

***TBX15* rs98422, *DNM3* rs1011731, *RAD51B* rs8017304, and rs2588809 Gene Polymorphisms and Associations With Pituitary Adenoma**

GABIJA JUKNYTĖ¹, INGA LAURINAITYTĖ¹, ALVITA VILKEVIČIŪTĖ², GRETA GEDVILAITĖ², BRIGITA GLEBAUSKIENĖ², LORESA KRIAUCIŪNIENĖ² and RASA LIUTKEVIČIENĖ²

¹*Medical Academy, Lithuanian University of Health Sciences, Kaunas, Lithuania;*

²*Neuroscience Institute, Medical Academy, Lithuanian University of Health Sciences, Kaunas, Lithuania*

Abstract. *Background:* Pituitary adenoma (PA) is a benign tumor of parenchymal cells in the adenohypophysis, and its development is strongly associated with genetic factors. This study aim was to find whether *TBX15* rs98422, *DNM3* rs1011731, *RAD51B* rs8017304, and rs2588809 single nucleotide polymorphisms can be associated with pituitary adenoma. While the *TBX15* gene belongs to the T-box family of genes and is a transcription factor involved in many developmental processes, the *DNM3* encodes a protein that is a member of the dynamin family with mechanochemical properties involved in actin-membrane processes, predominantly in membrane budding, and the *RAD51B* gene plays a significant role in homologous recombination in DNA repair for genome stability. *Materials and Methods:* The study enrolled 113 patients with pituitary adenoma and 283 healthy control subjects. DNA samples were extracted and purified from peripheral blood leukocytes. Genotyping was carried out using real-time polymerase chain reaction. The results were assessed using binomial logistic regression. *Results:* Our study revealed that *RAD51B* rs2588809 TT genotype could be associated with PA development in the co-dominant (OR=6.833; 95% CI=2.557-18.262; $p<0.001$) and recessive (OR=7.066; 95% CI=2.667-18.722; $p<0.001$) models. The same results were observed in females but not in males and PA without recurrence, while in PA with recurrence, no statistically significant results were obtained.

Conclusion: *RAD51B* rs2588809 TT genotype may increase the odds of PA development in women; it may also be associated with non-recurrent PA development.

Pituitary adenoma (PA) is an intracranial tumor localized in the bone cavity (*sella turcica*) surrounded by multiple neural, vascular, endocrine, and bone structures, which further may contribute to an assortment of tumor types (1-7). PA accounts for approximately 15 to 20 percent of primary brain tumors with a prevalence of 77.6-97.6 PA cases per 100,000 individuals. Clinically significant PAs occur in one out of 1064 individuals (5-11). PA can occur insidiously – most patients do not realize they have it until specifically investigated. This tumor can manifest in two ways: an endocrine imbalance or pressure on the surrounding structures. The latter is the most common form of macroadenoma manifestation (12). Six to ten percent of all PAs expand into the cavernous sinus (13, 14). The optic chiasm is directly above the pituitary gland, so a prolonged compression of the chiasm can cause primary optic nerve atrophy and result in visual function defects, such as decreased visual acuity and visual field defects or impaired color vision (15). The earlier the tumor is diagnosed, the more likely it is to be removed and the visual function to be preserved. Endocrine changes may be due to the overexpression of tumor hormones or hypoexpression, when the tumor compresses the pituitary gland (16).

The etiology and pathogenesis of PA are complex and still poorly understood. PA represents a heterogeneous disease whose pathogenesis is a multifactorial process that involves both environmental and genetic factors. Therefore, a better understanding of the PA pathogenesis requires a comprehensive research of this disease's biological and genetic markers. Plenty of possible molecular markers, as well as interleukin 9 variant rs1859430, which might be incorporated in the tumorigenesis of PAs, are currently under investigation (17). Recent studies focus on genetic

This article is freely accessible online.

Correspondence to: Alvita Vilkeviciute, Laboratory of Ophthalmology, Neuroscience Institute, Lithuanian University of Health Sciences, Eiveniu st. 2, Kaunas, LT-50161, Lithuania. Tel: +37 062424461, e-mail: alvita.vilkeviciute@lsmuni.lt

Key Words: Pituitary adenoma, prolactinoma, *TBX15*, *DNM3*, *RAD51B*, gene polymorphisms.

Table I. *Characteristics of study subjects.*

Characteristics	Subjects with PA (group I) n=113	Control group (group II) n=283	<i>p</i> -Value
Men, n (%)	45 (39.8)	100 (35.3)	0.438
Women, n (%)	68 (60.2)	183 (64.7)	
Age, median (IQR)	54 (22.5)	55.5 (27)	0.426

markers for cancer development, so we aimed to elucidate the role of four *TBX15*, *DNM3* and *RAD51B* single nucleotide variants in PA development. The elevated genes has been reported in a variety of cancers, including prostate, ovarian cancers. However, the data regarding *TBX15*, *DNM3* and *RAD51B* genes and PA is still lacking (18-20).

TBX15, a T-box family member, is possibly involved in cancer cell transformation because of its antiapoptotic function (18). It also is known that T-box genes are involved in in carcinogenesis (21-23). *TBX15* is associated with prostate (24), thyroid cancer (19, 25, 26), ovarian carcinoma (20).

The other marker dynamin 3 (*DNM3*) is a candidate tumor suppressor gene. This gene encodes a member of the dynamin family, which possesses mechanochemical properties to tabulate and sever membranes (27). However, few reports describe the relationship between *DNM3* and malignant diseases (28, 29). *DNM3* has been found mainly in the brain (at a lower level than *DNM1*) and testicles, and less frequently in the lungs and heart (30). The importance of the *DNM3* gene has been investigated in gliomas (31, 32), hepatocellular carcinoma (33-35), colon cancer (36), and papillary thyroid carcinoma (37). Few studies have investigated the association between *DNM3* and hepatocellular carcinoma, breast cancer, T-cell lymphoma, colon cancer (28-30, 33-34, 38, 39). Additionally, the importance of *DNM3* was investigated in brain tumors glioblastomas (31-32).

RAD51B plays a role in homologous DNA pairing and strand exchange in DNA double-strand break repair (38, 40). The importance of *RAD51B* has previously been investigated in the breast, ovarian, lung cancer and uterine leiomyomas (31, 41-43). Also, some studies have been carried out to look for the possible association between the *RAD51* gene variants and pancreatic (44-47), prostate cancer (48, 49), malignant melanoma (50), colorectal adenocarcinoma (51), endometrial cancer (52), soft tissue sarcoma (53) and glioblastoma (54).

Our study aimed to determine associations between *TBX15* rs98422, *DNM3* rs1011731, *RAD51B* rs8017304, rs2588809 single nucleotide polymorphisms and pituitary adenoma invasiveness, development, and recurrence.

These findings support the hypothesised role of *TBX15*, *DNM3* and *RAD 51* as tumour promoters. Based on the *TBX15*, *DNM3* and *RAD51B* associations with cancerous

processes we selected four widely described SNPs located in these genes. According to the dbSNP database (<https://www.ncbi.nlm.nih.gov/snp/>) the minor allele frequencies of these intronic variants (*TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304, rs2588809) are more than 0.1 in the Europe population, and none of these variants have been studied with PA development, invasiveness, PA activity and recurrence. The aim of the present study was to determine these associations.

Materials and Methods

Patients and selection. This study was carried out at the Department of Ophthalmology, Hospital of Lithuanian University of Health Sciences and Laboratory of Ophthalmology, Neuroscience Institute, Lithuanian University of Health Sciences. The Ethics Committee for Biomedical Research at Lithuanian University of Health Sciences (LUHS) approved the study (number BE-2-47). All subjects provided written informed consent under the Declaration of Helsinki. Based on our inclusion and exclusion criteria (55), two groups were formed in the study: the PA group (*n*=113) and the control group (*n*=283).

