

REVIEW

Biological drug and drug delivery-mediated immunotherapy



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Received 25 September 2020; received in revised form 3 November 2020; accepted 15 November 2020

Abbreviations: α 1D-AR, α 1D-adrenergic receptor; AAs, amino acids; ACT, adoptive T cell therapy; AHC, Chlamydia pneumonia; ALL, acute lymphoblastic leukemia; AP, ascorbyl palmitate; APCs, antigen-presenting cells; ApoA-I, apolipoprotein A-I; ApoB LPs, apolipoprotein-B-containing lipoproteins; AS, atherosclerosis; ASIT, antigen-specific immunotherapy; bDMARDs, biological DMARDs; BMPR-II, bone morphogenetic protein type II receptor; Bregs, regulatory B lymphocytes; CAR, chimeric antigen receptor; CCR9–CCL25, CC receptor 9–CC chemokine ligand 25; CD, Crohn's disease; CETP, cholesterol ester transfer protein; CpG ODNs, CpG oligodeoxynucleotides; CTLA-4, cytotoxic T-lymphocyte-associated protein-4; CX3CL1, CX3C-chemokine ligand 1; CXCL 16, CX3-chemokine ligand 16; CXCR 2, CXC-chemokine receptor 2; DAMPs, danger-associated molecular patterns; DCs, dendritic cells; DDS, drug delivery system; Dex, dexamethasone; DMARDs, disease-modifying antirheumatic drugs; DMPC, 1,2-dimyristoyl-sn-glycero-3-phosphatidylcholine; DSS, dextran sulfate sodium; ECs, endothelial cells; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; EPR, enhanced permeability and retention effect; ET-1, endothelin-1; ETAR, endothelin-1 receptor type A; FAO, fatty acid oxidation; GM-CSF, granulocyte–macrophage colony-stimulating factor; HA, hyaluronic acid; HDL, high density lipoprotein; HER2, human epidermal growth factor-2; hsCRP, high-sensitivity C-reactive protein; IBD, inflammatory bowel diseases; ICOS, inducible co-stimulator; ICP, immune checkpoint; IFN, interferon; IL, interleukin; IT-hydrogel, inflammation-targeting hydrogel; JAK, Janus kinase; LAG-3, lymphocyte-activation gene 3; LDL, low density lipoprotein; LPS, lipopolysaccharide; LTB4, leukotriene B4; mAbs, monoclonal antibodies; MCP-1, monocyte chemotactic protein-1; MCT, monocrotaline; MDSC, myeloid-derived suppressor cell; MHCs, major histocompatibility complexes; MHPC, 1-myristoyl-2-hydroxy-sn-glycero-phosphocholine; MIF, migration inhibitory factor; MM, multiple myeloma; MMP, matrix metalloproteinase; mPAP, mean pulmonary artery pressure; MOF, metal–organic framework; MPO, myeloperoxidase; MSCs, mesenchymal stem cells; nCmP, nanocomposite microparticle; NF- κ B, nuclear factor κ -B; NK, natural killer; NPs, nanoparticles; NSAIDs, nonsteroidal anti-inflammatory drugs; PAECs, pulmonary artery endothelial cells; PAH, pulmonary arterial hypertension; PASMcs, pulmonary arterial smooth muscle cells; PBMCs, peripheral blood mononuclear cells; PCSK9, proprotein convertase subtilisin kexin type 9; PD-1, programmed death protein-1; PD-L1, programmed cell death-ligand 1; PLGA, poly lactic-co-glycolic acid; RA, rheumatoid arthritis; rHDL, recombinant HDL; rhTNFRFc, recombinant human TNF- α receptor II-IgG Fc fusion protein; ROS, reactive oxygen species; scFv, single-chain variable fragment; SLE, systemic lupus erythematosus; SMCs, smooth muscle cells; Src, sarcoma gene; SHP-2, Src homology 2 domain-containing tyrosine phosphatase 2; TCR, T cell receptor; Teff, effector T cell; TGF- β , transforming growth factor β ; Th17, T helper 17; TILs, tumor-infiltrating lymphocytes; TIM-3, T-cell immunoglobulin mucin 3; TLR, Toll-like receptor; TNF, tumor necrosis factor; T_{ph}, T peripheral helper; TRAF6, tumor necrosis factor receptor-associated factor 6; Tregs, regulatory T cells; UC, ulcerative colitis; VEC, vascular endothelial cadherin; VEGF, vascular endothelial growth factor; VISTA, V-domain immunoglobulin-containing suppressor of T-cell activation; YCs, yeast-derived microcapsules.

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Peer review under responsibility of Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association.

<https://doi.org/10.1016/j.apsb.2020.12.018>

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KEY WORDS

Inflammatory diseases;
Cancer immunotherapy;
Atherosclerosis;
Pulmonary artery
hypertension;
Biologics;
Adoptive cell transfer;
Immune targets;
Drug delivery

Abstract The initiation and development of major inflammatory diseases, *i.e.*, cancer, vascular inflammation, and some autoimmune diseases are closely linked to the immune system. Biologics-based immunotherapy is exerting a critical role against these diseases, whereas the usage of the immunomodulators is always limited by various factors such as susceptibility to digestion by enzymes *in vivo*, poor penetration across biological barriers, and rapid clearance by the reticuloendothelial system. Drug delivery strategies are potent to promote their delivery. Herein, we reviewed the potential targets for immunotherapy against the major inflammatory diseases, discussed the biologics and drug delivery systems involved in the immunotherapy, particularly highlighted the approved therapy tactics, and finally offer perspectives in this field.

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1. Introduction

Inflammatory diseases including cardiovascular diseases, cancer, allergies, autoimmune, and neuropsychiatric diseases commonly feature dysregulation of immune response¹. For instance, atherosclerosis (AS) starting with dysfunctional alternation in the endothelium demonstrates increased recruitments of immune cells encompassing lymphocytes, antigen-presenting cells (APCs) and monocytes/macrophages². Immunotherapy refers to disease treatment through activating or inhibiting the immune system. Unlike traditional treatments, the immunotherapy exerts therapeutic efficacy through modifying the endogenous immune response, or reversing the immunosuppression conditions of diseases³, always characterized by changed infiltration of immune cells and modified expression of immune factors at the allergen⁴. Immunotherapy possesses significant advantages over traditional treatment regimes, such as higher efficacy against disease with fewer off-target effects and more durable response. Increasing evidence demonstrates immunotherapy is potent to treat malignant diseases, and cancer immunotherapy is becoming widely accepted in the clinic⁵. Furthermore, immunotherapy has promising potential to treat other inflammatory disorders, *e.g.*, AS⁶, rheumatoid arthritis (RA)⁷, intestinal inflammation⁸, and pulmonary arterial hypertension (PAH)⁹.

To target the pathogenesis of diseases, innate or adaptive immune system^{10,11}, numerous biological drugs were approved, including anti-tumor necrosis factor (anti-TNF) agents (etanercept, adalimumab, infliximab)¹², immune checkpoint (ICP) blockers (ipilimumab, tremelimumab, pembrolizumab)¹³, and interleukin (IL)-family agonists (nivolumab) or antagonists (tocilizumab)^{14,15}. The approved immunotherapy is present in Table 1^{16–24}. However, the use of immunotherapies is always limited by several factors. For instance, repeated administration of the immunomodulators at high dose is always required and, as a result, may induce a series of autoimmune-mediated side effects²⁵, such as flu-like reactions, and vascular leak syndrome²⁶, and show a significant individual variation. Furthermore, most immunomodulators belong to biological drugs separated from or manufactured by biological systems and always are characterized by increased size, low stability, poor penetration into the diseased site and limited ability to cross cell membrane. Drug delivery using carriers including liposomes, hydrogels, living cells, microspheres, inorganic materials, polymeric micelles, drug crystals, and protein vehicles is robust to improve the treatment efficacy of

biological drugs^{27–32}, owing to the advantages including extended blooded circulation, improved accumulation, promoted penetration in diseased tissues, increased uptake, and high drug-loading ability, large surface areas, and easy decoration of physicochemical properties^{1,27,33–36}. Especially, more than 65 nanoscale drug delivery systems (DDSs) were marked for commercial use. In this review, we summarize the immunotherapy implications in major inflammatory diseases, highlight the biopharmaceuticals and DDS utilized to improve the delivery of immunomodulators, and finally provide perspectives in this field.

2. Cancer immunotherapy

2.1. Potential targets for cancer immunotherapy

Immunotherapy of cancer is attracting huge attention for its remarkable success in the clinic. *Via* targeting the immune system and overcoming the acquired drug resistance, the immunologically “cold” tumors lacking immune infiltrate can be converted into “hot” tumors having dense T cell infiltrate through efficient approaches³⁷, exhibiting as mobilization of the immune cells and eliminating the cancer cells¹⁰. In general, the immune response could be prompted by modulating the production of immune factors and ICPs and motivating the immune cells (Fig. 1).

2.1.1. Cytokines and vaccines

Cytokines are potent to modulate the immune system. Three main types of cytokine are involved in cancer immunotherapy, including ILs such as IL-2, IL-12, IL-15, and IL-21, interferons (IFNs), and granulocyte–macrophage colony-stimulating factor (GM-CSF)³⁸. The recombinant cytokine IFN α is the first approved for clinical use in 1986³⁹, followed by recombinant IL-2⁴⁰. However, a high dose of cytokines was required for effective treatment efficacy, and frequently leads to a series of unwanted effects, *e.g.*, capillary-leak syndrome and cytokine-release syndrome⁴¹. The combined use of cytokines with checkpoint inhibitors or anticancer monoclonal antibodies (mAbs) might improve anti-tumor efficacy⁴².

Therapeutic cancer vaccines whose activities closely rely on cytotoxic T cells combat the disease *via* abolishing cancer cells or abnormal cells⁴³. The cancer vaccines are divided into four classes: peptide vaccines, cell-based vaccines, viral vector vaccines, and nucleic acid vaccines⁴⁴. APCs, especially dendritic cells (DCs), are essential to the vaccination because they are

Table 1 Approved immunotherapy for clinical use.

| Approved therapy | Active agent | Administration route | Manufacturer | Trade name | Therapy implication | Approved year | Ref. |
|-------------------------------------|----------------------------|----------------------|-------------------------------------|--------------|--|---------------|-------|
| mAbs | Abciximab | IV injection | Johnson & Johnson/Lilly | ReoPro® | Cardiovascular disease | 1994 | 16 |
| | Rituximab | IV injection | Genentech Inc. | Rituxan® | NHL, RA | 1997 | 16 |
| | Infliximab | IV injection | Johnson & Johnson | Remicade® | CRD, RA | 1998 | 16 |
| | Trastuzumab | IV injection | Genentech Inc. | Herceptin® | Breast cancer | 1998 | 16 |
| | Etanercept | SC injection | Amgen | Enbrel® | RA | 1998 | 16 |
| | Gemtuzumab ozogamicin | IV injection | Wyeth | Mylotarg® | Leukemia | 2000 | 16 |
| | Alemtuzumab | IV injection | Genzyme | Campath -1H® | Leukemia | 2001 | 16 |
| | Adalimumab | SC injection | CAT, Abbott | Humira® | RA, CRD | 2002 | 16 |
| | Abatacept | IV infusion | BMS | Orencia® | RA | 2005 | 16 |
| | Panitumumab | IV infusion | Amgen | Vectibix® | Colorectal cancer | 2006 | 16 |
| | Golimumab | SC injection | Centocor/Johnson & Johnson | Simponi® | RA | 2009 | 16 |
| | Certolizumab pegol | SC injection | UCB Inc. | Cimzia® | RA | 2009 | 16 |
| | Ofatumumab | IV injection | Novartis | Arzerra® | MCD, RA | 2009 | 16 |
| | Ipilimumab | IV injection | Bristol-Myers Squibb | Yervoy | Metastatic melanoma | 2011 | 17 |
| | Mogamulizumab | IV injection | Kyowa Hakko Kirin | POTELIGEO® | ATL | 2012 | 16 |
| | Pertuzumab | IV injection | Genentech | Perjeta® | Breast cancer | 2012 | 16 |
| | Ziv-aflibercept | IV injection | Sanofi/Regeneron | ZALTRAP® | MCRC | 2012 | 16 |
| | Trastuzumab emtansine | IV injection | Roche/Genentech | Kadcyla® | Breast cancer | 2013 | 16 |
| | Obinutuzumab | IV injection | Genentech | Gazyva® | CLL | 2013 | 16 |
| | Pembrolizumab | IV injection | Merck Sharp & Dohme Corp. | KEYTRUDA® | advanced melanoma, HNSCC | 2014, 2016 | 17,18 |
| | Nivolumab | IV injection | Bristol-Myers Squibb | Opdivo® | NSCLC, HNSCC, renal cell carcinoma | 2015, 2016 | 17,18 |
| Adoptive cell therapy | Atezolizumab | IV injection | Genentech Inc | Tecentriq | NSCLC, Urothelial Carcinoma | 2016 | 18 |
| | Avelumab | IV injection | Pfizer/Merck KGaA (EMD Serono)/Dyax | Bavencio™ | Merkel cell carcinoma | 2017 | 16 |
| | Durvalumab | IV injection | AstraZeneca UK Limited | Imfinzi | Urothelial carcinoma | 2017 | 19 |
| | CAR-T therapy | | Novartis | Kymriah® | B cell ALL | 2017 | 20 |
| | | | Kite Pharma/Gilead Sciences | Yescarta | Large B cell lymphoma | 2017 | 21 |
| | | | Kite Pharma | Yescarta® | Relapsed or refractory large B-cell lymphoma | 2017 | 22 |
| Cytokine-or chemokine-based therapy | Tofacitinib | Oral | Pfizer | Xeljanz | RA | 2012 | 23 |
| | Ruxolitinib Phosphate | Oral | Incyte corp | JAKAFI | RA | 2011 | 24 |
| | Daclizumab | SC injection | Biogen | Zinbryta | Multiple sclerosis | 2016 | 24 |
| | Sarilumab | SC injection | Sanofi Synthelabo | Kevzara | RA | 2017 | 21 |
| | Ribociclib | Oral | Novartis | Kisqali | Breast cancer | 2017 | 21 |
| | Brigatinib | Oral | Ariad Pharmaceuticals/Takeda | Alunbrig | ALK-positive NSCLC | 2017 | 21 |
| | Neratinib | Oral | Puma Biotechnology | Nerlynx | HER2-overexpressed breast cancer | 2017 | 21 |
| | Copanlisib dihydrochloride | IV injection | Bayer | Aliqopa | Follicular lymphoma | 2017 | 21 |
| | Baricitinib | Oral | Eli Lilly | OLUMIANT | RA | 2017 | 21 |
| | Sarilumab | SC injection | Sanofi Synthelabo | KEVZARA | RA | 2018 | 24 |
| | Tofacitinib citrate | Oral | Pfizer | XELJANZ XR | RA | 2019 | 24 |

ALL, acute lymphoblastic leukemia; ALK, anaplastic lymphoma kinase; ATL, adult T-cell leukemia/lymphoma; CLL, chronic lymphocytic leukemia; CRD, Crohn's disease; HNSCC, head and neck squamous cell carcinoma; MCD, multicentric Castleman's disease; MCRC, metastatic colorectal cancer; NSCLC, non-small cell lung cancer; NHL, non-hodgkin lymphoma; RA, rheumatoid arthritis; IV Injection, intravenous injection; SC Injection, subcutaneous injection; IV Infusion, intravenous infusion.

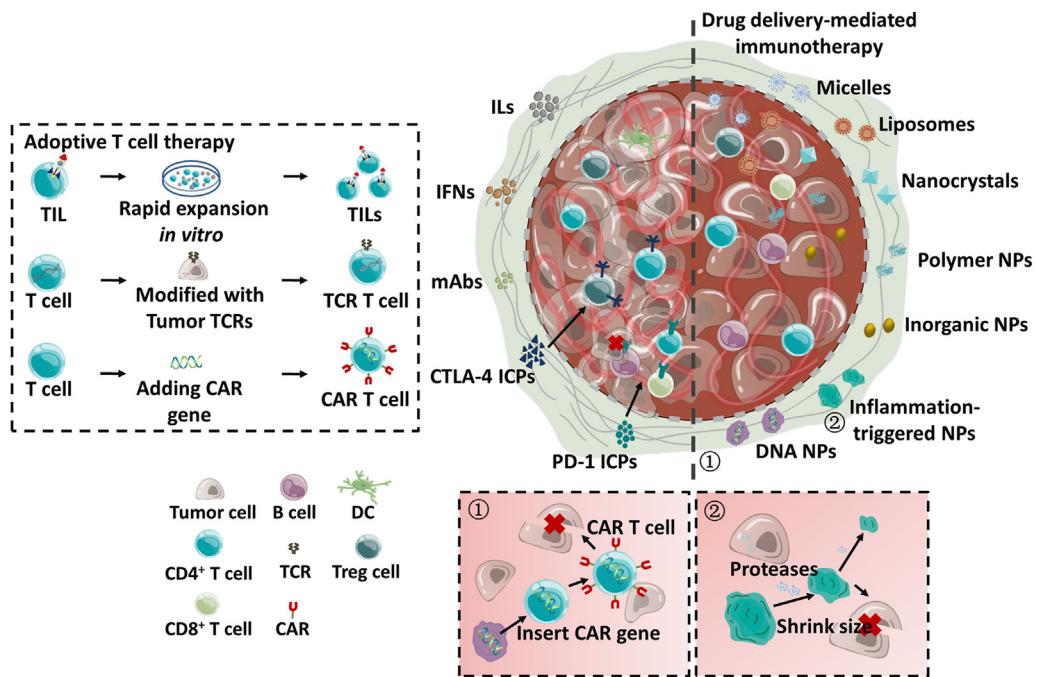


Figure 1 Immunotherapy for cancer and the used drug delivery systems (DDSs).

efficient to catch, refine, and present antigens to T cells⁴³. Most reported cancer vaccines belong to the DC-based vaccines. Effective vaccine-elicited CD8⁺ T cells should have properties as follows: (i) efficiently binding T-cell receptor, (ii) possessing robust T-cell affinity to the major histocompatibility complexes (MHCs) on cancer cells, (iii) producing a high level of granzymes and perforin (IV) potently recruiting T cells to site of tumor, and (V) modulating the release of costimulatory and inhibitory molecules⁴³. Three cancer vaccines, Gardasil, Cervarix, and Sipuleucel-T were commercially marketed. However, the mutation of antigens is always unique to individuals and, therefore, compromises the treatment efficacy of the commonly used vaccine. The personalized vaccine is a potential route to overcome the shortage.

