23(4), 542–556, 2021 | doi:10.1093/neuonc/noaa283 | Advance Access date 18 December 2020

# **Mechanisms of imipridones in targeting mitochondrial metabolism in cancer cells**

#### **Erin R. Bonner, Sebastian M. Waszak, Michael A. Grotzer, Sabine Mueller, and Javad Nazarian**

*Center for Genetic Medicine, Children's National Health System, Washington, DC (E.R.B, J.N.); Institute for Biomedical Sciences, The George Washington University School of Medicine and Health Sciences, Washington, DC, (E.R.B., J.N.); Centre for Molecular Medicine Norway (NCMM), Nordic EMBL Partnership, University of Oslo and Oslo University Hospital, Oslo, Norway (S.M.W.); Department of Pediatric Research, Division of Paediatric and Adolescent Medicine, Rikshospitalet, Oslo University Hospital, Oslo, Norway (S.M.W.); Department of Oncology, University Children's Hospital Zürich, Zürich, Switzerland (M.A.G., S.M., J.N.); Department of Neurology, Neurosurgery and Pediatrics, University of California San Francisco, San Francisco, California (S.M.)*

**Corresponding Author:** Javad Nazarian, PhD, Department of Oncology, Children's Research Center, University Children's Hospital Zürich, Balgrist Campus, Lengghalde 5, 8008 Zürich, Switzerland [\(Javad.Nazarian@kispi.uzh.ch](mailto:Javad.Nazarian@kispi.uzh.ch?subject=)).

#### **Abstract**

ONC201 is the first member of the imipridone family of anticancer drugs to enter the clinic for the treatment of diverse solid and hematologic cancers. A subset of pediatric and adult patients with highly aggressive brain tumors has shown remarkable clinical responses to ONC201, and recently, the more potent derivative ONC206 entered clinical trials as a single agent for the treatment of central nervous system (CNS) cancers. Despite the emerging clinical interest in the utility of imipridones, their exact molecular mechanisms are not fully described. In fact, the existing literature points to multiple pathways (e.g. tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) signaling, dopamine receptor antagonism, and mitochondrial metabolism) as putative drug targets. We have performed a comprehensive literature review and highlighted mitochondrial metabolism as the major target of imipridones. In support of this, we performed a meta-analysis of an ONC201 screen across 539 human cancer cell lines and showed that the mitochondrial caseinolytic protease proteolytic subunit (ClpP) is the most significant predictive biomarker of response to treatment. Herein, we summarize the main findings on the anticancer mechanisms of this potent class of drugs, provide clarity on their role, and identify clinically relevant predictive biomarkers of response.

#### **Keywords**

#### ClpP | imipridone | ONC201 | ONC206 | ONC212

The anticancer imipridone drug family has emerged as promising candidates for treating a diverse range of solid and he-matologic cancers.<sup>[1](#page-12-0)[,2](#page-12-1)</sup> ONC201, the parent imipridone ([Table 1\)](#page-1-0), exhibits cytotoxicity across a spectrum of preclinical cancer models and has entered phase 1 and 2 clinical trials for treating patients diagnosed with cancers including leukemia, lymphomas, colon, prostate, breast, and central nervous system (CNS) tumors [\(Table 2\)](#page-3-0). Clinically, ONC201 has demonstrated a favorable safety profile and encouraging performance at prolonging patient survival, even in patients with advanced treatment-refractory solid tumors.<sup>3</sup> Moreover, ONC201 demonstrates CNS tumor penetration and encouraging response rates

in a subset of both adult and pediatric brain cancer patients, $4-7$  $4-7$ including children with H3K27M-mutant diffuse midline glioma (DMG)[,6](#page-12-5),[7](#page-12-4) warranting further clinical study. These positive clinical observations have catalyzed the synthesis of imipridone derivatives that share ONC201's core chemical structure but harbor modifications conferring enhanced potency and signaling capabilities in preclinical models $8-12$  ([Table 1](#page-1-0)). However, the precise anticancer mechanisms of imipridones remain elusive. These drugs may selectively antagonize G protein-coupled receptor (GPCR) proteins, most notably the D(2) dopamine receptor (D2R), widely expressed across cancer cells.<sup>13</sup> In addition, imipridones may induce tumor necrosis factor (TNF)-related

© The Author(s) 2020. Published by Oxford University Press on behalf of the Society for Neuro-Oncology. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com

ì

<span id="page-1-0"></span>



<span id="page-3-0"></span>





cancer; AML, acute myeloid leukemia; HGG, high grade glioma; GBM, glioblastoma multiforme; CNS, central nervous system. aThe table summarizes the current state of clinical trials involving the lead imipridones ONC201 (*n* = 19 clinical trials) and ONC206 (n = 1). Included are trials that are recruiting (R, *n* = 13), temporarily not available (TNA) (*n* = 1), completed (C, *n* = 2), not yet recruiting (NYR, *n* = 2), or active and not recruiting (ANR,  $n = 2$ ), as registered on Clinicaltrials.gov. Trials are arranged by cancer type, and the trial phase, status, study start/estimated completion dates, and patient criteria are listed.

apoptosis-inducing ligand (TRAIL) signaling, leading to the extrinsic pathway of cell death.<sup>[1](#page-12-0)[,2](#page-12-1)</sup> However, emerging data indicate that imipridones' primary mechanism is to potently activate the mitochondrial ATP-dependent caseinolytic protease (Clp)proteolytic subunit (ClpP), leading to enhanced degradation of mitochondrial proteins and impaired tumor cell metabolism.<sup>[11](#page-13-3),14</sup> This observation has reduced support for ONC201's drug interaction with D2R and has emphasized instead its potential role in targeting disease bioenergetics. In this Review, we focus on the molecular mechanisms of the most well-characterized imipridones ONC201, ONC206 and ONC212. We provide an overview of imipridones' putative biological targets, highlight data from clinical trials supporting the therapeutic efficacy of these drugs, and note combinatorial approaches in preclinical studies that hold promise for future clinical translation. Finally, we discuss outstanding questions on the molecular mechanisms of imipridones. The encouraging preclinical and clinical findings, which have catalyzed a surge of new clinical trials using ONC201 and the first-in-human ONC206 trial, merit a timely synthesis of the existing literature on the anticancer activity of this novel drug family.

## **Imipridones and cancer**

ONC201 was discovered as an anticancer agent in 2013 in a bioluminescence reporter screen for inducers of TRAIL,<sup>[1](#page-12-0)</sup> and was thus named "TRAIL-inducing compound 10" (TIC10)[.1](#page-12-0) ONC201 achieves sustained upregulation of TRAIL across a multitude of human cancer cell lines and induces apoptosis in malignant cells without affecting healthy cells.<sup>1,[2](#page-12-1)</sup> ONC201's structure was first reported incorrectly as imidazo[1,2-a]pyrido[4,3-d]pyrimidine derivative based on

mass spectrometry<sup>1</sup> but was soon corrected to be an angular [3,4-e], rather than linear [4,3-d], isomer,<sup>18</sup> confirmed by NMR and X-ray structural analysis.<sup>19</sup> The angular struc-ture seems to be highly essential for ONC201's activity.<sup>[18](#page-13-7),[19](#page-13-8)</sup>

## **Antagonism of G-protein coupled receptors**

Imipridones selectively antagonize the D(2) dopamine receptor (D2R, encoded by *DRD2*),[13](#page-13-1),[15](#page-13-4) a GPCR that is widely expressed across cancers[.13,](#page-13-1)[20](#page-13-9),[21](#page-13-10) *DRD2* expression correlates with ONC201 response in cancers including glio-blastomas.<sup>[5](#page-12-7)[,13,](#page-13-1)20</sup> However, this receptor is not essential for ONC201 sensitivity: CRISPR/Cas9-mediated knockout of *DRD2* in colorectal and breast cancer cell lines does not abrogate ONC201 response.<sup>13</sup> Modified derivatives of ONC201 may have additional GPCR targets beyond D2R; for example, ONC212 selectively activates the GPCR GPR132, which is highly expressed in leukemias and lymphomas, and in turn activates Gαq signaling in acute myeloid leukemia (AML) preclinical models.<sup>12</sup> A list of imipridones and their putative GPCR targets is provided in [Table 1.](#page-1-0)