Evaluation of PA hormonal activity, invasiveness, recurrence and DNA extraction and genotyping. The analysis of all pituitary adenomas was based on histopathological findings of PA and hormone levels in the blood serum before surgery. All PA subjects were categorized into two groups – active and inactive PA (56).

Since some of the subjects had already had surgery in recent years, we categorized them by recurrence of pituitary adenoma into two groups – PA with and without recurrence.

Pituitary adenoma recurrence was diagnosed when enlargement of a residual tumor or a new growth was documented on follow-up magnetic resonance imaging (MRI) after surgical resection during the period of this study. The residual tumor was considered stable if there no signs of tumor progression on follow-up MRI. Most prolactinomas were surgically treated because of the remaining pressure effects of surrounding structures or ineffective medical treatment.

PA invasiveness has been described previously (55). The suprasellar extension and sphenoid sinus invasion by PA were classified according to the Hardy classification modified by Wilson, and the degree of suprasellar and parasellar extensions was graded as stages A–E. The degree of sellar floor erosion was graded as grades I-IV. Grade III shows localized sellar perforation, and grade IV shows diffuse destruction of sellar floor, which are the signs of invasive PA. The Knosp classification system was used to quantify the invasion of the cavernous sinus. Grade 3 and 4 pituitary tumors were considered to be invasive.

DNA was extracted from 200 µL venous blood (white blood cells) using the silica-based membrane technology utilizing a genomic DNA extraction kit (GeneJET Genomic DNA Purification Kit, Thermo Scientific, MA, USA), according to the manufacturer’s recommendations. The genotyping of *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304 and rs2588809 was carried out using the real-time PCR. SNPs were genotyped on the Step One Plus real-time PCR system (Applied Biosystems, Foster City, CA, USA). The TaqMan® SNP genotyping assays (Thermo Scientific) for all SNPs were performed according to the manufacturer’s protocol. The Allelic Discrimination program was used during the

Table II. *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 genotype and allele frequencies in the PA patient and control groups.

SNP	Genotype/ allele	Group		p-Value
		Control group n=283 n (%)	PA group n=113 n (%)	
<i>TBX15</i> rs984222	Genotype			
	G/G	141 (49.8)	65 (57.5)	0.341
	G/C	120 (42.4)	42 (37.2)	
	C/C	22 (7.8)	6 (5.3)	
	In total	283 (100)	113 (100)	
Allele	G	402 (71.02)	172 (76.11)	0.148
	C	164 (28.98)	54 (23.89)	
<i>DNMT3</i> rs1011731	Genotype			
	A/A	90 (31.8)	34 (30.1)	0.919
	G/A	142 (50.2)	57 (50.4)	
	G/G	51 (18.0)	22 (19.5)	
	In total	283 (100)	113 (100)	
Allele	G	322 (56.89)	125 (55.31)	0.685
	A	244 (43.11)	101 (44.69)	
<i>RAD51B</i> rs8017304	Genotype			
	AA	130 (45.94)	49 (43.36)	0.609
	AG	116 (40.98)	52 (46.02)	
	GG	37 (13.08)	12 (10.62)	
	In total	283 (100)	113 (100)	
Allele	A	376 (66.43)	150 (66.37)	0.987
	G	190 (33.57)	76 (33.63)	
<i>RAD51B</i> rs2588809	Genotype			
	CC	198 (69.96)	74 (65.49)	0.024
	CT	70 (24.74)	24 (21.24)	
	TT	15 (5.30)	15 (13.27)	
	In total	283 (100)	113 (100)	
Allele	C	466 (82.33)	172 (76.12)	0.045
	T	100 (17.67)	54 (23.88)	

PA: Pituitary adenoma; p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$.

real-time PCR. The program determined individual genotypes according to the fluorescence intensity rate of different detectors (VIC and FAM).

Statistical analysis. The age of study participants was presented as the median and interquartile range (IQR). It was compared between both study groups using the nonparametric Mann-Whitney *U*-test. All categorical variables of *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304 and rs2588809 genotypes and alleles were expressed as absolute numbers with percentages in brackets and compared using the Pearson's χ^2 and Fisher's exact test (when $n < 50$) in both groups. Binomial logistic regression analysis was performed to evaluate the genotype and allele impact on PA development and reported as odds ratios (ORs) with 95% confidence intervals (CIs). The lowest values of the Akaike

Table III. Binary logistic regression analysis of *RAD51B* rs2588809.

Model	Genotype	OR (95% CI)	p-Value	AIC
<i>RAD51B</i> rs2588809				
Co-dominant	C/T	0.873 (0.509; 1.497)	0.622	459.716
	T/T	6.833 (2.557; 18.262)	<0.001	
Dominant	C/T+T/T	1.332 (0.833; 2.129)	0.232	474.155
Recessive	T/T	7.066 (2.667; 18.722)	<0.001	457.963
Overdominant	C/T	0.763 (0.449; 1.298)	0.318	474.546
Additive	T	1.627 (1.135; 2.334)	0.008	468.686

OR: Odds ratio; AIC: Akaike information criterion; p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant p-Values are shown in bold.

information criterion (AIC) showed the best genetic models. Statistically significant differences were reported when $p < 0.05$, but for multiple comparisons, the Bonferroni correction was applied with the $p < 0.05/4$ (since we analyzed four different SNPs).

Results

A total of 396 individuals were included in the study. Two groups of subjects were formed during the study. The first one included patients with pituitary adenoma, the second included healthy subjects (control group). The characteristics of the subjects are presented in Table I. The first group consisted of 113 individuals, of whom 45 (39.8%) were men, and 68 (60.2%) were women. The median age of this group was 54 years. The control group consisted of 100 (35.3%) men and 183 (64.7%) women. In total, the control group consisted of 283 individuals with a median age of 55.5 years.

TBX15 rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 genotype frequencies in the pituitary adenoma and healthy population groups. Hardy Weinberg analysis was performed to compare the observed and expected frequencies of *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 using the χ^2 test in the control group. The genotype distribution of the polymorphisms matched the Hardy-Weinberg equilibrium. ($p > 0.001$) (57). *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 genotypes and allele frequencies did not significantly differ between the PA and control groups. The results are shown in Table II.

Binomial logistic regression analysis was performed to estimate the impact of genotypes and alleles on PA development. Binomial logistic regression analysis of *RAD51B* rs2588809 revealed that the TT genotype was associated with about 7-fold increased odds of PA development in the co-dominant (OR=6.833; 95% CI=2.557-18.262; $p < 0.001$) and recessive (OR=7.066; 95% CI=2.667-

Table IV. *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 genotype and allele frequencies in PA patients and controls by gender.

Genotype/allele	Males		p-Value	Females		p-Value
	PA group n=45 n (%)	Control group n=100 n (%)		PA group n=68 n (%)	Control group n=183 n (%)	
<i>TBX15</i> rs984222						
GG	25 (55.6)	45 (45.0)	0.499	40 (58.8)	96 (52.5)	0.532
GC	17 (37.8)	47 (47.0)		25 (36.8)	73 (39.9)	
CC	3 (6.7)	8 (8.0)		3 (4.4)	14 (7.7)	
Allele						
G	67 (74.4)	137 (68.5)	0.305	105 (77.20)	265 (72.40)	0.277
C	23 (25.6)	63 (31.5)		31 (22.80)	101 (27.60)	
<i>DNM3</i> rs1011731						
AA	12 (26.7)	35 (35.0)	0.583	22 (32.4)	55 (30.1)	0.923
AG	25 (55.6)	51 (51.0)		32 (47.1)	91 (49.7)	
GG	8 (17.7)	14 (14.0)		14 (20.6)	37 (20.2)	
Allele						
A	49 (54.4)	121 (60.5)	0.333	76 (55.88)	201 (54.91)	0.847
G	41 (45.6)	79 (39.5)		60 (44.12)	165 (45.09)	
<i>RAD51B</i> rs8017304						
AA	19 (42.22)	38 (38.0)	0.890	30 (44.12)	92 (50.27)	0.340
AG	19 (42.22)	45 (45.0)		33 (48.53)	71 (38.25)	
GG	7 (15.56)	17 (17.0)		5 (7.35)	20 (11.48)	
Allele						
A	57 (63.33)	121 (60.5)	0.646	93 (68.38)	255 (69.67)	0.780
G	33 (36.67)	79 (39.5)		43 (31.62)	111 (30.33)	
<i>RAD51B</i> rs2588809						
CC	27 (60.0)	75 (75.0)	0.179	47 (69.12)	123 (67.21)	0.011
CT	13 (28.89)	19 (19.0)		11 (16.18)	51 (27.87)	
TT	5 (11.11)	6 (6.0)		10 (14.7)	9 (4.92)	
Allele						
C	67 (74.44)	169 (84.5)	0.042	105 (77.21)	297 (81.15)	0.325
T	23 (25.56)	31 (15.5)		31 (22.79)	69 (18.85)	

PA: Pituitary adenoma; p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant p-Values are shown in bold.

