ROYAL SOCIETY OPEN SCIENCE

royalsocietypublishing.org/journal/rsos

Research



Cite this article: Louis M, Skovrind M, Garde E, Heide-Jørgensen MP, Szpak P, Lorenzen ED. 2021 Population-specific sex and size variation in longterm foraging ecology of belugas and narwhals. *R. Soc. Open Sci.* **8**: 202226. https://doi.org/10.1098/rsos.202226

Received: 7 December 2020 Accepted: 11 January 2021

Subject Category:

Ecology, conservation, and global change biology

Subject Areas:

ecology

Keywords: stable isotopes, Arctic cetaceans, carbon, nitrogen

Authors for correspondence:

Marie Louis e-mail: marie.louis@sund.ku.dk Eline D. Lorenzen e-mail: elinelorenzen@sund.ku.dk

Electronic supplementary material is available online at https://doi.org/10.6084/m9.figshare.c. 5280416.

THE ROYAL SOCIETY PUBLISHING

Population-specific sex and size variation in long-term foraging ecology of belugas and narwhals

Marie Louis¹, Mikkel Skovrind¹, Eva Garde², Mads Peter Heide-Jørgensen², Paul Szpak³ and Eline D. Lorenzen¹

¹Globe Institute, University of Copenhagen, Denmark ²Greenland Institute of Natural Resources, Copenhagen, Denmark ³Trent University, Peterborough, Ontario, Canada

ML, 0000-0002-4611-5503; MS, 0000-0002-5430-5884; EG, 0000-0003-3016-5722; MPH-J, 0000-0003-4846-7622; PS, 0000-0002-1364-6834; EDL, 0000-0002-6353-2819

Intraspecific variation in resource use by individuals of different age, sex or size may reflect differing energetic requirements and physiological constraints. Males and females often show differences in diet owing to sexual size dimorphism, different life histories and/or habitat use. Here, we investigate how sex and size influence the long-term foraging ecology of belugas and narwhals in Greenland, using stable isotopes of carbon and nitrogen from bone collagen. We show that males have a higher trophic level and a larger ecological niche than females in West Greenland belugas and in East Greenland narwhals. In addition, for these two populations, we find that δ^{15} N increases with size, particularly in males. We hypothesize that sexual size dimorphism together with strong maternal investment drive these differences. By contrast, we find no differences in foraging ecology between sexes in West Greenland narwhals and observe no influence of size on trophic level. This may reflect the influence of interspecific competition in West Greenland, where the distributions of belugas and narwhals overlap, and/or geographical resource partitioning among different summer aggregations of narwhals. Our results suggest that sex and size variations in diet are population dependent, and probably the result of varying ecological interactions.

1. Introduction

The foraging ecology of a population may be driven by the abundance and distribution of resources, interspecific competition,

@ 2021 The Authors. Published by the Royal Society under the terms of the Creative Commons Attribution License http://creativecommons.org/licenses/by/4.0/, which permits unrestricted use, provided the original author and source are credited.



West Greenland (WG) belugas

Figure 1. (*a*) Sample localities (a: Qaanaaq, b: Melville Bay, c: Kullorsuaq, d: Nuussuaq, e: Uummannaq, f: Qeqertarsuaq, *g*: Scoresbysund) with sample size and distribution of belugas and narwhals [29]. The WG narwhal population comprises samples from four localities. Coastline data are from [30]. (*b*) Bone δ^{13} C and δ^{15} N for male and female WG belugas ($n_M = 14$, $n_F = 9$), WG narwhals ($n_M = 19$, $n_F = 20$) and EG narwhals ($n_M = 22$, $n_F = 16$). Solid circles indicate Bayesian standard ellipse areas (SEA_B). Mean (square) and s.d. (error bars) are indicated.

and age and sex structure within the population [1–4]. Animals of different age, sex or size may show variations in resource use such as preferred prey or foraging habitat, owing to differing energetic requirements and physiological capabilities [5–7]. This may act to reduce intraspecific competition [2]. In mammals, strong sexual size dimorphism and/or different life histories, including maternal care, between males and females, can often lead to dietary differences [7]. Females raise their young for several weeks to years and adjust their foraging behaviour to ensure offspring survival [8].

In the marine environment, differences in foraging ecology between sexes are particularly strong in pinnipeds [8–11], where males are often heavier and larger than females [12]. It is thought that larger individuals can target larger prey at higher trophic levels, and that their greater diving capabilities [13] also enable them to target more benthic species. These differences may be accentuated by spatial segregation linked to maternal investment, such as females regularly returning to land to feed their pups [8,10,14]. For many seal species, changes in size-related diving capability [15] and experience may explain ontogenetic shifts in foraging ecology [3,9,11,16,17]. The same factors could also explain age-related changes in cetacean diets [18–20]. However, in cetaceans, size differences between males and females are usually smaller, and despite maternal care over several years, many cetacean species show similar foraging ecology between sexes [20–25] (although differences have been observed in sperm whales and killer whales [26–28]).