2.1.2. *mAb and ICP suppressors*

By targeting surface antigens differentially expressed on cancer cells, such as CD20, CD33, CD52, human epidermal growth factor-2 (HER2), vascular endothelial growth factor (VEGF), epidermal growth factor receptor (EGFR), the antibody exerts cancer immunotherapy *via* means including the antibody-dependent cellular cytotoxicity and complement-mediated cytotoxicity⁴⁵. mAbs represent the most frequently employed cancer immunotherapy in the clinic, and over 30 products were approved.

ICPs are regulators often expressed on lymphocytes and classified into inhibitors and stimulators, such as cytotoxic T-lymphocyte-associated protein-4 (CTLA-4), programmed death protein-1 (PD-1), programmed cell death-ligand 1 (PD-L1), lymphocyte-activation gene-3 (LAG-3), OX40 (a potent costimulatory receptor), B7-H3 belonging to a member of B7 superfamily, 4-1BB categorized into a member of TNF receptor superfamily, V-domain immunoglobulin-containing suppressor of T-cell activation (VISTA), T-cell immunoglobulin mucin 3 (TIM-3),

and inducible co-stimulator (ICOS)⁴⁶. Several inhibitors of ICPs, *i.e.*, anti-PD-1, anti-PD-L1, and anti-CTLA-4, were approved for the clinical use^{47,48}. Nonetheless, their application may be limited for acquired-resistance to monotherapy⁴⁹.

2.1.2.1. CTLA-4. T cells are activated *via* binding their surface CD28 with B7-1 (CD80) or B7-2 (CD86) on the APCs⁵⁰. However, the CD28 homolog CTLA-4, possesses a significantly greater binding affinity toward B7⁵¹ and, as a result, leads to blockade of T cell upregulation and activation. Anti-CTLA-4 acts through blocking the connection between B7 and CTLA-4. Human CTLA-4 antibodies, ipilimumab, was approved to treat advanced metastatic cancer; and while another CLTLA-4 blockade, tremelimumab, is under clinical trial. The long-lasting anti-tumor response always occurs after dosing, yet accompanying unwanted effects, such as enterocolitis, inflammatory hepatitis, and dermatitis; however, it was argued these toxicities could be discounted by using corticosteroids and whereas did not reduce the anti-tumor effects⁵².

2.1.2.2. PD-1 and PD-L1. PD-1 is categorized into the CD28 superfamily as well, whereas PD-L1 and PD-L2 are classified as the B7 family. The expression of the PD-1 was found predominantly on three immune cells in the periphery, activated CD8⁺ and CD4⁺ T cells and B cells⁵³. The binding with the ligand, PD-L1 or PD-L2, allow PD-1 to recruit the sarcoma gene (Src) homology 2 domain-containing tyrosine phosphatase 2 (SHP-2) and inhibit the T-cell activities⁵⁴, *e.g.*, T-cell expansion and effector functions including release of IFN- γ and cytotoxicity⁵⁵. It should be noteworthy that PD-1 mainly reduce effector T-cell functions at the later-phase of immune reaction while CTLA-4 engages at the early stage⁵⁶.

By targeting the PD-1, PD-L1, or PD-1PD-L1 axis, numerous mAbs were fabricated, including nivolumab, pembrolizumab,

tisrelizumab, camrelizumab and sintilimab, durvalumab, avelumab, and atezolizumab⁵⁷. Nivolumab (Opdivo®) and pembrolizumab (Keytruda®) have been commercially marked.

Blockades of CTLA-4 or PD-1-based signaling are effective to combat cancer, however, monotherapy may induce adverse effects occasionally probably due to the individual variations^{25,58}. Combinatorial treatment, *e.g.*, anti-CTLA-4 plus anti-PD-1 and PD-1 plus PD-L1 antibodies, provides the potential to eliminate or alleviate the side effects.

2.1.3. Adoptive T cell therapy (ACT)

ACT refers to the transfer of isolated T cells from the patient that are genetically engineered *in vitro* to the same patient⁵⁹, including tumor-infiltrating lymphocytes (TILs), T cell receptor (TCR) T cells, and chimeric antigen receptor (CAR) T cells⁶⁰ (Fig. 1). The FDA has approved CAR therapy for adult patients with leukemia and lymphoma.

For TIL therapy, the TILs are extracted from the separated tumors, sorted with endogenous TCRs, purified, and ultimately undergo a rapid expansion protocol *in vitro* using with IL-2 and CD3 antibody⁶¹. For TCR therapy, TCR composed of an α- and a β-chains is anchored on T cells through noncovalently binding with CD3 complex. T cells become cytotoxic T cells when the anchored TCR is recognized and binds with the MHC on APCs or tumor cells⁶⁰. Rapoport et al.⁶² developed NY-ESO-1/LAGE-1 TCR-engineered T cells to treat multiple myeloma (MM). The engineered T cells were infused into twenty patients with MM at a cell number of 2.4×10^9 two days after dosing autologous stem cell⁶². The results indicated that the engineered T cells could proliferate, move to the marrow, and kill the cancer cells selectively, with clinical response of up to 80% and median survival of 19.1 months⁶².

To overcome the limitations of TCR therapy, *e.g.*, the requirement of MHC expression, MHC identity, and costimulation, CAR therapy was developed *via* adding CAR genes on T cells comprised of an extracellular single-chain variable fragment (scFv), a transmembrane spacer, and intracellular signaling/activation domain(s)⁶³.

ACT therapy has demonstrated its success in the treatment of cancers and several products were approved for clinical use. However, increasing limitations of the therapy are revealed as well, *i.e.*, on-target/off-tumor toxicity, cytokine-release syndrome, neurologic toxicities, off-targeting reactivity, complicated fabrication, extremely high cost, etc.^{64–66}.

2.2. Drug delivery-mediated cancer immunotherapy

To promote the immune response toward cancer, repeated administration of immunomodulators at a high dose is always required because most immunostimulants are unstable in physiological conditions, have the poor tumor-targeting ability, and can't translocate the plasma membrane, etc.²⁷. Such a dosing approach frequently induces side effects and compromises the patient's compliance. As a result, dosing the immunomodulators in a controllable and safe way is highly expected. Drug carriers, *e.g.*, liposomes⁶⁷ and dendrimers⁶⁸, are effective to improve the delivery due to their well-known advantages (Fig. 1). Numerous drug carriers were reported to improve the efficacy of immunotherapeutic agents such as T cell activators, ICP inhibitors, and cytokines⁶⁹, *via* increasing their circulation time⁷⁰ and target-

ability to immune cells⁷¹. Several reviews summarized the use of a drug delivery strategy to improve cancer immunotherapy^{69,72–75}. The most commonly used carriers include polymer nanoparticles (NPs), inorganic NPs, and lipid-based NPs^{72,76,77}. Recently, to lower the cost and facilitate the expansion of T lymphocytes for CAR therapy without complicated procedures, Smith et al.⁷⁸ developed plasmid DNA-loaded polymeric nanocarriers decorated with T-cell-targeting anti-CD3e f(ab')2 fragments to deliver leukemia-specific CAR genes into host T cells *in situ*. They found that the 155-nm NPs were able to rapidly and selectively program circulating T cells *in vivo* and demonstrated improved leukemia regression over the treatment with conventional CAR therapy⁷⁸. This work represents a new use of the DDS aiming to reduce the cost of ACT and avoid complications of clinical-scale manufacturing. Furthermore, changing the basic properties of NPs, *e.g.*, diameter, shape and surface charge, are potent to modulate the immunotherapy⁷³. For instance, smaller NPs (<50 nm) have enhanced ability to elicit the immune activities over the large NPs (>100 nm) because the smaller ones tend to traffic to lymph nodes *via* DCs, whereas the larger ones are difficult to move once accumulating at the diseased site⁷². The NPs with a diameter of over 500 nm can target macrophages and are internalized *via* phagocytosis⁷⁴.

Another significant advantage of using drug carriers is the efficiency to facilitate the combinational therapy. The durable immune response is only indicated in limited cancer types when an immunostimulant is used alone. Such that immunomodulator and other anti-tumor inhibitors are always combined for use in the clinic; however, their active targets are spatio-temporally discrepant and, as a result, often leads to sub-optimized treatment efficacy. Drug carriers have a remarkable potential to deliver the agents to their respective active sites precisely by co-delivery approach or physicochemical triggering means. For example, by using nanoclews based on long-chain single-strand DNA as a carrier that could be enzymatically degraded in inflammation conditions, CpG oligodeoxynucleotides (CpG ODNs) and anti-PD-1 antibody were released in a sustained pattern⁷⁹. The results demonstrated that the codelivery system synergistically induced long-lasting anti-tumor T lymphocyte responses in a melanoma model⁷⁹.

The nanocarriers demonstrated their promising potential to promote the treatment efficacy of cancer immunotherapy. Nonetheless, few of them are translated, mainly owing to the poor reproducibility and scalability, unpredictable toxicity *in vivo*, etc⁸⁰. Several techniques such as bubble blown assembly, capillary-force-assisted assembly, electric-field-assisted assembly, and Langmuir–Blodgett assembly were developed to scale up the nanomaterials⁸¹. Yet, it is difficult to acquire a commonly used approach to fabricate the devices since they always have their unique features. Furthermore, many studies for cancer immunotherapy was performed on the mouse models while there are huge discrepancy between the animal and human immune systems, the efficacy appeared on mouse may have poor correlation with human patients⁸².

3. Immunotherapy for autoimmune disease therapy

Autoimmune diseases encompassing RA, multiple sclerosis, inflammatory bowel diseases (IBD), mainly results from dysregulation of the T cell checkpoint pathways⁸³. Especially, the helper T

cells have profound effect on the progression of these diseases, since they often affect the function of other immune cells, *e.g.*, regulatory T cells (Tregs), monocytes and macrophages⁸⁴.

3.1. RA

RA is a chronic inflammation and frequently demonstrates damage of both articular cartilage and bone⁸⁵. The exact pathological mechanism of RA is unclear, but it is well accepted that RA is closely linked to the breakdown of immune tolerance⁸⁶. The immunotherapy by modulating the differentiation of lymphocytes and secretion of cytokines may combat RA (Fig. 2).

3.1.1. Implications for immunotherapy in RA

3.1.1.1. Regulation of lymphocytes. Four lymphocyte subpopulations, Tregs, T helper 17 (Th17) cells, and regulatory B lymphocytes (Bregs), affect the process of RA⁸⁷. Furthermore, Lamas et al.⁸⁸ discovered that the activation extends of peripheral blood mononuclear cells (PBMCs) and the disease activity allowed for immunomodulatory effect of bone marrow-derived mesenchymal stem cells (MSCs) on T-cell activation. Accordingly, the immunotherapy should concentrate on the modulation of these lymphocytes.

A deficit of Tregs was demonstrated to promote the RA development and increasing proliferation of Tregs *via* anti-TNF treatment benefits to the suppression of RA⁸⁹. Consequently, the activation of Tregs is the potential to ameliorate RA. These activators include IL-2⁹⁰, T cell superagonists (CD28SAs), and non-depleting anti-CD4 mAbs⁹¹. Second, the subsets of B cells, *i.e.*, Bregs⁹², memory B cells (CD24^{hi}CD27⁺ phenotype)⁹³, and B10⁺ cells⁹⁴, are potential targets to treat RA due to increased secretion

of IL-10⁹², improved proliferating of Tregs⁹⁴, or reduced expansion of Th1, Th17, TNF α ⁺ T cells⁹⁵. However, it was reported that, in patients with RA, the CD24^{hi}CD27⁺ and the CD24^{hi}CD38^{hi} B cells may not enhance the Treg's proliferation or decrease Th1 and TNF α ⁺ T cells although the abundance of the two sets is similar to that in the healthy⁹⁶. In this situation, the adoptive transfer of Bregs has the potential to alleviate the symptoms of the disease⁹⁷. Besides, the synovial macrophages advance the process of RA *via* the secreting IL-6 and TNF- α and the resultant damage of the joint¹.

Overall, *via* reeducating or depleting the autoreactive cells, the process of RA can be inhibited *via* inducing immune tolerance to self-antigens⁹⁸. The antigen-specific immunotherapy (ASIT) using peptides, antibodies, vaccines, etc. is extensively employed to target the autoreactive cells, *i.e.*, T and B cells and DCs⁹⁸. Recently, a pcDNA-CCOL2A1 DNA vaccine was developed to treat collagen-induced RA⁹⁹. The administration *via* intramuscular injection at 300 μ g/kg pcDNA-CCOL2A1 enabled decreased percentages of CD4⁺CD29⁺ and transferred Th1 to Th2 and Tc1 to Tc2, along with the reduced level of Th1 cytokines and downregulation of proinflammatory modulators IL-10 and transforming growth factor β (TGF- β) derived from Th2 and Th3, respectively⁹⁹.

3.1.1.2. Cytokines and chemokines. Cytokines and chemokines have a robust ability to regulate intercellular interactions, cell activation, localization, and phenotype in the lymphoid environment¹⁰⁰. The cytokines, in particular TNF and IL-6¹⁰¹, IL family, and GM-CSF¹⁰¹, promote the process of RA¹⁰².

TNF, a multifunctional cytokine, often exacerbates inflammation *via* increasing T-cell proliferation and differentiation

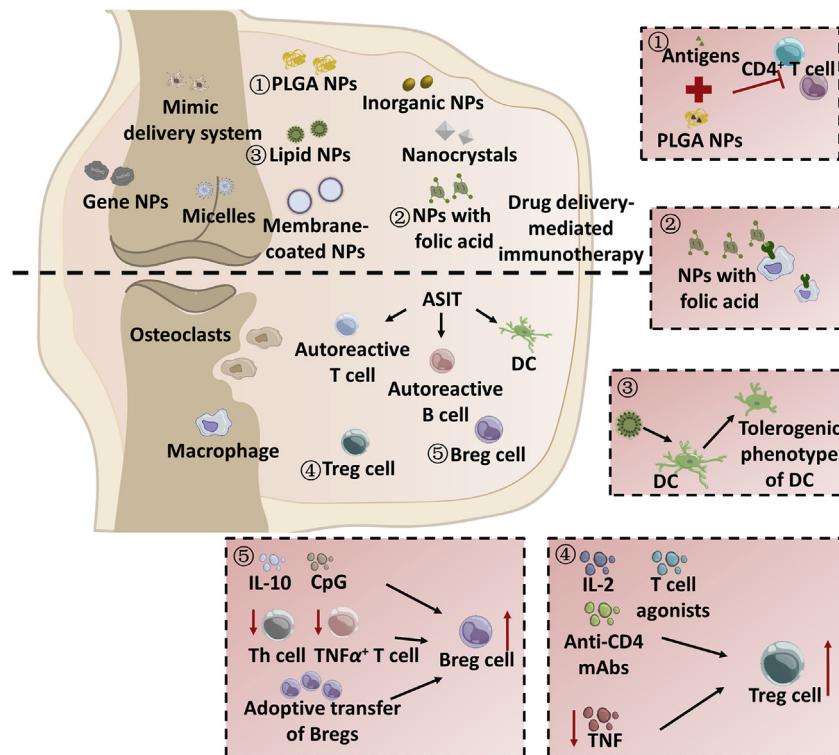


Figure 2 Immunotherapy for rheumatoid arthritis (RA) and the used DDSs.

at various stages¹⁰³, *i.e.*, single positive thymocytes and CD3/CD4/CD8 (triple-negative) T cells¹⁰⁴, and activating the immune system *via* the control of secondary lymphoid organs structures¹⁰³. Anti-TNF- α treatment is potent to treat RA *via* increasing Treg proportion and suppressing effector T cell (T_{eff})¹⁰⁵, affecting T peripheral helper (T_{ph}) cells that may prevent the differentiation of plasma-blasts¹⁰⁶, decreasing activated B cells, and expanding regulatory B10 cells¹⁰⁷.

IL-6, an activator of B and T cells, facilitates the differentiation of B cells into Ig-producing plasmablasts, directs the expansion of antigen inexperienced CD4 $^{+}$ T cells, as a consequence, promotes the transition of innate immunity to adaptive immunity¹⁰⁸. IL-6 inhibitors, such as IL-6 mAbs and miRNA targeting IL-6¹⁰⁸, toll-like receptor (TLR) 4 inhibitor¹⁰⁹, demonstrated promising inhibition of RA¹⁰⁹. In particular, the IL-6 mAbs exhibited outstanding efficacy against RA¹⁰⁸.

So far, some mAbs capable of neutralizing TNF- α have been approved for the clinical use, including etanercept, infliximab, certolizumab pegol, golimumab, adalimumab, and other blockers such as mAbs IL-6 (tocilizumab) and IL-1R (anakinra)¹¹⁰. The turbulence of immune cells, and the immune response is closely linked to RA progression. Consequently, the regulation of immune cells or immune response is promising to alleviate RA. However, individual treatment should not be ignored, since the gene expression and sensitivities are differentiated among person to person.