18.722; $p < 0.001$) models. Each copy of the T allele was associated with increased odds of PA development (OR=1.627; 95% CI=1.135-2.334; $p=0.008$) (Table III). Analysis of *TBX15* rs984222, *DNM3* rs1011731, and *RAD51B* rs8017304 did not show any statistically significant results (Supplementary material).

Comparison of TBX15 rs984222, DNM3 rs101173, RAD51B rs8017304, and rs2588809 polymorphisms in pituitary adenoma patients by gender. Statistical analysis was also performed to compare the *TBX15* rs984222, *DNM3* rs1011731, and *RAD51B* rs8017304 genotype and allele

frequencies between the patients with PA and control group subjects by their gender (Table IV). The analysis of *RAD51B* rs2588809 showed a statistically significant difference in the CC, CT, and TT genotype distributions between females with PA and control females (69.12%, 16.18%, and 14.7% vs. 67.21%, 27.87%, and 4.92%, respectively, $p=0.011$). The results are presented in Table IV.

Binominal logistic regression was performed to evaluate these polymorphisms' impact on the PA development in men and women, separately. Binominal logistic regression analysis in the women's group showed that the TT genotype was associated with 6.7-fold higher odds of PA development in

Table V. Binary logistic regression analysis of *RAD51B* rs2588809 in females.

Model	Genotype	OR (95% CI)	p-Value	AIC
<i>TBX15</i> rs984222				
<i>RAD51B</i> rs2588809				
Co-dominant	C/T	0.572 (0.275; 1.188)	0.134	282.117
	T/T	6.744 (2.021; 22.583)	0.002	
Dominant	C/T+T/T	1.013 (0.554; 1.852)	0.966	295.251
Recessive	T/T	7.716 (2.331; 25.533)	0.001	282.517
Overdominant	C/T	0.486 (0.236; 1.000)	0.050	291.045
Additive	T	1.425 (0.905; 2.245)	0.126	292.969

OR: Odds ratio; AIC: Akaike information criterion; p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant p-Values are shown in bold.

the co-dominant model (OR=6.744; 95% CI=2.021-22.583; $p=0.002$) and with 7.7-fold increased odds of PA development in the recessive model (OR=7.716; 95% CI=2.332-25.533; $p=0.001$). The results are shown in Table V. The *TBX15* rs984222, *DNM3* rs1011731, and *RAD51B* rs8017304 were not associated with female PA development (Supplementary material). Also, no statistically significant variables were found in the men's group (Supplementary material).

Association of TBX15 rs984222, DNM3 rs1011731, RAD51B rs8017304, and RAD51B rs2588809 polymorphisms with clinical and morphological features of PA. One of our study's objectives was to determine if there is a relationship between *TBX15*, *DNM3*, and *RAD51B* gene polymorphisms with PA's clinical and morphological features. Comparing the distribution of genotypes and alleles of *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 between the PA groups by recurrence and the control group, we obtained statistically significant differences in the rs2588809 CC, CT, and TT genotype distributions between PA without-recurrence patients and healthy controls (67.03%, 17.58% and 15.39% vs. 69.96%, 24.73%, and 5.31%, respectively; $p=0.005$). The results are shown in Table VI. Regarding PA recurrence, we performed binominal logistic regression to evaluate the impact of *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 polymorphisms on the development of PAs with and without recurrence. We found that the *RAD51B* rs2588809 TT genotype was associated with approximately 8-fold increased odds of development of PA without recurrence in the co-dominant (OR=7.842; 95% CI=2.890-21.277; $p < 0.001$) and recessive model (OR=8.394; 95% CI=3.122-22.571; $p < 0.001$). Also, each T allele was associated with 1.7-fold increased odds of development of PA without recurrence in the additive model (OR=1.676; 95% CI=0.114-2.457; $p=0.008$). The data

are presented in Table VII. No associations were found in the recurrent PA group (Supplementary material). The *TBX15* rs984222, *DNM3* rs1011731, and *RAD51B* rs8017304 were not associated with PA recurrence (Supplementary material).

TBX15 rs984222, *DNM3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 genotypes and allele frequencies were compared between the active and inactive PA and healthy control groups. We found that the *RAD51B* rs8017304 G allele was detected significantly more frequently in the inactive PA group vs. the control group (48.13% vs. 33.57%; $p=0.004$) (Table VIII).

Binominal logistic regression revealed that the *RAD51B* rs2588809 TT genotype was associated with increased odds of active PA development in the codominant (OR=6.058; 95% CI=2.146-19.734; $p=0.001$) and recessive (OR=7.103; 95% CI=2.366-21.320; $p < 0.001$) models (Table IX). Also, the *RAD51B* rs2588809 TT genotype was associated with increased odds of inactive PA development in the codominant (OR=7.247; 95% CI=2.29-22.906; $p=0.001$) and recessive (OR=7.260; 95% CI=2.260-21.840; $p=0.001$) models. Each T allele at rs2588809 was associated with 1.9-fold increased odds of inactive PA development in the additive model (OR=1.865; 95% CI=1.154-3.014; $p=0.011$). These data are presented in Table IX. The *TBX15* rs984222, *DNM3* rs1011731 and *RAD51B* rs8017304 were not associated with PA hormonal activity (Supplementary material).

We then compared the distribution of *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304, and rs2588809 genotypes and alleles in patients with invasive and non-invasive PAs vs. healthy controls. The *RAD51B* rs2588809 genotypes (CC, CT, and TT) were distributed significantly differently in patients with non-invasive PA and healthy subjects (59.09%, 22.72% and 18.19% vs. 69.96%, 24.73%, and 5.31%, respectively, $p=0.008$) (Table X). Also, the T allele occurred more frequently in patients with non-invasive PA than in control subjects (29.55% vs. 17.67%, $p=0.008$). The results are presented in Table X.

Binominal logistic regression was performed in patients with PA by its invasiveness. It was revealed that the *RAD51B* rs2588809 TT genotype was associated with about 5-fold increased odds of invasive PA in the codominant (OR=4.881; 95% CI=1.570-15.172; $p=0.006$) and recessive (OR=5.212; 95% CI=1.693-16.050; $p=0.004$) models (Table XI). Also, the *RAD51B* rs2588809 TT genotype was associated with increased odds of non-invasive PA development in the codominant (OR=10.513; 95% CI=3.381-32.688; $p < 0.001$) and recessive (OR=10.259; 95% CI=3.368-31.255; $p < 0.001$) models. Each T allele was associated with 2.2-fold increased odds of non-invasive PA development in the additive model (OR=2.222; 95% CI=1.352-3.652; $p=0.002$). The results are shown in Table XI. The *TBX15* rs984222, *DNM3* rs1011731, and *RAD51B* rs8017304 were not associated with PA invasiveness (Supplementary material).

