

Neurovascular Pathways and Alzheimer Amyloid β -peptide

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According to the prevailing amyloid cascade hypothesis, the onset and progression of a chronic neurodegenerative condition in Alzheimer disease (AD) is initiated by the amyloid β -peptide ($A\beta$) accumulation in brain and consequent neuronal toxicity. Recent emphasis on co-morbidity of AD and cerebrovascular disease and the recognition that cerebrovascular dysregulation is an important feature of AD, has shed new light on neurovascular dysfunction as a possible contributor to cognitive decline and Alzheimer neurodegeneration. In the same time, this association has raised a question as to whether there is a causal relationship between cerebrovascular dysregulation and $A\beta$ -initiated pathology, and whether influencing targets in the neurovasculature may prevent different forms of $A\beta$ brain accumulation and/or lower pre-existing accumulates in a later stage of the disease. Pathogenic cascades which operate to dissociate normal transport exchanges between central and peripheral pools of $A\beta$, and decreased vascular competence leading to brain hypoperfusion and impaired $A\beta$ clearance are discussed. We suggest that there is a link between neurovascular dysfunction and elevated brain $A\beta$ which provides a new scenario for therapeutic interventions to control Alzheimer mental deterioration.

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INTRODUCTION

A central feature of Alzheimer disease (AD) is a chronic neurodegenerative process which according to the prevailing amyloid cascade hypothesis is initiated by the amyloid β -peptide ($A\beta$) accumulation in brain and consequent neuronal toxicity (24). The neurofibrillary tangle (NFT) resulting from hyperphosphorylated tau is also a neuropathological feature of AD (55). The study of these pathological traits of AD has dominated the field for the last 3 decades. Indeed, much of the therapeutic armamentarium for AD is directed at developing strategies to control $A\beta$ -related and tau-related pathology in hopes of improving cognitive decline associated with AD (24, 51). It is important to stress, however, that recent emphasis on co-morbidity of AD and cerebrovascular disease (21), the link between AD and atherosclerosis (6, 44), and the realization that cerebrovascular dysregulation is an important feature of AD (3, 26), has shed new light on neurovascular dysfunction as a likely contributor

to dementia and a chronic neurodegenerative condition in AD (60).

Based on a growing body of literature demonstrating that vascular risk factors add to cognitive decline in AD (21), that brain microvascular system is significantly altered in AD (3, 20), and that disrupted cerebral blood flow (CBF) and hypoperfusion to the brain parenchyma are seen in both animal models of AD, as well as in human patients with AD (5, 22, 26, 54), it has been proposed that at least some forms of AD may develop primarily due to cerebrovascular changes and should be classified as vasocognopathies (15). Since the neurovascular unit maintains tightly controlled chemical composition of neuronal internal "milieu" by regulating local CBF and blood-brain barrier (BBB) molecular transport, pathological changes in the vasculature are likely to undermine the normal physiological function of this critical unit (60). However, this also raises a question of whether there is a causal relationship between $A\beta$ accumulates and $A\beta$ -initiated pathology and neurovascular dysfunction, and whether

influencing neurovascular pathways may regulate $A\beta$ brain homeostasis to ultimately lower brain $A\beta$?

IS CEREBRAL β -AMYLOIDOSIS DISORDER OF $A\beta$ CLEARANCE OR $A\beta$ PRODUCTION?

There is little evidence that normal brain aging results in local overexpression of the $A\beta$ precursor protein (APP) and overproduction of $A\beta$ (37). In addition, a relatively small number of AD patients have increased $A\beta$ production in the CNS which is limited to familial forms of the disease caused by inherited mutations in the APP gene nearby the $A\beta$ coding region (ie, Swedish mutation) or presenilins 1 or 2 genes (47). The majority of patients with so-called late-onset "non-genetic" AD, as well as patients with familial forms of cerebrovascular β -amyloidosis, do not have increased $A\beta$ production or increased APP expression in the CNS. Therefore, these patients are likely to exhibit a failure in $A\beta$ clearance from the CNS that could be either due to dysregulated transport mechanisms that control vascular efflux of $A\beta$ across the BBB (61, 63) and/or due to its faulty degradation in the CNS (47). Alternatively, an increased influx of circulating $A\beta$ into the CNS across the BBB may result in accumulation of soluble neurotoxic $A\beta$ in brain interstitial fluid (ISF) and/or its subsequent aggregation and deposition in the CNS (11, 59). In sum, it is likely that cerebrovascular β -amyloidosis in AD develops slowly as a result of a progressive $A\beta$ clearance disorder rather than due to its uncontrolled production from APP. It is also likely that decreased vascular competence associated with reduced CBF and diminished brain capillary surface area available for $A\beta$ efflux from the brain may further amplify the failure of the vascular system to filter $A\beta$ from the brain into the

blood for its systemic clearance and excretion through the liver, kidney and possibly some other excretory organs.

