

Phenotypic Spectrum of Seizure Disorders in MBD5-Associated Neurodevelopmental Disorder

Kenneth A. Myers, MD, PhD, Carla Marini, MD, PhD, Gemma L. Carvill, PhD, Amy McTague, PhD, Julie Panetta, MBBS, Chloe Stutterd, MBBS, Thorsten Stanley, MBChB, Samantha Marin, MD, John Nguyen, BSc, Carmen Barba, MD, PhD, Anna Rosati, MD, PhD, Richard H. Scott, MD, Heather C. Mefford, MD, PhD, Renzo Guerrini, MD, FRCP, and Ingrid E. Scheffer, MBBS, PhD

Correspondence
Dr. Myers
sfuken1@gmail.com

Neurol Genet 2021;7:e579. doi:10.1212/NXG.0000000000000579

Abstract

Objective

To describe the phenotypic spectrum in patients with MBD5-associated neurodevelopmental disorder (MAND) and seizures; features of MAND include intellectual disability, epilepsy, psychiatric features of aggression and hyperactivity, and dysmorphic features including short stature and microcephaly, sleep disturbance, and ataxia.

Methods

We performed phenotyping on patients with MBD5 deletions, duplications, or point mutations and a history of seizures.

Results

Twenty-three patients with MAND and seizures were included. Median seizure onset age was 2.9 years (range 3 days–13 years). The most common seizure type was generalized tonic-clonic; focal, atypical absence, tonic, drop attacks, and myoclonic seizures occurred frequently. Seven children had convulsive status epilepticus and 3 nonconvulsive status epilepticus. Fever, viral illnesses, and hot weather provoked seizures. EEG studies in 17/21 patients were abnormal, typically showing slow generalized spike-wave and background slowing. Nine had drug-resistant epilepsy, although 3 eventually became seizure-free. All but one had moderate-to-severe developmental impairment. Epilepsy syndromes included Lennox-Gastaut syndrome, myoclonic-atonic epilepsy, and infantile spasms syndrome. Behavioral problems in 20/23 included aggression, self-injurious behavior, and sleep disturbance.

Conclusions

MBD5 disruption may be associated with severe early childhood-onset developmental and epileptic encephalopathy. Because neuropsychiatric dysfunction is common and severe, it should be an important focus of clinical management.

From the Research Institute of the McGill University Health Centre (K.M.), Montreal, PQ; Division of Child Neurology (K.M.), Department of Pediatrics, Montreal Children's Hospital, McGill University, Montreal, PQ; Department of Neurology & Neurosurgery (K.M.), Montreal Children's Hospital, McGill University, Montreal, PQ; Child Neurology and Psychiatry (C.M.), Salesi Pediatric Hospital, United Hospitals of Ancona, Ancona, Italy; Division of Genetic Medicine (G.L.C., J.N., H.C.M.), Department of Pediatrics, University of Washington, Seattle, WA; Department of Neurology (A.M.), Great Ormond Street Hospital for Children, London, UK; Developmental Neurosciences Programme (A.M.), UCL Great Ormond Street Institute of Child Health, London, UK; Neurology Network Melbourne (J.P.), Melbourne, Victoria, Australia; Murdoch Children's Research Institute (C.S., I.E.S.), Parkville, Victoria, Australia; Department of Paediatrics and Child Health (T.S.), School of Medicine and Health Sciences, University of Otago, Wellington, New Zealand; Division of Neurology (S.M.), Department of Pediatrics, Rady Faculty of Health Sciences, University of Manitoba, Winnipeg, Manitoba, Canada; Neurology Unit and Neurogenetic Laboratories (C.B., A.R., R.G.), Meyer Children's Hospital, Florence, Italy; Department of Clinical Genetics (R.H.S.), Great Ormond Street Hospital, London, UK; Epilepsy Research Centre (I.E.S.), Department of Medicine, The University of Melbourne, Austin Health, Heidelberg, Victoria, Australia; Department of Paediatrics (I.E.S.), Royal Children's Hospital, The University of Melbourne, Parkville, Victoria, Australia; and The Florey Institute of Neuroscience and Mental Health (I.E.S.), Heidelberg, Victoria, Australia.

Go to [Neurology.org/NG](https://www.neurology.org/NG) for full disclosures. Funding information is provided at the end of the article.

The Article Processing Charge was funded by the authors.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND), which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Glossary

DS = Dravet syndrome; **FS+** = febrile seizures plus; **LGS** = Lennox-Gastaut syndrome; **MAE** = myoclonic-atonic epilepsy; **MAND** = *MBDS*-associated neurodevelopmental disorder.

MBDS (methyl-CpG-binding domain protein 5; OMIM #611472), located on chromosome 2q23.1, belongs to a family of genes involved in DNA methylation and chromatin remodeling.¹ Disruption of this gene through heterozygous deletion or point mutation leads to *MBDS*-associated neurodevelopmental disorder (MAND), with some patients described as having a Kleefstra syndrome phenotypic spectrum.² Intellectual disability occurs in all patients, with seizures, dysmorphic features including short stature and microcephaly, sleep disturbance, ataxia, aggressive behavior, and hyperactivity frequently observed.^{3–5} *MBDS* deletions are not especially rare, found in 0.05% (1 in 2,000) of 17,477 samples that underwent clinical microarray testing.³ Deletions or mutations are almost always de novo although inheritance from mildly affected or mosaic parents has been reported.⁶

Seizures occur in over 80% of patients with MAND; however, the epileptology has not yet been delineated.^{3–5,7} Here, we analyzed the phenotypic spectrum in 23 patients with heterozygous deletion, duplication, or point mutation of *MBDS* and a history of seizures.

