

ORIGINAL ARTICLE

Genomic profiling of multifocal intrahepatic cholangiocarcinoma reveals intraindividual concordance of genetic alterations

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Abstract

In multifocal intrahepatic cholangiocarcinoma (IHC), intrahepatic metastases (IM) represent a contraindication to surgical resection, whereas satellite nodules (SN) do not. However, no consensus criteria exist to distinguish IM from SN. The purpose of this study was to determine genetic alterations and clonal relationships in surgically resected multifocal IHC. Next-generation sequencing of 34 spatially separated IHC tumors was performed using a targeted panel of 201 cancer-associated genes. Proposed definitions in the literature were applied of SN located in the same liver segment and ≤ 2 cm from the primary tumor; and IM located in a different liver segment and/or > 2 cm from the primary tumor. Somatic point mutations concordant across tumors from individual patients included BAP1, SMARCA4 and IDH1. Small insertions and deletions (indels) present at the same genome positions among all tumors from individuals included indels in DNA repair genes, CHEK1, ERCC5, ATR and MSH6. Copy number alterations were also similar between all tumors in each patient. In this cohort of multifocal IHC, genomic profiles were concordant across all tumors in each patient, suggesting a common progenitor cell origin, regardless of the location of tumors in the liver. The decision to perform surgery should not be based upon a perceived distinction between IM and SN.

Introduction

Intrahepatic cholangiocarcinoma (IHC) is the second most common primary liver cancer, following hepatocellular carcinoma, and has a rising incidence globally (1). Surgery remains the only potentially curative treatment, with 5-year overall survival rates between 20 and 35% (2). However, two-thirds of patients suffer postoperative disease recurrence, most commonly in the remnant liver. Approximately 30% of patients undergo resection of multiple anatomically separate tumors and have significantly

worse survival compared with patients undergoing resection of solitary tumors (1,3). In multifocal IHC, individual tumors are thought to represent disseminated metastases or regional extensions of the primary tumor. Peritumoral satellite nodules (SN) are considered potentially resectable, whereas intrahepatic metastases (IM) represent a contraindication to surgical resection (4,5). However, radiologic or pathologic criteria that distinguish between regional SN and disseminated IM are lacking.

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Abbreviations

CNA	copy number alteration
IHC	intrahepatic cholangiocarcinoma
IM	intrahepatic metastases
SN	satellite nodules

Previous reports have proposed classifying SN and IM based on tumor location and distribution in the liver (6–9). To our knowledge, no prior study has examined molecular differences between SN and IM. To address this gap, we aimed to determine whether IM and SN are genomically distinct entities to inform clinical decision-making. Here, we report that somatic point mutations, copy number alterations (CNAs) and small insertions and deletions (indels) are concordant across all tumors in individual patients, indicative of a shared origin from the same initiating cell. These findings suggest that multiple intrahepatic tumors in IHC represent metastases, regardless of their location in the liver.

Materials and methods

Patient samples

Thirty-four tumor samples and nine matched control samples of adjacent non-tumoral liver were collected from nine patients who underwent resection of multifocal IHC at the University of Texas MD Anderson Cancer Center from 2010 to 2017. Hematoxylin and eosin stained slides were reviewed by a gastrointestinal pathologist to determine the proportion of malignant cells relative to necrosis, fibrosis and inflammatory infiltrate. Five μm thick sections were prepared from formalin-fixed, paraffin-embedded tissue. Slides were deparaffinized, and tissue was scraped and placed in Eppendorf tubes. Genomic DNA was extracted from all samples as described previously (10).

Tumor number and size were recorded from pathology reports. The collection and analysis of patient samples were approved by the Institutional Review Board.

Intrahepatic metastases versus satellite nodules

Preoperative radiologic imaging and surgical pathology reports were used to classify SN and IM, based on a proposed definition in the literature (6,11). SN were defined as smaller lesions located in the same liver segment, ≤ 2 cm from the larger primary tumor (Figure 1). IM were defined as lesions in a different liver segment and/or > 2 cm from the primary tumor.

Targeted DNA sequencing

T200 is a deep targeted sequencing platform that comprises 4874 exons encoding 938 607 bases, with median coverage of 300 reads in 201 cancer-related genes (10). This platform was chosen because it has been optimized for formalin-fixed, paraffin-embedded specimens and reliably detects low-frequency mutations and CNAs with a false discovery rate of 1%. The 201 genes were selected for their biological relevance in cancer based on mutational data in the Catalogue of Somatic Mutations in Cancer and The Cancer Genome Atlas.