3.1.1.3. Janus kinase (JAK) inhibitors. The JAK pathway also links to the development of diverse immune-dependent disorders, *e.g.*, RA and IBD, by promoting the signal transduction of immunostimulators¹¹¹. The JAK, mainly composed of JAK1, JAK2, JAK3, and TYK2, acts through the receptors of type I and II. Type-I receptor generally associates with ILs, hormones and colony-stimulating factors, whereas Type-II receptor binds with IL-10-family cytokines including IL-10, IL-19, IL-20, IL-22 and IL-26¹¹² and IFNs. Two inhibitors of JAK, tofacitinib and baricitinib, were marked in 2018 and 2012, respectively, to treat RA¹¹³.

3.1.2. Drug delivery-mediated immunotherapy in RA

The used drugs against RA mainly consist of nonsteroidal anti-inflammatory drugs (NSAIDs), corticosteroids, disease-modifying antirheumatic drugs (DMARDs), and biological DMARDs (bDMARDs), and are always dosed *via* oral delivery or injection. Various DDSs were adopted to improve the delivery of modulators to the immune cells, *e.g.*, lipid-based NPs^{114–116}, polymeric NPs^{117,118}, hydrogels¹¹⁹, gold NPs¹²⁰, pH-sensitive NPs¹¹⁸, and biological membrane-coated NPs^{121–123}. Surface modification with ligands or peptide modification allows for improved targeting-ability to immune cells¹²⁴, and cytokines and chemokines pathways, such as nuclear factor κ -B (NF- κ B), ERK signal pathway, IL related pathway, etc⁹¹. Nonetheless, the most frequently reported ligand is folate receptor β that is overexpressed on the activated macrophages¹²⁵.

Liposomes or liposome-like NPs are the most widely used DDS for RA treatment due to their excellent encapsulation-ability and biocompatibility¹²⁶. Recently, lipidoid-polymer hybrid NPs were designed to deliver siRNA against IL-1 β to the activated macrophages to inhibit the pathogenesis of RA induced by collagen antibody¹¹⁵. Dosing *via* intravenous injection of the NPs enabled the efficient delivery of siRNA to macrophages inhibiting in the arthritic joints, downregulation of inflammation-induced

modulators in the joints, and a significant reduction in the cartilage destruction, swelling of ankle and bone damage¹¹⁵. Furthermore, such DDS facilitates codelivery for combined therapy. For instance, hybrid-NPs consisted of calcium phosphate/liposomes were developed to deliver methotrexate and NF- κ B-specific siRNA to the lipopolysaccharide (LPS)-activated macrophages at the diseased site¹²⁷. Egg phosphatidylcholine liposomes were used to co-load an antigen, OVA or methylated BSA, and a water-insoluble inhibitor of NF- κ B, curcumin or quercetin, and targeted APCs¹¹⁶, demonstrating as suppressing the response of the cells to proinflammatory pathway and promoting the proliferation of Ag-specific Foxp3 $^{+}$ Tregs¹¹⁶.

Biological membrane from living cells, *e.g.*, red blood cells, platelet, neutrophils, and macrophages, is always rich in various biomarkers and receptors; and as a result, its coating is able to alter the biological properties of DDSs and elevate their cell-targeting ability¹²¹. Motivated by the association of platelet with RA, platelet-like NPs were fabricated to deliver the anti-inflammatory tacrolimus to the joints of a collagen-induced arthritis¹²². Through GVPI recognition and P-selectin, these NPs allowed for efficient drug accumulation in the joint and inhibited RA's development¹²². Interestingly, drug-free neutrophil membrane-coated poly lactic-*co*-glycolic acid (PLGA) based NPs were developed recently¹²³. *Via* neutralizing the inflammation-induced TNF- α and IL-1 β , these neutrophil-NPs exhibited synovial inflammation, robust chondroprotection against joint damage, and enhanced penetration into the cartilage matrix¹²³. Furthermore, their anti-RA effectiveness in both human-transgenic arthritic model and collagen-induced model is comparable to that from the treatment with anti-TNF- α or anti-IL-1 β ¹²³. These biomimetic-targeting DDSs, due to their natural targeting-ability to the inflamed sites, represent a promising approach against RA and may have the potential of clinical translation because of the simple preparation process. Nevertheless, their translation is still limited by the scalability of DDS.

3.2. Intestinal inflammation

3.2.1. Potential targets for IBD immunotherapy

IBD is always characterized by long-lasting inflammation and divided into ulcerative colitis (UC) and Crohn's disease (CD). The IBD pathogenesis has not been illustrated fully, however, is often characterized by an imbalance between the mucosal immune system and the commensal ecosystem¹²⁸. The regulatory immune cells including intestinal intraepithelial lymphocytes, T and B cells, macrophages, DCs and innate lymphoid cells could affect the progression of IBD¹²⁹. DCs contribute the maintaining of immune environment homeostasis in the intestine *via* connecting humoral and cellular immune response. Especially, Tregs play a critical role in limiting the populations of Teffs and innate inflammatory signaling¹³⁰. Antigen-specific T-helper cells and natural killer (NK) cells contribute to inflammation in IBD as well and their influx can be used as a potential treatment target¹³¹. In addition, agitations in intestinal epithelial cells, in particular Paneth cells, may initiate intestinal inflammation¹³¹. The therapy strategies toward IBD are classified into anti-inflammatory treatment with mesalazine and glucocorticoids, antibiotics therapy using ciprofloxacin and metronidazole, gene therapy, and immunotherapy with immunosuppressants and anti-TNF agents⁸. The immunotherapy is acquired through interfering with IL-12/23 axis, JAK and TGF- β /Smad 7 pathways, and modulating IL-6, IL-13, chemokines and chemokine receptors CC receptor 9–CC

chemokine ligand 25 (CCR9–CCL25)¹³², and cell adhesion and leukocyte recruitment¹³³. The IBD immunotherapy can also be achieved by using adoptive cell transfer, such as MSCs and engineered Tregs⁸. Previous reviews summarized the use of biological drugs for IBD immunotherapy^{133,134}. Currently, about seven mAbs were approved for IBD immunotherapy, including ustekinumab, natalizumab, infliximab, vedolizumab, golimumab, certolizumab pegol and adalimumab. The implication of immunotherapy for IBD is illustrated in Fig. 3.

3.2.2. Drug delivery-mediated immunotherapy in IBD

The mAbs are effective to treat IBD; however, the response rate to initial treatment is only 50% and their use is always limited by systematic side-effects including immunogenicity, the induction of anti-drug antibodies, serum sickness, etc., induced by administration *via* intravenous injection¹³⁵. DDS-mediated therapy may elevate treatment outcomes and reduce systemic toxicity. A previous review summarized various approaches and devices for targeting treatment of IBD using DDS, encompassing meanings of ligand-receptor-, charge-, degradation-, size- and microbiome-mediation¹³⁶. Another review discussed intestine targeting strategies, *e.g.*, conventional DDS-mediated treatment, disease-mediated delivery of active agents by synthetic and biological DDS¹³⁷. Given that oral administration is a well-accepted delivery route for both patients and physicians¹³⁸, herein we mainly discuss targeted oral delivery of immunomodulators to the inflamed sites in the large intestine. The site-specific DDSs are often designed according to (i) the physiological changes in the gastrointestinal tract such as pH, microflora, transit time, pressure, and osmotic potential^{139,140} or (ii) disease-induced alterations, *i.e.*, increased permeability, changes in tight junctions and mucus composition and amount, reduced antimicrobial secretions and numbers of secretory cells and loss of the area of ulcerated epithelium^{141–145}. The recently reported DDSs include hydrogel platform^{146–149}, redox- and pH-sensitive NPs^{150–155}, hyaluronic-based NPs^{148,156,157}, macrophage-targeted DDSs^{1,156,158–160}, polyphenol-based delivery^{161,162}, etc.

Recently, an oral inflammation-targeting hydrogel (IT-hydrogel) assembled from ascorbyl palmitate (AP) was developed to treat IBD¹⁴⁹. IT-hydrogel microfibers encapsulating corticosteroid dexamethasone (Dex) could adhere to the inflamed mucosa from animal and human colon and exhibited increased drug release at the inflammation site due to the degradation by the enzymes secreted from active-macrophages and other immune cells. In clitic ulcerative mice administrated *via* a single enema with free Dex as control, dosing with the drug-loaded IT-hydrogel enabled significant reduction of colon weight, myeloperoxidase (MPO) activity, and expression of TNF in the distal colon, and lowered the systemic drug exposure¹⁴⁹. Another reactive oxygen species (ROS) responsive assembles prepared from HA-bilirubin conjugate were fabricated to combat dextran sulfate sodium (DSS)-induced acute colitis¹⁵⁶. *In vitro*, the assembles dissociated rapidly after exposure to ROS, were well taken up by macrophages and granulocytes due to the hyaluronic acid (HA)-CD44 affinity, polarized pro-inflammatory M1 macrophages into the M2 phenotype. *In vivo*, in contrast with treatment with clinically used drugs, the treatment had remarkably boosted efficiency to combat DSS-induced acute colitis *via* decreasing the impairment of colon and MPO activity, recovering the body weight, and keeping the length of colon¹⁵⁶. In addition, the treatment markedly reduced the infiltration of pro-inflammatory phenotypes, CD11b⁺Ly6C⁺Ly6G⁺ neutrophils and CD11b⁺Ly6C⁺Ly6G[−] monocytes, in the layer of lamina propria in DSS-induced model, and increased the accumulation of anti-inflammatory phenotypes including CD3⁺CD4⁺Foxp3⁺ Tregs, MHCII⁺CD11c⁺CD11b[−] DCs and CD11b⁺Ly6C[−]Ly6G-MHCII⁺ tissue-resident macrophages¹⁵⁶. Overall, increasing oral DDSs against IBD is emerging, such as polymer-drug prodrug formulations^{152,163}, microspheric vehicles to suppress TNF- α ^{154,164}, thermoreversible mucoadhesive polymer-drug dispersion with prolonged retention at the inflamed site¹⁶⁵, and biomimetic NPs, *i.e.*, cell membrane-coated NPs and liposomes engineered with cell membrane proteins¹⁶⁶. These oral inflammation-targeting DDSs represent a promising strategy to treat IBD, owing to their scalability, biocompatibility, and potent therapeutic efficacy.

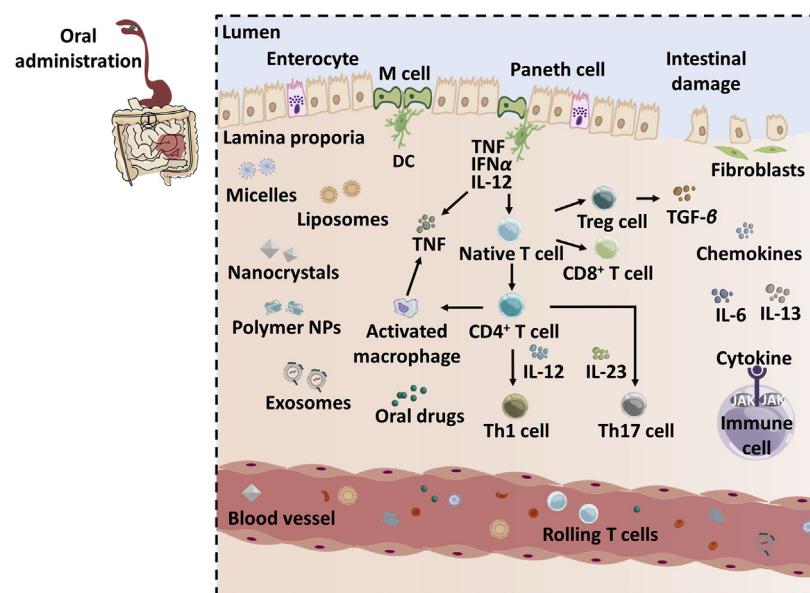


Figure 3 Immunotherapy for inflammatory bowel diseases (IBD) and the used DDSs.

Numerous DDSs were designed to treat IBD *via* targeting macrophages^{167,168}, whereas exosomes isolated from the TGF- β 1 gene-modified DCs was demonstrated to inhibit the progression of IBD *via* eliciting immunosuppression¹⁶⁹. The 50–100-nm exosomes can efficiently block the advance of DSS-triggered IBD *via* inducing Tregs through the TGF- β 1 pathway and reducing the Th17 in the inflammatory site, mesentery lymph nodes¹⁶⁹.

4. Immunotherapy for vascular disease

4.1. AS

In AS, the immune cells encompassing monocytes, T cells, DCs, neutrophils, NK cells and innate lymphoid cells are recruited to these sites^{170–173}, due to the elicited local inflammation by apolipoprotein-B-containing lipoproteins (ApoB LPs) that deposit in the artery wall and are liable to modification by oxidation, enzymes and aggregation¹⁷⁴. The recruited monocytes constantly differentiate into macrophages, a major cell population in the atherosclerotic plaques, and finally become cholesterol-loaded macrophage foam cells, facilitating the plaque formation^{175,176}. Whereas the factors, *e.g.*, pro-inflammatory modulators, cholesterol crystals, oxidative stress, oxidized lipids, and danger-associated molecular patterns (DAMPs) predominantly stemmed from the macrophage foam cells, construct a complicated micro-environment that maintains the local inflammation¹⁷⁷. A previous review highlighted the role of macrophages in AS development¹. Here, we mainly focus on other immune cells involved in AS and related treatment approaches.

4.1.1. Implications for AS immunotherapy

According to the pathological mechanism of AS, the treatment with anti-hypertensive or cholesterol-lowering drugs is far from to cure AS. Several experimental results demonstrated the essential roles of immune activation in the AS development^{178,179}. Always, the innate response to AS mediated by stimulating macrophages and endothelial cells (ECs) in the walls of the coronary arteries allows for adaptive immune reactions to the antigens presented to Teff by APCs, *i.e.*, DCs¹⁸⁰. As a result, targeting inflammation to modulate immune responses against plaque antigens may treat AS fundamentally. The potential targets and used DDSs are displayed in Fig. 4.

4.1.1.1. Cytokine-based therapy. Two types of cytokines are involved in AS progression, pro-atherogenic and anti-atherogenic cytokines¹⁸¹. Pro-atherogenic cytokines always promote the development of AS, including various ILs such as IL-4, IL-6, IL-8, IL-12, IL-15, IL-18, IL-20, IL-21, IL-23 and IL-32, GM-CSF, TNF- α , monocyte chemotactic protein-1 (MCP-1), IFN- α , β , and γ , etc. Whereas the anti-atherogenic ones can inhibit AS development, such as IL-5, IL-10, IL-13, IL-19, IL-27, IL-33, IL-35, IL-37, TGF- β , etc¹⁸¹. In general, cytokine-based treatment drugs are mainly categorized into broad-based immunomodulatory agents, blockade of pro-inflammatory cytokines and activators to induce anti-inflammatory cytokines¹⁸². Clinical trials uncovered the administration of Canakinumab, a mAb targeting IL-1 β , at 150-mg dose every 3 months reduced the inflammation and rate of cardiovascular events, though, did not lower the lipid-level¹⁸³. Another clinical test demonstrated that dosing Canakinumab with the same regimen allowed for decreased levels of IL-6 and

inflammatory biomarker high-sensitivity C-reactive protein (hsCRP), an indicator that the mAb works *via* inhibiting the IL-1 β –IL-6 signaling of innate immunity¹⁸⁴. Accordingly, pro-inflammatory cytokines can be effective targets for AS therapy and IL-1 β –IL-6–CRP signaling axis is a credible AS-associated inflammatory pathway¹⁸⁵.

4.1.1.2. ICPs. Due to a surplus of ICPs, *e.g.*, CD27, CD28, CTLA-4, CD40, CD40L, CD70, CD80/86, Ox40, Ox40 L, PD-1, PD-L1/2, the costimulatory molecules derived from T cells, CD30 and CD137L, can be induced and facilitate atherogenesis¹⁸⁶. For instance, blocking the CD80/86–CD28 axis alleviates the symptoms of AS that have occurred or are about to occur in both mice and humans^{187–189}. In addition, the dyad CD40L–CD40 is closely associated with plaque's vulnerability and formation^{190–193}. Treatment with anti-CD40L or CD40 allowed for plaque suppression¹⁹¹.

4.1.1.3. Chemokines. Over 20 chemokines produced mainly from ECs, smooth muscle cells (SMCs), leukocytes¹⁹⁴ and their receptors are involved in AS progression¹⁹⁵. The chemokines, CCL5, CCL2, CXC-chemokine receptor 2 (CXCR 2) and CXCR3 and their ligands, CXXXC-chemokine ligand 1 (CX3CL1) and CXC-chemokine ligand 16 (CXCL16), CXCL12/CXCR4 axis, and macrophage migration inhibitory factor (MIF), are linked to the plaque development¹⁹⁶. The main AS-treatment strategies based on the chemokines are divided into small molecule chemokine receptor antagonists, modified chemokine, chemokine-neutralizing protein and chemokine heteromer formation-antagonists¹⁹⁶. For example, the treatment with CCR5 antagonist enabled size reduction of plaque in *ApoE*^{-/-} mice¹⁹⁷. Furthermore, inhibition of CXCL12 is promising to prevent and alleviate AS-associated diseases. CXCL12 inhibitors, AMD3100, AMD3465, and POL551, showed inhibition of CXCL12-damaged vascular wall^{198,199}.