## **Induction of TRAIL-mediated apoptosis**

ONC201 is thought to induce TRAIL signaling by reducing levels of phosphorylated protein kinase B (Akt) and extracellular signal regulated kinase (ERK) and, in turn, phos-phorylation of their target transcription factor Foxo3a.<sup>[1](#page-12-0)</sup> Through the proposed pathway, ONC201 treatment leads to dephosphorylation and subsequent translocation of

Foxo3a to the nucleus, where Foxo3a binds the TRAIL promoter to induce p53-independent TRAIL expression.<sup>[1](#page-12-0)</sup> TRAIL protein then localizes to the cell membrane and binds to death receptor 4 (DR4) or 5 (DR5), activating the extrinsic pathway of apoptosis involving the initiator caspase-8[.22](#page-13-11) Indeed, ONC201 induces TRAIL upregulation in vitro across cancer cell lines including  $CNS$ ,<sup>[1](#page-12-0)</sup> lung,<sup>1,[23](#page-13-12)</sup> breast, ovarian, prostate,<sup>1,[2](#page-12-1)</sup> colorectal,<sup>[24](#page-13-13)</sup> and lymphoma<sup>[25](#page-13-14)</sup> cells, without affecting healthy fibroblasts.<sup>1</sup> In vivo, ONC201 upregulates TRAIL and induces tumor regression in colon and triple-negative breast cancer (TNBC) models, and crosses the intact blood-brain barrier (BBB) of CNS tumor-bearing mice to significantly prolong survival[.1](#page-12-0) In support of TRAIL signaling as ONC201's target, Foxo3a knockdown inhibits TRAIL upregulation and apoptosis in ONC201-treated colon cancer cells, and TRAIL and DR5 are required for ONC201 sensitivity in breast adeno-carcinoma<sup>[1](#page-12-0)</sup> and lung cancer cells.<sup>1,[23](#page-13-12)</sup> In addition, TRAIL and caspase-8 expression increase in uterine serous carcinoma cell lines treated with ONC201.<sup>[26](#page-13-15)</sup> However, in vitro kinase activity assays revealed no direct interaction between ONC20[1](#page-12-0) and Akt or ERK,<sup>1</sup> and the precise mechanism by which imipridones activate TRAIL signaling is uncertain.

Recent studies have cast doubt on TRAIL signaling as ONC201's primary targeted pathway. These studies revealed that ONC201 elicits apoptosis in the absence of Akt/ERK inhibition<sup>27</sup> and TRAIL or DR5 upregulation in breast,<sup>27</sup> pancreatic,<sup>9</sup> prostate,<sup>16</sup> hematologic cancer,<sup>11,[28](#page-13-19),[29](#page-13-20)</sup> and glioblastoma $30$  cells. In fact, in contrast to the observed upregulation of TRAIL in solid tumors,<sup>1</sup> TRAIL upregulation is limited or absent following imipridone treatment in most hematologic cancer cells.<sup>[25](#page-13-14),[28](#page-13-19)</sup> Indeed, TRAIL-resistant, caspase-8-deficient leukemic T cells are equally sensitive to ONC201-induced apoptosis as their wild-type counterparts.<sup>28</sup> ONC201 treatment does not affect Akt/ERK/Foxo3a or TRAIL levels in hematologic cancer cell lines, and Foxo3a knockdown fails to diminish drug sensitivity in AML cells.<sup>28</sup> In pediatric non-Hodgkin's lymphoma, although imipridone treatment upregulated TRAIL and DR5, antibody-mediated sequestration of TRAIL only partially inhibited apoptosis, $25$  indicating a limited role for TRAIL.

Similar observations have been reported in solid cancers. In prostate cancer cells, 4 of 5 cell lines tested did not show significant TRAIL upregulation, and there was no correlation between TRAIL mRNA levels and ONC201 sensitivity.<sup>[16](#page-13-18)</sup> TRAIL-resistant TBNC cells remain sensitive to ONC201,<sup>31</sup> and only modest and insignificant changes to TRAIL, DR4 and DR5 levels occur in ONC201-treated breast adenocarcinoma cells.<sup>27</sup> Breast and endometrial cancer cell lines treated with ONC201 undergo apoptosis in the presence of the pan-caspase inhibitor Z-VAD-FMK, indicating caspase-independent cell death.<sup>[27](#page-13-16)</sup> Knockdown of DR5 or caspase-8, or treatment with caspase-8 inhibitor (Z-IETD-FMK), does not abrogate glioblastoma cell death, $30$  further suggesting the absence of extrinsic, caspase-8-mediated apoptosis. Moreover, only half of the patients with advanced solid tumors treated with ONC201 exhibited a modest (20%) increase in serum TRAIL.<sup>3</sup> Together, these data indicate that while imipridones may induce TRAIL in certain cancers, their anticancer activity is not entirely dependent on TRAIL activation. Below, we expand on alternative mechanisms of drug action.

# **Activation of the mitochondrial Clp protease**

Imipridones are potent agonists of the mitochondrial Clp pro-tease proteolytic subunit (ClpP)<sup>11,14</sup> [\(Fig. 1\)](#page-6-0). ClpP localizes to the mitochondrial matrix and is essential for the normal turnover of mitochondrial proteins, including mitochondrial ribosomal subunits (*e.g.* Era like 12S mitochondrial rRNA chaperone 1 [ERAL1]) and metabolic enzymes such as electron transport chain (ETC) components. $32,33$  $32,33$  ClpP activity is tightly regulated by its chaperone protein ClpX, which recognizes and unfolds specific proteins, then feeds them into ClpP's proteolytic chamber for degradation.<sup>34</sup> Several cancers overexpress ClpP, including AML (45% of cases).<sup>35</sup> ClpP over-activation using natural acyldepsipeptides (ADEPs) results in impaired mitochondrial oxidative phosphorylation (OXPHOS) and intrinsic cell death, and has been proposed as an anticancer strategy.<sup>[36](#page-13-27)[,37](#page-13-28)</sup>

A screen for ClpP activators identified imipridones (ONC201 and ONC212) as potent activators of ClpP.<sup>[11](#page-13-3)</sup> Imipridones non-covalently bind to ClpP at the interface with ClpX, as demonstrated by isothermal titration calorimetry and co-crystallization studies.<sup>11</sup> ONC201 molecules occupy the hydrophobic pockets between adjacent ClpP subunits, involving extensive hydrophobic contact and hydrogen bonding.<sup>[11](#page-13-3)</sup> The precise interactions between ONC201 and ClpP induce the opening of ClpP's axial entrance pore, which basally is opened only under the regulation of ClpX.<sup>11</sup> ONC201 causes ClpP's entrance pore radius to enlarge (from 12 to 17 Å), its N-terminal residues to exhibit increased dynamics, and its active site to change conformation, altering the placement of its catalytic triad residues.<sup>[11](#page-13-3)</sup> As a result, imipridones activate ClpP in the ab-sence of ClpX.<sup>[11](#page-13-3)</sup> Compared to ONC201, ONC212 interacts with even higher affinity and structural complementarity to ClpP.<sup>11</sup> ONC212's highly electronegative p-CF3-benzyl substituent extends into ClpP's apolar pocket and may form additional multipolar bonds with the protease.<sup>11</sup> The precise binding properties between ClpP and other imipridones have yet to be characterized.