Table VI. *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 genotype and allele frequencies in patients grouped by PA recurrence and healthy subjects.

SNP	Genotype/ Allele	Frequency					
		PA without recurrence n (%) n=91	Control group n (%) n=283	<i>p</i> -Value	PA with recurrence n (%) n=22	Control group n (%) n=283	<i>p</i> -Value
<i>TBX15</i> rs984222	GG	49 (53.85)	141 (49.82)	0.786	15 (68.18)	141 (49.82)	0.252
	GC	36 (39.56)	120 (42.40)		6 (27.27)	120 (42.40)	
	CC	6 (6.59)	22 (7.78)		1 (4.55)	22 (7.78)	
	Allele						
	G	134 (73.63)	402 (71.02)	0.498	36 (81.82)	402 (71.02)	0.125
	C	48 (26.37)	164 (28.98)		8 (18.18)	164 (28.98)	
<i>DNM3</i> rs1011731	AA	28 (30.77)	90 (31.80)	0.929	6 (27.27)	90 (31.80)	0.900
	AG	45 (49.45)	142 (50.18)		12 (54.55)	142 (50.18)	
	GG	18 (19.78)	51 (18.02)		4 (18.18)	51 (18.02)	
	Allele						
	A	101 (55.49)	322 (56.89)	0.741	24 (54.55)	322 (56.89)	0.762
	G	81 (44.51)	244 (43.11)		20 (45.45)	244 (43.11)	
<i>RAD51</i> rs8017304	AA	41 (45.05)	130 (45.94)	0.941	8 (36.36)	130 (45.94)	0.203
	AG	39 (42.86)	116 (40.99)		13 (59.09)	116 (40.99)	
	GG	11 (12.09)	37 (13.07)		1 (4.55)	37 (13.07)	
	Allele						
	A	121 (66.48)	376 (66.43)	0.989	29 (65.90)	376 (66.43)	0.943
	G	61 (33.52)	190 (33.57)		15 (34.09)	190 (33.57)	
<i>RAD51B</i> rs2588809	CC	61 (67.03)	198 (69.96)	0.005	13 (59.09)	198 (69.96)	0.484
	CT	16 (17.58)	70 (24.73)		8 (36.36)	70 (24.73)	
	TT	14 (15.39)	15 (5.31)		1 (4.55)	15 (5.31)	
	Allele						
	C	138 (75.82)	466 (82.33)	0.052	34 (77.27)	466 (82.33)	0.401
	T	44 (25.18)	100 (17.67)		10 (22.73)	100 (17.67)	

PA: Pituitary adenoma; *p*-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

Discussion

Our study analyzed the *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 gene polymorphisms in PA patients ($n=113$) and healthy control subjects ($n=283$). The results were compared by gender, age, and the clinical course of the disease. Studies of these polymorphisms analyzing PA association with rs984222, rs1011731, rs8017304, and rs2588809 have not been performed yet, to the best of our knowledge.

The role of TBX family genes (*TBX2* and *TBX3*) in oncogenic processes was associated with an increase of their expression level, as they have been found to be overexpressed in different types of cancer, including breast, cervical, ovarian, pancreatic, liver, and bladder cancer (58, 59). *TBX15* hypermethylation has been evaluated in prostate and ovarian carcinomas (19, 20). No studies have been performed in association with any brain tumors, including PA. Our study was the first to find that the C allele of *TBX15* rs984222 polymorphism reduced PA's recurrence ($p=0.037$).

Table VII. *RAD51B* rs2588809 association with PA without recurrence.

Model	Genotype	OR (95% CI)	<i>p</i> -Value	AIC
Co-dominant	C/T	0.747 (0.405; 1.378)	0.350	398.342
	T/T	7.842 (2.890; 21.277)		
Dominant	C/T+T/T	1.293 (0.777; 2.150)	0.323	418.081
Recessive	T/T	8.394 (3.122; 22.571)	<0.001	397.245
Overdominant Additive	C/T	0.637 (0.349; 1.164)	0.143	414.772
	T	1.676 (0.114; 2.457)		

OR: Odds ratio; AIC: Akaike information criterion; *p*-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

DNM3 has been shown to be involved in various malignancies (28-37). Marino *et al.* have reported *DNM3* expression in the brain and testicles and less often in the lungs and heart (29). Inokawa *et al.* and Shen *et al.* have found that *DNM3* is hypermethylated in hepatocellular cancer (HCC) (33-34). Zhang *et al.* have also studied the

Table VIII. *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 genotype and allele frequencies in patients grouped by PA hormonal activity and healthy subjects.

SNP	Genotype/ Allele	Frequency					
		Inactive PA group n=53 n (%)	Control group n=283 n (%)	<i>p</i> -Value	Active PA group n=60 n (%)	Control group n=283 n (%)	<i>p</i> -Value
<i>TBX15</i> rs984222	GG	30 (56.60)	141 (49.82)	0.636	35 (58.33)	141 (49.82)	0.252
	GC	20 (37.74)	120 (42.40)		22 (36.67)	120 (42.40)	
	CC	3 (5.6)	22 (7.78)		3 (5)	22 (7.78)	
	Allele						
	G	80 (75.47)	402 (71.02)	0.351	92 (76.67)	402 (71.02)	0.125
	C	26 (24.53)	164 (28.98)		28 (23.33)	164 (28.98)	
<i>DNMT3</i> rs1011731	AA	16 (30.19)	90 (31.81)	0.804	18 (30)	90 (31.81)	0.900
	AG	29 (54.72)	142 (50.17)		28 (46.67)	142 (50.17)	
	GG	8 (15.09)	51 (18.02)		14 (23.33)	51 (18.02)	
	Allele						
	A	61 (57.55)	322 (56.89)	0.900	64 (53.33)	322 (56.89)	0.762
	G	45 (42.45)	244 (43.11)		56 (46.67)	244 (43.11)	
<i>RAD51B</i> rs8017304	AA	24 (45.28)	130 (45.94)	0.996	25 (41.67)	130 (45.94)	0.358
	AG	22 (41.51)	116 (40.99)		30 (50.0)	116 (40.99)	
	GG	7 (13.21)	37 (13.07)		5 (8.33)	37 (13.07)	
	Allele						
	A	55 (51.87)	376 (66.43)	0.004	80 (66.67)	376 (66.43)	0.960
	G	51 (48.13)	190 (33.57)		40 (33.33)	190 (33.57)	
<i>RAD51B</i> rs2588809	CC	33 (62.26)	198 (69.96)	0.098	41 (68.33)	198 (69.96)	0.060
	CT	13 (24.53)	70 (24.73)		11 (18.33)	70 (24.73)	
	TT	7 (13.21)	15 (5.31)		8 (13.34)	15 (5.31)	
	Allele						
	C	79 (74.53)	466 (82.33)	0.059	93 (77.5)	466 (82.33)	0.215
	T	27 (25.47)	100 (17.67)		27 (22.5)	100 (17.67)	

p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

Table IX. *RAD51B* rs2588809 associations with PA hormonal activity.

