# RESEARCH ARTICLE

# Pineocytoma and Pineal Parenchymal Tumors of Intermediate Differentiation Presenting Cytologic Pleomorphism: A Multicenter Study

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#### Keywords

pineal parenchymal tumors; pineocytoma; pleomorphic; neuronal differentiation.

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Received 10 October 2007; accepted 12 November 2007.

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doi:10.1111/j.1750-3639.2008.00128.x

#### Abstract

# Cytologic pleomorphism has been described in a limited number of benign pineal tumors, namely pineocytoma (PC) and pineal parenchymal tumors (PPTs) of intermediate differentiation (PPTID). We examined the clinicopathologic features in a retrospective series of 14 cases (seven females and seven males aged from 10 to 65 years) of pleomorphic PPT. Seven cases were PC, with no mitoses and with areas of tumoral cells forming large pineocytomatous rosettes and other areas with giant cells containing hyperchromatic nuclei. The other seven were PPTID, presenting few mitoses ( $\leq 2$ ), a Ki67 proliferation index between 3% and 7%, and predominantly composed of small neoplastic cells and scattered giant cells, sometimes multinucleated. In the 14 tumors, the proportion of pleomorphic areas was variable. Most tumoral cells showed extensive neuronal differentiation with strong expression of neuron-specific enolase, synaptophysin and neurofilaments. Some of the neoplastic cells expressed S100 protein. The follow-up period ranged from 1.2 to 13 years and only one PC and one PPTID progressed after stereotactic biopsy or incomplete resection. The lack of

invasiveness and the low proliferation index of these tumors suggest a benign clinical course

despite the marked pleomorphism, the latter of which can lead to upgrading.

# INTRODUCTION

Pineal parenchymal tumors (PPTs) represent about 30% of pineal region neoplasms and are rare tumors accounting for less than 0.1% of all brain tumors. They are subdivided into pineocytoma (PC), pineoblastoma and PTT of intermediate differentiation (PPTID) (9, 15, 16). In PC, cytologic pleomorphism accompanied by ganglion cells intensely expressing neuronal markers has been described (9, 11). Moreover, cytologic pleomorphism has been recently reported in one case of PPTID (20). However, because of the rarity of PPT, these are the only tumors with cytologic pleomorphism reported in the literature. The presence of pleomorphic cells may lead to upgrading of tumors. In this study, we report 14 cases

of PPT presenting cytologic pleomorphism from our cooperative series of more than 100 patients presenting with PPT.

# **MATERIALS AND METHODS**

#### **Patients**

One hundred and ten out of 433 tumor samples of the pineal region obtained between 1979 and 2006 in the different centers were retained as PPT after histopathological review. Fourteen of these showed pleomorphism and were studied using light microscopy and immunohistochemistry. Five of these cases have been included in a previous study analyzing the correlation of histological features with prognosis in 66 cases of PPT (9).

#### Light microscopy

Surgically removed tissues were fixed in 4% paraformaldehyde/ 15% picric acid, formol or formol-zinc and embedded in paraffin, then cut into 4  $\mu$ m sections. Routine staining was performed using hemalin phloxin saffron, hemalin eosin saffron or hemalin eosin. Mitoses were counted in 10 randomly selected fields at high magnification (×400) and expressed as the number of mitoses in 10 high-power fields (HPFs). The number of vessels was scored as 0, + or ++ when, respectively, absent, moderate or high. For a subset of tumors, cytological imprints were prepared and stained with toluidine blue.

#### Immunohistochemistry

With the exception of those intended for testing for the presence of S100 protein, the sections were heated for  $2 \times 5$  minutes in citrate buffer in a microwave oven, then, stained with the specific antiserum. Immunohistochemical staining was performed using the avidin-biotin technique and either polyclonal antiserum against S100 protein or monoclonal antibodies against neuron-specific enolase (NSE), synaptophysin (SYN), neurofilament (NF, 70 and 200 kDa forms), chromogranin A (ChgA), NeuN, vimentin (VIM), glial fibrillary acidic protein (GFAP), or CD34 (see Table 2 for sources). The intensity of labeling was scored as 0 when absent or +/- or + when, respectively, moderate or high. Immunohistochemistry with anti-Ki67 antibody (MIB1 clone, Dako) was performed on samples fixed in formol or formol-zinc in order to estimate cell proliferation.

