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Optimal dosing of Meropenem in a small cohort of critically ill children receiving continuous renal replacement therapy

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Abstract

Severe sepsis is an important cause of mortality and morbidity in critically ill children. Meropenem is a broad spectrum antibiotic commonly used to treat sepsis. Current meropenem dosage recommendations for children on continuous renal replacement therapy are extrapolated from pharmacokinetics (PK) studies done in adults. Our study aims to determine the optimal dosing in critically ill septic children receiving CRRT. A prospective single-center PK study was performed in 9 ICU children on CRRT. Meropenem concentrations were measured from blood and effluent fluid samples. A population PK model was developed using nonlinear mixed effects modeling software (NONMEM®). Monte Carlo simulations were performed. The PK/ pharmacodynamic (PD) target aimed for plasma concentrations above minimum inhibitory concentration (MIC) of 4 mg/L for 100% of dosing interval (100% fT>MIC). A two-compartment model best characterized meropenem PK. Mean (range) clearance and elimination half-life was 0.091 L/hr/kg (0.04–0.157) and 3.9 hr (2.1–7.5) respectively. Dosing of 40mg/kg/dose q12h over 30-mins achieved PK/PD target in only 32% while 20mg/kg q8h over 4-hour or 40mg/kg q8h over 2-hour achieved 100% fT>MIC target for at least 90% of simulated patients.

Keywords

Meropenem; pharmacokinetics; pediatrics; critical care; renal replacement therapy

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DATA AVAILABILITY STATEMENT:

The data that support the findings of this study are available from the corresponding author upon reasonable request.

INTRODUCTION

Severe sepsis is an important cause of morbidity and mortality in critically ill children.^{1,2} Early appropriate antimicrobial therapy is crucial.^{3–5} Acute kidney injury during sepsis requiring continuous renal replacement therapy (CRRT) is not uncommon.^{6,7} Critically ill children on CRRT often have altered pharmacokinetics (PK) due to drug extraction by the CRRT circuit and the underlying critical illness.^{8,9} Drug extraction by CRRT depends on drug characteristics, the pore size of the dailysis filter membranes, the type of CRRT modality employed and the changes in blood or dialysate flow rate.^{10,11,12} Fluid overload also affects drug disposition,^{10,13}

Meropenem, a commonly used carbapenem for sepsis treatment, is effective for highly resistant pathogens including extended-spectrum β -lactamase (ESBL) producing *Enterobacteriaceae* and *Pseudomonas aeruginosa*.¹⁴ It exhibits primarily time-dependent antimicrobial activity, and the pharmacokinetic-pharmacodynamic (PK/PD) index that best predicts the in-vivo antimicrobial activity is the fraction of time of the dosing interval during which the free serum concentration of meropenem remains above the minimum inhibitory concentration (MIC) of the pathogen ($f_{T>MIC}$).^{15,16} Traditionally a PK/PD target of percentage time above MIC of 40% of the dosing interval (40% $f_{T>MIC}$) is defined as the primary threshold.¹⁷ For critically ill patients with severe gram-negative sepsis, recent studies have proposed PK/PD target of $f_{T>MIC} > 100\%$ for meropenem, using susceptibility breakpoint of MIC of 4 mg/L.^{10,18–21}

Meropenem is primarily eliminated by the kidneys. Its small molecular size, small volume of distribution (V) and insignificant protein binding (<2%) predisposes it to extensive clearance by CRRT.^{9,22,18,20,23} Current meropenem dosage recommendations for paediatric patients on CRRT are extrapolated from PK studies done in adult CRRT patients.^{6,24} This is a lack of meropenem population PK (PopPK) modelling study that includes critically ill children on CRRT who are less than five years of age.

Our study aims to (i) evaluate the pharmacokinetics of intravenous meropenem in critically ill children receiving meropenem for the treatment of presumed or proven sepsis while on CRRT and (ii) determine the optimal meropenem dosing regimens in critically ill children on CRRT using PopPK modelling and dosing simulations.

METHODS

This study is an open-label was performed at a 16-bed PICU at KK Women's and Children's Hospital on patients who received intravenous meropenem for proven or presumed sepsis and supported with either CVVH or CVVHDF for acute renal failure (ARF). This trial was approved by the institutional review boards for Duke (Protocol ID: Pro00084313) and KK Women's and Children's Hospital (CIRB Ref No: 2011/538/E) with written consent prior to enrollment.