4.1.1.4. Metabolic regulation of immune cells. Besides the specific target substances, the altered metabolism of cells in AS may be used as therapeutic potential. The changed metabolism includes upregulated inflammatory activities, the elevated vulnerability of plaque, downregulated fatty acid oxidation (FAO), increased consumption of amino acids (AAs) and upregulated glycolysis in plaque²⁰⁰. Abnormal glycolysis always fosters the production of the inflammation-stimulated IL-1 β and IL-6^{201,202}. Accordingly, supplementary of FAO, during the activation of M2 macrophages²⁰³ and T cells²⁰⁴, may stimulate anti-inflammation signals directly or make CD8 $^{+}$ T cells exert indirect anti-inflammatory activity^{203–205}. Reduced metabolism of AAs may decrease foam cell formation and reduce plaque size^{206,207}.

4.1.1.5. Vaccination against low density lipoprotein (LDL) particles. AS does not belong to an autoimmune disease, however, ApoB is always known as AS antigens. As a result, autoimmune responses against ApoB *via* vaccination can be a potential therapeutic implication for AS¹⁷¹. The vaccination therapy includes using mAbs against the cholesterol ester transfer protein (CETP) or proprotein convertase subtilisin kexin type 9 (PCSK9) and induction of antigen-specific Tregs. PCSK9 is able to damage the LDL receptor and raise plasma LDL cholesterol level²⁰⁸, whereas CETP strengthens the change of high density lipoprotein (HDL)-LDL²⁰⁹. Consequently, anti-PCSK9 allows for reductions

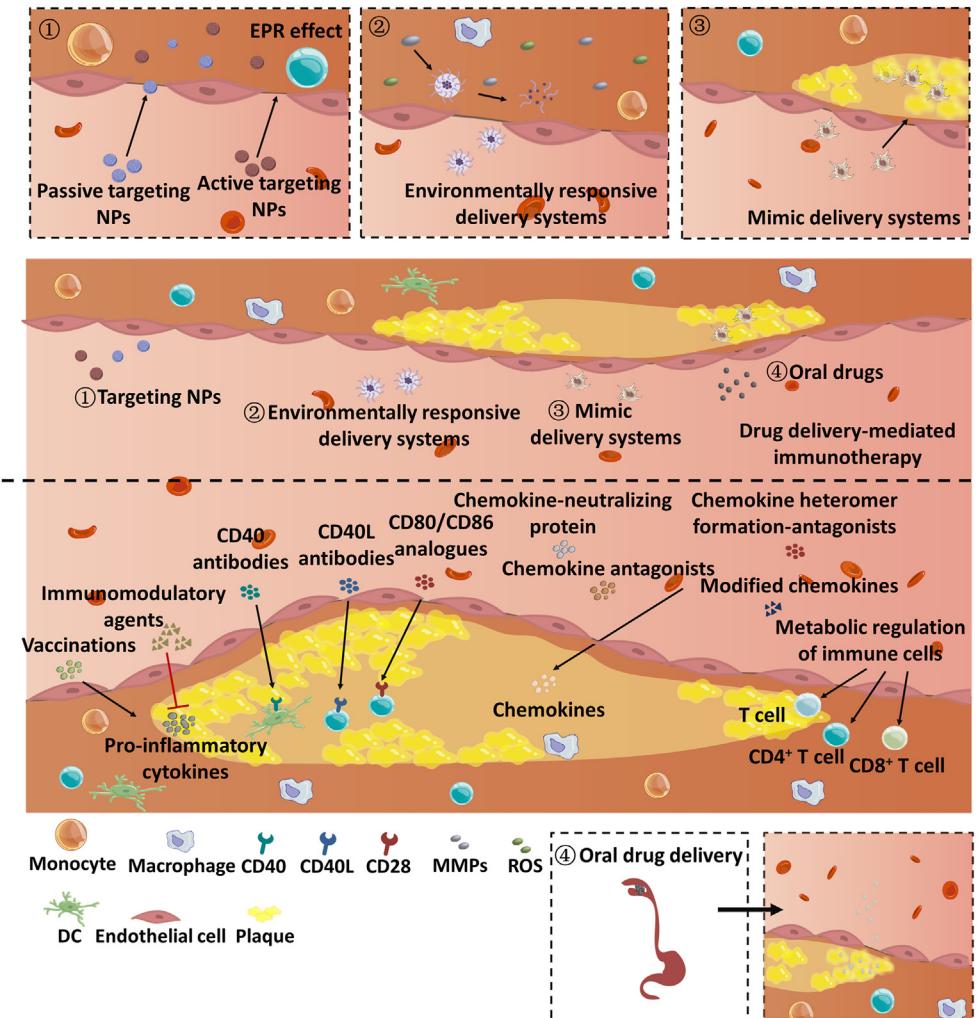


Figure 4 Immunotherapy for atherosclerosis (AS) and the used DDSs.

of LDL cholesterol^{210,211}, and vaccination with CETP could promote HDL cholesterol levels and decrease the plaque size²¹². In addition, Tregs can suppress the plaque development through limiting Teffs expansion, especially Th1 cells, and reduced production of inflammatory cytokine^{213,214}. Accordingly, stimulation of the Tregs may inhibit AS progression by suppressing the activities of immune cells including T cells, NK cells, monocytes, B cells, and DCs and by inducing suppressive modulators such as IL-10, TGF- β and IL-35²¹⁵.

4.1.2. Drug delivery-mediated immunotherapy in AS

The formation of plaque offers numerous opportunities for targeting therapy of AS by using DDS. The commonly reported DDSs are summarized in Fig. 4. The targeting approaches include enhanced accumulation of DDS in the plaque through enhanced permeability and retention effect (EPR)-like or biomimetic mechanism and promoted drug release from DDS in the plaque by microenvironment-responsive strategy. Beldman and co-workers²¹⁶ used a kind of HA-NPs to investigate the EPR effect in the plaque of AS progression. They found that the endothelial junction architecture normalized at the later period of AS compared with early AS and the accumulated HA-NPs was decreased. However, the HA-NPs can enter the plaque *via* endothelial junctions, distribute throughout the extracellular matrix (ECM) and eventually phagocytized by plaque-associated macrophages.

A recent study indicated the unusual and condensed cell morphology and junction irregularities in arterial endothelial layer of *ApoE*^{-/-} mouse observed by transmission electron microscopy²¹⁶. In the endothelial junctions of AS, the distance between vascular endothelial cadherin (VEC) units is up to 3 μm ²¹⁶. Whereas, in the normal vascular endothelial layer, the VEC units were firmly ranked and the space between the VEC units is only about 0.5 μm ²¹⁶. Nonetheless, advanced plaques have a small recovery of endothelial junction²¹⁶. Whereas, in the normal vascular endothelial layer, the VEC units were firmly ranked and the space between the VEC units is only about 0.5 μm ²¹⁶. However, the stage of AS affects the accumulation and the trafficking pathway of NPs. In advanced plaque, the accumulation of NPs at the AS lesions was closely relied on the transcellular route and was reduced around 30% compared with that in early plaque²¹⁶. Anyway, the findings rationalize the use of nanoscaled DDS for target treatment of AS. So far, DDSs have been widely utilized for AS immunotherapy, such as liposomes^{217,218}, recombinant HDL (rHDL) NPs²¹⁹, nanofiber²²⁰, membrane-coated NPs²²¹, polymersomes²²², and injectable filamentous hydrogel loaded micelles⁸¹.

Targeting macrophages are the most commonly used strategy in AS immunotherapy^{1,167,219}. The ligand signaling of CD40–CD40L is a widely known enhancer of AS and other chronic inflammatory diseases; consequently, its inhibition allows

for inhibition of AS²²³. Whereas tumor necrosis factor receptor-associated factor 6 (TRAF6) is potent to boost CD40's signaling cascade inside monocytes and macrophages¹⁹⁰. As a result, disruption of CD40-TRAF6 interactions can reduce monocyte recruitment to plaques and inhibition the formation of plaque¹⁹⁰. Recently, to suppress the interplay between CD40 and TRAF6 in macrophages and monocytes, a CD40-TRAF6 inhibitor 6877002 was loaded into 20-nm TRAF6i-rHDL NPs consisted of apolipoprotein A-I (ApoA-I), the 1,2-dimyristoyl-sn-glycero-3-phosphatidylcholine (DMPC) and 1-myristoyl-2-hydroxy-sn-glycero-phosphocholine (MHPC)²²⁴. The results demonstrated that TRAF6i-HDL NPs could bind well to monocytes and macrophages in the lesion site. The one-week treatment decreased the content of plaque macrophage content and plaque inflammation through the reduction of monocyte accumulation instead of decrement of local macrophage proliferation²²⁴. The results demonstrated that TRAF6i-HDL NPs could bind well to monocytes and macrophages in the lesion site. The one-week treatment decreased the content of plaque macrophage content and plaque inflammation through the reduction of monocyte accumulation instead of decrement of local macrophage proliferation²¹⁹. TRAF-STOPS enabled AS inhibition by limiting chemokine-induced accumulation of leukocyte to the plaques and suppressing release of cytokine from macrophages. Upon encapsulation in the rHDL NPs, their treatment efficacy was improved, displayed as reduced administration times and total dose over treatment with free TRAF-STOP²¹⁹.

Targeting Tregs or DCs are promising for AS immunotherapy^{215,225–227}. DCs have an extremely lower concentration compared with monocytes and macrophages within AS plaque, however, they have an unignored role in promoting the inflammation and AS advance^{228–230}. Yi et al.²²² prepared three types NPs, 20 nm-micelles, 100-nm polymersomes, and 50 nm × micron-length filomicelles using PEG-bl-PPS block copolymers. The results demonstrated that, among the NPs, the 100-nm polymersomes had the highest targeting ability to DCs in the arterial wall and lymphoid organs of animal model²²². By surface decoration with a P-D2 peptide, the polymersomes could well target DCs and enhance the cytosolic delivery of anti-inflammatory agent 1,25-dihydroxy vitamin D3-(aVD)²³¹. Low-dose intravenous administration for a week markedly inhibits the progression of plaque in a high-fat-diet-fed *ApoE*^{-/-} mice²³¹.

AS immunotherapy acquired via activating Tregs include adoptive transfer Tregs, AS relevant antigens such as x-LDL, HSP60, and ApoB100, pharmacological agents such as rapamycin, vitamin D3 and cholesterol-lowering drugs, and antibodies and cytokines such as IL-2 and anti-CD3 antibody²¹⁵. To improve the delivery of the immunostimulators to Tregs, liposomes formulated with the anionic phospholipid 1,2-distearoyl-sn-glycero-3-phosphoglycerol²³², pH-responsive NPs²³³ and filomicelles⁸¹ were developed to selectively deliver LDL-derived peptide antigen, miR-33, and vitamin D to Tregs.

AS immunotherapy displays promising treatment efficacy in preclinical studies, and some of them are undergoing clinical trials²³⁴. However, their translation is still hampered by various factors, e.g., unknown side effects and immunogenicity, poor scalability of DDS, high cost, and poor patient compliance because the injection is always required. AS immunotherapy obtained via oral administration benefits the translation, whereas it is challenged to prevent the degradation of the biopharmaceuticals or DDS in the gastrointestinal tract. Nonetheless, several oral formulations for AS immunotherapy have been investigated in

preclinical studies, such as yeast-derived microcapsules (YCs) encapsulated an inhibitor of MCP-1/CCL2 bindarit²³⁵, recombinant *Mycobacterium smegmatis*, a live bacterial vector, that allowed to produce cloned Chlamydia pneumonia (AHC) antigen and induce regulatory immune response to self-proteins^{236,237}, low-dose oral cyclophosphamide formulation²³⁸, algae-based vaccine²³⁹, and carrot-cell vaccine platform²⁴⁰.

4.2. PAH

PAH is characterized by an average pulmonary arterial pressure of >25 mmHg while a capillary wedge pressure of ≤15 mmHg. PAH is an uncommon and serious disease demonstrated as pulmonary vascular remodeling, endothelial abnormality, vasoconstriction and *in situ* inflammation and thrombosis²⁴¹. Most immune cells, i.e., T cells, DCs, NK cells, macrophages, B cells, mast cells, and eosinophils, are involved in the progression of PAH^{1,242}. Accordingly, these immunity pathways can be potential targets for treat PAH. Implications of immunotherapy in PAH are illustrated in Fig. 5.

4.2.1. Implications for immunotherapy in PAH

4.2.1.1. Cell-based therapy. Increasing evidence indicates Tregs involve in all stages of PAH pathogenesis, and reducing their number and activities always excrete PAH^{243,244}. A previous review summarized the function of Tregs in PAH²⁴⁵. Tregs inhibit the development of PAH through producing cytokines and chemokines such as IL-10, bone morphogenetic protein type II receptor (BMPR-II) and CXCL12–CXCR4, relating with other immune cells to suppress the immune activity and, as a result, repair injured pulmonary artery endothelial cells (PAECs), control proliferation and apoptosis of pulmonary arterial smooth muscle cells (PASMCs), limit proliferation and activation of fibroblast, and stay immune homeostasis²⁴⁵. Consequently, Tregs is a potential treatment target against PAH. The Tregs-targeted treatment includes adoptive Treg therapy acquired by exogenous Treg transplantation and expansion of intrinsic Tregs induced by stimulators including Liver kinase B1²⁴⁶, IL-2^{247,248}, vitamin D²⁴⁹, and CD28 superagonist²⁵⁰. Of the stimulators, IL-2 is most frequently applied and is demonstrated rous ability to promote the proliferation of Tregs. Until now, approximately fifty one clinical tests using Treg therapy have been recorded in Clinical-Trials.gov²⁵¹. A phase I trial demonstrated that the infused Tregs in patients with T1D could last one year²⁵², indicating safety and tolerance. Recently, adoptive Treg cell therapy was used on a patient with systemic lupus erythematosus (SLE)²⁵³. The results displayed that the treatment could increase activated Treg cells in inflamed skin and promote a shift from Th1 toward Th17 reactions²⁵³. Another clinical trial using rituximab to delete B cells for PAH immunotherapy is ongoing²⁵⁴.

Myeloid-derived suppressor cells (MDSCs) were reported to be involved in the development of PAH and several inflammatory diseases²⁵⁵. PD-L1 is overexpressed on MDSC from PAH patients²⁵⁵, and PD-1/PD-L1 interactions exacerbate the inflammation in PAH in animal model²⁵⁶. A report displayed that therapy using anti-PD-1 or PD-L1 might inhibit MDSC and alleviate the progression of PAH²⁵⁶.

4.2.1.2. Cytokine- and chemokine-based and vaccination therapy. The cytokines, e.g., IL-6, IL-8, IL-10, IL-13, IL-18, IL-1β and TNFα, are intimately associated with development of

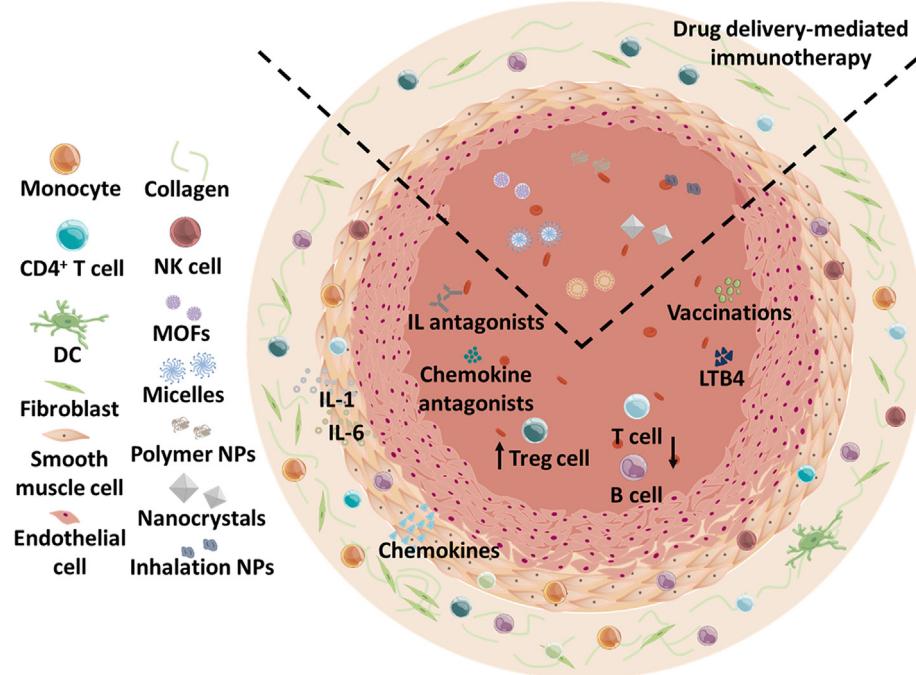


Figure 5 Immunotherapy for pulmonary arterial hypertension (PAH) and the used DDSs.

PAH^{257–259}. Inhibition of the proinflammatory cytokines is potential to treat PAH. Clinical test was performed to study the effectiveness against PAH using a IL-6 receptor antagonist, tocilizumab²⁶⁰. The results revealed that the treatment with tocilizumab was safe and improved pulmonary hemodynamic parameters²⁶⁰. Dosing a TNF- α antagonist, recombinant human TNF- α receptor II-IgG Fc fusion protein (rhTNFRFc), alleviated PAH *via* lowering mean pulmonary artery pressure (mPAP) and inhibiting pulmonary vascular remodeling²⁶¹. Furthermore, disruption of the IL-6/Th17/IL-21 pathway is promising to selectively treat PAH²⁶².

