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# Structure-activity relationships of noncovalent immunoproteasome β5i-selective dipeptides

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# Abstract

The immunoproteasome (i-20S) has emerged as a therapeutic target for autoimmune and inflammatory disorders and hematological malignancies. Inhibition of the chymotryptic  $\beta$ 5i subunit of i-20S inhibits T cell activation, B cell proliferation and dendritic cell differentiation in vitro and suppresses immune responses in animal models of autoimmune disorders and allograft rejection. However, cytotoxicity to immune cells has accompanied the use of covalently reactive  $\beta$ 5i inhibitors, whose activity against the constitutive proteasome (c-20S) is cumulative with time of exposure. Herein we report a structure-activity relationship study of a class of noncovalent proteasome inhibitors with picomolar potencies and thousands-fold selectivity for i-20S over c-20S. Furthermore, these inhibitors are specific for  $\beta$ 5i over the other 5 active subunits of i-20S and c-20S, providing useful tools to study functions of  $\beta$ 5i in immune responses. The potency of these compounds in inhibiting human T cell activation suggests that they may have therapeutic potential.

# **Graphical Abstract**

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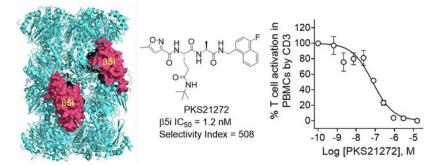
Author Contributions

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Synthesis and compound characterization of all new compounds and Table S1.

The authors declare the following competing financial interest(s): Cornell University has obtained a United States patent on these immunoproteasome inhibitors, patent number 9,988,421, as well as corresponding foreign patents. G. Lin, C. Nathan and X. Ma are listed as inventors on these patents.



#### Keywords

Immunoproteasome;  $\beta$ 5i inhibitor; noncovalent; isoform selectivity; T cell activation; immune response

# INTRODUCTION

Since the approval of Bortezomib for treatment of once largely untreatable multiple myeloma, components of the ubiquitin proteasome system (UPS) of protein degradation have become attractive drug targets for various malignancies and autoimmune disorders. The UPS degrades cellular proteins via a highly regulated process by which protein substrates are polyubiquitinated through a cascade of reactions catalyzed by a ubiquitin activating enzyme (E1), a ubiquitin conjugating enzyme (E2) and a ubiquitin ligase (E3). Polyubiquitinated proteins are recognized, unfolded and translocated by the proteasome regulatory particle (19S) into the 20S core particle for degradation by three proteolytic  $\beta$ subunits in the 20S core:  $\beta$ 1,  $\beta$ 2 and  $\beta$ 5.<sup>1, 2</sup> Through this ordered degradation process, the UPS regulates numerous cellular activities ranging from removal of unfolded or damaged proteins to cell cycle progression, transcription factor activation and immune surveillance by providing antigenic peptides for presentation by MHC class I molecules.<sup>2, 34</sup> Panproteasome inhibitors limit the supply of peptides for MHC class I molecules and thus block antigen presentation.<sup>5</sup> Bortezomib is highly toxic to plasma cells<sup>6</sup> and has been used to prevent antibody mediated graft rejection of transplanted kidneys, although the effect is modest at the limited doses that are considered tolerable in patients who do not have an incurable malignancy.<sup>7, 8</sup> Species selective inhibitors of proteasomes of pathogenic microbes have been developed for treatment of infections by Mycobacterium tuberculosis,9-12 *Plasmodia*,<sup>13–16</sup> *Leishmania* and *Trypanosoma*,<sup>17, 18</sup> suggesting further potential therapeutic utility of proteasome inhibitors.

To harness the benefit of proteasome inhibitors in the therapy of autoimmune disorders without the toxicity of pan-proteasome inhibition, it is necessary to develop proteasome inhibitors that selectively target one or more of the proteasome subunits that predominate in the immune system rather than those expressed in non-immune cells. There are two major isoforms of proteasome in humans that differ in each of the three proteolytically active subunits: constitutive proteasome (c-20S) and immunoproteasome (i-20S). Recent proteomic studies have found that i-20S predominates in the cells of the human innate and adaptive

immune systems.<sup>19</sup> The i-20S is also upregulated in other cells at sites of inflammation and upon exposure to interferon- $\gamma$ .

Selective inhibitors of i-20S  $\beta$ 5i over c-20S  $\beta$ 5c have been reported, such as the peptide epoxyketone ONX0914, which produced similar immunoregulatory effects in mice as BTZ but with much reduced toxicity.<sup>20–26</sup> A recent study reported an improvement on ONX0914 with up to 600-fold selectivity.<sup>27</sup> Reports on ONX0914 were followed by reports of other selective i-20S inhibitors (Figure 1); due to space limitations, only reviews and the most recent primary publications are cited.<sup>27–33</sup> Most reported inhibitors are peptide epoxyketones or sulfonyl fluorides that act covalently and irreversibly with the active site. Irreversible covalent binding can result in unpredictable liver toxicity, even if off-target binding occurs at a low level,<sup>34</sup> and irreversible inhibitors often show diminished selectivity over time.<sup>35</sup> Compounds reported in patents also include amino boronates from Merck-Serono and substituted thiazoles and peptide  $\alpha$ -keto amides from Roche (Figure 1). These three classes of i-20S inhibitors are either covalent but reversible or noncovalent. They represent alternative directions for developing i-20S selective inhibitors.

Recently, our labs reported three new classes of immunoproteasome inhibitors (IPIs): 1,3,4oxathiazol-2-ones;<sup>36</sup>  $\beta$ -amino acid-based *N*,*C*-capped diepeptidomimetics;<sup>37</sup> and *N*,*C*capped dipeptides.<sup>35</sup> We also discovered asparagine-ethylenediamine IPIs (AsnEDAs) (Figure 1).<sup>38</sup> The latter three classes are noncovalent inhibitors and highly  $\beta$ 5i- selective. We have demonstrated that compounds from each class inhibit  $\beta$ 5i in cells and restrict immunological responses of plasmacytoid dendritic cells, B cells and T cells in vitro.<sup>35, 38</sup> We reported that DPLG3 prolonged the survival of MHC-mismatched mouse cardiac allografts in a heterotopic heart transplant model.<sup>35</sup> However, DPLG3 lacks drug-like properties. Herein we report our further structure-activity relationship studies of the *N*,*C*capped dipeptides.

# **RESULTS AND DISCUSSION**

#### **Design rationale**

DPLG3 was developed based on DPLG2, which was designed for species selective inhibition of the *Mycobacterium tuberculosis* proteasome (Mtb20S) over i-20S and c-20S.<sup>39</sup> We have shown that i-20S and Mtb20S share similar substrate preferences, especially at the S1 site (Figure 2), which prefers bulky hydrophobic moieties as visualized in the structures of Mtb20S and i-20S.<sup>9, 10, 12, 38, 39</sup> We postulated that by anchoring a bulky P1 (*N*-cap) in the *N*,*C*-capped dipeptide, we could explore the P2 (R<sub>2</sub>), P3 (R<sub>3</sub>) and *N*-cap positions to improve potency and in the meantime maintain selectivity and improve pharmacokinetic properties. DPLG3 is a highly selective immunoproteasome  $\beta$ 5i inhibitor with single-digit nanomolar potency, but it suffers from poor physicochemical properties and cell permeability.<sup>35</sup> We reported that replacing P2 *p*-F-phenylalanine of DPLG3 with O-methylserine (PKS2086) maintained potency, but the isoform selectivity ( $\beta$ 5c / $\beta$ 5i) over c-20S was reduced to 68-fold. Replacing P2 O-methyl-serine in PKS2086 with P2 Ala (PKS2058) improved isoform selectivity ( $\beta$ 5c/ $\beta$ 5i = 230) and maintained comparable inhibition activity. These preliminary structure-activity analyses suggested that even partially exposing S2 to

the solvent could be used to tune the physicochemical properties and selectivity profile of *N*,*C*-capped dipeptides, inviting further SAR studies (Table 1).

#### C-cap optimization

The *C*-cap binds to the S1 pocket, which is immediately adjacent to the active site. Structural studies of i-20S and c-20S have revealed that this S1 site dictates the selectivity of an inhibitor between  $\beta$ 5i and  $\beta$ 5c. We have shown that the P1 naphthyl ring is the selectivity driver, but the naphthyl ring is hydrophobic. To improve solubility and microsomal stability, we replaced the naphthalenemethyl ring in PKS2058 with the following substituents: 4-quinolinmethyl (PKS2068), 5-quinolinmethyl (PKS2073) and 4-indolemethyl (PKS2083). Quinolinyl analogues eliminated inhibitory activity against both  $\beta$ 5i and  $\beta$ 5c, while the indolyl analog decreased potency against  $\beta$ 5i and increased activity against  $\beta$ 5c. Replacing 1-naphthalenemethyl on the *C*-cap of PKS2086 with 8-quinolinmethyl (PKS2220) and 8-tetrahydroquinolinmethyl (PKS2226) led to 19-fold and 114-fold reductions in  $\beta$ 5i activities, respectively, further suggesting that a basic P1 moiety is detrimental to its interaction with the hydrophobic S1 pocket of  $\beta$ 5i. Using substituted benzyl groups as *C*-caps, as in PKS2160 and PKS2211, abolished selectivity. Therefore, we kept the naphthalenemethyl moiety as the *C*-cap for further SAR studies.

#### N-cap optimization-

Subsequent exploration of the *N*-cap with small, focused libraries revealed that a hydrophobic *N*-cap helps maintain selectivity. Extending the phenylpropionate group on the *N*-cap of PKS2058 yielded PKS2127. Phenylbutanoate-bearing PKS2127 was >5076-fold selective for  $\beta$ 5i over  $\beta$ 5c and yet with only 3-fold lower potency than PKS2058. Likewise, the abridged analog PKS2142 showed an 1840-fold preference for  $\beta$ 5i. Replacement of phenylpropionate in PKS2058 with the smaller trifluoroacetate in PKS2106 resulted in diminished activity with decent selectivity ( $\beta$ 5c / $\beta$ 5i = 229). Conversely, polar *N*-caps were not tolerated, as exemplified by the pyrazine PKS2098, the morpholine PKS2099, the thiazole PKS2102 and the isoxazole PKS2105, all of which suffered a significant reduction in  $\beta$ 5i inhibitory potency and compromised selectivity compared to PKS2058.

#### R3 optimization-

We have detailed the role of the defined S3 pocket in the species-selective binding of *N*,*C*-capped dipeptide inhibitors to the Mtb20S<sup>28</sup>, but we have found no reports exploring interactions with the S3 pocket for improving  $\beta$ 5i potency and selectivity for  $\beta$ 5i over  $\beta$ 5c. Accordingly, we changed R3 from branched chain amino acids to asparagine and glutamine. Replacing the R3 Asn-O<sup>t</sup>Bu with *O*-methyl-Ser (PKS2048) or Asn (PKS21083) abolished the inhibitory activities against both  $\beta$ 5i and  $\beta$ 5c, whereas the Asn-neopentyl analogue (PKS21066) was tolerated, suggesting that Asn coupled with a bulky moiety on the side chain would be a potency driver for *N*,*C*-capped dipeptide proteasome inhibitors. Replacement of R3 Asn-O<sup>t</sup>Bu with Asn-oxetane (PKS2130) reduced potency against  $\beta$ 5i over 100-fold compared to PKS2058, further testifying to the critical role of the bulky O<sup>t</sup>Bu group in inhibition of  $\beta$ 5i activity. Replacing the R3 Asn-O<sup>t</sup>Bu (PKS2086) with Asn amide with a fully substituted N atom (PKS2209, 2230BP, 2205 and 2208) resulted in significant loss of affinity for both  $\beta$ 5i and  $\beta$ 5c. However, the bulky Asn piperidine in PKS2206 did not

impose a great loss in potency and retained selectivity. Removal of the O atom from Asn-O<sup>t</sup>Bu of PKS2086 provided PKS2230. While their potencies and selectivity profiles are similar, the physicochemical properties of 'amide' PKS2230 would be presumably better than those of the '*N*-alkoxyamide' PKS2086. Improvement in  $\beta$ 5i potency was observed for the R3 Gln-<sup>t</sup>Bu analogue PKS2243 (IC<sub>50</sub> = 0.0023  $\mu$ M), suggesting that Gln-O<sup>t</sup>Bu fits the S3 pocket better than Asn-O<sup>t</sup>Bu, despite their comparable lengths. The trend was further attested by the observation that replacing R3 Asn-O<sup>t</sup>Bu of PKS2058 (IC<sub>50</sub> = 0.0057  $\mu$ M) with Gln-O<sup>t</sup>Bu (PKS2255, IC<sub>50</sub> = 0.0015  $\mu$ M) boosted  $\beta$ 5i potency by 3.8-fold. However, potency against  $\beta$ 5c was enhanced slightly more, impairing  $\beta$ 5i selectivity by 2-to 3-fold (PKS2255 vs. PKS2058, PKS2243 vs. PKS2086). We thus chose Asn-<sup>t</sup>Bu (selectivity) or Gln-<sup>t</sup>Bu (potency) as R3 for further optimization.

Introduction of a 4-F substituent on the C-cap naphthalenemethyl of PKS2255, which was expected to block metabolism, gave PKS21210, an equipotent  $\beta$ 5i inhibitor with about 2fold improvement in selectivity relative to PKS2255. The isoform selectivity ( $\beta 5c/\beta 5i = 204$ ) of PKS21271, a hybrid compound incorporating both the 4-F naphthalenemethyl group and the R3 Asn-<sup>t</sup>Bu, was slightly improved relative to that of PKS2255 ( $\beta$ 5c / $\beta$ 5i = 152), whereas the potency was compromised by over 30-fold. Because 3-indoleglyoxylate was reported to be an optimal N-cap for dipeptide inhibitors of the human 20S proteasome with a P3 threonine residue,  $^{40}$  we incorporated it into the scaffold of current  $\beta$ 5i inhibitor. Combining 3-indoleglyoxylate N-cap with R3 Asn-<sup>t</sup>Bu yielded a  $\beta$ 5i inhibitor with pM IC50 and over 100-fold selectivity (PKS3053). Additionally, we revisited the 5methylisoxazole-3-carboxylate N-cap with an aim to further improve the selectivity profile. Replacing the phenylpropionate group of PKS21210 and PKS21217 with 5methylisoxazole-3-carboxylate improved selectivity indices from 152- and 204-fold to 508-(PKS21272) and 984-fold (PKS21213), and unexpectedly, also improved \$\beta5\$i inhibition by 1.3- and 2.7- fold. Still better, 765- and 1187- fold isoform selectivities were recorded for two R2 O-methyl-serine analogs, PKS21245 and PKS21244. A 4-F substituent on the C-cap naphthalenemethyl contributed over 4-fold improvement to the noteworthy isoform selectivity of PKS21244 compared to that of PKS2244.

Next, we assessed the influence of a sulfonyl *N*-cap on asparagine and glutamine derivatives (Table 2). We replaced phenylpropionate with tosyl on the *N*-cap of PKS2255, yielding PKS2256. IC50s were determined to be 400 pM against  $\beta$ 5i and 424 nM against  $\beta$ 5c, affording 3 orders of magnitude of selectivity. As in compounds with the selectivity-enhancing 4-F substituents described above, a further 2-fold improvement in selectivity was observed for PKS21209 ( $\beta$ 5c / $\beta$ 5i = 2329). However, PKS21209 exhibited a 6-fold reduction in potency against  $\beta$ 5i compared to PKS2256. Again, replacing the Gln-<sup>t</sup>Bu of PKS21209 with Asn-<sup>t</sup>Bu (PKS21206) lost 6.8-fold in potency of inhibition of  $\beta$ 5i, but improved selectivity to 3274-fold over inhibition of  $\beta$ 5c. This pattern of impact on  $\beta$ 5i activity was reinforced in the R2 O-methyl-serine analogs PKS21265 and PKS21266, where the Gln-<sup>t</sup>Bu PKS21265 (IC<sub>50</sub> = 0.8 nM) exhibited 70-fold greater activity over Asn-<sup>t</sup>Bu PKS21266 (IC<sub>50</sub> = 56 nM) (Figure 3).

We further determined  $\beta 1$  or  $\beta 2$  activities of 6 select compounds in either i-20S or c-20S, and all exhibited <50% inhibition against  $\beta 1i$ ,  $\beta 1c$ ,  $\beta 2i$ , or  $\beta 2c$  at a concentration of 33.3  $\mu$ M, suggesting they are *bona fide*  $\beta 5i$  specific inhibitors (Table S1).

Blackburn and others have shown that N,C-capped dipeptides are generally specific inhibitors of proteasome over a panel of proteases.<sup>40</sup> We next tested these compounds against  $\alpha$ -chymotrypsin, cathepsin L and trypsin with the compounds at 100, 33.3 and 11.1  $\mu$ M. As shown in Table S2, all the compounds were inactive against all three proteases, except for weak inhibition of  $\alpha$ -chymotrypsin and cathepsin L by PKS3053. Thus, in agreement with the previous study,<sup>40</sup> these compounds were highly specific for proteasome inhibition.

#### Inhibition kinetics of i-20S β5i

The kinetics of proteasome inhibition can be complicated by the duplication of each active site in different rings of the proteasome.<sup>41</sup> We have observed that an active site binding inhibitor can behave like a non-competitive inhibitor.<sup>38</sup> We selected PKS21265 for elucidation of the kinetics of inhibition of  $\beta$ 5i. The Lineweaver-Burk plot of 1/V versus 1/S in the presence of different concentrations of PKS21265 yielded a series of straight lines that had the same x-intercept, suggesting a typical non-competitive inhibition kinetics (Figure 3B).

#### Intracellular proteasome inhibition

To investigate if these *N*,*C*-capped dipeptides are cell permeable, we selected three compounds with *N*-cap amides and 3 compounds with *N*-cap sulfonamides and tested their intracellular proteasome inhibition in Karpas-1106P cells (which express i-20S but not c-20S) and HeLa cells (which express c-20S but not i-20S). These compounds dose-dependently inhibited i-20S  $\beta$ 5i in Karpas-1106P and c-20S  $\beta$ 5c in Hela cells (Figure 4A), and in situ potencies were correlated well with their activities determined *in vitro* with purified enzymes (Table 3).

#### Inhibition of T cell activation and proliferation

We have shown that  $\beta$ 5i-selective  $\beta$ -amino acid based dipeptides inhibit the activation and proliferation of human T cells in peripheral blood mononuclear cells.<sup>37</sup> We tested the same 6 compounds as above for their inhibitory activity against primary human T cells stimulated by anti-CD3 antibody (Figure 4B). PKS3053 was two times more active than PKS21210 and > 3 times more active than PKS21272, and the activity tracked with their IC50s against  $\beta$ 5i. With its *N*-cap sulfonamide, PS21265 was the most potent compound, with 2-fold greater potency compared to PKS21209 and 5-fold compared to PKS21266 (Table 3).