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Study Design

This was an open-label study that enrolled children supported with CRRT and prescribed meropenem by their treating physician. Meropenem dosing was determined by the treating physician based on institutional guidelines of 20mg/kg/dose or 40mg/kg/dose dosed q12h over a 30-minute infusion for children on CRRT. According to the hospital protocol, IV meropenem (MeronemTM, 500mg powder) was administered as 30-minute infusions at a maximum concentration of 50mg/ml. CRRT was performed using the PrismaflexTM for continuous venovenous hemofiltration (CVVH) or hemodiafiltration (CVVHDF) with either a 0.2m² polyarylethersulfone filter (PAES, HF 20, Gambro) for patient weight 10kg, or 0.6m² and 0.9m² polyacrylonitrile filter (AN69, Gambro, Deerfield, IL) for patient weight 11 to 30kg and >30kg, respectively. The CRRT program prescribed Hemosol BO (Baxter) for replacement and dialysate fluids except for the patients who were on regional citrate anticoagulation when Prismocal (Gambro) was used instead. All replacement fluids were administered pre-filter. Patients' actual body weight on admission was used for PK calculations. Demographic, clinical, laboratory and CRRT parameters were collected. Hematology and biochemistry laboratory data were obtained for the tests done within 24 hours of the sampling.

Sample Collection and Analysis

Arterial blood samples from indwelling catheters in the patients and effluent fluid samples from the dialysis machine were collected at specified time points: prior to infusion, at the end of infusion, 1, 2, 4, 8 and 12 hours post infusion. The total volume of urine and effluent were also measured. The supernatant plasma and effluent samples were frozen at -7° C. Meropenem concentrations in plasma and the effluent were quantified using a commercial, validated high-performance liquid chromatography-tandem mass spectrometry bioanalytical assay.²² Accuracy and precision were assessed using 4 determinations at theoretical levels of 0.050, 0.100, 0.500, 1.0, 5.0, 10.0, 50.0 and 100.0 mcg/mL within the Food and Drug Administration (FDA) bioanalytical assay validation criteria (e.g. \pm 15%). The lower limit of quantification for meropenem was 0.050 mcg/L. Because meropenem has neglible protein binding²⁵, total and unbound concentration were assumed to be equivalent.

Pharmacokinetic Analysis

A PopPK model was developed based on the measured meropenem concentrations in plasma using nonlinear mixed effects modeling software (NONMEM, version 7.2). First-order conditional estimation method with interaction was used for all model runs. Run management was performed using Pirana (version 2.8.1). Visual predictive checks and bootstrap methods were performed with Perl-speaks-NONMEM (version 3.6.2).²⁶ Data manipulation and visualization were performed using Stata software (version 13.1, College Station, TX), R (version 3.4.1, R Foundation for Statistical Computing, Vienna, Austria) and RStudio (version 0.97.551, RStudio, Boston, MA, US). Based on visual inspection of the PK data and a review of the primary literature,^{18–20,23,27–31} both one and two compartment models were evaluated using the ADVAN2 TRANS2 and ADVAN4 TRANS4 subroutines, respectively. CRRT clearance model using effluent and plasma concentrations, and CRRT clearance model using effluent concentrations only were also evaluated using ADVAN6

TRANS1. In addition, two CRRT clearance models were also evaluated using ADVAN6 TRANS1 (Supplementary Figure S1).

After the base structural model was identified, covariate analysis was performed using a forward inclusion and backward elimination approach to evaluate the statistical significance of the following covariates: postnatal age, total bilirubin, serum creatinine, albumin, dialysate rate, total ultrafiltration rate, CRRT type, sex, and extracorporeal membrane oxygenation (ECMO) on clearance. Actual body weight (WT) was assumed to be a significant covariate for both clearance and volume of distribution parameters and was included in the base model prior to assessment of other covariates.

A decrease in the OFV with p value < 0.01 and an increase in the OFV with p value < 0.001 were accepted as statistically significant in the forward inclusion and backward elimination steps, respectively, to obtain the final PK irreducible model.

The PK analysis dataset was generated and formatted by merging clinical database data (dosing, demographics, laboratory data) with raw concentration values received from the central laboratory (OpAns).

