Viruses and Rickettsiae

Margaret M Esiri

Department of Neuropathology and Clinical Neurology, Radcliffe Infirmary, Oxford

In tiffs review I shall try to provide a brief, up-todate, account of the neuropathology of those viral and rickettsial diseases that are particularly prevalent in tropical regions. These diseases are not, however, exclusive to the tropics. Some, such as AIDS, are common in temperate regions as well, though others are closer to being exclusively tropical, such as some of the arthropod-borne (ARBO) virus encephalides. The latter are dependent for their dissemination on an existence during part of their infectious cycle in insects which are, in turn, climatically and seasonally sensitive. This necessarily limits their geographical distribution. Factors that influence some of the other diseases are less closely dependent on climate and geography and reflect more the social or cultural conditions under which people live. Thus, diseases that depend for their spread on forms of human behavior such as promiscuity or drug abuse (AIDS), or poor hygiene and living conditions (polio, rickettsial diseases) or on contact with domestic and other animals (rabies) may occur in a more widespread distribution, for the tropics are not the only places that afford opportunities for these diseases to flourish.

I shall select for discussion aspects of the pathology of these diseases that are currently undergoing investigation but will aim to present these against the backdrop of more established aspects of their pathology. Recent reviews of the pathology of viral encephalitis can be found in Hamilton and Wiley (33) and Esiri and Kennedy (20) and of HIV-1 infection in Price & Sidtis (78) and Scaravilli (85).

Retrovirus Infections

Retroviruses are enveloped RNA viruses which replicate in cells by means of a reverse transcriptase enzyme that allows synthesis of a complementary

Corresponding author:

e-mail: mmesiri@clneuro.ox.ac.uk

DNA which is integrated into the host cell DNA. They remain in the cell for its lifetime and cause lifetime infections in humans.

The recently investigated retroviral infections human immunodeficiency virus (HIV) - 1, HIV-2 and human T lymphotropic virus (HTLV) - 1 are those to be discussed here. HIV-1 is the cause of the world epidemic of AIDS, first reported in the USA in 1981. HIV-2 is a similar but molecularly and serologically distinct virus from HIV-1 which has been described as a pathogen principally in West Africa. HTLV-1 is a virus that causes endemic infection in many parts of the world, but particularly in the tropics, which causes adult T cell leukemia and a form of chronic myelopathy.

HIV-1 Infection and AIDS

HIV-1 infection is the most common central nervous system (CNS) viral infection in tropical countries. There are estimated to be over 18 million adults and 1.5 million children infected with this virus world-wide (103). About 3 million of these people are living in sub-Saharan Africa and 4 million in South and South-East Asia. Over 6 million of those infected with HIV-1 have developed AIDS, the clinical manifestation of HIV-1 infection and its consequences, and AIDS is now the commonest cause of death in those aged 15-49 years in many African localities (80).

HIV-1 infection is associated with a remarkably wide range of neuropathological changes, some of which are due to opportunistic infections resulting from the failure of immune function which HIV-1 infection promotes. Others are neoplastic (lymphoma) or directly attributable to the effects of HIV-1 on the nervous system. It is only with the direct effects of HIV-1 infection that this review is concerned. There are three different manifestations of HIV-1 infection to consider: HIV-1 encephalitis, myelopathy and peripheral neuropathy. For the most part, all are seen relatively late in the course of HIV-1 infection. While the prevalence of opportunistic CNS infections in AIDS sufferers in the tropics is quite high and comparable to that in Western countries the prevalence of HIV-1 encephalitis in tropical countries is lower than in the West, probably because fewer of the systemic infections that occur in AIDS are treated effectively and patients therefore die

Margaret M. Esiri, Neuropathology Dept, Radcliffe Infirmary, Oxford OX2 6HE, UK, Tel.: 01865 224403 Fax: 01865 224508

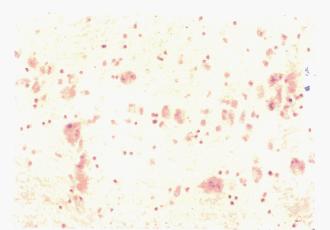


Figure 1. Focus of macrophages and macrophage-derived multinucleated cells in the basal ganglia from a case of HIV-1 encephalitis. H & E stain.



Figure 2. Myelin-stained section through the frontal and anteror temporal lobes of a case of HIV-1 encephalitis. Note diffuse pallor of myelin in the frontal lobe white matter. Luxol fast blue stain. (Reproduced with permission from Esiri MM. Oppenheimer's Diagnostic Neuropathology, 2nd Ed. Blackwell Science 1996).

at an earlier stage of the disease, before HIV-1 encephalitis has had an opportunity to develop. Thus, HIV-1 encephalitis was seen in only 3% of sub-

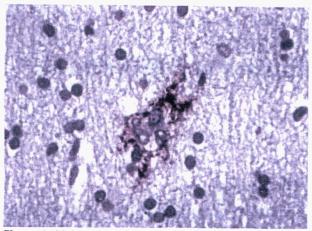


Figure 3. Multinucleated giant cell in cerebral white matter containing HIV-1 p24 core protein antigen (red reaction product). Haematoxylin counterstain. (Immunostained section prepared by Dr CS Morris).

jects with AIDS undergoing autopsy in Abidjan, West Africa (60) compared with 23-41% in recent autopsy series in Europe (9, 61, 63).

A recent prospective study in the USA showed that the incidence of clinical AIDS dementia depended on the severity of depression of the CD4 lymphocyte count and ranged from 7.3 to 0.5 cases per 100 person years (5). About 20% of cases of advanced AIDS in Western countries can be expected to develop dementia. The prevalence of HIV-1 encephalitis at autopsy varies depending on whether HIV-infection has been treated (9, 32, 61).

No reliable figures for the prevalence of myelopathy and peripheral neuropathy complicating HIV-1 infection in tropical countries are available. In selected series from the USA the prevalence of myelopathy has been estimated as 2-22%, while in an unselected prospective study the incidence of sensory peripheral neuropathy was found to be 1.5/100 person years (5).

HIV-1 Encephalitis

Early in the course of the AIDS epidemic it was noted that HIV-1 can be detected in the brains from some sufferers (89). The presence of the virus in the brain in adults is associated with the clinical development of a diffuse encephalopathy termed the AIDS dementia complex (77) or HIV-1 - associated cognitive/ motor complex (42). This usually occurs towards the terminal stages of infection but it is not seen in all subjects with the pathology of HIV-1 encephalitis. Death usually occurs within 6 months of the development of severe symptoms. Infants and children infected with HIV-1 before or at the time of birth or later are also susceptible to developing HIV-1 encephalitis (88).

HIV-1 encephalitis presents a unique pathological picture. Because the patients are severely immunosuppressed by the effects of HIV-1 on the immune

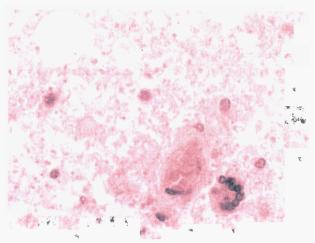


Figure 4. HIV-1 myelopathy. Lateral column white matter showing vacuolar change and a multinucleated macrophage adjacent to a blood vessel. H & E stain.

system there is remarkably little lymphocyte infiltration in the brain and the pathology can be overlooked if it is not specifically sought. There is a diffuse or multifocal accumulation of microglia, macrophages and, in most cases, macrophagederived multinucleated cells (Fig 1). These are found in a widespread distribution in the brain but with a predilection for the cerebral white matter and deep grey matter. The white matter, in addition, shows diffuse myelin pallor (Fig 2) and reactive astrocytosis. HIV-1 structural proteins can be demonstrated immunocytochemically in multinucleated cells, macrophages and microglia (Fig 3). Because multinucleated cells are not always evident and the other pathology is non-specific, HIV-1 encephalitis cannot be definitely excluded without performing immunocytochemical or in situ hybridization reactions for detection of HIV-1. In addition to these alterations in white matter and deep grey matter that are clearly associated with the local presence of virus, more subtle changes not so clearly localized to sites of demonstrable HIV-1 infection, have also been described. Thus, pyramidal neuron loss has been found in parts of the cerebral cortex (21, 22, 101, 105). Subcortical nuclei including the cerebellar dentate nuclei and inferior olives have also been shown to lose neurons (1). These studies of neuron loss have relied on morphometric analysis and simple inspection of sections rarely shows clear evidence of cell loss. In cerebral cortex reduction of dendritic spines on cortical neurons, reduction in synaptic density (64) and apoptosis of neurons (3, 27, 75) have also been found.

The clinical features of the AIDS dementia-complex (73, 79) are predominantly those of a subcortical dementia with slowed responses, apathy, and poor judgement and concentration. This suggests that the subcortical rather than cortical pathology contributes more to the clinical syndrome. Motor impairment can range from mild slowing or clumsiness of rapid or complex movements to severe weakness. HIV-1 infections of the brain in infants can be associated with severely impaired brain development and delayed or curtailed mental or motor development. The brain weight is normally reduced. The histological features seen in affected infants resemble those seen in adults with the added common feature of mineralization in the walls of blood vessels or as extracellular deposits particularly in the basal ganglia and frontal lobe white matter. Some lesions may be necrotic (30).

