#### REVIEW

# Dihydroartemisinin as a Sensitizing Agent in Cancer Therapies

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**Abstract:** Cancer is one of the major threats to human health. Although humans have struggled with cancer for decades, the efficacy of treatments for most tumors is still very limited. Dihydroartemisinin (DHA) is a derivative of artemisinin, a first-line antimalarial drug originally developed in China. Beyond the anti-malarial effect, DHA has also been reported to show anti-inflammatory, anti-parasitosis, and immune-modulating properties in vitro and in vivo. Furthermore, an increasing number of studies report that DHA possesses anticancer activities on a wide range of cancer types both in vitro and in vivo, as well as enhances the efficacy of chemotherapy, targeted therapy, and even radiotherapy. However, the mechanisms of DHA on different tumors differ in various ways. In this review, we intend to summarize how DHA sensitizes cancer cells to anti-cancer therapies, highlight its molecular mechanisms and pharmacological effects in vitro and in vivo as well as in current clinical trials, and discuss potential issues concerning DHA. Hopefully, more attention will be paid to DHA as a sensitizer for cancer therapy in the future.

**Keywords:** dihydroartemisinin, anti-tumor drugs, sensitizer, molecular mechanism

#### **Introduction**

<span id="page-0-3"></span><span id="page-0-2"></span>Cancer is one of the leading causes of death worldwide, with an estimated [1](#page-7-0)8.1 million new cases and 9.6 million deaths globally in  $2018<sup>1</sup>$ . Artemisinin is a colorless needle-like crystal extracted from the stems and leaves of the composite inflorescence plant *Artemisia annua*, which is a sesquiterpene lactone compound containing a peroxy group. The peroxy group in its molecular structure has alternating O-C-O-C segments that can be fractured for chemical synthesis of artemisinin derivatives. Dihydroartemisinin (DHA) is an artificial semi-synthetic derivative obtained after the reduction of artemisinin. DHA retains the antimalarial active group and contains a hydroxyl group, which greatly enhances its antimalarial effects.<sup>2</sup> DHA has many advantages compared to artemisinin, such as higher water solubility, higher efficacy, easier absorption, wider distribution, quicker excretion and metabolism, higher efficiency, and lower toxicity. The oral bioavailability of DHA is 10 times more than that of artemisinin, and its antimalarial effect is 4–8 times greater compared with artemisinin. $3$  In the presence of ferrous ions, the peroxide bridge in DHA breaks and results in the production of cytotoxic reactive oxygen species (ROS), which is considered to be one of the mechanisms behind the anti-tumor and antimalarial effects of DHA.<sup>4</sup> Data shows that DHA can kill tumor cells by inducing cell

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<span id="page-1-3"></span><span id="page-1-2"></span><span id="page-1-1"></span>cycle arrest or apoptosis,  $5,6$  $5,6$  preventing tumor angiogenesis, $\frac{7}{10}$  $\frac{7}{10}$  $\frac{7}{10}$  inhibiting tumor invasion and metastasis.[8](#page-7-7) Moreover, no obvious toxicity has been found in normal cells treated with DHA, which reveals that DHA is a potential ideal anti-tumor drug for cancer therapy. Recent studies showed that DHA revealed anticancer activities in various tumors and was used as a sensitizer for some cancer therapies as well.

In this review, we summarized studies on the synergistic effect of DHA in tumor therapies to explain that DHA has the potential value to improve the therapeutic effect of clinical anti-cancer drugs, including chemotherapy ([Figure 1\)](#page-1-0), targeting therapy, and radiotherapy ([Figure 2](#page-2-0)).

## **In vitro Evidence of DHA Positive Effects on Chemosensitization** DHA Enhances the Anti-Cancer Activity of Alkylating Agents

Alkylating agents, also known as biological alkylating agents, are a class of cell cycle non-specific drugs. The most used alkylating agents in cancer patients are temozolomide, nitrogen mustard, etc. The alkyl group in the molecular structure is the common feature of this kind of drug. Alkyl

<span id="page-1-0"></span>

**Figure 1** Sensitization of DHA to chemotherapy drugs. Red arrows ↑ and ↓ indicate proteins or pathways up-regulated and down-regulated by DHA to elevate the toxicity of other chemotherapy drugs to cancer cells.