During the popPK model-building process, successful minimization, diagnostic plots, plausibility, and precision of parameter estimates, as well as objective function and shrinkage values, were used to assess model appropriateness. Diagnostic plots generated included individual predictions and population predictions (PREDs) vs observations, conditional weighted residuals vs PRED and time.

Parameter precision for the final popPK model was evaluated using non-parametric bootstrapping (1000 replicates) to generate the 95% CIs for parameter estimates. Visual predictive checks were performed using the final model to generate 1000 Monte Carlo simulation replicates per time point of meropenem exposure. Simulated results were compared at the participant level with those observed in the study by calculating and plotting the percentile of each observed concentration in relation to its 1000 simulated observations derived from the final model.³²

Extracorporeal Analysis

Area under the curve to the last sample collection time $(AUC\tau)$ for both plasma $(AUC\tau pl)$ and effluent $(AUC\tau uf)$ was determined for each patient using non-compartmental PK analysis in Phoenix WinNonlin (version 6.3, Pharsight Corporation).

The following PK parameters were derived:

1. Sieving/saturation coefficient (Sc/Sa) for CVVH/CVVHDF:

$$Sc \, or \, Sa = \frac{AUC\tau_{uf}}{AUC\tau_{pl}}$$

where $AUC\tau_{uf}$ is the exposure of meropenem in the ultrafiltrate and $AUC\tau_{pl}$ is the exposure of drug in the plasma of simultaneously collected patients' blood specimens.

2. Clearance by CVVH/CVVHDF:

$$CL_{HF} = Q_f * Sc$$

where CL_{HF} is the hemofilter clearance and Q_f is the ultrafiltration rate for CVVH and ultrafiltrate plus the dialysate flow rates for CVVHDF.

3. Fraction of total CL contributed by CRRT (FractEC):

 $FractEC = \frac{CL_{HF}}{CL}$

Extracorporeal clearance was defined as being significant if the Fract_{EC} was greater than 0.3^{33} .

Probability of target attainment (PTA)

We used the PK-Sim[®] (version 7.2; Open Systems Pharmacology Suite, open-systemspharmacology.com) population generator to generate a population of virtual children (n=1000) for simulations. For each simulated individual, we used the final NONMEMgenerated PK model to generate empirical Bayes estimates (EBEs) for clearance and volume of distribution and used Monte Carlo simulations to simulate concentration-time curves at steady state for each dose. Dosing regimens simulated and evaluated included 10mg/kg q8h, 20mg/kg q8h, 40mg/kg q8h (maximum 6grams per day), 20mg/kg q12h, and 40mg/kg q12h infused over 30mins, 2hours and 4hours. We then determined the PTA for the PK/PD targets of 40% $f_{T>MIC}$ and 100% $f_{T>MIC}$ for various MICs (0.5–32mg/L). The PK/PD target was set to achieve meropenem plasma concentrations above MIC of 4 mg/L for 100% of the dosing interval (100% $f_{T>MIC}$), with the assumption that >98% of the drug is unbound. Optimal meropenem dosing was selected when greater than 90% of simulated children achieved the PK/PD target.

RESULTS

Nine patients were enrolled in the study. Their demographic and clinical data are summarized in Table 1. None of the enrolled patients had chronic or end-stage renal failure prior to PICU admission. All patients were were anuric or had neglible urine output at the time of enrolment. All were critically ill with 4 patients on ECMO support and the rest either with multi-organ failure or on inotropic support. The underlying diseases were a mix of postoperative congenital heart disease, hemato-oncology disease and pulmonary disease. The mean duration of CRRT and the mean number of doses of meropenem given before the dose sampling were 5.9 days (2 - 18) and 3.9 doses (1 - 8), respectively.

A total of 53 plasma and 38 dialysate samples contributed to the development of the meropenem PopPK model. A 2-compartment model provided the overall best fit for the data (Supplementary Figure S2).

The mean (range) of combined hemofiltration and dialysate corrected to 1.73m² body surface area (BSA), the mean serum albumin level and mean total bilirubin levels were 2689.5ml/h (940.2–4201.4), 29.9 g/L (21–52) and 77.2 umol/L (18–219), respectively (Supplementary Table S3). We included postnatal age, total bilirubin, serum creatinine, albumin, dialysate rate, total hemofiltration rate, CRRT type, gender and extracorporeal membrane oxygenation (ECMO) on clearance but none were retained as statistically significant in the final population PK irreducible model (Supplementary S4) except for the allometric relationship with total WT with fixed estimates of 0.75 for clearance and 1.0 for volume of distribution. The goodness-of-fits plots are shown in Figure 1.

