The effectiveness of locally injected platelet-rich plasma on orthodontic tooth movement acceleration: *A systematic review of animal studies*

Ziyu Li^a; Jialiang Zhou^a; Song Chen^b

ABSTRACT

Objectives: To critically appraise the current evidence available from animal studies pertaining to the effectiveness of platelet-rich plasma (PRP) in accelerating orthodontic tooth movement.

Materials and Methods: Electronic searches of nine databases were conducted up to June 2020, followed by a hand search of the reference list of eligible studies. The study design required was prospective controlled animal studies. The primary outcome assessed was the rate of orthodontic tooth movement. The secondary outcome evaluated was histological changes after PRP application. Following study retrieval and selection, relevant data were extracted. Risk-of-bias (RoB) assessment was done using the Systematic Review Center for Laboratory Animal Experimentation's Risk of Bias Tool (SYRCLE's Risk of Bias Tool). Two review authors conducted the work of searching, study selection, and quality assessment independently and in duplicate.

Results: Of 193 studies, 5 animal studies were included in this systematic review. Three studies found a positive correlation between PRP injection and tooth movement acceleration, along with corresponding histological changes. Two studies detected no significant difference in tooth movement rate after PRP application.

Conclusions: Based on the current limited evidence, the efficacy of PRP on tooth movement acceleration remains debatable. More well-designed randomized controlled trials involving humans are called for to obtain more clinically significant conclusions. (*Angle Orthod.* 2021;91:391–398.)

KEY WORDS: Orthodontics; Tooth movement techniques; Platelet-rich plasma; Acceleration

INTRODUCTION

Orthodontic treatment often requires 2 years and even longer for extraction cases and other complicated cases.¹ Side effects including caries, periodontal

Corresponding author: Dr Song Chen, State Key Laboratory of Oral Diseases, Department of Orthodontics and Pediatric Dentistry, West China Hospital of Stomatology, Sichuan University, Section 3, No. 14, Renmin South Road, Chengdu, Sichuan 610041, China

(e-mail: songchen882002@hotmail.com)

Accepted: November 2020. Submitted: June 2020.

 $\ensuremath{\textcircled{\sc 0}}$ 2021 by The EH Angle Education and Research Foundation, Inc.

disease, and root resorption are more common in prolonged treatment. Therefore, accelerating tooth movement and shortening the treatment time could be of great importance.

Many techniques have been developed to reduce the duration of treatment. Surgical-assisted approaches such as micro-osteoperforation and piezocision have been proven to be clinically effective.^{2,3} However, they require surgical injury to bone and tissue.⁴ Research concerning the effects of biological agents, including prostaglandin, parathyroid hormone, and vitamin D3, on tooth movement acceleration has shown conflicting results.⁵ However, the use of hormones or other allogenic products can result in irreversible systemic effects.⁶

Platelet-rich plasma (PRP), an autologous concentration of platelets in a minute volume of plasma, is a rich source of autologous growth factors and cytokines.⁷ Since these growth factors and cytokines can stimulate both osteoblastic and osteoclastic activity, PRP can thus interfere with the alveolar boneremodeling process.^{8,9} The successful application of

^a Postgraduate student, State Key Laboratory of Oral Diseases, Department of Orthodontics and Pediatric Dentistry, West China Hospital of Stomatology, Sichuan University, Chengdu, Sichuan, China.

^b Professor, State Key Laboratory of Oral Diseases, Department of Orthodontics and Pediatric Dentistry, West China Hospital of Stomatology, Sichuan University, Chengdu, Sichuan, China.

Published Online: February 10, 2021

PRP has been reported in oral surgery and regenerative dentistry.^{10,11} These characteristics support the potential for PRP to accelerate orthodontic tooth movement, which is caused by the gradual modeling and remodeling of the surrounding alveolar bone. Liou¹² reported that PRP achieved an accelerating effect in different types of tooth movements clinically. Recently, more studies have been conducted to confirm the effectiveness of PRP on orthodontic tooth movement acceleration, mostly in animal models. However, no systematic review has been performed to summarize and critically analyze the studies conducted. This systematic review aimed to evaluate the available knowledge, comprehensively, from animal studies regarding the influence of PRP injection on the rate of orthodontic tooth movement.