In imipridone-treated glioblastoma cells, ClpP significantly depletes nearly half of its target mitochondrial pro-teins, including strong depletion of ETC components<sup>[11](#page-13-3)</sup> explaining the resulting OXPHOS impairment<sup>38</sup> and decrease in enzymatic activity of respiratory chain complexes I, II, and IV.<sup>11</sup> In breast cancer cells, treatment reduces levels of mitochondrial transcription factor A, mitochondrial (TFAM) and Tu translation elongation factor, mitochondrial (TUFM), an effect that is abolished by ClpP knockdown[.14](#page-13-6) Imipridone treatment also depletes mitochondrial proteins including ClpX (which regulates several other proteins in addition to ClpP), the PAM complex component GRPEL1, ribosomal subunits MRPS7/MRPS22, and metabolic and detoxifying enzymes.<sup>17</sup> While mitochondrial proteins are the hardest hit by ClpP hyper-activation, non-mitochondrial proteins are also targeted, including regulators of cell

<span id="page-6-0"></span>

**Fig. 1** Anticancer mechanism of imipridones. Data from the existing literature indicates that imipridones exert their anticancer effects primarily by binding to and potently activating the mitochondrial Clp protease proteolytic subunit ClpP, causing ClpP to lose its dependence on the chaperone protein ClpX (**Step 1**). Hyper-active Clp protease then depletes its target substrates including components of the respiratory complex chain, most strongly complex I and II proteins (**Step 2**). In turn, OXPHOS is impaired and cellular ATP is depleted (**Step 2**). Mitochondrial structural damage and distress occurs, concomitant with the state of energy deprivation leading to integrated stress response (ISR) activation (**Step 3**). The ISR is relayed to the nucleus through an undefined mechanism, involving the typical (phospho-eIF2α-dependent) or atypical (phospho-eIF2α-independent) pathway. ISR activation causes global translational attenuation, including reduced levels of cyclin D1 leading to cell cycle arrest (**Step 4A**). In conditions of prolonged stress, ATF4 and CHOP are upregulated, and together these transcription factors increase the expression of their target genes including GADD34, which promotes further protein synthesis and stress (thus further activating the ISR); the TRAIL receptor DR5, which can promote TRAIL-mediated extrinsic cell death; and pro-apoptotic Bcl-2 family proteins, which promote the intrinsic, mitochondrial cell death program (**Step 4B**). Created with BioRender.com.

division and cytokinesis (e.g., aurora kinase A [AURKA], cyclin D3 [CCND3], cell division cycle 20 [CDC20]),<sup>[17](#page-13-5)</sup> indicating broad and destructive effects of ClpP agonism both within and beyond the mitochondria.

Consistent with the hyper-degradation of mitochondrial proteins, imipridones induce mitochondrial structural damage in  $AML<sup>11</sup>$  $AML<sup>11</sup>$  $AML<sup>11</sup>$  and breast cancer cells by 3 h post treatment.<sup>27</sup> Severe swelling, matrix lysis, cristae membrane disruption, and disintegration are evident within 6  $h<sub>1</sub><sup>27</sup>$  $h<sub>1</sub><sup>27</sup>$  $h<sub>1</sub><sup>27</sup>$  and by 24 h, mitochondrial fragmentation and fis-sion occur in breast cancer cells.<sup>[27](#page-13-16)</sup> Structural damage to the mitochondria is accompanied by decreased oxygen consumption rate (OCR), ATP, mtDNA, and levels of mitochondrial-encoded genes and nuclear-encoded mitochondrial genes involved in OXPHOS; as well as increased mitochondrial ROS.<sup>11,[27](#page-13-16)</sup>

# **Targeting of mitochondrial bioenergetics**

Metabolic reprogramming is a hallmark of cancer cells, and targeting of metabolic abnormalities presents a therapeutic opportunity. While cancer cells have long been thought to depend primarily on glycolysis for energy production even in the presence of oxygen (known as the Warburg effect<sup>39</sup>), it is now clear that many cancer cells upregulate mitochondrial OXPHOS for adenosine triphosphate (ATP) production.<sup>40</sup> By hyper-activating ClpP, imipridones disrupt OXPHOS and induce a state of energy deprivation in cancer cells. Some studies have indicated that imipridones suppress both OXPHOS and glycolysis, given an observed reduction of extracellular acidification rate (ECAR, a marker of glycolysis), reduction of glycolysis-related proteins (e.g., HK2, LDH1, GLUT1), and accumulation of glycolytic metabolites in imipridonetreated glioblastoma cells.<sup>10</sup> Importantly, treatment with ONC201, ONC206 or ONC212 significantly reduces levels of OXPHOS complexes, basal OCR, ATP, and maximal respiration in glioblastoma cells, indicating strong suppression of OXPHOS.<sup>10</sup> Mitochondrial DNA (mtDNA) copy number levels and mitochondrial membrane potential are also depleted.<sup>10</sup> Other findings indicate that imipridones specifically target mitochondrial OXPHOS:

First, breast cancer cells grown in glucose-containing medium and treated with ONC201 show a reduction in OCR and ATP levels, indicating suppression of OXPHOS;

but also show a slight increase in ECAR, suggesting a compensatory shift towards glycolysis<sup>27</sup> rather than suppression of both pathways. However, when cells are grown in galactose-containing medium, a stronger inhibition of OCR is observed, without a compensatory increase in ECAR,<sup>[27](#page-13-16)</sup> given that the cells cannot metabolize galactose without functioning OXPHOS machinery. Pruss and colleagues showed that imipridone-treated cancer cells strongly downregulate respiratory chain complex I and II proteins, concomitant with OCR downregulation and increased ECAR, $38$  again demonstrating a shift from OXPHOS to glycolysis. Importantly, cells that do not depend on mitochondrial respiration (e.g. cancer cells with reduced mtDNA quantities and those with fumarate hydratase deficiency) are ONC201-resistant.<sup>27</sup> Together, these findings support a mechanism by which imipridones specifically target OXPHOS.

Second, the transition from OXPHOS to glycolysis renders cells vulnerable to glucose depletion: when glucose levels are decreased, imipridone treatment significantly reduces glioblastoma cell viability.<sup>38</sup> This effect is further exacerbated when combining ONC201 with the glucose analog and glycolysis inhibitor 2-deoxyglucose (2-DG), a combination that reduces cell viability, depletes ATP and induces G2/M arrest.<sup>38</sup> With the exception of a few protein kinases (e.g., AMPK $\alpha$ 1, which is phosphorylated upon ATP depletion), ONC201 plus 2-DG treatment results in a state of hypo-phosphorylation across major kinases including mTOR, EGFR and PDGFR $β$ , as well as Akt and ERK $38$ the first kinases implicated in imipridone response $1$ indicating global downregulation of energy metabolism rather than targeting of a specific pathway (e.g., Akt/ERK-Foxo3a-TRAIL signaling).

Third, in response to imipridone treatment, cells upregulate a compensatory energy production pathway, the serine one-carbon cycle, glycine synthesis (SOG) pathway,<sup>10,[38](#page-13-29)</sup> including the enzymes phosphoglycerate dehydrogenase (PHGDH) and phosphoserine aminotransferase 1 (PSAT1). Moreover, ONC212 synergizes with the PDGDH inhibitor (NCT-503 and CBR-5884) to enhance apoptosis in glioblastoma and colon carcinoma cells in vitro, and reduce tumor size in in vivo models.<sup>10</sup> These findings provide support for coupling imipridones with drugs that inhibit alternate metabolic pathways, to effectively target tumor cell metabolism.

## **ISR pathway activation**

Imipridone treatment induces gene expression profiles consistent with the unfolded protein response (UPR) and integrated stress response (ISR) activation, mainly by upregulating the expression of the activating transcription factor-4 (ATF4).<sup>41</sup> ISR is a protective mechanism in response to stressors including nutrient deprivation and misfolded protein aggregation.<sup>42[,43](#page-13-34)</sup> ATF4, the main mediator of the ISR, plays a protective role by upregulating the expression of genes that restore cellular homeostasis.<sup>[44](#page-13-35)</sup> ISR activation may follow a typical (phospho-eIF2αdependent) or atypical (phospho-eIF2α-independent) pathway. In typical ISR, the serine/threonine kinases GCN2, PKR, HRI, or PERK are activated by specific stressors and phosphorylate the eukaryotic translation initiation factor-2-alpha (eIF2 $\alpha$ ) protein.<sup>42</sup> eIF2 $\alpha$  phosphorylation induces global translational attenuation, including reduced cyclin D1 levels causing G1/S cell cycle arrest.<sup>45</sup> Only certain mRNAs, including ATF4, ATF5, CCAAT-enhancer-binding protein homologous protein (CHOP), and growth arrest and DNA-damaging inducible (GADD34), are preferentially translated to protein <sup>[45](#page-13-36),[46](#page-13-37)</sup>