The model was evaluated using a 1000-set bootstrap analysis. The median of bootstrap parameter estimates were within 10% of population estimates from the original data set for all parameters (Supplementary Table S5). The prediction corrected visual predictive check revealed a reasonable fit between the observed and predicted meropenem concentrations with only 5.7% (3/53) of observed concentrations outside of the 90% prediction interval (Supplementary Figure S6). Median (range) weight adjusted CL from the EBEs obtained from the final model was 0.096 L/hr/kg (0.040–0.157). Summaries for other EBE PK parameters are shown in Table 2. The median (range) AUC τp and AUC τeff were 196.6 (39.6–515.8) and 160.8 (42.0–575.8) respectively. Median (range) Sc/Sa was 0.958 (0.737–1.1) and mean Fract_{EC} was 0.816 (0.457–1.3).

Based on simulations, meropenem regimens of 20mg/kg/dose q8h over 4hours or 40mg/kg/ dose q8H over 2hours achieved the target attainment of \geq 90% patients achieving concentration above MIC of 4mg/L for 100% of the dosing interval (100% $f_{T>MIC}$) (Table 3, Supplementary Figure S7).

DISCUSSION

Optimal meropenem dosage regimen in critically ill patients with acute renal failure on CRRT is lacking especially in young children. Inadequate dosing to achieve the desirable PK targets can lead to treatment failure and emergence of antibiotic resistance. Our results show that longer infusions of meropenem in this group of patients are required to achieve adequate PK targets.

We demonstrated that a 2-compartment model provided the overall best fit for our meropenem PK data which is consistent with other meropenem PK studies in patients on CRRT.^{18–20,27–30} The mean Sc/Sa of approximately 1 meant that meropenem was freely filtered by the hemofilter/dialysis membrane. This was consistent with the values reported. ^{18–20} The population estimates of CL in children on CRRT normalized to 70 kg (median [range] of 4.1 L/hour [2.6 – 6.3]) differed substantially from the CL estimates of 14.1 L/hour based on the FDA label for children with normal renal function.²⁵ This was likely due to low or negligible renal clearance of meropenem in these patients and the CRRT clearance of

meropenem was lower compared to normal renal clearance.³⁴ The mean meropenem CRRT clearance of 80% exceeded the suggested 30% threshold required for dosing adjustment while on CRRT.³³

Of note, we included a very sick group of patients in PICU with almost half of our study population on ECMO support. ECMO can alter PK by increasing volume of distribution through adsorption and hemodilution.³⁵ We explored ECMO as a covariate in our analysis but in our final irreducible model, ECMO was not retained as a covariate.

Recent evidence suggests improved outcome when the percentage of T>MIC is 100% of the dosing interval (100% $f_{T>MIC}$) for critically ill or neutropenic patients on meropenem.³⁶ Based on this surrogate endpoint for efficacy, current meropenem dosing recommendations from FDA label may be sub-therapeutic in critically ill children on CRRT. A population PK simulation in 9 critically ill children with normal renal function also suggested that current dosing recommendation was inadequate and a meropenem dosage regimen of 120 to 160mg/kg/day as continuous infusion was necessary to achieve 80% $f_{T>MIC}$ (MIC 2mg/L).²⁹ However, continuous infusions can be a challenge in critically children when there is often competing needs for limited venous access and when constituted meropenem at room temperature is only stable for 4 hours. Other studies had suggested a PK target based on meropenem concentration >4X MIC for 75% to 100% of the time was required in ill patients.^{37,38} In our study, only one patient had a blood culture positive for E.coli (ESBL) but there was no data on MIC available. During the study period, the institution's antibiogram showed a 100% sensitivity to meropenem for Enterobacteriaceae and 80-100% sensitivity to meropenem for *Pseudomonas aeruginosa* using a susceptibility breakpoint of 2mg/L for both. The Clinical and Laboratory Standards Institute (CLSI) MIC breakpoints for Pseudomonas aeruginosa and Enterobacteriaceae are 2 mg/L and 1 mg/L, respectively.³⁹ With this, we based our PTA on the PK/PD target to achieve 100% $f_{T>MIC}$ (MIC=4 mg/L as breakpoint) in 90% of the critically ill children on CRRT. Our simulations suggest either a 20mg/kg/dose q8h over 4-hour infusion or a 40-mg/kg/dose q8h over 2-hour infusion is required. The dosing regimen differed from an earlier study by Nehus et al²⁰ that suggested that meropenem dosing of 20mg/kg q12h over 30mins was adequate to achieve the same targets.²⁰ One explanation could be that our study involved younger children and younger children may require more frequent dosing to achieve the same therapeutic target compared to older children.¹⁹