MATERIALS AND METHODS

Protocol and Registration

This review was registered in the PROSPERO database under protocol CRD42020188837, in accordance with the PRISMA checklist of systematic reviews and meta-analyses.¹³ This review was conducted in accordance with the Cochrane Handbook for Systematic Review of Interventions and Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA).^{14,15}

Eligibility Criteria

The PICOS (population, intervention, comparison, outcome, study design) format was used to formulate the clinical question with defined inclusion and exclusion criteria (Table 1). Experimental prospective controlled animal studies involving healthy animals under active orthodontic treatment were included. The primary outcome assessed was the rate of orthodontic tooth movement, and the secondary outcome was histological changes after PRP application.

Information Source and Search Strategy

PubMed, Cochrane Central Register of Controlled Trials (CENTRAL), Embase, Web of Science, China National Knowledge Infrastructure (CNKI), China Biology Medicine disc (CBM), System for Information on Grey Literature in Europe (SIGLE), Clinical Trial.gov, and ProQuest Dissertations & Theses were searched without language or time restrictions until June 2020. The details of the database search are summarized in Table 2. The reference lists of the eligible studies were also checked for additional relevant studies. This process was conducted independently and in duplicate.

Study Selection

The study selection process was done by two reviewers independently and in duplicate. First, the titles and abstracts of articles were assessed for eligibility. Then, full-text reports were considered for articles that seemed to have met the inclusion criteria.

Data Collection and Data Items

Relevant data from the included studies were extracted by two reviewers in previously developed and piloted forms: author and year, study design, subject characteristics, orthodontic procedures, the injections applied in both the intervention group and control group, intervals of outcome assessment, and the details of PRP application.

Risk of Bias in Individual Studies

The risk of bias (RoB) of the included studies was assessed using the Systematic Review Center for Laboratory Animal Experimentation's Risk of Bias tool (SYRCLE's Risk of Bias Tool).¹⁶ The summary of RoB within a study was assessed according to Higgins and Green.¹⁷ This process was done independently and in duplicate by two authors.

Summary Measures and Synthesis of Results

Random effects meta-analysis was planned to determine pooled estimates and relative effects of PRP on the rate of tooth movement. However, the differences in the study design and interventions employed precluded quantitative data synthesis.

RoB Across Studies and Additional Analyses

Analyses for small-study effects, publication bias, and exploratory subgroup analyses were planned if an adequate number of studies were identified. The quality of evidence from the retrieved studies was assessed using the Grading of Recommendation Assessment, Development, and Evaluation (GRADE) approach.¹⁸

RESULTS

Study Selection

A total of 192 studies were initially identified through database searching, and 1 study was obtained by manual search. After removing duplicates, 146 studies remained. After reading the titles and abstracts, 12 studies remained for further assessment. Eventually, 5 studies were included in the qualitative analysis.^{19–23} Procedures of the electronic search are presented in Figure 1.

| Table 1. Eligibility Criteria for the Present Systematic Re | eview |
|---|-------|
|---|-------|

| Domain | Inclusion Criteria | Exclusion Criteria |
|--------------|---|---|
| Participants | Healthy animal subjects undergoing active orthodontic tooth movement | In vitro or ex vitro studies, in silico study, human studies |
| Intervention | Local injection of platelet-rich plasma, with or without activation agents | Simultaneous injection of agents other than platelet-rich plasma and activation agents |
| Comparison | Placebo intervention or no intervention | |
| Outcome | Primary outcome: quantitative data regarding the rate of orthodontic tooth movement (ie, the amount of tooth | Qualitative assessments regarding the rate of orthodontic tooth movement |
| | movement in a specific period of time) measured by various ways | Inadequate definition of outcomes |
| | Secondary outcome: histological changes | |
| Study design | Experimental prospective controlled animal study | Noncomparative studies Reviews, systematic reviews and meta-analyses, opinions and editorials |

Study Characteristics

The characteristics of each included study are presented in Table 3. Among the five included studies, three studies used a split-mouth design.^{19,20,23} Apart from the comparison of a PRP injection group with a noninjection group, Akbulut et al.22 also compared a PRP group with a platelet-poor plasma (PPP) group, and Gülec et al.¹⁹ compared a high-platelet-concentration injection group with a moderate-platelet-concentration injection group. Orthodontic tooth movement was induced by the placement of coil springs, except for one study,²¹ in which tooth movement was initiated by placing a rubber separator between incisors. The concentrations of platelets in PRP in the included studies ranged from 2.45 to 6.6 times that of the whole blood. The details of the PRP application are presented in Table 4.