Here, we investigate sex and size variation in the long-term foraging ecology of the two Arctic toothed cetaceans, belugas (*Delphinapterus leucas*) and narwhals (*Monoceros monoceros*), in Greenland. The two Monodontidae species coexist in West Greenland, while only narwhals are distributed in East Greenland (figure 1*a*). Both species can occur in sex-specific and mixed-sex groups [31–34], and show important maternal investment [35] and sexual size dimorphism. Male belugas and narwhals are 40–100 cm and approximately 60 cm longer than females, respectively [31,36,37], and 40–45% heavier [36,37]. Male narwhals furthermore have an erupted left tooth, the tusk, which is believed to be a secondary sexual trait [38].

The foraging ecology of belugas and narwhals has been studied using stomach contents and stable isotopes of carbon (δ^{13} C) and nitrogen (δ^{15} N) in soft tissues, which have turnover rates of several months [39–41], and are limited to reflect seasonal (often spring/summer) diet [42–44]. δ^{13} C reflects the feeding habitat (pelagic versus benthic, offshore versus coastal) and δ^{15} N the trophic level [45–48]. In Baffin Bay (West Greenland/eastern Canada), Arctic cod (*Boreogadus saida*), polar cod (*Arctogadus glacialis*) and capelin (*Mallotus villosus*) appear to be the main summer prey of both whale species [31,42,43,49]. Interspecific competition between belugas and narwhals is therefore likely in West Greenland, where their distributions overlap [29]. Capelin represent a large proportion of the summer diet of narwhals in East Greenland [43]. Both whale species undertake seasonal migrations, following the distribution of sea ice [29,50]. During winter, they stay in ice-covered areas inaccessible to humans,

limiting dietary information, although Greenland halibut (*Reinhardtius hippoglossoides*), capelin and squid (*Gonatus fabricii*) are believed to be important prey [42,44,49,51]. For both species, sexes show no or little dietary differences (0.2‰) in stable isotope compositions of soft tissues [42,43].

To gain insights into the longer term foraging ecology of beluga and narwhal populations in Greenland, we analysed bone collagen δ^{13} C and δ^{15} N, which reflect an individual's diet over multiple years [52,53]. Studies examining variation in foraging ecology on the basis of isotopic measurements of tissues with rapid turnover rates reflect only a limited portion of the year and, for Arctic species, are consistently biased towards the spring and summer months. By contrast, isotopic analyses of bone collagen offer an opportunity to examine the average diet across seasons. Thus, this approach provides insight into sustained foraging patterns that persist over years, circumventing variation caused by short-term dietary variation. To elucidate differences related to sex and size, we investigated whether beluga and narwhal foraging ecology differs across space.

2. Material and methods

2.1. Laboratory analyses

We obtained 300 mg of bone powder from the skulls of 27 belugas and 40 narwhals from West Greenland (WG), and 39 narwhals from East Greenland (EG, figure 1*a*, electronic supplementary material, table S1). All samples represented sub-adults or adults. δ^{13} C and δ^{15} N values for 14 WG belugas and 11 WG narwhals were from [54]. The skulls were collected between 1990 and 2007 during subsistence hunting, and are housed at the Natural History Museum of Denmark, University of Copenhagen. Sex information for 100 out of the 106 samples was determined previously by molecular methods [55], an inspection of sexual organs, or the presence/absence of a tusk in narwhals [56]. We followed the laboratory procedures described in [54] and in the electronic supplementary material, text.

2.2. Statistical analyses

We ran all statistical analyses in R v. 3.6.1 [57]. Our data satisfied normality and homogeneity of variance for all subdivisions. To test for dietary differences among the three populations (WG belugas, WG narwhals, EG narwhals), we compared their δ^{13} C and δ^{15} N using ANOVA and Tukey's post hoc tests using the package multcomp [58].

To test for dietary differences between males and females, we compared δ^{13} C and δ^{15} N within populations using Student's *t*-tests. The effects of location and sampling date on our results are presented in the electronic supplementary material, text; they had limited impact on our conclusions.

To investigate the association between the size of individuals and their isotopic compositions, we ran a linear regression between skull length (used as a proxy for body length [59]) and δ^{13} C or δ^{15} N within each population. A linear regression was also run separately for males and females (termed groups) within populations. Owing to some of the skulls being fragmented or otherwise broken, we were able to obtain skull measurements for 81 of the 106 belugas and narwhals investigated (see the electronic supplementary material, text for details).

We compared isotopic niche (a proxy for an ecological niche) among populations, and between groups (males/females) within each population, using Bayesian multivariate ellipse-based metrics implemented in the packages SIBER and rjags [60–63]. We calculated standard ellipse areas corrected for sample size (SEA_C), and Bayesian standard ellipses (SEA_B) for each population/group. We estimated SEA_B using 10^5 posterior draws, a burning of 10^3 and a thinning of 10, and used SEA_B to test for differences in niche width among populations/groups (i.e. the proportion (p) of draws of the posterior distribution of the SEA_B in which the area of one population/group was smaller than the other). We evaluated isotopic niche similarity between two populations/groups as the proportion (%) of the non-overlapping area of the maximum-likelihood fitted ellipses of the two.

