

Serum Neurofilament Levels and PML Risk in Patients With Multiple Sclerosis Treated With Natalizumab

Nicolás Fissolo, PhD, Beatrice Pigolet, PhD, Jordi Rio, MD, PhD, Patrick Vermersch, MD, PhD, Aurélie Ruet, MD, PhD, Jerome deSèze, MD, PhD, Pierre Labauge, MD, PhD, Sandra Vukusic, MD, PhD, Caroline Papeix, MD, Laurent Martinez-Almoyna, MD, Ayman Tourbah, MD, PhD, Pierre Clavelou, MD, PhD, Thibault Moreau, MD, PhD, Jean Pelletier, MD, PhD, Christine Lebrun-Frenay, MD, PhD, Bertrand Bourre, MD, Gilles Defer, MD, PhD, Xavier Montalban, MD, David Brassat, MD, PhD, and Manuel Comabella, MD, PhD

Correspondence: Dr. Fissolo
nicolas.fissolo@vhir.org

Neurol Neuroimmunol Neuroinflamm 2021;8:e1003. doi:10.1212/NXI.0000000000001003

Abstract

Objectives

The study aimed to assess the potential for serum neurofilament light chain (NFL) levels to predict the risk of progressive multifocal leukoencephalopathy (PML) in natalizumab (NTZ)-treated patients with multiple sclerosis (MS) and to discriminate PML from MS relapses.

Methods

NFL levels were measured with single molecule array (Simoa) in 4 cohorts: (1) a prospective cohort of patients with MS who developed PML under NTZ therapy (pre-PML) and non-PML NTZ-treated patients (NTZ-ctr); (2) a cohort of patients whose blood was collected during PML; (3) an independent cohort of non-PML NTZ-treated patients with serum NFL determinations at 2 years (replication cohort); and (4) a cohort of patients whose blood was collected during exacerbations.

Results

Serum NFL levels were significantly increased after 2 years of NTZ treatment in pre-PML patients compared with NTZ-ctr. The prognostic performance of serum NFL levels to predict PML development at 2 years was similar in the NTZ-ctr group and replication cohort. Serum NFL levels also distinguished PML from MS relapses and were 8-fold higher during PML compared with relapses.

Conclusions

These results support the use of serum NFL levels in clinical practice to identify patients with relapsing-remitting MS at higher PML risk and to differentiate PML from clinical relapses in NTZ-treated patients.

Classification of Evidence

This study provides Class I evidence that serum NFL levels can identify NTZ-treated patients with MS who will develop PML with a sensitivity of 67% and specificity of 80%.

MORE ONLINE

→ Class of Evidence

Criteria for rating therapeutic and diagnostic studies

[NPub.org/coe](https://www.npub.org/coe)

From the Servei de Neurologia-Neuroimmunologia (N.F., M.C.), Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Institut de Recerca Vall d'Hebron (VHIR), Hospital Universitari Vall d'Hebron, Universitat Autònoma de Barcelona, Spain; CRC-SEP Neurosciences Centre Hospitalier Universitaire Toulouse (B.P., D.B.), CPTP INSERM UMR 1043 CNRS UMR 5282 et Université de Toulouse III, UPS, France; Servei de Neurologia-Neuroimmunologia (J.R., X.M.), Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Hospital Universitari Vall d'Hebron, Barcelona, Spain; Univ. Lille (P.V.), Inserm U1172, CHU Lille, FHU Imminent, France; Université (A.R.), Bordeaux; CHU de Bordeaux (A.R.), INSERM-CHU CIC-P 0005, & Services de Neurologie; Neurocentre Magendie (A.R.), INSERM U1215; Department of Neurology (J.dS), Hôpital Civil, Strasbourg; Department of Neurology (P.L.), CHU Montpellier; Department of Neurology CHU Lyon (S.V.); Department of Neurology (C.P.), Hôpital de la Salpêtrière, Paris; Chi Aix en Provence (L.M.-A.); Department of Neurology and Faculté de Médecine de Reims (A.T.), CHU de Reims, URCA; LPN EA2027 Université Paris VIII (A.T.), Saint-Denis; Department of Neurology (P.C.), CHRU Clermont Ferrand; Department of Neurology (T.M.), CHU Dijon; Aix-Marseille Univ (J.P.), APHM, Hôpital de la Timone, Pôle de Neurosciences Cliniques, Service de Neurologie, CNRS, CRMBM UMR 7339, Marseille; Service de Neurologie (C.L.-F.), CHU de Nice Pasteur2, Université Nice Côte d'Azur UR2CA URRIS, Nice; Neurologie (B.B.), CHU Rouen; and Neurologie (G.D.), CHU Caen, France.