Model	Genotype	OR (95% CI)	<i>p</i> -Value	AIC	Model	Genotype	OR (95% CI)	<i>p</i> -Value	AIC
<i>RAD51B</i> rs2588809					<i>RAD51B</i> rs2588809				
Active PA					Inactive PA				
Co-dominant	C/T	0.678 (0.323; 1.421)	0.303	309.171	Co-dominant	C/T	1.122 (0.559; 2.249)	0.746	286.264
	T/T	6.508 (2.146; 19.734)	0.001			T/T	7.247 (2.293; 22.906)	0.001	
Dominant	C/T+T/T	1.126 (0.612; 2.074)	0.702	319.894	Dominant	C/T+T/T	1.593 (0.862; 2.942)	0.137	292.775
Recessive	T/T	7.103 (2.366; 21.320)	<0.001	308.292	Recessive	T/T	7.260 (2.260; 21.840)	0.001	284.368
Overdominant	C/T	0.597 (0.288; 1.239)	0.166	317.966	Overdominant	C/T	0.970 (0.491; 1.917)	0.931	294.917
Additive	T	1.513 (0.949; 2.414)	0.082	317.159	Additive	T	1.865 (1.154; 3.014)	0.011	288.799

OR: Odds ratio; AIC: Akaike information criterion; *p*-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

mechanism of DNMT3 in HCC (60). Teicher *et al.* have reported liposarcoma 1q24.3 amplifications involving DNMT3 (29) while low DNMT2 expression has been associated with tumor invasion and metastasis in cervix carcinoma and up-

regulation of matrix metalloproteinase 2 (MMP-2) expression (61). The DNMT3 gene has also been investigated as a possible molecular marker for diagnosis and gene therapy of malignant diseases (38). Yang *et al.* have

Table X. *TBX15* rs984222, *DNM3* rs1011731, *RAD51B* rs8017304 and *RAD51B* rs2588809 genotype and allele frequencies in patients grouped by PA invasiveness and healthy subjects.

SNP	Genotype/ Allele	Frequency					
		Non-invasive PA group n=44 n (%)	Control group n=283 n (%)	<i>p</i> -Value	Invasive PA group n=69 n (%)	Control group n=283 n (%)	<i>p</i> -Value
<i>TBX15</i> rs984222	GG	26 (59.09)	141 (49.82)	0.300	39 (56.52)	141 (49.82)	0.600
	GC	17 (38.64)	120 (42.40)		25 (36.23)	120 (42.40)	
	CC	1 (2.27)	22 (7.78)		5 (7.25)	22 (7.78)	
	Allele						
	G	69 (78.41)	402 (71.02)	0.151	103 (74.64)	402 (71.02)	0.398
	C	19 (21.59)	164 (28.98)		35 (25.36)	164 (28.98)	
<i>DNM3</i> rs1011731	AA	14 (31.82)	90 (31.81)	0.732	20 (28.99)	90 (31.81)	0.868
	AG	20 (45.45)	142 (50.17)		37 (53.62)	142 (50.17)	
	GG	10 (27.73)	51 (18.02)		12 (17.39)	51 (18.02)	
	Allele						
	A	48 (54.55)	322 (56.89)	0.679	77 (55.79)	322 (56.89)	0.816
	G	40 (45.45)	244 (43.11)		61 (44.21)	244 (43.11)	
<i>RAD51B</i> rs8017304	AA	15 (34.09)	130 (45.94)	0.232	34 (49.28)	130 (45.94)	0.773
	AG	24 (54.55)	116 (40.99)		28 (40.58)	116 (40.99)	
	GG	5 (11.36)	37 (13.07)		7 (10.14)	37 (13.07)	
	Allele						
	A	54 (61.36)	376 (66.43)	0.351	96 (69.57)	376 (66.43)	0.482
	G	34 (38.64)	190 (33.57)		42 (30.43)	190 (33.57)	
<i>RAD51B</i> rs2588809	CC	26 (59.09)	198 (69.96)	0.008	48 (69.47)	198 (69.96)	0.280
	CT	10 (22.72)	70 (24.73)		14 (20.29)	70 (24.73)	
	TT	8 (18.19)	15 (5.31)		7 (10.14)	15 (5.31)	
	Allele						
	C	62 (70.45)	466 (82.33)	0.008	110 (79.71)	466 (82.33)	0.473
	T	26 (29.55)	100 (17.67)		28 (20.29)	100 (17.67)	

p-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

Table XI. *RAD51B* gene rs2588809 association with PA invasiveness.

Model	Genotype	OR (95% CI)	<i>p</i> -Value	AIC	Model	Genotype	OR (95% CI)	<i>p</i> -Value	AIC
<i>RAD51B</i> rs2588809					<i>RAD51B</i> rs2588809				
Inactive PA					Non-invasive PA				
Co-dominant	C/T	0.755 (0.387; 1.473)	0.410	343.800	Co-dominant	C/T	1.095 (0.503; 2.382)	0.819	248.474
	T/T	4.881 (1.570; 15.172)	0.006			T/T	10.513 (3.381; 32.688)	<0.001	
Dominant	C/T+T/T	1.073 (0.600; 1.919)	0.813	350.311	Dominant	C/T+T/T	1.820 (0.945; 3.503)	0.073	257.197
Recessive	T/T	5.212 (1.693; 16.050)	0.004	342.503	Recessive	T/T	10.259 (3.368; 31.255)	<0.001	244.526
Overdominant	C/T	0.693 (0.358; 1.342)	0.277	349.125	Overdominant	C/T	0.878 (0.413; 1.868)	0.736	260.187
Additive	T	1.357 (0.857; 2.148)	0.193	348.735	Additive	T	2.222 (1.352; 3.652)	0.002	250.945

OR: Odds ratio; AIC: Akaike information criterion; *p*-Value: Bonferroni corrected level of significance, differences are considered statistically significant when $p < 0.05/4$. Significant *p*-Values are shown in bold.

discussed the importance of the *DNM3* gene in gliomas. As the *DNM3* gene is the target of miR-221, the overexpression of *DNM3* could reverse its tumor-promoting effect (31-32). Based on these findings, we sought to examine whether a

polymorphism in the *DNM3* promoter could impact PA development risk. Unfortunately, in our study, we did not find any statistically significant differences analyzing *DNM3* rs1011731 gene polymorphism in relation to PA.

Concerning the other two gene polymorphisms, we found that the *RAD51B* rs2588809 CC genotype and the rs8017304 AG genotype might increase the probability of PA recurrence and invasiveness. Also, we proved that the *RAD51B* rs2588809 TT genotype might increase the odds of PA development in women and may be associated with PA development without recurrence. The *RAD51B* gene has previously been studied in other tumor types (breast, ovarian, and lung cancers (32, 41) but not in brain tumors, so we could not compare our results with the results of other authors.

RAD51B has been previously evaluated as a candidate gene for breast cancer predisposition, but no mutation was detected in a study of 188 multiple-case breast cancer families (62). Previous studies have identified chromosomal rearrangements disrupting *RAD51B* in benign tumors, particularly uterine leiomyomas (42, 43). In addition, the findings by Golmard and colleagues must be interpreted in the context of two genome-wide association studies (GWAS), which identified the minor allele of single nucleotide polymorphisms in *RAD51B* acting as a low-risk factor for breast cancer: the rs999737 (63) and rs1314913 (64), located in *RAD51B* introns 10 and 7, respectively. Results by Mengyin *et al.* also suggest that *RAD51B* could be a candidate prognostic factor for non-small cell lung cancer patients (41).

Overall, the present study of the *TBX15* rs984222, *DNMT3* rs1011731, *RAD51B* rs8017304, and *RAD51B* rs2588809 gene polymorphisms requires future replication in studies with higher sample sizes to confirm the association of *RAD51B* rs2588809 with PA.

Conclusion

The *RAD51B* rs2588809 TT genotype was more common in women with PA than in healthy women, and the T allele was less frequent in men with PA than in healthy men. The *RAD51B* rs2588809 T allele increased the potential for PA invasiveness and PA activity. The likelihood of PA recurrence was reduced by the TT genotype and each T allele.

Data Availability

The genotyping data used to support the findings of this study is available from the corresponding author upon request.

Supplementary Material

Available at: <https://docs.google.com/document/d/1U5Za-8j3e9mbHEye0LJIEcwVPL1f7-uSt7HlwRplqM/edit?usp=sharing>.

Conflicts of Interest

None of the Authors has any proprietary interests or conflicts of interest related to this submission.

Authors' Contributions

Conceptualization, R.L., and B.G.; Data curation, A.V., G.G., and B.G.; Writing-Original draft preparation, I.L., G.J., and R.L.; Methodology, A.V., G.G., B.G., L.K., and R.L.; Investigation, A.V., G.G., G.J., I.L., and L.K.; Validation, L.K., and R.L.; Supervision, R.L., and B.G.; Writing-Reviewing and Editing, R.L.