HIV Associated Myelopathy

Difficulty walking due to an ataxic-spastic gait and incontinence in AIDS sufferers is usually due to a myelopathy. Like HIV-1 encephalitis this myelopathy tends to occur late in the course of AIDS and there is a weak association with HIV-1 encephalitis (71). The pathology consists of vacuolar change and myelin degeneration in the lateral and posterior column white matter, maximal in the thoracic region (74, 87, 91). The myelin damage is associated with focal macrophage infiltrates. The pathology is reminiscent of that of subacute combined degeneration of the cord due to vitamin B_{12} or folate deficiency. Occasionally multinucleated macrophages are also present (Fig 4), and HIV-1 can be demonstrated in such cells (17).

Peripheral Sensory Neuropathy

The commonest form of peripheral neuropathy in AIDS is a progressive distal sensory neuropathy, sometimes accompanied by pain, seen late in the course of the disease. The pathology consists of a 'dying-back' type of axonal degeneration, but in some cases there may also be evidence of segmental demyelination and inflammation. HIV-1 has occasionally been demonstrated in nerves in such cases (6).

Other Findings

Neurological symptoms can occur during the latent phase of HIV-1 infection, before the onset of AIDS. Peripheral demyelinating neuropathy, either acute or chronic, of the immune type that also occurs in uninfected people as acute or chronic inflammatory polyneuropathy, is more common in HIV-1 infected subjects. Mononeuritis multiplex and cranial neuropathy also occur in this phase and have been attributed to immune complex vasculitis (29). In the terminal stages a sensory ganglionitis and autonomic neuropathy are also seen occasionally. In the CNS in the pre-AIDS phase of HIV-1 infection the brain usually appears normal but it may show mild mononuclear cell infiltrates in meninges and perivascular spaces and increased microglial cell activation (3).

Myopathy or myositis may also occur in subjects with AIDS. In polymyositis HIV-1 has been demon-

698

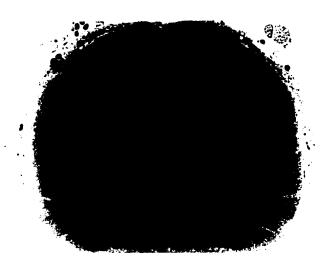


Figure 5. Myelin-stained section from the lumbar spinal cord from a chronic case of tropical spastic paraparesis (TSP). There is pallor of myelin staining, which is also accompanied by axonal loss, in the lateral white matter columns. Inflammation in this case was relatively sparse. Weil stain for myelin.

strated in lymphocytic infiltrates in muscle but not in muscle fibers. Zidovudine therapy for AIDS is associated with a myopathy which is reversible on discontinuing the drug and which appears to damage mitochondria with development of 'ragged red' muscle fibers (90).

Pathogenesis of HIV-1 associated neuropathology

Understanding is still lacking for many aspects of the pathology of HIV-1 encephalitis, myelopathy and neuropathy. It is clear from studies of cerebrospinal fluid (CSF) that HIV-1 reaches the meninges if not the brain in most subjects at an early stage of HIV-1 infection but the relevance of this finding for the development of brain or spinal cord damage is unclear. Symptoms of CNS disease are not correlated with the amount of HIV-1 RNA detectable in CSF but are correlated with the amount of HIV-1 RNA detectable in serum and with the severity of depression of the blood CD4 lymphocyte count (15). This suggests that severe immunosuppression and high HIV-1 levels systemically are necessary conditions for the development of HIV-1 associated CNS damage. However, since not all terminal AIDS sufferers develop CNS disease these conditions cannot on their own be sufficient to cause CNS disease, and it seems likely that an additional requirement is the evolution of the virus. Infectious virus differs in culture requirements and genotype. Genetic variants of the original infecting virus develop in any individual under the stimulus of immune attack during the latent phase of the infection. In particular, the development of variants that are macrophage tropic may be important to the development of CNS disease, since macrophage lineage cells are overwhelmingly the most common site of detection of HIV-1 in the brain. Furthermore,

the most likely mechanism for entry of HIV-1 into the brain is as an infection of monocytes traversing the blood brain barrier (37, 94). Since the quantity of HIV DNA in brain was not correlated with prospectively assessed measures of dementia in one recent study (46) it was suggested that the quality rather than the quantity of HIV-1 that is present may be more important for promoting the CNS damage that underlies dementia (46). Furthermore, certain amino-acid changes in the V3 Loop of the envelope protein appear more commonly in brain-derived DNA from neurologically affected than non-neurologically affected individuals (76). Thus, a macrophage tropic variant of the virus with distinctive envelope protein structure and severe immunosuppression late in AIDS are the likely requirements of HIV-1-induced CNS brain damage.

There remains considerable uncertainty over the means by which HIV-1 infected macrophages in the brain bring about the alterations in grey and white matter that have been found in HIV-encephalitis. Recent studies using the highly sensitive methods of polymerase chain reaction (PCR) and PCR - in situ hybridization have suggested that other cells in the CNS, particularly astrocytes, are subject to infection by HIV-1, albeit at a much lower level and with more highly restricted expression than macrophages (72, 81, 84, 95). The significance of such low-level infection still needs to be assessed. The much more abundant macrophage infection has been suggested to be capable of causing neuron and myelin damage via a number of different mechanisms. These have been largely investigated in vitro and the relative importance of each in vivo is not yet clear. Such mechanisms include synthesis and release of cytokines (97), release of soluble neurotoxic viral products such as the envelope protein gp120, and interference with production of growth factors (reviewed by 2, 59, 78).

The likelihood is that similar mechanisms to those operating in HIV-1 encephalitis are also responsible for HIV-associated myelopathy and neuropathy. The similarities between HIV-associated myelopathy and subacute combined degeneration of the cord have prompted speculation that metabolic factors such as local biotin deficiency, possibly exacerbated by its high local consumption by macrophages, may also play a part (92).

HIV-2 Infection

HIV-1 and HIV-2 viruses share approximately 40% of their genome in common. HIV-1 infections are far more widely distributed than HIV-2 which has mainly been described from West Africa. However, cases of HIV-2 infection have also been recently identified in India (8). In many regions of Africa facilities are not available for accurately distinguishing between HIV-1 and HIV-2 infections. The disease produced by HIV-2 closely resembles that produced by HIV-1, the main difference being that the natural

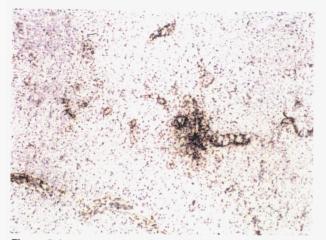


Figure 6. Low power view of the spinal cord inflammatory cell infiltrate in a case of TSP, shown here with a reaction for leucocyte common antigen (LCA). The inflammation is both perivascular and parenchymal in distribution. Counterstained with hematoxylin. (Section prepared by Dr CS Morris).

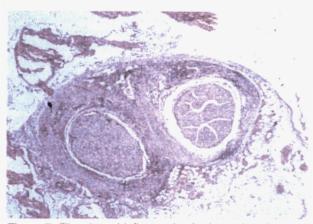


Figure 7. Fibrosis and inflammation in the perineurium surrounding these spinal nerve roots from a case of TSP. H & E stain.

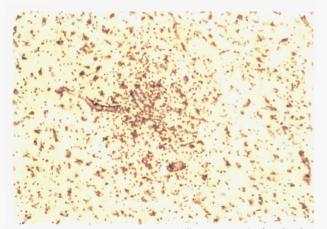


Figure 8. Exceptionally severe inflammatory lesion in the claustrum from the case of TSP illustrated in Fig 6. Reactive microglial cells and macrophages demonstrated by use of the lectin Ricinis Communis agglutinin 1. Counterstained with hematoxylin. (Section prepared by Dr CS Morris).

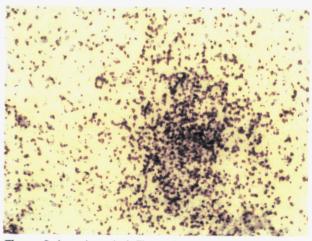


Figure 9. Lymphocytic infiltrate inthe same lesion as that shown in Fig 8, demonstrated with LCA antibody. Counterstained with hematoxylin. (Section prepared by Dr CS Morris).

history of HIV-2 infection seems to be rather more prolonged (102). An autopsy study from Africa suggested that multinucleated giant cell encephalitis is more common there in HIV-2 infection than in HIV-1 infection. A possible explanation for this difference is that the prolonged natural history of HIV-2 infections allows time for the encephalitis to develop (60).

HTLV-1 Infection

HTLV-1 is a human retrovirus found in many parts of the world, particularly in southern Japan, the Caribbean, Africa and South America. In some parts of these areas that harbor endemic virus the seroprevalence rate can be as high as 30% with the majority of those infected being totally asymptomatic (41). However, in about 3% of those infected an aggressive T cell leukemia develops and in about 0.25% of those infected a paralytic disease, tropical spastic paraparesis (TSP), also known as HTLV-1-associated myelopathy (HAM), develops. The incubation period between infection and development of TSP. when known, can vary from months to many years. Patients with TSP have high serum antibody titres to HTLV-1 - a feature that first drew attention to the possible link between HTLV-1 and TSP (28). Clinically TSP is characterized by slowly progressive spastic weakness of the lower limbs, sensory disturbances and difficulties with bladder control.