3. Results and discussion

3.1. Niche differentiation among populations

Our results indicate resource partitioning of belugas and narwhals in West Greenland. The species occupy distinct, yet slightly overlapping (9%), ecological niches, with belugas showing higher bone



Figure 2. Size of the standard ellipse areas (SEA, $\%^2$) for male and female West Greenland (WG) belugas ($n_M = 14$, $n_F = 9$), WG narwhals ($n_M = 22$, $n_F = 16$) and East Greenland (EG) narwhals ($n_M = 19$, $n_F = 20$), with the black dot indicating the mode of the size of the SEA_R, the red cross the mean for the SEA_C and the box edges the 50, 75 and 95% credible intervals.

collagen δ^{13} C and δ^{15} N (p < 0.01, electronic supplementary material, figure S3 and table S3). Detailed results are provided in the electronic supplementary material, text. This observed niche partitioning may be the result of interspecific competition, or of differences in dietary preference. Our findings are consistent with studies indicating that—despite prey overlap between species [28,41–43,46]—belugas eat less squid, which are at lower trophic levels [43], than narwhals, and also forage on Atlantic cod (*Gadus morhua*) and other fishes such as redfish (*Sebastes mentella*) [64], which feed at higher trophic levels [65]. Although it is not possible to test for this using our stable isotope data, our findings may reflect that belugas feed on larger individuals, representing higher trophic levels, than narwhals (if both species feed on Greenland halibut or Arctic/polar cod [31,42,43,49,51]).

We did not include prey isotopic data, as these are available from soft tissue samples only [43,44,65] and reflect the short-term seasonal diet of the prey species, while our cetacean stable isotope data from bone collagen reflect the consumer diet over several years. We attempted to incorporate prey data [43,44,65] but the resulting mixing polygon was inconsistent with the isotopic compositions observed in belugas and narwhals after correcting for the Suess effect, which was done for the consumers following Szpak *et al.* 2019 [66] and trophic enrichment [67]. This suggests that the prey isotopic data are not representative of what would have been consumed by these cetaceans either because isotopic compositions are inaccurate, certain prey sources are missing, or a combination of the two. Despite belugas being considered a more generalist feeder [68], we find no significant differences in isotopic niche size between species (electronic supplementary material, figures S3, S4 and table S3).

For narwhals, we find regional differences, with no niche overlap and higher δ^{13} C and δ^{15} N in West Greenland than in East Greenland (p < 0.01; electronic supplementary material, table S3 and figure S3; figure 1*b*). This finding supports the long-term habitat segregation of narwhals west and east of Greenland, as has been shown by telemetry studies [50], and is also in agreement with stable isotope analysis of skin tissue; Watt *et al.* 2013 showed that Baffin Bay narwhals, whose main prey are benthic fish species including halibut, Arctic cod and polar cod, have a less pelagic diet than East Greenland narwhals, which consume a larger proportion of capelin [43]. Higher δ^{13} C and δ^{15} N in West relative to East Greenland is also reported in fishes [43] and other marine mammal species [69], suggesting regional differences at the base of the food web.

3.2. Sex and size differences in foraging ecology

While δ^{13} C does not differ significantly between sexes in any population (p > 0.05), our results indicate sex and size differences in the foraging ecology of West Greenland belugas and East Greenland narwhals based on δ^{15} N (figure 1*b*; electronic supplementary material, figure S5). Males have significantly higher δ^{15} N (WG belugas p = 0.03, EG narwhals p = 0.01) and larger ecological niches than females ($p \ge 0.99$; electronic supplementary material, table S3; figures 1*b* and 2). This contrasts with the analysis of δ^{13} C and δ^{15} N obtained from skin samples, which did not show any difference in niche size between sexes in narwhals in both Baffin Bay and East Greenland [43]. These discrepancies between stable isotope values estimated from bone and skin probably reflect the more rapid turnover rate of skin (months versus years), and therefore short-term versus long-term ecological differences between sexes owing to their differing energetic and physiological needs (see below). Moreover, bone collagen stable isotope data reflect year-round foraging, averaged over several years, including the winter, where most of the



royalsocietypublishing.org/journal/rsos

R.

Soc. Open Sci. 8: 202226

Figure 3. Variation in δ^{13} C and δ^{15} N from bone samples according to skull length (cm) for male and female (*a*) WG belugas ($n_M = 12$, $n_F = 9$), (*b*) WG narwhals ($n_M = 11$, $n_F = 14$) and (*c*) EG narwhal ($n_M = 15$, $n_F = 16$). The regression lines for each sex and their confidence intervals (shading) are plotted, with females in a lighter colour shade than males. Beluga and narwhal illustrations are from Uko Gorter.

foraging occurs for narwhals [49], and which is not reflected in stable isotope data from skin tissue collected in the summer.

Larger belugas and East Greenland narwhals tend to have higher δ^{13} C (p = 0.05 and p = 0.01, respectively) and higher δ^{15} N (p < 0.01; electronic supplementary material, figure S5). The pattern is stronger than that reported using skin tissue from belugas sampled in southeast Baffin Island, where older individuals show marginally higher (<0.5%) δ^{13} C and δ^{15} N [42], again possibly owing to differences in integration times between skin and bone collagen.