References

- 1 Mete O, Ezzat S and Asa SL: Biomarkers of aggressive pituitary adenomas. *Mol Endocrinol* 49(2): R69-78, 2012. PMID: 22822048. DOI: 10.1530/JME-12-0113
- 2 Drummond JB, Ribeiro-Oliveira A Jr and Soares BS: Non-Functioning Pituitary Adenomas. 2018 Nov 28. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, de Herder WW, Dungan K, Grossman A, Hershman JM, Hofland J, Kaltsas G, Koch C, Kopp P, Korbonits M, McLachlan R, Morley JE, New M, Purnell J, Singer F, Stratakis CA, Trencle DL, Wilson DP, editors. *Endotext* [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000-. PMID: 30521182.
- 3 Raverot G, Jouanneau E and Trouillas J: Clinicopathological classification and molecular markers of pituitary tumours for personalized therapeutic strategies. *Eur J Endocrinol* 170(4): R121-132, 2014. PMID: 24431196. DOI: 10.1530/EJE-13-1031
- 4 Kovacs K, Horvath E and Vidal S: Classification of pituitary adenomas. *J Neuro-Oncol* 54(2): 121-127, 2001. PMID: 11761429. DOI: 10.1023/a:1012945129981
- 5 Altay T, Krisht KM and Couldwell WT: Sellar and parasellar metastatic tumors. *Int J Surg Oncol* 2012: 647256, 2012. PMID: 22312541. DOI: 10.1155/2012/647256
- 6 Vandeva S, Jaffrain-Rea M, Daly AF, Tichomirowa M, Zacharieva S and Beckers A: The genetics of pituitary adenomas. *Best Pract Res Clin Endocrinol Metab* 24(3): 461-476, 2010. PMID: 20833337. DOI: 10.1016/j.beem.2010.03.001
- 7 Dworakowska D and Grossman AB: The pathophysiology of pituitary adenomas. *Best Pract Res Clin Endocrinol Metab* 23(5): 525-541, 2011. PMID: 19945021. DOI: 10.1016/j.beem.2009.05.004
- 8 Lake MG, Krook LS and Cruz SV: Pituitary adenomas: an overview. *Am Fam Physician* 88(5): 319-327, 2013. PMID: 24010395.
- 9 Gruppeta M, Mercieca C and Vassallo J: Prevalence and incidence of pituitary adenomas: a population based study in Malta. *Pituitary* 16(4): 545-553, 2013. PMID: 23239049. DOI: 10.1007/s11102-012-0454-0.
- 10 Agustsson TT, Baldvinsdottir T, Jonasson JG, Olafsdottir E, Steinhorsdottir V, Sigurdsson G, Thorsson AV, Carroll PV, Korbonits M and Benediktsson R: The epidemiology of pituitary adenomas in Iceland, 1955-2012: a nationwide population-based study. *Eur J Endocrinol* 173(5): 655-664, 2015. PMID: 26423473. DOI: 10.1530/EJE-15-0189
- 11 Day PF, Loto MG, Glerean M, Picasso MF, Lovazzano S and Giunta DH: Incidence and prevalence of clinically relevant pituitary adenomas: retrospective cohort study in a Health Management Organization in Buenos Aires, Argentina. *Arch Endocrinol Metab* 60(6): 554-561, 2016. PMID: 27982201. DOI: 10.1590/2359-399700000195
- 12 Bertolossi M, Linta L, Seufferlein T, Kleger A and Liebau S: A fresh look on T-Box factor action in early embryogenesis (T-Box