Pathologically, the most striking feature of TSP is a myelopathy which affects the white and grey matter of the cord, particularly at thoracic and lumbar levels (Fig 5). The pathology varies depending to some extent on the length of history with a well developed inflammatory response being seen in the spinal cord, leptomeninges and spinal roots in cases with shorter historles, and less inflammatory, more gliotic spinal cord lesions and fibrotic meningeal and nerve root lesions in those with a very long history. The inflammation affects both grey and white matter of the cord and consists of perivascular and more widespread lymphocyte and macrophage infiltration (Fig 6) accompanied by increased numbers of MHC Class II - expressing microglial cells and astrocytes. The majority of lymphocytes are CD8 T cells. The inflammatory lesions extend into the proximal nerve roots and leptomeninges around the cord (Fig 7). HTLV-1 p19 core protein has been demonstrated in spinal cord and proviral DNA has been detected in infiltrating lymphocytes (34, 66) and in occasional astrocytes (53) but not in neurons. White matter of the lateral and posterior columns of the spinal cord shows loss of myelin and to a lesser extent of axons (Fig 5). Similar inflammatory and demyelinating lesions may also be seen in the brain occasionally (Figs 8 and 9) and in the optic nerves. The meningeal inflammatory infiltrate extends to the meninges around the brain stem not uncommonly.

In attempting to explain why a tiny fraction of subjects infected with HTLV-1 should develop this progressive neurological disease studies have been performed on peripheral blood lymphocytes infected with the virus and on the host immune response to the virus. It seems to be generally accepted that TSP sufferers have a higher frequency of CD8 + HTLV-1specific cytotoxic lymphocyte responses in both blood and CSF than do healthy carriers of the virus (18, 40, 48). There is also a greater amount of HTLV-1 proviral DNA detectable in peripheral blood lymphocytes of TSP subjects than in healthy carriers (52). The evidence suggests that HTLV-1 is present in the spinal cord in TSP and that a brisk immune response to it takes place locally possibly damaging the spinal cord indirectly through release of cytokines and directly by cytotoxic killing of neuroglial cells harboring the virus. At present there is no clear evidence for a specifically neurotropic strain of virus (7).

HTLV-1 infection is associated with the development of polymyositis as well as TSP. In polymyositis, which clinically resembles the same disease in uninfected individuals and is thought to be immunologically mediated, HTLV-1;virus has been detected in the infiltrating lymphocytes in muscle but not in muscle fibers themselves (57).

Polio

Polio viruses are members of the picornavirus family, part of the enterovirus group of human pathogenic viruses, the group of viruses that most frequently infects the central nervous system in the form of aseptic meningitis. Polio virus, a singlestranded RNA virus, which occurs as three different serological subtypes, has long been recognized to be capable of causing paralytic disease in a small minority of subjects whom it infects. Initially described as a sporadic disease in young children this pattern of disease gave way to epidemics of infection particularly in late summer and autumn in Western countries in the early part of this century. Infection occurs by the fecal-oral route and in the 1-2% in which paralytic disease develops this takes the form of an initially non-specific febrile illness with sore throat and headache followed by back pain, stiffness, vomiting and muscle weakness. Limb involvement is often asymmetric and in a minority of subjects bulbar and respiratory muscles are also involved.

This form of disease carries a high mortality rate. Whilst paralytic polio has become exceedingly rare in Western countries as a result of effective immunization policies, it still remains relatively common in less developed countries of the tropics (38).

The usual site of infection with polio virus is the gastrointestinal tract and its associated lymphoid tissue. Confined to this region, the infection is commonly subclinical or trivial. How polio virus reaches the CNS to infect and kill motor neurons remains poorly understood. Early experimental studies in monkeys showed that a phase of viraemia was involved and that introduction of circulating antibody could prevent the infection reaching the CNS (11). A wealth of anecdotal and systematic evidence has recorded an excess susceptibility to paralysis due to polio in muscles that have recently been strenuously exercised, injured or injected (35, 106). Experiments on infection in monkeys some decades ago demonstrated the capacity of the virus to spread along axonal pathways (12). When these observations are coupled with the recent finding that polio virus receptors are present at neuromuscular junctions (56) a likely route for virus to take from blood to CNS would seem to be via muscle (particularly mildly inflamed muscle) and motor axons to spinal cord anterior horn cells. Such a route is supported by recent experiments in transgenic mice expressing human polio virus receptor which could be shown to have the development of paralysis after intramuscular inoculation with polio virus aborted by section of the sciatic nerve (82). The earlier explanation put forward to account for the enhanced susceptibility of injured muscles to paralysis was the unconvincing one that the blood supply to motor neurons innervating these muscles was increased (70). According to this hypothesis the selective vulnerability of motor neurons was assumed to be due to expression of polio virus receptors being confined to motor neurons. However, this has not been found to be the case. The polio virus receptor has now been identified, cloned and sequenced and found to be a member of the immunoglobulin superfamily (65). Its function other than as a polio virus receptor is not known. Northern blot analysis has indicated that its mRNA can be found in many human tissues in addition to brain (49) and the protein has likewise been detected in a wide variety of tissues (26). Immunocytochemical analysis suggests that the receptor is present in the brain not only in motor

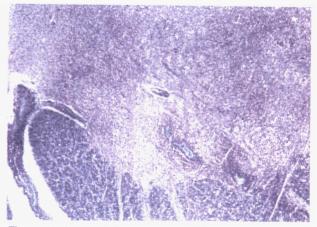


Figure 10. Intense inflammatory cell infiltrates in the anterior horn of the spinal cord in a case of acute polio. Luxol fast blue/cresyl violet-stain.

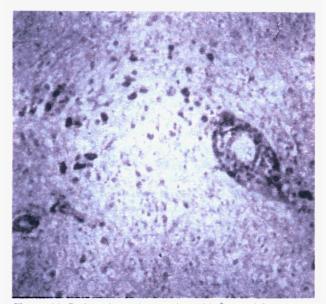


Figure 11. Perivacular and parenchymal IgG-containing plasma cells in the brain stem of a case of bulbar polio. Counterstained with hematoxylin.

cortex and cerebellum (known to be susceptible to infection by polio viruses) but also in olfactory bulb (thought to be refractory to infection) (25). Thus, the distribution of receptors, as recently analysed, does not readily explain the selective susceptibility of CNS cells to infection by polio virus, and this underlines the need to examine alternative explanations such as the route via muscle suggested above.

In the acute phase, polio is characterized pathologically by congestion and a severe inflammatory reaction concentrated particularly on the anterior horns of the spinal cord and, in cases with bulbar symptoms, in motor nuclei of the brain stem. Motor neurons show swelling or necrosis, accompanied by neuronophagia (Fig 10). A little later a severe loss of motor neurons at affected cord levels is detected, with continuing severe inflammation and microglial nodules. By 4-6 weeks following onset of symptoms many plasma cells can also be seen in the same areas (Fig 11) (19). The leptomeninges also participate in the inflammatory process. In the brain inflammatory cell infiltrates are not confined to motor cortex but may also be seen in the pontine and medullary reticular formation, midbrain, deep grey matter of the cerebral hemispheres, and cortex or dentate nuclei of the cerebellum.

Recent studies using *in situ* reverse transcriptase PCR of archival material have shown that polio virus RNA is present exclusively in motor neurons in the spinal cord anterior horns of subjects dying at the time of the acute phase of the disease (39), confirming earlier similar findings using immunocytochemistry and in situ hybridization in experimentally infected monkeys (16, 35).

In subjects dying after an attack of acute polio the inflammation has usually subsided and the sites of damage are marked by anterior horns largely devoid of motor neurons at levels corresponding to sites of origin of the innervation of severely paralyzed muscles and with increased numbers of glial cells in their place. In severe cases the whole anterior half of the cord tends to be severely atrophied and anterior roots wasted. Recent studies undertaken in the context of increasing recognition of a post polio syndromes in which further muscle fatigue and weakness may develop many years after an attack of acute polios have revealed persistent perivascular inflammation in some cases (47). Experiments to examine the possibility that polio virus may persist long after an attack of acute polio have produced suggestive evidence that it may, based on some positive PCR studies of CSF from post-polio syndrome patients (55, 58, 68). The significance of this recent finding for the development of the post polio syndrome remains to be assessed.

Rabies

Rabies is a unique CNS viral infection of human and other mammals due to infection with rabies virus, the most prominent of the lyssa viruses which are members of the rhabdo (rod-shaped) virus family. The virus has a bullet shape, being rounded at one end and flat at the other. It is an enveloped virus with a protruding surface transmembrane G glycoprotein, which is a target for immune responses, and a matrix protein lining the inner surface of the envelope. The matrix protein differs between different rabies virus strains whereas the G protein is shared in common. The envelope encloses the nucleocapsids or infectious components of the virus. They contain a single-stranded RNA genome coding for envelope and additional proteins including the RNA polymerase required for its own transcription.