Methods

We searched our epilepsy genetics research databases for patients with pathogenic variants involving *MBDS* and identified 9 individuals. Two patients were identified through the Epi4K research testing program.⁸ Fourteen additional families volunteered to participate after social media patient groups brought attention to our research. Patients were ascertained from Australia, Italy, New Zealand, Finland, Canada, Germany, the United Kingdom, and the United States. We conducted personal interviews with all patients' families and reviewed medical records, EEG, neuroimaging, and genetic testing results. For medication response, we classified drugs as effective if caregivers reported a clear reduction in seizure frequency, even if seizures were not completely controlled. Wherever possible, epilepsy syndromes were classified according to the International League Against Epilepsy classification.^{9,10} Genetic variants were classified per American College of Medical Genetics and Genomics guidelines.^{11,12}

Standard Protocol Approvals, Registrations, and Patient Consents

Written informed consent was provided for all patients by a parent or legal guardian. This study was approved by the Human Research Ethics Committee, Austin Health, or the local ethics committee.

Data Availability

Anonymized data will be shared by request from any qualified investigator.

Results

Twenty-three patients from 22 families were identified with a history of seizures in the context of a *MBDS* molecular lesion.

Genetic Findings

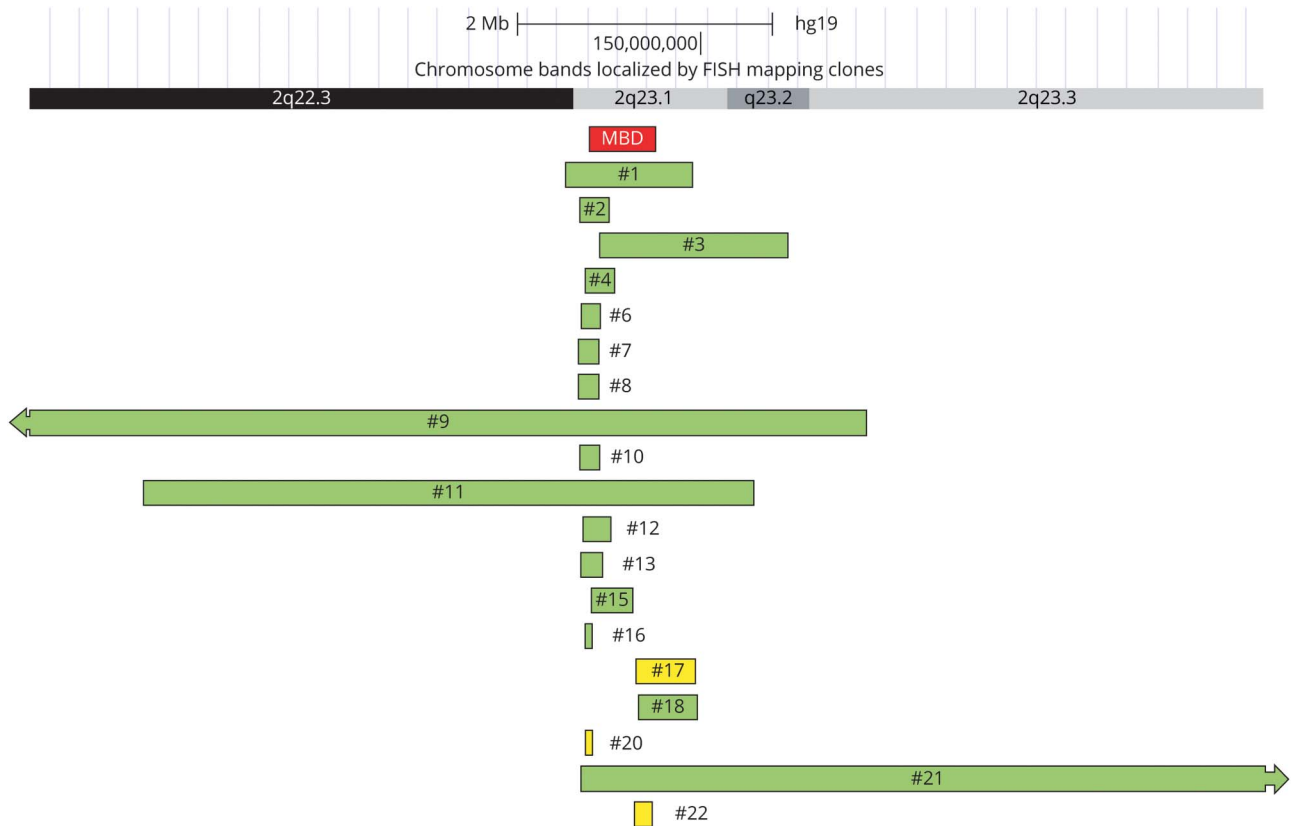
Nineteen of the 23 patients had heterozygous 2q23.1 deletions: 15 resulted in partial deletion of *MBDS* (11 proximal, 3 distal, and 1 data not available) and 4 had complete loss of *MBDS* (figure 1, table 1). Two patients had 2q23 micro-duplications with partial *MBDS* duplication (1 distal and 1 intragenic). The remaining 2 patients had point mutations resulting in truncation: p.Arg200* and p.Thr157Glnfs*4 (previously published¹³).

Pathogenic variants were de novo in 9 patients, with results from both parents not available in 10. Mutations were inherited from a mosaic parent in 4 patients from 3 families (figure 2). Patients 7 and 8 were brothers whose mosaic mother (20%–30% mosaicism in blood) had mild-moderate intellectual disability, but no history of seizures. Patient 13 had a mosaic carrier father (36% mosaicism in blood) of normal intellect and no history of seizures. Her older brother also inherited the deletion and had autism spectrum disorder and intellectual disability, without seizures. Patient 19 had parents who both tested negative for his mutation on blood-derived DNA; however, chorionic villus sampling of their next pregnancy showed that the fetus carried the mutation and the pregnancy was terminated. This suggested that 1 parent must be mosaic, with the mutation possibly limited to gonadal tissue. Patient 12 was mosaic for his deletion, affecting ~65% of cells in saliva. The results of other genetic testing performed are given in table e-1, [links.lww.com/NXG/A402](https://www.lww.com/NXG/A402).