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**Figure 2** Sensitization of DHA to targeted drugs and radiotherapy. Red arrows ↑ and ↓ indicate proteins or pathways up-regulated and down-regulated by DHA to elevate the toxicity of targeting drugs or radiotherapy to cancer cells.

groups can be alkylated with nucleophilic groups in DNA, RNA, or proteins in cancer cells, which can form cross-links or cause a depurination effect and finally induce DNA damage. In the next cell cycle, the miscoding of the base pair results in DNA structural and functional damage and eventually induces cell death in severe cases.

<span id="page-2-3"></span><span id="page-2-2"></span><span id="page-2-1"></span>Temozolomide (TMZ) is an imidazolotetrazine alkylating agent with antitumor activity. In the physiological condition (PH=7.4), TMZ spontaneously converts to 5- (3-Methyltriazen-1yl) imidazole-4-carboxamide  $(MTIC)^9$ which exerts cytotoxic effects mainly through alkylation/ methylation at the O6 and N2 sites of DNA guanine. Studies show that TMZ induces apoptosis of gliomas by up-regulating caspase-3 and down-regulating AKT and HSP90 on protein levels. $10,11$  $10,11$  TMZ also induces AMPK activation, which is involved in TMZ-induced glioblastoma cell apoptosis by promoting p53 activation, inhibiting mTORC1 signaling, altering the expression levels of apoptosis-related proteins such as p21, Noxa, Bax, and Bcl-2.<sup>[12](#page-7-11)</sup> However, TMZ is prone to drug resistance, and the current study demonstrated that TMZ resistance was the result of a combination of factors, such as DNA damage repair systems like O6-methylguanine DNA methyltransferase, mismatch repair, and base excision repair as well as autophagy or cancer stem cells.<sup>[13](#page-8-0)</sup> DHA can induce autophagy and apoptosis in cancer cells, and a synergistic effect was seen when DHA was combined with TMZ for cancer therapies. DHA combined with TMZ

<span id="page-2-5"></span><span id="page-2-4"></span>inhibited U87MG cell proliferation and induced apoptosis. $14$  The mechanism was mainly down-regulating the phosphorylated mitogen extracellular kinase (MEK) and extracellular signal-regulated protein kinase (ERK) proteins, and up-regulating the expression of P53 and down-regulating the expression of Bcl-2 and Mcl-1, which finally initiated mitochondrial apoptosis in cancer cells. DHA enhanced the toxic response of cells to TMZ and inhibited the proliferation of glioma cells through autophagy. [15](#page-8-2) In addition, DHA has also been found to enhance the cytotoxic effect of temozolomide (TMZ) on rat C6 glioma cells by ROS generation. When the concentration of DHA was 5 µmol/L, the cytotoxic effect of TMZ was increased by 321% and significantly enhanced TMZ-induced apoptosis and necrosis.<sup>[16](#page-8-3)</sup> Additionally, TMZ alone reduced tumor size by about 55%, while the combination of DHA with TMZ resulted in a tumor size of less than 20% compared to untreated animals.<sup>17</sup> In summary, DHA enhances the cytotoxic effect of TMZ by inducing autophagy and ROS, subsequently enhancing the antiproliferative effect of TMZ on cancer cells in vitro and in vivo. Moreover, DHA could also enhance the antitumor effect of TMZ by inducing cancer cell apoptosis.