RoB Within Studies

Two studies^{19,21} were determined to be at high RoB, and three studies^{20,22,23} were of unclear RoB. The methods of sequence generation, allocation concealment, blinding of caregivers and/or investigators during the intervention, and random outcome assessment were generally inadequately reported. Dropouts in one study were clearly outlined.²¹ Finally, there was no sufficient information to determine categorically the presence of any additional problems that could possibly increase the RoB. The RoB assessment of the included studies is presented in Table 5.

Effect of PRP on the Rate of Orthodontic Tooth Movement

Three studies supported the effectiveness of PRP injection in accelerating the rate of orthodontic tooth movement. Güleç et al.19 reported that, on day 21, the amount of tooth movement induced by high concentrations of PRP was 1.7 times more than that on the control side and 1.4 times more than that induced by moderate concentrations of PRP. Rashid et al.²⁰ also reported a higher percentage of mean changes in tooth movement than the control group, with a percentage change ratio of 2.13:1. Nakornnoi et al.23 found greater cumulative tooth movement in the L-PRP group than in the control group at all observation times, and a significantly greater rate of tooth movement was observed in the L-PRP group in the first 14 days (0-7 days: control side: 0.94 \pm 0.09 mm, intervention side: 1.04 \pm 0.05 mm; 7–14 days: control side: 0.45 \pm 0.12 mm, intervention side: 0.58 ± 0.09 mm, P < .05).

However, the other two studies found that PRP injection was not efficient in tooth movement acceleration. Akbulut et al.²² discovered that the amount of molar mesialization was less in the PRP group than that in the control group on day 3 (PRP group: 0.287 \pm 0.176 mm, PPP group: 0.482 \pm 0.128 mm, control

 Table 2.
 Search Strategy for Each Database

| Database | Search Strategy |
|--|---|
| PubMed | (orthodontics[Mesh] OR orthodontic*) AND (Tooth movement techniques[Mesh] OR tooth movement OR movement OR move OR moving OR retract*) AND (platelet-rich plasma [Mesh] OR platelet concentrate OR platelet gel OR PRP OR platelet) |
| Embase, Web of Science, CENTRAL | (orthodontics OR orthodontic*) AND (Tooth movement techniques OR tooth movement OR movement OR move OR moving OR retract*) AND (platelet-rich plasma OR platelet concentrate OR platelet gel OR PRP OR platelet) |
| Proquest CNKI, CBM, Clinical Trial.gov, SIGLE | orthodontic*, tooth movement, platelet-rich plasma tooth movement, platelet-rich plasma |

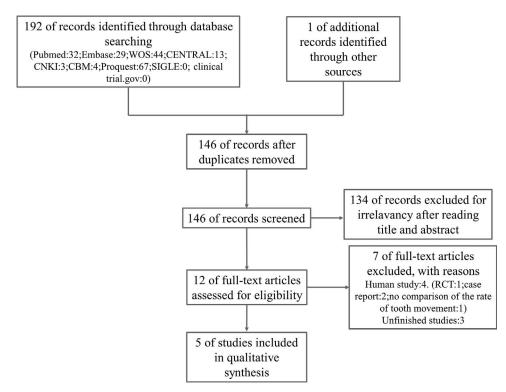


Figure 1. Systematic search and selection strategy.

group: 0.625 \pm 0.028 mm, P = .01), and no significant difference was observed among the groups on days 1, 7, or 14. Sufarnap et al.²¹ found that tooth movement was similar with or without PRP injection on days 6, 9, 12, and 24 after rubber separator placement.