CEREBROVASCULAR FACTOR IN A β ACCUMULATION

The exact physiological role of A β in the brain and in the body during development and aging still remains a mystery. Is A β just a waste product of APP metabolism with powerful cellular toxic actions which require its immediate removal from brain, or alternatively, is A β a regulatory molecule of neurogenesis (57) and cerebral angiogenesis (41, 42) with important roles in development and neurovascular repair? Nevertheless, a number of pathways exist to maintain A β homeostasis precisely in the body and within the CNS. These pathways seem to be well orchestrated during normal physiology, but are completely unsynchronized in diseases associated with dementia and storage of A β in the CNS. It is also interesting to note that amyloid lesions and NFTs can both be triggered by vascular mechanisms such as hypertension and ischemic brain injury (3, 15, 26), which frequently affect elderly individuals with Alzheimer-type dementia (53). It is possible that ischemic brain episodes in AD patients and “mini strokes” develop because of chronic brain hypoperfusion due to atherosclerotic changes in their cerebral arteries (6, 44). These novel findings raise a possibility that a cerebrovascular disorder may precipitate and/or amplify the A β - and tau-related pathology, and therefore could be an important pathogenic mechanism contributing to cognitive decline and neuropathology in AD.

CNS AND SYSTEMIC PATHWAYS REGULATING BRAIN A β

The CNS A β homeostasis is controlled by numerous pathways which interact with each other and may play important roles in early and later stage of the disease process resulting in A β accumulation and/or deposition in brain. These include: *i*) systemic and brain A β production, and its systemic clearance mechanisms; *ii*) rapid regulation of A β soluble brain interstitial fluid (ISF) pool by receptor-mediated transport across the BBB from brain to blood via the low density lipoprotein receptor related protein-1 (LRP1) (10, 16, 48), or under certain pathological conditions from blood