Rabies virus is capable of infecting a wide variety of hosts, including man, though human infection can be regarded as incidental to infection of other

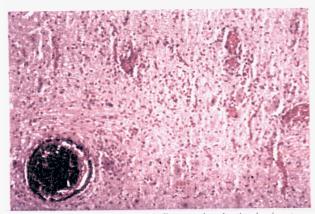


Figure 12. Congestion and inflammation in the brain stem from a case of rabies. H & E stain.

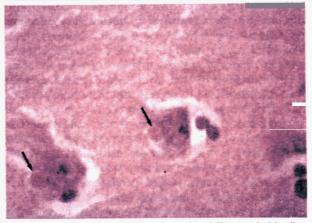


Figure 13. Negri bodies (arrows), eosinophilic rounded bodies in perivascular cytoplasm of two hippocampal pyramidal neurons from a case of rabies. H & E stain.

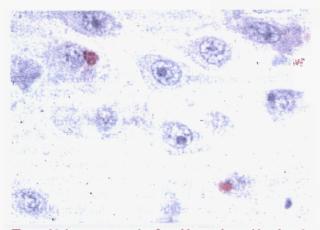


Figure 14. Immmunoreaction for rabies nucleocapid antigen in hippocampal pyramidal cells from a case of rabies. Counterstained with hematoxylin. (Photograph courtesy of Prof G Gosztonyi).

wild and domestic mammals. There are a few island communities that have eliminated the risk of rabies by quarantine procedures that exclude infected animals from their territory. Elsewhere, in different geo-

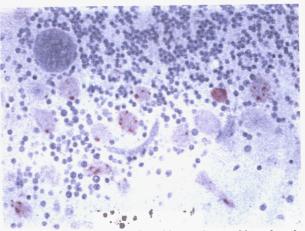


Figure 15. Immunoreaction for rabies nucleocapsids antigen in cerebellar Purkinje cells from a case of rabies. Counterstained with hematoxylin (photograph courtesy of Prof G Gosztonyi).

graphical regions of the world the threat of human infection is associated with infection in different groups of animals which act as reservoirs for the disease. For example, in India and South-East Asia the main threat is from infected dogs, in the USA from skunks, in Europe from foxes, in the central plateau of South Africa from the yellow mongoose, and in South America from bats. Infected animals start to secrete the virus in saliva a few days before developing symptoms and may continue to do so throughout the symptomatic period and beyond. Infection is usually spread by bite from an infected animal, whose tendency to bite is enhanced by aggression induced by the infection, but sometimes licking will suffice to transmit the virus or, if the source is bats, there may be no direct contact at all required for the disease to be spread. Many thousands of human infections occur world-wide each year (67).

Following infection due to a bite an unusually variable, often long, incubation period ensues before symptoms in the human develop. The modal length for such an incubation period is 40 days but it tends to be shorter in children or if the bite was on the head or neck. First symptoms are non-specific, consisting of fever, headache and nausea but there may also be paraesthesiae at the site of the bite. These are followed by meningism, convulsions, unwillingness to swallow due to a tendency to develop pharyngeal spasms, agitation and eventual coma, cardio-respiratory disturbance and death. The mortality rate in untreated infection with no prophylactic intervention is 100%.

Experimental infection following intramuscular injection of virus has clarified the events leading to CNS infection (69) although the reason for the prolonged incubation period is not well understood. The virus proliferates initially locally after being taken into muscle fibers probably by receptors provided by the acetylcholine receptors at motor end plates (54). From this site it spreads via axons to the

Disease	Glial nodules	Perivascular infiltrate	Foci of rarefaction necrosis without inflammation	Meningeal infiltrate	Sites of most severe damage	Vasculitis	Foci of tissue necrosis with inflammation	Neuronophagia	White matter lesions	References
EEE	Yes	Neurophils, mononuclear	Yes (may be late calcification)	Neurophils, mononuclear	Basal ganglia, substantia nigra, cerebral cortex, hippocampus	Yes, with thrombi	Many, especially in grey matter	Yes, neurophils, macrophages	Only near affected grey matter	5, 12, 22, 38
WEE	Yes	Mononuclear	Yes	Sparse, mononuclear	Striatum, thalamus, cerebral and cerebellar cortex, base of pons	Not usually	No	Yes, macrophages	No	2, 7, 12, 22, 25, 26, 37, 38
VEE	Yes	Sparse, mononuclear	No	Sparse, mononuclear	Putamen	No	No	Rare	No	12, 22, 28, 35
SLE	Yes	Mononuclear	Rare	Mononuclear	Thalamus, basal ganglia, brain stem, especially midbrain	No	No	Rare	Mild inflammation	4, 12, 18, 19, 22, 29, 31, 34, 36
JBE	Yes	Neurophils only in fulminating cases. Mononuclear	Yes (may be late calcification)	Mononuclear	Thalamus, substantia nigra, cerebral cortex, hippocampus, cerebellar cortex, spinal cord	No	Yes (may be late calcification(Yes	Only near affected grey matter	12-16, 21, 22, 32, 39, 40
MVE	Yes	Neurophils only in fulminating cases. Mononuclear	Yes (may be late calcification)	Mononuclear	Cerebral and cerebellar cortex, basal ganglia, brain stem	No	Yes	Rare	No	1, 8 , 22, 27, 30
TBE	Yes	Mononuclear	Yes	Mononuclear	Upper spinal cord, brain stem, cerebellum, thalamus, basal ganglia	Yes	Yes	Yes	No	3, 6, 9-12, 17, 20, 22, 25, 33
CalE	Yes	Mononuclear	No	Mononuclear	Cerebral cortex, basal ganglia, thalamus, mid- brain, pons	Yes	Few	Yes	No	16

Summary of pathological features in various forms of arbovirus encephalitis (reproduced from Ref. 13, with permission)

 Table 1. EEE: Eastern equine encephalitis; WEE: Western equine encephalitis; VEE: Venezuelan equine encephalitis; SLE: St Louis encephalitis; JBE: Japanese ß encephalitis; MVE: Murray Valley encephalitis; TBE: tick-borne encephalitis; CalE: Californian encephalitis.

1. Anderson SG (1954) J Hyg 52, 447. 2. Baker AB, Noran HH (1942) Archs Neurol Psychiat 47:565. 3. Bednar B (1961) In Encephalitides (eds van Bogaert et al), p17, Elsevier, Amsterdam. 4. Broun GO (1958) Neurology 8:883. 5. Farber S et al (1940) JAMA 114:1725. 6. Fingerland A, Vortel V (1961) In Encephalitides (eds van Bogaert et al) pp 23, 60 Elsevier, Amsterdam. 7. Finley K, Hollister AC (1951) Cal Med 74:225. 8. Garven AK et al (1952) Med J Austr 2:623. 9. Greenfield JG, Matthews WB (1954) J Neurol Neurosurg Psychiat 17:50. 10. Grinschgl G (1955) Bull Wid Hith Org 12:535. 11. Grinschgl G et al (1961) In Encephalitides (eds van Bogaert et al) p3, Elsevier, Amsterdam. 12. Haymaker W (1961) In Encephalitides (eds van Bogaert et al) p38, Elsevier, Amsterdam. 13. Haymaker W, Sabin AB (1947) Archs Neurol Psychiat 57:673. 14. Ishii T et al (1977) Acta Neuropath 38:181. 15. Iver CGS, Hadley GG (1957) Ind J Med Sci 11:227. 16. Kalfayan B (1983) Prog Clin Biol Res 123:179. 17. Kornyey S (1978) Acta Neuropath 43:179. 18. Leech RW, Harris JC (1977) J Neuropath Exp Neurol 36:611. 19. McCordock HA et al (1934) JAMA, 103:822. 20. McLean DM, Donahue WL (1959) Canad Med Ass J 80:708. 21. Matsuyama H (1955) Keio J Med 4:11. 22. Nieberg KC, Blumberg JM (1972) In Pathology of the Nervous System, Vol 3 (ed Minckler j) p2269, McGraw-Hill, NY. 23. Noran HH, Baker AB (1943) Archs Neurol Psychiat 49:398. 24. Noran HH, Baker AB (1945) J Neuropath Exp Neurol 4:269. 25. Osetowska E (1977) Tissue Neuropathology of Viral and Allergic Encephalitides, p93, Springfiield, Va. 26. Peers JH (1942) Archs Path 34:1050. 27. Perdau JR (1936) J Path Bacteriol 43:59. 28. Randall R, Mills JW (1944) Science 99:225. 29. Reves MG et al (1981) Archs Neurol 38:329. 30. Robertson EG (1952) Med J Austr 1:107. 31. Shinner JJ (1963) Archs Path 75:309. 32. Shiraki H et al (1963) Rev Neurol 180:633. 33. Silber LA, Soloview VD (1946) Am Rev Soviet Med (special suppl). 34. Suzuki M, Phillips CA (1966) Archs Path 81:47. 35. Tigertt WD, Downs WG (1962) Am J Trop Med 11:822. 36. Weil A (1934) Archs Neurol Psychiat 31:1139. 37. Weil A, Breslich PJ (1942) J Neuropath Exp Neurol 1:49. 38. Wolf A (1950) In The Pathogenesis and Pathology of Viral Diseases (ed. Kidd JG) p194, Columbia Press, NY. 39. Zimmerman HM (1946) Am J Path 22:965. 40. Zimmerman HM (1948) J Neuropath Exp Neurol 7:106.

spinal cord. Within the axons the virus is transported as a nucleocapsid, without its envelope (93). Having reached the neuronal perikaryon virus proliferation proceeds. Nucleocapsids are initially formed and later acquire their envelopes at sites on internal cell cisternal membrane or at the cell surface. From the spinal cord, infection spreads rapidly via axonal pathways to all parts of the CNS and, eventually, as enveloped particles at this stage, back into the peripheral nerves. One method of diagnosing the disease is to perform immunofluorescence on sections of skin in which dermal nerves can be seen to harbor the virus. This spread back into the periphery allows infection of salivary glands to occur, hence affording opportunity for spread of virus in saliva. Infection of limbic areas of the brain is said to generate altered behavior with aggression and a tendency A myocarditis also not uncommonly to bite. develops, and this may be fatal. At no stage does a viraemia develop and an immune response is not generated until the infection reaches the CNS.