Histological Analysis

Histologic analysis was performed in four studies. Two studies^{19,22} reported similar results between the PRP and non-PRP group. One study found that the number of osteoclasts and osteoblasts and the results of the immunohistochemistry evaluations with or without PRP injection were similar from day 1 to day 14.22 Surprisingly, despite an increased rate of tooth movement in the PRP groups, Güleç et al.¹⁹ reported that the osteoclast counts in both high concentrations of PRP and medium concentrations of PRP were greater than the controls (12.01 \pm 0.56 cells/mm² and 14.05 ± 0.46 cells/mm², respectively) only on day 3 and less than controls at all other observational days. In contrast, Rashid et al.20 and Nakornnoi et al.23 observed higher osteoclastic activity following PRP/ L-PRP injection. Rashid et al.²⁰ found significantly higher numbers of osteoblasts, osteoclasts, and cementoblasts in the PRP group as compared with the control group (osteoblasts: 16.2 \pm 1.30 vs 12.0 \pm 0.12 cells/10 μ m², *P* = .017; osteoclasts: 7.2 ± 1.30 vs 2.8 ± 0.84 cells/10 μ m², *P* = .006; cementoblasts: 21.8 \pm 1.30 vs 14.4 \pm 1.82 cells/10 μ m², P = .004). Nakornnoi et al.²³ found a much greater number of osteoclasts in the L-PRP group than in the control group on day 7 (10.6 \pm 2.07 vs 7.4 \pm 2.30 cells) and day 14 (16.2 \pm 3.03 vs 11.6 \pm 3.04 cells), but there was no significant difference on day 28 (4.2 \pm 1.78 vs 3.8 \pm 1.48 cells).

RoB Across Studies and Additional Analysis

It was not possible to conduct analyses for smallstudy effects, publication bias, or subgroup analyses. The quality of evidence of the rate of orthodontic tooth movement assessed in the systematic review was very low. The RoB, inconsistency, indirectness, and imprecision were rated as serious. The reasons were that two studies were of high RoB, inconsistent results were presented in the included studies, the results could not be directly applied to human clinical settings, and the number of subjects included was limited. Publication bias was unlikely in the retrieved studies.

DISCUSSION

Recently, there has been an increased interest in research focused on tooth movement acceleration without the use of invasive surgical procedures. PRP is considered a potential tooth movement acceleration agent. According to the results of this review, three studies showed a positive correlation between local injection of PRP/L-PRP and acceleration of orthodontic

Table 3. Characteristics of Included Studies^a

| Study ID | Study Design | Animals (Age/ Weight) | Orthodontic Procedure | Study Groups | Measurement Time | Outcome | |
|--|------------------------------------|---|--|---|---------------------------|--|--|
| Rashid et al. Split-mouth 2017 design | | blit-mouth Six male mongrel Maxillary sec | | Group A: test group (25 units of PRP with 25 units of thrombin- CaCl ₂ solution) Group B: control group (50 units of thrombin- CaCl ₂) | 9 wk | Distance of tooth movement; histological findings | |
| Akbulut et al. 2019 | Three arm | 54 Wistar male albino rats/6–8 wk, 150–200 g | Closed-coil springs were ligated to the maxillary incisors and first molars with stainless steel ligature wires Force: 50 g | Group A: PRP group (0.1 mL of PRP) Group B: PPP group (0.1 mL of PPP) Group C: control group (no injection) | 1, 3, 7, and 14 d | Distance of tooth movement; osteoblast and osteoclast cell counts, and ALF TRAP, and TGF β expressions | |
| Güleç et al. 2017 | Two-arm, split- mouth design | 81 young adult Sprague-Dawley rats/9–10 wk, 348 ± 29 g | Superelastic closed- coil springs were placed between the maxillary molars and incisors Force: 40 cN | Group A: hPRP group (0.01 mL) Group B: mPRP group (0.01 mL) Group C: hPRP control group (no injection) Group D: mPRP control group (no injection) | 3, 7, 14, 21, and 60 d | Distance of tooth movement; bone content of interradicular space; alveolar bone volume; osteoclastic activity | |
| Nakornnoi et al. 2019 | Two-arm, split- mouth design | 23 male New Zealand white rabbits/3–4 mo, 2.5–3 kg | Light-type nickel titanium closed- coil spring was placed between the maxillary first premolar and incisor on each side Force: 100 g | Group A: test group (0.5 mL L-PRP) Group B: control group (normal saline solution) Group C: blank control group (no injection) | 0, 3, 7, 14, 21, 28 d | Distance of tooth movement; osteoclast numbers; histological findings | |
| Sufarnap et al. 2017 | Two arm | 24 young Guinea pigs/2–3 mo, 250–400 g | Power-O (Ormco) as a rubber separator between central incisors after PRP injection | Group A: test group (0.5 mL PRP) Group B: control group (no injection) | 6, 9, 12, 24 d | Distance of tooth movement | |