Seizures

Seizures began at median age 2.9 years (range 3 days to 13 years; table 2). Bilateral tonic-clonic seizures occurred in 19/23 (83%) patients, with focal impaired awareness seizures (FIAS; 9/23; 39%), tonic (8/23; 35%), unclassified drop attacks (7/23; 30%), myoclonic (7/23; 30%), atypical absences (7/23; 30%), myoclonic-atonic (1/23; 4%), atonic (1/23; 4%), hemiclonic (1/23; 4%), unclassified staring spells (absences vs FIAS; 1/23; 4%), and epileptic spasms (1/23; 4%) also observed. Convulsive status epilepticus occurred in 7/23 (30%) patients and nonconvulsive status epilepticus in 3/23 (13%). Fever and viral illnesses provoked seizures in 11

Figure 1 Copy Number Variants of Patients With MBD5-Associated Neurodevelopmental Disorder



Green denotes deletion; yellow denotes duplication. This figure includes a screenshot from UCSC genome browser (genome.ucsc.edu).

patients; 1 patient's seizures were triggered by painful stimuli (e.g., mild accidental falls to the ground).

The most common interictal EEG findings were diffuse background slowing (11/23; 48%) and generalized spike-wave or polyspike-wave activity (10/23; 43%). Focal slowing and/or multifocal epileptiform discharges occurred in 9/23 (39%) patients. Epilepsy syndromes were defined in 7 patients: 3 had Lennox-Gastaut syndrome (LGS), 2 had myoclonic-atonic epilepsy (MAE), 1 had infantile spasms syndrome, and 1 had febrile seizures plus (FS+).

Ten patients had drug-resistant epilepsy. Although no drug was clearly superior, valproate showed the most consistent beneficial effect (12/14 cases), while carbamazepine exacerbated seizures in patient 2. Patient 2 became seizure-free during periods of illness and had dramatic reduction in seizure frequency on the ketogenic diet. Ketogenic diet therapy was also trialed in patient 5 with no benefit.

Twenty-one patients had available brain MRI results, with normal findings in 17/21 (85%). Of the patients with abnormal MRI, none had epileptogenic lesions (table 3). Patient 12 had a normal MRI, but fluorodeoxyglucose PET showed severe hypometabolism in the temporoparietal and occipital regions bilaterally.

Development and Behavior

Developmental impairment was present in all patients: severe in 14, moderate in 8, and mild in 1 (table 3). Regression with seizures occurred in 5 patients. The most dramatic regression occurred in patient 2 whose early developmental milestones were normal to mildly delayed (sat at 8 months, walked at 17 months, and first word at 12 months). When seizures began at age 2 years, there was marked developmental regression, particularly involving language. By age 4 years, he had only single words that were mostly unintelligible.

Marked behavioral difficulties were reported in 16 patients, with hyperactivity and aggression most common. Self-injurious behaviors occurred frequently, including finger and nail biting, scratching and picking lips until they bled. Behavioral difficulties were not controlled by stimulants, antipsychotics, and sedatives although methylphenidate elicited some benefit. Sleep disturbance during childhood was reported in 17 patients and involved frequent nocturnal awakenings. Nine patients had microcephaly.

Three patients had signs of metabolic dysfunction, which were considered coincidental. Patient 2 had decreased biotinidase activity suggesting a partial biotinidase deficiency; his parents reported some improvement in seizure control with biotin therapy. Patient 5 had borderline hypoglycorrhachia on

Table 1 Heterozygous Pathogenic Variants and Affecting *MBD5*

No.	<i>MBD5</i> effect; inheritance	Del/Dup size	Breakpoints (build)	Genes affected
1	Complete deletion; de novo	2.2 Mb	SNP-A-189490 to SNP-A-226411	<i>ACVR2A, ORC4, MBD5, EPC2</i>
2	Proximal deletion (exons 1–2); N/A	0.2 Mb	148734048-148932576 (Hg19)	<i>ORC4, MBD5</i>
3	Distal deletion (exons 3–15); N/A	1.5 Mb	148839546-150345992 (Hg19)	<i>MBD5, EPC2, KIF5C, LYPD6B, LYPD6</i>
4	Proximal deletion (exons 1–3); de novo	0.2 Mb	148489085-148678668 (Hg18)	<i>ORC4, MBD5</i>
5	p.Thr157Glnfs*4 truncation; de novo	N/A	N/A	<i>MBD5</i>
6	Proximal deletion (exons 1–2); de novo	0.1 Mb	148715661-148842706 (Hg19)	<i>ORC4, MBD5</i>
7	Proximal deletion (exons 1–2); inherited (mosaic mother)	0.1 Mb	148669363-148788392 (Hg19)	<i>ACVR2A, ORC4, MBD5</i>
8	Proximal deletion (exons 1–2); inherited (mosaic mother)	0.1 Mb	148669363-148788392 (Hg19)	<i>ACVR2A, ORC4, MBD5</i>
9	Complete del; N/A	10.1 Mb	141060000-151150000 (Hg18)	<i>LRP1B, KYNU, ARHGAP15, GTDC1, ZEB2, ACVR2A, ORC4, MBD5, EPC2, KIF5C, LYPD6B, LYPD6, MMADHC, RND3</i>
10	Proximal deletion (exons 1–2); de novo	0.1 Mb	148703861-148829749 (Hg19)	<i>ORC4, MBD5</i>
11	Complete deletion; N/A	3.7 Mb	146855669-150602070 (Hg19)	<i>ACVR2A, ORC4, MBD5, EPC2, KIF5C, LYPD6B, LYPD6, MMADHC</i>
12	Proximal deletion (exons 1–3); de novo (mosaic; 65% cells)	0.2 Mb	148758479-148954077 (Hg19)	<i>ORC4, MBD5</i>
13	Proximal deletion (exons 1–2); inherited mosaic father)	0.2 Mb	148734046-148897348 (Hg19)	<i>ORC4, MBD5</i>
14	Partial deletion; N/A	0.2 Mb	Not available	Not available
15	Proximal deletion (exons 1–4); N/A	0.3 Mb	148462331-148741534 (Hg18)	<i>ORC4, MBD5</i>
16	Proximal deletion (exons 1–2); N/A	0.05 Mb	148755020-148802565 (Hg19)	<i>ORC4, MBD5</i>