Cancer cells may leverage the protective effects of the ISR to facilitate survival during conditions of stress associated with rapid growth, proliferation and hypoxia, and to evade programmed cell death.<sup>45,[46](#page-13-37)</sup> However, during prolonged or severe stress, ATF4 induces apoptosis by upregulating the transcription factor CHOP. ATF4 and CHOP bind to promoters of genes involved in protein synthesis (e.g., tRNA synthetases, translation initiation factors) and the UPR.<sup>[47](#page-13-38)</sup> In addition, CHOP upregulates GADD34 thereby promoting eIF2 $\alpha$  dephosphorylation. Together, these changes increase protein synthesis, exacerbate cell stress, and ultimately trigger apoptosis. $36,38$  $36,38$  $36,38$  CHOP also downregulates antiapoptotic B-cell lymphoma 2 (Bcl-2) family members and upregulates pro-apoptotic proteins, as well as the TRAIL receptor DR5, to promote apoptosis.<sup>[45](#page-13-36)</sup>

Indeed, imipridone treatment upregulates ATF4 and CHOP across cancer cell lines. [8,](#page-12-6)[28](#page-13-19),[41](#page-13-32) CHOP upregulation increases the expression of target genes including GADD34 and DR5.<sup>28</sup> Interestingly, imipridones can activate either the typical or atypical ISR in a cell type-specific manner. Typical ISR pathway activation is observed in preclinical models of  $AML<sup>28</sup>$  colorectal, $41$  and breast $31$  cancers. In contrast, in MCL $41$  and cutaneous T-cell lymphoma (CTCL) cells,<sup>48</sup> imipridone treatment activates ATF4 via the atypical, phospho-eIF2α-independent ISR. Ishizawa and colleagues speculated that imipridones may do so by inhibiting eIF2B, which can induce ATF4 upregulation, or by inhibiting proteasomal degradation of ATF4 through a yet-undefined mechanism.<sup>28</sup> Notably, ATF4 activation can be triggered by mitochondrial stress, $49,50$  as described in the next section.

## **Downstream targets of ISR activation**

Imipridones agonize ClpP, leading to hyper-degradation of mitochondrial proteins, OXPHOS impairment, and ISR induction mediated by ATF4/CHOP (Fig. 1). However, the exact process by which mitochondrial distress relays to the nucleus to trigger the ISR is uncertain. Mitochondrial distress may trigger the atypical ISR independent of eIF2 $\alpha$  phosphorylation,<sup>49</sup> though this process is not fully understood. Another plausible pathway involves activation of the stress-induced mitochondrial metalloendopeptidase OMA1, which cleaves the inner mitochondrial membrane protein DAP3 binding cell death enhancer 1 (DELE1) leading to cy-tosolic accumulation of DELE1.<sup>[50](#page-14-1)</sup> DELE1 interacts with the eIF2 $\alpha$  kinase HRI, which phosphorylates eIF2 $\alpha$  triggering the ISR and ATF4 upregulation.<sup>50</sup> Alternatively, inhibition of ATP synthases leading to mitochondrial inner-membrane hyperpolarization can induce the ISR, through an incompletely defined mechanism.<sup>[51](#page-14-2)</sup> The ability of mitochondrial distress to evoke the typical or atypical ISR may explain why both pathways have been observed in imipridone-treated cells.

Imipridone treatment culminates in the intrinsic, mitochondrial pathway of apoptosis in many cancer types. This process is regulated by Bcl-2 proteins that control mitochondrial permeability and the release of cytochrome c into the cytosol. The intrinsic apoptotic pathway involves the initiator caspase-9 and executioner caspases-3 and -7.<sup>[52](#page-14-3)</sup> In some pancreatic cancer cells, imipridone treatment induces both the extrinsic, TRAIL/caspase-8-dependent, and the intrinsic, caspase-9-dependent apoptotic pathways.<sup>53</sup> In MCL cells, treatment upregulates pro-apoptotic Bcl-2 proteins Bax, Bak, Puma and Bim,<sup>[28](#page-13-19)</sup> and knockdown of Bax and Bak inhibits apoptosis in glioblastoma cells,  $30$ supporting an intrinsic apoptotic mechanism. In myeloma cells, imipridone treatment upregulates Bim, whereas Bim deficiency reduces drug sensitivity.<sup>29</sup> ONC212-treated AML cells downregulate the anti-apoptotic Bcl-2 protein MCL-1, and undergo apoptosis involving a reduction of mitochondrial membrane potential.<sup>12</sup> TRAIL/DR5 activation is not observed, but rather cell death is caused by the intrinsic mitochondrial pathway, as evidenced by upregulation of cleaved caspase-9, -3, and PARP.<sup>[29](#page-13-20)</sup>

## **Putative biomarkers of response to imipridones**

We have listed several possible predictive biomarkers of imipridone response in [Table 3](#page-9-0). To further investigate the



Abbreviations: ER AML, acute myeloid leukemia; EGFR, epidermal growth factor receptor *MIPEP,* mitochondrial intermediate peptidase; XIAP, X-linked inhibitor of apoptosis; mTOR, Mammalian Target of Rapamycin; BIP, binding immunoglobulin protein; *CLPP*, caseinolytic mitochondrial matrix peptidase proteolytic subunit.

<sup>a</sup>The table lists examples of biomarkers that may be associated with imipridone sensitivity in various cancer types.

<span id="page-9-1"></span>**Oncology Neuro-**

role of candidate biological targets in modulating sensitivity to imipridone treatment, we reanalyzed a newly gen-erated large-scale cell viability drug screening dataset.<sup>[54](#page-14-7)</sup> This study conducted a screen of 4,518 drugs across 578 genetically characterized human cancer cell lines representing 34 different cancer types from the Cancer Cell Line Encyclopedia. Our re-analysis of 539 cell lines ranked *CLPP* as the primary predictor of ONC201 sensitivity at pan-cancer level [\(Fig. 2A](#page-9-1)). More specifically, cell lines with high *CLPP* expression levels responded best to ONC201, whereas cancer cell lines with low *CLPP* expression levels responded weakly to ONC201 [\(Fig. 2B\)](#page-9-1). The association between ONC201 sensitivity and *CLPP* expression remained fully consistent within cancer types [\(Fig. 2D\)](#page-9-1). In contrast, *DRD2* expression levels demonstrated limited correlation with ONC201 sensitivity across and within cancer types [\(Fig. 2C](#page-9-1), [D](#page-9-1)). Notably, cancer cell lines without any detectable *DRD2* expression retained the same ONC201

sensitivity profile as compared to cell lines with detectable *DRD2* expression [\(Supplementary Figure 1\)](http://academic.oup.com/neuro-oncology/article-lookup/doi/10.1093/neuonc/noaa283#supplementary-data), further diminishing the role of D2R antagonism in imipridones' anticancer mechanism.

These findings are consistent with recently published work by Jacques and colleagues, wherein a genome-wide CRISPR/Cas9 gene knockout screen in human pre-B cell lymphocytic leukemia cells revealed that *CLPP* and the mitochondrial intermediate peptidase *MIPEP*, which regulates ClpP maturation, are essential genes for ONC201 and ONC212 sensitivity.[17](#page-13-5) Again, *DRD2* was not an essential gene for response, nor were other candidate targets including genes encoding TRAIL, DR5, and GPR132.[17](#page-13-5) Other resistance-associated genes encode primarily mitochondrial proteins (e.g. ETC components, mitochondrial transcription and translation factors), $17$  reinforcing the key role for mitochondrial function in defining cancer cell sensitivity to imipridones. Additional mitochondrial

<span id="page-9-0"></span>

**Fig. 2** Comprehensive cell viability screen of human cancer cell lines identified ClpP as the primary predictor of ONC201 response. Correlation between cell viability at 2.5 μM ONC201 and gene expression levels (log2 TPM) across 539 human cancer lines from the Cancer Cell Line Encyclopedia (**A**). Scatter plots and boxplots show ONC201 sensitivity compared to gene expression levels for *CLPP* (**B**) and *DRD2* (**C**). Box plots show Spearman's ρ between ONC201 sensitivity and gene expression levels for 16 individual cancer types. Calculations are based on 11 (rhabdoid) to 106 (lung) cancer cell lines (**D**). All gliomas among brain cancer cell lines are *IDH*-wild type. Primary drug screening data for human cancer cell lines was derived from the PRISM Repurposing Screen (1903)<sup>54</sup> and gene expression data from the Cancer Dependency Map (2003).<sup>[57](#page-14-8)</sup> Spearman's ρ and -log10 *P* values are shown. Boxplots show the first quartile, median value, third quartile, and minimum/maximum (whiskers) of the data. Blue lines show linear regression results between ONC201 sensitivity and gene expression levels. Classification of gene expression levels into low/high is based on median. DMSO, dimethylsulfoxide.

biomarkers, including mtDNA copy number, could also serve as predictors of response, under the assumption that the ISR is conditional on the availability of mitochondria in the tumor cell.