^a ALP indicates alkaline phosphatase; hPRP, high platelet concentration injection group; L-PRP, leukocyte-platelet-rich plasma; mPRP, moderate platelet concentration injection group; PPP, platelet-poor plasma; PRP, indicates platelet-rich plasma; NiTi, nickel titanium; TAD, temporary anchorage device; TGF-β, transforming growth factor–β; TRAP, tartrate-resistant acid phosphatase.

tooth movement.^{19,20,23} Their results were in agreement with those of El-Timamy et al.,²⁴ who showed PRP to be clinically feasible and effective in accelerating the rate of tooth movement in early stages of tooth movement.²⁵ Two studies reported no beneficial effects of PRP injection on the rate of tooth movement.^{21,22}

Histological analysis is beneficial for observing the effect of PRP on osteoblast and osteoclast activity, which regulates bone regeneration and thus regulates orthodontic tooth movement. Greater numbers of osteoclasts and osteoblasts were reported in two studies,^{20,23} whereas one study²² reported no significant difference in the cell counts at the early stage (14 days). Güleç et al.¹⁹ found that the osteoclast counts in the PRP injection groups were less than those of the control group at all observation times except day 3, despite recording an increased rate of tooth movement. Previous studies have shown conflicting results

Table 4. Details of the Application of PRP^a

| Study ID | Source of PRP | PRP Concentration | Injection Method | Injection Sites | Dose | Use of Activation Agent | Injection Interval |
|--------------------------|------------------|--|------------------------------|--|--------------|-------------------------------|---|
| Rashid et al. 2017 | Autologous | Unclear | Intraligamental injection | Middle of the distal side, distobuccal side, distopalatal side followed by buccal and palatal injections at the first premolar | 25 units | CaCl₂ and thrombin | 0, 21, and 42 d |
| Akbulut et al. 2019 | Donor | PRP: 3617 × 10³/μL PPP: 23 × 10³/ μL | Submucosal injection | Buccal vestibular mucosa next to the distal root of the maxillary right first molars | 0.1 mL PRP | Unclear | Injections were done only once at day 0 |
| Güleç et al. 2017 | Donor | hPRP: 2593.2 \pm 257 \times 10 ³ platelets/µL mPRP: 1220.4 \pm 154 \times 10 ³ platelets/µL | Submucosal injection | Buccal vestibular mucosa next to the mesial root of the right first molar | 0.01 mL PRP | Unclear | Injections were done only once at day 0 |
| Nakornnoi et al. 2019 | Autologous | PRP: platelet: 2314.44 ± 570.82 × 10 ³ cells/µL Leukocytes: 6.87 ± 2.29 × 10 ³ cells/µL | Submucosal injection | Buccal and lingual areas of the first maxillary premolar | 0.5 mL L-PRP | Unclear | Injections were done only once at day 0 |
| Sufarnap et al. 2017 | Donor | PRP: 507 × 10 ³ / μL | Unclear | Unclear | 0.5 mL PRP | Unclear | PRP was injected before rubber separator placement |

^a hPRP indicates high platelet concentration injection group; L-PRP, leukocyte-platelet-rich plasma; mPRP, moderate platelet concentration injection group; PPP, platelet-poor plasma; PRP, platelet-rich plasma.

in terms of bone remodeling. Most studies favor the idea that PRP can stimulate both osteoblast and osteoclast precursor cells to divide and differentiate and promote bone regeneration,^{26–28} while others found that PRP did not improve bone regeneration.^{29,30} Future studies are needed to confirm the effect of PRP on bone remodeling.