Sequence alignment and variant calling

Sequencing data were mapped to the human reference sequence GRCh37 (hg19) using the Burrows-Wheeler Aligner on default settings. Aligned reads were further processed following the Genome Analysis Toolkit Best Practices of duplicate removal, indels realignment and base recalibration. Platypus was used to call SNVs and indels. The MuTect algorithm was used to identify somatic mutations by comparing tumors with matched non-tumoral liver samples. Indels were identified using Pindel (12). In addition to build-in filters, the following filtering criteria were applied: (i) variant allele frequency in tumor DNA $> 5\%$; (ii) sequence depth > 10 ; (iii) presence of variant on both strands; (iv) variants rare in the population, with allele frequency < 0.01 in the Exome Aggregation Consortium Data and (v) removal of variants in positions

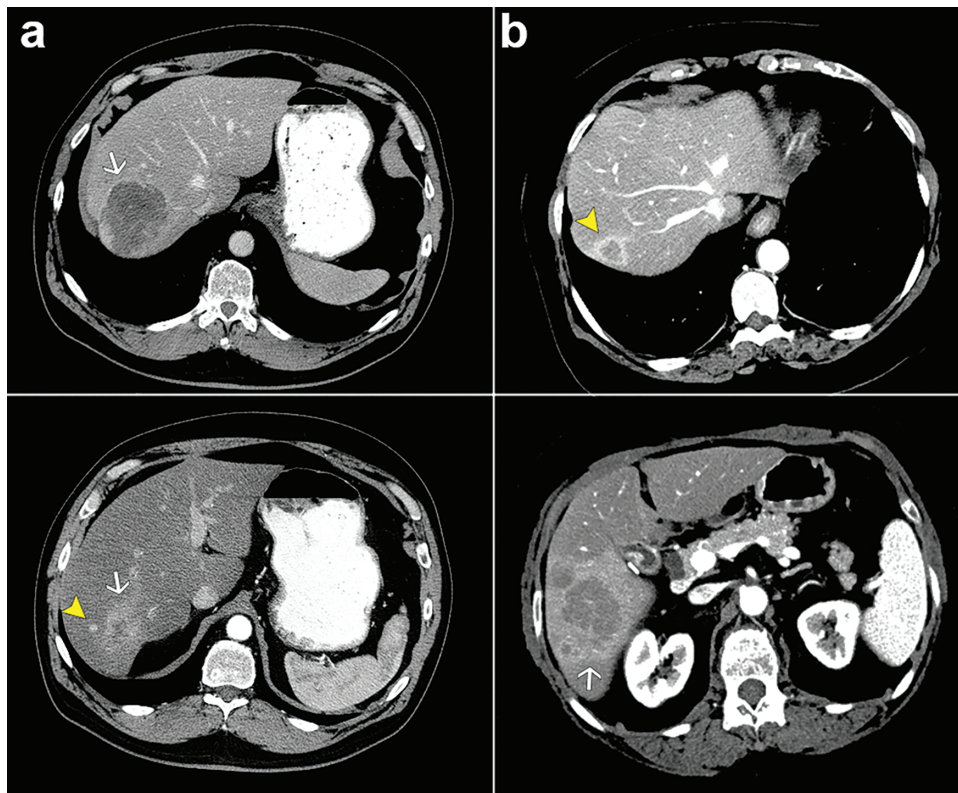


Figure 1. Representative preoperative computed tomography images showing main mass (white arrows) and smaller tumors (yellow arrowheads). (a) SN, patient 1. (b) IM, patient 9.

listed in the dbSNP 138 database. Substitutions and indels were annotated using Annotate Variation software (ANNOVAR) based on the University of Santa Cruz (USCS) Known Genes (<http://genome.ucsc.edu>).

Jaccard Index

Similarities in SNVs among samples from individual patients and between samples in different patients were calculated with the Jaccard

Index using the R package set (<https://cran.rproject.org/web/packages/sets>). All SNVs in 201 cancer-related genes were selected to estimate similarities.

Copy number alterations

Tumor and adjacent non-tumoral DNA was used to obtain tumor-specific, somatic CNAs from T200 data using an in-house R package. The log2 ratios

