95% confidence interval error bars) % area APP and (**A**)  $T_1$ - $T_2$ , (**B**) MD- $T_1$ , and (**C**) MD- $T_2$  injury SCs show positive and significant correlation with APP density. Both  $T_1$ - $T_2$  and MD- $T_2$  injury SCs demonstrated specificity exclusively towards TAI (all grey matter and normal-appearing white matter had negligible intensities). The conventional MRI metrics (**D**–**G**) did not result in strong and significant correlations with % area APP.

resulting dehydration. First, the fixation process by itself changes the tissue properties, such that a direct comparison with *in vivo* data is not possible. In the case of human tissue, delayed fixation (i.e. post-mortem interval) affects the measured MRI parameters as well (Shepherd *et al.*, 2009; Roebroeck *et al.*, 2019), which further complicates comparison with *in vivo* human results. The post-mortem interval in this study was reasonably homogeneous, with 7 of 11 cases  $\leq 12$  h, and the rest  $\leq 30$  h. Moreover, histopathology results indicate preservation of APP and other immunostains in all samples, regardless of post-mortem interval. Although this study was designed to include both fatal and non-fatal TBI outcomes, this design results in inherently heterogeneous

survival time, which presents a limitation. Glial reactivity, which is highly dependent on the time from injury (Ramlackhansingh *et al.*, 2011), cannot be controlled for in our study, and cannot be ruled out as a contributing factor to our MRI findings. Future studies, in which the whole brain is imaged and processed, may yield additional information regarding TAI localization and severity in the brain following different types of trauma.

Here, we would like to emphasize the  $T_1$ - $T_2$  MRI sequence results because of its potential ease of migration towards clinical applications. Although technological progress is being made (Wu *et al.*, 2016; Hutter *et al.*, 2018), substantial gaps remain between preclinical MRI, like the



**Figure 6 Proposed explanation of T<sub>1</sub> and T<sub>2</sub> shortening in TAI lesions.** Schematic representation of (**A**) healthy neuron and (**B**) traumatized neuron. In healthy neurons, APP and other proteins transported through axons via microtubules by fast axonal transport without reaching detectable levels. In traumatized neurons, microtubules undergo mechanical failure resulting in transport disruption that leads to accumulation of APP and possibility other proteins, which causes local axonal swelling. As known T<sub>1</sub> and T<sub>2</sub> relaxation enhancers, proteins accumulate and affect the surrounding water by shortening their observed T<sub>1</sub> and T<sub>2</sub> values.

one used here, and clinical MRI systems, and achieving satisfactory diffusion encoding in the latter is a particularly challenging task (Mukherjee *et al.*, 2008; Jones *et al.*, 2018). Conversely,  $T_1$ - $T_2$ -weighted acquisition is more clinically feasible (Deoni *et al.*, 2008; Shepherd *et al.*, 2020), especially considering the relatively small number of  $T_1$ - $T_2$ -weighted images that were required in our study, i.e. 56. Here we used roughly half of the amount of data that has been previously used in the human brain *in vivo* (Kim *et al.*, 2020), which would have reduced the acquisition time of a singleslice dataset from 20 to 10 min. Because  $T_1$ ,  $T_2$ , and diffusion dynamics are different in fixed tissue compared with living systems, further research will be needed to establish whether and how the  $T_1$ - $T_2$ -MD range of multidimensional magnetic resonance parameters is altered *in vivo*.

A non-invasive method to detect DAI pathology with high specificity and in vivo would be a major medical contribution. Here, we have shown that DAI due to trauma has a multidimensional spectral signature, which can be used to generate injury biomarker images that closely follow APP histopathology: axonal injury image intensity scales with pathological APP accumulation, and importantly, lack of APP corresponds to negligible intensity. Our results indicate that the  $T_1$ - $T_2$ -MD range we identified is directly linked to microscopic tissue alterations due to DAI. We believe that the improved sensitivity of these novel injury biomarkers towards DAI is advancing the neuroimaging field closer towards non-invasive quantitative 'histology'. This neuroimaging tool provides an injury-only image that may help clinicians and clinical investigators to detect and visualize microscopic DAI lesions in the brain. As evidence that indicates DAI is a likely pathological substrate for concussion is mounting (Smith and Stewart, 2020), the importance of developing a means to detect it clinically becomes clear.

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# **Competing interests**

The authors report no competing interests.

# Supplementary material

Supplementary material is available at Brain online.

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