## **Effect on immune signaling**

The role of imipridones in immune signaling is another area of active investigation. In CTCL cell lines, imipridones downregulate the pro-inflammatory cytokine IL-32[.48](#page-13-39) In colorectal cancer mouse models, imipridone treatment induces activation and accumulation of CD3+, CD4+, and CD8+ T cells and NK cells in tumors, and CD3+ and NK cells in the blood and spleen.<sup>58</sup> Treatment also increases NK and CD3+ T cells in the blood of healthy, non-tumor-bearing mice, and cells that are resistant to ONC201 in vitro become more sensitive in in vivo models. $58$  These findings suggest that ONC201 treatment may induce an immune response, perhaps independent of the other actions described (e.g. OXPHOS impairment). Depletion of NK cells, but not T cells, affects imipridone response.<sup>58</sup> NK cells may act by secreting TRAIL, and indeed treatment with the TRAIL-sequestering antibody RIK-2 reduces, but does not abolish, the cytotoxic effect of  $ONC201,58$  indicating that the drug's effect on NK activity expands beyond TRAIL secretion.

In clinical trials, granzyme B+ and CD56+ cells, suggestive of infiltrating NK cells, were detected in an on-treatment lymph node biopsy from a metastatic prostate cancer patient receiving ONC201[.59](#page-14-10) Serum immune cytokine and effector molecule profiling revealed a strong immune cytokine induction during the first 2 treatment cycles, followed by a strong effector induction after the second cycle. $59$  Stronger immune responses, defined as >50% serum perforin induction (a component of CTLs and NK cells), correlated to prolonged progres-sion free survival (PFS).<sup>[59](#page-14-10)</sup> The first patient with MCL to receive ONC201 underwent a rectal biopsy after 6 months of treatment, revealing an increase in CHOP, CD45+ lymphocytes, and CD8+ T cells, but not NK or granzyme  $B+$  cells.<sup>[60](#page-14-11)</sup> More studies are needed to understand the immune responses evoked by imipridones across cancer types, and clinical trials (Table 2) are incorporating the assessment of immune responses, including evaluation of NK cell and cytokine profiles, into their outcome measures.

## **Drug interactions**

Imipridones have been tested in combination with numerous FDA-approved small molecule drugs in preclinical studies [\(Table 4\)](#page-11-0), and several promising drug combinations have emerged, some of which have entered clinical trials ([Table 2](#page-3-0)). For example, ONC201 shows increased anticancer efficacy when coupled with Bcl-2 inhibitors in glioblastoma models.<sup>30</sup> Bcl-2 overexpression may protect against ISR signaling and imipridone-induced cell death, as shown in leukemia and lymphoma cells.<sup>28</sup> Combining ONC201 with the Bcl-2 antagonist ABT-199 increases apoptosis in AML cells, including in high Bcl-2 expressing cells and in cells that are resistant to both ONC201 and ABT-199 as single agents.<sup>28</sup>

## **Imipridones for CNS cancers**

CNS tumors are molecularly subtyped based on genomic mutations and copy number variations, $62$  which may activate distinct signaling pathways and shape the metabolic dependencies of the tumor – emphasizing the need to assess imipridone response in the context of CNS tumor molecular subtype. For example, the WHO 2016 integrated classification system defines 5 adult diffuse glioma subtypes incorporating isocitrate dehydrogenase (*IDH)* mutation and 1p19q co-deletion status.<sup>63</sup> *IDH* mutation status influences the metabolic profile of the tumor, with *IDH1* mutations causing increased reliance on OXPHOS, 64 whereas *IDH*-wild-type glioma cells exhibit higher glucose uptake rates[.65](#page-14-15) *IDH*-wild-type glioblastoma is the most common primary malignant brain tumor in adults, and a subset of these tumors harbor *EGFR* amplification, 62-66 which may promote increased glycolysis.<sup>[67](#page-14-17),68</sup> Importantly, recent data suggests that EGFR overexpressing adult glioblastoma cell lines are resistant to ONC201, whereas low EGFR expressing cell lines are sensitive to ONC201 treatment.<sup>55</sup> Given that imipridones' primary mechanism is to agonize ClpP and impair mitochondrial function, the extent to which the tumor is reliant on OXPHOS is likely to influence tumor response to treatment, potentially explaining the insensitivity of EGFR overexpressing glioblastomas to ONC201.[55](#page-14-5) These findings provide support for a recently announced phase 2 clinical trial of ONC201 for adults with EGFR-low glioblastoma [\(Table 2\)](#page-3-0). Notably, primary glioblastomas can acquire *EGFR* mutations or gene amplification during therapy, or exhibit heterogeneity at diagnosis,<sup>69-[71](#page-14-20)</sup> underscoring the need to consider evolving responses to imipridones over the course of disease. More research is needed to assess imipridone response across clinical subtypes of adult and pediatric gliomas.

Although CNS tumors frequently exhibit a shift towards aerobic glycolysis to meet their high energy demands, as reviewed elsewhere, $72$  it is becoming increasingly clear that mitochondrial OXPHOS plays a key role in sustaining CNS tumors. For example, glioblastomas consist of heterogeneous cell populations with different metabolic demands, and OXPHOS is essential to maintain the population of glioblas-toma cancer stem cells.<sup>73,[74](#page-14-23)</sup> Proteomic and phosphoproteomic analyses of *IDH*-wild-type glioblastoma indicate 2 distinct subtypes marked by expression of OXPHOS-related proteins. $75$  One group shows a protein expression signature consistent with aerobic glycolysis and overexpression of neural stem cell markers, while the second group shows an OXPHOS-dependent protein signature and overexpression of oligodendrocyte and astrocyte markers.<sup>75</sup> Thus, protein expression-based subtyping may be helpful for identifying good candidates for imipridone treatment. Indeed, glioblastomas $76$  and pediatric diffuse intrinsic pontine gliomas (DIPGs[\)77](#page-14-26) overexpress OXPHOS-related proteins, including respiratory complex I components—which are among the

**Oncology Neuro-**



[61](#page-14-27)

[53](#page-14-4)

[38](#page-13-29)

[10](#page-13-2)

[12](#page-13-0)

<span id="page-11-0"></span>

Abbreviations: mTOR, Mammalian Target of Rapamycin; AML, acute myeloid leukemia.

<sup>a</sup>The table lists promising drug combinations with imipridones in preclinical cancer models, which may hold potential for future clinical translation.

strongest downregulated proteins in imipridone-treated cells[.38](#page-13-29) Moreover, DIPG cells are sensitive to mitochondrial targeting agents, particularly when combined with PI3K/AKT/ mTOR pathway inhibitors.<sup>77</sup> The importance of both aerobic glycolysis and OXPHOS in sustaining CNS tumors highlights the need for dual targeting of multiple bioenergetics pathways to effectively shut down CNS tumor cell metabolism.