We hypothesize the sex-specific differences in foraging ecology observed in West Greenland belugas and East Greenland narwhals are driven by a combination of sexual size dimorphism and maternal investment, as has been observed in pinnipeds [3,8,9,16]. Larger individuals may be able to dive deeper, as they have a greater capacity to store oxygen in their tissues [15,70], allowing them to target higher trophic-level benthic prey. Larger belugas spend significantly less time at the surface than smaller individuals [71]. Their larger size may also enable them to catch and handle larger individuals of the same prey species that are at higher trophic levels (e.g. halibut or cod). Alternatively, fine-scale habitat segregation between sexes, as observed in Beaufort Sea belugas, could explain sex differences in diet [72].

The significantly positive correlation between trophic level and size is observed in male belugas and male East Greenland narwhals (p < 0.01; figure 3). The absence of a similar pattern in females may reflect their maternal investment. Despite female odontocetes raising their calves for several years and investing significant energy resources in their growth and survival, many species show no apparent sex-based dietary differences [20–25]. The offshore and deep-diving lifestyle of both belugas and narwhals may exacerbate sex-specific differences in feeding ecology, as females, whatever their size, may (at least for periods of time) adjust their diving and foraging behaviour to that of their calves, which have lower aerobic capabilities [73]. Both belugas and narwhals are predated by killer whales [74], and females probably invest in protecting their calves from this predator. Our hypothesis correlates well with studies of diving behaviour. In eastern Canada, female belugas with calves spend a higher proportion of time at the surface than females without calves and have lower diving rates [71].

Maternal investment may explain the more pronounced sex differences in foraging ecology obtained from stable isotope analysis of bone collagen than of skin tissue [42,43]. All females may not be accompanied by a calf during the particular season (often spring/summer) represented by a soft

tissue sample, whereas most adult females probably have had a calf (or multiple calves) over the integration of time represented by bone collagen.

3.3. The lack of sex and size variation in diet in West Greenland narwhals

We find no evidence of differentiated foraging ecology between males and females in narwhals in West Greenland (p > 0.05), and the size of the ecological niche is similar between sexes (p = 0.11; figures 1*b* and 2). Furthermore, we find a correlation between length and δ^{13} C (p = 0.01, electronic supplementary material, figure S5) only, in particular in females (figure 3), with no size differences in δ^{15} N (p = 0.79; figure 3; electronic supplementary material, figure S5), suggesting sex and size variation in diet may be population specific.

The pattern may reflect interspecific competition with belugas in areas where the species distributions overlap. The niche of a species and within-population variation in ecology may be constrained by interspecific competition [2]. Belugas may potentially target larger prey at higher trophic levels, and hence being a larger male narwhal would present no advantage for accessing higher trophic-level resources. Alternatively, a lack of differentiation in stable isotope values does not necessarily imply there are no dietary differences, as different prey can have similar isotopic values. Therefore, there could be sex and size differences in diet, linked to the same factors as for the other populations, including maternal investment, but which we do not detect with our data.

We investigated whether combining sampling across four locations (Qaanaq, Melville Bay, Uummannaq and Qeqertarsuaq, figure 1*a*) for West Greenland narwhals influenced our results (electronic supplementary material, text). We do not observe any differences between sexes within the one locality (Qaanaaq) from which we had a sufficient sample size for sex comparison (see the electronic supplementary material, text). Thus, including several localities probably does not influence our findings of a lack of sex differences in the foraging ecology of West Greenland narwhals. Among certain localities, we recover variation in δ^{15} N, although we observe no variation in δ^{13} C (electronic supplementary material, figure S6). This may reflect geographical partitioning within the large West Greenland (i.e. Baffin Bay) population, which is composed of several stocks [29]; every year, different stocks of narwhals in West Greenland migrate between various coastal summer localities, and offshore in Baffin Bay during winter [50]. We suggest such geographical resource partitioning may be another mechanism to decrease intraspecific competition.

4. Conclusion

We report long-term differences in foraging ecology between sexes in West Greenland belugas and East Greenland narwhals. These findings probably reflect a combination of sexual size, dimorphism, maternal investment and a deep-diving lifestyle. However, we find no differences in diet between sexes in West Greenland narwhals, indicating that sex differences in foraging ecology are population specific, and may be driven by intra- and interspecific competition. Our study highlights the applicability of stable isotope analysis of bone collagen for revealing long-term dietary differences among individuals with different life histories or physiological needs, which may remain undetected in the analysis of soft tissues with shorter turnover.

Data accessibility. Electronic supplementary material, table S1.

Authors' contributions. E.D.L. conceived the study. M.L., P.S. and E.D.L. conceived the analyses. E.G. and M.P.H.-J. collected samples. M.L. and M.S. sampled the skulls. P.S. ran the isotopic laboratory analyses. M.L. performed the statistics. M.L. and E.D.L. wrote the manuscript with input from all co-authors.

Competing interests. We have no competing interests.

Funding. This research was supported by the Carlsberg Foundation Distinguished Associate Professor Fellowship, grant CF16-0202 to E.D.L.

Acknowledgements. We thank fieldworkers and hunters involved in specimen collection. We are grateful to Uko Gorter for the illustrations, Julie Lorenzen for sampling advice and Rene Swift for figure 1*a*. We also would like to thank three anonymous reviewers for their input.