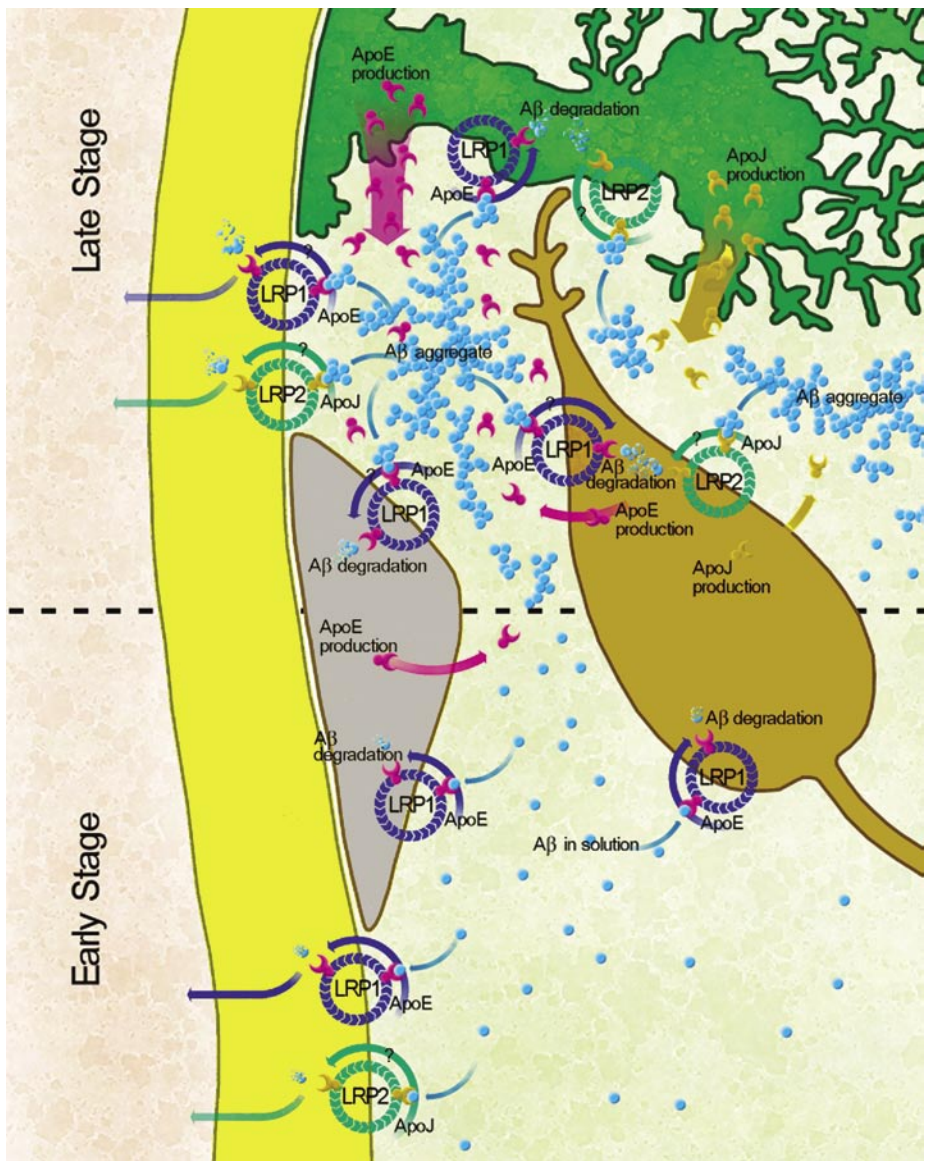


Figure 1. The proposed role of lipoprotein receptors LRP1 and LRP2 and of A β transport proteins ApoE and apoJ in controlling brain A β creates a new scene of potential therapeutic opportunities for AD. **Late stage.** In the late stage of the disease LRP1 on astrocytes could mediate removal of diffuse A β aggregates (30). Astrocytes (green) produce ApoE, an LRP1 ligand, which is also produced to a lesser degree by neuronal cells (brown). Injured astrocytes and neurons in AD also express LRP2 and produce apoJ, an LRP2 ligand (32). Deletion of the apoJ gene has remarkable effect on development of A β -related pathology in APP mice lacking the ApoE gene (18) suggesting its probable major role in clearing A β . Whether ApoE secreted by astrocytic microdomains nearby the endothelial blood-brain barrier (BBB) (yellow) can co-localize diffuse capillary A β aggregates for clearance via LRP1 at the BBB, and/or whether apoJ may co-localize A β aggregates for clearance via LRP2 which is expressed at the BBB (62), is not known. There is also a possibility that vascular smooth muscle cells and pericytes (gray) which are in close proximity to the endothelial barrier, and express LRP1 and secrete ApoE and apoJ (25), may help clearing diffuse A β aggregates. The role of LRP1 and LRP2 on neurons with respect to clearing aggregated and/or oligomeric A β is not known, except for the fact that oligomeric A β species are extremely neurotoxic (24). **Early stage.** Before significant A β accumulates develop, LRP1 at the BBB can directly clear soluble A β (10), whereas LRP1 on neurons most likely regulates the intracellular vs. extracellular redistribution of soluble A β in brain and does not have an effect on net clearance from brain (58). How ApoE and apoJ binding to A β influences clearance of their respective complexes at the BBB still remains unknown.

to brain via the receptor for advanced glycation end products (RAGE) (12); *iii*) control of soluble A β pool sizes in the extracellular body fluids and plasma, and in brain ISF and cerebrospinal fluid (CSF) via A β binding proteins, eg, apolipoprotein E

(ApoE), apoJ, α 2-macroglobulin (α 2M), transthyretin, and albumin (18, 19, 51), which may also regulate transport exchanges of their respective complexes with A β across the BBB and blood-CSF barrier (51, 62); *iv*) A β metabolism by different

systemic and brain enzymes including neprilysin (29), insulin degrading enzyme, coagulation-related enzymes (eg, plasmin, tissue plasminogen activator), or different matrix metalloproteinases (47); *v*) removal of deposited A β by different brain cell types including astrocytes (56) which may depend on ApoE-directed co-localization of deposits (30) and via microglia (1); *vi*) slow removal of A β via the ISF-CSF bulk flow (50); and *vii*) oligomerization and aggregation of A β alone and/or with its binding proteins (24, 51). Figure 1 summarizes a hypothetical diagram on some selected lipoprotein receptors-mediated pathways in different brain cells and of their ligands/A β binding proteins, which could potentially be involved in removal of aggregated A β and/or soluble A β from the brain and across the BBB.