Despite receiving substantial attention, there is still a lack of standardization in PRP treatment protocols. According to Marx,⁷ autologous blood was critical for achieving effective outcomes with the use of PRP, whereas the use of donor animal blood platelets could cause an overt immune reaction and lead to false-negative results. In addition, differences in the spinning

| Table 5. | Summary of Risk-of-Bias | Assessment According to | the SYRCLE RoB Tool |
|----------|-------------------------|-------------------------|---------------------|
|----------|-------------------------|-------------------------|---------------------|

| | Signaling Questions ^a | | | | | | | | | | |
|-----------------------|----------------------------------|---|---|---|---|---|---|---|---|----|---------|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | Summary |
| Rashid et al. 2017 | ? | + | ? | + | + | + | + | ? | + | ? | ? |
| Akbulut et al. 2019 | ? | + | ? | + | ? | ? | ? | ? | + | ? | ? |
| Güleç et al. 2017 | - | ? | - | ? | ? | ? | + | + | + | ? | - |
| Nakornnoi et al. 2019 | ? | + | ? | ? | ? | ? | ? | + | + | ? | ? |
| Sufarnap et al. 2017 | ? | ? | ? | ? | ? | ? | ? | - | + | ? | • |

^a Signaling questions: 1: Was the allocation sequence adequately generated and applied? 2: Were the groups similar at baseline or were they adjusted for confounders in the analysis? 3: Was the allocation adequately concealed? 4: Were the animals randomly housed during the experiment? 5: Were the caregivers and investigators blinded to the intervention that each animal received? 6: Were animals selected at random for outcome assessment? 7: Was the outcome assessor blinded? 8: Were incomplete outcome data adequately addressed? 9: Are reports of the study free of selective outcome reporting? 10: Was the study apparently free of other problems that could result in high risk of bias?

•:low risk of bias; ?:unclear-:high risk of bias.

techniques can cause different concentrations of platelets, leukocytes, and growth factors in PRP.³¹ Therefore, the heterogeneity of this systematic review might be partially attributable to the differences in the methods for PRP production, as three of the animal studies^{19,21,22} applied homologous blood from donors and only one study²⁰ claimed to use the spinning technique as recommended by Marx.

There has also been no consensus on the optimal concentration of PRP. A minimum platelet concentration of 1 \times 10⁶ platelets/µL was defined by Marx.³² Sufarnap et al.²¹ used PRP at a concentration of 507 \times 10³ platelets/µL, less than the lower limit. In addition, some studies reported that high concentrations of PRP suppressed the viability and proliferation of alveolar bone cells and could be harmful for the healing process.^{33,34} Therefore, it is important to identify therapeutic concentration ranges for effective and harmless PRP application.

In the included studies, Nakornnoi et al.²³ used L-PRP instead of pure PRP, claiming that leukocytes could lead to an initial burst release of proinflammatory growth factors, initiating cellular and molecular events in the early phase after orthodontic force application.³⁵ However, whether to include leukocytes in the injected concentration has been a controversial topic. Yin et al.²⁵ reported that pure PRP was more effective than L-PRP in terms of osteogenic differentiation of bone marrow stem cells in rats. Xu et al.³⁶ also found that L-PRP could activate NF- κ B and result in adverse effects on cartilage regeneration. Additional clinical trials are needed to justify leukocyte inclusion.

This was the first systematic review to evaluate the effect of PRP on orthodontic tooth movement acceleration. However, the limitations cannot be ignored. The foremost limitation of this review was that all data were drawn from animal experiments and cannot be directly extrapolated to humans. The studies included were of an unclear or even high RoB, and the overall quality was very low. In addition, quantitative analysis was not performed because of the significant heterogeneity in the designs of the included studies. Therefore, more high-quality studies with stricter designs are needed to draw more clinically significant conclusions.

CONCLUSIONS

- Based on the current evidence, the effectiveness of PRP on orthodontic tooth movement acceleration cannot be confirmed.
- It seems that the effect of PRP on the rate of tooth movement is affected by the standards of PRP

acquisition, PRP concentrations, and the methods of PRP application.