References

1. Schoener TW. 1974 Resource partitioning in ecological communities. *Science* **185**, 27–39. (doi:10.1126/science. 185.4145.27)

 Bolnick DI, Svanbäck R, Fordyce JA, Yang LH, Davis JM, Hulsey CD, Forister ML. 2003 The

ecology of individuals: incidence and implications of individual specialization. *Am. Nat.* **161**, 1–28. (doi:10.1086/343878)

- Bjorkland RH, Pearson SF, Jeffries SJ, Lance MM, Acevedo-Gutiérrez A, Ward EJ. 2015 Stable isotope mixing models elucidate sex and size effects on the diet of a generalist marine predator. *Mar. Ecol. Prog. Ser.* 526, 213–225. (doi:10.3354/meps11230)
- Hutchinson GE. 1957 Concluding remarks. Cold Spring Harb. Symp. Quant. Biol. 22, 415–427. (doi:10.1101/SQB.1957.022.01.039).
- Clutton-Brock TH, Iason GR, Albon SD, Guinness FE. 1982 Effects of lactation on feeding behaviour and habitat use in wild red deer hinds. *J. Zool.* 198, 227–236. (doi:10. 1111/j.1469-7998.1982.tb02072.x)
- Breed GA, Bowen WD, McMillan JI, Leonard ML. 2006 Sexual segregation of seasonal foraging habitats in a non-migratory marine mammal. *Proc. R. Soc. B* 273, 2319–2326. (doi:10.1098/ rspb.2006.3581)
- Belovsky GE. 1978 Diet optimization in a generalist herbivore: the moose. *Theor. Popul. Biol.* 14, 105–134. (doi:10.1016/0040-5809(78)90007-2)
- Bailleul F, Authier M, Ducatez S, Roquet F, Charrassin J-B, Cherel Y, Guinet C. 2010 Looking at the unseen: combining animal bio-logging and stable isotopes to reveal a shift in the ecological niche of a deep diving predator. *Ecography* 33, 709–719. (doi:10.1111/j.1600-0587.2009.06034.x)
- Jones KA, Ratcliffe N, Votier SC, Newton J, Forcada J, Dickens J, Stowasser G, Staniland IJ. 2020 Intraspecific niche partitioning in Antarctic fur seals, *Arctocephalus gazella. Sci. Rep.* **10**, 3238. (doi:10. 1038/s41598-020-59992-3)
- Kernaléguen L, Cazelles B, Arnould JPY, Richard P, Guinet C, Cherel Y. 2012 Long-term species, sexual and individual variations in foraging strategies of fur seals revealed by stable isotopes in whiskers. *PLoS ONE* 7, e32916. (doi:10.1371/journal.pone.0032916)
- Tucker S, Bowen WD, Iverson SJ. 2007 Dimensions of diet segregation in grey seals *Halichoerus grypus* revealed through stable isotopes of carbon (δ¹³C) and nitrogen (δ¹⁵N). *Mar. Ecol. Prog. Ser.* **339**, 271–282. (doi:10. 3354/meps339271)
- Le Boeuf BJ, Laws RM. 1994 Elephant seals: population ecology, behavior, and physiology. Berkeley, CA: University of California Press.
- Costa DP, Kuhn CE, Weise MJ, Shaffer SA, Arnould JPY. 2004 When does physiology limit the foraging behaviour of freely diving mammals? *Int. Congress Ser.* **1275**, 359–366. (doi:10.1016/j.ics.2004.08.058)
- de Lima RC, de Lima RC, Franco-Trecu V, Vales DG, Inchausti P, Secchi ER, Botta S. 2019 Individual foraging specialization and sexual niche segregation in South American fur seals. *Mar. Biol.* 166, 1–2. (doi:10.1007/s00227-019-3480-x)
- Schreer JF, Kovacs KM. 1997 Allometry of diving capacity in air-breathing vertebrates. *Can. J. Zool.* 75, 339–358. (doi:10.1139/z97-044)
- 16. Hanson NN, Wurster CM, Bird MI, Reid K, Boyd IL. 2009 Intrinsic and extrinsic forcing in life

histories: patterns of growth and stable isotopes in male Antarctic fur seal teeth. *Mar. Ecol. Prog. Ser.* **388**, 263–272. (doi:10.3354/meps08158)