Table 1 Heterozygous Pathogenic Variants and Affecting *MBD5* (continued)

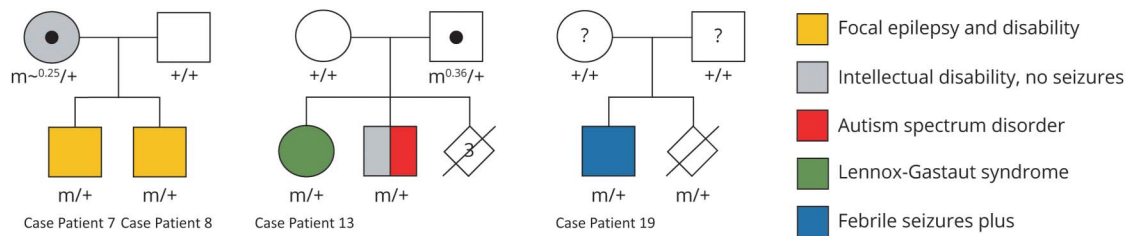
No.	<i>MBD5</i> effect; inheritance	Del/Dup size	Breakpoints (build)	Genes affected
17	Distal duplication (exons 7–15); N/A	0.4 Mb	149218851-149655290 (Hg19)	<i>MBD5, EPC2, KIF5C</i>
18	Distal deletion (exons 7–15); de novo	0.6 Mb	149219863-149796844 (Hg19)	<i>MBD5, EPC2, KIF5C</i>
19	c.598C > T; p.Arg200* (truncation); inherited (parent mosaic)	N/A	N/A	<i>MBD5</i>
20	Proximal deletion (exons 1–2); N/A	0.02 Mb	148764203-148786336 (Hg19)	<i>ORC4, MBD5</i>
21	Complete deletion; de novo	6.5 Mb	148438001-154962504 (Hg19)	<i>ACVR2A, ORC4, MBD5, EPC2, KIF5C, LYPD6B, LYPD6, MMADHC, RND3, RBM43, NMI, TNFAIP6, RIF1, NEB, ARL5A, CACNB4, STAM2, FMNL2, PRPF40A, ARL6IP6, RPRM, GALNT13</i>
22	Intragenic duplication (exon 5); de novo	0.085 Mb	149052433-149137634 (Hg19)	<i>MBD5</i>
23	Distal deletion (exons 6–15); de novo	0.789 Mb	148914820-149703672 (Hg18)	<i>MBD5, EPC2, KIF5C, LYPD6B</i>

Abbreviations: N/A = inheritance unknown; MLPA = multiplex ligation-dependent probe amplification, WES = whole exome sequencing, WGS = whole genome sequencing.

lumbar punctures at 8 and 10 years (CSF glucose 2.9 and 2.2 mM with CSF:serum ratios of 0.41 and 0.43, respectively). Patient 20 was diagnosed with galactosemia on newborn screening.

Although our cohort is relatively small, we looked for genotype-phenotype correlations. Surprisingly, phenotypic severity was not more severe in patients with deletions affecting multiple genes. For example, patient 5 with a simple *MBD5* truncation point mutation had one of the most severe phenotypes with a refractory developmental and epileptic encephalopathy and severe behavioral disturbance. The lack of strong genotype-phenotype correlation was further emphasized by the 2 families in which sibling pairs inherited the same deletion from a mosaic parent but had marked phenotypic differences, with 1 not having epilepsy at all. We did, however, observe greater phenotypic severity in patients with complete deletion of *MBD5* compared with those with only partial gene deletion. Of those with partial deletions, the mildest phenotypes (patients 10, 16, and 20) were seen in individuals with loss of the proximal end of *MBD5*.

Figure 2 Pedigrees of Families With Inheritance From Mosaic Parent



The parents of patient 19 (right) both tested negative for the mutation on blood sequencing; however, chorionic villus sampling of their second pregnancy showed that the fetus carried the same mutation, so one of the parents is assumed to have low-level (likely gonadal) mosaicism. “m” = deletion/truncation mutation affecting *MBDS*; “m^{0.x}” = mosaic with x% of cells having deletion; “+” = wild type. Central black circle indicates mosaic carrier; central question mark indicates possible mosaic carrier.

Discussion

With increasing access to molecular testing including high-resolution chromosomal microarray, more patients with disruption of *MBDS* resulting in MAND will be identified. Understanding the epileptology of this genetic disease is critical for prompt diagnosis and optimal management. We analyzed the phenotypes of a global cohort of patients with *MBDS* pathogenic variants, which often resulted in a severe early childhood-onset developmental and epileptic encephalopathy.

A broad spectrum of phenotypes was observed, and no genotype-phenotype correlation could be identified. The size of the cohort limits this analysis, and as the epileptology of further cases is described, correlations may become apparent. Phenotypes were more severe in patients with complete *MBDS* gene deletion and milder in those with deletion of only the proximal end of the gene. This phenomenon could be explained by the deletion of fewer and different contiguous genes in the patients with proximal deletions. However, substantial phenotypic differences were noted in siblings carrying the same deletion, suggesting that variable expressivity in MAND reflects the involvement of modifier genes, epigenetic, or environmental influences.