# CHEMISTRY

The synthesis of dipeptide analogs with different *N*-caps is shown in Scheme 1. PKS2058 was synthesized by condensation of PKS21059<sup>35</sup> with PKS2026.<sup>35</sup> The reaction gave poor yield (6%) due to intramolecular cyclization of PKS21059 giving *N*-tert-butoxysuccinimide byproduct. Other *N*-cap analogs were synthesized from PKS21054.<sup>35</sup> Debenzylation of

PKS21054 and coupling with Ala-OBn followed by debenzylation and coupling with 1methylnaphthylamine gave PKS2095. Finally, BOC-deprotection and coupling with appropriate carboxylic acids gave the *N*-cap analogs. *C*-cap analogs were synthesized from PKS21059 (Scheme 2). Coupling of PKS21059 with Ala-OBn and subsequent debenzylation gave PKS2067. Different amines were coupled with PKS2067 to give *C*-cap analogs. PKS2048 was synthesized from PKS2032<sup>36</sup> by BOC-deprotection and coupling with 3-phenylpropanoic acid.

The synthesis of compounds where P2 is methylserine is shown in Scheme 3. The commercially available O-methylserine was coupled with 3-methoxybenzylamine and subsequent BOC-deprotection gave compound PKS2158. The condensation of PKS21059 with PKS2158 to provide compound PKS2160. Similarly, compound PKS2220 was synthesized starting from O-methylserine and 8-quinolinemethylamine. Compound PKS2211 was synthesized by condensation of O-methylserine with 2,3dimethoxybenzylamine. The condensation product was subjected to BOC-deprotection and coupled with BOC-Asp(OBn)-OH to give PKS2191. Again BOC-deprotection and coupling with 3-phenylpropanoic acid yielded PKS2200. The debenzylation of PKS2200 and condensation with O-tertbutylhydroxylamine gave the compound PKS2211. Similarly, following the same set of reactions compound PKS2226 was synthesized starting from Omethylserine and 8-quinolinemethylamine. Condensation of PKS2124<sup>10</sup> with 3aminooxetane yielded compound PKS2130. Similarly, compound PKS2230 and PKS2209 were synthesized by condensation of PKS220110 with tert-butylamine and azetidine respectively. PKS2244 was synthesized from PKS2131<sup>10</sup> by converting to *tert*-butylamide followed by BOC-deprotection and coupling with 5-methylisoxazole-3-carboxylic acid.

Condensation of BOC-Glu-OBn with *tert*-butylamine followed by BOC-deprotection gave **PKS2233**. It was then coupled with 3-phenylpropanoic acid and subsequent debenzylation gave PKS2239. PKS21202 was synthesized by condensation of BOC-Ala-OSu with 4-fluoromethyl-1-naphthylmethylamine and subsequent BOC-deprotection. Condensation of PKS2239 with PKS2082,<sup>35</sup> PKS2026 and PKS21202 gave compounds PKS2243, PKS2255 and PKS21210 respectively (Scheme 4). PKS3052<sup>37</sup> was coupled with PKS2066 and PKS21013<sup>35</sup> was coupled with PKS21202 to provide compounds PKS3035 and PKS21271 respectively. *O*-methylserine was coupled with 4-fluoromethyl-1-naphthylmethylamine and subsequently BOC was deprotected to give compound PKS21238. PKS21178<sup>37</sup> was coupled with PKS21202 and PKS21238 to give compounds PKS21213 and PKS21244 respectively. 5-methylisoxazole-3-carboxylic acid was subjected to condensation with PKS2233 and subsequent debenzylation furnished the compound PKS21231. PKS21233 was coupled with PKS21202 and PKS21238 to give compounds PKS21245 respectively. Finally, compounds with *N*-sulfonyl cap were synthesized by HATU mediated amide formation of corresponding acids and amines (Scheme 6)

# CONCLUSIONS

We found that several factors significantly contribute to potency and selectivity of *N*,*C*-capped dipeptides: 1) for inhibition, neither  $\beta$ 5i nor  $\beta$ 5c prefers benzopyridinyl at P1, but an indole moiety is acceptable, suggesting the hydrophobic nature of the S1 pocket and H-bond

acceptor presented; 2) a bulky P1 (*C*-cap) is the major determinant of selective inhibition of  $\beta$ 5i over  $\beta$ 5c; 2) P3 glutamine with <sup>*t*</sup>-butyl amide side chain generally improved potency by 3- to 50-fold for  $\beta$ 5i and by 30- to 40-fold for  $\beta$ 5c compared to P3 asparagine, without affecting the selectivity; 4) a hydrophobic *N*-cap is generally preferred; 5) these inhibitors act only on the chymotryptic  $\beta$ 5 subunits.

Recent studies of  $\beta$ 5i selective inhibitors have presented several observations: 1) inhibition of both  $\beta$ 5i and  $\beta$ 5c without inhibition of  $\beta$ 2i/c and  $\beta$ 1i/c can kill cancer cells;<sup>42</sup> however, whether inhibition of only  $\beta$ 5i or  $\beta$ 5c can induce cell death is still understudied; 2) selective inhibition of  $\beta$ 5i by peptide boronates does not lead to cell death of plasma cells and pDCs; <sup>43</sup> 3) inhibition of both  $\beta$ 5 and  $\beta$ 2 was synergistic for death of breast cancer cell line cells and inhibition of both  $\beta$ 5 and  $\beta$ 1 was additive.<sup>44, 45</sup> In contrast, non-cytotoxic inhibition of  $\beta$ 5i with compounds that showed no inhibitory activity against  $\beta$ 2c,  $\beta$ 2i,  $\beta$ 1c or  $\beta$ 1i sufficed to inhibit human T cell proliferation and might afford partial inhibition of the function of many types of immune cells with the cumulative effect of therapeutic efficacy in chronic autoimmune states.

# **EXPERIMENTAL SECTION**

Suc-LLVY-AMC, Ac-ANW-AMC, Z-VLR-AMC, Ac-LLE-AMC, Ac-PAL-AMC, human c-20S (RBC) and i-20S (PBMC) were from Boston Biochem (Cambridge, MA). Enzymatic assays were recorded on a Molecular Devices SpectraMax M5 plate readers. The human blood samples were sourced ethically and their research use was in accord with the terms of the informed consent. Purity of all final compounds were > 95%.

#### Synthetic Methods

**Chemicals and Spectroscopy:** All commercially available materials were purchased from Bachem, Aldrich, P3 BioSystems, or other vendors and were used as received unless stated otherwise. All non-aqueous reactions were performed under argon in oven-dried glasswares. Reactions were monitored using Waters Acquity Ultra Performance Liquid Chromatography (UPLC). All HPLC purifications were done by Varian PrepStar HPLC system or Waters Autopure (mass directed purification system) using Prep C18 5µm OBD (19 X 150 mm) column. <sup>1</sup>H- and <sup>13</sup>C- NMR spectra were acquired on a Bruker DRX-500 spectrometer. Chemical shifts  $\delta$  are expressed in parts per million, with the solvent resonance as an internal standard (chloroform-d, <sup>1</sup>H: 7.26; <sup>13</sup>C: 77.16 ppm; DMSO-d6, <sup>1</sup>H: 2.50 ppm; <sup>13</sup>C: 39.52 ppm). Hexafluorobenzene was used as internal standard for <sup>19</sup>F NMR. NMR data are reported as following: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, b = broad), coupling constant, and integration. We followed our reported procedure for HATU or EDC mediated amide bond formation, *boc*-deprotection and *O*-debenzylation.<sup>37,38</sup> All final compounds are > 95% pure.

**PKS2058:** Synthesized by following the general procedure for EDC mediated amide bond formation. Yield = 6%. <sup>1</sup>H NMR (500 MHz, DMSO- $d_{6}$ ) & 10.40 (s, 1H), 8.46 (t, *J* = 5.8 Hz, 1H), 8.20 - 8.16 (m, 1H), 8.15 (d, *J* = 7.2Hz, 1H), 8.10 - 8.02 (m, 1H), 7.96 - 7.90 (m, 1H), 7.86 - 7.80 (m, 1H), 7.56 - 7.50 (m, 2H), 7.47 - 7.41 (m, 2H), 7.28 - 7.24 (m, 2H), 7.20 - 7.13 (m, 3H), 4.70 - 4.67 (m, 2H), 4.62 - 4.56 (m, 1H), 4.29 - 4.22 (m, 1H), 2.82 - 2.75 (m, 2H), 7.86 - 7.80 (m, 1H), 7.86 - 7.80 (m, 2H), 7.80 (m, 2H), 7.80 (m, 2H), 7.80 (m, 2H), 7.80

2H), 2.55 - 2.48 (m, 1H), 2.44 - 2.38 (m, 2H), 2.36 - 2.30 (m, 1H), 1.24 (d, J = 7.2 Hz, 3H), 1.22 & 1.10 (s, rotamers, 9H). LCMS calc. for  $C_{31}H_{39}N_4O_5$  [M+H]<sup>+</sup> : 547.3. Found: 547.4.

**PKS2076:** Synthesized by following the general procedure for *O*-debenzylation of **PKS21054** (592 mg, 1.5 mmol). After completion of reaction (5 h), the mixture was filtered through celite and filtrate was evaporated to give product (450 mg, 98%). <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  9.28 & 8.98 (s, rotamers, 1H), 6.92 & 5.76 (b, rotamers, 1H), 4.59 – 4.47 (m, 1H), 3.22 – 2.69 (m, 2H), 1.44 (s, 9H), 1.27 & 1.24 (s, rotamers, 9H). LCMS calc. for C<sub>13</sub>H<sub>23</sub>N<sub>2</sub>O<sub>6</sub> [M–H]<sup>-</sup>: 303.2. Found: 303.5.

**PKS2081:** Synthesized by following the general procedure for HATU mediated coupling of **PKS2076** (304 mg, 1 mmol) and *O*-benzylalanine hydrochloride (237 mg, 1.1 mmol). Isolated crude product was purified by recrystallization with ethanol-water to give pure product (276 mg, 59%). <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  8.20 (s, 1H), 7.51 – 7.34 (m, 6H), 6.13 & 5.89 (b, rotamers, 1H), 5.20 (d, *J* = 12.4 Hz, 1H), 5.15 (d, *J* = 12.4 Hz, 1H), 4.60 – 4.45 (m, 2H), 2.79 – 2.66 (m, 1H), 2.49 – 2.45 (m, 1H), 1.46 (s, 9H), 1.41 (d, *J* = 7.2 Hz, 3H), 1.26 (s, 9H). LCMS calc. for C<sub>23</sub>H<sub>36</sub>N<sub>3</sub>O<sub>7</sub> [M+H]<sup>+</sup> : 466.3. Found: 466.5.

**PKS2092:** Synthesized by following the general procedure for *O*-debenzylation of **PKS2081** (150 mg, 0.32 mmol). After completion of reaction, mixture was filtered through celite and evaporated to give product (120 mg, quant.). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  12.96 (b, 1H), 10.37 (s, 1H), 7.94 (d, *J* = 6.7 Hz, 1H), 7.01 (d, *J* = 6.6 Hz, 1H), 4.28 – 4.23 (m, 1H), 4.10 – 4.04 (m, 1H), 2.39 (dd, *J* = 14.4, 4.4 Hz, 1H), 2.27 (dd, *J* = 14.4, 9.9 Hz, 1H), 1.37 (s, 9H), 1.23 (d, *J* = 7.2 Hz, 3H), 1.14 (s, 9H).

**PKS2095:** Synthesized following the general procedure for HATU mediated coupling of **PKS2092** (120 mg, 0.32 mmol) and 1-naphthylmethylamine (56 µl, 0.38 mmol). After completion of reaction (6 h), the mixture was precipitated with water. The precipitate was filtered and dried to give product (153 mg, 93%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.26 (s, 1H), 8.47 (t, J = 5.7 Hz, 1H), 8.06 – 8.04 (m, 1H), 8.01 (d, J = 7.4 Hz, 1H), 7.96 – 7.94 (m, 1H), 7.86 – 7.84 (m, 1H), 7.57 – 7.52 (m, 2H), 7.48 – 7.43 (m, 2H), 6.94 (d, J = 8.1 Hz, 1H), 4.74 (d, J = 5.8 Hz, 2H), 4.32 – 4.26 (m, 2H), 2.46 (dd, J = 14.6, 5.5 Hz, 1H), 2.29 (dd, J = 14.6, 8.5 Hz, 1H), 1.37 (s, 9H), 1.24 (d, J = 7.0 Hz, 3H), 1.13 (s, 9H).

**PKS2097:** Synthesized by following the general procedure for *boc*-deprotection of **PKS2095** (118 mg, 0.23 mmol). Isolated crude was triturated with diethyl ether to give product (120 mg, 98%). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.68 (s, 1H), 8.71 (d, *J* = 7.3 Hz, 1H), 8.58 (t, *J* = 5.7 Hz, 1H), 8.16 (s, 3H), 8.07 – 8.05 (m, 1H), 7.97 – 7.95 (m, 1H), 7.87 – 7.85 (m, 1H), 7.58 – 7.53 (m, 2H), 7.49 – 7.44 (m, 2H), 4.79 – 4.71 (m, 2H), 4.39 – 4.34 (m, 1H), 4.13 (m, 1H), 2.70 (dd, *J* = 16.4, 4.9 Hz, 1H), 2.56 (dd, *J* = 16.4, 8.1 Hz, 1H), 1.28 (d, *J* = 7.0 Hz, 3H), 1.15 (s, 9H). LCMS calc. for C<sub>22</sub>H<sub>31</sub>N<sub>4</sub>O<sub>4</sub> [M+H]<sup>+</sup> : 415.2. Found: 415.5.

**PKS2098:** Synthesized by following the general procedure for HATU mediated coupling of pyrazine-2-carboxylic acid (2.5 mg, 0.02 mmol) and **PKS2097** (10.6 mg, 0.02 mmol). The crude was purified by HPLC to give 10.3 mg (99%) of product. <sup>1</sup>H NMR (500 MHz,

DMSO- $d_6$ ) 810.45 (s, 1H), 9.19 (d, J = 1.4 Hz, 1H), 8.94 (d, J = 8.1 Hz, 1H), 8.92 (d, J = 2.5 Hz, 1H), 8.78 (t, J = 2.0 Hz, 1H), 8.50 (t, J = 5.8 Hz, 1H), 8.37 (d, J = 7.3 Hz, 1H), 8.07 – 8.05 (m, 1H), 7.95 – 7.93 (m, 1H), 7.85 – 7.83 (m, 1H), 7.56 – 7.52 (m, 2H), 7.48 – 7.44 (m, 2H), 4.85 – 4.81 (m, 1H), 4.75 (d, J = 5.8 Hz, 2H), 4.36 – 4.30 (m, 1H), 2.68 (dd, J = 14.7, 7.2 Hz, 1H), 2.63 (dd, J = 14.7, 5.2 Hz, 1H), 1.27 (d, J = 7.1 Hz, 3H), 1.06 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) 8 172.3, 170.2, 168.0, 162.8, 148.3, 144.5, 143.9, 134.9, 133.7, 131.2, 128.9, 127.9, 126.6, 126.2, 125.8, 125.6, 123.8, 81.1, 50.5, 49.2, 40.6, 35.4, 26.6, 18.4. LCMS calc. for C<sub>27</sub>H<sub>33</sub>N<sub>6</sub>O<sub>5</sub> [M+H]<sup>+</sup> : 521.3. Found: 521.5.

**PKS2099:** Synthesized by following the general procedure for HATU mediated coupling of morpholine-4-acetic acid (3.0 mg, 0.02 mmol) and **PKS2097** (10.6 mg, 0.02 mmol). The crude was purified by HPLC to give 10.6 mg (98%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  10.38 (s, 1H), 8.50 (t, J = 5.8 Hz, 1H), 8.20 (d, J = 7.3 Hz, 1H), 8.08 – 8.06 (m, 1H), 7.98 – 8.02 (m, 1H), 7.96 – 7.94 (m, 1H), 7.85 (dd, J = 7.0, 2.6 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.48 – 7.44 (m, 2H), 4.73 (d, J = 5.7 Hz, 2H), 4.62 – 4.57 (m, 1H), 4.31 – 4.27 (m, 1H), 3.61 (t, J = 4.6 Hz, 4H), 2.94 (s, 2H), 2.50 – 2.43 (m, 6H), 1.25 (d, J = 7.2 Hz, 3H), 1.11 (s, 9H). LCMS calc. for C<sub>28</sub>H<sub>40</sub>N<sub>5</sub>O<sub>6</sub> [M+H]<sup>+</sup>: 542.3. Found: 542.5.

**PKS2102:** Synthesized by following the general procedure for HATU mediated coupling of 2-methylthiazole-3-carboxylic acid (2.9 mg, 0.02 mmol) and **PKS2097** (10.6 mg, 0.02 mmol). The crude was purified by HPLC to give 8.4 mg (77%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.44 (s, 1H), 8.52 (t, J = 5.8 Hz, 1H), 8.32 (d, J = 7.3 Hz, 1H), 8.29 (d, J = 8.1 Hz, 1H), 8.12 (s, 1H), 8.08 – 8.06 (m, 1H), 7.96 – 7.94 (m, 1H), 7.84 (dd, J = 7.0, 2.5 Hz, 1H), 7.56 – 7.52 (m, 2H), 7.48 – 7.44 (m, 2H), 4.78 – 4.74 (m, 3H), 4.35 – 4.29 (m, 1H), 2.71 (s, 3H), 2.63 (dd, J = 14.6, 7.3 Hz, 1H), 2.58 (dd, J = 14.6, 5.3 Hz, 1H), 1.26 (d, J = 7.1 Hz, 3H), 1.07 (s, 9H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ) & 172.4, 170.4, 168.0, 166.7, 160.4, 149.2, 134.9, 133.7, 131.3, 128.9, 127.9, 126.6, 126.2, 125.9, 125.6, 124.7, 123.8, 81.1, 50.3, 49.1, 40.6, 35.6, 26.6, 19.2, 18.5. LCMS calc. for C<sub>27</sub>H<sub>34</sub>N<sub>5</sub>O<sub>5</sub>S [M+H]<sup>+</sup>: 540.2. Found: 540.5.