The importance of A β exchanges between cerebral vascular compartment and brain ISF compartment has been well documented by several recent studies in animal models of AD-like brain β -amyloidosis. For example, it has been shown that A β efflux across the BBB in AD mice can predict the volume of brain amyloid burden (16), whereas the development of senile plaques directs the A β transport equilibrium towards the brain (17). Similar findings have been reported in aged non-human primates which develop either cerebral β -amyloidosis (Rhesus monkey) (38) or cerebral β -amyloid angiopathy, CAA (Squirrel monkey) (2).

A recent study reported that the apoJ gene is essential for preventing A β accumulation and deposition in APP overexpressing mice (18). Deletion of the apoJ gene accelerated A β -related pathology in APP mice lacking the ApoE gene (18), raising the possibility that apoJ could be a major apolipoprotein mediating A β efflux from brain. The role of apoJ in blood-to-brain and brain-to-blood transport exchanges of A β is still unclear. An earlier report suggested A β complexed to apoJ in the circulation may utilize the lipoprotein receptor at the BBB, gp330/megalin or LRP2, for its transport into brain ISF and/or CSF across the BBB and blood-CSF barrier of the choroid plexus epithelium, respectively (62). It has been noted that LRP2 at the BBB is completely saturated by relatively high levels of circulating apoJ ($\sim 1 \mu\text{M}$); whereas, the in vivo determined half-saturation

transport constants (K_m) at the BBB for A β 40-apoJ complexes were in the range $<1 \text{ nM}$ (62). It remains unknown whether LRP2 can clear A β -apoJ complexes across the BBB and the blood-CSF barrier into the blood, nor whether apoJ is as effective in transporting the pathogenic A β 42 across the biological membranes as it is in mediating A β 40 trafficking. On the other hand, it is not clear whether LRP2 expressed on different brain cells in AD brains, including injured neurons and astrocytes (32), could be involved in the removal of aggregated A β by the apoJ-directed co-localization in a fashion similar to that reported for the ApoE/LRP1-mediated removal of aggregated A β via astrocytes (30). Whether vascular smooth muscle cells and pericytes which express LRP1 and secrete ApoE (25) have a role in removing aggregated A β , and can brain endothelial cells can be as effective in removing A β aggregates as they are in transporting soluble A β from brain to blood is not known (Figure 1).