# **Clinical implications**

Clinical trials using imipridones were first launched in 2014, shortly after ONC201's emergence as an anticancer agent. ONC201 reached micromolar therapeutic plasma concentrations and caused only mild (grade I) adverse treatment-related events including fever, nausea, and em-esis, each of which were reversed.<sup>[3](#page-12-2)</sup> The strongest clinical responses occurred in advanced-stage prostate and endometrial cancer patients with involvement of lymph nodes, bone, and lung.<sup>3</sup> Following the demonstrated benign safety profile of ONC201 in its pilot trial, $3$  an extended phase 1 study was launched, continuing ONC201 treatment for 20 patients with heavily pretreated solid tumors (prostate, colon, endometrial cancers, and glioblastoma) resistant to standard-of-care treatment.<sup>59</sup> Five patients with metastatic prostate or endometrial cancers experienced prolonged stable disease for over 6 months.<sup>59</sup>

ONC201 has since expanded into numerous phase 1 and 2 clinical trials, and ONC206 recently entered a phase 1 trial for adults with recurrent and rare primary CNS neoplasms ([Table 2](#page-3-0)). In total, 5 trials are centered on patients

with CNS tumors, 2 of which are open to pediatric patients. In adults with recurrent glioblastoma, ONC201 is well tolerated and achieves CNS penetration with intratumoral concentrations and pharmacodynamics responses, including elevated ATF4 and DR5, and apoptosis induction relative to archival specimens.<sup>4</sup> The positive response in one patient diagnosed with H3K27M-mutant DMG<sup>5</sup> resulted in an expansion of access program enrolling patients harboring H3K2[7](#page-12-4)M-mutant DMG, including DIPG.<sup>7</sup> Two children with DIPG experienced PFS of 13 and 20 months,<sup>7</sup> markedly exceeding the median PFS (7 months) and overall survival (11 months) of this devastating disease.<sup>78</sup> These children showed radiographic evidence of regression and improvement in disease-related neurological symptoms.<sup>7</sup> In a case report of an H3K27M-mutant DIPG patient receiving ONC201, the patient showed a remarkable response with reduced tumor size, a 22-month survival from diagnosis (censored at the time of publication), and no adverse ONC201-related events[.6](#page-12-5)

## **Conclusions**

Imipridones, and in particular ONC201 and ONC206, have emerged as promising therapeutic agents for clinical use in a wide range of cancers. The observed low toxicity profile and BBB penetration of these drugs provide an ideal opportunity for single and combination use for the treatment of malignant CNS cancers. Several questions remain to be clarified as to the anticancer activity of these drugs, including the role of the tumor microenvironment and immune signaling in modulating response across cancer types. Additionally, strategies for overcoming acquired resistance to imipridones should be explored, given the relative ease of developing resistance to small molecule anticancer drugs. Further studies are also needed to address how imipridones trigger the ISR, and how this response plays out across different cancer types. Our collective review of existing data suggests that the sensitivity of cancer cells to imipridones is likely contingent on their relative dependencies on mitochondrial OXPHOS and ability to shift to glycolysis. As described, combinatorial strategies coupling imipridones with other agents that inhibit alternate metabolic pathways may be of interest for targeting tumor cell metabolism, particularly in the context of CNS tumors. Together, the existing preclinical and clinical data supports the promise of these drugs for treating a diverse range of pediatric and adult cancers by targeting common metabolic vulnerabilities.

## **Supplementary Material**

Supplementary material is available online at *Neuro-Oncology* [\(http://neuro-oncology.oxfordjournals.org/](http://neuro-oncology.oxfordjournals.org/)).

## **Funding**

This work was supported by funding from Charlie Kerr and Isabella Kerr Molina Foundation, the Kortney Rose Foundation (Oceanport, NJ), Swifty Foundation (Woodridge, IL), Michael Mosier Defeat DIPG Foundation (Bethesda, MD), ChadTough Foundation (Woodridge, IL), Smashing Walnuts Foundation (Middleburg, VA), Gabriella Miller Kids First Data Resource Center (Philadelphia, PA), the Matthew Larson Foundation (Franklin Lake, NJ), the Lilabean Foundation for Pediatric Brain Cancer Research (Silver Spring, MD), the Research Council of Norway (187615), the South-Eastern Norway Regional Health Authority, and the University of Oslo.

## **Acknowledgements**

The authors would like to acknowledge the generosity of all patients and their families.

**Conflict of interest statement.** The authors declare no conflicts of interest.

**Authorship statement.** Reviewed articles, prepared manuscript text, created figures, and tables: ERB. Edited and revised manuscript text, performed data re-analysis, and contributed to preparation of figures: SMW. Edited and revised manuscript text: SM and MAG. Edited and revised manuscript text, figures and tables, and served as corresponding author: JN.

## **References**

- <span id="page-12-0"></span>1. Allen JE, Krigsfeld G, Mayes PA, et al. Dual inactivation of Akt and ERK by TIC10 signals Foxo3a nuclear translocation, TRAIL gene induction, and potent antitumor effects. *Sci Transl Med.* 2013;5(171):171ra117.
- <span id="page-12-1"></span>2. Allen JE, Crowder RN, Crowder R, El-Deiry WS. First-in-class small molecule ONC201 induces DR5 and cell death in tumor but not normal cells to provide a wide therapeutic index as an anti-cancer agent. *PLoS One.* 2015;10(11):e0143082.
- <span id="page-12-2"></span>3. Stein MN, Bertino JR, Kaufman HL, et al. First-in-human clinical trial of oral ONC201 in patients with refractory solid tumors. *Clin Cancer Res.* 2017;23(15):4163–4169.
- <span id="page-12-3"></span>4. Arrillaga-Romany I, Chi AS, Allen JE, Oster W, Wen PY, Batchelor TT. A phase 2 study of the first imipridone ONC201, a selective DRD2 antagonist for oncology, administered every three weeks in recurrent glioblastoma. *Oncotarget.* 2017;8(45):79298–79304.
- <span id="page-12-7"></span>5. Arrillaga-Romany I, Odia Y, Prabhu VV, et al. Biological activity of weekly ONC201 in adult recurrent glioblastoma patients. *Neuro Oncol.* 2020;22(1):94–102.
- <span id="page-12-5"></span>6. Hall MD, Odia Y, Allen JE, et al. First clinical experience with DRD2/3 antagonist ONC201 in H3 K27M-mutant pediatric diffuse intrinsic pontine glioma: a case report. *J Neurosurg Pediatr.* 2019:1–7.
- <span id="page-12-4"></span>7. Chi AS, Tarapore RS, Hall MD, et al. Pediatric and adult H3 K27Mmutant diffuse midline glioma treated with the selective DRD2 antagonist ONC201. *J Neurooncol.* 2019;145(1):97–105.
- <span id="page-12-6"></span>8. Wagner J, Kline CL, Ralff MD, et al. Preclinical evaluation of the imipridone family, analogs of clinical stage anti-cancer small molecule

ONC201, reveals potent anti-cancer effects of ONC212. *Cell Cycle.* 2017;16(19):1790–1799.