Go to [Neurology.org/NN](https://www.neurology.org/NN) for full disclosures. Funding information is provided at the end of the article.

The Article Processing Charge was funded by the authors.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND), which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Glossary

AUC = area under the ROC curve; **MS** = multiple sclerosis; **NFL** = neurofilament light chain; **NTZ** = natalizumab; **PML** = progressive multifocal leukoencephalopathy; **RRMS** = relapsing-remitting multiple sclerosis.

Despite its proved efficacy for patients with highly active relapsing-remitting multiple sclerosis (RRMS),¹ the use of natalizumab (NTZ) is limited due to the increased risk of progressive multifocal leukoencephalopathy (PML).² In addition to existing PML risk stratification algorithms based on NTZ treatment duration, previous immunosuppressive therapies, and JC virus index,^{2,3} other biomarkers may certainly contribute to estimate the risk of PML at an individual level.

Numerous studies suggest that the concentration of neurofilament light chain (NFL) in peripheral blood and CSF is a promising biomarker in MS.^{4,5} In a recent study, the serum NFL levels measured with an electrochemiluminescence assay were proposed as a biomarker for early identification of PML in patients with MS under NTZ treatment.⁶ In the present study, we aimed to expand on the potential for serum NFL levels measured with single molecule array (Simoa) to predict the risk of PML in a prospective cohort of NTZ-treated patients. We also aimed to discriminate PML from MS relapses based on serum NFL levels.

Methods

Patient Cohorts

Four different cohorts of patients with RRMS were included in the study:

1. A multicentric prospective cohort of patients treated with NTZ (BIONAT cohort; ClinicalTrials.gov identifier: NCT00942214)⁷ was used to evaluate the association between serum NFL levels at baseline and at 1 and 2 years of NTZ treatment and PML development. Patients belonging to this cohort were classified into 2 groups: patients who did not develop PML after a follow-up longer than 5 years (NTZ controls; NTZ-ctr) and patients who developed PML (pre-PML) under NTZ treatment.
2. A cohort of patients whose blood was drawn during the PML condition was included for comparison of NFL levels with the pre-PML group after 2 years of treatment (during PML cohort).
3. An independent cohort of patients treated with NTZ who did not develop PML after more than 5 years of follow-up was included to assess the reproducibility between centers of serum NFL measurements after 2 years of NTZ treatment (replication cohort).
4. A cohort of patients whose blood was collected at the time of an acute relapse was included to investigate the potential for serum NFL levels to discriminate between the PML condition and MS relapses (relapsing cohort). Twenty-seven percent of these patients were receiving treatment with interferon-beta at the time of exacerbations.

The table summarizes the main demographic and baseline clinical characteristics of patients included in the study.

Standard Protocol Approvals, Registrations, and Patient Consents

Written informed consent was obtained from each participant. The study was approved by the local hospital ethics committees, BIONAT cohort; ClinicalTrials.gov identifier: NCT00942214, and Vall d'Hebron Hospital (EPA(AG)57/2013(3834)).