**PKS2105:** Synthesized by following the general procedure for HATU mediated coupling of 5-methylisoxazole-3-carboxylic acid (2.5 mg, 0.02 mmol) and **PKS2097** (10.6 mg, 0.02 mmol). The crude was purified by HPLC to give 10.2 mg (97%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  10.37 (s, 1H), 8.62 (d, J= 8.0 Hz, 1H), 8.48 (t, J= 5.8 Hz, 1H), 8.27 (d, J= 7.4 Hz, 1H), 8.06 – 8.04 (m, 1H), 7.96 – 7.94 (m, 1H), 7.85 (dd, J= 7.4, 1.9 Hz, 1H), 7.56 – 7.52 (m, 2H), 7.48 – 7.43 (m, 2H), 6.54 (s, 1H), 4.79 – 4.76 (m, 1H), 4.74 (d, J= 5.8 Hz, 2H), 4.34 – 4.28 (m, 1H), 2.60 (dd, J= 14.7, 5.5 Hz, 1H), 2.55 (dd, J= 14.7, 8.0 Hz, 1H), 2.47 (s, 3H), 1.25 (d, J= 7.1 Hz, 3H), 1.09 (s, 9H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ )  $\delta$  171.9, 171.4, 169.7, 167.5, 158.4, 158.4, 134.4, 133.2, 130.8, 128.5, 127.5, 126.2, 125.8, 125.4, 125.2, 123.4, 101.3, 80.6, 50.0, 48.6, 40.1, 34.6, 26.2, 18.1, 11.8. LCMS calc. for C<sub>27</sub>H<sub>34</sub>N<sub>5</sub>O<sub>6</sub> [M+H]<sup>+</sup>: 524.3. Found: 524.5.

**PKS2106:** Substrate (**PKS2097**) (10.6 mg, 0.02 mmol) was dissolved in 1 mL tetrahydrofuran and the solution was cooled to 0 °C. The mixture was basified with *N*-methylmorpholine (4.4 mL, 0.04 mmol). Trifluoroacetic anhydride (2.8 mL, 0.02 mmol) was

added and mixture was stirred for one hour at 0 °C. Solvent was evaporated and crude was purified by HPLC to give pure product (8.3 mg, 81%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  10.38 (s, 1H), 9.67 (b, 1H), 8.48 (t, J = 5.7 Hz, 1H), 8.41 (d, J = 7.3 Hz, 1H), 8.06 – 8.04 (m, 1H), 7.96 – 7.94 (m, 1H), 7.85 (d, J = 7.6 Hz, 1H), 7.57 – 7.53 (m, 2H), 7.48 – 7.43 (m, 2H), 4.74 – 4.70 (m, 3H), 4.33 – 4.27 (m, 1H), 2.63 (dd, J = 15.2, 5.0 Hz, 1H), 2.54 – 2.49 (m, 1H), 1.26 (d, J = 7.1 Hz, 3H), 1.12 (s, 9H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ )  $\delta$  172.4, 169.5, 167.4, 134.9, 133.7, 131.2, 128.9, 127.9, 126.6, 126.2, 125.9, 125.6, 123.8, 81.1, 50.7, 49.2, 40.6, 34.6, 26.6, 18.5. <sup>19</sup>F NMR (471 MHz, DMSO, C<sub>6</sub>F<sub>6</sub> external reference)  $\delta$  –71.81. LCMS calc. for C<sub>24</sub>H<sub>30</sub>F<sub>3</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 511.2. Found: 511.4.

**PKS2127:** Synthesized by following the general procedure for HATU mediated coupling of 4-phenylbutanoic acid (5.4 mg, 0.033 mmol) and **PKS2097** (17.3 mg, 0.033 mmol). The crude was purified by HPLC to give 15.7 mg (85%) of product. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.34 (s, 1H), 8.46 (t, *J* = 5.9 Hz, 1H), 8.11 – 8.06 (m, 3H), 7.95 – 7.93 (m, 1H), 7.85 – 7.83 (m, 1H), 7.56 – 7.52 (m, 2H), 7.45 (d, *J* = 5.0 Hz, 2H), 7.28 – 7.25 (m, 2H), 7.19 – 7.16 (m, 3H), 4.75 (dd, *J* = 15.3, 5.9 Hz, 1H), 4.70 (dd, *J* = 15.3, 5.7 Hz, 1H), 4.60 – 4.56 (m, 1H), 4.30 – 4.24 (m, 1H), 2.56 – 2.50 (m, 3H), 2.34 (dd, *J* = 14.8, 7.8 Hz, 1H), 2.13 (t, *J* = 7.4 Hz, 2H), 1.80 – 1.74 (m, 2H), 1.24 (d, *J* = 7.0 Hz, 3H), 1.10 (s, 9H). <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  172.4, 172.4, 171.0, 168.1, 142.3, 134.9, 133.7, 131.3, 128.9, 128.8, 128.7, 127.9, 126.6, 126.2, 126.2, 125.8, 125.7, 123.8, 81.00, 50.2, 49.0, 35.2, 35.1, 35.0, 27.5, 26.7, 18.5. LCMS calc. for C<sub>32</sub>H<sub>41</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 561.3. Found: 561.5.

**PKS2142:** Synthesized by following the general procedure for HATU mediated coupling of phenylacetic acid (4.9 mg, 0.036 mmol) and **PKS2097** (17.3 mg, 0.033 mmol). The crude was purified by HPLC to give 12.1 mg (68%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  10.38 (s, 1H), 8.47 (t, *J* = 5.9 Hz, 1H), 8.36 (d, *J* = 7.9 Hz, 1H), 8.18 (d, *J* = 7.4 Hz, 1H), 8.09 – 8.07 (m, 1H), 7.95 – 7.93 (m, 1H), 7.85 – 7.83 (m, 1H), 7.56 – 7.52 (m, 2H), 7.45 (d, *J* = 5.0 Hz, 2H), 7.29 – 7.19 (m, 5H), 4.76 (dd, *J* = 15.3, 6.0 Hz, 1H), 4.69 (dd, *J* = 15.3, 5.8 Hz, 1H), 4.60 – 4.56 (m, 1H), 4.29 – 4.23 (m, 1H), 3.46 (s, 2H), 2.56 – 2.51 (m, 1H), 2.37 (dd, *J* = 14.9, 7.3 Hz, 1H), 1.22 (d, *J* = 7.1 Hz, 3H), 1.10 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  172.4, 170.9, 170.5, 168.1, 136.6, 134.9, 133.7, 131.3, 129.5, 128.9, 128.6, 127.9, 126.8, 126.6, 126.2, 125.8, 125.7, 123.9, 81.0, 50.2, 49.1, 42.4, 40.6, 35.3, 26.7, 18.4. LCMS calc. for C<sub>30</sub>H<sub>37</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 533.3. Found: 533.5.

**PKS2063:** Synthesized by following the general procedure for HATU mediated coupling of **PKS21059** (33.6 mg, 0.1 mmol) and H-Ala-OBn.HCl (21.5 mg, 0.1 mmol). After completion of reaction, the mixture was precipitated by the addition of cold water. The precipitate was filtered and dried to give the product (27 mg, 54%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.20 (s, 1H), 8.31 (d, J= 7.0 Hz, 1H), 8.07 (d, J= 8.2 Hz, 1H), 7.40 – 7.32 (m, 5H), 7.28 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 5.11 (s, 2H), 4.66 (td, J= 9.0, 4.9 Hz, 1H), 4.33 – 4.27 (m, 1H), 2.80 – 2.76 (m, 2H), 2.41 – 2.37 (m, 3H), 2.26 (dd, J= 14.8, 9.5 Hz, 1H), 1.29 (d, J= 7.2 Hz, 3H), 1.14 (s, 9H).

**PKS2067:** Synthesized by following the general procedure for *O*-debenzylation of **PKS2063** (25.0 mg, 0.05 mmol). After completion of reaction (5 h), the mixture was filtered

through celite. The filtrate was evaporated and dried under vacuum to give the product (20 mg, quant.). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  12.35 (b, 1H), 9.75 (b, 1H), 7.80 (b, 1H), 7.20 – 7.17 (m, 2H), 7.12 – 7.09 (m, 1H), 7.01 (d, J = 7.4 Hz, 2H), 4.32 – 4.28 (m, 1H), 3.67 – 3.61 (m, 1H), 2.67 – 2.58 (m, 2H), 2.43 – 2.37 (m, 1H), 2.34 – 2.28 (m, 1H), 2.23 – 2.21 (m, 2H), 1.15 (d, J = 6.8 Hz, 3H), 1.12 (s, 9H).

**PKS2068:** Synthesized by following the general procedure for HATU mediated coupling of **PKS2067** (5.0 mg, 0.0123 mmol) and quinolin-4-ylmethylamine dihydrochloride (2.8 mg, 0.0123 mmol). The crude was purified by HPLC to give 2.0 mg (30%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.39 (s, 1H), 8.84 (d, J = 4.5 Hz, 1H), 8.60 (t, J = 6.0 Hz, 1H), 8.29 (d, J = 7.1 Hz, 1H), 8.19 – 8.15 (m, 2H), 8.05 (dd, J = 8.5, 1.3 Hz, 1H), 7.80 – 7.76 (m, 1H), 7.66 – 7.63 (m, 1H), 7.40 (d, J = 4.4 Hz, 1H), 7.27 – 7.24 (m, 2H), 7.20 – 7.15 (m, 3H), 4.80 – 4.78 (m, 2H), 4.64 – 4.59 (m, 1H), 4.30 – 4.27 (m, 1H), 2.80 – 2.77 (m, 2H), 2.56 – 2.51 (m, 1H), 2.44 – 2.40 (m, 2H), 2.35 (dd, J = 14.9, 7.1 Hz, 1H), 1.29 (d, J = 7.1 Hz, 3H), 1.06 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) & 172.4, 171.4, 170.7, 167.7, 150.1, 147.3, 144.6, 141.2, 129.4, 129.3, 128.3, 128.1, 126.6, 125.9, 125.8, 123.5, 118.9, 80.5, 64.9, 49.6, 48.8, 36.7, 34.8, 30.9, 26.2, 17.7. HRMS calc. for C<sub>30</sub>H<sub>38</sub>N<sub>5</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 548.2873. Found: 548.2857.

**PKS2073:** The title compound was prepared by following the general procedure for HATU mediated coupling of **PKS2067** (5.0 mg, 0.0123 mmol) and quinolin-5-ylmethylamine (2.0 mg, 0.0123 mmol). The crude was purified by HPLC to give 4.7 mg (70%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  10.39 (s, 1H), 8.92 – 8.91 (m, 1H), 8.55 (d, J = 8.6 Hz, 1H), 8.51 (t, J = 6.0 Hz, 1H), 8.18 – 8.15 (m, 2H), 7.94 (d, J = 8.4 Hz, 1H), 7.70 (dd, J = 8.5, 7.0 Hz, 1H), 7.57 – 7.54 (m, 2H), 7.27 – 7.24 (m, 2H), 7.19 – 7.15 (m, 3H), 4.79 (dd, J = 15.4, 6.1 Hz, 1H), 4.70 (dd, J = 15.4, 5.7 Hz, 1H), 4.61 – 4.56 (m, 1H), 4.27 – 4.21 (m, 1H), 2.80 – 2.76 (m, 2H), 2.53 – 2.48 (m, 1H), 2.43 – 2.39 (m, 2H), 2.33 (dd, J = 14.8, 7.2 Hz, 1H), 1.23 (d, J = 7.1 Hz, 3H), 1.10 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  172.4, 171.9, 171.0, 168.1, 150.6, 148.4, 141.7, 136.0, 132.5, 129.4, 128.9, 128.7, 128.6, 126.4, 126.3, 126.2, 121.7, 81.0, 70.2, 50.0, 49.1, 37.2, 35.3, 31.4, 26.7, 18.3. HRMS calc. for  $C_{30}H_{38}N_5O_5$  [M+H]<sup>+</sup>: 548.2867. Found: 548.2879.

**PKS2083:** Synthesized by following the general procedure for HATU mediated coupling of **PKS2067** (5.0 mg, 0.0123 mmol) and 4-(aminomethyl)indole (1.8 mg, 0.0123 mmol). The crude was purified by HPLC to give 5.8 mg (88%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  11.10 (s, 1H), 10.35 (s, 1H), 8.36 (t, J = 5.9 Hz, 1H), 8.16 (d, J = 8.0 Hz, 1H), 8.05 (d, J = 7.4 Hz, 1H), 7.32 – 7.25 (m, 4H), 7.20 – 7.16 (m, 3H), 7.01 (t, J = 7.6 Hz, 1H), 6.89 (d, J = 7.1 Hz, 1H), 6.52 – 6.51 (m, 1H), 4.61 – 4.48 (m, 3H), 4.28 – 4.24 (m, 1H), 2.81 – 2.77 (m, 2H), 2.53 – 2.49 (m, 1H), 2.46 – 2.38 (m, 2H), 2.33 (dd, J = 14.8, 7.6 Hz, 1H), 1.23 (d, J = 7.1 Hz, 3H), 1.13 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  172.1, 171.8, 170.8, 168.1, 141.7, 136.2, 130.5, 128.7, 128.6, 128.6, 126.6, 126.3, 125.3, 121.1, 117.8, 110.8, 81.0, 50.1, 49.0, 41.0, 37.2, 35.3, 31.4, 26.7, 18.7. HRMS calc. for C<sub>29</sub>H<sub>37</sub>N<sub>5</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup>: 558.2687. Found: 558.2698.

**PKS2038:** Synthesized by following the general procedure for *boc*-deprotection of **PKS2032** (95 mg, 0.22 mmol). After completion of reaction, dichloromethane and excess trifluoroacetic acid were evaporated and crude was triturated with diethyl ether. The mixture was filtered to give product (60 mg, 61%) as white powder. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.73 (d, *J* = 7.6 Hz, 1H), 8.51 (t, *J* = 5.7 Hz, 1H), 8.18 (b, 3H), 8.05 – 8.03 (m, 1H), 7.97 – 7.95 (m, 1H), 7.87 (d, *J* = 8.0 Hz, 1H), 7.58 – 7.54 (m, 2H), 7.49 – 7.43 (m, 2H), 4.80 – 4.72 (m, 2H), 4.45 – 4.39 (m, 1H), 4.03 (m, 1H), 3.65 (dd, *J* = 10.7, 3.8 Hz, 1H), 3.56 (dd, *J* = 10.7, 7.2 Hz, 1H), 3.25 (s, 3H), 1.28 (d, *J* = 7.0 Hz, 3H).

**PKS2048:** Synthesized following the general procedure for EDC coupling of 3phenylpropanoic acid (16.5 mg, 0.11 mmol) with **PKS2038** (44.3 mg, 0.1 mmol). The crude was purified by silica gel column chromatography to give 44.1 mg (87%) of product. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.24 – 8.18 (m, 2H), 8.15 (d, J = 7.8 Hz, 1H), 8.04 – 8.02 (m, 1H), 7.96 – 7.94 (m, 1H), 7.86 – 7.84 (m, 1H), 7.56 – 7.53 (m, 2H), 7.45 – 7.40 (m, 2H), 7.27 – 7.24 (m, 2H), 7.20 – 7.16 (m, 3H), 4.75 (d, J = 5.7 Hz, 2H), 4.50 (dt, J = 7.9, 6.0 Hz, 1H), 4.36 – 4.30 (m, 1H), 3.45 – 3.41 (m, 2H), 3.12 (s, 3H), 2.79 (t, J = 7.9 Hz, 2H), 2.47 – 2.44 (m, 2H), 1.26 (d, J = 7.1 Hz, 3H). LCMS calc. for C<sub>27</sub>H<sub>32</sub>N<sub>3</sub>O<sub>4</sub> [M+H]<sup>+</sup>: 462.2. Found: 462.3.

**PKS2154:** Synthesized following the general procedure of HATU mediated coupling of *boc*- $\beta$ -methoxyalanine dicyclohexylamine (100 mg, 0.25 mmol) and 3-methoxybenzyl amine (36 mL, 0.275 mmol). The product was isolated by ethyl acetate extraction and purified by silica-gel column chromatography (yield 74 mg, 87%). <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.26 – 7.22 (m, 1H), 6.86– 6.84 (m, 1H), 6.82 – 6.80 (m, 2H), 6.75 (t, *J* = 6.2 Hz, 1H), 5.42 (b, 1H), 4.47 – 4.46 (m, 2H), 4.28 (m, 1H), 3.85 (dd, *J*= 9.2, 3.8 Hz, 1H), 3.80 (s, 3H), 3.51 (dd, *J*= 9.2, 6.2 Hz, 1H), 3.38 (s, 3H), 1.44 (s, 9H).

**PKS2158:** Synthesized by following the general procedure for *boc*-deprotection of **PKS2154** (74 mg, 0.219 mmol). Isolated crude was dried under vacuum to give product (70 mg, 91%). <sup>1</sup>H NMR (500 MHz, Chloroform-*d*) & 9.13 (b, 1H), 8.01 (b, 2H), 7.56 (t, *J* = 5.8 Hz, 1H), 7.22 (t, *J* = 7.9 Hz, 1H), 6.80 (dd, *J* = 8.3, 2.5 Hz, 1H), 6.75 (d, *J* = 7.6 Hz, 1H), 6.73 – 6.72 (m, 1H), 4.41 – 4.35 (m, 2H), 4.29 (dd, *J* = 15.0, 5.5 Hz, 1H), 3.75 (s, 3H), 3.73 (dd, *J* = 10.4, 4.7 Hz, 1H), 3.67 (dd, *J* = 10.4, 5.3 Hz, 1H), 3.32 (s, 3H).

**PKS2160:** Synthesized following the general procedure for HATU mediated coupling of **PKS21059** (15.8 mg, 0.047 mmol) and **PKS2158** (18.4 mg, 0.052 mmol). The product was purified by HPLC (yield 16.2 mg, 62%).<sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.39 (s, 1H), 8.51 (t, *J* = 6.1 Hz, 1H), 8.20 (d, *J* = 7.9 Hz, 1H), 8.09 (d, *J* = 7.7 Hz, 1H), 7.27 – 7.25 (m, 2H), 7.21 – 7.15 (m, 4H), 6.82 – 6.81 (m, 2H), 6.77 (dd, *J* = 8.1, 2.4 Hz, 1H), 4.67 – 4.63 (m, 1H), 4.40 – 4.37 (m, 1H), 4.30 – 4.21 (m, 2H), 3.72 (s, 3H), 3.61 (dd, *J* = 9.8, 5.9 Hz, 1H), 3.50 (dd, *J* = 9.8, 4.6 Hz, 1H), 3.25 (s, 3H), 2.80 – 2.76 (m, 2H), 2.53 – 2.48 (m, 1H), 2.43 – 2.39 (m, 2H), 2.34 (dd, *J* = 14.8, 7.6 Hz, 1H), 1.11 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  172.0, 171.4, 169.7, 168.1, 159.8, 141.7, 141.3, 129.7, 128.8, 128.6, 126.4, 119.6, 112.9, 112.6, 81.1, 72.3, 58.8, 55.4, 53.7, 50.1, 42.5, 37.3, 35.3, 31.5, 26.7. HRMS calc. for C<sub>29</sub>H<sub>40</sub>N<sub>4</sub>O<sub>7</sub>Na [M+Na]<sup>+</sup>: 579.2789. Found: 579.2774.