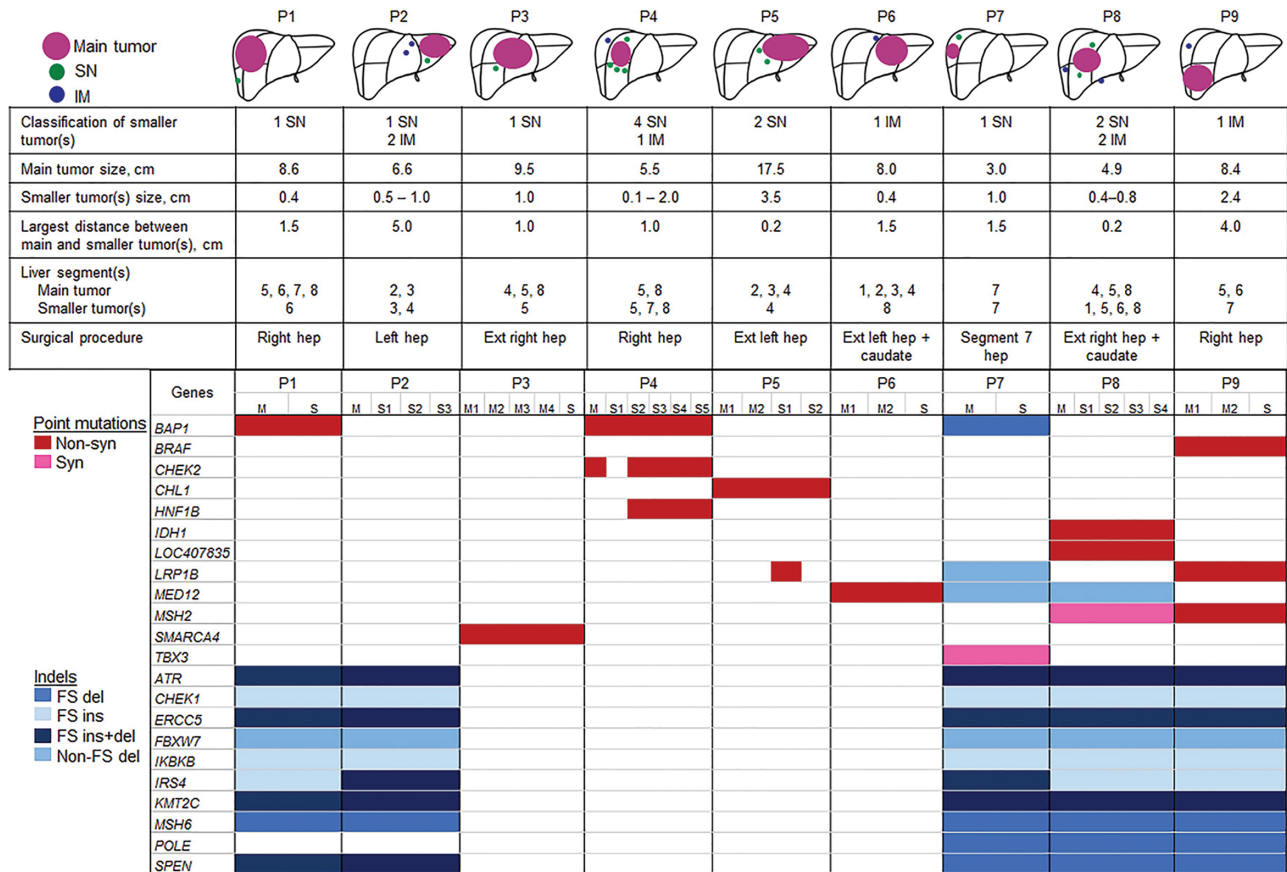


Figure 2. Schematic representations, clinicopathologic factors and mutation heat map of nine patients undergoing resection of multifocal IHC. Mutated genes comprise rows, and columns represent patient samples. The presence and type of mutations are indicated by different colors, as shown, and the absence of mutation indicated by white. del, deletion; ext, extended; FS, frameshift; hep, hepatectomy; indel, small insertion and deletion; ins, insertion; M, main tumor; P, patient; S, smaller tumor(s); syn, synonymous.