 More well-designed randomized controlled trials involving humans are needed to draw more clinically significant conclusions.

ACKNOWLEDGMENT

This systematic review was funded by the National Natural Science Foundation of China (81671021).

REFERENCES

- Fisher MA, Wenger RM, Hans MG. Pretreatment characteristics associated with orthodontic treatment duration. *Am J Orthod Dentofacial Orthop.* 2010;137:178–186.
- Alikhani M, Raptis M, Zoldan B, et al. Effect of microosteoperforations on the rate of tooth movement. *Am J Orthod Dentofacial Orthop.* 2013;144:639–648.
- Yi J, Xiao J, Li Y, Li X, Zhao Z. Efficacy of piezocision on accelerating orthodontic tooth movement: a systematic review. *Angle Orthod*. 2017;87:491–498.
- Chang J, Chen PJ, Dutra EH, Nanda R, Yadav S. The effect of the extent of surgical insult on orthodontic tooth movement. *Eur J Orthod*. 2019;41:601–608.
- Nimeri G, Kau CH, Abou-Kheir NS, Corona R. Acceleration of tooth movement during orthodontic treatment—a frontier in orthodontics. *Prog Orthod*. 2013;14:42.
- Mangal U. Influence of platelet rich plasma on orthodontic tooth movement: a review. *Biomed Pharmacol J.* 2017;10: 1463–1468.
- Marx RE. Platelet-rich plasma: evidence to support its use. J Oral Maxillofac Surg. 2004;62:489–496.
- Suaid FF, Carvalho MD, Santamaria MP, et al. Platelet-rich plasma and connective tissue grafts in the treatment of gingival recessions: a histometric study in dogs. *J Periodont*. 2008;79:888–895.
- 9. Andrade I, Taddei SRA, Souza PEA. Inflammation and tooth movement: the role of cytokines, chemokines, and growth factors. *Semin Orthod*. 2012;18:257–269.
- 10. Xu J, Gou L, Zhang P, Li H, Qiu S. Platelet-rich plasma and regenerative dentistry. *Aust Dent J.* 2020;65:131–142.
- 11. Franchini M, Cruciani M, Mengoli C, et al. The use of platelet-rich plasma in oral surgery: a systematic review and meta-analysis. *Blood Transf.* 2019;17:357–367.
- 12. Liou EJW. The development of submucosal injection of platelet rich plasma for accelerating orthodontic tooth movement and preserving pressure side alveolar bone. *APOS Trends Orthod.* 2016;6:5–11.
- Shamseer L. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation (vol 350, g7647, 2015). *BMJ*. 2016;354:1.
- 14. Liberati A, Altman DG, Tetzlaff J, et al. The PRISMA statement for reporting systematic reviews and metaanalyses of studies that evaluate health care interventions: explanation and elaboration. *J Clin Epidemiol.* 2009;62:e1– 34.
- Higgins JPT, Thomas J, Chandler J, Cumpston M, Welch VA. Cochrane Handbook for Systematic Reviews of Interventions. Version 6.0 (updated July 2019). London: Cochrane Collaboration; 2019. Available at: www.training. cochrane.org/handbook.