- factors in early development). *Stem Cells Dev* 24(16): 1833-1851, 2015. PMID: 25952667. DOI: 10.1089/scd.2015.0102
- 13 Destrieux C, Kakou MK, Velut S, Lefrancq T and Jan M: Microanatomy of the hypophyseal fossa boundaries. *J Neurosurg* 88: 743-752, 1998. PMID: 9525722. DOI: 10.3171/jns.1998.88.4.0743
 - 14 Harris FS and Rhoton AL: Anatomy of the cavernous sinus: a microsurgical study. *J Neurosurg* 45: 169-180, 1976. PMID: 939976. DOI: 10.3171/jns.1976.45.2.0169.
 - 15 Glebauskienė B, Liutkeviciene R, Zlatkute E, Kriauciuniene L and Zaliuniene D: Association of retinal nerve fibre layer thickness with quantitative magnetic resonance imaging data of the optic chiasm in pituitary adenoma patients. *J Clin Neurosci* 50: 1-6, 2018. PMID: 29398198. DOI: 10.1016/j.jocn.2018.01.005
 - 16 Ferrante E, Ferraroni M, Castrignanò T, Menicatti L, Anagni M, Reimondo G, Del Monte P, Bernasconi D, Loli P, Faustini-Fustini M, Borretta G, Terzolo M, Losa M, Morabito A, Spada A, Beck-Peccoz P and Lania AG: Non-functioning pituitary adenoma database: a useful resource to improve the clinical management of pituitary tumors. *Eur J Endocrinol* 155(6): 823-829, 2006. PMID: 17132751. DOI: 10.1530/eje.1.02298
 - 17 Mickevicius T, Vilkeviciute A, Glebauskienė B, Kriauciuniene L and Liutkeviciene R: Do TRIB1 and IL-9 gene polymorphisms impact the development and manifestation of pituitary adenoma? *In Vivo* 34(5): 2499-2505, 2020. PMID: 32871778. DOI: 10.21873/invivo.12066.
 - 18 Arribas J, Giménez E, Marcos R and Velázquez A: Novel antiapoptotic effect of *TBX15*: overexpression of *TBX15* reduces apoptosis in cancer cells. *Apoptosis* 20(10): 1338-1346, 2015. PMID: 26216026. DOI: 10.1007/s10495-015-1155-8
 - 19 Kron K, Pethe V, Briollais L, Sadikovic B, Ozelic H, Sunderji A, Venkateswaran V, Pinthus J, Fleshner N, van der Kwast T and Bapat B: Discovery of novel hypermethylated genes in prostate cancer using genomic CpG island microarrays. *PLoS One* 4(3): e4830, 2009. PMID: 19283074. DOI: 10.1371/journal.pone.0004830
 - 20 Gozzi G, Chelbi ST, Manni P, Alberti L, Fonda S, Saponaro S, Fabbiani L, Rivasi F, Benhattar J and Losi L: Promoter methylation and downregulated expression of the *TBX15* gene in ovarian carcinoma. *Oncol Lett* 12(4): 2811-2819, 2016. PMID: 27698863. DOI: 10.3892/ol.2016.5019
 - 21 Peres J, Davis E, Mowla S, Bennett DC, Li JA, Wansleben S and Prince S: The highly homologous T-box transcription factors, *TBX2* and *TBX3*, have distinct roles in the oncogenic process. *Genes Cancer* 1(3): 272-282, 2010. PMID: 21779450. DOI: 10.1177/1947601910365160
 - 22 Yu J, Ma X, Cheung KF, Li X, Tian L, Wang S, Wu CW, Wu WK, He M, Wang M, Ng SS and Sung JJ: Epigenetic inactivation of T-box transcription factor 5, a novel tumor suppressor gene, is associated with colon cancer. *Oncogene* 29(49): 6464-74, 2010. PMID: 20802524. DOI: 10.1038/onc.2010.370
 - 23 Papaioannou VE: The T-box gene family: emerging roles in development, stem cells and cancer. *Development* 141(20): 3819-3833, 2014. PMID: 25294936. DOI: 10.1242/dev.104471
 - 24 Kron K, Liu L, Trudel D, Pethe V, Trachtenberg J, Fleshner N, Bapat B and van der Kwast T: Correlation of ERG expression and DNA methylation biomarkers with adverse clinicopathologic features of prostate cancer. *Clin Cancer Res* 18(10): 2896-2904, 2012. PMID: 22452941. DOI: 10.1158/1078-0432.CCR-11-2901
 - 25 Pacifico F and Leonardi A: Role of NF-kappaB in thyroid cancer. *Mol Cell Endocrinol* 321(1): 29-35, 2010. PMID: 19879919. DOI: 10.1016/j.mce.2009.10.010
 - 26 Xing M: Molecular pathogenesis and mechanisms of thyroid cancer. *Nat Rev Cancer* 13(3): 184-199, 2013. PMID: 23429735; PMCID: DOI: 10.1038/nrc3431
 - 27 Orth JD and McNiven MA: Dynamin at the actin-membrane interface. *Curr Opin Cell Biol* 15(1): 31-39, 2003. PMID: 12517701. DOI: 10.1016/s0955-0674(02)00010-8
 - 28 Booken N, Gratchev A, Utikal J, Weiss C, Yu X, Qadoumi M, Schmith M, Sepp N, Nashan D, Rass K, Tüting T, Assaf C, Dippel E, Stadler R, Klemke CD and Goerdts S: Sézary syndrome is a unique cutaneous T-cell lymphoma as identified by an expanded gene signature including diagnostic marker molecules *CDO1* and *DNM3*. *Leukemia* 22(2): 393-399, 2008. PMID: 18033314. DOI: 10.1038/sj.leu.2405044
 - 29 Teicher BA: Searching for molecular targets in sarcoma. *Biochem Pharmacol* 84(1): 1-10, 2012. PMID: 22387046. DOI: 10.1016/j.bcp.2012.02.009
 - 30 Marino N, Collins JW, Shen C, Caplen NJ, Merchant AS, Gökmen-Polar Y, Goswami CP, Hoshino T, Qian Y, Sledge GW Jr. and Steeg PS: Identification and validation of genes with expression patterns inverse to multiple metastasis suppressor genes in breast cancer cell lines. *Clin Exp Metastasis* 31(7): 771-786, 2014. PMID: 25086928. DOI: 10.1007/s10585-014-9667-0
 - 31 Yang JK, Yang JP, Tong J, Jing SY, Fan B, Wang F, Sun GZ and Jiao BH: Exosomal miR-221 targets *DNM3* to induce tumor progression and temozolomide resistance in glioma. *J Neurooncol* 131(2): 255-265, 2017. PMID: 27837435. DOI: 10.1007/s11060-016-2308-5
 - 32 Yang JK, Song J, Huo HR, Zhao YL, Zhang GY, Zhao ZM, Sun GZ and Jiao BH: *DNM3*, p65 and p53 from exosomes represent potential clinical diagnosis markers for glioblastoma multiforme. *Ther Adv Med Oncol* 9(12): 741-754, 2017. PMID: 29449895. DOI: 10.1177/1758834017737471
 - 33 Inokawa Y, Nomoto S, Hishida M, Hayashi M, Kanda M, Nishikawa Y, Takeda S, Fujiwara M, Koike M, Sugimoto H, Fujii T, Nakayama G, Yamada S, Tanaka C, Kobayashi D and Kodera Y: Dynamin 3: a new candidate tumor suppressor gene in hepatocellular carcinoma detected by triple combination array analysis. *Oncotargets Ther* 6: 1417-1424, 2013. PMID: 24143113. DOI: 10.2147/OTT.S51913
 - 34 Shen J, Wang S, Zhang YJ, Kappil M, Wu HC, Kibriya MG, Wang Q, Jasmine F, Ahsan H, Lee PH, Yu MW, Chen CJ and Santella RM: Genome-wide DNA methylation profiles in hepatocellular carcinoma. *Hepatology* 55(6): 1799-1808, 2012. PMID: 22234943. DOI: 10.1002/hep.25569
 - 35 Gu C, Yao J and Sun P: Dynamin 3 suppresses growth and induces apoptosis of hepatocellular carcinoma cells by activating inducible nitric oxide synthase production. *Oncol Lett* 13(6): 4776-4784, 2017. PMID: 28599479. DOI: 10.3892/ol.2017.6057
 - 36 Ma Y, Guan L, Han Y, Zhou Y, Li X, Liu Y, Zhang X, Zhang W, Li X, Wang S and Lu W: siPRDX2-elevated *DNM3* inhibits the proliferation and metastasis of colon cancer cells *via* *AKT* signaling pathway. *Cancer Manag Res* 11: 5799-5811, 2019. PMID: 31388312. DOI: 10.2147/CMAR.S193805
 - 37 Lin S, Tan L, Luo D, Peng X, Zhu Y and Li H: Linc01278 inhibits the development of papillary thyroid carcinoma by regulating miR-376c-3p/*DNM3* axis. *Cancer Manag Res* 11: 8557-8569, 2019. PMID: 31572010. DOI: 10.2147/CMAR.S217886