Numerous chemokines are involved in inflammation and pulmonary vascular remodeling, including CXCL8/CXCR1/CXCR2, CXCL10/CXCR3, CXCL12/CXCR4/ACKR3, CCL2/CCR2, CCL5/CCR5/CCR1, CX3CL1/CX3CR1, etc.²⁶³. In particular, leukotriene B4 (LTB4) has robust ability to promote inflammatory immune response *via* increasing neutrophil recruitment and, as a result, induce apoptosis of ECs^{264,265}. Inhibition of LTB4 with bestatin allows reversing established PAH *via* increasing the numbers of open arterioles and reducing arteriolar wall thickness and muscularization²⁶⁴.

Endothelin-1 (ET-1) receptor type A (ETAR) can activate the endothelin system and facilitate the initiation and development of PAH²⁶⁶. A vaccine against ETAR was designed by conjugating an ETR-002 peptide with a Q β bacteriophage virus-like particle²⁶⁷. The vaccination approach has potent efficacy to combat PAH in the monocrotaline (MCT)-induced- and Sugen/hypoxia-induced models by suppressing the pulmonary arterial remodeling and the RV hypertrophy through inhibition of Ca²⁺-dependent signal transduction events²⁶⁷. In addition, disruption of the α 1D-adrenergic receptor (α 1D-AR) might be a vaccination strategy against hypertension by using ADRQ β -004 vaccine²⁶⁸.

4.2.2. Drug delivery-mediated immunotherapy in PAH

Numerous DDSs are applied to improve immunotherapy; however, their use in PAH immunotherapy moves forward slowly. The

most commonly used DDSs to elevate pulmonary delivery are liposomes^{152,269,270} and polymeric NPs^{269,271–273} dosed *via* intravenous injection or inhalation²⁷⁴. Loss of endothelial BMPR-II facilitates the initiation and development of PAH, enabling BMPR-II to be a therapeutic target²⁷⁵. Tacrolimus, an immuno-suppressor, is able to activate BMPR-II and is allowed to repair the endothelial function in PAH patient cells and inhibit the remodeling of the pulmonary artery in animal model²⁷⁶. A clinical test demonstrated that administration of tacrolimus at a low dose to three patients with advanced PAH for twelve months, which the trough concentration was 1.5–5 ng/mL, upregulated BMPR-II in PBMCs and, as a result, ameliorated PAH through elevating heart function, prolonging 6-min walk distance, and inducing N-terminal pro-brain natriuretic peptide²⁷⁷. Another clinical trial displayed that this administration regimen was safe and could promote the expression of BMPR-II in subsets of PAH patients²⁷⁸. To improve the pulmonary delivery of tacrolimus, nanocomposite microparticles (nCmPs) were prepared by formulating 200-nm drug-loaded polymeric NPs into microparticles through spray drying. After administration *via* inhalation, the nCmP could deposit in the lung regions, penetrate through the mucus barrier, and control drug release over time²⁷⁹. In addition, other immunomodulators, *e.g.*, rapamycin, everolimus, anti-TNF α , TGF- β antagonist, rituximab, and tocilizumab²⁸⁰, were used to combat PAH as well.

5. Conclusions and outlook

We summarized the potential immune targets in several major inflammatory diseases, reviewed the biological drugs and DDSs used for immunotherapy. Immunotherapy is updating the concept of disease treatment and has acquired rapid development in the past five years, evident by that several products such as mAbs and adoptive cell transfer were approved for clinical use. In particular, immunotherapy is being developed as a most effective strategy

against cancer. For RA and IBD immunotherapy, the progression is being promoted smoothly, along with several mAbs against TNF- α and IL-6 and two JAK inhibitors, baricitinib and tofacitinib, being marked, whereas there are seven mAbs approved for IBD immunotherapy. For immunotherapy of vascular diseases such as AS and PAH, the clinical test demonstrated promising potential, *e.g.*, the treatment efficacy against AS with a mAb targeting IL-1 β can persist three months¹⁸³, bringing significant connivance to patients who have to take lipid-lowering drugs daily. So far, there is no report regarding clinical trials to ameliorate PAH. Always, the patients with the advanced PAH possess poor response to the frequently applied vasodilator agents probably due to the loss of elasticity in the remodeling pulmonary arteries. Such that the utilization of immunotherapy may reverse PAH; however, the rationalization is required from the physician. Overall, immunotherapy against serious vascular diseases don't move forward smoothly compared with cancer immunotherapy, mainly owing to the factors: (1) in pathogenesis lacking sufficiently understanding toward the immune pathways involved in these diseases; (2) absence of legitimacy from the clinic; (3) the potential immune-related adverse events²⁸¹; (4) remarkably high cost compared with the conventionally used treatment regimens; (5) patients' compliance because dosing *via* injection is always required in most immunotherapy.

In general, the strategies for immunotherapy are predominantly categorized into several types, including mAbs against cytokines or chemokines, inhibitory ICPS, JAK inhibitors, adoptive cell transfer, metabolic regulation of immune cells and vaccination. mAb-immunotherapy is the most widely applied approach and has gained huge success, evident by over seventy-four formulations have entered the market. Second, ACT, especially T cell-based transplantation, is attracting increasing attention and advances rapidly. The milestone event of this technique is the approval use of CAR therapy to treat for relapsed/refractory acute lymphoblastic leukemia (ALL)²⁸². After that, other ACTs are constantly emerging, such as TIL-, TCR-, NK Cell-, Treg-, and MDSC-therapies. Although several problems regarding ACT, such as safety, efficacy, and persistence, are needed to be addressed, the ACT will be concentrated continuously and an increasing number of commercial products will be approved, mainly due to its advantages such as simple composition and controllable scalability. Particularly, Tregs are demonstrating potent efficacy to combat serious inflammation and over 50 of Treg techniques were registered for clinical trial²⁵¹. This technique deserves much more attention and we believe increasing products will be marked for clinical usage.

Most of the immunomodulators belong to biological drugs and their application is always limited by their large size, poor stability, humble penetration ability across physiological barriers, rapid clearance by the reticuloendothelial system, etc. To improve immunotherapy, repeatedly dosings at high doses of the biological drugs *via* intravenous injection are always required, leading to safety concerns and significantly reducing patient's compliance. The drug delivery approach through engineering biomaterials is robust to enhance delivery of the biologics to the targeted site. Tremendous DDSs demonstrated their amazing immunotherapy efficacy toward various inflammatory diseases in pre-clinical studies. Nonetheless, extremely limited DDS-mediated immunotherapy is approved frequently due to DDS's potential toxicity to the body, unknown *in vivo* fate²⁸³, modest scale-up ability, and unfriendly dosing route. In this case, the selection of DDS is critical to the translation. As a result, DDSs with excellent safety

and promising industrial perspectives may be an optional choice for the DDS immunotherapy. These DDSs include liposomes or liposome-like NPs, degradable polymeric carriers such as PLGA-NPs or microspheres, albumin-based NPs, cell carriers like red blood cells, etc. In addition, the dosing routes are of the essence to the translation, and well-accepted delivery pathways should be first choice, encompassing oral, buccal, transdermal, nasal, inhalation and subcutaneous routes²⁸⁴.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (Nos. 81872823 and 82073782), the Double First-Class (CPU2018PZQ13, China) of the China Pharmaceutical University, the Shanghai Science and Technology Committee (No. 19430741500, China), the Key Laboratory of Modern Chinese Medicine Preparation of Ministry of Education of Jiangxi University of Traditional Chinese Medicine (TCM-201905, China), and the Start-up Grant from City University of Hong Kong (No. 9610472, China).

Author contributions

Wei He conceived the work. Qingqing Xiao, Xiaotong Li, Yi Li, Zhengfeng Wu, Chenjie Xu, Zhongjian Chen, and Wei He co-wrote the paper. Xiaotong Li prepared the figures. All of the authors discussed the results and commented on the manuscript. All of the authors have read and approved the final manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

- He W, Kapate N, IV CWS, Mitragotri S. Drug delivery to macrophages: a review of targeting drugs and drug carriers to macrophages for inflammatory diseases. *Adv Drug Deliv Rev* 2020;**165-166**:15–40.
- Garn H, Bahn S, Baune BT, Binder EB, Bisgaard H, Chatila TA, et al. Current concepts in chronic inflammatory diseases: interactions between microbes, cellular metabolism, and inflammation. *J Allergy Clin Immunol* 2016;**138**:47–56.
- Galluzzi L, Chan TA, Kroemer G, Wolchok JD, López-Soto A. The hallmarks of successful anticancer immunotherapy. *Sci Transl Med* 2018;**10**:eaat7807.
- Till SJ, Francis JN, Nouri-Aria K, Durham SR. Mechanisms of immunotherapy. *J Allergy Clin Immunol* 2004;**113**:1025–34.
- Tan SZ, Li DP, Zhu X. Cancer immunotherapy: pros, cons and beyond. *Biomed Pharmacother* 2020;**124**:109821.
- Steffens S, Weber C. Immunotherapy for atherosclerosis—novel concepts. *Thromb Haemostasis* 2019;**119**:515–6.
- Ahmed M, Bae Y-S. Dendritic cell-based immunotherapy for rheumatoid arthritis: from bench to bedside. *Immune Netw* 2016;**16**:44–51.
- Catalan-Serra I, Brenna Ø. Immunotherapy in inflammatory bowel disease: novel and emerging treatments. *Hum Vaccines Immunother* 2018;**14**:2597–611.
- Nicolls MR, Voelkel NF. The roles of immunity in the prevention and evolution of pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2017;**195**:1292–9.
- Sharma P, Hu-Lieskovian S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell* 2017;**168**:707–23.

11. Law AMK, Lim E, Ormandy CJ, Gallego-Ortega D. The innate and adaptive infiltrating immune systems as targets for breast cancer immunotherapy. *Endocr Relat Cancer* 2017;**24**:R123–44.
12. Silva LCR, Ortigosa LCM, Benard G. Anti-TNF- α agents in the treatment of immune-mediated inflammatory diseases: mechanisms of action and pitfalls. *Immunotherapy* 2010;**2**:817–33.
13. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 2012;**12**:252–64.
14. Zhang C, Wu Z, Li JW, Zhao H, Wang GQ. Cytokine release syndrome in severe COVID-19: interleukin-6 receptor antagonist tocilizumab may be the key to reduce mortality. *Int J Antimicrob Agents* 2020;**55**:105954.
15. Hara Y, Nagaoka S. *Nivolumab (Opdivo)*. Singapore: Springer Singapore; 2019.
16. Wills S, Hochmuth LK, Bauer KS, Durvalumab Deshmukh R. A newly approved checkpoint inhibitor for the treatment of urothelial carcinoma. *Curr Probl Cancer* 2019;**43**:181–94.
17. Subklewe M, von Bergwelt-Baildon M, Humpe A. Chimeric antigen receptor T cells: a race to revolutionize cancer therapy. *Transfus Med Hemotherapy* 2019;**46**:15–24.
18. Strohl WR. Current progress in innovative engineered antibodies. *Protein cell* 2018;**9**:86–120.
19. Schwartz DM, Bonelli M, Gadina M, O’Shea JJ. Type I/II cytokines, JAKs, and new strategies for treating autoimmune diseases. *Nat Rev Rheumatol* 2016;**12**:25–36.
20. Sanjabi S, Oh SA, Li MO. Regulation of the immune response by TGF- β : from conception to autoimmunity and infection. *Cold Spring Harb Perspect Biol* 2017;**9**:a022236.
21. Mullard A. 2017 FDA drug approvals. *Nat Rev Drug Discov* 2018; **17**:81–5.
22. Mullard A. 2012 FDA drug approvals. *Nat Rev Drug Discov* 2013; **12**:87–90.
23. Alsaab HO, Sau S, Alzhrani R, Tatiparti K, Bhise K, Kashaw SK, et al. PD-1 and PD-L1 checkpoint signaling inhibition for cancer immunotherapy: mechanism, combinations, and clinical outcome. *Front Pharmacol* 2017;**8**:561.
24. Agarwala SS. Practical approaches to immunotherapy in the clinic. *Semin Oncol* 2015;**42**:S20–7.
25. Pföhler C, Eichler H, Burgard B, Krecké N, Müller CSL, Vogt T. A case of immune thrombocytopenia as a rare side effect of an immunotherapy with PD1-blocking agents for metastatic melanoma. *Transfus Med Hemotherapy* 2017;**44**:426–8.
26. Vial T, Descotes J. Immune-mediated side-effects of cytokines in humans. *Toxicology* 1995;**105**:31–57.
27. He W, Xing XY, Wang XL, Wu D, Wu W, Guo JL, et al. Nanocarrier-mediated cytosolic delivery of biopharmaceuticals. *Adv Funct Mater* 2020;1910566. n/a.
28. Wu W, Li TL. Unraveling the *in vivo* fate and cellular pharmacokinetics of drug nanocarriers. *Adv Drug Deliv Rev* 2019;**143**:1–2.
29. Zhao ZM, Ukidve A, Krishnan V, Mitragotri S. Effect of physico-chemical and surface properties on *in vivo* fate of drug nanocarriers. *Adv Drug Deliv Rev* 2019;**143**:3–21.
30. Xiao QQ, Zhu X, Yuan YT, Yin LF, He W. A drug-delivering-drug strategy for combined treatment of metastatic breast cancer. *Nanomed-Nanotechnol* 2018;**14**:2678–88.
31. Jin K, Luo ZM, Zhang B, Pang ZQ. Biomimetic nanoparticles for inflammation targeting. *Acta Pharm Sin B* 2018;**8**:23–33.
32. Mao YS, Zou CF, Jiang YJ, Fu DL. Erythrocyte-derived drug delivery systems in cancer therapy. *Chin Chem Lett* 2021;**32**:990–8.
33. Donahue ND, Acar H, Wilhelm S. Concepts of nanoparticle cellular uptake, intracellular trafficking, and kinetics in nanomedicine. *Adv Drug Deliv Rev* 2019;**143**:68–96.
34. Su C, Liu YZ, Li RZ, Wu W, Fawcett JP, Gu JK. Absorption, distribution, metabolism and excretion of the biomaterials used in nanocarrier drug delivery systems. *Adv Drug Deliv Rev* 2019;**143**:97–114.
35. Zhu YF, Yu XR, Thamphiwatana SD, Zheng Y, Pang ZQ. Nanomedicines modulating tumor immunosuppressive cells to enhance cancer immunotherapy. *Acta Pharm Sin B* 2020;**10**:2054–74.
36. Lu Y, Li Y, Wu W. Injected nanocrystals for targeted drug delivery. *Acta Pharm Sin B* 2016;**6**:106–13.
37. Corrales L, Glickman LH, McWhirter SM, Kanne DB, Sivick KE, Katibah GE, et al. Direct activation of STING in the tumor microenvironment leads to potent and systemic tumor regression and immunity. *Cell Rep* 2015;**11**:1018–30.
38. Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, et al. Cytokines in clinical cancer immunotherapy. *Br J Cancer* 2019;**120**:6–15.
39. Quesada JR, Hersh EM, Manning J, Reuben J, Keating M, Schnipper E, et al. Treatment of hairy cell leukemia with recombinant alpha-interferon. *Blood* 1986;**68**:493–7.
40. Rosenberg SA. IL-2: the first effective immunotherapy for human cancer. *J Immunol* 2014;**192**:5451–8.
41. Rosenberg SA, Lotze MT, Mule LM, Chang AE, Avis FP, Leitman S, et al. A progress report on the treatment of 157 patients with advanced cancer using lymphokine-activated killer cells and interleukin-2 or high-dose interleukin-2 alone. *N Engl J Med* 1987; **316**:889–97.
42. Waldmann TA. Cytokines in cancer immunotherapy. *Cold Spring Harb Perspect Biol* 2018;**10**:a028472.
43. Palucka K, Banchereau J. Dendritic-cell-based therapeutic cancer vaccines. *Immunity* 2013;**39**:38–48.
44. Jahanafrooz Z, Baradaran B, Mosafer J, Hashemzaei M, Rezaei T, Mokhtarzadeh A, et al. Comparison of DNA and mRNA vaccines against cancer. *Drug Discov Today* 2020;**25**:552–60.
45. Kimiz-Geboglu I, Gulce-Iz S, Biray-Avcı C. Monoclonal antibodies in cancer immunotherapy. *Mol Biol Rep* 2018;**45**:2935–40.
46. Jafari S, Molavi O, Kahroba H, Hejazi MS, Maleki-Dizaji N, Barghi S, et al. Clinical application of immune checkpoints in targeted immunotherapy of prostate cancer. *Cell Mol Life Sci* 2020;**77**:3693–710.
47. Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. *EMBO J* 1992;**11**:3887–95.
48. Brunet JF, Denizot F, Luciani MF, Roux-Dosseto M, Suzan M, Mattei MG, et al. A new member of the immunoglobulin superfamily-CTLA-4. *Nature* 1987;**328**:267–70.
49. Aspeslagh S, Postel-Vinay S, Rusakiewicz S, Soria J-C, Zitvogel L, Marabelle A. Rationale for anti-OX40 cancer immunotherapy. *Eur J Cancer* 2016;**52**:50–66.
50. Buchbinder EI, Desai A. CTLA-4 and PD-1 pathways: similarities, differences, and implications of their inhibition. *Am J Clin Oncol* 2016;**39**:98–106.
51. Chambers CA, Kuhns MS, Egen JG, Allison JP. CTLA-4-mediated inhibition in regulation of T cell responses: mechanisms and manipulation in tumor immunotherapy. *Annu Rev Immunol* 2001;**19**:565–94.
52. Postow MA, Sidlow R, Hellmann MD. Immune-related adverse events associated with immune checkpoint blockade. *N Engl J Med* 2018;**378**:158–68.
53. Iwai Y, Okazaki T, Nishimura H, Kawasaki A, Yagita H, Honjo T. Microanatomical localization of PD-1 in human tonsils. *Immunol Lett* 2002;**83**:215–20.
54. Patsoukis N, Duke-Cohan JS, Chaudhri A, Aksoylar H-I, Wang Q, Council A, et al. Interaction of SHP-2 SH2 domains with PD-1 ITSM induces PD-1 dimerization and SHP-2 activation. *Commun Biol* 2020;**3**:128.
55. Iwai Y, Hamanishi J, Chamoto K, Honjo T. Cancer immunotherapies targeting the PD-1 signaling pathway. *J Biomed Sci* 2017;**24**:26.
56. Parry RV, Chemnitz JM, Frauwirth KA, Lanfranco AR, Braunstein I, Kobayashi SV, et al. CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. *Mol Cell Biol* 2005;**25**:9543–53.