As the sensitivity patterns of *Enterobacteriaceae* and *Pseudomonas aeruginosa* to meropenem may differ in different settings, and PK targets also differ based on site and severity of infection, we also express the target attainment rates for T>MIC of 40% and T>MIC of 100% using different dosing regimen and different MIC breakpoints (Table 3).

Meropenem is generally well tolerated but its serum concentration has been shown to correlate with acute neurotoxicity and nephrotoxicity in adults, with trough thresholds of 64mg/L and 44mg/L, respectively.⁴⁰ We did not simulate trough levels for the dosing regimen of 20mg/kg/dose q8h over 4-hour infusion or a 40-mg/kg/dose q8h over 2-hour infusion. However, a study demonstrated a low proportion of simulated troughs exceeding toxic levels using a meropenem regimen of 2g/kg q8h (equivalent to 40mg/kg for a 50-kg

individual) over 2- or 4-hour infusions in adults with hematological malignancies including those with creatinine clearance $50\text{ml/min/1.73m}^{2.41}$ With the median population estimates of CL in children on CRRT normalized to 1.73 m^2 in our study to be 60.0 ml/min (range 41.2 - 91.3), we postulate that our dosing regimen to be at low risk of attaining toxic trough levels. There are a few limitations to this investigation. Firstly, the study was conducted in a single centre with a small sample size (n = 9). The patients had heterogeneous underlying diseases. This could affect the reliability and generalizability of the data and is a possible reason why no covariates were retained in the final model. Secondly, we have assumed a low protein binding of 2% for meropenem. A study has recently shown a wide variability of protein binding for meropenem and the measurement of total meropenem level in our study could have overestimated the unbound fraction.⁴² Thirdly, as dialysis clearance is influenced by the physicochemical properties of the dialysis filter, caution must be exercised when extrapolating these results to other types of dialysis membranes.

In conclusion, our findings support the use of longer meropenem infusions in critically ill patients. Due to the limitations of this study, larger prospective clinical studies are needed to validate this dosing regimen.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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All other authors has no financial disclosure to declare.

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(A) Observed versus Individual Predictions for the Final Meropenem Model



The solid lines represent lines of identity. The red dashed lines represent smooth lines by loess fit.

Inter-individual variability (IIV) on CL was 27.1%, and the residual (proportional) error was 25.7%. Only IIV for CL due to unreliability of the data and high shrinkage values (> 60%) for the other random-effects could be estimated parameters

(B) Weighted Residuals (CWRES) vs Population Predictions and Time After Dose for the Final Meropenem PK Model Sh



The solid lines represent CWRES=0. The middle dashed line represents a smooth line by loess fit.

Shrinkage estimates were less than 10% for all random-effect parameters.

Figure 1.

Goodness-of-fits plots for the construction model.

TABLE 1.

Clinical Parameters of Study Population

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Concurrent medications	Erythromycin,
Dialysis/ Replacement Fluid	
Other concurrent extracorporeal therapy	
Mode of Renal placement	