It has also been suggested that P-glycoprotein at the luminal side of the BBB may reduce the levels of A β in the brain endothelial cells by promoting its efflux into blood (33). Our work based on A β 40 transport kinetics has suggested that RAGE and LRP1 are dominant transport pathways creating almost instantaneous changes in the levels of soluble A β in brain ISF by regulating its rapid bi-directional trafficking across the BBB (11, 51).

RAGE, a multiligand receptor in the immunoglobulin superfamily, mediates transcytosis of A β across the BBB under pathologic conditions associated with a neuroinflammatory response, increased cytokine expression and oxidant stress, and neuronal co-localization of circulating A β (12).

LRP1, an endocytic and signaling multiligand receptor for several molecules associated with development of AD pathology including A β , ApoE, $\alpha 2\text{M}$ and APP, is linked to AD genetically and biochemically through regulation of APP processing and metabolism and A β clearance (11, 25, 51). Originally, it has been suggested that LRP1 on neurons may clear A β , but more recent work in APP mice overexpressing the LRP1 mini-receptor on neurons demonstrated accumulation of soluble A β in brain (51), arguing against the role of neuronal LRP1 in A β clearance. Since LRP1 binds and in-

ternalizes free A β (10), it is possible that neuronal LRP1 plays a role in distributing soluble A β between the intracellular versus extracellular neuronal milieu, but without a net effect on A β clearance from brain. In contrast to neurons, LRP1 on brain capillary endothelium interacts directly with free A β and clears preferentially A β 40 into the blood, whereas the affinity of other A β isoforms for LRP1 is greatly reduced with an increasing β -sheet content in A β compared to A β 40 (10, 48). These findings suggest that low affinity of certain soluble A β species for LRP1 at the BBB may result in accumulation of these A β isoforms in brain, as recently demonstrated in the Dutch/Iowa transgenic APP model (9, 10). Whether LRP1 on brain endothelium may clear aggregated A β via an ApoE-dependent mechanism as shown for astrocytes (30), or via an apoJ-dependent co-localization to LRP2 on vascular endothelial cells, is not known. Whether brain endothelium can clear oligomeric forms of A β , and whether chaperone proteins can influence BBB clearance of oligomeric A β is not known.

THERAPEUTIC INTERVENTIONS FOR AD

The core of research for potential breakthrough therapeutic interventions to control cognitive decline in AD patients has been and is still centered on A β , yet we are awaiting for an approved A β -based therapy for this devastating brain disorder. Currently approved therapies for AD are mainly symptomatic directed at either compensating for the neurotransmitter deficit in brain resulting from neuronal and synaptic loss, or at effecting the blood flow to the brain, and/or at controlling certain symptomatic neurological features of the disease. However, no known treatment can arrest the progression of AD.

Approved therapies. In addition to Aricept, there are 4 other approved therapies for AD. For some people in the early and middle stages of the disease, the drugs tacrine (Cognex), rivastigmine (Exelon), or galantamine (Reminyl) may help slowing down the progression of symptoms for a limited time. While each of these drugs is regarded as commercially successful, it is difficult to distinguish between them in terms of efficacy since they all have a similar mechanism of action, ie, the inhibition of acetyl cholinesterase activity (15, 52). A re-

cently approved new drug memantine (Namenda) marketed by Forest Laboratories Inc, is an N-methyl D-aspartate (NMDA) antagonist. The main effect of memantine is to delay progression of the symptoms of moderate to severe AD which may allow some patients to maintain certain daily functions a little longer (52). Other drugs not initially targeted or labeled for AD patients may help controlling some behavioral symptoms of AD such as sleeplessness, agitation, wandering, anxiety, and depression. Treating these symptoms often makes patients more comfortable and makes their care easier. Again, these drugs do not arrest or alter the progression of the disease.

A β -based experimental therapies. The prevailing concept states deficient A β clearance has a major role in late onset, non-genetic AD, (>98% of AD cases) (11, 13, 45, 47, 51, 59). A β clearance strategies had remarkable results in preclinical immunization trials (46), but were less conclusive in humans (40), although improvements in cognitive decline have been reported. The A β clearance pathways delineate new therapeutic opportunities including activation of several A β -degrading enzymes (47, 51), which may help preventing A β accumulation. Strategies to directly enhance the binding of A β to LRP1 (10) or to its lipoprotein receptors ligands/A β chaperones (51), may offer promising therapeutic avenues in future giving the prevailing role of LRP1 in A β clearance across the BBB (10, 48) and on astrocytes (30). It has been shown that some agents, eg, statins (11) and proteasome inhibitors (10), may increase LRP1 expression at the BBB and perhaps enhance its activity. It has also been shown that blocking A β /RAGE interaction at the BBB by systemic treatment with soluble form of the receptor (sRAGE) prevents A β accumulation in PDAPP mice if treatment is initiated before 6 months of age (12). This effect of sRAGE is most likely created through a peripheral binding of A β in the circulation which in turn would allow efflux mechanisms to prevail. In addition, blocking A β /RAGE interaction may help increase LRP1 levels at the BBB (11). It is important to note that the efficacy of many A β peripheral binding agents, eg, immunoglobulins (16, 46, 49), sRAGE (12), sLRP1 (10), gelsolin (39), may critically depend on the availability of clearance mechanisms