- <span id="page-13-17"></span>9. Lev A, Lulla AR, Wagner J, et al. Anti-pancreatic cancer activity of ONC212 involves the unfolded protein response (UPR) and is reduced by IGF1-R and GRP78/BIP. *Oncotarget.* 2017;8(47):81776–81793.
- <span id="page-13-2"></span>10. Ishida CT, Zhang Y, Bianchetti E, et al. Metabolic reprogramming by dual AKT/ERK inhibition through imipridones elicits unique vulnerabilities in glioblastoma. *Clin Cancer Res.* 2018;24(21):5392–5406.
- <span id="page-13-3"></span>11. Ishizawa J, Zarabi SF, Davis RE, et al. Mitochondrial ClpP-mediated proteolysis induces selective cancer cell lethality. *Cancer Cell.* 2019;35(5):721–737.e729.
- <span id="page-13-0"></span>12. Nii T, Prabhu VV, Ruvolo V, et al. Imipridone ONC212 activates orphan G protein-coupled receptor GPR132 and integrated stress response in acute myeloid leukemia. *Leukemia.* 2019;33(12):2805–2816.
- <span id="page-13-1"></span>13. Kline CLB, Ralff MD, Lulla AR, et al. Role of dopamine receptors in the anticancer activity of ONC201. *Neoplasia.* 2018;20(1):80–91.
- <span id="page-13-6"></span>14. Graves PR, Aponte-Collazo LJ, Fennell EMJ, et al. Mitochondrial protease ClpP is a target for the anticancer compounds ONC201 and related analogues. *ACS Chem Biol.* 2019;14(5):1020–1029.
- <span id="page-13-4"></span>15. Madhukar NS, Khade PK, Huang L, et al. A Bayesian machine learning approach for drug target identification using diverse data types. *Nat Commun.* 2019;10(1):5221.
- <span id="page-13-18"></span>16. Lev A, Lulla AR, Ross BC, et al. ONC201 targets AR and AR-V7 signaling, reduces PSA, and synergizes with everolimus in prostate cancer. *Mol Cancer Res.* 2018;16(5):754–766.
- <span id="page-13-5"></span>17. Jacques S, van der Sloot AM, C Huard C, et al. Imipridone anticancer compounds ectopically activate the ClpP protease and represent a new scaffold for antibiotic development. *Genetics.* 2020;214(4):1103–1120.
- <span id="page-13-7"></span>18. Jacob NT, Lockner JW, Kravchenko VV, Janda KD. Pharmacophore reassignment for induction of the immunosurveillance cytokine TRAIL. *Angew Chem Int Ed Engl.* 2014;53(26):6628–6631.
- <span id="page-13-8"></span>19. Wagner J, Kline CL, Pottorf RS, et al. The angular structure of ONC201, a TRAIL pathway-inducing compound, determines its potent anti-cancer activity. *Oncotarget.* 2014;5(24):12728–12737.
- <span id="page-13-9"></span>20. Prabhu VV, Madhukar NS, Gilvary C, et al. Dopamine receptor D5 is a modulator of tumor response to dopamine receptor D2 antagonism. *Clin Cancer Res.* 2019;25(7):2305–2313.
- <span id="page-13-10"></span>21. Peters MAM, Meijer C, Fehrmann RSN, et al. Serotonin and dopamine receptor expression in solid tumours including rare cancers. *Pathol Oncol Res.* 2020;26(3):1539–1547.
- <span id="page-13-11"></span>22. Yuan X, Gajan A, Chu Q, Xiong H, Wu K, Wu GS. Developing TRAIL/ TRAIL death receptor-based cancer therapies. *Cancer Metastasis Rev.* 2018;37(4):733–748.
- <span id="page-13-12"></span>23. Feng Y, Zhou J, Li Z, Jiang Y, Zhou Y. Small molecular TRAIL inducer ONC201 induces death in lung cancer cells: a preclinical study. *PLoS One.* 2016;11(9):e0162133.
- <span id="page-13-13"></span>24. Prabhu VV, Allen JE, Dicker DT, El-Deiry WS. Small-molecule ONC201/ TIC10 targets chemotherapy-resistant colorectal cancer stem-like cells in an Akt/Foxo3a/TRAIL-dependent manner. *Cancer Res.* 2015;75(7):1423–1432.
- <span id="page-13-14"></span>25. Talekar MK, Allen JE, Dicker DT, El-Deiry WS. ONC201 induces cell death in pediatric non-Hodgkin's lymphoma cells. *Cell Cycle.* 2015;14(15):2422–2428.
- <span id="page-13-15"></span>26. Fang Z, Wang J, Clark LH, et al. ONC201 demonstrates anti-tumorigenic and anti-metastatic activity in uterine serous carcinoma in vitro. *Am J Cancer Res.* 2018;8(8):1551–1563.
- <span id="page-13-16"></span>27. Greer YE, Porat-Shliom N, Nagashima K, et al. ONC201 kills breast cancer cells in vitro by targeting mitochondria. *Oncotarget.* 2018;9(26):18454–18479.
- <span id="page-13-19"></span>28. Ishizawa J, Kojima K, Chachad D, et al. ATF4 induction through an atypical integrated stress response to ONC201 triggers p53-independent

apoptosis in hematological malignancies. *Sci Signal.* 2016;9( 415):ra17.

- <span id="page-13-20"></span>29. Tu YS, He J, Liu H, et al. The imipridone ONC201 induces apoptosis and overcomes chemotherapy resistance by up-regulation of bim in multiple myeloma. *Neoplasia.* 2017;19(10):772–780.
- <span id="page-13-21"></span>30. Karpel-Massler G, Bâ M, Shu C, et al. TIC10/ONC201 synergizes with Bcl-2/Bcl-xL inhibition in glioblastoma by suppression of Mcl-1 and its binding partners in vitro and in vivo. *Oncotarget.* 2015;6(34):36456–36471.
- <span id="page-13-22"></span>31. Yuan X, Kho D, Xu J, Gajan A, Wu K, Wu GS. ONC201 activates ER stress to inhibit the growth of triple-negative breast cancer cells. *Oncotarget.* 2017;8(13):21626–21638.
- <span id="page-13-23"></span>32. Szczepanowska K, Maiti P, Kukat A, et al. CLPP coordinates mitoribosomal assembly through the regulation of ERAL1 levels. *EMBO J.* 2016;35(23):2566–2583.
- <span id="page-13-24"></span>33. Fischer F, Langer JD, Osiewacz HD. Identification of potential mitochondrial CLPXP protease interactors and substrates suggests its central role in energy metabolism. *Sci Rep.* 2015;5:18375.
- <span id="page-13-25"></span>34. Baker TA, Sauer RT. ClpXP, an ATP-powered unfolding and proteindegradation machine. *Biochim Biophys Acta.* 2012;1823(1):15–28.
- <span id="page-13-26"></span>35. Cole A, Wang Z, Coyaud E, et al. Inhibition of the mitochondrial protease ClpP as a therapeutic strategy for human acute myeloid leukemia. *Cancer Cell.* 2015;27(6):864–876.
- <span id="page-13-27"></span>36. Wong KS, Mabanglo MF, Seraphim TV, et al. Acyldepsipeptide analogs dysregulate human mitochondrial ClpP protease activity and cause apoptotic cell death. *Cell Chem Biol.* 2018;25(8):1017–1030.e1019.
- <span id="page-13-28"></span>37. Dougan DA, Hantke I, Turgay K. Dysregulating ClpP: from antibiotics to anticancer? *Cell Chem Biol.* 2018;25(8):929–930.
- <span id="page-13-29"></span>38. Pruss M, Dwucet A, Tanriover M, et al. Dual metabolic reprogramming by ONC201/TIC10 and 2-Deoxyglucose induces energy depletion and synergistic anti-cancer activity in glioblastoma. *Br J Cancer.* 2020;122(8):1146–1157.
- <span id="page-13-30"></span>39. Warburg O, Wind F, Negelein E. The metabolism of tumors in the body. *J Gen Physiol.* 1927;8(6):519–530.
- <span id="page-13-31"></span>40. Ashton TM, McKenna WG, Kunz-Schughart LA, Higgins GS. Oxidative phosphorylation as an emerging target in cancer therapy. *Clin Cancer Res.* 2018;24(11):2482–2490.
- <span id="page-13-32"></span>41. Kline CL, Van den Heuvel AP, Allen JE, Prabhu VV, Dicker DT, El-Deiry WS. ONC201 kills solid tumor cells by triggering an integrated stress response dependent on ATF4 activation by specific eIF2alpha kinases. *Sci Signal.* 2016;9(415):ra18.
- <span id="page-13-33"></span>42. Pakos-Zebrucka K, Koryga I, Mnich K, Ljujic M, Samali A, Gorman AM. The integrated stress response. *EMBO Rep.* 2016;17(10):1374–1395.
- <span id="page-13-34"></span>43. Melber A, Haynes CM. UPRmt regulation and output: a stress response mediated by mitochondrial-nuclear communication. *Cell Res.* 2018;28(3):281–295.
- <span id="page-13-35"></span>44. Quirós PM, Prado MA, Zamboni N, et al. Multi-omics analysis identifies ATF4 as a key regulator of the mitochondrial stress response in mammals. *J Cell Biol.* 2017;216(7):2027–2045.
- <span id="page-13-36"></span>45. Rozpedek W, Pytel D, Mucha B, Leszczynska H, Diehl JA, Majsterek I. The role of the PERK/eIF2 $\alpha$ /ATF4/CHOP signaling pathway in tumor progression during endoplasmic reticulum stress. *Curr Mol Med.* 2016;16(6):533–544.
- <span id="page-13-37"></span>46. Ramirez MU, Hernandez SR, Soto-Pantoja DR, Cook KL. Endoplasmic reticulum stress pathway, the unfolded protein response, modulates immune function in the tumor microenvironment to impact tumor progression and therapeutic response. *Int J Mol Sci.* 2019;21(1):169.
- <span id="page-13-38"></span>47. Han J, Back SH, Hur J, et al. ER-stress-induced transcriptional regulation increases protein synthesis leading to cell death. *Nat Cell Biol.* 2013;15(5):481–490.
- <span id="page-13-39"></span>48. Ni X, Zhang X, Hu CH, et al. ONC201 selectively induces apoptosis in cutaneous T-cell lymphoma cells via activating pro-apoptotic integrated

stress response and inactivating JAK/STAT and NF-κB pathways. *Oncotarget.* 2017;8(37):61761–61776.