#### RESULTS

#### **Clinical data**

The main clinical data are summarized in Table 1. Seven cases, with a male predominance (six males and one female), were pleomorphic PC without mitoses and considered as low grade tumors (grade I). The other seven cases, with a female predominance (one male and six females), were tumors with  $\leq 2$  mitoses and a Ki67 proliferation index between 3% and 7% and were considered as PPTID grade II. The patients' ages ranged from 10 to 65 years for the PCs (mean  $\pm$  SEM = 47.4  $\pm$  7.1), and from 24 to 60 years for the PPTIDs (mean  $\pm$  SEM = 42.0  $\pm$  6.1).

On computer tomography (CT), the tumors were non invasive, well-circumscribed masses that were isodense to hypodense on non-contrast CT. Homogeneous enhancement was seen with the use of contrast. On magnetic resonance imaging, the tumors exhibited low isointensity or mixed signal intensity on T1-weighted images (Figure 1A). Homogeneous to slightly heterogeneous contrast enhancement was seen on the contrast enhanced images (Figure 1B).

Tumor size (maximal diameter as assessed in preoperative imaging studies) ranged from 20 to 50 mm and was not different between the PCs ( $25.0 \pm 2.2$ ) and PPTIDs ( $35.0 \pm 4.4$ ). Complete resection of the tumor was achieved in nine patients; in the other five cases, only incomplete resection of the tumor was feasible or

surgery remained restricted to diagnostic biopsy. Nine patients had no treatment after surgery, while five received radiotherapy, focally in three cases.

Detailed follow-up information was available for 13 patients. The follow-up period ranged from 1.2 to 13 years. Progression was only seen in two patients, one 8 years after PC was diagnosed by stereotactic biopsy and one PPTID patient 1.4 years after incomplete resection. Of the seven patients so far followed up for 5 years or known to have died in this period, four out of four with PC survived (one dying at 7 years and one at 8 years) and two out of three with PPTID survived, one dying of pneumopathy at 1.9 years.

# **HISTOPATHOLOGICAL FEATURES**

The pleomorphic PCs presented areas with tumoral cells forming large pineocytomatous rosettes of variable number and size and areas with giant cells with hyperchromatic nuclei (Figure 2A). The cytological imprint showed pinealocytes with small round nuclei and other larger cells with pleomorphic nuclei (Figure 2A inset). The pleomorphic PPTIDs were predominantly composed of small neoplastic cells with a high nuclear to cytoplasmic ratios, dense chromatin and irregularly shaped nuclei along with scattered giant cells, which were sometimes multinucleated (Figure 2B). The proportion of pleomorphic areas varied between tumors and could represent more than half of the tumor. Only three tumors presented any mitotic figures and these were few (1–2 per 10 HPFs). Despite the marked pleomorphism, neither endothelial proliferation nor necrosis was found. In all tumors, numerous vessels were seen.

The immunohistochemical results are reported in Table 2 and illustrated in Figure 2. The tumor cells were positive for the neuronal markers SYN (Figure 2C,D), NSE, and NF (Figure 2E,F). Some giant cells were intensively labeled, especially with anti-NF antibodies (Figure 2E,F). In some cases, the neoplastic cells were reactive for ChgA (Figure 2G). In a subset of tumors analyzed (one PC and four PPTIDs), no NeuN immunoreactivity was seen. Of the two tumors analyzed, a pleomorphic PC expressed CD34 only in vascular endothelial cells, while a PPTID also expressed this glycoprotein in giant cells (Figure 2H). In some tumors, vimentin expression was restricted to vessels, while, in others, it was also seen in tumor cells. Only interstitial and glial cells were labeled with anti-GFAP antibody. Giant cells in some tumors expressed S100 protein (Figure 2I). The Ki67 labeling index varied from 3% to 7% (4.7  $\pm$  0.5) in the maximally stained areas in the PPTIDs (Figure 2J).