Our molecular data highlight the importance of mosaicism, both in patient 12 and, even more critically, in 3/22 (14%) families who had 2 affected pregnancies. In 1 family, antenatal testing identified a second affected child despite negative testing of parental blood, strongly implicating gonadal mosaicism or low-level mosaicism in 1 parent that was missed on conventional sequencing. This suggests that inheritance from parents with low-level mosaicism may be more frequent than previously thought with key implications for reproductive counseling.¹⁴

Multiple seizure types were usual, including both generalized and focal, with tonic-clonic, focal, absence, atonic, myoclonic, and tonic seizures. EEG studies showed generalized and

multifocal epileptiform activity. Seizures were often initially medically refractory but sometimes spontaneously resolved in childhood (age 4–7 years). Neuropsychiatric and developmental features were prominent including moderate-to-severe developmental impairment, language deficits, sleep disturbance, hyperactivity, and aggression.

Fever provoked seizures in 10 patients in our cohort, a pattern reported in 4 published cases.^{4,5,15,16} An additional study described hemiclonic seizures with alternating sides beginning at 10 months of age, a feature classically associated with Dravet syndrome (DS), a well-recognized developmental and epileptic encephalopathy associated with *SCN1A* mutations.^{17,18} Our cases, together with those reported, suggest that epilepsy in MAND sometimes has phenotypes on the genetic epilepsy with febrile seizures plus spectrum, including DS, MAE, FS+, and febrile seizures.¹⁹

MAND is typically associated with normal neuroimaging or thin corpus callosum with mild hypomyelination in rare cases. There are rare reports of focal cerebral malformations with *MBDS* pathogenic variants, but these were associated with relatively large heterozygous deletions involving loss of many genes other than *MBDS*.^{4,20,21}

The neuropsychiatric and behavioral abnormalities commonly observed in *MBDS* pathogenic variants included sleep disturbance, developmental disability, language impairment, aggressive, and hyperactive behavior.⁴ When these occur, families should be counseled that these features are likely intrinsic to the genetic syndrome rather than secondary to medications or uncontrolled seizures. This is an important observation because some patients may undergo unnecessary investigations or medication changes, potentially jeopardizing seizure control, when the etiology of these behaviors is poorly understood.

Our findings should, however, be considered with some caution, given that this study had several limitations. Given the small size of the cohort, it was not possible to conduct statistical analyses or to make meaningful comments

Table 2 Epilepsy Features

No./ Sex/ Age	Sz onset	Seizure types (initial seizure type in bold)	Seizure triggers	SE?	EEG	Electroclinical syndrome	Epilepsy course	Effective meds	Ineffective meds
1/ M/ 10 y	2 y	GTC, atonic, myoclonic, myoclonic-atic	Hot weather, febrile illnesses	Yes ^a	Diffuse slowing; generalized SW/PSW	MAE	Refractory initially, Sz-free since age 4 y	PHT, CLN, TPM, LEV, VPA	—
2/ M/4 y	2 y	Atypical absence, DA, GTC, myoclonic, tonic	None	Yes ^a	Diffuse slowing; generalized ShW, GSW, increase in sleep	—	Refractory	VPA, LTG, KD, biotin	CBZ (worsened)
3/ M/5 y	4 m	GTC, myoclonic, tonic	Viral illnesses	Yes	Diffuse slowing; focal slowing (L temporal), generalized and multifocal ShW, SW at 4 m (not present at 4 y)	—	Only 3 sz clusters but 2 involved SE	LEV, CLN, TPM	—
4/F/ 6 y	3 y	Atypical absence, GTC	Pain	No	Diffuse slowing; generalized SW	—	Controlled with LTG	LTG	—
5/F/ 26 y	6 m	Atypical absence, DA, FIAS, GTC, myoclonic, tonic	Hot weather, febrile illnesses	Yes	Diffuse slowing; parieto-occipital spikes independent bilaterally; generalized SW/PSW	—	Refractory	LTG, PHT, VPA, CLB, LEV	GAB, TPM, KD
6/F/ 10 y	3 y	FS, GTC	Fever, sleep	No	Normal background; centrottemporal or diffuse SW	—	2 sz total; sz-free for >3 y	VPA	—
7/ M/ 10 y	4 y	FS, FBTC, FIAS	Febrile illnesses	No	Normal background; diffuse epileptiform discharges with TPO predominance	Focal epilepsy	Refractory initially; now sz-free	VPA, LEV, TPM	—
8/ M/ 11 y	7 y	FBTC, FIAS	No	No	Normal background; diffuse epileptiform discharges with TPO predominance	Focal epilepsy	Sz-free	LEV	—
9/ M/ 13 y	10 m	Hemiclonic, F	Sleep	Yes	Diffuse and multifocal epileptiform discharges	Focal epilepsy	Refractory initially; Sz-free since 4 y	VPA, TPM, CLB, PHT	VIG, LTG, LEV
10/ F/ 3.5 y	2.5 y	FS	Febrile illness	No	Normal	FS	Only one event; not treated	—	—
11/ F/11 y	0.3 y	ES, GTC, FIAS	Hot weather	No	Hypsarrhythmia with ES; normal when other sz types emerged	West	Responded well to medication	ACTH, VPA, OXC	TPM, VGB, CLN
12/ M/ 13 y	2.5 y	GTC, FIAS, myoclonic, tonic, DA, atypical absence	Sleep	Yes	Diffuse slowing; multifocal epileptiform discharges; generalized slow SW and PSW; PFA	LGS	Refractory	VPA, ETX, LTG, LEV, TPM	CLB
13/ F/28 y	2.5 y	GTC, tonic, atypical absence, myoclonic, DA, FIAS	Sleep, illness	Yes	Generalized 1.5 Hz SW, PSW, ShW; diffuse slowing	LGS	Refractory	LTG, VPA, CLB, CLN, LEV	TPM, ETX
14/ M/5 y	1.5 y	FS, GTC, DA, myoclonic	Febrile illness, sleep	Yes	Diffuse slowing; generalized slow SW, PSW	MAE	Refractory	VPA, CLB, LTG	TPM, LEV