## <span id="page-2-7"></span><span id="page-2-6"></span>DHA Improves the Activities of Antimetabolite Anti-Cancer Drugs

Currently, the widely used antimetabolite chemotherapeutic drugs include gemcitabine, 5-fluorouracil (5-FU), cytarabine, and capecitabine. These drugs work by interfering with the essential biosynthetic pathways, perturbing the synthesis of DNA/RNA, or forming deoxyribonucleic acid reductase and deoxyribonucleic acid polymerase, as well as incorporating pseudo structural analogues of pyrimidine/purine into DNA[.18](#page-8-5)

<span id="page-3-3"></span><span id="page-3-2"></span><span id="page-3-1"></span><span id="page-3-0"></span>Gemcitabine (GEM) is a novel cytosine nucleoside derivative, activated by deoxycytidine kinase after entering the body and further metabolized by cytosine nucleoside deaminase, which is finally incorporated into DNA and arrests the cell cycle at the G1/S phase. Gemcitabine resistance may be related to epithelial-mesenchymal transition (EMT), DNA damage and repair, abnormal signaling pathways, and the related coding genes, etc.<sup>19</sup> Deoxycytidine kinase (*dCK*) is a key protein that transforms GEM into an effective active form and exerts its efficacy. The mutation in *dCK* is an important mechanism for cancer cells to acquire drug resistance towards GEM. The mutation E197K in *dCK* is a key event of GEM resistance, resulting in DNA damage in cancer cells.<sup>20</sup> The combination of DHA and GEM has a synergistic antiovarian cancer effect. $^{21}$  $^{21}$  $^{21}$  ROS generated from DHA inhibits the expression of cellular cytidine deaminase (CDA), reduces GEM inactivation, and finally enhances the anticancer activity of GEM. Zhao and colleagues proved that DHA enhanced the anti-cancer activity of GEM to A549 cells by producing  $ROS<sup>22</sup> Notably, they demonstrated$ that, although both GEM and DHA increased ROS production, respectively, the combination of the two drugs did not enhance the ROS level, while the synergistic induction of apoptosis was obvious through the Bak-mediated intrinsic apoptotic pathway and the FAS-caspase-8-mediated extrinsic apoptotic pathway. In addition, they discovered that DHA had a more potent pro-apoptotic effect in gemcitabine resistant A549 cells (A549GR), in which Bax and caspases were involved. $^{23}$  $^{23}$  $^{23}$  Wang and colleagues demonstrated that DHA increased the anti-cancer activity of GEM both in vitro and in vivo. $24$  In pancreatic cancer cells, BxPC-3 and PANC-1, DHA enhanced GEMinduced growth inhibition and apoptosis by blocking GEM-induced NF-κB activation, reducing c-myc, cyclin D1, Bcl-2, and Bcl-xL expression in vitro. In in vivo pancreatic cancer xenograft experiments, the tumor volume was significantly reduced and apoptosis was sig-nificantly increased in the combination group.<sup>[25](#page-8-12)</sup> Some researchers revealed that the combination of DHA and GEM could significantly reduce the expression of VEGF and MVD and decrease the tumor microvessel density, <span id="page-3-7"></span>inhibit tumor neovascularization and proliferation.<sup>[26](#page-8-13)</sup> These studies revealed that DHA could elevate the chemotherapeutic effect of GEM mainly by increasing ROS production to inhibit chemoresistance and induce apoptosis. Moreover, DHA might also inhibit tumor neovascularization to enhance the antitumor effect of GEM.

<span id="page-3-9"></span><span id="page-3-8"></span>5-FU has no biological activities until it has been converted into 5-fluorouracil deoxynucleotide in vivo, which then combines with thymine synthase (TS) and formyltetrahydrofolate to form a stable triple complex and finally inhibits the activity of TS. So far, inhibition of TS by the active metabolites of 5-FU as the main mechanism for its anti-tumor effects has been well understood. $27$  Recent studies found that DHA could act synergistically with 5-FU in inhibiting the growth of cancer cells. DHA synergistically inhibited the growth of gastric cancer cells by enhancing the pro-apoptotic effect of 5-FU, which sensitized gastric cancer cells to the 5-FU action by inhibiting Bcl-XL expression and ATF2 phosphorylation and elevating the release of cytochrome C activation of caspase- $3.28$  $3.28$ Moreover, a combination of DHA and 5-FU significantly promoted gastric cancer cell, SGC7901, apoptosis through inhibiting the expression of SIRT1 and promoting ROS generation as well as phosphorylation of ASK1 and JNK.[29](#page-8-16) Furthermore, Yao et al showed that DHA effectively reversed the anticancer effect of 5-FU on p53 gene knockout colorectal cancer HCT116 cells (HCT116, TP53-/-) through ROS-mediated apoptosis in DHA treated  $HCT116$  cells.<sup>30</sup> The molecular mechanisms might be associated with the up-regulation of Bcl-2 and the downregulation of Bax. Overall, DHA exerts synergistic antitumor effects by inducing apoptosis to enhance the sensitivity of tumor cells to 5-FU.