#### DISCUSSION

Pleomorphic variants of PPT were first described in PC, with the presence of giant cells with irregular, hyperchromatic nuclei, sometimes multinucleated, in tumors also presenting characteristic pineocytomatous rosettes (9, 11). Cytologic pleomorphism has also been reported in one case of PPTID with moderate proliferative activity (20). An accurate histological diagnosis is essential for these rare tumors because optimal therapeutic management depends on the tumor grading. Our large cooperative series of PPTs made it possible for us to collect 14 clinically followed cases of pleomorphic PPT presenting features of either PC or PPTID. Except for one young boy, the PC patients were older than 40 years. More than half of the patients with PPTID were younger than 40

GaseTuron sizeTuron sizeComplementary adjuantFollow-upFollow-upKonser/10 HPSKosser/10 HPS <t< th=""><th>numb€</th><th>er of vessels; RT =</th><th>radiotherapy.</th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th></t<>	numb€	er of vessels; RT =	radiotherapy.								
1   10M   20   Complete resection   PC   1   No   0	Case	Age (years)/sex	Tumor size (mm)	Operation	Histological type	Grade	Complementary adjuvant treatment	Follow-up	Mitoses/10 HPFs	Vessels	Ki67 (%)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	-	10/M	20	Complete resection	РС	_	No	No recurrence, alive at 13 years	0	0	DN
	2	42/M	30	Complete resection	PC	_	No	Lost to follow-up	0	+	ND
	с	44/M	30	Stereotactic biopsy	PC	_	Focal RT	Progression at 8 years, death	0	+	DN
5 $4M$ 30Complete resectionPC1NoAlive at 4 years0+ND7 $64/F$ 20Complete resectionPC1NoNo recurrence, alive at0+ND7 $65/M$ 20Complete resectionPC1FC1No recurrence, alive at0+ND8 $24/F$ NAVentriculoscopy and biopsyPPTID1RTNo recurrence, alive at0+ND9 $24/F$ 40BiopsyPPTID1NoNo recurrence, alive at0+4810 $33/F$ 50Complete resectionPPTID1NoNo recurrence, alive at0+4811 $37/F$ 50Complete resectionPPTID1NoNo recurrence, alive at0+4611 $37/F$ 50Complete resectionPPTID1NoNo recurrence, alive at0+4612 $57/F$ NAVenescific loopsyPPTID1NoNo recurrence, alive at0+4613 $37/F$ 30Endoscopic loopsyPPTID1NoNo recurrence, alive at0+4613 $37/F$ NAStereosticic loopsyPPTID1NoNo recurrence, alive at0+6613 $37/F$ NAStereosticic loopsyPPTID1NoNo	4	53/M	AN	Ventriculoscopy Stereotactic biopsy Complete resection	PC	_	Focal RT	Alive at 2 years	0	+	0
	വ	54/M	30	Complete resection	PC	_	No	Alive at 4 years	0	+	ND
7 $65/M$ 20Complete resectionPCIFocal RTNo recurrence, death at0 $-+$ NO8 $24/F$ NAVentriculoscopy and biopsyPPTIDIRT $7 \text{ years}$ 1 $++$ 39 $24/F$ 40BiopsyPPTIDINoNo recurrence, alive at0 $++$ 410 $33/F$ 50Complete resectionPPTIDINoNo recurrence, alive at0 $++$ 411 $37/F$ 30Endoscopic biopsyPPTIDINoNo recurrence, alive at0 $++$ 411 $37/F$ 30Endoscopic biopsyPPTIDINoNo recurrence, alive at0 $+$ 412 $57/F$ NANo recurrence, alive at0 $+$ 14 $+$ 412 $57/F$ NAEndoscopic biopsyPPTIDINo $-1.2 \text{ years}$ 0 $+$ 413 $59/M$ 29Endoscopic biopsyPPTIDINo $-1.2 \text{ years}$ 2 $+$ 414 $60/F$ 26Ventriculositer resectionPPTIDINo $-1.2 \text{ years}$ 2 $+$ 614 $60/F$ 26Ventriculositer resectionPPTIDINo $-1.2 \text{ years}$ 2 $+$ 615 $60/F$ 26Ventriculositer resectionPPTIDINo $-1.2 \text{ years}$ 1 $+$ 714 $60/F$ <td>9</td> <td>64/F</td> <td>20</td> <td>Complete resection</td> <td>РС</td> <td>_</td> <td>No</td> <td>No recurrence, alive at 13 years</td> <td>0</td> <td>+</td> <td>DN</td>	9	64/F	20	Complete resection	РС	_	No	No recurrence, alive at 13 years	0	+	DN