57. Lee HW, Cho KJ, Park JY. Current status and future direction of immunotherapy in hepatocellular carcinoma: what do the data suggest?. *Immune Netw* 2020;20:e11.
58. Faruki H, Mayhew GM, Serody JS, Hayes DN, Perou CM, Lai-Goldman M. Lung adenocarcinoma and squamous cell carcinoma gene expression subtypes demonstrate significant differences in tumor immune landscape. *J Thorac Oncol* 2017;12:943–53.
59. Ö Met, Jensen KM, Chamberlain CA, Donia M, Svane IM. Principles of adoptive T cell therapy in cancer. *Semin Immunopathol* 2019;41:49–58.
60. June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* 2018;359:1361–5.
61. Andersen R, Borch TH, Draghi A, Gokuldass A, Rana MAH, Pedersen M, et al. T cells isolated from patients with checkpoint inhibitor-resistant melanoma are functional and can mediate tumor regression. *Ann Oncol* 2018;29:1575–81.
62. Rapoport AP, Stadtmauer EA, Binder-Scholl GK, Goloubanova O, Vogl DT, Lacey SF, et al. NY-ESO-1-specific TCR-engineered T cells mediate sustained antigen-specific antitumor effects in myeloma. *Nat Med* 2015;21:914–21.
63. Newick K, O'Brien S, Moon E, Albelda SM. CAR T cell therapy for solid tumors. *Annu Rev Med* 2017;68:139–52.
64. Zhang J, Wang L. The emerging world of TCR-T cell trials against cancer: a systematic review. *Technol Cancer Res Treat* 2019;18:1533033819831068.
65. Barrett DM, Grupp SA, June CH. Chimeric antigen receptor- and TCR-modified T cells enter main street and wall street. *J Immunol* 2015;195:755–61.
66. Reddy ST, Rehor A, Schmoekel HG, Hubbell JA, Swartz MA. *In vivo* targeting of dendritic cells in lymph nodes with poly(propylene sulfide) nanoparticles. *J Control Release* 2006;112:26–34.
67. He HS, Lu Y, Qi JP, Zhu QG, Chen ZJ, Wu W. Adapting liposomes for oral drug delivery. *Acta Pharm Sin B* 2019;9:36–48.
68. Da Silva CG, Rueda F, Löwik CW, Ossendorp F, Cruz LJ. Combinatorial prospects of nano-targeted chemoimmunotherapy. *Biomaterials* 2016;83:308–20.
69. Caster JM, Callaghan C, Seyedin SN, Henderson K, Sun B, Wang AZ. Optimizing advances in nanoparticle delivery for cancer immunotherapy. *Adv Drug Deliv Rev* 2019;144:3–15.
70. Mishra P, Nayak B, Dey RK. PEGylation in anti-cancer therapy: an overview. *Asian J Pharm Sci* 2016;11:337–48.
71. Schmid D, Park CG, Hartl CA, Subedi N, Cartwright AN, Puerto RB, et al. T cell-targeting nanoparticles focus delivery of immunotherapy to improve antitumor immunity. *Nat Commun* 2017;8:1747.
72. Buabeid MA, Arafa EA, Murtaza G. Emerging prospects for nanoparticle-enabled cancer immunotherapy. *J Immunol Res* 2020;2020:9624532.
73. Ke X, Howard GP, Tang H, Cheng B, Saung MT, Santos JL, et al. Physical and chemical profiles of nanoparticles for lymphatic targeting. *Adv Drug Deliv Rev* 2019;151–152:72–93.
74. Moon JJ, Huang B, Irvine DJ. Engineering nano- and microparticles to tune immunity. *Adv Mater* 2012;24:3724–46.
75. Riley RS, June CH, Langer R, Mitchell MJ. Delivery technologies for cancer immunotherapy. *Nat Rev Drug Discov* 2019;18:175–96.
76. Man F, Gawne PJ, TMdR R. Nuclear imaging of liposomal drug delivery systems: a critical review of radiolabelling methods and applications in nanomedicine. *Adv Drug Deliv Rev* 2019;143:134–60.
77. Peng JR, Yang Q, Shi K, Xiao Y, Wei XW, Qian ZY. Intratumoral fate of functional nanoparticles in response to microenvironment factor: implications on cancer diagnosis and therapy. *Adv Drug Deliv Rev* 2019;143:37–67.
78. Smith TT, Stephan SB, Moffett HF, McKnight LE, Ji W, Reiman D, et al. *In situ* programming of leukaemia-specific T cells using synthetic DNA nanocarriers. *Nat Nanotechnol* 2017;12:813–20.
79. Wang C, Sun WJ, Wright G, Wang AZ, Gu Z. Inflammation-triggered cancer immunotherapy by programmed delivery of CpG and anti-PD1 antibody. *Adv Mater* 2016;28:8912–20.
80. Jia YP, Ma BY, Wei XW, Qian ZY. The *in vitro* and *in vivo* toxicity of gold nanoparticles. *Chin Chem Lett* 2017;28:691–702.
81. Yin JF, Huang YX, Hameed SM, Zhou RY, Xie LJ, Ying YB. Large scale assembly of nanomaterials: mechanisms and applications. *Nanoscale* 2020;12:17571–89.
82. Sang W, Zhang Z, Dai YL, Chen XY. Recent advances in nanomaterial-based synergistic combination cancer immunotherapy. *Chem Soc Rev* 2019;48:3771–810.
83. He XS, Gershwin ME, Ansari AA. Checkpoint-based immunotherapy for autoimmune diseases – opportunities and challenges. *J Autoimmun* 2017;79:1–3.
84. Wraith D. Antigen-specific immunotherapy. *Nature* 2016;530:422–3.
85. Hilkens CM, Isaacs JD. Tolerogenic dendritic cell therapy for rheumatoid arthritis: where are we now?. *Clin Exp Immunol* 2013;172:148–57.
86. Weyand CM, Goronzy JJ. Immunometabolism in the development of rheumatoid arthritis. *Immunol Rev* 2020;294:177–87.
87. Salomon S, Guignant C, Morel P, Flahaut G, Brault C, Gourguechon C, et al. Th17 and CD24^{hi}CD27⁺ regulatory B lymphocytes are biomarkers of response to biologics in rheumatoid arthritis. *Arthritis Res Ther* 2017;19:33.
88. Lamas JR, Muciientes A, Lajas C, Fernández-Gutiérrez B, López Y, Marco F, et al. Check-control of inflammation displayed by bone marrow mesenchymal stem cells in rheumatoid arthritis patients. *Immunotherapy* 2019;11:1107–16.
89. Ehrenstein MR, Evans JG, Singh A, Moore S, Warnes G, Isenberg DA, et al. Compromised function of regulatory T cells in rheumatoid arthritis and reversal by anti-TNF α therapy. *J Exp Med* 2004;200:277–85.
90. Xu L, Song XL, Su LL, Zheng Y, Li R, Sun J. New therapeutic strategies based on IL-2 to modulate Treg cells for autoimmune diseases. *Int Immunopharm* 2019;72:322–9.
91. Semerano L, Minichiello E, Bessis N, Boissier M-C. Novel immunotherapeutic avenues for rheumatoid arthritis. *Trends Mol Med* 2016;22:214–29.
92. Rosser EC, Blair PA, Mauri C. Cellular targets of regulatory B cell-mediated suppression. *Mol Immunol* 2014;62:296–304.
93. Veen Wvd, Stanic B, Wirz OF, Jansen K, Globinska A, Akdis M. Role of regulatory B cells in immune tolerance to allergens and beyond. *J Allergy Clin Immunol* 2016;138:654–65.
94. Mielle J, Audo R, Hahne M, Macia L, Combe B, Morel J, et al. IL-10 producing B cells ability to induce regulatory T cells is maintained in rheumatoid arthritis. *Front Immunol* 2018;9.
95. Daien CI, Gailhac S, Mura T, Audo R, Combe B, Hahne M, et al. Regulatory B10 cells are decreased in patients with rheumatoid arthritis and are inversely correlated with disease activity. *Arthritis Rheum* 2014;66:2037–46.
96. Flores-Borja F, Bosma A, Ng D, Reddy V, Ehrenstein MR, Isenberg DA, et al. CD19⁺ CD24^{hi}CD38^{hi} B cells maintain regulatory T cells while limiting TH1 and TH17 differentiation. *Sci Transl Med* 2013;5: 173ra23-ra23.
97. Mauri C, Gray D, Mushtaq N, Londei M. Prevention of arthritis by interleukin 10-producing B cells. *J Exp Med* 2003;197:489–501.
98. Pozsgay J, Szekanecz Z, Sármay G. Antigen-specific immunotherapies in rheumatic diseases. *Adv Drug Deliv Rev* 2017;13:525–37.
99. Zhao X, Long J, Liang F, Liu N, Sun YY, Xi YZ. Vaccination with a novel antigen-specific tolerizing DNA vaccine encoding CCOL2A1 protects rats from experimental rheumatoid arthritis. *Hum Gene Ther* 2018;30:69–78.
100. McInnes IB, Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. *Nat Rev Immunol* 2007;7:429–42.
101. Eleamam NM, Hannawi S, Maghazachi AA. Role of chemokines and chemokine receptors in rheumatoid arthritis. *ImmunoTargets Ther* 2020;9:43–56.

102. McInnes IB, Buckley CD, Isaacs JD. Cytokines in rheumatoid arthritis—shaping the immunological landscape. *Nat Rev Rheumatol* 2016;12:63–8.
103. Davignon J-L, Rauwel B, Degboé Y, Constantin A, Boyer J-F, Kruglov A, et al. Modulation of T-cell responses by anti-tumor necrosis factor treatments in rheumatoid arthritis: a review. *Arthritis Res Ther* 2018;20:229.
104. Basetta JG, Stutman O. TNF regulates thymocyte production by apoptosis and proliferation of the triple negative (CD3⁻CD4⁻CD8⁻) subset. *J Immunol* 2000;165:5621–30.
105. Huang ZC, Yang B, Shi YY, Cai B, Li Y, Feng WH, et al. Anti-TNF- α therapy improves Treg and suppresses Teff in patients with rheumatoid arthritis. *Cell Immunol* 2012;279:25–9.
106. Rao DA, Gurish MF, Marshall JL, Slowikowski K, Fonseka CY, Liu YY, et al. Pathologically expanded peripheral T helper cell subset drives B cells in rheumatoid arthritis. *Nature* 2017;542:110–4.
107. Bankó Z, Pozsgay J, Gáti T, Rojkovich B, Ujfalussy I, Sármay G. Regulatory B cells in rheumatoid arthritis: alterations in patients receiving anti-TNF therapy. *Clin Immunol* 2017;184:63–9.
108. Narazaki M, Tanaka T, Kishimoto T. The role and therapeutic targeting of IL-6 in rheumatoid arthritis. *Expet Rev Clin Immunol* 2017;13:535–51.
109. Samarpita S, Kim JY, Rasool MK, Kim KS. Investigation of toll-like receptor (TLR) 4 inhibitor TAK-242 as a new potential anti-rheumatoid arthritis drug. *Arthritis Res Ther* 2020;22:16.
110. Olsen IC, Lie E, Vasilescu R, Wallenstein G, Strengtholt S, Kvien TK. Assessments of the unmet need in the management of patients with rheumatoid arthritis: analyses from the NOR-DMARD registry. *Rheumatology* 2019;58:481–91.
111. Yamaoka K. Janus kinase inhibitors for rheumatoid arthritis. *Curr Opin Chem Biol* 2016;32:29–33.
112. Fragoulis GE, McInnes IB, Siebert S. JAK-inhibitors. New players in the field of immune-mediated diseases, beyond rheumatoid arthritis. *Rheumatology* 2019;58:i43–54.
113. Nakayama S, Kubo S, Iwata S, Tanaka Y. Chemical JAK inhibitors for the treatment of rheumatoid arthritis. *Expet Opin Pharmacother* 2016;17:2215–25.
114. Pujol-Autonell I, Mansilla M-J, Rodriguez-Fernandez S, Cano-Sarabia M, Navarro-Barriuso J, Ampudia R-M, et al. Liposome-based immunotherapy against autoimmune diseases: therapeutic effect on multiple sclerosis. *Nanomedicine* 2017;12:1231–42.
115. Song P, Yang CX, Thomsen JS, Dagnæs-Hansen F, Jakobsen M, Brüel A, et al. Lipidoid-siRNA nanoparticle-mediated IL-1 β gene silencing for systemic arthritis therapy in a mouse model. *Mol Ther* 2019;27:1424–35.
116. Capini C, Jaturapinyo M, Chang H-I, Mutualik S, McNally A, Street S, et al. Antigen-specific suppression of inflammatory arthritis using liposomes. *J Immunol* 2009;182:3556–65.
117. Kishimoto TK, Ferrari JD, LaMothe RA, Kolte PN, Griset AP, O'Neil C, et al. Improving the efficacy and safety of biologic drugs with tolerogenic nanoparticles. *Nat Nanotechnol* 2016;11:890–9.
118. Khan D, Qindeel M, Ahmed N, Khan AU, Khan S, Au Rehman. Development of novel pH-sensitive nanoparticle-based transdermal patch for management of rheumatoid arthritis. *Nanomedicine* 2020;15:603–24.
119. Mohammadi M, Li Y, Abebe DG, Xie YR, Kandil R, Kraus T, et al. Folate receptor targeted three-layered micelles and hydrogels for gene delivery to activated macrophages. *J Control Release* 2016;244:269–79.
120. Lee H, Lee MY, Bhang SH, Kim BS, Kim YS, Ju JH, et al. Hyaluronate–gold nanoparticle/tocilizumab complex for the treatment of rheumatoid arthritis. *ACS Nano* 2014;8:4790–8.
121. Zou SJ, Wang BL, Wang C, Wang QQ, Zhang LM. Cell membrane-coated nanoparticles: research advances. *Nanomedicine* 2020;15:625–41.
122. He YW, Li RX, Liang JM, Zhu Y, Zhang SY, Zheng ZC, et al. Drug targeting through platelet membrane-coated nanoparticles for the treatment of rheumatoid arthritis. *Nano Res* 2018;11:6086–101.
123. Zhang QZ, Dehaini DN, Zhang Y, Zhou JL, Chen XY, Zhang LF, et al. Neutrophil membrane-coated nanoparticles inhibit synovial inflammation and alleviate joint damage in inflammatory arthritis. *Nat Nanotechnol* 2018;13:1182–90.
124. Gorantla S, Singhvi G, Rapalli VK, Waghule T, Dubey SK, Saha RN. Targeted drug-delivery systems in the treatment of rheumatoid arthritis: recent advancement and clinical status. *Ther Deliv* 2020;11:269–84.
125. Nogueira E, Gomes AC, Preto A, Cavaco-Paulo A. Folate-targeted nanoparticles for rheumatoid arthritis therapy. *Nanomed Nanotechnol Biol Med* 2016;12:1113–26.
126. Lyu YQ, Xiao QQ, Yin LF, Yang L, He W. Potent delivery of an MMP inhibitor to the tumor microenvironment with thermosensitive liposomes for the suppression of metastasis and angiogenesis. *Signal Transduct Tar* 2019;4:26.
127. Duan WF, Li H. Combination of NF- κ B targeted siRNA and methotrexate in a hybrid nanocarrier towards the effective treatment in rheumatoid arthritis. *J Nanobiotechnol* 2018;16:58.
128. Graham DB, Xavier RJ. Pathway paradigms revealed from the genetics of inflammatory bowel disease. *Nature* 2020;578:527–39.
129. Sun M, He C, Cong Y, Liu Z. Regulatory immune cells in regulation of intestinal inflammatory response to microbiota. *Mucosal Immunol* 2015;8:969–78.
130. Maloy KJ, Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature* 2011;474:298–306.
131. Cader MZ, Kaser A. Recent advances in inflammatory bowel disease: mucosal immune cells in intestinal inflammation. *Gut* 2013;62:1653–64.
132. Trivedi PJ, Adams DH. Chemokines and chemokine receptors as therapeutic targets in inflammatory bowel disease; pitfalls and promise. *J Crohns Colitis* 2018;12:S641–52.
133. Raad MA, Chams NH, Sharara AI. New and evolving immunotherapy in inflammatory bowel disease. *Inflammatory Intestinal Diseases* 2016;1:85–95.
134. Danese S, Vuitton L, Peyrin-Biroulet L. Biologic agents for IBD: practical insights. *Nat Rev Gastroenterol Hepatol* 2015;12:537–45.
135. Griffiths OR, Landon J, Coxon RE, Morris K, James P, Adams R. Chapter Five - inflammatory bowel disease and targeted oral anti-TNF α therapy. *Adv Protein Chem Str* 2020;119:157–98.
136. Zhang SF, Langer R, Traverso G. Nanoparticulate drug delivery systems targeting inflammation for treatment of inflammatory bowel disease. *Nano Today* 2017;16:82–96.
137. Lautenschläger C, Schmidt C, Fischer D, Stallmach A. Drug delivery strategies in the therapy of inflammatory bowel disease. *Adv Drug Deliv Rev* 2014;71:58–76.
138. Vass P, Démuth B, Hirsch E, Nagy B, Andersen SK, Vigh T, et al. Drying technology strategies for colon-targeted oral delivery of biopharmaceuticals. *J Control Release* 2019;296:162–78.
139. Li X, Lu C, Yang YY, Yu CH, Rao YF. Site-specific targeted drug delivery systems for the treatment of inflammatory bowel disease. *Biomed Pharmacother* 2020;129:110486.
140. Friend DR. New oral delivery systems for treatment of inflammatory bowel disease. *Adv Drug Deliv Rev* 2005;57:247–65.
141. Zhang YY, Thanou MY, Villasaliu D. Exploiting disease-induced changes for targeted oral delivery of biologics and nanomedicines in inflammatory bowel disease. *Eur J Pharm Biopharm* 2020;155:128–38.
142. Courthion H, Mugnier T, Rousseaux C, Möller M, Gurny R, Gabriel D. Self-assembling polymeric nanocarriers to target inflammatory lesions in ulcerative colitis. *J Control Release* 2018;275:32–9.
143. Xiao B, Chen QB, Zhang Z, Wang LX, Kang YJ, Denning T, et al. TNF α gene silencing mediated by orally targeted nanoparticles combined with interleukin-22 for synergistic combination therapy of ulcerative colitis. *J Control Release* 2018;287:235–46.
144. Nguyen T-HT, Trinh N-T, Tran HN, Tran HT, Le PQ, Ngo D-N, et al. Improving silymarin oral bioavailability using silica-installed redox nanoparticle to suppress inflammatory bowel disease. *J Control Release* 2021;331:515–24.