Quantification of Serum NFL Levels

Peripheral blood was collected by standard venipuncture and allowed to clot spontaneously for 30 minutes. Serum was obtained by centrifugation and stored frozen at -80°C until used. Levels of NFL were measured in serum samples using commercially available NFL immunoassay kits (Quanterix, cat#103186) run on the fully automated ultrasensitive Simoa HD-1 Analyzer (Quanterix). Samples were run in duplicate in accordance with manufacturers' instructions with appropriate standards and internal controls. The intra-assay and interassay coefficients of variation were 5% and 9%, respectively.

Classification of Evidence

Our primary research question was to ascertain whether serum NFL levels can identify NTZ-treated patients with MS who will develop PML. The classification of evidence assigned to this question is Class I.

Statistical Analyses

Statistical analysis was performed by using the IBM SPSS Statistics version 22. The distribution of serum NFL levels was tested for normality with a Kolmogorov-Smirnov test. Afterward, paired and unpaired nonparametric tests were applied for comparisons of mean NFL levels among groups. When needed, analysis was adjusted by age and disease duration. Quantitative data are presented as mean values \pm SD unless otherwise stated. Differences were considered statistically significant when p values were below 0.05. Receiver operating characteristic (ROC) curve analyses were used to determine the best cutoff values based on serum NFL levels and the respective sensitivities and specificities.

Data Availability

All data analyzed during this study will be shared anonymized by request of a qualified investigator to the corresponding author.

Results

Serum NFL Levels Are Elevated After 2 Years of NTZ Treatment in Pre-PML Patients and During PML

At baseline, no significant differences were observed in serum NFL levels between pre-PML and NTZ-ctr patients. At 1 and

Table Demographic and Baseline Clinical Characteristics of Patients With RRMS Treated With NTZ

Baseline characteristics	BIONAT cohort		During PML cohort ^a	Replication cohort ^b	Relapsing cohort ^c
	NTZ-ctr ^d	Pre-PML ^e			
n	37	17	13	29	30
Age (y)	36.2 (8.1)	38.6 (8.0)	37.6 (4.0)	37.1 (11.0)	32.3 (9.5)
Female/male (% women)	29/8 (78.4)	13/4 (76.5)	10/3 (76.9)	19/10 (65.5)	23/7 (76.7)
Duration of disease (y)	9.5 (6.4)	11.4 (6.1)	11.4 (6.4)	9.0 (6.6)	7.2 (6.2)
EDSS score at baseline^f	3.0 (2.0–4.2)	3.7 (2.6–5.4)	4.2 (1.6–5.4)	4.0 (2.5–5.2)	3.0 (2.4–3.6) ^{g,h}
JCV status (+/–/unknown) at baseline	23/14/0	13/0/4	4/0/9	2/4/23	—
Duration of NTZ treatment/time to PML (y)ⁱ	6.1 (1.6)	3.7 (1.4)	3.2 (0.7)	7.1 (3.7)	—
Serum NFL levels (pg/mL)^f	8.9 (6.4–18.4)	16.0 (10.2–21.5)	171.1 (130.0–231.0)	6.2 (5.0–8.3)	13.9 (7.6–21.8)

Abbreviations: EDSS = Expanded Disability Status Scale; JCV = JC virus; NFL = neurofilament light chain; NTZ = natalizumab; PML = progressive multifocal leukoencephalopathy; RRMS = relapsing-remitting multiple sclerosis.

Data are expressed as mean (SD) unless otherwise stated.

^a During PML: refers to patients whose blood was collected during the PML condition (4 patients in this group are shared with the pre-PML group). In this group, information on the time between PML onset and blood collection was not available in for 4 patients.

^b Replication cohort: refers to an independent cohort of patients treated with NTZ who did not develop PML, and serum NFL levels were measured after 2 years of treatment.

^c Relapsing cohort: refers to a cohort of patients with MS whose blood was collected at the time of a clinical relapse. In this group, 8 patients (26.7%) were receiving treatment with interferon-beta during relapses, but none of the patients were treated with corticosteroids before blood extraction.