8   24/F   NA   Ventriculoscopy and biopsy Complete resection   PTID   I   RT   Alive at 6 years   1   +   3     9   24/F   40   Biopsy   PPTID   I   No   No recurrence, alive at   0   +   4     10   33/F   50   Biopsy   PPTID   I   No   No recurrence, alive at   0   +   4   4     11   37/F   50   Complete resection   PPTID   I   No   No recurrence, alive at   0   +   6     11   37/F   50   Complete resection   PPTID   I   No   No recurrence, alive at   0   +   6     12   37/F   30   Endoscopic biopsy   PPTID   I   No   No recurrence, alive at   0   +   4     13   57/F   NA   Steneostatic biopsy   PPTID   I   No   No recurrence, alive at   0   +   4     13   59/M   29   Steneostat	2	65/M	20	Complete resection	РС	_	Focal RT	No recurrence, death at 7 years	0	+	DN
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	00	24/F	AN	Ventriculoscopy and biopsy Complete resection	DITID	=	RT	Alive at 6 years	<del>, -</del>	+	с
10   33/F   50   Complete resection   PPTID   I   No   No recurrence, alive at   0   +   6     11   37/F   30   Endoscopic biopsy   PPTID   I   No   1.2 years   1   4   4     12   37/F   30   Endoscopic biopsy   PPTID   I   No   1.4 years   0   4   4     12   57/F   NA   Stereostatic biopsy   PPTID   I   No   1.4 years   2   ++   4     13   59/M   29   Stereostatic biopsy   PPTID   I   No   1.4 years   2   ++   4     13   59/M   29   Stereostatic biopsy   PPTID   I   No   1.4 years   2   ++   5     14   60/F   26   Venticulosity   11   No   1.9 years   2   ++   5     14   60/F   26   Venticulosity   1   1.9 years   1.9 years   1.4   1.7	0	24/F	40	Biopsy Resection	DITIO	=	No	No recurrence, alive at 3.5 years	0	+	4
11   37/F   30   Endoscopic biopsy Incomplete resection   PPTID   II   No   Progression, alive at   0   0   4     12   57/F   NA   Stereostatic biopsy Incomplete resection   II   RT   1.4 years   1.4   4     13   59/M   29   Stereostatic biopsy Endoscopic biopsy Incomplete resection   II   No   Death (pneumopathy) at   0   ++   4     13   59/M   29   Stereostatic biopsy Endoscopic biopsy Incomplete resection   II   No   Death (pneumopathy) at   0   ++   5     14   60/F   26   Ventriculocisternostomy   PTID   II   No   1.9 years   2   ++   7     14   60/F   26   Ventriculocisternostomy   PTID   II   No   Alive at 1.7 years   2   ++   7	10	33/F	50	Complete resection	DPTID	=	No	No recurrence, alive at 1.2 years	0	+	9
12 57/F NA Stereostatic biopsy PPTID II RT Alive at 6.6 years 2 ++ 4   13 59/M 29 Stereostatic biopsy PPTID II No Death (pneumopathy) at 0 ++ 5   13 59/M 29 Stereostatic biopsy PPTID II No 0 ++ 5   14 60/F 26 Ventriculocisternostomy PPTID II No Alive at 1.7 years 2 ++ 7   14 60/F 26 Ventriculocisternostomy PPTID II No Alive at 1.7 years 2 ++ 7	11	37/F	30	Endoscopic biopsy Incomplete resection	DPTID	=	No	Progression, alive at 1.4 years	0	0	4
13   59/M   29   Stereostatic biopsy   II   No   Death (pneumopathy) at   0   ++   5     Redoscopic biopsy   1.9 years   1.9 years   1.9 years   1.9 years   ++   7     14   60/F   26   Ventriculocisternostomy   PTID   II   No   Alive at 1.7 years   +   7	12	57/F	AN	Stereostatic biopsy Incomplete resection	DPTID	=	RT	Alive at 6.6 years	2	‡	4
14 60/F 26 Ventriculocistemostomy PPTID II No Alive at 1.7 years 2 + 7 Complete resection	13	59/M	29	Stereostatic biopsy Endoscopic biopsy Incomplete resection	DITIP	=	No	Death (pneumopathy) at 1.9 years	0	‡	Q
	14	60/F	26	Ventriculocisternostomy Complete resection	PPTID	=	No	Alive at 1.7 years	2	+	7