in the brain, which depend on the levels of LRP1 at the BBB. The LRP1 levels in brain capillaries, however, are substantially reduced in APP overexpressing animals at 9 to 12 months of age and in late-stage AD patients and Dutch patients with cerebrovascular β -amyloidosis (10). Therefore, early treatment with A β -binding agents may be a key to success of such peripheral A β sequestration-based therapies. Metal chelators may destabilize β -amyloid deposits and attenuate cognitive decline in AD patients (51).

In addition to the clearance strategies, considerable efforts by major pharmaceutical companies in the past decade have been focused on developing inhibitors of γ -secretase and β -secretase, enzymes responsible for proteolytic cleavage of A β -precursor protein (APP) and formation of A β (43, 47). Studies with secretase inhibitors have demonstrated that inhibiting APP production in transgenic models designed to overproduce A β in brain will result in improved cognitive decline and reduced A β pathology (43). If these strategies were able to influence normal A β production at physiological APP expression levels in brain, they could also have the potential to reduce brain A β in late onset AD patients, who do not have increased APP levels in brain. However, as with all strategies for A β as discussed above, the side effects of the inhibition of secretases would still remain, at least for the present, an important concern.

Finally, studies have used small molecule libraries to screen for compounds that either interfere with assembly of A β particles into fibrils (14, 34) or disaggregate existing fibrils (4).

Tau-based experimental therapies. Since the microtubule-associated tau protein is hyperphosphorylated in NFT, efforts have been directed towards interfering with the phosphorylation of tau either by inhibiting various protein kinases or promoting phosphatase activities (28, 35). High-throughput screening assays are currently underway to define drugs that interfere with the hyperphosphorylated tau, as well as its associated sequestration of normal tau (27). A recent report described a small molecule (N744, <700 Da) that dose-dependently inhibited tau filament nucleation and fibrillization in

vitro, making this a promising candidate to test in animal models (8).

Neurovascular protection strategies. We now appreciate that neurovascular dysfunction is a major hallmark of AD (15, 26, 60). Brain ischemia and athero-thrombotic disorder could therefore be important therapeutic targets in AD (6, 44). Agents which exhibit direct cytoprotective activity on stressed and ischemic brain endothelium and neurons, and have substantial anti-inflammatory and pro-angiogenic activity, such as the serine protease activated protein C (7, 23, 36), may hold the potential to protect the neurovascular unit from combined A β , hypoxic and excitotoxic injuries as seen in AD. Altered physiology of vascular smooth muscle cells and altered brain endothelium in AD could be important therapeutic targets for controlling arterial and brain capillary disorder in AD, but at present their role is still poorly understood.

CONCLUSIONS

The accumulating body of evidence suggests that neurodegenerative disorder in AD may have a complex multifactorial origin which may not necessarily be primarily centered on neurons and/or originate from a primary neuronal disorder. It became evident that in contrast to valuable animal models of AD-like cerebral β -amyloidosis, which are created by overexpressing APP in neurons associated with A β neuronal overproduction, the actual disease in patients does not mimic this situation in models since the accumulation of A β in late onset AD is not related to APP neuronal overproduction (47, 51, 59). Moreover, studies demonstrating that vascular dysregulation and brain hypoperfusion may result in increased APP expression/A β production and tau pathology (3, 26) further support the view that observed neuronal changes may be rather secondary to cerebrovascular problems associated with AD. Recent experimental studies in transgenic mice producing low levels of poorly cleared vasculotropic Dutch/Iowa A β mutants confirmed impaired vascular clearance of A β across the BBB leads to an increased A β brain capillary deposition and formation of amyloid lesions (9, 10). It has also been suggested that dense A β plaques in Alzheimer Tg2576 APP overexpressing mice and double transgenic PS1/APP Tg2576 mice

(31), as well as in AD brains (60), may have vascular origin.

At this point, we would argue that a more in depth understanding of multiple neurovascular cascades that operate to dissociate normal neurovascular coupling, induce brain hypoperfusion and reduce vascular competence resulting in impaired A β clearance from brain may open new avenues to link neurovascular dysfunction to A β brain accumulation and AD pathology. Although numerous pathways may initially precipitate the onset of the disease, at a later stage in the disease process, these cascades may work synergistically to form interrelated and irreversible pathogenic circles to devastate the brain. For example, A β oligomers are directly neurotoxic (24), whereas A β aggregates are anti-angiogenic (41, 42), synergistically destroying the neurovascular unit leaving brain without its important clearance mechanism. Therefore, combined therapies to enhance A β clearance, improve neurovascular repair and protect brain cells from different types of stress may hold potential to control cognitive dysfunction associated with AD.