**PKS2184:** Synthesized following the general procedure of HATU mediated coupling of *boc*-β-methoxyalanine dicyclohexylamine (200 mg, 0.5 mmol) and 2,3-dimethoxybenzyl amine (83 µL, 0.55 mmol). The product was isolated by ethyl acetate extraction and purified by silica-gel column chromatography (yield = 184 mg, quant.). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ 8.24 (t, J = 5.9 Hz, 1H), 7.00 – 6.97 (m, 1H), 6.93 (dd, J = 8.3, 1.7 Hz, 1H), 6.86 (d, J = 8.3 Hz, 1H), 6.81 (dd, J = 7.6, 1.7 Hz, 1H), 4.31 – 4.23 (m, 2H), 4.20 – 4.15 (m, 1H), 3.79 (s, 3H), 3.72 (s, 3H), 3.48 – 3.46 (m, 2H), 3.24 (s, 3H), 1.38 (s, 9H).

**PKS2190:** Synthesized by following the general procedure for *boc*-deprotection of **PKS2184** (180 mg, 0.49 mmol). Crude product was dried under vacuum to give viscous paste, which upon standing turned solid. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.78 (t, *J* = 5.7 Hz, 1H), 8.21 (b, 3H), 7.05 – 7.02 (m, 1H), 6.97 (dd, *J* = 8.2, 1.6 Hz, 1H), 6.83 (dd, *J* = 7.6, 1.6 Hz, 1H), 4.36 (dd, *J* = 15.1, 5.7 Hz, 1H), 4.31 (dd, *J* = 15.1, 5.6 Hz, 1H), 4.04 – 4.01 (m, 1H), 3.80 (s, 3H), 3.75 (s, 3H), 3.66 (d, *J* = 5.1 Hz, 2H), 3.30 (s, 3H).

**PKS2191:** Synthesized following the general procedure of HATU mediated coupling of *N*-(*tert*-butoxycarbonyl)-L-aspartic acid 4-benzyl ester (159 mg, 0.49 mmol) and PKS2190 (0.49 mmol, from previous step). The product was isolated by ethyl acetate extraction and recrystallized with ethanol-water (yield = 245 mg, 87% for 2 steps). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.29 (t, *J* = 5.8 Hz, 1H), 7.86 (d, *J* = 7.9 Hz, 1H), 7.38 – 7.31 (m, 5H), 7.28 (d, *J* = 8.0 Hz, 1H), 7.00 – 6.97 (m, 1H), 6.92 (dd, *J* = 8.2, 1.6 Hz, 1H), 6.78 (dd, *J* = 7.7, 1.6 Hz, 1H), 5.08 (d, *J* = 12.6 Hz, 1H), 5.05 (d, *J* = 12.6 Hz, 1H), 4.45 – 4.37 (m, 2H), 4.31 – 4.23 (m, 2H), 3.78 (s, 3H), 3.71 (s, 3H), 3.56 (dd, *J* = 9.7, 5.6 Hz, 1H), 3.46 (dd, *J* = 9.7, 5.1 Hz, 1H), 3.24 (s, 3H), 2.82 – 2.78 (m, 1H), 2.61 (dd, *J* = 16.3, 8.9 Hz, 1 H), 1.37 (s, 9H).

**PKS2197:** Synthesized by following the general procedure for *boc*-deprotection of **PKS2191** (115 mg, 0.2 mmol). The crude was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.81 (d, J = 7.8 Hz, 1H), 8.37 (t, J = 5.9 Hz, 1H), 8.26 (b, 3H), 7.39 – 7.34 (m, 5H), 6.99 (t, J = 7.9 Hz, 1H), 6.93 (dd, J = 8.3, 1.6 Hz, 1H), 6.78 (dd, J = 7.7, 1.6 Hz, 1H), 5.14 (d, J = 12.4 Hz, 1H), 5.11 (d, J = 12.4 Hz, 1H), 4.55 – 4.51 (m, 1H), 4.33 – 4.23 (m, 3H), 3.78 (s, 3H), 3.71 (s, 3H), 3.59 (dd, J = 9.9, 6.2 Hz, 1H), 3.52 (dd, J = 9.9, 4.8 Hz, 1H), 3.27 (s, 3H), 3.02 (dd, J = 17.5, 4.1 Hz, 1H), 2.82 (dd, J = 17.5, 8.8 Hz, 1H).

**PKS2200:** Synthesized following the general procedure of HATU mediated coupling of 3phenylpropanoic acid (33 mg, 0.22 mmol) and **PKS2197** (0.2 mmol, from previous step). The reaction mixture was precipitated with water and the precipitate was filtered and dried to give product (110 mg, 91% for 2 steps). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.30 (d, J =7.9 Hz, 1H), 8.24 (t, J = 5.9 Hz, 1H), 8.04 (d, J = 7.9 Hz, 1H), 7.38 – 7.32 (m, 5H), 7.28 – 7.25 (m, 2H), 7.18 – 7.15 (m, 3H), 6.99 (t, J = 7.9 Hz, 1H), 6.92 (dd, J = 8.0, 1.6 Hz, 1H), 6.79 (dd, J = 7.5, 1.6 Hz, 1H), 5.04 (s, 2H), 4.74 (td, J = 8.1, 5.7 Hz, 1H), 4.42 (dt, J = 7.8, 5.4 Hz, 1H), 4.31 – 4.24 (m, 2H), 3.78 (s, 3H), 3.72 (s, 3H), 3.56 (dd, J = 9.8, 5.8 Hz, 1H), 3.47 (dd, J = 9.8, 5.0 Hz, 1H), 3.24 (s, 3H), 2.85 – 2.77 (m, 3H), 2.59 (dd, J = 16.2, 8.3 Hz, 1H), 2.41 – 2.38 (m, 2H).

**PKS2204:** Synthesized by following general procedure for *O*-debenzylation. The reaction was not complete and isolated crude was purified by HPLC to give 40 mg product (50% bsrm; 16.0 mg starting material was recovered). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 12.34 (s, 1H), 8.27 – 8.24 (m, 2H), 7.95 (d, *J* = 7.9 Hz, 1H), 7.28 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 6.99 (t, *J* = 7.9 Hz, 1H), 6.93 (dd, *J* = 8.2, 1.5 Hz, 1H), 6.80 (dd, *J* = 7.7, 1.5 Hz, 1H), 4.66 – 4.60 (m, 1H), 4.43 – 4. 38 (m, 1H), 4.28 (d, *J* = 5.9 Hz, 2H), 3.78 (s, 3H), 3.72 (s, 3H), 3.56 (dd, *J* = 9.7, 5.8 Hz, 1H), 3.47 (dd, *J* = 9.7, 5.0 Hz, 1H), 3.24 (s, 3H), 2.79 (t, *J* = 7.9 Hz, 2H), 2.68 (dd, *J* = 16.6, 5.9 Hz, 1H), 2.48 – 2.39 (m, 3H).

**PKS2211:** Synthesized following the general procedure for HATU mediated coupling of *O*-*tert*-butyl hydroxylamine hydrochloride (6.6 mg, 0.0525 mmol) and **PKS2204** (18.0 mg, 0.035 mmol). After completion of reaction (1 h), mixture was diluted with water and extracted with ethyl acetate. The organic layer was evaporated and purified by HPLC to give the product (11.0 mg, 54%). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.37 (s, 1H), 8.37 (t, *J* = 6.0 Hz, 1H), 8.20 (d, *J* = 7.9 Hz, 1H), 8.05 (d, *J* = 7.7 Hz, 1H), 7.28 – 7.25 (m, 2H), 7.19 – 7.15 (m, 3H), 6.98 (t, *J* = 7.9 Hz, 1H), 6.92 (dd, *J* = 8.1, 1.6 Hz, 1H), 6.82 (d, *J* = 7.7 Hz, 1H), 4.67 – 4.63 (m, 1H), 4.40 (dt, *J* = 7.6, 5.3 Hz, 1H), 4.31 – 4.24 (m, 2H), 3.78 (s, 3H), 3.72 (s, 3H), 3.59 (dd, *J* = 9.8, 6.0 Hz, 1H), 3.49 (dd, *J* = 9.8, 4.6 Hz, 1H), 3.24 (s, 3H), 2.80 – 2.76 (m, 2H), 2.52 – 2.48 (m, 1H), 2.43 – 2.39 (m, 2H), 2.33 (dd, *J* = 14.9, 7.8 Hz, 1H), 1.11 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  172.0, 171.3, 169.7, 168.0, 152.6, 146.5, 141.7, 132.8, 128.8, 128.6, 126.3, 124.1, 120.3, 112.0, 81.0, 72.3, 60.4, 58.7, 56.1, 53.6, 50.1, 37.4, 37.2, 35.3, 31.4, 26.7. HRMS calc. for C<sub>30</sub>H<sub>42</sub>N<sub>4</sub>O<sub>8</sub>Na [M+Na]<sup>+</sup>: 609.2895. Found: 609.2897.

**PKS2175:** Synthesized following the general procedure for HATU coupling of boc-βmethoxyalanine dicyclohexylamine (80 mg, 0.02 mmol) and quinolin-8-ylmethylamine dihydrochloride (46 mg, 0.2 mmol) in 2 mL dimethylformamide. (*Note: reaction mixture was not soluble in dimethylformamide*) After completion of reaction (3 h), water was added to reaction mixture (reaction mixture turned transparent) and extracted twice with chloroform. The combined organic layer was washed with water followed by brine, dried over anhydrous sodium sulfate and evaporated. The crude product was purified by HPLC to give 68.5 mg (95%) of pure product. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ 8.96 – 8.94 (m, 1H), 8.46 (t, *J* = 6.2 Hz, 1H), 8.39 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.88 (d, *J* = 8.1 Hz, 1H), 7.64 (d, *J* = 7.1 Hz, 1H), 7.58 (dd, *J* = 8.3, 4.2 Hz, 1H), 7.54 (t, *J* = 7.6 Hz, 1H), 6.96 (d, *J* = 8.1 Hz, 1H), 4.96 – 4.87 (m, 2H), 4.26 – 4.22 (m, 1H), 3.54 – 3.53 (m, 2H), 3.27 (s, 3H), 1.39 (s, 9H).

**PKS2181:** Synthesized by following the general procedure for boc-deprotection of **PKS2175** (68.5 mg, 0.19 mmol). The crude was used in next step. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.98 (dd, J = 4.2, 1.8 Hz, 1H), 8.95 (t, J = 5.9 Hz, 1H), 8.42 (dd, J = 8.2, 1.8 Hz, 1H), 8.22 (b, 3H), 7.93 (dd, J = 8.1, 1.5 Hz, 1H), 7.67 (dd, J = 7.1, 1.5 Hz, 1H), 7.62 – 7.58 (m, 2H), 5.02 (dd, J = 15.9, 5.9 Hz, 1H), 4.96 (dd, J = 15.9, 5.7 Hz, 1H), 4.13 – 4.10 (m, 1H), 3.75 – 3.69 (m, 2H), 3.32 (s, 3H).

**PKS2188:** Synthesized following the general procedure for HATU mediated coupling of *N*-(*tert*-butoxycarbonyl)-L-aspartic acid 4-benzyl ester (61.4 mg, 0.19 mmol) and PKS2181

(0.19 mmol, from previous step). After completion of reaction (1 h), reaction mixture was diluted with water and extracted twice with ethyl acetate. Ethyl acetate layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated. The crude was dried under high vacuum and used in next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.96 (dd, J = 4.3, 1.8 Hz, 1H), 8.49 (t, J = 5.9 Hz, 1H), 8.43 (dd, J = 8.3, 1.8 Hz, 1H), 7.95 (d, J = 7.8 Hz, 1H), 7.90 (dd, J = 8.0, 1.5 Hz, 1H), 7.63 (dd, J = 7.1, 1.5 Hz, 1H), 7.61 – 7.55 (m, 2H), 7.37 – 7.30 (m, 6H), 5.09 (d, J = 12.7 Hz, 1H), 5.05 (d, J = 12.7 Hz, 1H), 4.96 (dd, J = 16.4, 6.0 Hz, 1H), 4.89 (dd, J = 16.4, 5.9 Hz, 1H), 4.51 – 4.48 (m, 1H), 4.43 (td, J = 8.5, 5.0 Hz, 1H), 3.63 (dd, J = 9.7, 5.5 Hz, 1H), 3.52 (dd, J = 9.7, 5.1 Hz, 1H), 3.28 (s, 3H), 2.85 – 2.81 (m, 1H), 2.62 (dd, J = 16.2, 8.9 Hz, 1H), 1.38 (s, 9H).

**PKS2194:** Synthesized by following the general procedure for boc-deprotection of **PKS2188** (0.19 mmol, from previous step). After completion of reaction (3 h), excess trfluoroacetic acid and dichloromethane were evaporated. Crude paste was washed twice with diethyl ether to give product as off-white solid (yield = 111 mg, 84% for 3 steps). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.94 (dd, J = 4.2, 1.8 Hz, 1H), 8.87 (d, J = 7.8 Hz, 1H), 8.53 (t, J = 6.0 Hz, 1H), 8.39 (dd, J = 8.4, 1.8 Hz, 1H), 8.27 (b, 3H), 7.88 (dd, J = 8.1, 1.5 Hz, 1H), 7.61 (dd, J = 7.2, 1.5 Hz, 1H), 7.58 – 7.54 (m, 2H), 7.40 – 7.31 (m, 5H), 5.13 (J = 12.5 Hz, 1H), 5.10 (d, J = 12.5 Hz, 1H), 4.97 (dd, J = 16.4, 6.1 Hz, 1H), 4.90 (dd, J = 16.4, 5.9 Hz, 1H), 4.62 – 4.58 (m, 1H), 4.27 (m, 1H), 3.66 (dd, J = 9.8, 6.0 Hz, 1H), 3.57 (dd, J = 9.8, 4.8 Hz, 1H), 3.30 (s, 3H), 3.04 (dd, J = 17.5, 3.9 Hz, 1H), 2.85 – 2.80 (m, 1H). LCMS calc. for C<sub>25</sub>H<sub>28</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 464.2. Found: 464.3.

**PKS2198:** Synthesized following the general procedure for HATU mediated coupling of 3-phenylproapnoic acid (26.4 mg, 0.176 mmol) and **PKS2194** (111 mg, 0.16 mmol). After completion of reaction (3 h), the mixture was precipitated with water. The precipitate was filtered and dried to give the product (77 mg, 81%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.94 (dd, J = 4.2, 1.8 Hz, 1H), 8.43 (t, J = 6.1 Hz, 1H), 8.38 (dd, J = 8.3, 1.8 Hz, 1H), 8.32 (d, J = 8.0 Hz, 1H), 8.12 (d, J = 7.8 Hz, 1H), 7.87 (dd, J = 8.1, 1.5 Hz, 1H), 7.61 (dd, J = 7.2, 1.5 Hz, 1H), 7.57 – 7.53 (m, 2H), 7.35 – 7.30 (m, 5H), 7.27 – 7.24 (m, 2H), 7.19 – 7.15 (m, 3H), 5.03 (s, 2H), 4.95 (dd, J = 16.4, 6.1 Hz, 1H), 4.90 (dd, J = 16.4, 5.9 Hz, 1H), 4.77 (td, J = 8.2, 5.7 Hz, 1H), 4.49 (dt, J = 7.8, 5.3 Hz, 1H), 3.62 (dd, J = 9.7, 5.8 Hz, 1H), 3.52 (dd, J = 9.7, 5.0 Hz, 1H), 3.28 (s, 3H), 2.85 – 2.77 (m, 3H), 2.59 (dd, J = 16.2, 8.4 Hz, 1H), 2.42 – 2.38 (m, 2H).

**PKS2202:** Synthesized by following general procedure for O-debenzylation of **PKS2198** (73 mg, 0.122 mmol). The mixture was filtered through celite, evaporated and purified by HPLC to give the product (19 mg, 30%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  12.34 (s, 1H), 8.26 – 8.23 (m, 2H), 7.99 (d, J = 7.8 Hz, 1H), 7.28 – 7.25 (m, 2H), 7.21 – 7.15 (m, 3H), 6.81 (dd, J = 7.5, 1.5 Hz, 1H), 6.76 (dd, J = 7.5, 1.5 Hz, 1H), 6.40 (t, J = 7.4 Hz, 1H), 4.63 (td, J = 7.8, 5.8 Hz, 1H), 4.40 (dt, J = 7.7, 5.4 Hz, 1H), 4.08 (dd, J = 15.3, 6.1 Hz, 1H), 4.03 (dd, J = 15.3, 5.9 Hz, 1H), 3.56 (dd, J = 9.8, 5.8 Hz, 1H), 3.47 (dd, J = 9.8, 5.0 Hz, 1H), 3.24 (s, 3H), 3.22 (t, J = 5.5 Hz, 2H), 2.80 (t, J = 7.9 Hz, 2H), 2.70 – 2.65 (m, 3H), 2.48 – 2.39 (m, 3H), 1.79 – 1.74 (m, 2H).

**PKS2226:** Synthesized following the general procedure for HATU mediated coupling of **PKS2202** (19.0 mg, 0.037 mmol) and *O-tert*-butylhydroxylamine hydrochloride (5.1 mg, 0.041 mmol). The product was purified by HPLC (17.9 mg, 83%) <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.40 (s, 1H), 8.41 (t, J = 6.1 Hz, 1H), 8.18 (d, J = 7.9 Hz, 1H), 8.10 (d, J = 7.8 Hz, 1H), 7.28 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 6.84 (d, J = 7.5 Hz, 1H), 6.75 (d, J = 7.5 Hz, 1H), 6.38 (t, J = 7.4 Hz, 1H), 5.24 – 5.20 (m, 1H), 4.67 – 4.62 (m, 1H), 4.40 – 4.37 (m, 1H), 4.10 (dd, J = 15.1, 6.3 Hz, 1H), 3.24 (s, 3H), 3.22 – 3.21 (m, 2H), 2.80 – 2.76 (m, 2H), 2.66 (t, J = 6.4 Hz, 2H), 2.52 – 2.48 (m, 1H), 2.42 – 2.39 (m, 2H), 2.35 (dd, J = 14.8, 7.5 Hz, 1H), 1.79 – 1.74 (m, 2H), 1.14 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) & 171.5, 170.9, 169.5, 167.6, 142.5, 141.2, 128.3, 128.1, 128.0, 126.5, 125.8, 121.3, 120.0, 114.6, 80.6, 71.7, 58.3, 53.1, 49.6, 41.3, 36.8, 34.8, 31.00, 27.2, 26.3, 21.3. HRMS calc. for C<sub>31</sub>H<sub>43</sub>N<sub>5</sub>O<sub>6</sub>Na [M+Na]<sup>+</sup>: 604.3106. Found: 604.3105.