10 Cander Rate (array) Weigh (array) Under/joint (array) Positive (array) 1 Positive (array) 1 2 1 </th <th></th> <th></th> <th></th> <th></th> <th></th> <th></th>										
D Gender Race (rous) Mage (rous) Description (topologic) Notified (topologic) Notified (topologic) Notified (topologic) Distribution (topologic) Dis	Concurrent medications	Erythromycin, Amphotericin B, Rocuronium, Midazolam, Fentanyl, Omeprazole	Ampicillin, Neomycin, Rocuronium, Midazolam, Fentanyl, Ranitidine, Oseltamivir, Dopamine, Mirlinone, Hydrocortisone	Vancomycin, Fluconazole, Morphine, Dopamine, Epinephrine, Norepinephrine,	Vancomycin, Fluconazole, Morphine, Dopamine, Epinephrine, Norepinephrine, Heparin Arbisome, Caspofungin, Morphine, Midazolam, Omeprazole, Dopamine, Norepinephrine,					
ID Gender Rate (version) Age (version) Weight (version) Byr of the seat Wood flow (version) Other (version) <	Dialysis/ Replacement Fluid	Hemosol BO	Hemosol BO	Hemosol BO	Hemosol BO PrismOCal					
D Gender Rate Results Mage (varies) (varies) Weight (varies) (varies) Day of (varies) (varies) Not efficie (varies) Urite Results Mode of (varies)	Other concurrent extracorporeal therapy	VA ECMO	VA ECMO	IN	IN	IN				
D Gender Race Age (sensis) Weight (sensis) Day of (sensis) No. of prior (sensis) Underlying (sensis) Positive (sensis) Positive (sensis) Underlying (sensis) Positive (sensis) Positive (sensitive (sensitive) Underlying (sensis) Positive (sensitive) Underlying (sensis) Positive (sensitive) Underlying (sensitive) Positive (sensitive) O 3 Fennale Chinese 0.1 2.6 6 2 Positive(sensitive) 0 9 4 Male Chinese 16 3.2.7 2 1 Positive(sensitive) 0 5 Male Chinese 9.6 41.5 2 Sensitive(sensitive) 0 5	Mode of Renal Replacement	CVVHDF	СVVН	СЛИН	CVVHDF	СVVН				
ID Gender Race Age (sers) Weight (sers) Day of (ses) No. of prior (doese of meropenen Underlying (assed) Positive meropenen 1 Fennale Myramnese 0.1 3.3 6 8 Critical More Positive meropenen 2 Male Chinese 0.1 3.3 6 3 3 Critical Myrocardits, Myrocardits, and live Protocardits, Pertuasis 3 Fennale Chinese 0.1 2.6 3 3 and live Influence 3 Fennale Chinese 0.1 2.6 6 2 herroletarial failure Myrocardits, failure NYA: 3 Fennale Chinese 0.1 2.6 6 2 herroletarial failure Myrocardits, failure 4 Male Chinese 16 5.7 2 1 germaturity severe conflicter, K. 5 Male Chinese 9.6 41.5 2 severed failure conflicter, K. 5 Male Chinese 9.6 41.5 2 severed failure conflicter, K.	Urine output (ml/kg /h)	0	0	0	<0.1	0				
ID Gender Race Age (years) Weight (kg) Day of CRRT No. of Prior doess of meropenen Underlying doess of meropenen 1 Female Myaannese 0.1 3.3 6 8 Postoperative acute renail 2 Male Chinese 0.3 6 3 3 Terrel acute renail 3 Female Myaannese 0.1 3.3 6 3 3 3 Female Underlying doese of meropenen 0.3 6 3 3 4 Male Chinese 0.1 2.6 6 2 Postoperative complex 3 Female Chinese 0.1 2.6 6 2 Postoperative complex 4 Male Chinese 16 52.7 2 1 Postoperative complex 5 Male Chinese 9.6 41.5 2 Supreselar transplant 5 Male Chinese 9.6 41.5 5 Relapsed Relapsed Relapsed Relapsed Relapsed	Positive Microbiolgy Results	NPA: Pertussis	NPA: Influenza A		Blood culture: E. Coli (ESBL), Candida krusei ETT culture: A. baumanii	Blood: CMV PCR + NPA: Parainfluenza I				
IDGenderRaceAge (years)Weight (kg)Day of (Rs)No. of prior doss of accopenen1FemaleMyannese0.13.3682MaleChinese0.36333FemaleChinese0.36334MaleChinese0.12.6624MaleChinese0.12.6625MaleChinese165.7214MaleChinese9.641.5255MaleChinese9.641.525	Underlying Illness	Critical pertussis, MOF	Myocarditis, acute renal and liver failure	Postoperative complex cyanotic heart disease, prematurity, severe capillary leak	Relapsed suprasellar germinoma post stem cell transplant	Relapsed ALL post stem cell transplant, CMV pneumonitis				
IDGenderRaceAge (years)Weight (kg)Day of CRRT1FemaleMyanmese0.13.361FemaleMyanmese0.13.362MaleChinese0.3633FemaleChinese0.12.663FemaleChinese0.12.664MaleChinese0.12.665MaleChinese9.641.525MaleChinese9.641.52	No. of prior doses of meropenem	œ	£	7	I	'n				
IDGenderRaceAge (years)Weight (kg)1FemaleMyanmese0.13.31FemaleMyanmese0.13.32MaleChinese0.363FemaleChinese0.12.64MaleChinese0.12.65MaleChinese9.15.775MaleChinese9.641.5	Day of CRRT	9	£	9	7	2				
ID Gender Race Age (years) 1 Female Myannese 0.1 2 Male Chinese 0.3 3 Female Chinese 0.1 4 Male Chinese 0.1 5 Male Chinese 9.6	Weight (kg)	e 33		2.6	52.7	41.5				
ID Gender Race 1 Female Myannese 2 Male Chinese 3 Female Chinese 5 Male Chinese	Age (years)	0.1		0.1	16	9.6				
ID Gender 1 Female 2 Male 3 Female 4 Male 5 Male	Race	Myanmese	Myanmese Chinese		Myanmese Chinese		Chinese	Chinese		
v B B	Gender	Female	Male	Female	Male	Male				
	A	1	7	3	4	Ś				