## <span id="page-3-11"></span><span id="page-3-10"></span><span id="page-3-4"></span>DHA Promotes the Anti-Cancer Activities of Antibiotic Chemotherapeutic Drugs

<span id="page-3-6"></span><span id="page-3-5"></span>An antibiotic chemotherapeutic drug is a kind of antibiotic with a killing effect on tumor cells; it belongs to nonspecific cell cycle drugs. Antibiotic chemotherapeutic drugs include two categories according to their different mechanisms. One type is antibiotics, which directly crossbind to DNA, disrupt its structure and function, and finally prevent its replication, such as mitomycin and bleomycin. The other drug intercalates into the DNA double strand to interfere with its duplication and inhibit RNA synthesis, such as doxorubicin, daunorubicin, and actinomycin D.

<span id="page-4-0"></span>Doxorubicin (DOX) is an anti-tumor antibiotic that inhibits the synthesis of RNA and DNA, which has a stronger inhibitory effect on RNA. DOX has a widely anti-tumor spectrum and possesses a killing effect on tumor cells in various cell cycles; it is a cell cycle nonspecific drug. P-glycoprotein (P-gp) is a transmembrane glycoprotein with a molecular weight of 170kD. P-gp binds to both drugs and ATP, so that intracellular drugs can be pumped out of the cells, and finally reduces the intracellular drug concentration and develops drug resistance. The higher expression of P-gp on the endothelial cell surface could reduce the penetration of chemotherapy drugs to specific sites. DHA could restrain the expression of P-gp by inhibiting the p53 (R248Q)-ERK1/2-NF-κB signaling pathway, so that liver cancer cells embedding  $p53$  (R248O) are sensitized to doxorubicin, $31$  and reverse chemotherapy resistance. DHA elevated the sensitivity of human colon cancer resistant cells HCT8/ADR to DOX trough down-regulating the expression of Bcl-xl and inducing autophagy. [32](#page-8-19) Another study showed that DHA inhibited the activity of Bcl-2 by combining epirubicin, promoted the release of Beclin 1 and activated Bax to induce apoptosis, which finally led to type I programmed cell death of breast cancer cells. In addition, Beclin 1 initiated excessive autophagy and led to type II apoptosis of breast cancer cells.<sup>33</sup> Tai et al showed that the combination of DOX and DHA showed a synergistic effect on HeLa cells through the endogenous apoptosis pathway mediated by caspase-9 and caspase- $3^{34}$  $3^{34}$  $3^{34}$  Furthermore, in vivo experiments showed that the volume of the tumors significantly reduced after intratumoral injection of DHA combined with DOX, and no obvious toxicity was observed in the liver, spleen, kidney or heart of animals. Another study reported that arginine 8 (R8) modified epirubicin and DHA liposomes, significantly inhibited lung cancer migration by down-regulating the expression of VE-Cad, TGF-β1, MMP-2, and HIF-1a, elevated the concentration of chemotherapy drugs in the selective accumulation of tumor sites, and enhanced the targeted therapy effect. $35$  Taken together, DHA enhances the anti-tumor effect of DOX on tumor cells by down-regulating P-gp expression and/or promoting apoptosis, inducing autophagy, and inhibiting tumor migration.