- <span id="page-14-0"></span>49. Münch C, Harper JW. Mitochondrial unfolded protein response controls matrix pre-RNA processing and translation. *Nature.* 2016;534(7609):710–713.
- <span id="page-14-1"></span>50. Guo X, Aviles G, Liu Y, et al. Mitochondrial stress is relayed to the cytosol by an OMA1-DELE1-HRI pathway. *Nature.* 2020;579(7799): 427–432.
- <span id="page-14-2"></span>51. Mick E, Titov DV, Skinner OS, Sharma R, Jourdain AA, Mootha VK. Distinct mitochondrial defects trigger the integrated stress response depending on the metabolic state of the cell. *Elife.* 2020;9:e49178.
- <span id="page-14-3"></span>52. Wang C, Youle RJ. The role of mitochondria in apoptosis\*. *Annu Rev Genet.* 2009;43:95–118.
- <span id="page-14-4"></span>53. Zhang Q, Wang H, Ran L, Zhang Z, Jiang R. The preclinical evaluation of TIC10/ONC201 as an anti-pancreatic cancer agent. *Biochem Biophys Res Commun.* 2016;476(4):260–266.
- <span id="page-14-7"></span>54. Corsello SM, Nagari RT, Spangler RD, et al. Discovering the anti-cancer potential of non-oncology drugs by systematic viability profiling. *Nat Cancer.* 2020;1(2):235–248.
- <span id="page-14-5"></span>55. He Y, Li J, Koga T, et al. Epidermal growth factor receptor (EGFR) as a molecular determinant of glioblastoma response to dopamine receptor 2 (DRD2) inhibitors. *Neuro Oncol.* 2020.
- <span id="page-14-6"></span>56. Jin ZZ, Wang W, Fang DL, Jin YJ. mTOR inhibition sensitizes ONC201 induced anti-colorectal cancer cell activity. *Biochem Biophys Res Commun.* 2016;478(4):1515–1520.
- <span id="page-14-8"></span>57. DepMap: The Cancer Dependency Map Project at Broad Institute. DepMap 20Q3 Public. *Dataset.* 2020. doi[:10.6084/](https://doi.org/10.6084/m9.figshare.12931238.v1) [m9.figshare.12931238.v1](https://doi.org/10.6084/m9.figshare.12931238.v1).
- <span id="page-14-9"></span>58. Wagner J, Kline CL, Zhou L, et al. Dose intensification of TRAIL-inducing ONC201 inhibits metastasis and promotes intratumoral NK cell recruitment. *J Clin Invest.* 2018;128(6):2325–2338.
- <span id="page-14-10"></span>59. Stein MN, Malhotra J, Tarapore RS, et al. Safety and enhanced immunostimulatory activity of the DRD2 antagonist ONC201 in advanced solid tumor patients with weekly oral administration. *J Immunother Cancer.* 2019;7(1):136.
- <span id="page-14-11"></span>60. Romaguera JE, Lee HJ, Tarapore R, et al. Integrated stress response and immune cell infiltration in an ibrutinib-refractory mantle cell lymphoma patient following ONC201 treatment. *Br J Haematol.* 2019;185(1):133–136.
- <span id="page-14-27"></span>61. Allen JE, Prabhu VV, Talekar M, et al. Genetic and pharmacological screens converge in identifying FLIP, BCL2, and IAP proteins as key regulators of sensitivity to the TRAIL-inducing anticancer agent ONC201/ TIC10. *Cancer Res.* 2015;75(8):1668–1674.
- <span id="page-14-12"></span>62. Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization classification of tumors of the central nervous system: a summary. *Acta Neuropathol.* 2016;131(6):803–820.
- <span id="page-14-13"></span>63. Molinaro AM, Taylor JW, Wiencke JK, Wrensch MR. Genetic and molecular epidemiology of adult diffuse glioma. *Nat Rev Neurol.* 2019;15(7):405–417.
- <span id="page-14-14"></span>64. Grassian AR, Parker SJ, Davidson SM, et al. IDH1 mutations alter citric acid cycle metabolism and increase dependence on oxidative mitochondrial metabolism. *Cancer Res.* 2014;74(12):3317–3331.
- <span id="page-14-15"></span>65. Garrett M, Sperry J, Braas D, et al. Metabolic characterization of isocitrate dehydrogenase (IDH) mutant and IDH wildtype gliomaspheres uncovers cell type-specific vulnerabilities. *Cancer Metab.* 2018;6:4.
- <span id="page-14-16"></span>66. Verhaak RG, Hoadley KA, Purdom E, et al.; Cancer Genome Atlas Research Network. Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH1, EGFR, and NF1. *Cancer Cell.* 2010;17(1):98–110.
- <span id="page-14-17"></span>67. Makinoshima H, Takita M, Saruwatari K, et al. Signaling through the Phosphatidylinositol 3-Kinase (PI3K)/Mammalian Target of Rapamycin (mTOR) axis is responsible for aerobic glycolysis mediated by glucose transporter in epidermal growth factor receptor (EGFR)-mutated lung adenocarcinoma. *J Biol Chem.* 2015;290(28):17495–17504.
- <span id="page-14-18"></span>68. Yang W, Xia Y, Cao Y, et al. EGFR-induced and PKCε monoubiquitylationdependent NF-κB activation upregulates PKM2 expression and promotes tumorigenesis. *Mol Cell.* 2012;48(5):771–784.
- <span id="page-14-19"></span>69. Francis JM, Zhang CZ, Maire CL, et al. EGFR variant heterogeneity in glioblastoma resolved through single-nucleus sequencing. *Cancer Discov.* 2014;4(8):956–971.
- 70. Wang J, Cazzato E, Ladewig E, et al. Clonal evolution of glioblastoma under therapy. *Nat Genet.* 2016;48(7):768–776.
- <span id="page-14-20"></span>71. Eskilsson E, Røsland GV, Solecki G, et al. EGFR heterogeneity and implications for therapeutic intervention in glioblastoma. *Neuro Oncol.* 2018;20(6):743–752.
- <span id="page-14-21"></span>72. Agnihotri S, Zadeh G. Metabolic reprogramming in glioblastoma: the influence of cancer metabolism on epigenetics and unanswered questions. *Neuro Oncol.* 2016;18(2):160–172.
- <span id="page-14-22"></span>73. Janiszewska M, Suvà ML, Riggi N, et al. Imp2 controls oxidative phosphorylation and is crucial for preserving glioblastoma cancer stem cells. *Genes Dev.* 2012;26(17):1926–1944.
- <span id="page-14-23"></span>74. Hoang-Minh LB, Siebzehnrubl FA, Yang C, et al. Infiltrative and drugresistant slow-cycling cells support metabolic heterogeneity in glioblastoma. *EMBO J.* 2018;37(23):e98772.
- <span id="page-14-24"></span>75. Oh S, Yeom J, Cho HJ, et al. Integrated pharmaco-proteogenomics defines two subgroups in isocitrate dehydrogenase wild-type glioblastoma with prognostic and therapeutic opportunities. *Nat Commun.* 2020;11(1):3288.
- <span id="page-14-25"></span>76. Park J, Shim JK, Kang JH, et al. Regulation of bioenergetics through dual inhibition of aldehyde dehydrogenase and mitochondrial complex I suppresses glioblastoma tumorspheres. *Neuro Oncol.* 2018;20(7):954–965.
- <span id="page-14-26"></span>77. Tsoli M, Liu J, Franshaw L, et al. Dual targeting of mitochondrial function and mTOR pathway as a therapeutic strategy for diffuse intrinsic pontine glioma. *Oncotarget.* 2018;9(7):7541–7556.
- <span id="page-14-28"></span>78. Cooney T, Lane A, Bartels U, et al. Contemporary survival endpoints: an International Diffuse Intrinsic Pontine Glioma Registry study. *Neuro Oncol.* 2017;19(9):1279–1280.