Continued

Table 2 Epilepsy Features (*continued*)

No./ Sex/ Age	Sz onset	Seizure types (initial seizure type in bold)	Seizure triggers	SE?	EEG	Electroclinical syndrome	Epilepsy course	Effective meds	Ineffective meds
15/ M/9 y	1.5 y	Tonic, DA GTC, atypical absence, gelastic	Febrile illness, sleep	Yes	Diffuse slowing; generalized and multifocal (R and L frontocentral) slow 1.5–2 Hz SW, PSW	LGS	Refractory	OXC, LEV, CLB, NIT	—
16/ M/15 y	13 y	GTC, atypical absence	—	No	Normal background; 3–3.5 Hz generalized SW	—	Only 3 GTC	VPA	LTG
17/ F/7 y	3 y	FS, hemiclonic, DA	Febrile illness	Yes	Official reports not available; parents reported ESES diagnosis at age 3 y	—	Refractory	LEV, CLN, IVIG	—
18/ M/11 y	2 y	FS	Febrile illness	No	Not performed	FS	Only one febrile seizure; not medicated	—	—
19/ M/3.3 y	9 m	FS, GTC	Febrile illness	No	Normal	FS+	Seizure-free on LEV	LEV	VPA
20/ M/3.3 y	2.9 y	Staring spells (absences vs FIAS)	—	No	Normal (at 6 m)	—	3 events; not medicated	—	—
21/ M/2.3 y	3 d	FIAS, tonic	—	No	Normal	—	Cluster of events in first week of life, then 2 likely FIA seizures at 26 m; not medicated	—	—
22/ F/12.5 y	8 y	FIAS, GTC, tonic	—	No	Focal SW and PSW right frontocentral region	—	Sz controlled on OXC	OXC	—
23/ M/11 y	18 m	GTC	—	No	Diffuse slowing	—	Sz controlled on VPA and LEV	VPA, LEV	—

Abbreviations: ACTH = adrenocorticotropic hormone; CBZ = carbamazepine; CLB = clobazam; CLN = clonazepam; DA = drop attacks; ES = epileptic spasms; F = focal; FBTC = focal to bilateral tonic-clonic; FIAS = focal with impaired awareness seizures; FS = febrile seizures; FS+ = febrile seizures plus; G = gelastic; GAB = gabapentin; GTC = generalized tonic-clonic; IVIG = intravenous immunoglobulin; KD = ketogenic diet; LEV = levetiracetam; LGS = Lennox-Gastaut syndrome; LTG = lamotrigine; MAE = myoclonic-atonic epilepsy; NCSE = nonconvulsive status epilepticus; NIT = nitrazepam; OXC = oxcarbazepine; PB = phenobarbital; PFA = paroxysmal fast activity; PHT = phenytoin; PSW = polyspike-wave; RUF = rufinamide; ShW = sharp-slow wave; SW = spike-wave; Sz = seizure; TPM = topiramate; TPO = temporoparieto-occipital; VGB = vigabatrin; VPA = valproic acid.

Medications were classified as “effective” if there was reported to be at least partial improvement in seizure control, and “ineffective” if there was no apparent improvement.

^aStatus epilepticus was the initial presentation of seizures.

regarding genotype-phenotype correlation. As a retrospective study in which many patients self-referred, there are potential biases, most notably selection and recall biases. There was a preponderance of patients with copy number variants in our cohort because only 2 individuals had truncating *MBDS* variants. This may relate to the accessibility of different investigations because CGH microarrays are far more readily available than gene panels around the world so that point mutations may be being missed in patients who lack access to next-generation sequencing.

In summary, patients with *MBDS* pathogenic variants and seizures may have a range of phenotypes, including early childhood-

onset developmental and epileptic encephalopathy. Epilepsy syndromes include infantile spasms syndrome, LGS, and MAE. Convulsive status epilepticus and nonconvulsive status epilepticus are fairly frequent, and seizures are often provoked by fever or environmental hyperthermia. Parental genetic testing should be offered because inheritance from mosaic parents may be more common than currently appreciated with important implications for genetic counseling.

Disclosure

K.A. Myers has received a travel grant from Zynerba and receives/has received research support from Fonds de