- Chaigne A, Authier M, Richard P, Cherel Y, Guinet C. 2013 Shift in foraging grounds and diet broadening during ontogeny in southern elephant seals from Kerguelen Islands. *Mar. Biol.* 160, 977–986. (doi:10.1007/s00227-012-2149-5)
- Giménez J, Cañadas A, Ramírez F, Afán I, García-Tiscar S, Fernández-Maldonado C, Castillo JJ, de Stephanis R. 2017 Intra- and interspecific niche partitioning in striped and common dolphins inhabiting the southwestern Mediterranean Sea. *Mar. Ecol. Prog. Ser.* 567, 199–210. (doi:10. 3354/meps12046)
- Newsome SD, Etnier MA, Monson DH, Fogel ML. 2009 Retrospective characterization of ontogenetic shifts in killer whale diets via δ¹³C and δ¹⁵N analysis of teeth. *Mar. Ecol. Prog. Ser.* 374, 229–242. (doi:10.3354/meps07747)
- Riccialdelli L, Goodall N. 2015 Intra-specific trophic variation in false killer whales (*Pseudorca crassidens*) from the southwestern South Atlantic Ocean through stable isotopes analysis. *Mamm. Biol.* **80**, 298–302. (doi:10. 1016/j.mambio.2015.01.003)
- de Stephanis R, de Stephanis R, García-Tíscar S, Verborgh P, Esteban-Pavo R, Pérez S, Minvielle-Sebastia L, Guinet C. 2008 Diet of the social groups of long-finned pilot whales (*Globicephala melas*) in the Strait of Gibraltar. *Mar. Biol.* **154**, 603–612. (doi:10.1007/s00227-008-0953-8)
- Riccialdelli L, Newsome SD, Dellabianca NA, Bastida R, Fogel ML, Goodall RNP. 2013 Ontogenetic diet shift in Commerson's dolphin (*Cephalorhynchus commersonii commersonii*) off Tierra del Fuego. *Polar Biol.* 36, 617–627. (doi:10.1007/s00300-013-1289-5)
- Rossman S, McCabe EB, Barros NB, Gandhi H, Ostrom PH, Stricker CA, Wells RS. 2015 Foraging habits in a generalist predator: sex and age influence habitat selection and resource use among bottlenose dolphins (*Tursiops truncatus*). *Mar. Mamm. Sci.* **31**, 155–168. (doi:10.1111/ mms.12143)
- Jackson-Ricketts J, Iliana Ruiz-Cooley R, Junchompoo C, Thongsukdee S, Intongkham A, Ninwat S, Kittiwattanawong K, Hines EM, Costa DP. 2019 Ontogenetic variation in diet and habitat of Irrawaddy dolphins (*Orcaella brevirostris*) in the Gulf of Thailand and the Andaman Sea. *Mar. Mamm. Sci.* **35**, 492–521. (doi:10.1111/mms.12547)
- Troina G, Botta S, Secchi ER, Dehairs F. 2016 Ontogenetic and sexual characterization of the feeding habits of franciscanas, *Pontoporia blainvillei*, based on tooth dentin carbon and nitrogen stable isotopes. *Mar. Mamm. Sci.* 32, 1115–1137. (doi:10.1111/mms.12316)
- Mendes S, Newton J, Reid RJ, Frantzis A, Pierce GJ. 2007 Stable isotope profiles in sperm whale teeth: variations between areas and sexes. J. Mar. Biol. Assoc. UK 87, 621–627. (doi:10. 1017/S0025315407056019)
- Reisinger RR, Gröcke DR, Lübcker N, McClymont EL, Hoelzel AR, de Bruyn PJN. 2016 Variation in the diet of killer whales *Orcinus orca* at Marion

Island, Southern Ocean. *Mar. Ecol. Prog. Ser.* **549**, 263–274. (doi:10.3354/meps11676)

- Pirotta E, Vighi M, Brotons JM, Dillane E, Cerdà M, Rendell L. 2020 Stable isotopes suggest finescale sexual segregation in an isolated, endangered sperm whale population. *Mar. Ecol. Prog. Ser.* 654, 209–218. (doi:10.3354/ meps13502)
- Hobbs RC *et al.* 2019 Global review of the conservation status of monodontid stocks. *Mar. Fish. Rev.* 81, 3–4.
- Wessel P, Smith WHF. 1996 A global, selfconsistent, hierarchical, high-resolution shoreline database. J. Geophys. Res. (Solid Earth) 101, 8741–8743. (doi:10.1029/96JB00104)
- Heide-Jørgensen MP, Teilmann J. 1994 Growth, reproduction. age structure and feeding habits of white whales (*Delphinapterus leucas*) in West Greenland waters. *Studies of White Whales* (*Delphinapterus leucas*) and Narwhals (Monodon monoceros) in Greenland and Adjacent Waters 39, 195–212.
- Marcoux M. 2009 Social behaviour, vocalization and conservation of narwhals. ARCTIC 61, 456–460. (doi:10.14430/arctic90)
- Marcoux M, Auger-Méthé M, Humphries MM. 2009 Encounter frequencies and grouping patterns of narwhals in Koluktoo Bay, Baffin Island. *Polar Biol.* **32**, 1705–1716. (doi:10.1007/ s00300-009-0670-x)
- Colbeck GJ, Duchesne P, Postma LD, Lesage V, Hammill MO, Turgeon J. 2013 Groups of related belugas (*Delphinapterus leucas*) travel together during their seasonal migrations in and around Hudson Bay. *Proc. R. Soc. B* 280, 20122552. (doi:10.1098/rspb.2012.2552)
- Matthews CJD, Ferguson SH. 2015 Weaning age variation in beluga whales (*Delphinapterus leucas*). J. Mamm. 96, 425–437. (doi:10.1093/ jmammal/gyv046)
- Michaud R. 2005 Sociality and ecology of the odontocetes. In Sexual segregation in vertebrates: ecology of the two sexes (eds KE Ruckstuhl, P Neuhaus), pp. 303–326. Cambridge, UK: Cambridge University Press.
- Garde E, Heide-Jørgensen MP, Hansen SH, Nachman G, Forchhammer MC. 2007 Agespecific growth and remarkable longevity in narwhals (*Monodon monoceros*) from West Greenland as estimated by aspartic acid racemization. J. Mamm. 88, 49–58. (doi:10. 1644/06-mamm-a-056r.1)
- Graham ZA, Garde E, Heide-Jørgensen MP, Palaoro AV. 2020 The longer the better: evidence that narwhal tusks are sexually selected. *Biol. Lett.* 16, 20190950. (doi:10.1098/ rsbl.2019.0950)
- Aubin DJS, Smith TG, Geraci JR. 1990 Seasonal epidermal molt in beluga whales, *Delphinapterus leucas. Can. J. Zool.* 68, 359–367. (doi:10.1139/z90-051)
- Wild LA, Chenoweth EM, Mueter FJ, Straley JM. 2018 Evidence for dietary time series in layers of cetacean skin using stable carbon and nitrogen isotope ratios. *Rapid Commun. Mass Spectrom.* 32, 1425–1438. (doi:10.1002/rcm.8168)
- Giménez J, Ramírez F, Almunia J, Forero MG, de Stephanis R. 2016 From the pool to the sea: applicable isotope turnover rates and diet to