<span id="page-4-5"></span><span id="page-4-4"></span><span id="page-4-3"></span>Rapamycin (RAPA) is a specific inhibitor of the mTORC1 complex. RAPA indirectly promotes the occurrence of autophagy by inhibiting the mTORC1 complex.<sup>36</sup> Studies showed that RAPA significantly increased the expression of caspase-3 in tumor tissues, which promoted <span id="page-4-8"></span><span id="page-4-7"></span><span id="page-4-6"></span>the apoptosis of tumor cells.<sup>[37](#page-8-24)</sup> RAPA resistance may be related to the negative feedback activation of phosphatidylinositol 3-kinase-protein kinase B (PI3K-AKT) after mTOR suppression. $38$  A previous study showed that DHA effectively inhibited the expression of PI3K and p-Akt.[39](#page-8-26) A combination of DHA with RAPA resulted in dual inhibition of the PI3K/AKT signaling pathway, which further prevented the reactivation of PI3K. Thongchot et al showed that DHA induced the expression of autophagyrelated genes ATG12, BNIP3, and ULK1 in cholangiocarcinoma and reduced the expression of mTOR that inhibits autophagy. [40](#page-8-27) Liu et al demonstrated that DHA and RAPA synergistically promote the apoptosis and death of breast cancer MDA-MB-231 cells by up-regulating DAPK and  $ATG7<sup>41</sup>$  $ATG7<sup>41</sup>$  $ATG7<sup>41</sup>$  Collectively, the synergistic anti-tumor effect of DHA combined with RAPA probably works through the PI3K/AKT/mTOR pathway inhibition and apoptosis promotion.

## <span id="page-4-10"></span><span id="page-4-9"></span><span id="page-4-1"></span>DHA Elevates the Anti-Cancer Activities of Plant-Derived Chemotherapeutic Drugs

<span id="page-4-2"></span>Plant-derived chemotherapeutic drugs are composed of alkaloids and natural products, which could inhibit mitosis or enzymes of tumor cells, preventing the synthesis of proteins that are necessary for cell regeneration. The clinical first-line used plant-derived chemotherapeutic drugs are paclitaxel, curcumin, and vincristine.

<span id="page-4-13"></span><span id="page-4-12"></span><span id="page-4-11"></span>Paclitaxel (PTX) is an anti-cancer agent derived from the North American Pacific yew tree (*Taxus brevifolia*) that specifically acts on M-phase cells and belongs to the cell cycle-specific anti-cancer drugs. PTX could promote the polymerization of tubules into stable microtubules, inhibit their depolymerization, damage the network structure of microtubules, and inhibit mitosis of cells. PTX achieved its anti-tumor effect on cervical cancer by inhibiting cancer cell mitosis and affecting the activity of mTOR signaling pathway.<sup>42</sup> Huang et al showed that the inhibition rate of breast cancer cell line MCF-7 was significantly higher in cells treated with DHA combined with PTX compared with that treated with DHA or PTX alone.[43](#page-8-30) Moreover, DHA combined with docetaxel showed a synergistic effect on prostate cancer cells, which prevented or delayed docetaxel resistance to prostate cancer cells.[44](#page-8-31) To alleviate the side effects of high-dose PTX in patients, researchers synthesized PEGylated PTX and DHA nanoparticles to evaluate its anti-cancer effect in

colorectal cancer cells in vitro and in vivo, and the results showed that DHA-PEG-PTX nanosystems (PD@PPD) showed remarkably increased apoptosis in HT-29 cells, as compared to free drug treatment. More importantly, DHA minimized the side effects of PTX and enhanced anti-tumor efficacy. [45](#page-8-32)

<span id="page-5-1"></span><span id="page-5-0"></span>Curcumin (CUR) is a yellow pigment extracted from the rhizomes of turmeric and other ginger plants; it is an acidic polyphenol with unsaturated aliphatic and aromatic groups, which shows anti-inflammatory and anti-oxidation pharmacological effects. Studies revealed that curcumin has an anticancer effect by inhibiting cell proliferation, promoting apoptosis, preventing tumor angiogenesis and metastasis, and inducing autophagy. [46–50](#page-8-33) Zhao et al demonstrated that the combination of DHA and curcumin synergistically up-regulated the expression of miR-124, subsequently reduced the expression of midkine(MK) and increased cytotoxicity, and finally reduced SKOV3 ovarian cancer cell survival by inducing cell cycle arrest and promoting apoptosis. $51$ 