**Table 1.** Clinical and morphological findings in pleomorphic PPT. Abbreviations: HPF = high-power field; NA = not available; ND = not done; 0 = no or very low; + = moderate number; ++ = high

Brain Pathology **18** (2008) 354–359 © 2008 The Authors; Journal Compilation © 2008 International Society of Neuropathology

Cases 1, 2, 3, 6 and 7 have been included in a previous study (9).

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<b>ble 2.</b> List and sources of antibodies used id immunohistochemical results obtained. obreviations: NSE = neuron-specific enolase; (N = synaptophysin; NF = neurofilament;	Antigen	Antibody source	Number of tumors analyzed	- (%)	+/- (%)	+ (%)
ngA = chromogranin A; VIM = vimentin;	NSE	H14 clone, Dako (Trappes, France)	8	0	0	100
FAP = glial fibrillary acidic protein.	SYN	SY38 clone, Dako (Trappes, France)	14	0	0	100
	NF	2F11 clone, Dako (Trappes, France)	14	0	0	100
	ChgA	Dak A3 clone, Dako (Trappes, France)	13	23	8	69
	NeuN	Clone A60, Chemicon, Euromedex (Mundolsheim, France)	5	100	0	0
	CD34	Clone QBEND10, Immunotech (Marseille, France)	2	50	0	50
	S100-P	Biomeda 23/11 (Paris) or Dako (Trappes, France)	12	8	17	75
	VIM	V9 clone, Dako (Trappes, France)	11	36	36	27

6F2 clone, Dako (Trappes, France)

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years and this tumor seems to be observed in younger patients, in accordance with the previously reported case (20). The male predominance in PC and the female predominance in PPTID remain to be verified with a higher number of cases, as no sex predilection has been reported for PPT (15, 16). Tumor recurrence was only observed in two cases. Moreover, the disease progression in the patients presenting with PC could be due to incomplete tumor resection. Based on our previous experience, completeness of excision seems to be the most important factor for avoiding disease progression.

GFAP

Seven cases were diagnosed as PC because of the presence of underlying typical areas with an isomorphic cytology and large fibrillary pineocytomatous rosettes and immunoreactivity for a variety of neuronal markers (8, 9), which make it possible to distinguish these tumors from glioneuronal neoplasms (4, 18, 22), which generally express glial markers. The large number of cells labeled for neuronal markers, including many pleomorphic cells, confirmed previous findings that a neuronal immunophenotype is common in PC (5, 9, 11). Moreover, we previously showed by microarray analysis that two variants of PC, one typical and one pleomorphic, had very similar gene profiles and the two tumors were clearly classified together by cluster analysis based on the entire gene set, notwithstanding the presence of pleomorphic cells (6). The other seven cases, diagnosed as PPTID in our series, were rather similar to the case presented by Sasaki et al (20) and to case

2 reported by Kuchelmeister et al (11). They had moderate proliferative activity, as indicated by only a few mitotic figures and slightly increased Ki67 index, and can be considered as grade II tumors, as reported in the recent PPT classification of the World Health Organisation (16), with a prognosis not as favorable as that of PC. Only one of these cases received radiation therapy and had a 6-year survival, but, as the follow-up of the other PPTID cases was quite short, the benefit of such treatment in preventing local tumor regrowth is still uncertain.