**PKS2220:** Synthesized following the general procedure for HATU mediated coupling of **PKS21059** (20.2 mg, 0.06 mmol) and **PKS2181** (32.2 mg, 0.066 mmol). The product was purified be HPLC (yield = 21.6 mg, 62%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 10.36 (s, 1H), 8.95 (dd, J = 4.2, 1.8 Hz, 1H), 8.57 (t, J = 6.1 Hz, 1H), 8.38 (dd, J = 8.3, 1.8 Hz, 1H), 8.22 (d, J = 8.0 Hz, 1H), 8.15 (d, J = 7.8 Hz, 1H), 7.87 (dd, J = 8.3, 1.5 Hz 1H), 7.63 (dd, J = 7.4, 1.5 Hz, 1H), 7.59 – 7.54 (m, 2H), 7.28 – 7.25 (m, 2H), 7.20 – 7.17 (m, 3H), 4.98 – 4.89 (m, 2H), 4.71 – 4.67 (m, 1H), 4.50 – 4.46 (m, 1H), 3.66 (dd, J = 9.7, 5.8 Hz, 1H), 3.55 (dd, J = 9.7, 4.7 Hz, 1H), 3.29 (s, 3H), 2.80 – 2.77 (m, 2H), 2.55 – 2.51 (m, 1H), 2.43 – 2.40 (m, 2H), 2.35 (dd, J = 14.7, 7.7 Hz, 1H), 1.06 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) & 172.0, 171.5, 170.0, 168.0, 150.1, 145.8, 141.7, 136.9, 136.8, 128.8, 128.6, 128.1, 127.1, 126.9, 126.7, 126.3, 121.9, 81.0, 72.3, 58.8, 53.7, 50.1, 39.2, 37.2, 35.3, 31.5, 26.7. HRMS calc. for  $C_{31}H_{39}N_5O_6Na$  [M+Na]<sup>+</sup>: 600.2793. Found: 600.2783.

**PKS2130:** Synthesized following the general procedure for HATU mediated coupling of **PKS2124** (12.4 mg, 0.026 mmol) and 3-aminoxetane (2.2 µL, 0.031 mmol). The mixture was purified by HPLC to give product (12.0 mg, 87%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.71 (d, J = 6.4 Hz, 1H), 8.44 (t, J = 5.8 Hz, 1H), 8.21 (d, J = 7.3 Hz, 1H), 8.15 (d, J = 7.9 Hz, 1H), 8.07 – 8.05 (m, 1H), 7.95 – 7.94 (m, 1H), 7.84 (d, J = 7.8 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.47 – 7.42 (m, 2H), 7.27 – 7.24 (m, 2H), 7.19 – 7.15 (m, 3H), 4.76 (dd, J = 15.4, 5.9 Hz, 1H), 4.70 (dd, J = 15.4, 5.8 Hz, 1H), 4.64 – 4.51 (m, 4H), 4.34 – 4.22 (m, 3H), 2.78 (t, J = 7.8 Hz, 2H), 2.58 (dd, J = 15.2, 7.5 Hz, 1H), 2.44 – 2.40 (m, 3H), 1.25 (d, J = 7.2 Hz, 3H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  172.4, 172.0, 171.0, 169.8, 141.7, 134.8, 133.7, 131.2, 128.9, 128.7, 128.6, 127.9, 126.6, 126.3, 126.2, 125.8, 125.6, 123.8, 77.4, 77.3, 50.0, 49.1, 44.3, 40.6, 37.8, 37.1, 31.4, 18.3. LCMS calc. for C<sub>30</sub>H<sub>35</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 531.3. Found: 531.3.

**PKS2209:** Synthesized following the general procedure for HATU mediated coupling of **PKS2201** (15.2 mg, 0.03 mmol) and azetidine hydrochloride (4.2 mg, 0.045 mmol). The mixture was purified by HPLC to give pure product (10.5 mg, 64%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.51 (t, *J* = 5.9 Hz, 1H), 8.29 (d, *J* = 7.7 Hz, 1H), 8.16 (d, *J* = 7.9 Hz, 1H), 8.08 – 8.06 (m, 1H), 7.95 – 7.93 (m, 1H), 7.84 – 7.82 (m, 1H), 7.55 – 7.52 (m, 2H), 7.46 – 7.41

(m, 2H), 7.28 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 4.80 – 4.62 (m, 3H), 4.43 – 4.39 (m, 1H), 4.07 (td, J= 8.7, 6.3 Hz, 1H), 4.00 (td, J= 8.7, 6.3 Hz, 1H), 3.72 – 3.56 (m, 3H), 3.53 (dd, J = 9.8, 4.3 Hz, 1H), 3.24 (s, 3H), 2.79 (t, J= 7.8 Hz, 2H), 2.47 – 2.36 (m, 3H), 2.29 (dd, J= 15.4, 5.8 Hz, 1H), 2.12 – 2.03 (m, 2H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) & 171.4, 171.0, 169.3, 169.2, 141.2, 134.2, 133.2, 130.7, 128.4, 128.2, 127.3, 126.1, 125.8, 125.7, 125.3, 124.9, 124.8, 123.4, 71.7, 58.2, 53.5, 49.6, 49.1, 47.3, 40.2, 36.6, 33.1, 31.0, 14.5. LCMS calc. for C<sub>31</sub>H<sub>37</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 545.3. Found: 545.2.

**PKS2230:** Synthesized following the general procedure for HATU mediated coupling of **PKS2201** (15.2 mg, 0.03 mmol) and *tert*-butylamine (9.5  $\mu$ L, 0.09 mmol). The product was purified by HPLC to give pure product (8.7 mg, 52%). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.66 (t, *J* = 5.9 Hz, 1H), 8.19 (d, *J* = 7.7 Hz, 1H), 8.12 (d, *J* = 8.0 Hz, 1H), 8.07 – 8.05 (m, 1H), 7.94 – 7.92 (m, 1H), 7.83 – 7.81 (m, 1H), 7.55 – 7.50 (m, 3H), 7.46 – 7.40 (m, 2H), 7.28 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 4.79 (dd, *J* = 15.6, 6.0 Hz, 1H), 4.70 (dd, *J* = 15.6, 5.8 Hz, 1H), 4.63 – 4.59 (m, 1H), 4.45 – 4.41 (m, 1H), 3.63 (dd, *J* = 9.8, 6.1 Hz, 1H), 3.54 (dd, *J* = 9.8, 4.4 Hz, 1H), 3.24 (s, 3H), 2.78 (t, *J* = 7.9 Hz, 2H), 2.55 – 2.51 (m, 1H), 2.43 – 2.34 (m, 3H), 1.14 (s, 9H).<sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  171.3, 171.2, 169.3, 169.1, 141.3, 134.2, 133.2, 130.7, 128.4, 128.3, 128.1, 127.3, 126.1, 125.8, 125.7, 125.3, 124.9, 123.3, 71.8, 58.2, 53.3, 50.1, 49.7, 40.2, 38.5, 36.7, 31.0, 28.4. HRMS calc. for C<sub>32</sub>H<sub>40</sub>N<sub>4</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup>: 583.2891. Found: 583.2877.

**PKS2230BP:** Isolated as a byproduct in the above reaction. <sup>1</sup>H NMR (500 MHz, DMSO $d_6$ ) & 8.44 (t, J = 6.0 Hz, 1H), 8.32 (d, J = 7.7 Hz, 1H), 8.15 (d, J = 8.0 Hz, 1H), 8.09 – 8.04 (m, 1H), 7.96 – 7.91 (m, 1H), 7.84 – 7.80 (m, 1H), 7.55 – 7.50 (m, 2H), 7.45 – 7.39 (m, 2H), 7.28 – 7.23 (m, 2H), 7.20 – 7.15 (m, 3H), 4.77 (dd, J = 15.6, 5.8 Hz, 1H), 4.74 – 4.64 (m, 2H), 4.42 – 4.37 (m, 1H), 3.66 (dd, J = 9.7, 6.3 Hz, 1H), 3.55 (dd, J = 9.7, 4.1 Hz, 1H), 3.24 (s, 3H), 2.87 (s, 3H), 2.79 (t, J = 7.8 Hz, 2H), 2.67 (dd, J = 16.2, 8.4 Hz, 1H), 2.54 – 2.60 (m, 1H), 2.57 (s, 3H), 2.46 – 2.37 (m, 2H). LCMS calc. for C<sub>30</sub>H<sub>37</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 533.3. Found: 533.5.

**PKS2237:** The title compound was prepared following the general procedure for HATU mediated coupling of **PKS2131** (118 mg, 0.25 mmol) and *tert*-butylamine (32 mL, 0.3 mmol). The product was isolated by ethylacetate extraction and purified by HPLC (yield = 37.0 mg). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.59 (t, J = 5.8 Hz, 1H), 8.05 – 8.03 (m, 1H), 7.98 – 7.93 (m, 2H), 7.84 – 7.82 (m, 1H), 7.55 – 7.51 (m, 2H), 7.46 – 7.43 (m, 3H), 6.93 (d, J = 8.1 Hz, 1H), 4.79 – 4.70 (m, 2H), 4.47 – 4.43 (m, 1H), 4.31 – 4.26 (m, 1H), 3.61 (dd, J = 9.7, 5.7 Hz, 1H), 3.51 (dd, J = 9.7, 4.8 Hz, 1H), 3.24 (s, 3H), 2.50 – 2.45 (m, 1H), 2.33 (dd, J = 14.9, 7.8 Hz, 1H), 1.36 (s, 9H), 1.18 (s, 9H). **PKS2242**: Synthesized by following the general procedure for *boc*-deprotection of **PKS2237** (34.4 mg, 0.065 mmol). Yield = 32 mg, 91%. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.76 (d, J = 7.7 Hz, 1H), 8.67 (t, J = 5.8 Hz, 1H), 8.14 (b, 3H), 8.04 – 8.02 (m, 1H), 7.95 – 7.93 (m, 1H), 7.88 (s, 1H), 7.85 – 7.83 (m, 1H), 7.56 – 7.52 (m, 2H), 7.47 – 7.43 (m, 2H), 4.80 – 4.72 (m, 2H), 4.53 (ddd, J = 7.7, 5.9, 4.6 Hz, 1H), 4.16 – 4.13 (m, 1H), 3.63 (dd, J = 9.8, 5.9 Hz, 1H), 3.53 (dd, J = 9.8, 4.6 Hz, 1H), 3.26 (s, 3H), 2.73 (dd, J = 16.7, 5.2 Hz, 1H), 2.60 – 2.55 (m, 1H), 1.21 (s, 9H). LCMS calc. for C<sub>23</sub>H<sub>33</sub>N<sub>4</sub>O<sub>4</sub> [M+H]<sup>+</sup>: 429.2. Found 429.5.

**PKS2244:** Synthesized following the general procedure for HATU mediated coupling of 5methylisoxazole-3-carboxylic acid (5.6 mg, 0.044 mmol) and **PKS2242** (21.7 mg, 0.04 mmol). Isolated crude was purified by HPLC to give pure product (18.6 mg, 87%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.62 (t, J = 5.8 Hz, 1H), 8.56 (d, J = 8.0 Hz, 1H), 8.25 (d, J = 7.8Hz, 1H), 8.05 – 8.03 (m, 1H), 7.95 – 7.93 (m, 1H), 7.85 – 7.82 (m, 1H), 7.54 – 7.51 (m, 3H), 7.45 – 7.43 (m, 2H), 6.54 (s, 1H), 4.80 – 4.74 (m, 3H), 4.50 – 4.46 (m, 1H), 3.60 (dd, J =9.8, 5.9 Hz, 1H), 3.53 (dd, J = 9.8, 4.9 Hz, 1H), 3.23 (s, 3H), 2.58 (d, J = 6.7 Hz, 2H), 2.47 (s, 3H), 1.16 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ ) & 171.4, 170.4, 169.2, 168.9, 158.5, 158.3, 134.2, 133.2, 130.7, 128.4, 127.4, 126.1, 125.7, 125.4, 125.0, 123.3, 101.3, 71.8, 58.2, 53.1, 50.2, 50.1, 40.3, 38.1, 28.3, 11.8. HRMS calc. for C<sub>28</sub>H<sub>35</sub>N<sub>5</sub>O<sub>6</sub>Na [M+Na]<sup>+</sup>: 560.2480. Found: 560.2478.

**PKS2231:** Synthesized by following the general procedure of EDC mediated coupling of Boc-Glu-OBn (5.06 g, 15.0 mmol) with *tert*-butylamine (2.37 mL, 22.5 mmol). After completion of reaction, water was added to the mixture. Mixture was extracted twice with ethyl acetate. Combined organic layer was washed with aq. NaHCO<sub>3</sub>, water, 1N HCl, water followed by saturated brine solution. Ethyl acetate layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to give product (5.78 g, 98%) as white solid. The product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*) & 7.43 – 7.29 (m, 5H), 5.58 (s, 1H), 5.27 (d, J= 8.3 Hz, 1H), 5.20 (d, J= 12.3 Hz, 1H), 5.13 (d, J= 12.3 Hz, 1H), 4.36 – 4.23 (m, 1H), 2.22 – 2.06 (m, 3H), 2.02 – 1.87 (m, 1H), 1.42 (s, 9H), 1.32 (s, 9H).

**PKS2233:** Synthesized by following the general procedure for boc-deprotection of **PKS2231** (3.68 g, 9.38 mmol). Crude was dried under high vacuum to give product (3.81 g, quant.) as colorless paste. Product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  8.44 (s, 3H), 7.42 – 7.29 (m, 5H), 6.07 (b, 1H), 5.27 (d, *J* = 11.9 Hz, 1H), 5.20 (d, *J* = 11.9 Hz, 1H), 4.24 – 4.16 (m, 1H), 2.53 – 2.44 (m, 1H), 2.43 – 2.34 (m, 1H), 2.34 – 2.24 (m, 1H), 2.24 – 2.13 (m, 1H), 1.30 (d, *J* = 2.1 Hz, 9H).

**PKS2234:** Synthesized by following the general procedure for HATU mediated coupling of 3-phenylpropanoic acid (82.6 mg, 0.55 mmol) and **PKS2233** (203.2 mg, 0.5 mmol). Crude was purified by column chromatography to give product (193 mg, 91%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ) & 8.28 (d, J = 7.4 Hz, 1H), 7.41 – 7.30 (m, 5H), 7.30 – 7.21 (m, 3H), 7.21 – 7.13 (m, 3H), 5.13 (d, J = 12.6 Hz, 1H), 5.10 (d, J = 12.6 Hz, 1H), 4.31 – 4.20 (m, 1H), 2.80 – 2.76 (m, 2H), 2.45 – 2.38 (m, 2H), 2.07 (t, J = 7.7 Hz, 2H), 1.97 – 1.84 (m, 1H), 1.79 – 1.67 (m, 1H), 1.22 (s, 9H).

**PKS2239:** Synthesized by following the general procedure for *O*-debenzylation of **PKS2234** (180 mg, 0.34 mmol). The product was isolated as a white solid (140 mg, 99%). <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  12.58 (s, 1H), 8.10 (d, J = 7.7 Hz, 1H), 7.37 (s, 1H), 7.27 – 7.13 (m, 5H), 4.19 – 4.11 (m, 1H), 2.81 (t, J = 7.9 Hz, 2H), 2.49 – 2.36 (m, 2H), 2.10 – 1.98 (m, 2H), 1.96 – 1.85 (m, 1H), 1.76 – 1.63 (m, 1H), 1.23 (s, 9H). LCMS calc. for C<sub>18</sub>H<sub>25</sub>N<sub>2</sub>O<sub>4</sub> [M–H]<sup>+</sup>: 333.2. Found: 333.4.

**PKS2243:** Synthesized by following the general procedure for HATU mediated coupling of **PKS2239** (20.1 mg, 0.06 mmol) and **PKS2082** (22.3 mg, 0.06 mmol). The crude was purified by HPLC to give product (18.8 mg, 54%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.55 (t, J = 5.8 Hz, 1H), 8.10 – 8.00 (m, 3H), 7.96 – 7.92 (m, 1H), 7.83 (dd, J = 6.1, 3.4 Hz, 1H), 7.56 – 7.50 (m, 2H), 7.47 – 7.43 (m, 2H), 7.35 (s, 1H), 7.28 – 7.23 (m, 2H), 7.21 – 7.13 (m, 3H), 4.75 (d, J = 5.7 Hz, 2H), 4.51 (dt, J = 7.7, 5.6 Hz, 1H), 3.24 (s, 3H), 2.79 (t, J = 7.9 Hz, 2H), 2.48 – 2.37 (m, 2H), 2.03 (t, J = 8.0 Hz, 2H), 1.92 – 1.78 (m, 1H), 1.77 – 1.60 (m, 1H), 1.22 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  171.6, 171.5, 171.2, 169.4, 141.3, 134.1, 133.2, 130.7, 128.4, 128.2, 128.1, 127.4, 126.1, 125.8, 125.7, 125.4, 124.9, 123.4, 71.9, 58.2, 52.6, 52.3, 49.8, 40.3, 36.8, 32.6, 31.0, 28.5, 28.3. HRMS calc. for C<sub>32</sub>H<sub>42</sub>N<sub>4</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup>: 597.3047. Found: 597.3027.

**PKS2255**: Synthesized by following the general procedure for HATU mediated coupling of **PKS2239** (18.4 mg, 0.055 mmol) and **PKS2026** (17.1 mg, 0.05 mmol). The crude was purified by HPLC to give product (20.2 mg, 74%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.45 (t, J = 5.7 Hz, 1H), 8.10 – 8.01 (m, 3H), 7.98 – 7.90 (m, 1H), 7.84 (dd, J = 7.5, 1.9 Hz, 1H), 7.58 – 7.50 (m, 2H), 7.50 – 7.41 (m, 2H), 7.35 (s, 1H), 7.29 – 7.22 (m, 2H), 7.22 – 7.11 (m, 3H), 4.76 (dd, J = 14.2, 4.6 Hz, 1H), 4.73 (dd, J = 14.2, 4.5 Hz, 1H), 4.37 – 4.27 (m, 1H), 4.27 – 4.18 (m, 1H), 2.79 (t, J = 7.9 Hz, 2H), 2.47 – 2.36 (m, 2H), 2.09 – 2.00 (m, 2H), 1.90 – 1.79 (m, 1H), 1.75 – 1.63 (m, 1H), 1.24 (d, J = 7.1 Hz, 3H), 1.22 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  172.1, 171.6, 171.2, 171.2, 141.3, 134.4, 133.2, 130.8, 128.5, 128.2, 128.2, 127.5, 126.2, 125.8, 125.8, 125.4, 125.1, 123.4, 52.3, 49.8, 48.3, 40.2, 36.7, 32.6, 31.0, 28.5, 28.3, 18.2. HRMS calc. for C<sub>32</sub>H<sub>40</sub>N<sub>4</sub>O<sub>4</sub>Na [M+Na]<sup>+</sup>: 567.2942. Found: 567.2937.