## <span id="page-5-2"></span>DHA Enhances the Anti-Cancer Efficacy of Platinum-Based Chemotherapeutic Drugs

<span id="page-5-6"></span><span id="page-5-5"></span><span id="page-5-4"></span><span id="page-5-3"></span>Platinum-based regimens play a vital role in cancer therapy. Platinum inhibits the division of cancer cells by inducing DNA replication disorders. Five kinds of platinum anti-tumor drugs are widely used in cancer patients, including cisplatin (the first generation), carboplatin (the second generation), and oxaliplatin and loplatin (the third generation). One study showed that most cell lines resistant to cisplatin exhibit a phenotype of reduced platinum accumulation, most likely due to reduced cisplatin uptake rather than increased drug efflux.<sup>[52](#page-9-1)</sup> Bcl-2 might also be associated with platinum chemoresistance in lung cancer.<sup>53</sup> Therefore, elevation of platinum concentration in tumor cells and autophagy induction might enhance the chemotherapeutic effect of platinum. A previous study showed that DHA increased the sensitivity of cisplatinresistant gastric cancer cell line SGC7901/DDP cells to cisplatin by inducing apoptosis, which was accompanied by P-gp down-regulation.<sup>54</sup> Feng et al showed that DHA significantly enhanced the cytotoxic effect of cisplatinresistant ovarian cancer cells SKOV3/DDP to cisplatin by inducing apoptosis and autophagy.<sup>55</sup> Moreover, another study in esophageal squamous cell carcinoma (ESCC) showed that continuous cisplatin treatment activated Shh signaling and induced cancer stem-like properties in ESCC

<span id="page-5-8"></span><span id="page-5-7"></span>patients, which subsequently resulted in reduction of cisplatin-induced cytotoxicity in KYSE510 cells. DHA could suppress activation of the Shh pathway and attenuate the cancer stem-like traits in ESCC cells and, finally, enhance the sensitivity of cisplatin on  $\text{ESCC.}^{56}$  $\text{ESCC.}^{56}$  $\text{ESCC.}^{56}$  Qin et al also revealed that DHA could enhance the chemo-sensitivity of HCC cells to cisplatin (DDP) and oxaliplatin  $(OXA)$ .<sup>[57](#page-9-6)</sup> The mechanism behind this enhancement was that DHA altered the morphology of HCC cells induced by DDP and OXA and the expression of EMT biomarkers in HCC cells through AKT-Snail signaling pathway, which improved the efficacy of chemotherapy in HCC patients. In addition, Zhang et al reported that DHA combined with carboplatin (CBP) had a stronger inhibitory effect on Lewis lung cancer (LLC) cells.<sup>58</sup> DHA sensitized LLC cells to CBP treatment and induced cell cycle arrest by activating the p38-MAPK signal pathway. In short, DHA could reduce platinum resistance in tumor cells by elevating the concentration of platinum in tumor cells and inhibiting the Shh signaling pathway, while enhancing platinum chemosensitivity by cell cycle arrest and apoptosis induction.

## <span id="page-5-9"></span>DHA Enhances the Anti-Cancer Activities of Targeted Therapies

Targeted therapy is a treatment aimed at identified carcinogenic sites at the cellular and molecular level. The corresponding therapeutic drugs enter the patient, specifically gather at the carcinogenic sites and specifically kill the tumor cells without damaging normal cells. As wellknown "biological missiles", the targeted chemotherapeutic drugs include sorafenib, gemfibrozil, and ABT-263, etc.