Table 3 Developmental Impairment, Behavioral Issues, and Other Clinical Features

#/Sex/ Age	Developmental milestones (sat/walked/first word) ID degree Regress with seizures?	Microcephaly?	Sleep issues	Behavioral issues	MRI brain	Musculoskeletal abnormalities
1/M/ 10 y	8 m/20 m/2 y Severe No	Y	Frequent waking from age 4 y	Agitation, hyperactivity, anxiety, aggression; self-mutilation	Normal at age 2.2 y; mild thickening of corpus callosum noted at 14 y	Idiopathic torticollis
2/M/4 y	8 m/17 m/12 m Severe Yes	N	Frequent waking with nocturnal tonic seizures	N	Normal (age 2.1 y)	N
3/M/5 y	18 m/4 y/3 y Severe No	Y	No	Irritability, aggression, violent tantrums, obsessiveness, inflexibility	Normal (age 4.5 mths)	Abnormal increased left foot and ankle tone with fixed, plantar-flexed, hindfoot varus, inverted position
4/F/6 y	6 m/18 m/3.5 y Severe No	N	Sleep initiation difficulty	ADHD, anxiety, tantrums, self-mutilation	Normal (age 3.2 y)	N
5/F/26 y	?/3 y/NV Severe No	N	Frequent waking	Compulsive behavior, severe agitation, violence, self-mutilation	"Global reduction in white matter" (age 10 y)	N
6/F/10 y	?/?/NV Severe No (behavior worsened at 6 y)	Y	Parasomnias, bruxism and sleep talking	Fluctuating aggressiveness, irritability	Chiari I malformation (age 6.5 y)	N
7/M/ 10 y	?/2 y/2.5 y Severe No	N	N	Hyperactivity	Normal (age 5 y)	N
8/M/ 11 y	?/2 y/4 y Severe No	Y	Pavor nocturnus	Hyperactivity	Normal (age 6 y)	N
9/M/ 13 y	20 m/NA/NV Severe No	Y	N	N	Mild hypomyelination, thin CC (age 3 y)	N
10/F/ 3.5 y	7 m/19 m/11 m Mild No	N	N	N	Not performed	N
11/F/ 11 y	17 m/23 m/NV Severe No	N	Frequent waking; "tears apart bedroom"	Bangs head, scratches arms; self-stimulation	Normal (age 5 mths)	N
12/M/ 13 y	8 m/15 m/13 m Moderate-severe Yes	N	Frequent waking and very active	N	Normal (age 3 y)	N
13/F/ 28 y	8 m/15 m/? Moderate Yes	Y	Frequent waking	Obsessive behaviors and tantrums	Normal (age 8 y)	N
14/M/ 5 y	10 m/19 m/NV Moderate-severe No	N	Frequent waking (average 10/night)	Bites self; bangs head against wall	Normal	N
15/M/ 9 y	9 m/17 m/17 m Moderate-severe Yes	N	Frequent waking	Tantrums; bites hands	Normal (ages 2.5 y and 10 y)	N

Continued

Table 3 Developmental Impairment, Behavioral Issues, and Other Clinical Features (continued)

#/Sex/ Age	Developmental milestones (sat/walked/first word) ID degree Regress with seizures?	Microcephaly?	Sleep issues	Behavioral issues	MRI brain	Musculoskeletal abnormalities
16/M/ 15 y	7 m/15 m/18 m Moderate Yes	N	Frequent waking	Misbehaves frequently	Normal	N
17/F/ 7 y	3 y/NA/NV severe No	Y	Frequent waking with nocturnal seizures	N	Not available	Congenital bilateral talipes equinovarus
18/M/ 11 y	1y/2y 3m/2y Severe No	N	Frequent waking from age 6 y	Poor attention and concentration, hyperactivity, repetitive hand mannerisms	Normal (age 2 y)	N
19/M/ 3.3 y	8m/16m/2y 4m Moderate No	N	N	Hyperactivity	Normal	N
20/M/ 3.3 y	12 m/21 m/2.5 y Moderate No	N	N	Pushing other children; pinching himself	Normal	N
21/M/ 2.3 y	2 y/NA/10 m Moderate No	N	Frequent waking	N	Normal (age 2.3 y)	N
22/F/ 12.5 y	10 m/2.5 y/4 y Moderate No	Y	Frequent waking	N	Normal (age 8.5 y)	N
23/M/ 11 y	6 m/2.5 y/16 m Severe No	Y	Frequent waking	Food-hoarding behaviors and other obsessions; had self-injurious behaviors when younger; now, hits and scratches others	Focal T2 hyperintensities in occipital white matter; mild cerebral and cerebellar atrophy (age 1.9 y)	N

Abbreviations: ADHD = attention deficit hyperactivity disorder; CC = corpus callosum; NA = non-ambulatory; NV = non-verbal.

Recherches du Québec—Santé, Citizens United for Research in Epilepsy (CURE), Research Institute of the McGill University Health Centre, the Savoy Foundation, Koolen-de Vries Foundation, and Dravet Canada. I.E. Scheffer serves on the editorial boards of *Neurology*[®] and *Epileptic Disorders*; may accrue future revenue on a pending patent re: Therapeutic compound; has received speaker honoraria from Athena Diagnostics, UCB, GSK, Eisai, and Transgenomics; has received scientific advisory board honoraria from Nutricia and GSK; has received funding for travel from Athena Diagnostics, UCB, and GSK; and receives/has received research support from the NHMRC, ARC, NIH, Health Research Council of New Zealand, March of Dimes, the Weizmann Institute, CURE, US Department of Defense, and the Perpetual Charitable Trustees. The other authors have indicated they have no relevant competing interests to disclose. Go to Neurology.org/NG for full disclosures.

Acknowledgment

The authors thank the patients and their families for their participation in this research. Amy Schneider assisted with data

collection. This study was supported by funding from the National Health and Medical Research Council and the Research Institute of the McGill University Health Centre.

Study Funding

NHMRC (1091593), Research Institute of the McGill University Health Centre.

Publication History

Received by *Neurology: Genetics* October 22, 2020. Accepted in final form February 11, 2021.

Appendix Authors

Name	Location	Contribution
Kenneth A. Myers, MD, PhD	McGill University, Montreal, Canada	Design and conceptualized study, analyzed the data, prepared figures, and drafted this article for intellectual content

Continued

Appendix (continued)

Name	Location	Contribution
Carla Marini, MD, PhD	Salesi Pediatric Hospital, Ancona, Italy	Major role in the acquisition of data
Gemma L. Carvill, PhD	University of Washington, Seattle	Major role in the acquisition of data, reviewed and revised this article
Amy McTague, PhD	Great Ormond Street Hospital for Children, London, UK	Major role in the acquisition of data
Julie Panetta, MBBS	Neurology Network Melbourne, Melbourne, Australia	Major role in the acquisition of data
Chloe Stutterd, MBBS	Murdoch Children's Research Institute, Parkville, Australia	Major role in the acquisition of data
Thorsten Stanley, MBChB	University of Otago, Wellington, New Zealand	Major role in the acquisition of data, reviewed and revised this article
Samantha Marin, MD	University of Manitoba, Winnipeg, Manitoba, Canada	Major role in the acquisition of data, reviewed and revised this article
John Nguyen, BSc	University of Washington, Seattle	Major role in the acquisition of data
Carmen Barba, MD, PhD	Meyer Children's Hospital, Florence, Italy	Major role in the acquisition of data