- 38 Zhang HJ, Yuan GL, Liang QL, Peng X, Cheng SA, Jiang L, Liu Q, Zhang XC, Huang Z and Zeng Y: Progress of dynamin 3 in tumors. *Int J Clin Exp Med* 10(11): 15060-15063, 2017.
- 39 Jiang L, Liang QL, Liang WM, Zhang HJ, Huang J, Yuan GL, Peng XX, Cheng SA, Huang ZG and Zhang XN: Construction of a recombinant eukaryotic expression vector containing DNMT3 gene and its expression in colon cancer cells. *Onco Targets Ther* 11: 6665-6671, 2017. PMID: 30349300. DOI: 10.2147/OTT.S176388
- 40 Meindl A, Hellebrand H, Wiek C, Erven V, Wappenschmidt B, Niederacher D, Freund M, Lichtner P, Hartmann L, Schaal H, Ramser J, Honisch E, Kubisch C, Wichmann HE, Kast K, Deissler H, Engel C, Müller-Myhsock B, Neveling K, Kiechle M, Mathew CG, Schindler D, Schmutzler RK and Hanenberg H: Germline mutations in breast and ovarian cancer pedigrees establish *RAD51C* as a human cancer susceptibility gene. *Nat Genet* 42(5): 410-414, 2010. PMID: 20400964. DOI: 10.1038/ng.569
- 41 Wu M, Sheng Z, Jiang L, Liu Z, Bi Y and Shen Y: Overexpression of RAD51B predicts a preferable prognosis for non-small cell lung cancer patients. *Oncotarget* 8: 91471-91480, 2017. PMID: 29207658. DOI: 10.18632/oncotarget.20676
- 42 Heim S, Nilbert M, Vanni R, Floderus UM, Mandahl N, Liedgren S, Lecca U and Mitelman F: A specific translocation, t(12;14)(q14-15;q23-24), characterizes a subgroup of uterine leiomyomas. *Cancer Genet Cytogenet* 32: 13-17, 1988. PMID: 3355995. DOI: 10.1016/0165-4608(88)90305-6
- 43 Schoenmakers EF, Huysmans C and Van de Ven WJ: Allelic knockout of novel splice variants of human recombination repair gene RAD51B in t(12;14) uterine leiomyomas. *Cancer Res* 59(1): 19-23, 1999. PMID: 9892177.
- 44 Nagathihalli NS and Nagaraju G: RAD51 as a potential biomarker and therapeutic target for pancreatic cancer. *Biochim Biophys Acta* 1816(2): 209-218, 2011. PMID: 21807066. DOI: 10.1016/j.bbcan.2011.07.004
- 45 Thacker J: The RAD51 gene family, genetic instability and cancer. *Cancer Lett* 219(2): 125-135, 2005. PMID: 15723711. DOI: 10.1016/j.canlet.2004.08.018
- 46 Zhang X, Ma N, Yao W, Li S and Ren Z: RAD51 is a potential marker for prognosis and regulates cell proliferation in pancreatic cancer. *Cancer Cell Int* 19: 356, 2019. PMID: 31889908. DOI: 10.1186/s12935-019-1077-6
- 47 Maacke H, Jost K, Opitz S, Miska S, Yuan Y, Hasselbach L, Lüttges J, Kalthoff H and Stürzbecher HW: DNA repair and recombination factor Rad51 is over-expressed in human pancreatic adenocarcinoma. *Oncogene* 19(23): 2791-2795, 2000. PMID: 10851081. DOI: 10.1038/sj.onc.1203578
- 48 Nowacka-Zawisza M, Wiśnik E, Wasilewski A, Skowrońska M, Forma E, Bryś M, Rózański W and Krajewska WM: Polymorphisms of homologous recombination RAD51, RAD51B, XRCC2, and XRCC3 genes and the risk of prostate cancer. *Anal Cell Pathol (Amst)* 2015: 828646, 2015. PMID: 26339569. DOI: 10.1155/2015/828646
- 49 Nowacka-Zawisza M, Raszkievicz A, Kwasiborski T, Forma E, Bryś M, Rózański W and Krajewska WM: RAD51 and XRCC3 polymorphisms are associated with increased risk of prostate cancer. *J Oncol* 2019: 2976373, 2019. PMID: 31186630. DOI: 10.1155/2019/2976373
- 50 Krumm A, Barckhausen C, Kucuk P, Tomaszowski KH, Loquai C, Fahrer J, Krämer OH, Kaina B and Roos WP: Enhanced histone deacetylase activity in malignant melanoma provokes RAD51 and FANCD2-triggered drug resistance. *Cancer Res* 76(10): 3067-3077, 2016. PMID: 26980768. DOI: 10.1158/0008-5472.CAN-15-2680
- 51 Tennstedt P, Fresow R, Simon R, Marx A, Terracciano L, Petersen C, Sauter G, Dikomey E and Borgmann K: RAD51 overexpression is a negative prognostic marker for colorectal adenocarcinoma. *Int J Cancer* 132(9): 2118-2126, 2013. PMID: 23065657. DOI: 10.1002/ijc.27907
- 52 Michalska MM, Samulak D, Romanowicz H and Smolarz B: Association of polymorphisms in the 5' untranslated region of RAD51 gene with risk of endometrial cancer in the Polish population. *Arch Gynecol Obstet* 290(5): 985-991, 2014. PMID: 24930116. DOI: 10.1007/s00404-014-3305-6
- 53 Hannay JA, Liu J, Zhu QS, Bolshakov SV, Li L, Pisters PW, Lazar AJ, Yu D, Pollock RE and Lev D: Rad51 overexpression contributes to chemoresistance in human soft tissue sarcoma cells: a role for p53/activator protein 2 transcriptional regulation. *Mol Cancer Ther* 6(5): 1650-1660, 2007. PMID: 17513613. DOI: 10.1158/1535-7163.MCT-06-0636
- 54 Welsh JW, Ellsworth RK, Kumar R, Fjerstad K, Martinez J, Nagel RB, Eschbacher J and Stea B: Rad51 protein expression and survival in patients with glioblastoma multiforme. *Int J Radiat Oncol Biol Phys* 74(4): 1251-1255, 2009. PMID: 19545791. DOI: 10.1016/j.ijrobp.2009.03.018
- 55 Sidaraite A, Liutkeviciene R, Glebauskiene B, Vilkeviciute A and Kriauciuniene L: Associations of cholesteryl ester transfer protein (CETP) gene variants with pituitary adenoma. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 164(2): 189-195, 2020. PMID: 31012439. DOI: 10.5507/bp.2019.016
- 56 Liutkeviciene R, Vilkeviciute A, Morkunaite G, Glebauskiene B and Kriauciuniene L: SIRT1 (rs3740051) role in pituitary adenoma development. *BMC Med Genet* 20(1): 185, 2019. PMID: 31747893. DOI: 10.1186/s12881-019-0892-x
- 57 Donaldson P, Daly A, Ermini L and Bevitt D: Genetics of Complex Disease, 1st edition. Garland Science, Taylor Francis Group, pp. 151, 2015.
- 58 Rowley M, Grothey E and Couch FJ: The role of Tbx2 and Tbx3 in mammary development and tumorigenesis. *J Mammary Gland Biol Neoplasia* 9: 109-118, 2004. PMID: 15300007. DOI: 10.1023/B:JOMG.0000037156.64331.3f
- 59 Fan W, Huang X, Chen C, Gray J and Huang T: TBX3 and its isoform TBX3+2a are functionally distinctive in inhibition of senescence and are overexpressed in a subset of breast cancer cell lines. *Cancer Res* 64: 5132-5139, 2004. PMID: 15289316. DOI: 10.1158/0008-5472.CAN-04-0615
- 60 Zhang Z, Chen C, Guo W, Zheng S, Sun Z and Geng X: DNMT3 attenuates hepatocellular carcinoma growth by activating p53. *Med Sci Monit* 22(1): 197-205, 2016. PMID: 26784388. DOI: 10.12659/msm.896545
- 61 Lee YY, Do IG, Park YA, Choi JJ, Song SY, Kim CJ, Kim MK, Song TJ, Park HS, Choi CH, Kim TJ, Kim BG, Lee JW and Bae DS: Low dynamin 2 expression is associated with tumor invasion and metastasis in invasive squamous cell carcinoma of cervix. *Cancer Biol Ther* 10(4): 329-335, 2010. PMID: 20574164. DOI: 10.4161/cbt.10.4.12275
- 62 Johnson J, Healey S, Khanna KK and Chenevix-Trench G: Mutation analysis of *RAD51L1 (RAD51B/REC2)* in multiple-case, non-*BRCA1/2* breast cancer families. *Breast Cancer Res Treat* 129: 255-263, 2011. PMID: 21533530. DOI: 10.1007/s10549-011-1539-6

- 63 Thomas G, Jacobs KB, Kraft P, Yeager M, Wacholder S, Cox DG, Hankinson SE, Hutchinson A, Wang Z, Yu K, Chatterjee N, Garcia-Closas M, Gonzalez-Bosquet J, Prokunina-Olsson L, Orr N, Willett WC, Colditz GA, Ziegler RG, Berg CD, Buys SS, McCarty CA, Feigelson HS, Calle EE, Thun MJ, Diver R, Prentice R, Jackson R, Kooperberg C, Chlebowski R, Lissowska J, Peplonska B, Brinton LA, Sigurdson A, Doody M, Bhatti P, Alexander BH, Buring J, Lee IM, Vatten LJ, Hveem K, Kumle M, Hayes RB, Tucker M, Gerhard DS, Fraumeni JF Jr, Hoover RN, Chanock SJ and Hunter DJ: A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 14q24.1 (*RAD51LI*). *Nat Genet* 41(5): 579-584, 2009. PMID: 19330030. DOI: 10.1038/ng.353
- 64 Orr N, Lemnrau A, Cooke R, Fletcher O, Tomczyk K, Jones M, Johnson N, Lord CJ, Mitsopoulos C, Zvelebil M, McDade SS, Buck G, Blancher C; KConFab Consortium, Trainer AH, James PA, Bojesen SE, Bokmand S, Nevanlinna H, Mattson J, Friedman E, Laitman Y, Palli D, Masala G, Zanna I, Ottini L, Giannini G, Hollestelle A, Ouweland AM, Novaković S, Krajc M, Gago-Dominguez M, Castelao JE, Olsson H, Hedenfalk I, Easton DF, Pharoah PD, Dunning AM, Bishop DT, Neuhausen SL, Steele L, Houlston RS, Garcia-Closas M, Ashworth A and Swerdlow AJ: Genome-wide association study identifies a common variant in *RAD51B* associated with male breast cancer risk. *Nat Genet* 44(11): 1182-1184, 2012. PMID: 23001122. DOI: 10.1038/ng.2417

Received November 5, 2020

Revised January 7, 2021

Accepted January 11, 2021