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Concurrent medications	Ambisome, Anidulafingin, Morphine, Midazolam, Epinephrine, Tranexamic acid, Heparin	Levofloxacin, Fluconazole, Fentanyl, Midazolam, Acetaminophen	Doxycycline, Ambisome, Morphine, Midazolam, Acetaminophen, Dexamethasone, Tranexamic acid	Morphine, Midazolam, Omeprazole, Epinephrine, Milrinone, Amiodatone, Sodium valproate, Heparin, Trenaxemic acid,)F: Multiorgan
Dialysis/ Replacement Fluid	Hemosol BO	PrismOCal	PrismOCal	Hemosol BO	ta=lactamase; MC
Other concurrent extracorporeal therapy	VV ECMO	Nil	Nil	VA ECMO	ended spectrum be
Mode of Renal Replacement	CVVHDF	CVVHDF	СVVН	CVVHDF	lovirus; ESBL: Ext
Urine output (ml/kg /h)	0	0	<0.1	0	Cytomegal
Positive Microbiolgy Results	BAL culture: S. A. baumanii	Blood culture: S. pneumoniae ETT culture: A. baumanii	Stool culture: <i>E. coli</i> (ESBL)	ETT culture: E. cloacae, A. baumanii	Barr virus; CMV: (
Underlying Illness	Pulmonary hemosiderosis with ARDS	Streptococcus pneumonia with HUS	EBV related HLH, MOF	Postoperative Fontan	ate; EBV: Ebstein
No. of prior doses of meropenem	7	4	7	n	pharyngeal aspir
Day of CRRT	18	6	2	œ	NPA: naso
Weight (kg)	29	10.3	19	56.3	cheal tube;
Age (years)	13.8	1.8	4.9	18.9	TT: endotrac
Race	Malay	Chinese	Japanese	Malay	solar lavage; E1
Gender	Female	Female	Female	Male	Bronchoalve
D	6	7	œ	6	BAL: I

failure; ARDS: Acute respiratory distress syndrome; ALL: Acute lymphoblastic leukemia; HLH: Hemophagocytic lymphohistiocytosis; HUS: Hemolytic uremic syndrome; VA ECMO: Veno-arterial extra-corporeal membrane oxygenation; VV ECMO: Veno-venous extra-corporeal membrane oxygenation

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Indiv idual Empirical Baye s Post-hoc Parameter Estimates and Extracorporeal Parameters