skin discrimination factors for bottlenose dolphins (*Tursiops truncatus*). J. Exp. Mar. Biol. Ecol. **475**, 54–61. (doi:10.1016/j.jembe. 2015.11.001)

- Marcoux M, McMeans BC, Fisk AT, Ferguson SH. 2012 Composition and temporal variation in the diet of beluga whales, derived from stable isotopes. *Mar. Ecol. Prog. Ser.* **471**, 283–291. (doi:10.3354/meps10029)s
- Watt CA, Heide-Jørgensen MP, Ferguson SH. 2013 How adaptable are narwhal? A comparison of foraging patterns among the world's three narwhal populations. *Ecosphere* 4, 1–15. (doi:10.1890/es13-00137.1)
- Watt CA, Ferguson SH. 2015 Fatty acids and stable isotopes (δ¹³C and δ¹⁵N) reveal temporal changes in narwhal (*Monodon monoceros*) diet linked to migration patterns. *Mar. Mamm. Sci.* **31**, 21–44. (doi:10.1111/mms.12131)
- Peterson BJ, Fry B. 1987 Stable isotopes in ecosystem studies. Annu. Rev. Ecol. Syst. 18, 293–320. (doi:10. 1146/annurev.es.18.110187.001453)
- Newsome SD, Clementz MT, Koch PL. 2010 Using stable isotope biogeochemistry to study marine mammal ecology. *Mar. Mamm. Sci.* 26, 509–572. (doi:10.1111/j.1748-7692.2009.00354.x)
- France R, Loret J, Mathews R, Springer J. 1998 Longitudinal variation in zooplankton δ¹³C through the Northwest Passage: inference for incorporation of sea-ice POM into pelagic foodwebs. *Polar Biol.* **20**, 335–341. (doi:10. 1007/s003000050311)
- Hobson KA, Ambrose Jr WG, Renaud PE. 1995 Sources of primary production, benthic-pelagic coupling, and trophic relationships within the Northeast Water Polynya: insights from δ¹³C and δ¹⁵N analysis. *Mar. Ecol. Prog. Ser.* 128, 1–10. (doi:10.3354/meps128001)
- Laidre KL, Heide-Jorgensen MP. 2005 Winter feeding intensity of narwhals (Monodon monoceros). Mar. Mamm. Sci. 21, 45–57. (doi:10.1111/j.1748-7692.2005.tb01207.x)
- Heide-Jørgensen MP, Richard PR, Dietz R, Laidre KL. 2013 A metapopulation model for Canadian and West Greenland narwhals. *Anim. Conserv.* 16, 331–343. (doi:10.1111/acv.12000)
- Yurkowski DJ, Hussey NE, Fisk AT, Imrie KL, Tallman RF, Ferguson SH. 2017 Temporal shifts in intraguild predation pressure between beluga whales and Greenland halibut in a changing Arctic. *Biol. Lett.* **13**, 20170433. (doi:10.1098/ rsbl.2017.0433)
- Wild EM, Arlamovsky KA, Golser R, Kutschera W, Priller A, Puchegger S, Rom W, Steier P, Vycudilik W. 2000 ¹⁴C dating with the bomb peak: an application to forensic medicine. *Nucl.*

Instrum. Methods Phys. Res., Sect. B **172**, 944–950. (doi:10.1016/s0168-583x(00)00227-5)