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REFERENCES

1. Bacskai BJ, Kajdasz ST, McLellan ME, Games D, Seubert P, Schenk D, Hyman BT (2002). Non-Fc-mediated mechanisms are involved in the clearance of amyloid- β in vivo by immunotherapy. *J Neurosci* 22:7873-7878.
2. Bading JR, Yamada S, Mackic JB, Kirkman L, Miller C, Calero M, Ghiso J, Frangione B, Zlokovic BV (2002) Brain clearance of Alzheimer's amyloid- β 40 in the squirrel monkey: a SPECT study in a primate model of cerebral amyloid angiopathy. *J Drug Target* 10:359-368.
3. Bailey TL, Rivara CB, Rocher AB, Hof PR (2004) The nature and effects of cortical microvascular pathology in aging and Alzheimer's disease. *Neurol Res* 26:573-578.
4. Blanchard BJ, Chen A, Rozeboom LM, Stafford KA, Weigle P, Ingram VM (2004) Efficient reversal of Alzheimer's disease fibril formation and elimination of neurotoxicity by a small molecule. *Proc Natl Acad Sci U S A* 101:14326-14332.
5. Bookheimer SY, Strojwas MH, Cohen MS, Saunders AM, Pericak-Vance MA, Mazziotta JC, Small GW (2000) Patterns of brain activation in people at risk for Alzheimer's disease. *N Engl J Med* 343:450-456.
6. Casserly I, Topol E (2004) Convergence of atherosclerosis and Alzheimer's disease: inflammation, cholesterol, and misfolded proteins. *Lancet* 363:1139-1146.
7. Cheng T, Liu D, Griffin JH, Fernandez JA, Castellino F, Rosen ED, Fukudome K, Zlokovic BV (2003) Activated protein C blocks p53-mediated apoptosis in ischemic human brain endothelium and is neuroprotective. *Nat Med* 9:338-342.
8. Chirita C, Necula M, Kuret J (2004) Ligand-dependent inhibition and reversal of tau filament formation. *Biochemistry* 43:2879-2887.
9. Davis J, Xu F, Deane R, Romanov G, Previti ML, Zeigler K, Zlokovic BV, Van Nostrand WE (2004) Early-onset and robust cerebral microvascular accumulation of amyloid β -protein in transgenic mice expressing low levels of a vasculotropic Dutch/lowa mutant form of amyloid β -protein precursor. *J Biol Chem* 279:20296-20306.
10. Deane R, Wu Z, Sagare A, Davis J, Yan SD, Hamm K, Xu F, Parisi M, LaRue B, Hu HW, Spijkers P, Guo H, Song X, Lenting PJ, Van Nostrand WE, Zlokovic BV (2004) LRP/amyloid β -peptide interaction mediates differential brain efflux of a β isoforms. *Neuron* 43:333-344.
11. Deane R, Wu Z, Zlokovic BV (2004) RAGE (Yin) versus LRP (Yang) balance regulates Alzheimer amyloid β -peptide clearance through transport across the blood-brain barrier. *Stroke* 35[suppl 1]:2628-2631.
12. Deane R, Yan SD, Subramanian RK, LaRue B, Jovanovic S, Hogg E, Welch D, Manness L, Lin C, Yu J, Zhu H, Ghiso J, Frangione B, Stern A, Schmidt AM, Armstrong DL, Arnold B, Liliensiek B, Nawroth P, Hofman F, Kindy M, Stern D, Zlokovic BV (2003) RAGE mediates amyloid- β peptide transport across the blood-brain barrier and accumulation in brain. *Nat Med* 9:907-913.
13. De Felice FG, Ferreira ST (2002) Beta-amyloid production, aggregation, and clearance as targets for therapy in Alzheimer's disease. *Cell Mol Neurobiol* 22:545-563.
14. De Felice FG, Vieira MN, Saraiva LM, Figueroa-Villar JD, Garcia-Abreu J, Liu R, Chang L, Klein WL, Ferreira ST (2004) Targeting the neurotoxic species in Alzheimer's disease: inhibitors of Abeta oligomerization. *FASEB J* 18:1366-1372.
15. de la Torre JC (2004). Alzheimer's disease is a vasocognopathy: a new term to describe its nature. *Neurol Res* 26:517-524.
16. DeMattos RB, Bales KR, Cummins DJ, Paul SM, Holtzman DM (2002) Brain to plasma amyloid-efflux: a measure of brain amyloid burden in a mouse model of Alzheimer's disease. *Science* 295:2264-2267.
17. DeMattos RB, Bales KR, Parsadanian M, O'Dell MA, Foss EM, Paul SM, Holtzman DM (2002) Plaque-associated disruption of CSF and plasma amyloid- β equilibrium in a mouse model of Alzheimer's disease. *J Neurochem* 81:229-236.
18. DeMattos RB, Cirrito JR, Parsadanian M, May PC, O'Dell MA, Taylor JW, Harmony JAK, Aronow BJ, Bales KR, Paul SM, Holtzman DM (2004) ApoE and clusterin cooperatively suppress a β levels and deposition: evidence that ApoE regulates extracellular a β metabolism in vivo. *Neuron* 41:193-202.
19. Fagan AM, Watson M, Parsadanian M, Bales KR, Paul SM, Holtzman DM (2002) Human and murine ApoE markedly influence a β metabolism before and after plaque formation in a mouse model of Alzheimer's disease. *Neurobiol Dis* 9:305-318.
20. Farkas E, Luiten PG (2001) Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol* 64:575-611.
21. Gorelick PB (2004) Risk factors for vascular dementia and Alzheimer's disease. *Stroke* 35:2620-22.
22. Greenberg SM, Gurol ME, Rosand J, Smith EE (2004) Amyloid angiopathy-related vascular cognitive impairment. *Stroke* 35:2616-2619.
23. Guo H, Liu D, Gelbard H, Cheng T, Insalaco R, Fernandez JA, Griffin JH, Zlokovic BV (2004) Activated protein C prevents neuronal apoptosis via protease activated receptors 1 and 3. *Neuron* 41:563-572.
24. Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 297:353-356.
25. Herz J, Hui DY (2004) Lypoprotein receptors in the vascular wall. *Curr Opin Lipidol* 15:175-181.
26. Iadecola C (2004) Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Neurosci Rev* 5:347-360.
27. Iqbal K, Alonso Adel C, El-Akkad E, Gong CX, Haque N, Khatoun S, Pei JJ, Tanimukai H, Tsujio I, Wang JZ, Grundke-Iqbal I (2003) Alzheimer neurofibrillary degeneration: therapeutic targets and high-throughput assays. *J Mol Neurosci* 20:425-429.
28. Iqbal K, Grundke-Iqbal I (2004) Inhibition of neurofibrillary degeneration: a promising approach to Alzheimer's disease and other tauopathies. *Curr Drug Targets* 5:495-502.
29. Iwata N, Tsubuki S, Takaki Y, Shirota N, Lu B, Gerard NP, Gerard C, Hama E, Lee H-J, Saido TC (2001) Metabolic regulation of brain A β by neprilysin. *Science* 292:1550-1552.
30. Koistinaho M, Lin S, Wu X, Esterman M, Koger D, Hanson J, Higgs R, Liu F, Malkani S, Bales KR, Paul SM (2004) Apolipoprotein E promotes astrocyte colocalization and degradation of deposited amyloid- β peptides. *Nat Med* 10:719-726.
31. Kumar-Singh S, Dickson D, Pirici D, Serneels S, McGowan E, Duff K, Hardy J, Van Broeckhoven C (2004) Dense-core amyloid plaques in Tg2576 and PSAPP mice are centered on vascular wall and closely resemble Flemish Alzheimer's pathology. *Soc Neurosci Ann Meeting*, 23-27 October San Diego, CA.
32. LaFerla FM, Troncoso JC, Strickland DK, Kawas CH, Jay G (1997) Neuronal cell death in Alzheimer's disease correlates with ApoE uptake and intracellular A-beta stabilization. *J Clin Invest* 100:310-320.
33. Lam FC, Liu R, Lu P, Shapiro AB, Renoir J-M, Sharom FJ, Reiner PB (2001) β -Amyloid efflux me-