**PKS21201:** Boc-Ala-OSu (143 mg, 0.5 mmol) was dissolved in dichloromethane (5 mL) and (4-fluoro-1-naphthyl)methanamine (88 mg, 0.5 mmol) was added. The solution was cooled to 0 °C and triethylamine (51 mg, 0.5 mmol) was added. The reaction mixture was allowed to warm to room temperature and stirred at room temperature for 2 hours. After completion of reaction, dichloromethane was evaporated and water was added. Mixture was extracted with ethyl acetate twice. Combined organic layer was washed with 1N HCl followed by brine and dried over anhydrous sodium sulfate. Organic layer was evaporated to give product (170 mg, 98%) as white solid. Product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  8.16 – 8.09 (m, 1H), 7.96 (d, *J* = 7.8 Hz, 1H), 7.62 – 7.52 (m, 2H), 7.35 (dd, *J* = 7.8, 5.4 Hz, 1H), 7.07 (dd, *J* = 10.2, 7.9 Hz, 1H), 6.48 (b, 1H), 4.95 (b, 1H), 4.88 – 4.76 (m, 2H), 4.21 – 4.06 (m, 1H), 1.36 (d, *J* = 7.0 Hz, 3H), 1.33 (s, 9H). LCMS calc. for C<sub>19</sub>H<sub>24</sub>FN<sub>2</sub>O<sub>3</sub> [M+H]<sup>+</sup>: 347.2. Found 347.1.

**PKS21202:** Synthesized by following general procedure for boc-deprotection of **PKS21201**. Crude was dried under vacuum to give colorless paste (which turned off-white solid upon standing overnight at 0 °C). Product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO- $d_{d}$ )  $\delta$  8.89 (t, J= 5.6 Hz, 1H), 8.16 – 8.06 (m, 5H), 7.72 – 7.64 (m, 2H), 7.47 (dd, J= 7.9, 5.5 Hz, 1H), 7.32 (dd, J= 10.6, 7.9 Hz, 1H), 4.80 (dd,

J= 15.1, 5.6 Hz, 1H), 4.75 (dd, J= 15.1, 5.5 Hz, 1H), 3.92 – 3.80 (m, 1H), 1.35 (d, J= 6.9 Hz, 3H). LCMS calc. for C<sub>14</sub>H<sub>16</sub>FN<sub>2</sub>O [M+H]<sup>+</sup>: 247.1. Found 247.2.

**PKS21210:** Synthesized by following the general procedure for HATU mediated coupling of **PKS2239** (16.7 mg, 0.05 mmol) and **PKS21202** (18.0 mg, 0.05 mmol). Isolated mixture was purified by HPLC to give product (19.6 mg, 70%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_{o}$ )  $\delta$  8.44 (t, J = 5.6 Hz, 1H), 8.12 – 8.05 (m, 3H), 8.03 (d, J = 7.7 Hz, 1H), 7.68 – 7.62 (m, 2H), 7.42 (dd, J = 7.8, 5.6 Hz, 1H), 7.35 (s, 1H), 7.30 – 7.23 (m, 3H), 7.21 – 7.13 (m, 3H), 4.71 (d, J = 5.6 Hz, 2H), 4.34 – 4.26 (m, 1H), 4.25 – 4.17 (m, 1H), 2.78 (t, J = 7.8 Hz, 2H), 2.48 – 2.36 (m, 2H), 2.03 (t, J = 8.0 Hz, 2H), 1.89 – 1.78 (m, 1H), 1.74 – 1.62 (m, 1H), 1.28 – 1.20 (m, 12H). <sub>13</sub>C NMR (126 MHz, DMSO-  $d_{o}$ )  $\delta$  172.2, 171.6, 171.2, 171.2, 157.4 (d, J = 248.8 Hz), 141.3, 132.0 (d, J = 3.7 Hz), 130.8 (d, J = 3.2 Hz), 128.2, 128.1, 127.3, 126.6, 125.8, 125.2 (d, J = 8.9 Hz), 123.8, 122.9 (d, J = 16.3 Hz), 120.4 (d, J = 5.3 Hz), 108.9 (d, J = 19.8 Hz), 52.3, 49.8, 48.3, 39.8, 36.7, 32.6, 31.0, 28.5, 28.3, 18.2. HRMS calc. for C<sub>32</sub>H<sub>39</sub>FN<sub>4</sub>O<sub>4</sub>Na [M+Na]<sup>+</sup>: 585.2848. Found: 585.2843.

**PKS3053:** Synthesized by following the general procedure for HATU mediated coupling of **PKS3052** (7.2 mg, 0.02 mmol) and **PKS2026** (7.5 mg, 0.022 mmol). The mixture was purified by HPLC to give product (6.4 mg, 56%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  12.26 (d, J = 3.4 Hz, 1H), 8.77 (d, J = 3.2 Hz, 1H), 8.71 (d, J = 8.1 Hz, 1H), 8.52 (t, J = 5.8 Hz, 1H), 8.30 – 8.21 (m, 2H), 8.08 – 8.02 (m, 1H), 7.97 – 7.91 (m, 1H), 7.86 – 7.80 (m, 1H), 7.57 (s, 1H), 7.56 – 7.49 (m, 3H), 7.47 – 7.41 (m, 2H), 7.31 – 7.24 (m, 2H), 4.75 (d, J = 5.8 Hz, 2H), 4.70 – 4.60 (m, 1H), 4.38 – 4.25 (m, 1H), 2.66 – 2.54 (m, 2H), 1.27 (d, J = 7.1 Hz, 3H), 1.17 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  181.1, 171.9, 170.0, 169.0, 162.8, 138.6, 136.2, 134.4, 133.2, 130.8, 128.5, 127.5, 126.1, 125.7, 125.4, 125.1, 123.5, 123.3, 122.6, 121.3, 112.6, 112.1, 50.2, 50.1, 48.7, 40.2, 38.1, 28.4, 18.1. HRMS calc. for C<sub>32</sub>H<sub>35</sub>N<sub>5</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup>: 592.2530. Found: 592.2508.

**PKS21271:** Synthesized by following the general procedure for HATU mediated coupling of **PKS21013** (16.0 mg, 0.05 mmol) and **PKS21202** (18.0 mg, 0.05 mmol). Isolated mixture was purified by HPLC to give product (21.6 mg, 79%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_{6}$ ) & 8.57 (t, J = 6.1 Hz, 1H), 8.26 (d, J = 7.4 Hz, 1H), 8.16 – 8.04 (m, 3H), 7.69 – 7.61 (m, 2H), 7.53 (s, 1H), 7.46 – 7.39 (m, 1H), 7.29 – 7.22 (m, 3H), 7.21 – 7.12 (m, 3H), 4.76 (dd, J = 15.6, 5.9 Hz, 1H), 4.64 (dd, J = 15.6, 5.5 Hz, 1H), 4.55 – 4.47 (m, 1H), 4.29 – 4.18 (m, 1H), 2.78 (t, J = 8.0 Hz, 2H), 2.56 – 2.50 (m, 1H), 2.41 (t, J = 8.0 Hz, 2H), 2.35 (dd, J = 15.3, 6.1 Hz, 1H), 1.25 (d, J = 6.7 Hz, 3H), 1.12 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_{6}$ ) & 172.0, 171.3, 170.8, 169.3, 157.3 (d, J = 249.2 Hz), 141.3, 132.0 (d, J = 5.0 Hz), 130.8 (d, J = 5.3 Hz), 128.3, 128.2, 127.3, 126.5, 125.9, 125.1 (d, J = 8.7 Hz), 123.8, 122.9 (d, J = 16.3 Hz), 120.4 (d, J = 5.4 Hz), 108.8 (d, J = 19.2 Hz), 50.1, 49.8, 48.7, 39.8, 38.4, 36.7, 31.0, 28.3, 17.8. HRMS calc. for C<sub>31</sub>H<sub>37</sub>FN<sub>4</sub>O<sub>4</sub>Na [M+Na]<sup>+</sup>: 571.2691. Found: 571.2689.

**PKS21237:** Synthesized by following general procedure for HATU mediated amide bond formation of *boc*- $\beta$ -methoxyalanine dicyclohexylamine (401 mg, 1 mmol) with (4-fluoro-1-naphthyl)methanamine (175 mg, 1 mmol). Crude was isolated by ethyl acetate extraction

and purified by combi-flash to give product (337 mg, 90%) as off-white solid. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*) δ 8.16 – 8.08 (m, 1H), 7.95 (d, *J* = 7.8 Hz, 1H), 7.61 – 7.51 (m, 2H), 7.35 (dd, *J* = 7.9, 5.3 Hz, 1H), 7.11 – 7.01 (m, 1H), 6.71 (t, *J* = 5.6 Hz, 1H), 5.47 – 5.25 (m, 1H), 4.94 – 4.76 (m, 2H), 4.34 – 4.18 (m, 1H), 3.81 (dd, *J* = 9.3, 3.8 Hz, 1H), 3.51 – 3.43 (m, 1H), 3.27 (s, 3H), 1.37 (s, 9H).

**PKS21238:** Synthesized by following general procedure for boc-deprotection of **PKS21237** (335 mg, 0.9 mmol). Isolated crude was dried under vacuum to give product (347 mg, quant.) as off-white solid. Product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_6$ )  $\delta$  8.95 (b, 1H), 8.23 (b, 3H), 8.14 – 8.06 (m, 2H), 7.72 – 7.64 (m, 2H), 7.51 – 7.44 (m, 1H), 7.32 (dd, J= 10.3, 8.2 Hz, 1H), 4.82 (dd, J= 15.2, 5.7 Hz, 1H), 4.75 (dd, J= 15.2, 5.4 Hz, 1H), 4.09 – 3.98 (m, 1H), 3.72 – 3.59 (m, 2H), 3.27 (s, 3H). LCMS calc. for C<sub>15</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub> [M+H]<sup>+</sup>: 277.1. Found 277.7.

**PKS21218:** Synthesized by following the general procedure for HATU mediated coupling of 5-methylisoxazole-3-carboxylic acid (720 mg, 3.00 mmol) and **PKS2233** (880 mg, 3.00 mmol). After completion of reaction (2 h), water was added and mixture was stirred for 30 minutes. A white precipitate appeared. Precipitate was filtered, washed with water and dried in air to give product (1.03 g, 86%) as white solid.<sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.58 (d, *J* = 7.9 Hz, 1H), 7.39 – 7.30 (m, 5H), 6.41 (s, 1H), 5.47 (s, 1H), 5.20 (s, 2H), 4.80 – 4.72 (m, 1H), 2.47 (s, 3H), 2.35 – 2.25 (m, 1H), 2.23 – 2.04 (m, 3H), 1.31 (s, 9H). LCMS calc. for C<sub>21</sub>H<sub>28</sub>N<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 402.2. Found 402.3.

**PKS21233:** Synthesized by following the general procedure for *O*-debenzylation of **PKS21218** (1.02 g, 2.54 mmol). The product (780 mg, 99%) was isolated as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{\hat{o}}$ )  $\delta$  12.71 (s, 1H), 8.78 (d, *J* = 7.6 Hz, 1H), 7.41 (s, 1H), 6.55 (s, 1H), 4.35 – 4.27 (m, 1H), 2.47 (s, 3H), 2.16 – 2.10 (m, 2H), 2.10 – 2.00 (m, 1H), 1.97 – 1.86 (m, 1H), 1.21 (s, 9H). LCMS calc. for C<sub>14</sub>H<sub>20</sub>N<sub>3</sub>O<sub>5</sub> [M–H]<sup>+</sup>: 310.1. Found: 310.4.

**PKS21213:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS21178** (14.9 mg, 0.05 mmol) with **PKS21202** (18.0 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by HPLC to give product (18.6 mg, 71%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{o}$ )  $\delta$  8.54 (d, J = 7.9 Hz, 1H), 8.50 (t, J = 5.8 Hz, 1H), 8.27 (d, J = 7.3 Hz, 1H), 8.16 – 8.05 (m, 2H), 7.69 – 7.59 (m, 2H), 7.54 (s, 1H), 7.42 (dd, J = 7.8, 5.7 Hz, 1H), 7.27 (dd, J = 10.5, 8.0 Hz, 1H), 6.53 (s, 1H), 4.75 – 4.63 (m, 3H), 4.34 – 4.23 (m, 1H), 2.57 (d, J = 6.7 Hz, 2H), 2.47 (s, 3H), 1.25 (d, J = 7.1 Hz, 3H), 1.15 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_{o}$ )  $\delta$  171.9, 171.3, 170.0, 169.0, 158.5, 158.3, 157.3 (d, J = 249.3 Hz), 132.0 (d, J = 5.0 Hz), 130.8 (d, J = 5.2 Hz), 127.3, 126.5, 125.2 (d, J = 8.8 Hz), 123.8, 122.9 (d, J = 16.0 Hz), 120.4 (d, J = 5.9 Hz), 108.9 (d, J = 19.9 Hz), 101.3,50.2, 50.1, 48.7, 39.8, 38.0, 28.3, 18.0, 11.8. HRMS calc. for C<sub>27</sub>H<sub>32</sub>FN<sub>5</sub>O<sub>5</sub>Na [M +Na]<sup>+</sup>: 548.2280. Found: 548.2267.

**PKS21272:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS21233** (15.6 mg, 0.05 mmol) with **PKS21202** (18.0 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by HPLC to give product (20.7 mg, 77%)

as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_6$ )  $\delta$  8.54 (d, J = 7.7 Hz, 1H), 8.50 (t, J = 6.0 Hz, 1H), 8.26 (d, J = 7.3 Hz, 1H), 8.12 – 8.04 (m, 2H), 7.69 – 7.60 (m, 2H), 7.45 – 7.37 (m, 2H), 7.32 – 7.24 (m, 1H), 6.54 (s, 1H), 4.76 – 4.65 (m, 2H), 4.45 – 4.38 (m, 1H), 4.38 – 4.29 (m, 1H), 2.46 (s, 3H), 2.18 – 2.05 (m, 2H), 2.03 – 1.93 (m, 1H), 1.92 – 1.81 (m, 1H), 1.24 (d, J = 7.1 Hz, 3H), 1.21 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_6$ )  $\delta$  172.0, 171.2, 171.2, 170.4, 158.5, 158.5, 157.4 (d, J = 248.8 Hz), 132.0 (d, J = 5.3 Hz), 130.8 (d, J = 5.3 Hz), 127.4, 126.6, 125.2 (d, J = 10.2 Hz), 123.8, 122.9 (d, J = 16.3 Hz), 120.4 (d, J = 5.4 Hz), 108.9 (d, J = 19.5 Hz), 101.3, 52.7, 49.9, 48.3, 39.8, 32.6, 28.5, 27.9, 18.3, 11.8. HRMS calc. for C<sub>28</sub>H<sub>34</sub>FN<sub>5</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup>: 562.2436. Found: 562.2433.

**PKS21244:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS21178** (14.9 mg, 0.05 mmol) with **PKS21238** (19.5 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by preparative LCMS to give product (19.7 mg, 71%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{6}$ )  $\delta$  8.62 (t, J = 5.9 Hz, 1H), 8.55 (d, J = 7.9 Hz, 1H), 8.25 (d, J = 7.7 Hz, 1H), 8.13 – 8.04 (m, 2H), 7.68 – 7.60 (m, 2H), 7.53 (s, 1H), 7.46 – 7.38 (m, 1H), 7.31 – 7.22 (m, 1H), 6.53 (s, 1H), 4.81 – 4.74 (m, 1H), 4.74 – 4.66 (m, 2H), 4.50 – 4.41 (m, 1H), 3.63 – 3.55 (m, 1H), 3.52 (dd, J = 10.2, 4.7 Hz, 1H), 3.22 (s, 3H), 2.57 (d, J = 6.7 Hz, 2H), 2.47 (s, 3H), 1.15 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_{6}$ )  $\delta$  171.4, 170.4, 169.2, 168.9, 158.5, 158.3, 157.3 (d, J = 249.3 Hz), 131.9 (d, J = 4.4 Hz), 130.6 (d, J = 3.1 Hz), 127.3, 126.5, 125.1 (d, J = 8.6 Hz), 123.8, 122.9 (d, J = 16.4 Hz), 120.4 (d, J = 5.4 Hz), 108.8 (d, J = 19.7 Hz), 101.3, 71.8, 58.3, 53.2, 50.2, 50.1, 40.0, 38.1, 28.3, 11.8. HRMS calc. for C<sub>28</sub>H<sub>34</sub>FN<sub>5</sub>O<sub>6</sub>Na [M+Na]<sup>+</sup>: 578.2385. Found: 578.2377.

**PKS21245**: Synthesized following general procedure for HATU mediated amide bond formation of **PKS21233** (15.6 mg, 0.05 mmol) with **PKS21238** (19.5 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by preparative LCMS to give product (22.5 g, 79%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_0$ )  $\delta$  8.63 – 8.57 (m, 1H), 8.56 (d, J = 7.8 Hz, 1H), 8.25 (d, J = 7.8 Hz, 1H), 8.11 – 8.03 (m, 2H), 7.69 – 7.60 (m, 2H), 7.45 – 7.34 (m, 2H), 7.28 (dd, J = 10.6, 8.1 Hz, 1H), 6.54 (s, 1H), 4.72 (d, J = 5.6 Hz, 2H), 4.56 – 4.49 (m, 1H), 4.49 – 4.40 (m, 1H), 3.57 – 3.45 (m, 2H), 3.23 (s, 3H), 2.46 (s, 3H), 2.17 – 2.03 (m, 2H), 2.02 – 1.91 (m, 1H), 1.91 – 1.77 (m, 1H), 1.21 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_0$ )  $\delta$  171.2, 171.1, 170.8, 169.3, 158.5, 158.5, 157.4 (d, J = 248.9 Hz), 131.9, 130.5, 127.3, 126.5, 125.1 (d, J = 8.6 Hz), 123.8, 122.9 (d, J = 16.1 Hz), 120.4, 108.9 (d, J = 19.1 Hz), 101.3, 71.9, 58.2, 52.8, 52.6, 49.9, 39.9, 32.6, 28.5, 27.9, 11.8. HRMS calc. for C<sub>29</sub>H<sub>36</sub>FN<sub>5</sub>O<sub>6</sub>Na [M+Na]<sup>+</sup>: 592.2542. Found: 592.2537.