- Hedges REM, Clement JG, Thomas CDL, O'connell TC. 2007 Collagen turnover in the adult femoral mid-shaft: modeled from anthropogenic radiocarbon tracer measurements. *Am. J. Phys. Anthropol.* 133, 808–816. (doi:10.1002/ajpa.20598)
- Skovrind M, Castruita JAS, Haile J, Treadaway EC, Gopalakrishnan S, Westbury MV, Heide-Jørgensen MP, Szpak P, Lorenzen ED. 2019 Hybridization between two high Arctic cetaceans confirmed by genomic analysis. *Sci. Rep.* 9, 7729. (doi:10.1038/ s41598-019-44038-0)
- Palsbøll PJ, Heide-Jørgensen MP, Dietz R. 1997 Population structure and seasonal movements of narwhals, *Monodon monoceros*, determined from mtDNA analysis. *Heredity* **78**, 284–292. (doi:10.1038/hdy.1997.43)
- Wiig Ø, Heide-Jørgensen MP, Laidre KL, Garde E, Reeves RR. 2012 Geographic variation in cranial morphology of narwhals (*Monodon monoceros*) from Greenland and the eastern Canadian Arctic. *Polar Biol.* 35, 63–71. (doi:10.1007/s00300-011-1032-z)
- R Core Team. 2019 *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See https://www.R-project.org/.
- Hothorn T, Bretz F, Westfall P. 2008 Simultaneous inference in general parametric models. *Biom. J.* 50, 346–363. (doi:10.1002/ bimj.200810425)
- Westgate AJ. 2007 Geographic variation in cranial morphology of short-beaked common dolphins (*Delphinus delphis*) from the North Atlantic. J. Mamm. 88, 678–688. (doi:10.1644/ 06-MAMM-A-177R.1)
- Jackson AL, Inger R, Parnell AC, Bearhop S. 2011 Comparing isotopic niche widths among and within communities: SIBER - stable isotope Bayesian ellipses in R. J. Anim. Ecol. 80, 595–602. (doi:10.1111/j.1365-2656.2011.01806.x)
- Bearhop S, Adams CE, Waldron S, Fuller RA, MacLeod H. 2004 Determining trophic niche width: a novel approach using stable isotope analysis. *J. Anim. Ecol.* **73**, 1007–1012. (doi:10.1111/j.0021-8790.2004. 00861.x)
- Newsome SD, Martinez del Rio C, Bearhop S, Phillips DL. 2007 A niche for isotopic ecology. *Front. Ecol. Environ.* 5, 429–436. (doi:10.1890/ 060150.1)
- Plummer M. 2019 rjags: Bayesian graphical models using MCMC. *R package version* 4. See https://cran.r-project.org/package=rjags.

- Degerbøl M, Nielsen NL. 1930 Biologiske lagttagelser over og maalinger af hvidhvalen (Delphinapterus leucas (Pall.)) og dens Fostre. (with an English summary.) Meddr Grønland 77, 119–144.
- Hansen JH, Hedeholm RB, Sünksen K, Christensen JT, Grønkjær P. 2012 Spatial variability of carbon (δ¹³C) and nitrogen (δ¹⁵N) stable isotope ratios in an Arctic marine food web. *Mar. Ecol. Prog. Ser.* **467**, 47–59. (doi:10. 3354/meps09945)
- Szpak P, Julien MH, Royle TCA. 2020 Sexual differences in the foraging ecology of 19th century beluga whales (*Delphinapterus leucas*) from the Canadian High Arctic. *Mar. Mamm. Sci.* 36, 451–471. (doi:10.1111/ mms.12655)
- Szpak P, Orchard TJ, McKechnie I, Gröcke DR. 2012 Historical ecology of late Holocene sea otters (*Enhydra lutris*) from northern British Columbia: isotopic and zooarchaeological perspectives. J. Archaeol. Sci. **39**, 1553–1571. (doi:10.1016/j.jas.2011.12.006)
- Laidre KL, Stirling I, Lowry LF, Wiig O, Heide-Jørgensen MP, Ferguson SH. 2008 Quantifying the sensitivity of Arctic marine mammals to climate-induced habitat change. *Ecol. Appl.* 18, 597–125. (doi:10.1890/06-0546.1)
- Nelson DE, Møhl J, Heinemeier J, Arneborg J. 2012 Stable carbon and nitrogen isotopic measurements of the wild animals hunted by the Norse and the Neo-Eskimo people of Greenland. J. North Atlantic 2012, 40–50. (doi:10.3721/037.004.s304)
- Noren SR, Williams TM. 2000 Body size and skeletal muscle myoglobin of cetaceans: adaptations for maximizing dive duration. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **126**, 181–191. (doi:10.1016/S1095-6433(00)00182-3)
- Heide-Jørgensen MP, Hammeken N, Dietz R, Orr J, Richard PR. 2001 Surfacing times and dive rates for narwhals (*Monodon monoceros*) and belugas (*Delphinapterus leucas*). ARCTIC 54, 284–298. (doi:10.14430/arctic788)
- Loseto LL, Richard P, Stern GA, Orr J, Ferguson SH. 2006 Segregation of Beaufort Sea beluga whales during the open-water season. *Can. J. Zool.* 84, 1743–1751. (doi:10.1139/z06-160)
- Noren SR, Suydam R. 2016 Navigating under sea ice promotes rapid maturation of diving physiology and performance in beluga whales. J. Exp. Biol. 219, 2828–2836. (doi:10.1242/jeb.143644)
- Ferguson SH, Kingsley MCS, Higdon JW. 2012 Killer whale (*Orcinus orca*) predation in a multiprey system. *Popul. Ecol.* 54, 31–41. (doi:10. 1007/s10144-011-0284-3)