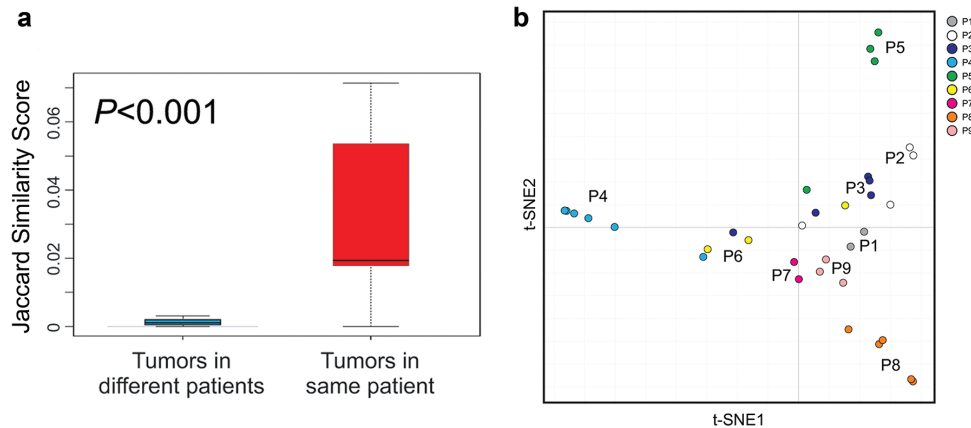


Figure 3. Concordance of single-nucleotide variants and CNAs across tumors from individual patients. (a) Jaccard similarity plot based on single-nucleotide variants showing that tumors within the same patient are similar, whereas tumors between patients are distinct. (b) Data from CNAs from all samples in each patient are plotted with t-SNE, a graph layout algorithm that gives each data point a location in a multidimensional map.

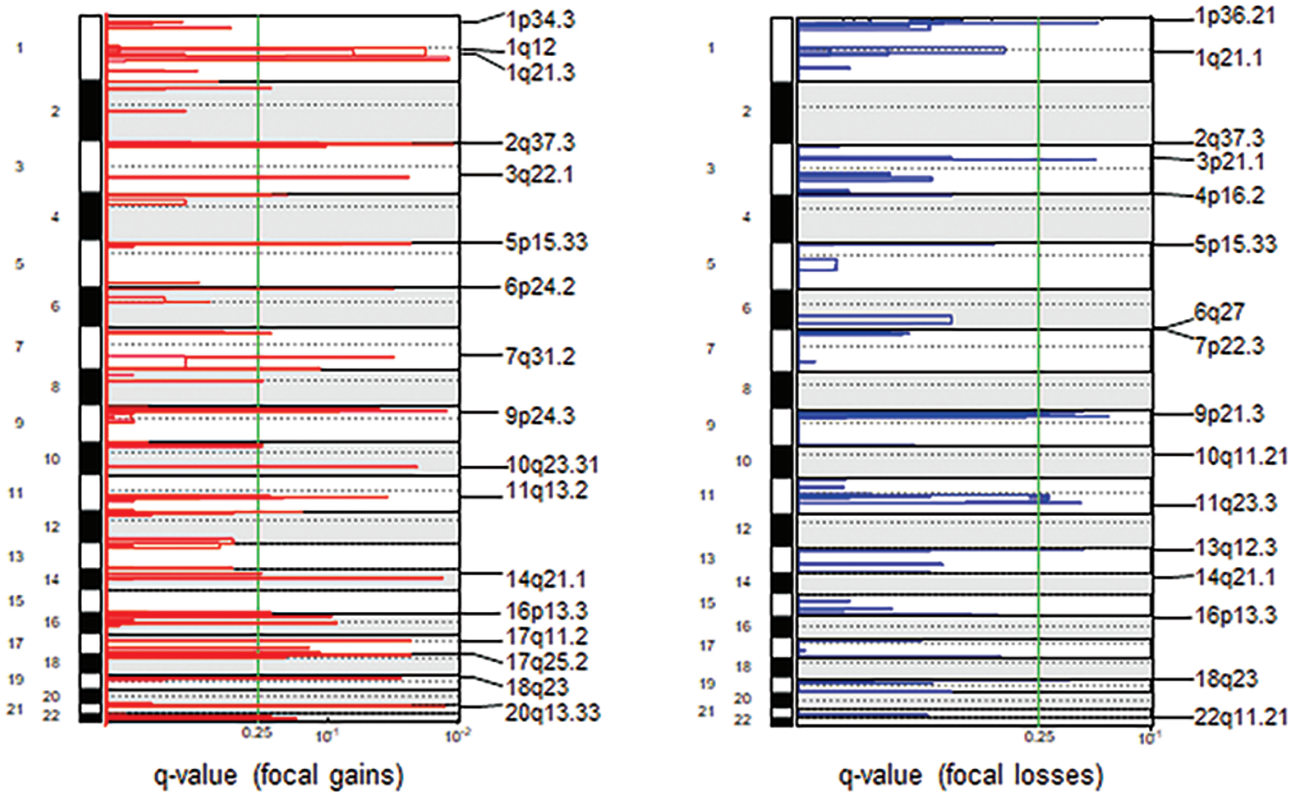


Figure 5. GISTIC2.0 analysis of CNAs for multiple samples in each patient. Regions of amplifications or deletions are designated with their chromosomal cytogenetic band.

The Jaccard Index, which measures similarity and diversity of sample sets, showed that mutational profiles were similar among multiple samples from the same individual and heterogeneous between patients (Figure 3a).