**PKS2238: PKS2233** (203.0 mg, 0.5 mmol) was dissolved in dichloromethane (6 mL) and triethylamine (140  $\mu$ L, 1.0 mmol) was added. After stirring the mixture for 10 minutes at room temperature, 4-toluenesulfonyl chloride (143.0 mg, 0.75 mmol) was added. The reaction mixture was stirred 2 hours at room temperature. Dichloromethane was evaporated and the crude was dissolved in ethyl acetate. The solution was washed with water, 1N HCl followed by brine. The product was purified by column chromatography to give product (177.0 mg, 79%) as white solid. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.66 (d, *J* = 8.5 Hz, 2H), 7.35 – 7.29 (m, 3H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.18 – 7.13 (m, 2H), 5.52 (s, 1H), 5.47 (d, *J* = 9.1 Hz, 1H), 4.91 (d, *J* = 12.2 Hz, 1H), 4.87 (d, *J* = 12.2 Hz, 1H), 3.93 – 3.84 (m, 1H),

2.38 (s, 3H), 2.33 – 2.11 (m, 3H), 1.88 – 1.77 (m, 1H), 1.35 (s, 9H). LCMS calc. for  $C_{23}H_{32}N_2O_5S$  [M+H]<sup>+</sup>: 447.2. Found 447.3.

**PKS2254:** Synthesized by following the general procedure for *O*-debenzylation of **PKS2238** (170 mg, 0.38 mmol). The product (135 mg, quant.) was isolated as white solid. <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.69 (d, *J* = 7.9 Hz, 2H), 7.26 (d, *J* = 7.9 Hz, 2H), 6.15 – 5.91 (m, 2H), 3.82 – 3.69 (m, 1H), 2.39 (s, 3H), 2.35 – 2.27 (m, 1H), 2.21 – 2.12 (m, 1H), 2.11 – 2.02 (m, 1H), 1.99 – 1.88 (m, 1H), 1.31 (s, 9H). LCMS calc. for C<sub>16</sub>H<sub>23</sub>N<sub>2</sub>O<sub>5</sub>S [M–H]<sup>+</sup>: 355.1. Found: 355.0.

**PKS2256**: Synthesized by following the general procedure for HATU mediated coupling of **PKS2254** (29.0 mg, 0.08 mmol) and **PKS2026** (27.4 mg, 0.08 mmol). The crude was purified by HPLC to give product (32.8 mg, 74%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  8.43 (t, J = 5.7 Hz, 1H), 8.09 (d, J = 7.3 Hz, 1H), 8.04 – 7.98 (m, 1H), 7.97 – 7.90 (m, 1H), 7.86 (b, 1H), 7.84 (d, J = 8.1 Hz, 1H), 7.63 (d, J = 8.3 Hz, 2H), 7.58 – 7.49 (m, 2H), 7.49 – 7.42 (m, 1H), 7.42 – 7.39 (m, 1H), 7.38 (s, 1H), 7.30 (d, J = 7.9 Hz, 2H), 4.75 (dd, J = 15.4, 5.8 Hz, 1H), 4.69 (dd, J = 15.4, 5.6 Hz, 1H), 4.11 – 4.01 (m, 1H), 3.74 – 3.67 (m, 1H), 2.34 (s, 3H), 2.11 – 1.93 (m, 2H), 1.77 – 1.66 (m, 1H), 1.66 – 1.56 (m, 1H), 1.21 (s, 9H), 1.07 (d, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (126 MHz, DMSO- $d_6$ )  $\delta$  171.8, 171.0, 169.9, 142.4, 138.0, 134.3, 133.2, 130.8, 129.2, 128.5, 127.5, 126.7, 126.2, 125.8, 125.4, 125.1, 123.4, 55.8, 49.8, 48.1, 40.1, 32.3, 29.2, 28.5, 20.9, 18.1. LCMS calc. for C<sub>30</sub>H<sub>39</sub>N<sub>4</sub>O<sub>5</sub>S [M+H]<sup>+</sup>: 567.3. Found: 567.1.

**PKS21206**: Synthesized following general procedure for HATU mediated amide bond formation of **PKS21241** (17.1 mg, 0.05 mmol) with **PKS21202** (18.0 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by HPLC to give product (24.0 mg, 84%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_0$ )  $\delta$  8.46 (t, J = 6.0 Hz, 1H), 8.21 (d, J = 7.2 Hz, 1H), 8.12 – 8.03 (m, 2H), 8.03 – 7.93 (m, 1H), 7.68 – 7.60 (m, 4H), 7.54 (s, 1H), 7.36 (dd, J = 7.9, 5.6 Hz, 1H), 7.32 (d, J = 7.9 Hz, 2H), 7.23 (dd, J = 10.6, 7.9 Hz, 1H), 4.72 (dd, J = 15.5, 6.1 Hz, 1H), 4.59 (dd, J = 15.5, 5.6 Hz, 1H), 4.05 – 3.95 (m, 1H), 3.91 – 3.81 (m, 1H), 2.49 – 2.45 (m, 1H), 2.35 (s, 3H), 2.29 (dd, J = 15.2, 5.5 Hz, 1H), 1.11 – 1.01 (m, 12H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_0$ )  $\delta$  171.8, 169.7, 168.7, 157.3 (d, J = 249.5 Hz), 142.4, 138.3, 131.9 (d, J = 5.2 Hz), 130.7 (d, J = 3.3 Hz), 129.2, 127.3, 126.6, 126.5, 125.0 (d, J = 8.0 Hz), 123.7, 122.9 (d, J = 16.4 Hz), 120.4 (d, J = 6.7 Hz), 108.8 (d, J = 18.4 Hz), 53.0, 50.1, 48.6, 39.7, 39.2, 28.2, 20.9, 17.4. HRMS calc. for C<sub>29</sub>H<sub>35</sub>FN<sub>4</sub>O<sub>5</sub>SNa [M+Na]<sup>+</sup>: 593.2204. Found: 593.2206.

**PKS21209:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS2254** (17.8 mg, 0.05 mmol) with **PKS21202** (18.0 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by HPLC to give product (17.0 mg, 58%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{o}$ )  $\delta$  8.42 (t, J = 5.7 Hz, 1H), 8.12 – 8.02 (m, 3H), 7.90 – 7.78 (m, 1H), 7.70 – 7.57 (m, 4H), 7.42 – 7.33 (m, 2H), 7.32 – 7.21 (m, 3H), 4.74 – 4.62 (m, 2H), 4.08 – 3.97 (m, 1H), 3.74 – 3.63 (m, 1H), 2.34 (s, 3H), 2.09 – 1.91 (m, 2H), 1.76 – 1.65 (m, 1H), 1.65 – 1.51 (m, 1H), 1.20 (s, 9H), 1.05 (d, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_{o}$ )  $\delta$  171.9, 171.1, 170.0, 157.4 (d, J = 248.9 Hz), 142.4, 138.0,

132.0 (d, J= 5.2 Hz), 130.7 (d, J= 5.1 Hz), 129.2, 127.4, 126.7, 126.5, 125.2 (d, J= 9.0 Hz), 123.8, 122.9 (d, J= 16.3 Hz), 120.4 (d, J= 5.4 Hz), 108.9 (d, J= 18.4 Hz), 55.8, 49.8, 48.1, 39.8, 32.3, 29.2, 28.5, 20.9, 18.1. HRMS calc. for C<sub>30</sub>H<sub>37</sub>FN<sub>4</sub>O<sub>5</sub>SNa [M+Na]<sup>+</sup>: 607.2361. Found: 607.2349.

**PKS21265**: Synthesized following general procedure for HATU mediated amide bond formation of **PKS2254** (17.8 mg, 0.05 mmol) with **PKS21238** (19.5 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by preparative LCMS to give product (24.5 g, 80%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_6$ ) & 8.59 – 8.48 (m, 1H), 8.11 (d, J = 7.8 Hz, 1H), 8.09 – 8.02 (m, 2H), 7.89 (s, 1H), 7.69 – 7.59 (m, 4H), 7.43 – 7.35 (m, 2H), 7.32 (d, J = 7.9 Hz, 2H), 7.30 – 7.22 (m, 1H), 4.75 – 4.64 (m, 2H), 4.31 – 4.21 (m, 1H), 3.81 – 3.70 (m, 1H), 3.31 – 3.23 (m, 2H), 3.19 (s, 3H), 2.35 (s, 3H), 2.10 – 2.00 (m, 1H), 2.00 – 1.90 (m, 1H), 1.75 – 1.64 (m, 1H), 1.64 – 1.50 (m, 1H), 1.20 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_6$ ) & 171.1, 170.5, 169.2, 157.4 (d, J = 248.9 Hz), 142.4, 138.0, 131.9 (d, J =5.2 Hz), 130.5 (d, J = 4.7 Hz), 129.3, 127.4, 126.7, 126.6, 125.1 (d, J = 8.8 Hz), 123.8, 122.9 (d, J = 16.3 Hz), 120.4 (d, J = 6.8 Hz), 108.9 (d, J = 20.0 Hz), 71.7, 58.3, 55.8, 52.3, 49.9, 39.9, 32.4, 29.3, 28.5, 21.0. HRMS calc. for C<sub>31</sub>H<sub>39</sub>FN<sub>4</sub>O<sub>6</sub>SNa [M+Na]<sup>+</sup>: 637.2467. Found: 637.2464.

**PKS21266:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS21241** (17.1 mg, 0.05 mmol) with **PKS21238** (19.5 mg, 0.05 mmol). After completion of reaction (1 h), mixture was purified by preparative LCMS to give product (26.8 mg, 89%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{6}$ ) & 8.57 (t, J = 6.0 Hz, 1H), 8.20 (d, J = 7.6 Hz, 1H), 8.12 – 8.03 (m, 2H), 7.93 (d, J = 8.6 Hz, 1H), 7.68 – 7.59 (m, 4H), 7.48 (s, 1H), 7.41 – 7.36 (m, 1H), 7.32 (d, J = 7.9 Hz, 2H), 7.23 (dd, J = 10.8, 8.2 Hz, 1H), 4.73 (dd, J = 15.7, 5.9 Hz, 1H), 4.63 (dd, J = 15.7, 5.5 Hz, 1H), 4.20 – 4.07 (m, 2H), 3.53 – 3.46 (m, 1H), 3.29 – 3.23 (m, 1H), 3.20 (s, 3H), 2.44 (dd, J = 15.6, 8.2 Hz, 1H), 2.36 (s, 3H), 2.26 (dd, J = 15.6, 6.2 Hz, 1H), 1.08 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_{6}$ ) & 170.3, 169.2, 168.5, 157.3 (d, J = 248.8 Hz), 142.4, 138.3, 131.9 (d, J = 3.8 Hz), 130.6 (d, J = 3.3 Hz), 129.3, 127.2, 126.6, 126.5, 125.0 (d, J = 7.5 Hz), 123.8, 122.8 (d, J = 16.2 Hz), 120.3 (d, J = 5.3 Hz), 108.8 (d, J = 19.5 Hz), 71.5, 58.3, 53.3, 52.9, 50.1, 39.9, 39.4, 28.3, 21.0 . HRMS calc. for C<sub>30</sub>H<sub>37</sub>FN<sub>4</sub>O<sub>6</sub>SNa [M+Na]<sup>+</sup>: 623.2310. Found: 623.2307.

**PKS21066:** Synthesized following general procedure for HATU mediated amide bond formation of **PKS2141** (22.8 mg, 0.04 mmol) with *neo*-pentylamine (9.4 µL, 0.08 mmol). After completion of reaction (1 h), mixture was purified by HPLC to give product (17.8 mg, 70%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{o}$ ) & 8.59 (t, J = 5.9 Hz, 1H), 8.21 (d, J = 8.2 Hz, 1H), 8.15 (d, J = 8.0 Hz, 1H), 8.07 (d, J = 7.9 Hz, 1H), 7.94 (d, J = 7.4 Hz, 1H), 7.89 – 7.81 (m, 2H), 7.57 – 7.51 (m, 2H), 7.42 (t, J = 7.6 Hz, 1H), 7.36 (d, J = 6.9 Hz, 1H), 7.30 – 7.13 (m, 7H), 7.04 – 6.95 (m, 2H), 4.78 (dd, J = 15.3, 5.9 Hz, 1H), 4.68 (dd, J = 15.3, 5.6 Hz, 1H), 4.59 – 4.52 (m, 1H), 4.50 – 4.42 (m, 1H), 3.09 (dd, J = 13.9, 4.7 Hz, 1H), 2.87 – 2.79 (m, 2H), 2.77 – 2.70 (m, 3H), 2.55 (dd, J = 15.0, 7.1 Hz, 1H), 2.41 (dd, J = 15.0, 7.2 Hz, 1H), 2.35 (t, J = 8.0 Hz, 2H), 0.76 (s, 9H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_{o}$ ) & 171.3, 171.0, 170.5, 169.7, 160.9 (d, J = 240.3 Hz), 141.3, 134.2, 134.0, 133.2, 131.0 (d, J = 7.7 Hz), 130.8, 128.5, 128.3, 128.1, 127.5, 126.2, 125.8, 125.7, 125.4, 125.3, 123.4, 114.7 (d, J

= 20.2 Hz), 54.3, 49.9, 49.7, 40.3, 37.5, 36.8, 36.2, 31.8, 31.0, 27.2. HRMS calc. for  $C_{38}H_{43}FN_4O_4Na \ [M+Na]^+: 661.3161$ . Found: 661.3154.

**PKS21076:** Synthesized by following general procedure for HATU coupling of Boc-Asn-OH (130 mg, 0.56 mmol) and **PKS2123** (245 mg, 0.56 mmol). After completion of reaction (1 h), water was added. A white precipitate was formed. Precipitate was filtered, washed with water and dried to give product (298 mg, 99%) as white solid. Product was next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_{o}$ ) & 8.55 (d, J = 5.9 Hz, 1H), 8.08 – 7.98 (m, 2H), 7.98 – 7.92 (m, 1H), 7.84 (d, J = 8.1 Hz, 1H), 7.60 – 7.49 (m, 2H), 7.47 – 7.40 (m, 1H), 7.40 – 7.28 (m, 2H), 7.28 – 7.15 (m, 2H), 7.11 – 6.97 (m, 2H), 6.97 – 6.87 (m, 2H), 4.72 (d, J = 5.6 Hz, 2H), 4.58 – 4.43 (m, 1H), 4.27 – 4.10 (m, 1H), 3.02 (dd, J = 13.9, 5.0 Hz, 1H), 2.85 (dd, J = 13.9, 8.9 Hz, 1H), 2.43 (dd, J = 15.3, 5.9 Hz, 1H), 2.32 (dd, J = 15.3, 7.9 Hz, 1H), 1.35 (s, 9H). LCMS calc. for C<sub>29</sub>H<sub>34</sub>FN<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup>: 537.2. Found: 537.4.

**PKS21078**: Synthesized by following general procedure for boc-deprotection of **PKS21076** (298 mg, 0.56 mmol). Isolated crude was triturated with diethyl ether to give a white solid. Diethyl ether was decanted and white solid was dried under high vacuum to give product (300 mg, 98%). Product was used in next step without further purification. <sup>1</sup>H NMR (500 MHz, DMSO-  $d_6$ ) & 8.74 (d, J = 8.0 Hz, 1H), 8.72 – 8.66 (m, 1H), 8.15 – 7.98 (m, 4H), 7.96 (d, J = 6.7 Hz, 1H), 7.86 (d, J = 8.2 Hz, 1H), 7.72 (s, 1H), 7.59 – 7.52 (m, 2H), 7.47 – 7.41 (m, 1H), 7.33 – 7.28 (m, 2H), 7.28 – 7.22 (m, 2H), 7.09 – 7.02 (m, 2H), 4.77 (dd, J = 15.5, 5.7 Hz, 1H), 4.69 (dd, J = 15.5, 5.4 Hz, 1H), 4.60 – 4.52 (m, 1H), 4.09 – 3.98 (m, 1H), 3.00 (dd, J = 14.1, 5.1 Hz, 1H), 2.85 – 2.72 (m, 2H), 2.58 (dd, J = 17.3, 9.0 Hz, 1H). LCMS calc. for C<sub>24</sub>H<sub>26</sub>FN<sub>4</sub>O<sub>3</sub> [M+H]<sup>+</sup>: 437.2. Found: 437.4.

**PKS21083:** Synthesized by following general procedure for HATU mediated amide bond formation of 3-phenylpropanoic acid (8.3 mg, 0.055 mmol) and **PKS21078** (27.5 mg. 0.05 mmol). After completion of reaction (1 h), the mixture was purified by HPLC to give product (21.6 mg, 76%) as white solid. <sup>1</sup>H NMR (500 MHz, DMSO- $d_0$ )  $\delta$  8.61 – 8.49 (m, 1H), 8.16 (d, J = 8.2 Hz, 1H), 8.07 (d, J = 7.9 Hz, 2H), 7.94 (d, J = 7.5 Hz, 1H), 7.84 (d, J = 8.2 Hz, 1H), 7.61 – 7.50 (m, 2H), 7.44 (t, J = 7.6 Hz, 1H), 7.40 (s, 1H), 7.36 (d, J = 7.2 Hz, 1H), 7.29 – 7.22 (m, 2H), 7.24 – 7.13 (m, 5H), 7.03 – 6.95 (m, 2H), 6.94 (s, 1H), 4.76 (dd, J = 15.6, 5.7 Hz, 1H), 4.70 (dd, J = 15.6, 5.7 Hz, 1H), 4.61 – 4.51 (m, 1H), 4.50 – 4.41 (m, 1H), 3.08 (dd, J = 14.3, 4.5 Hz, 1H), 2.83 (dd, J = 13.9, 9.7 Hz, 1H), 2.78 – 2.70 (m, 2H), 2.57 – 2.49 (m, 1H), 2.42 – 2.29 (m, 3H). <sup>13</sup>C NMR (126 MHz, DMSO-  $d_0$ )  $\delta$  171.8, 171.3, 170.9, 170.5, 160.9 (d, J = 240.0 Hz), 141.3, 134.2, 134.0, 133.2, 130.9 (d, J = 6.3 Hz), 130.7, 128.5, 128.3, 128.1, 127.4, 126.2, 125.8, 125.7, 125.4, 125.0, 123.4, 114.7 (d, J = 21.6 Hz), 54.3, 49.5, 40.2, 37.0, 36.7, 36.2, 31.0.<sup>19</sup>F NMR (471 MHz, DMSO- $d_0$ )  $\delta$  –119.31 (m). HRMS calc. for C<sub>33</sub>H<sub>33</sub>FN<sub>4</sub>O<sub>4</sub>Na [M+Na]<sup>+</sup>: 591.2378. Found: 591.2374.

**IC**<sub>50</sub> **Determination.**—IC50 values of all compounds against hu i-20S and c-20S were performed using a 96-well format assay, as described.<sup>37</sup>

**Cell-based proteasome**  $\beta$ **5 activity assay.**—Karpas-1106P (22,000 cells per well) or Hela (3,000 cells per well) were plated in 384-well plate and incubated with compound at

indicated concentrations for 2 h at 37 °C. The activity of the overall β5 activity including β5i and β5c in each was measured *in situ* after compound removal with Proteasome-Glo assay kit according to manufacturer's instructions. Luminescence was recorded on a SpectraMax M5 plate reader. Relative percentage of RLU was used to calculate the IC50s.