<span id="page-5-13"></span><span id="page-5-12"></span><span id="page-5-11"></span><span id="page-5-10"></span>Sorafenib (SRF) is an oral multi-kinase inhibitor, which not only inhibits the proliferation of tumor cells by targeting Raf kinase in the Raf/MEK/ERK signaling pathway, but also targets VEGFR2/3 and platelet derivation growth factor receptor β tyrosine kinase to both exert an anti-angiogenic effect and induce apoptosis in liver cancer cells.[59](#page-9-8) Overexpression of EGFR or ligands in HCC cells leads to sustained activation of EGFR downstream signaling and is related to SRF resistance.<sup>60</sup> Zhang et al elucidated that down-regulation of p-ERK may be associated with SRF resistance in liver cancer. [61](#page-9-10) Shimizu and colleagues found that SRF treatment resulted in autophagosomes accumulation and the activation of autophagic flux in Huh7, HLF, and PLC/PRF/5 cells, which promoted the survival of hepatoma cells by decreasing the efficiency of SRF. [62](#page-9-11) Therefore, the combination of drugs might reduce the resistance of tumor cells to sofiranib. Hou and colleagues demonstrated that the combination of DHA and SRF could significantly up-regulate APOA1 and MYH11 and down-regulate the expression of GALNT10 to inhibit tumor angiogenesis, migration, and invasion in liver cancer. [63](#page-9-12) Wang et al also proved that the combination of SRF and DHA had a synergistic anti-cancer effect on HepG2 cells, and low-density lipoprotein-based SRF/ DHA lipid nanoparticles (LD-SDN) were more effective in reducing cell survival and improving tumor-targeting effects compared with single administration. The antitumor response and delayed tumor growth phenomenon were more significant in LD-SDN-treated xenograft models as well.<sup>64</sup> Together, DHA could promote the anti-tumor effect of sorafenib mainly through inhibiting the invasion and migration, promoting the apoptosis, and reducing the resistance of tumor cells to sorafenib.

<span id="page-6-1"></span><span id="page-6-0"></span>Gefitinib is a selective epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor that blocks downstream signaling pathways by competing for the ATP binding site of the EGFR tyrosine kinase catalytic region, which inhibits the growth, metastasis and angiogenesis of tumor cells. Data showed that resistance to EGFR-TKIs was associated with impaired EGFR-TKI-mediated apoptosis.<sup>[65,](#page-9-14)[66](#page-9-15)</sup> Deng et al found that COX-2 mediated gefitinib resistance by interacting with EGFR and subsequently activating PI3K-AKT signaling in non-small cell lung cancer (NSCLC). $67$  Jin et al demonstrated that DHA combined with gefitinib induced NCI-H1975 cell cycle arrest at the G2/M phase and apoptosis, and inhibited cell migration and invasion by down-regulation of p-Akt, p-mTOR, p-STAT3, and Bcl-2 and up-regulation of Bax.<sup>68</sup> Song et al found that the combination of gemfibrozil and DHA significantly inhibited the proliferation and migration of lung adenocarcinoma cell A549, and arrested the cell cycle at the  $G0/G1$  phase.<sup>69</sup> The molecular mechanisms might be associated with the down-regulation of CDK4, cyclin D1, MMP2, and MMP9. Taken together, DHA might increase the anti-tumor effect of gefitinib by suppressing the PI3K/AKT/mTOR pathway and abolishing gemfibrozil resistance by inducing cell cycle arrest, apoptosis, migration, and invasion inhibition.

<span id="page-6-3"></span><span id="page-6-2"></span>ABT-263 (Navitoclax) is a targeted inhibitor against the Bcl-2 family of anti-apoptotic protein members such as Bcl-2, Bcl-xL, and Bcl-w. Many studies have shown that a high concentration of ABT-263 alone can induce tumor cell apoptosis in vivo and in vitro. Researchers found that activation of the endoplasmic reticulum stress response <span id="page-6-5"></span><span id="page-6-4"></span>pathway can mediate resistance of uveal melanoma cells to ABT-263.<sup>[70](#page-9-19)</sup> Another study showed that the interstitial phenotype of clear cell carcinoma of the ovary (OCCC) cells were resistant to ABT-263, and that pro-apoptotic protein BIM expression was insufficient.<sup>[71](#page-9-20)</sup> Mechanistically, EMT-induced transcription factor ZEB1 down-regulated the transcription of BIM by binding to the BIM promoter, and further developed the resistance of OCCC cells to ABT-263. Recently, some studies reported that DHA had a synergistic effect with ABT-263 and induced Bax-dependent apoptosis in NSCLC cells. DHA inhibited the activity of STAT3, down-regulated the expression of Mcl-1and survivin, up-regulated the expression of Bim, and finally worked as a sensitizer of ABT-263 to induce apoptosis of NSCLC cells with EGFR or RAS mutations.[72](#page-9-21) In leukemia, researchers demonstrated that ABT-263 combined with DHA efficiently improved the treatment of BCR-ABL<sup>+</sup>B-ALL. DHA inhibited cellular stress response activation, resulting in the down-regulation of Mcl-1, and increased the sensitivity of leukemia cells to ABT-263.[73](#page-9-22) Hence, DHA might promote ABT-263 induced apoptosis by reversing the resistance of tumor cells to ABT-263 through inhibition of STAT3 and upregulation of Bim.