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Ð	Meropenem dose per body weight (mg/kg)	Dosing interval	CL (L/hr/k g)	V (L/kg)	Vp (L/kg)	Q (L/kg/h r)	T ¹ / ₂ (hrs)	CL (L/hr)	AUCt(p) (mg*h/L)	AUCt(eff) (mg*h/L)	Sc/Sa	QE (L/h)	CLCRRT (QE*Sc) (L/h)	Fract _{EC} (CL (L/hr)
1	39.4	Q12H	0.157	0.194	0.209	0.277	2.1	0.519	39.6	42	1.1	0.525	0.557	1.1
2	20	Q12H	0.097	0.2	0.217	0.233	3.2	0.581	104.3	97.5	0.935	0.612	0.572	0.983
3	38.5	Q12H	0.102	0.194	0.209	0.294	3	0.265				0.099		
4	19	Q12H	0.04	0.194	0.209	0.139	7.5	2.1	81.7	70.9	0.867	3.3	2.9	1.3
5	24.1	Q12H	0.058	0.193	0.21	0.147	5.4	2.4	267.1	302.2	1.1	1.9	2.1	0.885
9	20	Q12H	0.096	0.193	0.21	0.162	3.5	2.8	155.9	114.9	0.737	1.8	1.3	0.476
7	39.8	Q12H	0.096	0.194	0.214	0.204	3.3	0.989	515.8	575.8	1.1	0.423	0.472	0.477
8	42.1	Q12H	0.082	0.195	0.211	0.179	3.9	1.6	280.2	274.8	0.981	1.2	1.2	0.747
6	35.5	Q12H	0.095	0.194	0.21	0.137	3.6	5.3	237.2	206.6	0.871	2.8	2.4	0.457
MEAN	29.8		0.091	0.194	0.211	0.197	3.9	1.8	210.2	210.6	0.962	1.4	1.4	0.806
SD	9.5		0.032	0.002	0.002	0.056	1.6	1.6	152.1	175.6	0.137	1.1	0.93	0.326

CL=plasma clearance;MAX=maximum; MIN=minimum; Q= I nter-compartmental clearance;T $\frac{1}{2}$ = half-life;

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Vc= Volume of distribution in central compartment; Vp= Volume of distribution in peripheral compartment

AUCt (eff)=Area under the curve from the last dose to last measurable concentration for effluent; AUCt(p=Area under the curve from the last dose to last measurable concentration for plasma; CL=plasma clearance; CL CRRT=continuous renal replacement therapy clearance;

FractEC=Fraction of contribution of CRRT CL to total CL; QE=effluent flow rate; SC/SA= sieving/saturation coefficient. [†]Patient with ID #3 had only 1 PK sample.

TABLE 3.

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		0mg/kg q8H	100	100	100	100	100	100	100			0mg/kg q8H	100	100	66	97	90	68		
		20mg/kg 4 q8H	100	100	100	100	100	100	100			20mg/kg 4 q8H	100	66	76	06	69	30	 	
	ırs infusion	10mg/kg q8H	100	100	100	100	100	100	91		ırs infusion	10mg/kg q8H	66	97	06	69	30	3	,	
	4 hou	40mgrkg q12H	100	100	100	100	100	100	66			4 hou	40mg/kg q12H	94	87	74	53	31	10	
		20mg/kg q12H	100	100	100	100	100	66	72			20mg/kg q12H	87	74	53	31	10	0		
		40mg/kg q8H	100	100	100	100	100	100	66			40mg/kg q8H	66	98	96	06	73	45		
	u	20mg/kg q8H	100	100	100	100	100	66	88		u	20mg/kg q8H	98	96	06	73	45	14		
	ours infusio	10mg/kg q8H	100	100	100	100	66	88	12		hours infusion	10mg/kg q8H	96	06	73	45	14	1		
	21	40mg/kg q12H	100	100	100	100	100	76	62		21	40mg/kg q12H	87	76	60	39	21	4		
		20mg/kg q12H	100	100	100	100	76	79	26			20mg/kg q12H	76	09	39	21	4	0		
		40mg/kg q8H	100	100	100	100	100	66	95		ion	40mg/kg q8H	98	96	92	82	59	31		
6	ninutes infusion	20mg/kg q8H	100	100	100	100	66	95	56	%		20mg/kg q8H	96	92	82	09	31	6		
>MIC of 40%		10mg/kg q8H	100	100	100	66	95	56	3	>MIC of 100	minutes infu	10mg/kg q8H	92	82	09	31	6	1	,	
Rates for T	301	40mg/kg q12H	100	100	100	100	86	68	56	Rates for T	301	40mg/kg q12H	81	67	51	32	13	3		
t Attainment		20mg/kg q12H	100	100	100	86	89	56	11	t Attainment		20mg/kg q12H	67	51	32	13	3	0		
Targe		MIC	0.5	1	2	4	8	16	32	Targe		MIC	0.5	1	2	4	8	16	;	

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MIC=Minimum Inhibitory Concentration (mg/L); q12h= every 12hours; q8h= every 8hours

Red box indicate the optimal target attainment rates (>90%) for 100% fT>MIC=4