145. Hua SS, Marks E, Schneider JJ, Keely S. Advances in oral nano-delivery systems for colon targeted drug delivery in inflammatory bowel disease: selective targeting to diseased versus healthy tissue. *Nanomed Nanotechnol Biol Med* 2015;11:1117–32.
146. Laroui H, Dalmasso G, Nguyen HTT, Yan YT, Sitaraman SV, Merlin D. Drug-loaded nanoparticles targeted to the colon with polysaccharide hydrogel reduce colitis in a mouse model. *Gastroenterology* 2010;138: 843-U77.
147. Knipe JM, Strong LE, Peppas NA. Enzyme- and pH-responsive microencapsulated manogels for oral delivery of siRNA to induce TNF-alpha knockdown in the intestine. *Biomacromolecules* 2016;17: 788–97.
148. Xiao B, Xu ZG, Viennois E, Zhang YC, Zhang Z, Zhang MZ, et al. Orally targeted delivery of tripeptide KPV via hyaluronic acid-functionalized nanoparticles efficiently alleviates ulcerative colitis. *Mol Ther* 2017;25:1628–40.
149. Zhang SF, Ermann J, Succi MD, Zhou A, Hamilton MJ, Cao B, et al. An inflammation-targeting hydrogel for local drug delivery in inflammatory bowel disease. *Sci Transl Med* 2015;7. 300ra128.
150. Vong LB, Mo J, Abrahamsson B, Nagasaki Y. Specific accumulation of orally administered redox nanotherapeutics in the inflamed colon reducing inflammation with dose-response efficacy. *J Control Release* 2015;210:19–25.
151. Li CW, Zhao Y, Cheng J, Guo JW, Zhang QX, Zhang XJ, et al. A proresolving peptide nanotherapy for site-specific treatment of inflammatory bowel disease by regulating proinflammatory microenvironment and gut microbiota. *Adv Sci* 2019;6:1900610.
152. Li SS, Xie AQ, Li H, Zou X, Zhang QX. A self-assembled, ROS-responsive janus-prodrug for targeted therapy of inflammatory bowel disease. *J Control Release* 2019;316:66–78.
153. Naeem M, Oshi MA, Kim J, Lee J, Cao JF, Nurhasni H, et al. pH-triggered surface charge-reversal nanoparticles alleviate experimental murine colitis via selective accumulation in inflamed colon regions. *Nanomed Nanotechnol Biol Med* 2018;14:823–34.
154. Xiao B, Laroui H, Ayyadurai S, Viennois E, Charania MA, Zhang YC, et al. Mannosylated bioreducible nanoparticle-mediated macrophage-specific TNF- α RNA interference for IBD therapy. *Biomaterials* 2013;34:7471–82.
155. Zeeshan M, Ali H, Khan S, Khan SA, Weigmann B. Advances in orally-delivered pH-sensitive nanocarrier systems; an optimistic approach for the treatment of inflammatory bowel disease. *Int J Pharm* 2019;558:201–14.
156. Lee Y, Sugihara K, Gilliland MG, Jon S, Kamada N, Moon JJ. Hyaluronic acid-bilirubin nanomedicine for targeted modulation of dysregulated intestinal barrier, microbiome and immune responses in colitis. *Nat Mater* 2020;19:118–26.
157. Vafaei SY, Esmaeili M, Amini M, Atyabi F, Ostad SN, Dinarvand R. Self assembled hyaluronic acid nanoparticles as a potential carrier for targeting the inflamed intestinal mucosa. *Carbohydr Polym* 2016;144:371–81.
158. Sun Y, Duan BC, Chen HH, Xu XJ. A novel strategy for treating inflammatory bowel disease by targeting delivery of methotrexate through glucan particles. *Adv Healthc Mater* 2020;9:1901805.
159. Zhang MZ, Xu CL, Liu DD, Han MK, Wang LX, Merlin D. Oral delivery of nanoparticles loaded with ginger active compound, 6-shogaol, attenuates ulcerative colitis and promotes wound healing in a murine model of ulcerative colitis. *J Crohns Colitis* 2018;12: 217–29.
160. Beloqui A, Coco R, Memvanga PB, Ucakar B, des Rieux A, Prétat V. pH-sensitive nanoparticles for colonic delivery of curcumin in inflammatory bowel disease. *Int J Pharm* 2014;473:203–12.
161. Singh AK, Cabral C, Kumar R, Ganguly R, Rana HK, Gupta A, et al. Beneficial effects of dietary polyphenols on gut microbiota and strategies to improve delivery efficiency. *Nutrients* 2019;11:2216.
162. Wang XY, Yan JJ, Wang LZ, Pan DH, Yang RL, Xu YP, et al. Rational design of polyphenol-poloxamer nanovesicles for targeting inflammatory bowel disease therapy. *Chem Mater* 2018;30:4073–80.
163. Kesharwani SS, Ahmad R, Bakkari MA, Rajput MKS, Dachineni R, Valiveti CK, et al. Site-directed non-covalent polymer-drug complexes for inflammatory bowel disease (IBD): formulation development, characterization and pharmacological evaluation. *J Control Release* 2018;290:165–79.
164. Huang Z, Gan JJ, Jia LX, Guo GX, Wang CM, Zang YH, et al. An orally administrated nucleotide-delivery vehicle targeting colonic macrophages for the treatment of inflammatory bowel disease. *Biomaterials* 2015;48:26–36.
165. Antonino RSCMQ, Nascimento TL, de Oliveira Junior ER, Souza LG, Batista AC, Lima EM. Thermoreversible mucoadhesive polymer-drug dispersion for sustained local delivery of budesonide to treat inflammatory disorders of the GI tract. *J Control Release* 2019; 303:12–23.
166. Brusini R, Varna M, Couvreur P. Advanced nanomedicines for the treatment of inflammatory diseases. *Adv Drug Deliv Rev* 2020;157: 161–78.
167. Teng C, Lin CS, Huang FF, Xing XY, Chen SY, Ye L, et al. Intracellular codelivery of anti-inflammatory drug and anti-miR 155 to treat inflammatory disease. *Acta Pharm Sin B* 2020;10:1521–33.
168. Zhao YG, Yang YT, Zhang JX, Wang R, Cheng BY, Kalambade D, et al. Lactoferrin-mediated macrophage targeting delivery and patchouli alcohol-based therapeutic strategy for inflammatory bowel diseases. *Acta Pharm Sin B* 2020;10:1966–76.
169. Cai ZJ, Zhang W, Yang F, Yu L, Yu Z, Pan JH, et al. Immunosuppressive exosomes from TGF- β 1 gene-modified dendritic cells attenuate Th17-mediated inflammatory autoimmune disease by inducing regulatory T cells. *Cell Res* 2012;22:607–10.
170. Cybulsky MI, Gimbrone MA. Endothelial expression of a mono-nuclear leukocyte adhesion molecule during atherosclerosis. *Science* 1991;251:788–91.
171. Gisterå A, Hansson GK. The immunology of atherosclerosis. *Nat Rev Nephrol* 2017;13:368–80.
172. Lievens D, von Hundelshausen P. Platelets in atherosclerosis. *J Thromb Haemostasis* 2011;106:827–38.
173. Newland SA, Mohanta S, Clément M, Taleb S, Walker JA, Nus M, et al. Type-2 innate lymphoid cells control the development of atherosclerosis in mice. *Nat Commun* 2017;8:1–11.
174. Borén J, Williams KJ. The central role of arterial retention of cholesterol-rich apolipoprotein-B-containing lipoproteins in the pathogenesis of atherosclerosis: a triumph of simplicity. *Curr Opin Lipidol* 2016;27:473–83.
175. Park YM, Febbraio M, Silverstein RL. CD36 modulates migration of mouse and human macrophages in response to oxidized LDL and may contribute to macrophage trapping in the arterial intima. *J Clin Invest* 2009;119:136–45.
176. Van Gils JM, Derby MC, Fernandes LR, Ramkhelawon B, Ray TD, Rayner KJ, et al. The neuroimmune guidance cue netrin-1 promotes atherosclerosis by inhibiting the emigration of macrophages from plaques. *Nat Immunol* 2012;13:136–43.
177. Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol* 2013;13:709–21.
178. Lichtman AH, Binder CJ, Tsimikas S, Witztum JL. Adaptive immunity in atherosclerosis: new insights and therapeutic approaches. *J Clin Invest* 2013;123:27–36.
179. Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol* 2011;12:204.
180. Taleb S. Inflammation in atherosclerosis. *Arch Cardiovasc Dis* 2016; 109:708–15.
181. Tousoulis D, Oikonomou E, Economou EK, Crea F, Kaski JC. Inflammatory cytokines in atherosclerosis: current therapeutic approaches. *Eur Heart J* 2016;37:1723–32.
182. Lüscher TF. Novel mechanisms of atherosclerosis and cardiovascular repair. *Eur Heart J* 2016;37:1709–11.
183. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017;377:1119–31.

184. Ridker PM, MacFadyen JG, Everett BM, Libby P, Thuren T, Glynn RJ, et al. Relationship of C-reactive protein reduction to cardiovascular event reduction following treatment with canakinumab: a secondary analysis from the CANTOS randomised controlled trial. *Lancet* 2018;391:319–28.
185. Ridker PM, Libby P, MacFadyen JG, Thuren T, Ballantyne C, Fonseca F, et al. Modulation of the interleukin-6 signalling pathway and incidence rates of atherosclerotic events and all-cause mortality: analyses from the canakinumab anti-inflammatory thrombosis outcomes study (CANTOS). *Eur Heart J* 2018;39:3499–507.
186. Rouwet E, Lutgens E. 2016 Jeffrey M. Hoog award lecture: immune checkpoints in atherosclerosis: toward immunotherapy for atheroprotection. *Aterio Thromb Vasc Biol* 2018;38:1678–88.
187. Doesch AO, Zhao L, Gleissner CA, Akhavanpoor M, Rohde D, Okuyucu D, et al. Inhibition of B7-1 (CD80) by RhuDex® reduces lipopolysaccharide-mediated inflammation in human atherosclerotic lesions. *Drug Des Dev Ther* 2014;8:447.
188. Meletta R, Herde AM, Dennler P, Fischer E, Schibli R, Krämer SD. Preclinical imaging of the co-stimulatory molecules CD80 and CD86 with indium-111-labeled belatacept in atherosclerosis. *EJNMMI Res* 2016;6:1.
189. Müller A, Mu LJ, Meletta R, Beck K, Rancic Z, Drandarov K, et al. Towards non-invasive imaging of vulnerable atherosclerotic plaques by targeting co-stimulatory molecules. *Int J Cardiol* 2014;174:503–15.
190. Lutgens E, Lievens D, Beckers L, Wijnands E, Soehnlein O, Zernecke A, et al. Deficient CD40-TRAF6 signaling in leukocytes prevents atherosclerosis by skewing the immune response toward an antiinflammatory profile. *J Exp Med* 2010;207:391–404.
191. Schönbeck U, Sukhova G, Shimizu K, Mach F, Libby P. Inhibition of CD40 signaling limits evolution of established atherosclerosis in mice. *P Natl Acad Sci USA* 2000;97:7458–63.
192. Mach F, Schönbeck U, Sukhova GK, Atkinson E, Libby P. Reduction of atherosclerosis in mice by inhibition of CD40 signalling. *Nature* 1998;394:200–3.
193. Lutgens E, Gorelik L, Daemen MJ, de Muinck ED, Grewal IS, Koteliansky VE, et al. Requirement for CD154 in the progression of atherosclerosis. *Nat Med* 1999;5:1313–6.
194. Zernecke A, Weber C. Chemokines in atherosclerosis: proceedings resumed. *Arterioscl Throm Vas* 2014;34:742–50.
195. EPCvd Vorst, Döring Y, Weber C. Chemokines and their receptors in atherosclerosis. *J Mol Med* 2015;93:963–71.
196. Noels H, Weber C, Koenen RR. Chemokines as therapeutic targets in cardiovascular disease. *Arterioscl Throm Vas* 2019;39:583–92.
197. Jones K, Maguire J, Davenport A. Chemokine receptor CCR5: from AIDS to atherosclerosis. *Br J Pharmacol* 2011;162:1453–69.
198. Hamesch K, Subramanian P, Li XF, Dembowsky K, Chevalier E, Weber C, et al. The CXCR4 antagonist POL5551 is equally effective as sirolimus in reducing neointima formation without impairing re-endothelialisation. *J Thromb Haemostasis* 2012;107:356–68.
199. Karshovska E, Zagorac D, Zernecke A, Weber C, Schober A. A small molecule CXCR4 antagonist inhibits neointima formation and smooth muscle progenitor cell mobilization after arterial injury. *J Thromb Haemostasis* 2008;6:1812–5.
200. Lutgens E, Atzler D, Doring Y, Duchene J, Steffens S, Weber C. Immunotherapy for cardiovascular disease. *Eur Heart J* 2019;40:3937–46.
201. Shirai T, Nazarewicz RR, Wallis BB, Yanes RE, Watanabe R, Hilhorst M, et al. The glycolytic enzyme PKM2 bridges metabolic and inflammatory dysfunction in coronary artery disease. *J Exp Med* 2016;213:337–54.
202. Moon JS, Hisata S, Park MA, DeNicola GM, Ryter SW, Nakahira K, et al. mTORC1-induced HK1-dependent glycolysis regulates NLRP3 inflammasome activation. *Cell Rep* 2015;12:102–15.
203. Vats D, Mukundan L, Odegaard JI, Zhang L, Smith KL, Morel CR, et al. Oxidative metabolism and PGC-1 β attenuate macrophage-mediated inflammation. *Cell Metabol* 2006;4:13–24.
204. Michalek RD, Gerriets VA, Jacobs SR, Macintyre AN, MacIver NJ, Mason EF, et al. Cutting edge: distinct glycolytic and lipid oxidative metabolic programs are essential for effector and regulatory CD4 $^{+}$ T cell subsets. *J Immunol* 2011;186:3299–303.
205. GJvd Windt, Everts B, Chang C-H, Curtis JD, Freitas TC, Amiel E, et al. Mitochondrial respiratory capacity is a critical regulator of CD8 $^{+}$ T cell memory development. *Immunity* 2012;36:68–78.
206. Ren B, Van Kampen E, Van Berkel TJ, Cruickshank SM, Van Eck M. Hematopoietic arginase 1 deficiency results in decreased leukocytosis and increased foam cell formation but does not affect atherosclerosis. *Atherosclerosis* 2017;256:35–46.
207. Cole JE, Astola N, Cribbs AP, Goddard ME, Park I, Green P, et al. Indoleamine 2, 3-dioxygenase-1 is protective in atherosclerosis and its metabolites provide new opportunities for drug development. *P Natl Acad Sci Usa* 2015;112:13033–8.
208. Peterson AS, Fong LG, Young SG. Errata. PCSK9 function and physiology. *J Lipid Res* 2008;49:1595–9.
209. Grooth GJd, Klerkx AH, Stroes ES, Stalenhoef AF, Kastelein JJ, Kuivenhoven JA. A review of CETP and its relation to atherosclerosis. *J Lipid Res* 2004;45:1967–74.
210. Koren MJ, Lundqvist P, Bolognesi M, Neutel JM, Monsalvo ML, Yang JY, et al. Anti-PCSK9 monotherapy for hypercholesterolemia: the MENDEL-2 randomized, controlled phase III clinical trial of evolocumab. *J Am Coll Cardiol* 2014;63:2531–40.
211. Crossey E, Amar MJ, Sampson M, Peabody J, Schiller JT, Chackerian B, et al. A cholesterol-lowering VLP vaccine that targets PCSK9. *Vaccine* 2015;33:5747–55.
212. Rittershaus CW, Miller DP, Thomas LJ, Picard MD, Honan CM, Emmett CD, et al. Vaccine-induced antibodies inhibit CETP activity *in vivo* and reduce aortic lesions in a rabbit model of atherosclerosis. *Arterioscl Throm Vas* 2000;20:2106–12.
213. Ait-Oufella H, Salomon BL, Potteaux S, Robertson A-KL, Gourdy P, Zoll J, et al. Natural regulatory T cells control the development of atherosclerosis in mice. *Nat Med* 2006;12:178–80.
214. Tv Es, GHv Puijvelde, Foks A, Habets K, Bot I, Gilboa E, et al. Vaccination against Foxp3 $^{+}$ regulatory T cells aggravates atherosclerosis. *Atherosclerosis* 2010;209:74–80.
215. Ou HX, Guo BB, Liu Q, Li YK, Yang Z, Feng WJ, et al. Regulatory T cells as a new therapeutic target for atherosclerosis. *Acta Pharmacol Sin* 2018;39:1249–58.
216. Beldman TJ, Malinova TS, Desclos E, Grootemaat AE, Misiak ALS, van der Velden S, et al. Nanoparticle-aided characterization of arterial endothelial architecture during atherosclerosis progression and metabolic therapy. *ACS Nano* 2019;13:13759–74.
217. Momtazi-Borjomi AA, Jaafari MR, Badiee A, Banach M, Sahebkar A. Therapeutic effect of nanoliposomal PCSK9 vaccine in a mouse model of atherosclerosis. *BMC Med* 2019;17:223.
218. Kiae N, Gorabi AM, Penson PE, Watts G, Johnston TP, Banach M, et al. A new approach to the diagnosis and treatment of atherosclerosis: the era of the liposome. *Drug Discov Today* 2020;25:58–72.
219. Seijkens TTP, van Tiel CM, Kusters PJH, Atzler D, Soehnlein O, Zarzycka B, et al. Targeting CD40-induced TRAF6 signaling in macrophages reduces atherosclerosis. *J Am Coll Cardiol* 2018;71:527–42.
220. Peters EB, Tsiliis ND, Karver MR, Chin SM, Musetti B, Ledford BT, et al. Atheroma niche-responsive nanocarriers for immunotherapeutic delivery. *Adv Healthc Mater* 2019;8:1801545.
221. Song YN, Huang ZY, Liu X, Pang ZQ, Chen J, Yang HB, et al. Platelet membrane-coated nanoparticle-mediated targeting delivery of rapamycin blocks atherosclerotic plaque development and stabilizes plaque in apolipoprotein E-deficient (ApoE $^{-/-}$) mice. *Nanomed-Nanotechnol* 2019;15:13–24.
222. Yi SJ, Allen SD, Liu YG, Ouyang BZ, Li X, Augsornworawat P, et al. Tailoring nanostructure morphology for enhanced targeting of dendritic cells in atherosclerosis. *ACS Nano* 2016;10:11290–303.
223. Schönbeck U, Libby P. CD40 signaling and plaque instability. *Circ Res* 2001;89:1092–103.