References

1. Talkowski ME, Mullegama SV, Rosenfeld JA, et al. Assessment of 2q23.1 microdeletion syndrome implicates MBD5 as a single causal locus of intellectual disability, epilepsy, and autism spectrum disorder. *Am J Hum Genet* 2011;89:551–563.
2. Kleefstra T, Kramer JM, Neveling K, et al. Disruption of an EHMT1-associated chromatin-modification module causes intellectual disability. *Am J Hum Genet* 2012;91:73–82.
3. Hodge JC, Mitchell E, Pillalamarri V, et al. Disruption of MBD5 contributes to a spectrum of psychopathology and neurodevelopmental abnormalities. *Mol Psychiatry* 2014;19:368–379.
4. van Bon BW, Koolen DA, Brueton L, et al. The 2q23.1 microdeletion syndrome: clinical and behavioural phenotype. *Eur J Hum Genet* 2010;18:163–170.
5. Wagenstaller J, Spranger S, Lorenz-Depiereux B, et al. Copy-number variations measured by single-nucleotide-polymorphism oligonucleotide arrays in patients with mental retardation. *Am J Hum Genet* 2007;81:768–779.
6. Tados S, Wang R, Waters JJ, et al. Inherited 2q23.1 microdeletions involving the MBD5 locus. *Mol Genet Genomic Med* 2017;5:608–613.
7. Mullegama SV, Mendoza-Londono R, Elsea SH. MBD5 haploinsufficiency. In: Adam MP, Ardinger HH, Pagon RA, et al, editors. *GeneReviews*[®], Seattle: University of Washington, 1993. Available at <https://www.ncbi.nlm.nih.gov/books/NBK1116/>.
8. Consortium EK. Epi4K: gene discovery in 4,000 genomes. *Epilepsia* 2012;53:1457–1467.

Appendix (continued)

Name	Location	Contribution
Anna Rosati, MD	Meyer Children's Hospital, Florence, Italy	Major role in the acquisition of data
Richard H. Scott, MD	Great Ormond Street Hospital for Children, London, UK	Major role in the acquisition of data
Heather C. Mefford, MD, PhD	University of Washington, Seattle	Major role in the acquisition of data, reviewed and revised this article
Renzo Guerrini, MD, FRCP	Meyer Children's Hospital, Florence, Italy	Major role in the acquisition of data
Ingrid E. Scheffer, MBBS, PhD	University of Melbourne, Melbourne, Australia	Design, conceptualization, and supervision of study; reviewed and revised the manuscript for intellectual content

9. Scheffer IE, Berkovic S, Capovilla G, et al. ILAE classification of the epilepsies: position paper of the ILAE commission for classification and terminology. *Epilepsia* 2017;58:512–521.
10. Fisher RS, Cross JH, French JA, et al. Operational classification of seizure types by the international League against epilepsy: position paper of the ILAE commission for classification and terminology. *Epilepsia* 2017;58:522–530.
11. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of medical genetics and Genomics and the association for molecular pathology. *Genet Med* 2015;17:405–424.
12. Riggs ER, Andersen EF, Cherry AM, et al. Technical standards for the interpretation and reporting of constitutional copy-number variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics (ACMG) and the Clinical Genome Resource (ClinGen). *Genet Med* 2020;22:245–257.
13. Carvill GL, Heavin SB, Yendle SC, et al. Targeted resequencing in epileptic encephalopathies identifies de novo mutations in CHD2 and SYNGAP1. *Nat Genet* 2013;45:825–830.
14. Myers CT, Hollingsworth G, Muir AM, et al. Parental mosaicism in "de novo" epileptic encephalopathies. *N Engl J Med* 2018;378:1646–1648.
15. Bravo-Oro A, Lurie IW, Elizondo-Cardenas G, et al. A novel interstitial deletion of 2q22.3 q23.3 in a patient with dysmorphic features, epilepsy, aganglionosis, pure red cell aplasia, and skeletal malformations. *Am J Med Genet A* 2015;167A:1865–1871.
16. Shichiji M, Ito Y, Shimojima K, et al. A cryptic microdeletion including MBD5 occurring within the breakpoint of a reciprocal translocation between chromosomes 2 and 5 in a patient with developmental delay and obesity. *Am J Med Genet A* 2013;161A:850–855.
17. Dravet C. The core Dravet syndrome phenotype. *Epilepsia* 2011;52(Suppl 2):3–9.
18. Motobayashi M, Nishimura-Tadaki A, Inaba Y, et al. Neurodevelopmental features in 2q23.1 microdeletion syndrome: report of a new patient with intractable seizures and review of literature. *Am J Med Genet A* 2012;158A:861–868.
19. Scheffer IE, Berkovic SF. Generalized epilepsy with febrile seizures plus. A genetic disorder with heterogeneous clinical phenotypes. *Brain* 1997;120:479–490.
20. Du X, An Y, Yu L, et al. A genomic copy number variant analysis implicates the MBD5 and HNRNPU genes in Chinese children with infantile spasms and expands the clinical spectrum of 2q23.1 deletion. *BMC Med Genet* 2014;15:62.
21. Jaillard S, Dubourg C, Gerard-Blanluet M, et al. 2q23.1 microdeletion identified by array comparative genomic hybridisation: an emerging phenotype with Angelman-like features?. *J Med Genet* 2009;46:847–855.