The macroscopic appearances of the brain in rabies are not diagnostic consisting only of swelling and severe congestion. The distinctive changes are present on the microscopy. There is a variable amount of inflammation concentrated mainly in matter with perivascular lymphocyte, grey macrophage and plasma cell cuffing, neuronophagia and microglial nodules (Fig 12). The most severely affected regions are the brain stem and spinal cord. In some cases the inflammation is not conspicuous. The most useful diagnostic feature is the presence of rabies virus antigen which is distributed widely in the brain and can be suspected from routinely stained sections by the presence of Negri bodies, intracytoplasmic, rounded eosinophilic inclusions which are most prominent in large cells such as Purkinje cells, large cortical and hippocampal pyra-

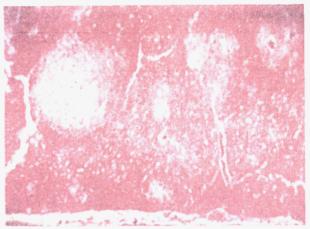


Figure 16. Foci of acellular necrosis in the cerebral cortex from a case of Japanese B encephalitis. H & E stain.

midal cells and spinal cord neurons (Fig 13). Ultrastructural studies have revealed Negri bodies to be ring-shaped aggregates of nucleocapsids (31), and they react immunocytochemically for rabies nucleocapsid antigen (Figs 14 and 15).

There are several puzzling aspects about the nature of rabies infection. One of these concerns the long incubation period. Another is the lack of an explanation for fatal brain stem involvement. This is not readily explicable solely on the basis of the distribution of the virus, for this is much more widespread and does not necessarily give rise to neuronal lysis at other sites. In tissue culture the virus is not in fact highly lytic. The disastrous consequences for the CNS of rabies infection may instead lie in the interference it produces to normal neurotransmitter regulation. Thus, binding capacity of acetylcholine receptors in rat brain was found to be reduced twofold by rables infection (50), and rables-infected neuroblastoma cells were shown to have a reduced affinity of opiate receptors and major disruption of aspects of cell metabolism controlled by opiate receptors (51). Another mechanism of neuronal damage that has been proposed is stimulation by rabies virus of production of nitric oxide by inducible nitric oxide synthase (23).

There is a further pattern of rabies pathology rather different from that described above in which the most severe pathology is found in the spinal cord. In this form of the disease, symptoms are predominantly those of an ascending paralysis resembling Guillain-Barré syndrome (14). About 10-20% of cases are of this type. The pathology in this form consists of inflammation, microglial nodules and neuronophagia in the anterior horns of the spinal cord and in the dorsal root ganglia with some extension of this pathology into the brain stem. The spinal leptomeninges are also inflamed. Negri bodies can usually be found in Purkinje cells and hippocampal neurons, as in the more common form of the disease.

Diagnosis during life depends on isolation of virus from saliva, demonstration of viral antigen in corneal cells or in nuchal dermal nerves, around hair follicles, or demonstration of rising serum and/or cerebrospinal fluid titres of anti-rabies antibody. Immunoperoxidase techniques can be used to detect virus in formalin fixed tissue (67). Post-exposure prophylactic immunization and administration of hyperimmune globulin can be provided before development of clinical symptoms and safe pre-exposure immunization is available to those at special risk. It needs to be emphasized that the possibility of rabies infection may be overlooked if a high level of clinical suspicion is not maintained. The diagnosis is not uncommonly made on post mortem examination, when again, the diagnosis may be missed if there is no history of exposure to rabies and a careful examination of the brain with this diagnosis in mind is not made.

Arthropod-borne (Arbo) Viruses

Arbovirus encephalitis (96) is an important cause of mortality and morbidity in many tropical regions of the world where the tendency of these viruses is to cause epidemics (13, 44, 104). The viruses responsible for these diseases are heterogeneous and classified among four main virus groups: the alpha virus subgroup of togaviruses (86) (responsible for Eastern, Western and Venezuelan equine encephalitis (VEE)), the flavivirus subgroup of togaviruses (responsible for St Louis, Japanese B (45) and Murray Valley encephalitides), bunyaviruses (responsible for California and La Crosse encephalitis) and a reovirus (responsible for Colorado tick fever encephalitis (43)). Many of these viruses circulate between mosquitoes and birds or mosquitoes and horses or small mammals and others between ticks and birds or small mammals. The viruses multiply both in the vertebrate and insect stages of their infectious cycles. Humans are incidentally infected, particularly under conditions in which insect breeding is prolific - conditions that were, for example, fulfilled before a recent epidemic of VEE after an absence of such epidemics for 20 years (99). Most cases of infection are subclinical but a minority of people (varying from around 1/100 to 1/1000) experience an acute, febrile illness of varying severity which may or may not be complicated by neurological involvement. Vaccines are available in some countries with populations at risk for arbovirus encephalitis but in other countries many thousands of cases continue to occur each year (104). Abortions and stillbirths are increased during and shortly after epidemics and virus may be recoverable from the brains of aborted fetuses (99).

The diseases are ushered in clinically with nonspecific flu-like symptoms lasting three or four days. This is quickly followed or simultaneously accompanied by features of neurological involvement, particularly seizures, confusion, drowsiness and

Main groups of rickettsial infections (adapted from ref 98)

Disense	Agent	Distribution	Transmission to man	
Rocky Mountain spotted fever	R. rickettsii	N Central and S America	Tick bite	
Boutonneuse fever	R. conorii	Mediterranean, Africa, Asia	Tick bite	
North Asia tick typhus	R. sibirica	Russia, China, Mongolia	Tick bite	
Oriental spotted fever	R. Japonica	Japan	Tick bite	
Queensland tick typhus	R. australis	E Australia	Tick bite	
Rickettsial pox	R. akari	USA, Ukraine, Croatia, possibly worldwide	Mite bite	
Typhus fevers				
Sylvate typhus	R. prowazeki	USA	Flea of flying squirrels to man	
Epidemic typhus	R. prowazeki	S America, Asia, Central America, Mexico	Louse feces scratched into skin	
Recrudescent typhus	R. prowazeki	Worldwide	None (re-activation of previous infection)	
Murine typhus	R. typhi	Worldwide predominantly tropical and subtropical	Flea feces scratched into skin, rubbed into conjunctiva or inhaled	
Scrub typhus	R. tsutsugamushi	Japan, S and E Asia, N Australia, islands of Western and Southwestern Pacific	Chigger bite	

Table 2.

meningism.

The pathological findings in fatal cases of arbovirus encephalitis have many features in common though there are also some important differences (Table 1). The viruses probably proliferate initially at local inoculation sites and are then spread in the blood to the brain and other organs. In the brain, virus infects neurons and, at least for some viruses, endothelial cells. Viral antigen can be detected in neurons early in the course of infection Perivascular infiltrates of predominantly (83). mononuclear inflammatory cells occur throughout the brain but with some predilection for grey matter in most forms of the disease, and for deep cerebral grey matter, cerebral cortex, hippocampus, substantia nigra and cerebellar cortex in particular (Table 1). The spinal cord may also be involved in some cases. Foci of acellular necrosis may be prominent (Fig 16) and a vasculitis with endothelial cell swelling or necrosis of small vessel walls and fibrin deposition in their lumen is seen. Neuronophagia, microglial nodules, astrocytes and patchy neuronal loss are common, but inclusion bodies are not seen. The leptomeninges participate in the inflammatory reaction. In experimental studies the immune response is important in controlling mortality for some viruses, eg, louping ill (107) but less important for others, eg Western and Eastern equine encephalitis (107), Sindbus virus (44) and Semliki Forest virus (100). Other factors such as age of host and strain of virus and its replication rate are also important determinants of disease severity (86).

Diagnosis depends on viral culture from serum or brain, or PCR analysis of such samples and on serological evidence of rising antibody titres during the course of the illness.