### <span id="page-6-7"></span><span id="page-6-6"></span>DHA Enhances the Efficacy of Radiotherapy

<span id="page-6-9"></span><span id="page-6-8"></span>Radiotherapy is a commonly used local and regional treatment for cancer. Studies have shown that the sensitivity of tumor cells to radiotherapy is tightly related to cell cycle status, whereby cells at the G2/M phase are the most sensitive to radiotherapy, followed by cells at the G0/G1 phase; cells at the S phase are insensitive. Moreover, the time and degree of arrest may also alter radiosensitivity.<sup>[74](#page-9-23)</sup> Zuo et al showed that DHA has a higher tumor suppressive effect and radiosensitizing effect on lung cancer GLC-82 cells transplanted in nude mice[.75](#page-9-24) The sensitization effect might be through arresting cell cycle at the G2/M phase and inducing apoptosis. In addition, they also proved that radiotherapy combined with DHA in human lung cancer cells could reduce the proportion of cells at the S phase, increase G0/G1 phase cells, and induce apoptosis. The mechanisms might be through functional retore of p53 and p21 to inhibit Bcl-2 expression to promote GLC-82 cell apoptosis[.76](#page-9-25) Moreover, PD-L1 expression of NSCLC was positively related with radiation resistance, while DHA could eliminate radiation resistance and synergistically enhance the anti-tumor effect of radiotherapy in NSCLC cells by inhibiting the PD-L1 expression trough

<span id="page-7-12"></span>inhibiting TGF-β, PI3K/Akt, and STAT3 signaling pathways[.77](#page-9-26)

# **Clinical Trials of DHA in Cancer Treatment**

In addition to in vivo and in vitro studies, two clinical trials of DHA for cancer treatment were identified from Chinadrugtrials.org.cn and Clinicaltrials.gov. The clinical trial in China (registration number: ChiCTR2100041652) investigated dihydroartemisinin combined with chemoradiotherapy in the treatment of locally advanced head and neck squamous cell carcinoma: a multicenter, open-label, randomized control study is at the prospective registration phase. The second trial was also from China (registration number: NCT03402464) and is titled Icotinib Combined with Dihydroartemisinin (DHA) Therapy in Patients with Advanced NSCLC; it is a Phase II study to explore the anti-tumor effect of icotinib combined with DHA on EGFR-mutant lung adenocarcinoma patients. Herein, more clinical trials are needed to further evaluate the clinical application of DHA in cancer treatment.

## **Conclusions**

Although varying amounts of anti-cancer agents are used as therapeutic drugs, drug-resistance remains intractable, which has been perplexing the cancer therapy research community for decades. Recently, researchers found that DHA could reverse drug resistance to chemotherapeutics or enhance anti-cancer activities when combined with some anti-cancer drugs. In this review, we summarized the potential molecular mechanisms or targets of DHA working as a sensitizer in tumor therapy to try to provide useful information for clinicians and patients, although how DHA exactly enhances the anti-cancer activity of tumor cells against chemotherapy, targeted therapy, and radiotherapy remains to be further explored. In addition, in vivo experiments should be applied as models for further target identification in DHA treatment. As a classic first-line anti-malarial drug, the efficacy and safety of DHA has been proven for years, while its role in cancer therapy needs further elucidation. Therefore, more clinical trials are needed to test the efficacy and safety of DHA in cancer patients. In addition, whether DHA could reduce the toxic side effects of anti-tumor drugs or radioactive rays on patients remains unclear. The elucidation of these key issues might be helpful to accelerate the clinical application of DHA as a sensitizer for cancer therapy.

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## **Disclosure**

The authors declare no conflicts of interest in this work.

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