- diated by p-glycoprotein. *J Neurochem* 76:1121-1128.
34. Lashuel HA, Hartley DM, Balakhaneh D, Aggarwal A, Teichberg S, Callaway DJ (2002) New class of inhibitors of amyloid-beta fibril formation. Implications for the mechanism of pathogenesis in Alzheimer's disease. *J Biol Chem* 277:42881-42890.
35. Lau LF, Schachter JB, Seymour PA, Sanner MA (2002) Tau protein phosphorylation as a therapeutic target in Alzheimer's disease. *Curr Top Med Chem* 2:395-415.
36. Liu D, Cheng T, Guo H, Fernandez JA, Griffin JH, Song X, Zlokovic BV (2004) Tissue plasminogen activator neurovascular toxicity is controlled by activated protein C. *Nat Med* 10:1379-1383.
37. Lu T, Pan Y, Kao SY, Li C, Kohane I, Chan J, Yankner BA (2004) Gene regulation and DNA damage in the ageing human brain. *Nature* 429:883-891.
38. Mackic JB, Bading J, Ghiso J, Walker L, Wisniewski T, Frangione B, Zlokovic BV (2002) Circulating amyloid- β peptide crosses the blood-brain barrier in aged monkeys and contributes to Alzheimer's disease lesions. *Vascul Pharmacol* 38:303-313.
39. Matsuoka Y, Saito M, LaFrancois J, Saito M, Gaynor K, Olm V, Wang L, Casey E, Lu Y, Shiratori C, Lemere C, Duff K (2003) Novel therapeutic approach for the treatment of Alzheimer's disease by peripheral administration of agents with an affinity to β -amyloid. *J Neurosci* 23:29-33.
40. Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO (2003). Neuropathology of human Alzheimer disease after immunization with amyloid- β peptide: a case report. *Nat Med* 9:448-452.
41. Paris D, Patel N, DelleDonne A, Quadros A, Smeed R, Mullan M (2004) Impaired angiogenesis in a transgenic mouse model of cerebral amyloidosis. *Neurosci Lett* 366:80-85.
42. Paris D, Townsend K, Quadros A, Humphrey J, Sun J, Brem S, Wotoczek-Obadia M, DelleDonne A, Patel N, Obregon DF, Crescentini R, Abdullah L, Coppola D, Rojiani AM, Crawford F, Sebt SM, Mullan M (2004) Inhibition of angiogenesis by A β peptides. *Angiogenesis* 7:75-85.
43. Roberts SB (2002) β -Secretase inhibitors and Alzheimer's disease. *Adv Drug Deliv Rev* 54:1579-1588.
44. Roher AE, Esh C, Rahman A, Kokjohn TA, Beach TG (2004) Atherosclerosis of cerebral arteries in Alzheimer's disease. *Stroke* 35:2623-2627.
45. Sacchettini JC, Kelly JW (2002) Therapeutic strategies for human amyloid diseases. *Nat Rev Drug Discov* 1:267-275.
46. Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandeventer C, Walker S, Wogulis M, Yednock T, Games D, Seubert P (1999). Immunization with amyloid- β attenuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature* 400:173-177.
47. Selkoe DJ (2001) Clearing the brain's amyloid cobwebs. *Neuron* 32:177-180.
48. Shibata M, Yamada S, Kumar SR, Calero M, Bading J, Frangione B, Holtzman DM, Miller CA, Strickland DK, Ghiso J, Zlokovic BV (2000) Clearance of Alzheimer's amyloid- β_{1-40} peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J Clin Invest* 106:1489-1499.
49. Sigurdsson EM, Scholtzova H, Mehta PD, Frangione B, Wisniewski T (2001) Immunization with a nontoxic/nonfibrillar amyloid-beta homologous peptide reduces Alzheimer's disease-associated pathology in transgenic mice. *Am J Pathol* 159:439-447.
50. Silverberg GD, Mayo M, Saul T, Rubenstein E, McGuire D (2003) Alzheimer's disease, normal-pressure hydrocephalus, and senescent changes in CSF circulatory physiology: a hypothesis. *Lancet Neurol* 2:506-511.
51. Tanzi RE, Moir RD, Wagner SL (2004) Clearance of Alzheimer's A β peptide: the many roads to perdition. *Neuron* 43:605-608.
52. Tariot PN, Farlow MR, Grossberg GT, Graham SM, McDonald S, Gergel I, Memantine Study Group (2004) Memantine treatment in patients with moderate to severe Alzheimer disease already receiving donepezil: a randomized controlled trial. *JAMA* 291:317-324.
53. Vermeer SE, Prins ND, den Heijer T, Hofman A, Koudstaal PJ, Breteler MMB (2003) Silent brain infarcts and the risk of dementia and cognitive decline. *N Engl J Med* 348:1215-1222.
54. Vinters HV, Farag ES (2003) Amyloidosis of cerebral arteries. *Adv Neurol* 92:105-112.
55. Wood JG, Mirra SS, Pollock NJ, Binder LI (1986) Neurofibrillary tangles of Alzheimer disease share antigenic determinants with the axonal microtubule-associated protein tau (tau). *Proc Natl Acad Sci U S A* 83:4040-4043.
56. Wyss-Coray T, Loike JD, Brionne TC, Lu E, Anankov R, Yan F, Silverstein SC, Husemann J (2003) Adult mouse astrocytes degrade amyloid- β in vitro and in situ. *Nat Med* 9:453-457.
57. Yankner BA, Duffy LK, Kirschner DA (1990) Neurotrophic and neurotoxic effects of amyloid-beta protein: reversal by tachykinin neuropeptides. *Science* 250:279-282.
58. Zerbinatti CV, Wozniak DF, Cirrito J, Cam JA, Osaka H, Bales KR, Zhuo M, Paul SM, Holtzman DM, Bu G (2004) Increased soluble amyloid β peptide and memory deficits in amyloid model mice overexpressing the LDL receptor-related protein. *Proc Natl Acad Sci U S A* 101:1075-1080.
59. Zlokovic BZ (2004) Clearing amyloid through the blood-brain barrier. *J Neurochem* 89:807-811.
60. Zlokovic BV (2005) Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci* in press.
61. Zlokovic BV, Frangione B (2003) Transport-clearance hypothesis for Alzheimer's disease and potential therapeutic implications. In: Saido TC, ed. *A β Metabolism in Alzheimer's Disease*. Georgetown, TX: Landes Bioscience 114-122.
62. Zlokovic BV, Martel CL, Matsubara E, McComb JG, Zheng G, McCluskey RT, Frangione B, Ghiso J (1996) Glycoprotein 330/megalin: probable role in receptor-mediated transport of apolipoprotein J alone and in a complex with Alzheimer's disease amyloid- β at the blood-brain and blood-cerebrospinal fluid barriers. *Proc Natl Acad Sci U S A* 93:4229-4234.
63. Zlokovic BV, Yamada S, Holtzman D, Ghiso J, Frangione B (2000) Clearance of amyloid-A β -peptide from brain: transport or metabolism? *Nature Med* 6:718-719