- Hooijmans CR, Rovers MM, de Vries RBM, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCL's risk of bias tool for animal studies. *BMC Med Res Methodol*. 2014;14:9.
- 17. Higgins JPT, Altman DG, Gotzsche PC, et al. The Cochrane Collaboration's tool for assessing risk of bias in randomised trials. *BMJ*. 2011;343:9.
- Guyatt GH, Oxman AD, Schunemann HJ, Tugwell P, Knottnerus A. GRADE guidelines: a new series of articles in the *Journal of Clinical Epidemiology*. J Clin Epidemiol. 2011;64:380–382.
- Güleç A, Bakkalbaşı B, Cumbul A, Uslu Ü, Alev B, Yarat A. Effects of local platelet-rich plasma injection on the rate of orthodontic tooth movement in a rat model: a histomorphometric study. *Am J Orthod Dentofacial Orthop.* 2017;151:92– 104.
- 20. Rashid A, ElSharaby FA, Nassef EM, Mehanni S, Mostafa YA. Effect of platelet-rich plasma on orthodontic tooth movement in dogs. *Orthod Craniofac Res.* 2017;20:102–110.
- Sufamap E, Sofyanti E, Ilyas S. The effect of platelet-rich plasma to orthodontic tooth movement. In: Abbot PV, Tseng PSK, Porto I, Lestari W, eds. *Proceedings of the International Dental Conference of Sumatera Utara 2017*. Vol 8. Paris, France: Atlantis Press; 2017;80–83.
- 22. Akbulut S, Yagci A, Yay AH, Yalcin B. Experimental investigation of effects of platelet-rich plasma on early phases of orthodontic tooth movement. *Am J Orthod Dentofacial Orthop.* 2019;155:71–79.
- 23. Nakornnoi T, Leethanakul C, Samruajbenjakun B. The influence of leukocyte-platelet-rich plasma on accelerated orthodontic tooth movement in rabbits. *Korean J Orthod.* 2019;49:372–380.
- 24. El-Timamy A, El Sharaby F, Eid F, El Dakroury A, Mostafa Y, Shakr O. Effect of platelet-rich plasma on the rate of orthodontic tooth movement: a split-mouth randomized trial. *Angle Orthod.* In press.
- Yin W, Qi X, Zhang Y, et al. Advantages of pure platelet-rich plasma compared with leukocyte- and platelet-rich plasma in promoting repair of bone defects. *J Transl Med.* 2016;14:73.
- 26. Gerard D, Carlson ER, Gotcher JE, Jacobs M. Effects of platelet-rich plasma at the cellular level on healing of

autologous bone-grafted mandibular defects in dogs. *J Oral Maxillofac Surg.* 2007;65:721–727.

- 27. Thorwarth M, Wehrhan F, Schultze-Mosgau S, Wiltfang J, Schlegel KA. PRP modulates expression of bone matrix proteins in vivo without long-term effects on bone formation. *Bone.* 2006;38:30–40.
- Otero L, Carrillo N, Calvo-Guirado JL, Villamil J, Delgado-Ruiz RA. Osteogenic potential of platelet-rich plasma in dental stem-cell cultures. *Br J Oral Maxillofac Surg.* 2017;55: 697–702.
- 29. Casati MZ, Gurgel BCD, Goncalves PF, et al. Platelet-rich plasma does not improve bone regeneration around periimplant bone defects: a pilot study in dogs. *Int J Oral Maxillofacial Surg.* 2007;36:132–136.
- Kilic SC, Gungormus M, Parlak SN. Histologic and histomorphometric assessment of sinus-floor augmentation with beta-tricalcium phosphate alone or in combination with pureplatelet-rich plasma or platelet-rich fibrin: a randomized clinical trial. *Clin Implant Dent Relat Res.* 2017;19:959–967.
- Oudelaar BW, Peerbooms JC, in't Veld RH, Vochteloo AJH. Concentrations of blood components in commercial plateletrich plasma separation systems: a review of the literature. *Am J Sports Med.* 2019;47:479–487.
- 32. Marx RE. Platelet-rich plasma (PRP): what is PRP and what is not PRP? *Implant Dent.* 2001;10:225–228.
- Yamaguchi R, Terashima H, Yoneyama S, Tadano S, Ohkohchi N. Effects of platelet-rich plasma on intestinal anastomotic healing in rats: PRP concentration is a key factor. *J Surg Res.* 2012;173:258–266.
- 34. Choi BH, Zhu SJ, Kim BY, Huh JY, Lee SH, Jung JH. Effect of platelet-rich plasma (PRIP) concentration on the viability and proliferation of alveolar bone cells: an in vitro study. *Int J Oral Maxillofacial Surg.* 2005;34:420–424.
- Alhashimi N, Frithiof L, Brudvik P, Bakhiet M. Orthodontic tooth movement and de novo synthesis of proinflammatory cytokines. *Am J Orthod Dentofacial Orthop*. 2001;119:307– 312.
- Xu ZL, Yin WJ, Zhang YL, et al. Comparative evaluation of leukocyte- and platelet-rich plasma and pure platelet-rich plasma for cartilage regeneration. *Sci Rep.* 2017;7:14.