Rickettsial diseases

Rickettsial diseases are a heterogeneous group of infections caused by gram-negative bacterial microorganisms of different serological subtypes (Table 2). The organisms are transmitted to humans by insect vectors whose prevalence in particular locations and seasons is a strong determinant of human disease. Most of the different forms of rickettsial disease are found in tropical regions and are generally under-recognized in these countries (98). Apart from the presence of infected insect vectors the other most important factor determining risk of rickettsial disease is the social and living conditions of the local population. Wars, famines and refugee migrations are liable to lead to severe outbreaks of rickettsial disease. Some of these diseases are spread by infected saliva inoculated by a tick or mite bite and others by insect (lice, fleas) feces scratched into skin (Table 2).

These diseases are a cause of substantial illness particularly in developing countries. They present as acute, febrile illnesses, often accompanied by a rash, and, if undiagnosed and untreated, have significant mortality rates. Neurological involvement may be absent or amount only to mild photophobia, headache and drowsiness in mild or moderately severe cases but is more marked in severe cases in which there may be confusion, coma, epilepsy, ataxia or focal neurological signs. A pleocytosis in CSF is sometimes present. The diseases are divided into different groups on the basis of epidemiology, mode of transmission and clinical features (98). However, they all share similar neuropathological features, though these vary in severity from one group to another. The neuropathology was reviewed by Manuelidis and Krigman (62).

After entry of the organisms through the skin they are spread to all parts of the body in the blood. They attach to endothelial cells, mainly in small blood vessels, and are taken into these cells by endocytosis. They proliferate in the cytosol until the cell bursts, allowing spread to other cells, both endothelial and others, including white blood cells. Experimental studies suggest that cells are damaged by lipid peroxidation of their membranes by a combination of free radical production, protease activity and rickettsial phospholipase A_2 production (98). Host cytokines are produced and an immune response generated which leads to focal, mixed inflammatory infiltrates consisting predominantly of T lymphocytes but with some neutrophils and later macrophages admixed with them. These cells accu-

mulate around and within the walls of affected vessels which often include those of the leptomeninges and brain. Other organs are also affected including lungs, heart, skin, liver, pancreas and skeletal muscle. Some rickettsia generate a more severe reaction than others; thus, Rocky Mountain spotted fever is associated with more severe, necrotizing lesions than typhus fever. The organisms themselves can be seen in endothelial cells using an immunofluorescence or immunoperoxidase technique. Cytokine-activated nitric oxide production by host cells in experimentally infected mice appears to be crucial in overcoming infection. Depletion of β -interferon and tumor necrosis factor - α in these animals resulted in fatal overwhelming infection by a normally non-fatal dose of Rickettsia conorii (24). At the inflammatory foci that develop around infected endothelial cells in small blood vessels platelet plugs are liable to form but as these are usually non-occlusive they are not usually associated with microinfarcts. However, in severe cases microinfarcts may be seen at post mortem examination. More commonly there are only congestion and petechial hemorrhage formation to be seen in the leptomeninges and brain on macroscopic examination.

Laboratory diagnosis can be made in the acute phase of the diseases by skin biopsy of any rash and demonstration of rickettsiae by immunofluorescence or immunoperoxidase techniques. Immunostaining of white blood cells may also enable a diagnosis to be made. Retrospectively a diagnosis can be confirmed by demonstrating a four-fold rise in antibody titre to rickettsial antigens between acute and convalescent sera. Effective antibiotic treatment is available and should be given without awaiting laboratory confirmation of diagnosis.

References

- Abe H, Mehraein P, Weis S (1996) Degeneration of the cerebellar dentate nucleus and the inferior olivary nuclei in HIV-1-infected brain: a morphometric analysis. *Acta Neuropathol* 90: 150-155
- 2. Achim CL, Wiley CA (1996) Inflammation in AIDS and the role of the macrophage in brain pathology. *Curr Op Neurol Neurosurg* 9: 221-225
- Adle-Biassette H, Levy Y, Colombel M, Poron F, Natchev S, Keohane C and Gray F (1995) Neuronal apoptosis in HIV infection in adults. *Neuropathol Appl Neurobiol* 21: 218-27
- An SF, Ciardi A, Giometto B, Scaravilli T, Gray F and Scaravilli F (1996) Investigation on the expression of major histocompatibility complex class II and cytokines and detection of HIV-1 DNA within brains of asymptomatic and symptomatic HIV-1 positive patients. Acta Neuropathol 91: 494-503
- Bacellar H, Munoz A, Miller E, Cohen BA, Besley D, Selnes OA, Becker JT, McArthur JC (1994) Temporal trends in the incidence of HIV-1 related neurologic diseases; multicenter AIDS cohort study 1985 - 1992. *Neurology* 44: 1892-1900

- Bailey RO, Baltch AL, Ventakesh R, Singh JK, Bishop MB (1988) Sensory motor neuropathy associated with AIDS. *Neurology* 38: 886-891
- Bangham CR (1993) Retrovirus infections of the nervous system. Curr Op Neurol Neurosurg 6: 176-181
- Barnett SW, Murthy KK. Herndier BG, Levy JA (1994) An AIDS-like condition induced in baboons by HIV-2. Science 266: 642-646
- Bell JE, Donaldson YK, Lowrie S, McKenzie CA, Elton RA, Chiswick A, Brettle RP, Ironside JW, Simmonds P (1996) Influence of risk group and Zidovudine therapy on the development of HIV encephalitis and cognitive impairment in AIDS patients. *AIDS* 10: 493-499
- Bell JE, Busuttil A, Ironside JW, Rebus S, Donaldson YK, Simmonds P, Peutherer JF (1993) Human immunodeficiency virus and the brain: investigation of virus load and neuropathologic changes in pre-AIDS subjects. J Inf Dis 169: 818-824
- Bodian D (1955) Emerging concept of poliomyelitis infection. Science 122: 105-108
- 12. Bodian D, Howe HA (1941) Experimental studies on intraneural spread of poliomyelitis virus. *Bull Johns Hopk Hosp* 68: 248-267
- Booss J, Esiri MM (1986) Viral encephalitis: pathology, diagnosis and management. Blackwell Scientific Publications: Oxford
- Chopra JS, Banerjee AK, Murthy JMK, Pal SR (1980) Paralytic rabies: a clinicopathologic study. *Brain* 103: 789-802
- Conrad AJ, Schind P, Syndulko K, Singer EJ, Nagro RM, Russell JJ, Tourtellotte WW (1995) Quantifying HIV-1 RNA using the polymerase chain reaction in cerebrospinal fluid and serum of seropositive individuals with and without neurologic abnormalities. J Acqui Immune Defic Syndr Hum Retrovir 10: 425-435
- Couderc T, Christodowlou C, Kepecka H, Marsden S, Taffs LF, Crainic R, Horand F (1989) Molecular pathogenesis of neural lesions induced by poliovirus type 1. J Gen Virol 70: 2907-2918
- Eilbott DJ, Peress N, Burger H, laNeve D, Orenstein J, Gendelman HT et al (1989) Human immunodeficiency virus type 1 in spinal cords of acquired immunodeficiency syndrome patients with myelopathy: expression and replication in macrophages. *Proceed Natl Acad Sci USA* 86: 3337-41
- Elovaara I, Koenigs S, Brewah A, Jacobson S (1993) High HTLV-I specific precursor cytotoxic T lymphocyte frequences in patients with HTLV-1 associated neurological disease. J Exp Med 177: 1567-1573
- Esiri MM (1980) Poliomyelitis: immunoglobulin-containing cells in the central nervous system in acute and convalescent phases of the human disease. *Clin Exp Immunol* 40: 42-48
- Esiri MM, Kennedy PGE (1996) Virus diseases. In: Greenfield's Neuropathology 6th ed Graham DI, Lantos PL (eds) Vol II chap 1 Arnold: London
- Everall IP, Luthert PJ and Lantos PL (1991) Neuronal loss in the frontal cortex in HIV infection. Lancet 1: 1119-21
- Everail IP, Luthert P, Lantos P (1993a) A review of neuronal damage in human immunodeficiency virus infection: its assessment, possible mechanism and relationship to dementia. J Neuropathol Exp Neurol 52: 561-66
- Fekadu M, Greer PW, Chandler FW, Sanderlin DW (1988) Use of the avidin-biotin peroxidase system to detect rables antigen in formalin-fixed paraffin embedded tissues. J Virol Meth 19: 91-96