13

The sometimes multinucleated giant cells are thought to be neoplastic and may derive from fusion of mononuclear tumor cells (20). Two types of cells have been described in the neonatal pineal gland (13) and we can hypothesize that both are capable of neoplastic transformation. The above authors showed that the two types of cells differentially express NSE and S100 protein and that the proportion of each type varies throughout postnatal evolution. At birth, the predominant cell type exhibits rosette formation and expresses S100 protein, but not NSE. The number of the second type of cell, which expresses NSE, but not S100 protein, increases gradually with age, and these cells are probably the origin of pineal parenchymal cells. As oncogenesis may recapitulate ontogeny of the human pineal gland (12), both types of cell might be seen in the tumor. Moreover, the presence of another type of cell, which is slightly larger than a pinealocyte and has non-homogeneously condensed nuclear chromatin, has been reported in the fetal pineal



Figure 1. A. Sagittal T1-weighted magnetic resonance imaging (MRI) scan in a patient with pleomorphic pineal parenchymal tumor (case 3). B. Sagittal T1-weighted MRI scan in the same patient after gadolinium injection, showing homogeneous enhancement.



Figure 2. Light microscopic and immunohistochemical features of pleomorphic pineal parenchymal tumors (PPTs). A. Pleomorphic pineocytoma (PC) with pineocytomatous rosettes and giant cells with hyperchromatic nuclei (hemalin phloxine saffron staining). Inset: cytologic imprint of a pleomorphic PC showing tumoral cells with small round nuclei and larger cells with pleomorphic nuclei (toluidine blue staining). B. Pleomorphic PPT of intermediate differentiation (PPTID) with marked cytologic pleomorphism (hemalin eosin staining). Synaptophysin immuno-labeling in a pleomorphic PC (C) and a pleomorphic PPTID (D). Expression of neurofilament in one pleomorphic PC (E) and in one pleomorphic PPTID ( $\mathbf{F}$ ), with intense labeling of giant cells. G. Chromogranin A labeling in a pleomorphic PC. H. CD34 labeling in a pleomorphic PC showing some positive giant cells. I. Nuclear and cytoplasmic S100 protein staining in giant cells of a pleomorphic PC. J. Areas of a pleomorphic PPTID showing nuclei stained for Ki67. Scale bars = 50  $\mu$ m in **A–I**, 100  $\mu$ m in **J**.

gland (14). The pleomorphism in PPT may derive from these different subtypes of cells found in the human pineal gland during development. The expression of S100 protein in some giant cells may be related to the presence of this protein in cells which may represent immature pinealocytes, as seen in the developing pineal gland (13). CD34, expressed in vascular endothelial cells, was also found in giant cells in one PPTID. This expression might correspond to an undifferentiated level of these tumoral cells. Indeed, this marker has been found in neoplastic and malformative lesions, most notably in gangliogliomas (2, 3) and pleomorphic xanthoastrocytomas (19), and seems to be associated with a displastic or non-differentiated cell type.

The pleomorphism encountered in PPT has been described in other low-grade tumors, such as astrocytoma in the pineal gland (17) and hypothalamus (1), pituicytoma (21) and pleomorphic xanthoastrocytoma (7, 10). This marked pleomorphism may lead to upgrading of these tumors and incorrect therapy.

In conclusion, PC and PPTID are rare tumors which can present with marked pleomorphism, potentially leading to the misclassification and upgrading of these tumors. The short postoperative period for some tumors did not allow assessment of the biological behavior of these tumors. However, the lack of invasiveness and the low number of proliferating cells suggest a benign clinical course for these pleomorphic neoplasms.

## ACKNOWLEDGMENTS

We thank the neurosurgeons B. Bataille (Poitiers), P. Cornu (Paris), P. Decq (Créteil), A. Goutelle (Lyon), S. Gaillard (Paris), F. Grisoli (Marseille), C. Lapras (Lyon), P. Mertens (Lyon), K. Mourier (Paris), C. Mottolese (Lyon), G. Robert (Paris), F.E. Roux (Toulouse), J.P. Sichez (Paris) and R. Van Effentere (Paris) for their contribution to the study.

We also thank E. Gros and M. Soutrenon for technical assistance, S. Badet for secretarial assistance, and T. Barkas for linguistic help.

This work was supported by INSERM, the Association for Cancer Research (ARC 3277) and the ADERTU.

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