224. Lameijer M, Binderup T, van Leent MMT, Senders ML, Fay F, Malkus J, et al. Efficacy and safety assessment of a TRAF6-targeted nanoimmunotherapy in atherosclerotic mice and non-human primates. *Nat Biomed Eng* 2018;2:279–92.
225. Ye ZS, Zhong L, Zhu SN, Wang YN, Zheng J, Wang SJ, et al. The P-selectin and PSGL-1 axis accelerates atherosclerosis via activation of dendritic cells by the TLR4 signaling pathway. *Cell Death Dis* 2019; 10:1–15.
226. Subramanian M, Thorp E, Hansson GK, Tabas I. Treg-mediated suppression of atherosclerosis requires MYD88 signaling in DCs. *J Clin Invest* 2013;123:179–88.
227. Foks AC, Lichtman AH, Kuiper J. Treating atherosclerosis with regulatory T cells. *Aterio Thromb Vasc Biol* 2015;35:280–7.
228. Paulson KE, Zhu SN, Chen M, Nurmohamed S, Jongstra-Bilen J, Cybulsky MI. Resident intimal dendritic cells accumulate lipid and contribute to the initiation of atherosclerosis. *Circ Res* 2010;106: 383–90.
229. Weber C, Meiler S, Döring Y, Koch M, Drechsler M, Megens RT, et al. CCL17-expressing dendritic cells drive atherosclerosis by restraining regulatory T cell homeostasis in mice. *J Clin Invest* 2011; 121:2898–910.
230. Niessner A, Weyand CM. Dendritic cells in atherosclerotic disease. *Clin Immunol* 2010;134:25–32.
231. Yi SJ, Zhang XH, Sangji MH, Liu YG, Allen SD, Xiao BX, et al. Surface engineered polymersomes for enhanced modulation of dendritic cells during cardiovascular immunotherapy. *Adv Funct Mater* 2019;29:1904399.
232. Benne N, van Duijn J, Vigario FL, Leboux RJ, van Veelen P, Kuiper J, et al. Anionic 1, 2-distearoyl-sn-glycero-3-phosphoglycerol (DSPG) liposomes induce antigen-specific regulatory T cells and prevent atherosclerosis in mice. *J Control Release* 2018;291:135–46.
233. Li CW, Dou Y, Chen YD, Qi YT, Li LL, Han SL, et al. Site-specific microRNA-33 antagonism by pH-responsive nanotherapies for treatment of atherosclerosis via regulating cholesterol efflux and adaptive immunity. *Adv Funct Mater* 2020;2002131.
234. Bermúdez V, Rojas-Quintero J, Velasco M. The quest for immunotherapy in atherosclerosis: CANTOS study, interleukin-1 β and vascular inflammation. *J Thorac Dis* 2017;10:64–9.
235. Yin LQ, Peng CP, Tang Y, Yuan YC, Liu JX, Xiang TT, et al. Biomimetic oral targeted delivery of bindarit for immunotherapy of atherosclerosis. *Biomater Sci* 2020;8:3640–8.
236. Deshpande V, Krishnan R, Philip S, Faludi I, Ponnusamy T, Thota LNR, et al. Oral administration of recombinant mycobacterium smegmatis expressing a tripeptide construct derived from endogenous and microbial antigens prevents atherosclerosis in ApoE $^{-/-}$ mice. *Cardiovasc Ther* 2016;34:314–24.
237. Thota LN, Ponnusamy T, Lu X, Mundkur L. Long-term efficacy and safety of immunomodulatory therapy for atherosclerosis. *Cardiovasc Drugs Ther* 2019;33:385–98.
238. Sato-Okabayashi Y, Isoda K, Heissig B, Kadoguchi T, Akita K, Kitamura K, et al. Low-dose oral cyclophosphamide therapy reduces atherosclerosis progression by decreasing inflammatory cells in a murine model of atherosclerosis. *IJC Heart & Vasculature* 2020;28: 100529.
239. Beltrán-López JI, Romero-Maldonado A, Monreal-Escalante E, Bañuelos-Hernández B, Paz-Maldonado LM, Rosales-Mendoza S. Chlamydomonas reinhardtii chloroplasts express an orally immunogenic protein targeting the p210 epitope implicated in atherosclerosis immunotherapies. *Plant Cell Rep* 2016;35:1133–41.
240. Arevalo-Villalobos JI, Alonso DOG, Rosales-Mendoza S. Using carrot cells as biofactories and oral delivery vehicles of LTB-Syn: a low-cost vaccine candidate against synucleinopathies. *J Biotechnol* 2020;309:75–80.
241. Schermuly RT, Ghofrani HA, Wilkins MR, Grimminger F. Mechanisms of disease: pulmonary arterial hypertension. *Nat Rev Cardiol* 2011;8:443.
242. Li C, Liu PP, Song R, Zhang YQ, Lei S, Wu SJ. Immune cells and autoantibodies in pulmonary arterial hypertension. *Acta Biochim Biophys Sin* 2017;49:1047–57.
243. Chu YB, XiangLi XY, Xiao W. Regulatory T cells protect against hypoxia-induced pulmonary arterial hypertension in mice. *Mol Med Rep* 2015;11:3181–7.
244. Zhu R, Chen L, Xiong YQ, Wang NN, Xie XC, Hong YQ, et al. An upregulation of CD8 $^{+}$ CD25 $^{+}$ Foxp3 $^{+}$ T cells with suppressive function through interleukin 2 pathway in pulmonary arterial hypertension. *Exp Cell Res* 2017;358:182–7.
245. Qiu HH, He Y, Ouyang F, Jiang P, Guo SH, Guo Y. The role of regulatory T Cells in pulmonary arterial hypertension. *J Am Heart Assoc* 2019;8:e014201.
246. Yang K, Blanco DB, Neale G, Vogel P, Avila J, Clish CB, et al. Homeostatic control of metabolic and functional fitness of Treg cells by LKB1 signalling. *Nature* 2017;548:602–6.
247. Whitehouse G, Gray E, Mastoridis S, Merritt E, Kodela E, Yang JH, et al. IL-2 therapy restores regulatory T-cell dysfunction induced by calcineurin inhibitors. *P Natl Acad Sci Usa* 2017;114:7083–8.
248. Wang H, Hou L, Kwak D, Fassett J, Xu X, Chen A, et al. Increasing regulatory T cells with interleukin-2 and interleukin-2 antibody complexes attenuates lung inflammation and heart failure progression. *Hypertension* 2016;68:114–22.
249. Marinho A, Carvalho C, Boleixa D, Bettencourt A, Leal B, Guimarães J, et al. Vitamin D supplementation effects on FoxP3 expression in T cells and FoxP3 $^{+}$ /IL-17A ratio and clinical course in systemic lupus erythematosus patients: a study in a Portuguese cohort. *Immunol Res* 2017;65:197–206.
250. Tabares P, Berr S, Römer PS, Chuvpilo S, Matskevich AA, Tyrsin D, et al. Human regulatory T cells are selectively activated by low-dose application of the CD28 superagonist TGN1412/TAB08. *Eur J Immunol* 2014;44:1225–36.
251. Ferreira LM, Muller YD, Bluestone JA, Tang Q. Next-generation regulatory T cell therapy. *Nat Rev Drug Discov* 2019;18:749–69.
252. Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, et al. Type 1 diabetes immunotherapy using polyclonal regulatory T cells. *Sci Transl Med* 2015;7:315ra18.
253. Dall'Era M, Pauli ML, Remedios K, Taravati K, Sandova PM, Putnam AL, et al. Adoptive Treg cell therapy in a patient with systemic lupus erythematosus. *Arthritis Rheum* 2019;71:431–40.
254. Zamanian R, Badesch D, Chung L, Domsic R, Medsger T, Pinckney A, et al. Late Breaking Abstract-Safety and efficacy of B-cell depletion with rituximab for the treatment of systemic sclerosis-associated pulmonary arterial hypertension. *Eur Respir J* 2019;54: RCT1884.
255. Bryant AJ, Fu Ch, Lu Y, Brantly ML, Mehrad B, Moldawer LL, et al. A checkpoint on innate myeloid cells in pulmonary arterial hypertension. *Palm Circ* 2018;9: 2045894018823528.
256. Nicolls MR, Voelkel NF. The roles of immunity in the prevention and evolution of pulmonary arterial hypertension. A perspective. *Am J Respir Crit Care Med* 2017;195:1292–9.
257. Humbert M, Monti G, Brenot F, Sitbon O, Portier A, Grangeot-Keros L, et al. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary hypertension. *Am J Respir Crit Care Med* 1995;151:1628–31.
258. Kim KS, Jung H, Shin IK, Choi BR, Kim DH. Induction of interleukin-1 beta (IL-1 β) is a critical component of lung inflammation during influenza A (H1N1) virus infection. *J Med Virol* 2015;87: 1104–12.
259. Groth A, Vrugt B, Brock M, Speich R, Ulrich S, Huber LC. Inflammatory cytokines in pulmonary hypertension. *Respir Res* 2014; 15:47.
260. Hernández-Sánchez J, Harlow L, Church C, Gaine S, Knightbridge E, Bunclark K, et al. Clinical trial protocol for TRANSFORM-UK: a therapeutic open-label study of tocilizumab in the treatment of pulmonary arterial hypertension. *Respir Res* 2017;8:2045893217735820.

261. Wang Q, Zuo XR, Wang YY, Xie WP, Wang H, Zhang MJ. Monocrotaline-induced pulmonary arterial hypertension is attenuated by TNF- α antagonists via the suppression of TNF- α expression and NF- κ B pathway in rats. *Vasc Pharmacol* 2013;58:71–7.
262. Nakaoka Y, Inagaki T, Shirai M. *Inflammatory cytokines in the pathogenesis of pulmonary arterial hypertension*. Singapore: Springer Singapore; 2020.
263. Mamazhakypov A, Viswanathan G, Lawrie A, Schermuly RT, Rajagopal S. The role of chemokines and chemokine receptors in pulmonary arterial hypertension. *Br J Pharmacol* 2019;195:1–18.
264. Tian W, Jiang XG, Tamoxiumiene R, Sung YK, Qian J, Dhillon G, et al. Blocking macrophage leukotriene B4 prevents endothelial injury and reverses pulmonary hypertension. *Sci Transl Med* 2013;5:200ra117.
265. Li SJ, Zhai C, Shi WH, Feng W, Xie XM, Pan YL, et al. Leukotriene B4 induces proliferation of rat pulmonary arterial smooth muscle cells via modulating GSK-3 β /catenin pathway. *Eur J Pharmacol* 2020;867:172823.
266. Galí N, Manes A, Branzi A. The endothelin system in pulmonary arterial hypertension. *Cardiovasc Res* 2004;61:227–37.
267. Dai Y, Chen X, Song XX, Chen XJ, Ma WR, Lin JB, et al. Immunotherapy of endothelin-1 receptor type a for pulmonary arterial hypertension. *J Am Coll Cardiol* 2019;73:2567–80.
268. Li C, Yan XL, Wu DY, Zhang K, Liang X, Pan YJ, et al. Vaccine-targeted alpha 1D-adrenergic receptor for hypertension. *Hypertension* 2019;74:1551–62.
269. Lee Y, Pai SB, Bellamkonda RV, Thompson DH, Singh J. Cerivastatin nanoliposome as a potential disease-modifying approach for the treatment of pulmonary arterial hypertension. *J Pharmacol Exp Therapeut* 2018;366:66–74.
270. Dhoble S, Patravale V. Development of anti-angiogenic erlotinib liposomal formulation for pulmonary hypertension: a QbD approach. *Drug Deliv Transl Re* 2019;9:980–96.
271. Kimura S, Egashira K, Chen L, Nakano K, Iwata E, Miyagawa M, et al. Nanoparticle-mediated delivery of nuclear factor B decoy into lungs ameliorates monocrotaline-induced pulmonary arterial hypertension. *Hypertension* 2009;53:877–83.
272. Chen L, Nakano K, Kimura S, Matoba T, Iwata E, Miyagawa M, et al. Nanoparticle-mediated delivery of pitavastatin into lungs ameliorates the development and induces regression of monocrotaline-induced pulmonary artery hypertension. *Hypertension* 2011;57:343–50.
273. Emami F, Yazdi SJM, Na DH. Poly(lactic acid)/poly(lactic-co-glycolic acid) particulate carriers for pulmonary drug delivery. *J Pharm Sci* 2019;49:427–42.
274. Lee W-H, Loo C-Y, Traini D, Young PM. Inhalation of nanoparticle-based drug for lung cancer treatment: advantages and challenges. *Asian J Pharm Sci* 2015;10:481–9.
275. Long L, Ormiston ML, Yang XD, Southwood M, Gräf S, Machado RD, et al. Selective enhancement of endothelial BMPR-II with BMP9 reverses pulmonary arterial hypertension. *Nat Med* 2015;21:777–85.
276. Spiekerkoetter E, Tian XF, Cai J, Hopper RK, Sudheendra D, Li CG, et al. FK506 activates BMPR2, rescues endothelial dysfunction, and reverses pulmonary hypertension. *J Clin Invest* 2013;123:3600–13.
277. Spiekerkoetter E, Sung YK, Sudheendra D, Bill M, Aldred MA, van de Veerdonk MC, et al. Low-dose FK506 (tacrolimus) in end-stage pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2015;192:254–7.
278. Spiekerkoetter E, Sung YK, Sudheendra D, Scott V, Del Rosario P, Bill M, et al. Randomised placebo-controlled safety and tolerability trial of FK506 (tacrolimus) for pulmonary arterial hypertension. *Eur Respir J* 2017;50:1602449.
279. Wang ZM, Cuddigan JL, Gupta SK, Meenach SA. Nanocomposite microparticles (nCmP) for the delivery of tacrolimus in the treatment of pulmonary arterial hypertension. *Int J Pharm* 2016;512:305–13.
280. Yang YC, Lin F, Xiao ZQ, Sun B, Wei ZY, Liu BY, et al. Investigational pharmacotherapy and immunotherapy of pulmonary arterial hypertension: an update. *Biomed Pharmacother* 2020;129:110355.
281. Pauken KE, Dougan M, Rose NR, Lichtman AH, Sharpe AH. Adverse events following cancer immunotherapy: obstacles and opportunities. *Trends Immunol* 2019;40:511–23.
282. Jiang XT, Xu J, Liu MF, Xing H, Wang ZM, Huang L, et al. Adoptive CD8 $^{+}$ T cell therapy against cancer: challenges and opportunities. *Cancer Lett* 2019;462:23–32.
283. Qi JP, Hu XW, Dong XC, Lu Y, Lu HP, Zhao WL, et al. Towards more accurate bioimaging of drug nanocarriers: turning aggregation-caused quenching into a useful tool. *Adv Drug Deliv Rev* 2019;143:206–25.
284. Anselmo AC, Gokarn Y, Mitragotri S. Non-invasive delivery strategies for biologics. *Nat Rev Drug Discov* 2019;18:19–40.