M.M. Esiri: Viruses and Rickettsiae

- Feng HM, Popov VL, Walker DH (1994) Depletion of gamma interferon and tumor necrosis factor alpha in mice with Rickettsia conorii - infected endothelium: impairment of rickettsicidal nitric oxide production resulting in fatal, overwhelming rickettsia disease. *Infect Imm* 62: 1952-1960
- Freistadt MS, Stoltz DA, Eberle KE (1995) Role of poliovirus receptor in the spread of infection. ANYAS 753 The post-polio syndrome. Dalakas MC, Bartfeld H, Kurland LT (eds) pp 27-47
- Freistadt MS, Kaplan G, Raccaniello R (1990) Heterogeneous expression of poliovirus receptor-related proteins in human cells and tissues. *Mol Cell Biol* 10: 5700-5706
- Gelbard HA, James HJ, Sharer LR, Perry SW, Saito Y, Kazee AM, Blumberg BM, Epstein LG (1995) Apoptotic neurons in brains from paediatric patients with HIV-1 encephalitis and progressive encephalopathy. *Neuropathol Appl Neurobiol* 21: 208-17
- Gessain A, Vernant JC, Maurs L, Barin F, Gout O, Calendar A, de The G (1985) Antibodies to human lymphotropic virus type-1 in patients with tropical spastic paraparesis. *Lancet* ii: 407-410
- 29. Therardi R, Lebargy F, Gaulard P, Mhiri C, Bernaudin JF, Gray F et al (1989) Necrotizing vasculitis and HIV replication in peripheral nerves. *N Engl J Med* 321: 685-6
- Giangaspero F, Scarabissi E, Baldacci MC, Betts CM (1989) Massive neuronal destruction in human immunodeficiency virus (HIV) encephalitis. A clinico-pathological study of pediatric cases. Acta Neuropathol 78: 662-5
- Gosztonyi G (1994) Reproduction of Lyssaviruses: structural composition of Lyssaviruses and functional aspects of pathogenesis. In: *Current topics in microbiology and immunology* Vol 187 Lyssaviruses, Rupprecht CE, Dietzschold B, Koprowski H (eds) pp 43-68, Springer: Berlin
- Gray F, Gelec L, Keohane C, de Trachis P, Clair B, Durigon M, Sobel A, Gherardi R (1994) Zidovudine therapy and HIV encephalitis: a 10-year neuropathological survey. *AIDS* 8: 489-93
- Hamilton R, Wiley CA (1997) Viral infection of the Nervous System, In: *Textbook of Neuropathology*. Davis RL, Robertson DM (eds) pp 927-1061 3rd Ed, Williams and Wilkins: Baltimore
- 34. Hara H, Morita M, Iwaki T, Hatae T, Itoyama Y, Kitamoto T, Akizuki S, Goto I, Watanabe T (1994) Detection of human T lymphotropic virus type 1 (HTLV-1) proviral DNA and analysis of T cell receptor Vβ CDR3 sequences in spinal cord lesions of HTLV-1-associated myelopathy / tropical spastic paraparesis. J Exp Med 180: 831-839
- 35. Hashimoto I, Hagiwara A, Komatsu T (1984) Ultrastructural studies on the pathogenesis of poliomyelitis in monkeys infected with poliovirus. Acta Neuropathol 64: 53-60
- Hill AB, Knowelden J (1950) Inoculation and poliomyelitis: a statistical investigation in England and Wales in 1949. Br Med J 2: 1-6
- Ho DD, Pomerantz RJ, Kaplan JC (1987) Pathogenesis of infection with human immunodeficiency virus. N Engl J Med 317: 278-86
- Horstmann DM (1982) Control of poliomyelitis: a continuing paradox. J Inf Dis 146: 540-51
- Isaacson SH, Sivalkumar K, Asher DM, Pomeroy KL, Ramos-Alvarez M, Gibbs CJ, Gajdusek DC, Dalakas MC (1995) Cellular localization of poliovirus RNA in the spinal cord during acute paralytic poliomyelitis. ANYAS Vol 753

The post polio syndrome, Dalakas MC, Bartfeld H, Kinland LT (eds) pp 194-200

- Jacobson S, Shuda H, McFarlin DE, Fanci AS, Koenig S (1990) Circulating CD8 + cytotoxic lymphocytes specific for HTLV-1 in patients with HTLV-1 associated neurological disease. *Nature* 348: 245-248
- Jacobson S (1995) Human T lymphotropic virus, type-1 myelopathy: an immunologically mediated chronic progressive disease of the central nervous system. *Curr Op Neurol Neurosurg* 8: 179-183
- Janssen RS, Cornblath DR, Epstein LG, Foa RP, McArthur JC, Price RW, Ashbury AK, Beckett A, Benson DF, Bridge TP, Leventhal CM, Spatz P, Saykin AJ, Sidtis JJ, Tross S (1991) Nomenclature and research case definition for neurological manifestations of human immunodeficiency virus type-1 (HIV-1) Infection. *Neurol* 41: 778-785
- 43. Johnson RT (1982) Viral infections of the nervous system. Raven Press: New York
- 44. Johnson RT (1982) Arboviruses in viral infections of the nervous system, pp 105-119, Raven Press: New York
- Johnson RT, Burke DS, Elwell M, Leake CJ, Nisalak A, Hoke CH, Lorsomrudee W (1985) Japanese encephalitis: immunocytochemical studies of viral antigen and inflammatory cells in fatal cases. *Ann Neurol* 18: 567-73
- Johnson RT, Glass JD, McArthur JC, Chesebro BW (1996) Quantitation of human immunodeficiency virus in brains of demented and non-demented patients with acquired immunodeficiency syndrome. *Ann Neurol* 39: 392-395
- Kaminski HJ, Tresser N, Hogan RE, Martin E (1995) Spinal cord histopathology in long-term survivors of poliomyelitis. *Muscle and Nerve* 18: 1208-209
- Koenig S, Woods R, Brewach AH, Newell A, Jones G, Boone E, Adelsberger JW, Baseter MW, Robinson SM Jacobson S (1993) Characterization of major histocompatibility complex - (MHC) class 1 restricted cytotoxic T cell (CT2) responses to tax in HTLV-1 infected patients with neurological disease. *J Immunol* 151: 3874-3883
- Koike S, Horie H, Ise I, Okitsu A, Yoshida M, Iizuka N, Takeuchi K, Takegami T, Nomoto A (1990) The poliovirus receptor protein is produced both as membrane-bound and secreted forms. *EMBO J* 9: 3217-3224
- Koprowski H, Zheng YM, Heber-Katz E, Fraser N, Rorke L, Fu ZF, Hanlon C, Dietzschold B (1993) In vivo expression of inducible nitric oxide synthase in experimentally induced neurological diseases. *Proc Nat Acad Sci (USA)* 90: 3024-7
- Koschel K, Munzel P 91984) Inhibitionof opiate receptormediated signal transmission by rabies virus in persistently infected NG-108-15 mouse neuroblastoma - rat glioma hybrid cells. Proc Nat Acad of Sci (USA) 81: 950-4
- Kubota R, Fujiyoshi T, Izumo S, Yashiki S, Marunyama I, Osame M, Sonada S (1993) Fluctuation of HTLV-1 proviral DNA in peripheral blood mononuclear cells of HTLV-1 associated myelopathy. J Neuroimmunol 43: 147-154
- Lehky TJ, Fox CH, Koenig S, Levin MC, Flerlage N, Izumo S, Sato E, Raine CS, Osame M, Jacobson S (1995) Detection of human T lymphotropic virus type-1 (HTLV-1) tax RNA in the central nervous system of HTLV-1 associated myelopathy/tropical spastic paraparesis by in situ hybridization. *Ann Neurol* 37: 246-254
- Lentz TL, Burrage TG, Smith AL, Crick J, Tignor GH (1982) Is the acetylcholine receptor a rabies virus receptor? *Science* 215: 182-184

- 55. Leon-Monzon M, Dalakas MC (1995) Detection of poliovirus antibodies and poliovirus genome in patients with the post-polio syndrome ANYAS Vol 753 *The Postpolio syndrome*. Dalakas MC, Bartfield H, Kurland LT (eds) pp 48-57
- Leon-Monzon M, Illa I, Dalakas C (1995) Expression of poliovirus receptor in human spinal cord and muscle. ANYAS Vol 753. *The post-polio syndrome*. Dalakas MC, Bartfield H, Kurland LT (eds) pp 48-57
- Leon-Monzon M, Illa I, Dalakas C (1994) Polymyositis in patients infected with human T cell lymphotropic virus type-1: the role of the virus in the cause of the disease. *Ann Neurol* 36: 643-649
- Leparc I, Kopecka H, Fuchs F, Janotova I, Aymarol M, Julian J (1995) Search for poliovirus in specimens from patients with the post-polio syndrome. ANYAS Vol 753 *The post-polio syndrome* Dalakas MC, Bartfield H, Kurland LT (eds) pp 233-236
- Lipton SA, Gendelman HE (1995) Dementia associated with the acquired immunodeficiency syndrome. N Engl J Med 332: 934-40
- Lucas SB, Hounnou A, Peacock C, Beaumel A, Djomand G, N'Gbidu J-M, Yeboue E Hondé M, Diomande M, Giordano L, Doorly R, Brattegaard K, Kestens L, Smithwick R, Kadio A, Ezani N, Yapi A, Decock KM (1993) Mortality and pathology of HIV infection in a West African city. *AIDS* 7: 1569-1579
- Maehlen J, Dunlop O, Liest CK, Doblong JH, Goplen AK, Torvik A (1995) Changing incidence of HIV-induced brain lesions in Oslo 1983 - 1994: effects of zidovudine treatment. *AIDS* 9: 1165-1169
- Manuelidis EE, Krigman MR (19??) Rickettsial encephalitides in pathology of the nervous system. Minckler J (ed) Vol 3 pp 2344-2362 McGraw-Hill: New York
- Martinez AJ, Mitrovics T, Stoltenburg-Didinger G, Inglesias-Rozas JR, Giraldo-Velasquez MA, Goxztonyi G, Schneider V, Cervos-Navarro J (1995) The neuropathology and epidemiology of AIDS: A Berlin experience. A review of 200 cases. *Pathol Res Pract* 191: 427-443
- Masliah E, Ge N, Morey M, Deteresa R, Tery RD, Wiley CA (1992) Cortical dendritic pathology in HIV encephalitis. Lab Invest 66: 286-291
- Mendelsohn CL, Wimmer E, Racaniello VR (1989) Cellular receptor for poliovirus: molecular cloning, nucleotide squence and expression of a new member of the immunoglobulin superfamily. *Cell* 56: 855-865
- Moore GRW, Traugott U, Scheinberg LC, Raine CS (1989) Tropical spastic paraparesis: a model of virusinduced, cytoxic T-cell mediated demyelination? Ann Neurol 26: 523-30
- Mrak RE, Young L (1994) Rabies encephalitis in humans: pathology, pathogenesis and pathophysiology. J Neuropathol Exp Neurol 53: 1-10
- Muir P, Nicholson F, Sharief MK, Thompson EJ, Cairns NJ, Lantos P, Spencer GT, Kaminski HJ, Banatvala JE (1995) Evidence for persistent enterovirus infection of the central nervous system in patients with previous paralytic poliomyelitis. ANYAS Vol 753 *The post-polio syndrome* Dalakas MC, Bartfield H, Kurland LT (eds) pp 219-232
- Murphy FA, Harrison AK, Winn WC, Bauer SP (1973) Comparative pathogenesis of rabies and rabies-like viruses. Infection of the central nervous system and centrifugal spread of virus to peripheral tissues. Lab Invest 29: 1-16
- 70. Nathanson N, Bodian D (1962) Experimental poliomyelitis following intramuscular virus injection 3. The effect of