**Human PBMC Proliferation Assay.**—IC<sub>50</sub> values of select compounds against T cell proliferation in PBMCs were determined as reported.<sup>37</sup>

**Kinetic assay of PKS21265 vs hu i-20S.**—The assay was performed on a SpectraMax Gemini plate-reader from Molecular Devices (Sunnyvale, CA). 1 µL of PKS21265 at 100x indicated concentrations in DMSO were spotted at the bottom of the wells of a black wall/ clear bottom 96-well plate. 99 µL of assay mixture containing 0.4 nM human i-20S and the indicated concentration of Suc-LLVY-AMC in HEPES buffer (25 mM HEPES, 0.5 mM EDTA, 0.02% Sodium dodecyl sulfate, pH 7.4) were dispensed into the designated wells. The reaction progress of each well was immediately recorded by monitoring fluorescence at Ex 360nm and Em 460 nm for 1.2 h at 37°C. *Ki* value for PKS21225 was derived by fitting data to equation  $V = (V_{\text{max}} / (1 + I / (\alpha * K_i))) * S / (K_{\text{M}} * (1 + I / K_i) / (1 + I / (\alpha * K_i)) + S)$  in Prism (GraphPad Software, Inc. La Jolla, CA).

# **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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# ABBREVIATIONS

i-20S	immunoproteasome
c-20S	constitutive proteasome

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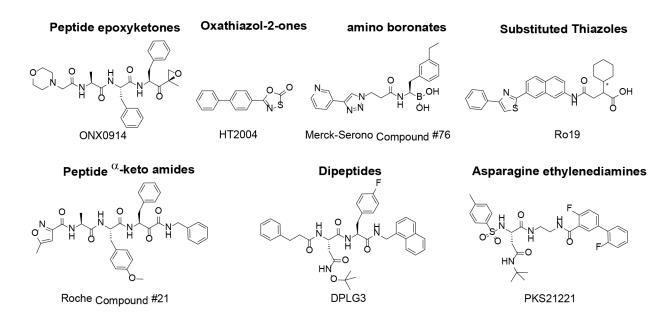
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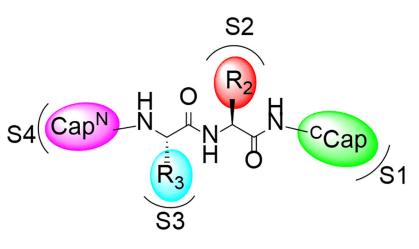
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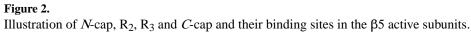
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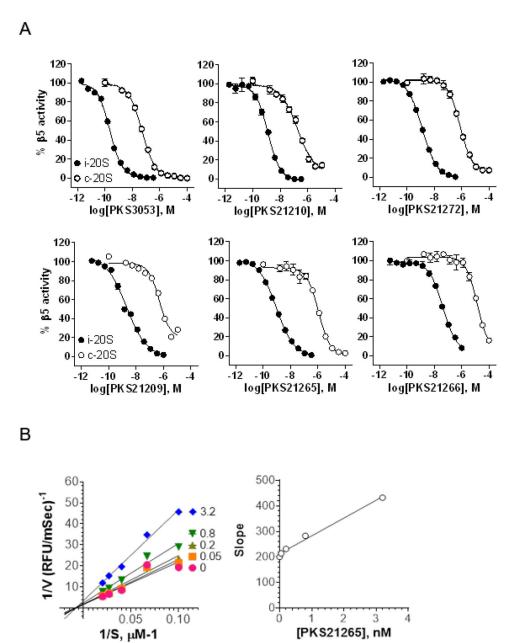


#### Figure 1.

Structures of some reported immunoproteasome inhibitors. ONX0914 was previously named PR957.<sup>24</sup> Merck-Serono  $\beta$ 5i selective inhibitor 76, Ro19 and Roche Compound 21 were reported in a patent review.<sup>17</sup> HT2004, DPLG3 and PKS21221 were recently reported as  $\beta$ 5i selective inhibitors.<sup>35, 36, 38</sup>

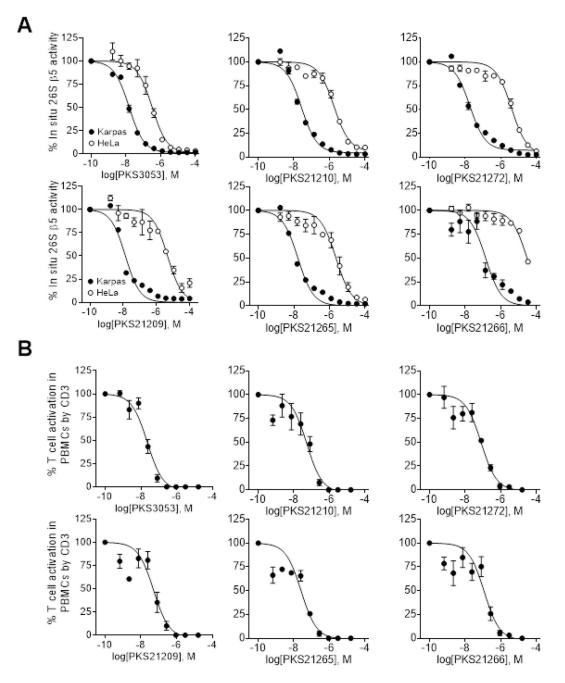






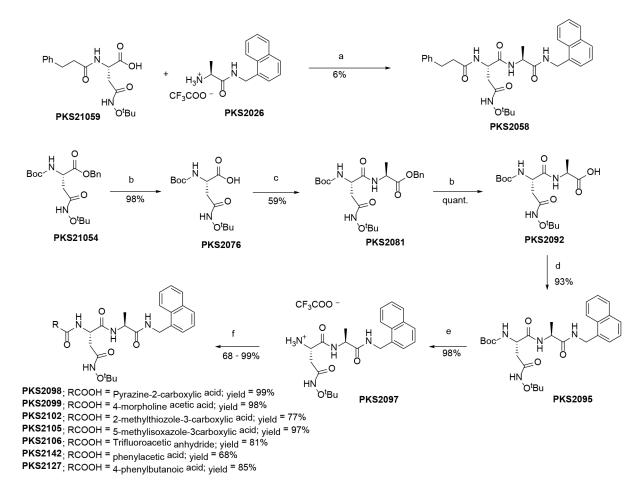
#### Figure 3.

Inhibition of  $\beta$ 5i and  $\beta$ 5c by select dipeptides. A) Dose-dependent inhibition of  $\beta$ 5i and  $\beta$ 5c by compounds indicted in the x-axial. B)  $K_i$  of PKS21265 was also determined with varying concentrations of PKS21265 and substrate suc-LLVY-AMC. Lineweaver-Burk plot was shown on the left. The plot of slopes against inhibitor concentration yield a  $K_i$  3.1 nM.



#### Figure 4.

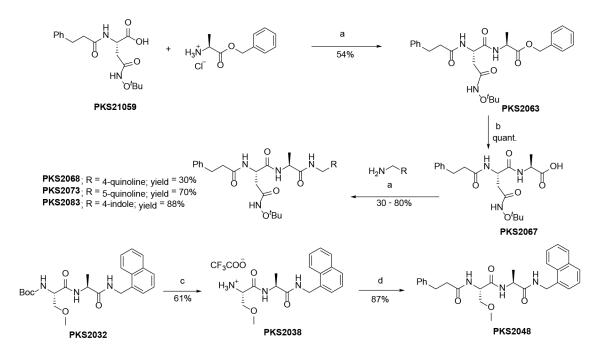
In situ inhibition of i-20S  $\beta$ 5i in Karpas-1106P and c-20S  $\beta$ 5c in Hela cells and dosedependent inhibition of T cell activation in peripheral blood mononuclear cells (PBMCs) stimulated by anti-CD3 antibody. All experiments were repeated three times. Representative results are shown and data summarized in Table 3.



#### Scheme 1.

Synthesis of N-Cap analogs.

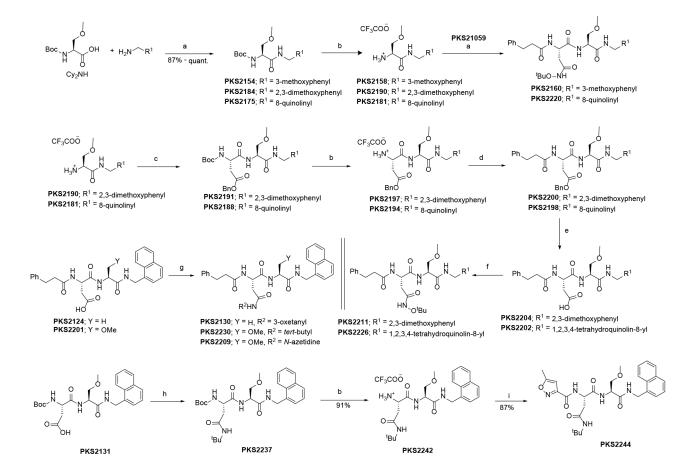
Reagents and conditions: a) EDCI, HOBt, Hünig's base; b) 10% Pd on Carbon, Methanol, H<sub>2</sub> balloon, 98%; c) *O*-benzylalanine hydrochloride, HATU, HOAt, Hünig's base, DMF, 59%; d) 1-naphthylmethylamine, HATU, HOAt, Hünig's base, DMF; e) 20% TFA in DCM; f) RCOOH, HATU, HOAt, Hünig's base, DMF



#### Scheme 2.

Synthesis of C-cap analogs.

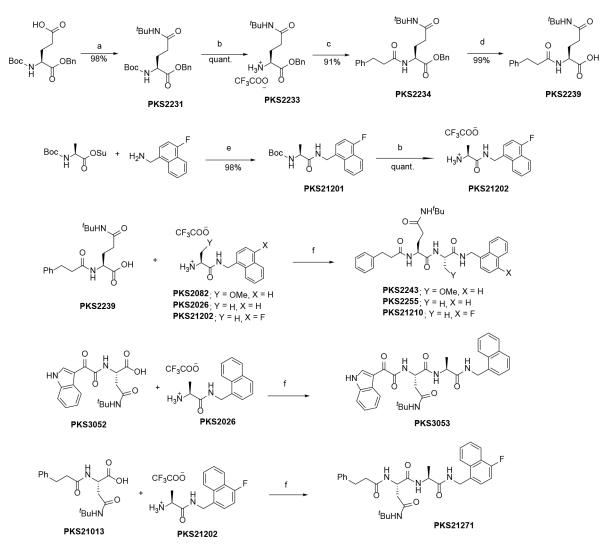
Reagents and conditions: a) HATU, HOAt, Hünig's base, DMF; b) 10% Pd on Carbon,  $H_2$  balloon, EtOH; c) TFA, DCM; d) 3-Phenylpropanoic acid, EDC, HOBt, Hünig's base, DMF



#### Scheme 3.

Synthesis of various P2 (methylserine) analogs.

Reagents and conditions: a) HATU, HOAt, Hünig's base, DMF; b) TFA, DCM; c) BOC-Asp(OBn)-OH, HATU, HOAt, Hünig's base, DMF; d) 3-phenylpropanoic acid, HATU, HOAt, Hünig's base, DMF; e) 10% Pd on carbon, methanol; f) *O*-tert-butylhydroxylamine, HATU, HOAt, Hünig's base, DMF; g) *tert*-butylamine or 3-aminooxitane, HATU, HOAt, Hünig's base, DMF; h) *tert*-butylamine, HATU, HOAt, Hünig's base, DMF; i) 5methylisoxazole-3-carboxylic acid, HATU, HOAt, Hünig's base, DMF.

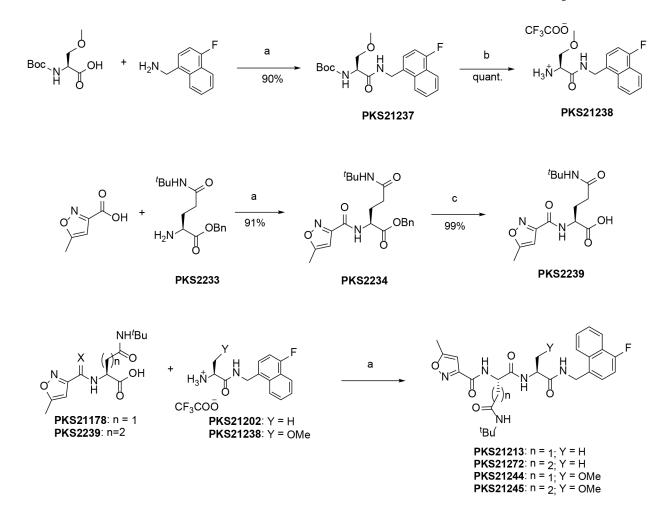


#### Scheme 4.

Reagents and conditions: a) *tert*-butylamine, EDC, HOBt, Hünig's base; b) TFA, DCM; c) 3-phenylpropanoic acid, HATU, HOAt, Hünig's base, DMF; d) 10% Pd on carbon, methanol, hydrogen balloon; e) triethylamine, DCM; f) HATU, HOAt, Hünig's base, DMF

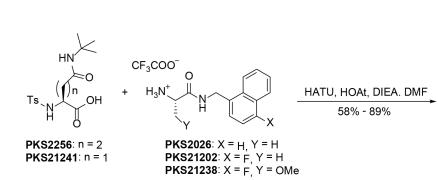
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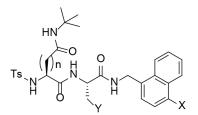
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#### Scheme 5.

Reagents and conditions: a) HATU, HOAt, Hünig's base, DMF; b) TFA, DCM; c) 10% Pd on carbon, methanol, hydrogen balloon





 $\begin{array}{l} \textbf{PKS2256: } n = 2, X = H, Y = H; \ yield = 74\% \\ \textbf{PKS21206: } n = 1, X = F, Y = H; \ yield = 84\% \\ \textbf{PKS21209: } n = 2, X = F, Y = H; \ yield 58\% \\ \textbf{PKS21265: } n = 2, X = F, Y = OMe; \ yield = 80\% \\ \textbf{PKS21266: } n = 1, X = F, Y = OMe; \ yield = 89\% \\ \end{array}$ 

Scheme 6. Synthesis of compounds with *N*-sulfonyl cap

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Table 1.

ID (DVS.)	$\begin{array}{c} H \stackrel{O}{\longrightarrow} R_2 \stackrel{H}{\longrightarrow} R_1 \\ R_4 \stackrel{N}{\longrightarrow} R_3 \stackrel{N}{\longrightarrow} N \stackrel{R_2}{\longrightarrow} R_1 \end{array}$				IC50 (µM)		Selectivity
(PKS-)	R4	R3	R2	R1	β5i	β5c	Index
DPLG3	O C C C		<i>p</i> -F-benzyl	*	0.0045*	32.4*	7200
2086	O		CH <sub>3</sub> OCH <sub>2</sub>	÷	0.0044*	0.3*	68
2058	O C C	HN O	CH <sub>3</sub>	₹\$	0.0057±0.0018	1.32±0.43	230
2068	O José	HN 0-(	CH <sub>3</sub>	+∕N	2.62±0.72	59.03±15.53	23
2073	O J J J J	HN O-	CH <sub>3</sub>	₹	1.16±0.27	12.73±0.35	11
2083	O L L L	HN O	CH <sub>3</sub>	₩	0.0284±0.0077	0.17±0.035	6
2220	O C C	HN O	CH <sub>3</sub> OCH <sub>2</sub>		0.0837±0.0302	1.04±0.159	12
2226	O C C	HN O	CH <sub>3</sub> OCH <sub>2</sub>	₩ HN	0.501±0.328	27.23±25.43	54
2160	O C C	HN O	CH <sub>3</sub> OCH <sub>2</sub>	₹<	0.0144±0.0051	0.101±0.0044	7
2211	O C C C	HN O	CH <sub>3</sub> OCH <sub>2</sub>		0.479±0.223	0.98±0.16	2
2127	O o	HN O	CH <sub>3</sub>	+	0.0197±0.0011	> 100	> 5076
2142	o	HN O	CH <sub>3</sub>	$\overset{+}{\bigcirc}$	0.0259±0.0024	47.67±30.19	1840
2106	O F F F	HN O	CH <sub>3</sub>	$\overset{+}{\bigcirc}$	0.0992±0.014	22.73±8.21	229
2098	O N N	HN O	CH <sub>3</sub>	*	2.812±0.512	> 100	> 35
2099	°⊂N N	HN O	CH <sub>3</sub>	ł	0.424±0.0497	41.08±1.962	104
2102	S N		CH <sub>3</sub>		1.179±0.189	90.63±17.40	77
2105		Med-Chem.	Author manus CH <sub>3</sub>	script: avail	able in PMC 2021 1 0.0194±0.004	November 12. 1.467±0.145	76

# Table 2.

Inhibition IC50s of compounds with P4 tosyl against human c-20S  $\beta$ 5c and i-20S  $\beta$ 5i.

ID (PKS-)		R₄ N H N H	$\mathbb{R}_{2}$ $\mathbb{H}_{\mathbb{N}_{2}}$ $\mathbb{R}_{1}$		IC50 (µM)		Selectivity
	R4	R3	R2	R1	β5i	β5c	
2256			CH <sub>3</sub>		$0.0004 \pm 0.0002$	$0.424 \pm 0.202$	1208
21209	-∕⊂∽-ö ö		CH <sub>3</sub>	÷ F	0.0024±0.0003	$5.52 \pm 0.928$	2329
21206	-∕⊂∽-ö ö	HN NO	CH <sub>3</sub>	÷ F	$0.0164 \pm 0.0022$	53.7 ± 13.3	3274
21265			CH <sub>3</sub> OCH <sub>2</sub>	÷ F	$0.0008 \pm 0.0003$	$1.456 \pm 0.829$	1881
21266	-∕⊂∑-ÿ-ÿ- Ö	HN K	CH <sub>3</sub> OCH <sub>2</sub>	÷ F	$0.056 \pm 0.002$	$13.74 \pm 1.866$	246

• Data were taken from reference 35.

## Table 3.

Inhibition of i-20S  $\beta$ 5i in Karpas-1106P and c-20S  $\beta$ 5c in Hela cells by select compounds and their inhibition of T cell proliferation in PBMCs upon stimulation by anti-CD3 antibody.

Compound	IC <sub>50</sub> (μΜ	• EC <sub>50</sub> (µM)	
	Karpas-1106P	HELA	20 4 /
PKS3053	0.015	0.293	0.021
PKS 21210	0.039	2.031	0.044
PKS 21272	0.053	4.306	0.073
PKS 21209	0.024	3.030	0.039
PKS 21265	0.028	1.861	0.018
PKS 21266	0.197	22.56	0.073