Copy number alterations

To investigate the clonal relationship between lesions, somatic CNA analysis was performed. Most CNAs were present in all samples sequenced from the same patient (Figure 4). Dimension reduction using t-distributed stochastic neighboring embedding (t-SNE) showed CNA profiles of samples from individual patients clustered together, indicating clonal similarity of multiple tumors from the same patient (Figure 3b).

Gene-level copy number status was determined via the GISTIC2.0 algorithm (Figure 5). Recurrent CNAs included amplifications in chromosomes 5p15.33 (patients 1, 2 and 8) and 11q13.2 (patients 2, 5 and 6). Recurrent deletions included chromosomes 9p21.3 (patients 4, 5 and 6) and 18q23 (patients 3, 4 and 7).

Discussion

In patients with multifocal IHC, a distinction is made between potentially resectable SN and unresectable IM. However, criteria that distinguish between SN and IM are lacking. In this study, molecular alterations were concordant across all intrahepatic tumors in individual patients. These data suggest that multiple tumors in IHC are derived from the same initiating cell and represent monoclonal metastatic seeding, regardless of tumor location or distribution in the liver. Our findings challenge the notion that SN and IM are different by providing preliminary evidence that all multifocal disease represents metastatic dissemination.

Thus, the decision to perform surgery in multifocal IHC should not be based upon a perceived biologic distinction between IM and SN.

Clinically actionable mutations in driver genes such as *IDH1* and *FGFR2* have been identified in up to 40% of patients with IHC (15,16). In this study, somatic point mutation analysis revealed that eight of the nine patients' tumors harbored driver mutations concordant across all samples in each patient. Shared non-synonymous mutations included *BRAF* and epigenetic regulators, *SMARCA4*, *MED12*, *IDH1* and *BAP1*. Analogous to the results for somatic point mutations, CNA profiles were concordant between tumors in each patient. Deletions in chromosome 9p21.3, a locus harboring *CDKN2A* and *CDKN2B* tumor suppressor genes, were identified in all samples from three of the nine patients. Recurrent *CDKN2A/B* deletions have been reported in 8–18% of patients with IHC and are associated with a poor prognosis (17–20). Three patients had amplifications in 11q13.2, which includes *CCND1*, *FGF* family members (*FGF3*, 4 and 19) and *ORAOV1*. Sia et al. reported amplification of 11q13.2 in IHC tumors characterized by activation of *RAS* and mitogen-activated protein kinase signaling pathways, which was associated with worse survival (18).

An unexpected finding of this study was concordant mutations in DNA repair genes in five of the nine patients. Indels present at the same genome positions in all samples from five patients included *CHEK1*, *ERCC5*, *ATR* and *MSH6*. In addition, three patients had shared indels in *POLE* and one patient had a non-synonymous point mutation in *MSH2*. Alterations in DNA repair genes have previously been identified in 12.5% of patients with IHC (17). This has important therapeutic implications, since tumors harboring mutations affecting DNA damage response and genomic instability can potentially be targeted with agents such as PARP inhibitors

(21). Furthermore, frameshift indels, shared among all samples in five of our patients, have been shown to trigger neoantigenic peptides highly distinct from self, leading to increased tumor immunogenicity and higher likelihood of response to immunotherapy (22). Clinical trials investigating immune checkpoint inhibitors in cholangiocarcinoma are ongoing (21).

Limitations of this study include small sample size and targeted sequencing of 201 genes, which underestimates the total burden of molecular alterations. However, the main finding of similar genomic profiles of samples from individual patients was consistent for all patients in the study. The sample size was insufficient to evaluate intratumoral heterogeneity of IHC. Another major limitation is the lack of analysis of FGFR2 fusions or rearrangements, which are reportedly present in 10–20% of patients with IHC and important targets for therapy (23–25). Furthermore, epigenomic and non-coding regulatory factors, which may contribute to differences between SN and IM, were not examined. This study was limited to patients who had resected, synchronous tumors and molecular alterations in patients with more advanced, unresectable or metachronous IHC were not analyzed. Future studies with more comprehensive molecular analysis and larger cohorts that include patients with unresectable and metachronous disease will be important to further elucidate the clinical impact of molecular alterations in multifocal IHC.

In conclusion, in this study of multifocal IHC, SNVs, indels and CNAs were concordant across spatially separated tumors within an individual and divergent between tumors from different individuals. These findings are clinically important because they challenge the current practice of differentiating between potentially resectable SN and unresectable IM. Future investigations are needed for adjuvant therapy strategies to eradicate intrahepatic micrometastases in multifocal IHC.

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Conflict of Interest Statement: None declared.

Data availability

The data underlying this article will be shared on reasonable request to the corresponding author.

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