passive antibody on paralysis and viraemia. Bull John Hopk Hosp 111: 198-220

- 71. Navia BA, Jordan BD, Price RW (1986) The AIDS dementia complex. Il Neuropathology. Ann Neurol 19: 525-35
- Nuovo G, Gallery F, MacConnel P, Braun A (1994) In situ detection of polymerase chain reaction - complified HIV-1 nucleic acids and tumor necrosis factor -α RNA in the central nervous system. Am J Pathol 144: 659-666
- Oldstone MBA, Vitkovic L (1995) HIV and dementia. Springer: Berlin
- Petito CK, Navia BA, Cho ES, Jordan BD, George DC, Price RW (1985) Vacuolar myelopathy pathologically resembling subacute combined degeneration in patients with acquired immunodeficiency syndrome. N Engl J Med 312: 874
- 75. Petito CK, Roberts B (1995) Evidence of apoptotic cell death in HIV encephalitis. Am J Pathol 146: 1121-30
- Power C, McArthur JC, Johnson RT, Griffin DE, Glass JD, Perryman S, Chesebro ? (1994) Demented and nondemented patients with AIDS differ in brain-derived human immunodeficiency virus type 1 envelope sequences. J Virol 68: 4643-4649
- 77. Price R, Sidtis J (1990) Early HIV infection and the AIDS dementia complex. *Neurol* 40: 323-326
- Price R, Sidtis JJ (1996) The cellular basis of central nervous system HIV-1 infection and the AIDS dementia complex. J Neuro AIDS 1: 1-173
- Price RW (1996) Neurological complications of HIV infection. Lancet 348: 445-452
- Quinn TC (1996) Global burden of the HIV pandemic. Lancet 348: 99-106
- Ranki A, Nyberg M, Ovod V, Haltia M, Elovaara I, Raminko R, Haapasalo H, Krohn K (1995) Abundant expression of HIV Nef and Rev proteins in brain astrocytes in vivo is associated with dementia. AIDS 9: 1001-1008
- Ren R, Racaniello VR (1992) Human poliovirus receptor gene expression and poliovirus tissue tropism in the transgenic mice. J Virol 66: 296-304
- Reyes MG, Gardner JJ, Poland JD, Month TP (1981) St Louis encephalitis: quantities histologic and immunofluorescent studies. Arch Neurol 38: 329-334
- Saito Y, Epstein LG, Michaels J, Mintz M, Louder M, Golding K, Cvetkovich TA, Blumberg BM (1994) Overexpression of nef as a marker for restricted HIV-1 infection. *Neurology* 44: 474-81
- 85. Scaravilli F (1993) *The neuropathology of HIV infection*. Springer: Berlin, London
- 86. Schlesinger RW (1980) The togaviruses; biology, structure and replication. Academic Press: New York
- Sharer LR, Epstein LG, Cho ES, Petito CK (1986) HTLV-III and vacuolar myelopathy in AIDS. N Engl J Med 315: 62-3
- Sharer LR, Mintz M (1953) Neuropathology of AIDS in children. In: Scaravilli F (ed) *The Neuropathology of HIV Infection.* Springer Verlag: London pp201-214
- Shaw GM, Harpers ME, Hahn BH, Epstein LG, Gajdusek DC, Price RW, Navia BA, Petito CK, O'Hara CJ, Groopman JE, Cho E-2, Oleske JM, Wong-Staal F, Gallo RC (1985) HTLV-1 infection in brains of children and adults with AIDS encephalopathy. *Science* 227: 177-182
- Simpson DM, Citak KA, Godfrey E, Godbold J, Wolfe D (1993) Myopathies associated with human immunodeficiency virus and zidorvudine: can their effect be distinguished? *Neurol* 43: 971-976

M.M. Esiri: Viruses and Rickettsiae

- Singh BM, Levine S, Yarrish RL, Hyland MJ, Jeanty D, Wormser GP (1986) Spinal cord syndromes in the acquired immune deficiency syndrome. Acta Neurol Scand 73: 590-8
- Smith I, Howells DW, Kendall B, Levinsky R, Hyland K (1987) Folate deficiency and demyelination in AIDS. *Lancet* 2: 215.
- 93. Sokol F, Schlumberger HD, Wiktor TJ, Koprowski H, Hummeler (1969) Biochemical and biophysical studies on the nucleocapsid and on the RNA of rabies virus. *Virology* 38:651-665
- 94. Stowring L, Haase AT, Petursson G, Georgsson G, Palsson P, Lutley R et al (1985) Detection of visna virus antigens and RNA in glial cells in foci of demyelination. *Virol* 141: 311-18
- Takahashi K, Wesselingh SL, Griffin DE, McArthur JC, Johnson RT, Glass JD (1996) Localisation of HIV-1 in human brain using polymerase chain reaction/in situ hybridization and immunocytochemistry. *Ann Neurol* 39: 705-711
- Thisyakorn U, Thisyakorn C (1994) Diseases caused by arboviruses-dengue haemorrhagic fever and Japanese B encephalitis. *Med J Aust* 160: 22-26
- Tyor WR, Wesselingh SL, Griffin JW, McArthur JC, Griffin DE (1995) Unifying hypothesis for the pathogenesis of HIV-associated dementia complex, vacuolar myelopathy and sensory neuropathy. J Ac Imm Defic. Syndr Hum Retrovir 9: 379-388
- Walker DH (1966) Rickettsial disease including ehrlichioses. In: Oxford Textbook of Medicine, Weatherall DJ, Ledingham JGG, Warrell DA (eds) pp 728-739 Oxford Univ Press Vol 1.
- Weaver SC, Salas R, Rico-Hesse R, Ludwig GV, Oberste MS, Boshell J, Tesh RB (1996) Re-emergence of epidemic Venezuelan equine encephalomyelitis in South America. Lancet 348: 436-440
- 100. Webb HE, Chew-Lim M, Jagelman S, Oaten SW, Pathak S, Suckling AJ, Mackenzie A (1979) Semliki Forest virus infections in mice as a model for studying acute and chronic central nervous system virus infections in man. In: *Clinical Neuroimmunology*, Rose FC (ed) pp 369-390. Blackwell: Oxford
- 101. Weis S, Haug H, Budka H (1993) Neuronal damage of the cerebral cortex in AIDS. Acta Neuropathol 85: 185-189
- 102. Whittle H, Morris M, Todd J, Corrah T, Sabally S, Bangah J, Ngom PT, Rolfe M, Wilkins A (1994) HIV-2-infected patients survive longer than HIV-1 patients. *AIDS* 8: 1617-1620
- 103. WHO (1995) Weekly epidemiological record 13th January.
- 104. WHO (1982) Report of a working group on Japanese encephalitis in *Centre for Disease Control Morbid Mortal* Wkly Rep 33: 119-125.
- 105. Wiley CA (1995) Quantitative neuropathologic assessment of HIV-1 encephalitis. *Curr Top Microbiol Immunol* 202: 55-61.
- 106. Wyatt HV (1982) Any questions? Tropical Doctor 12: 218
- 107. Zlotnik I, Smith CE, Grant DP, Peacock (1970) The effect of immunosuppression in viral encephalitis with special reference to cyclophosphamide. *Brit J Exp Path* 51: 434-439