^d NTZ-ctr: refers to patients with MS treated with NTZ who did not develop PML.

^e pre-PML: refers to patients with MS treated with NTZ who developed PML.

^f Data are expressed as median (interquartile range).

^g Refers to EDSS score during relapse.

^h Information on EDSS score was not available in 10 patients. Duration of disease is calculated from disease onset to baseline. Age, percentage of women, duration of disease, and EDSS score at baseline did not statistically significantly differ between the NTZ-ctr group, pre-PML group, during PML cohort, and replication cohort. A trend for older age was observed in the during PML cohort compared with the relapsing cohort ($p = 0.07$). Percentage of women, duration of disease, and EDSS score were not significantly different between these 2 cohorts of patients.

ⁱ Refers to mean time of NTZ treatment in the NTZ-ctr group and replication cohort and time to conversion to PML in the pre-PML group and during PML cohort.

2 years, serum NFL levels were significantly reduced by the effect of NTZ treatment both in the pre-PML and NTZ-ctr groups (figure 1). Comparisons of serum NFL levels at 1 and 2 years between pre-PML and NTZ-ctr patients revealed higher NFL levels only in those patients with MS who will develop PML after 2 years of NTZ treatment with mean values of 10.1 ± 5.9 pg/mL and 7.1 ± 2.5 pg/mL, respectively ($p = 0.03$ both unadjusted and after adjusting for age and disease duration; figure 1).

In the during PML cohort, blood was collected at a mean time from PML onset of 8.1 ± 16.3 days. Serum NFL levels during PML were higher compared with pre-PML patients after 2 years of NTZ treatment with mean values of 163.6 ± 153.8 pg/mL and 10.1 ± 5.9 pg/mL, respectively (unadjusted, $p = 7 \times 10^{-5}$; after adjusting for age and disease duration, $p = 0.02$; figure 1), which represents a 16-fold increase in serum NFL levels.

Serum NFL Levels Discriminate Between Pre-PML and NTZ-ctr Patients After 2 Years of NTZ Treatment

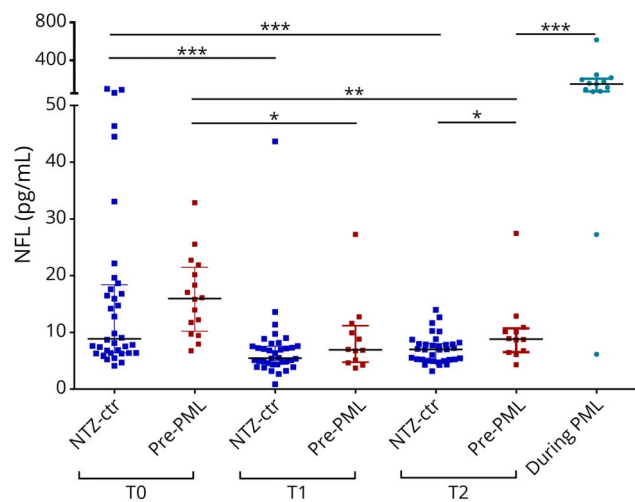
Figure 2A shows the prognostic performance of serum NFL levels to predict PML development at 2 years. The area under the ROC curve (AUC) was 71% ($p = 0.03$), and a serum NFL value of 8.4 pg/mL resulted in the best cutoff to classify pre-PML and NTZ-ctr patients after 2 years of NTZ treatment, with a sensitivity of 67% and specificity of 80%.

As shown in figure 2B, the distribution of serum NFL values in the replication cohort was similar to the NTZ-ctr group at 2 years of treatment, with mean NFL values of 6.9 ± 2.5 pg/mL and 7.1 ± 2.5 pg/mL, respectively. Performance of NFL levels in the replication cohort to predict PML was comparable to the NTZ-ctr cohort at 2 years, with an AUC of 71% ($p = 0.03$), and a serum NFL level of 8.1 pg/mL as the best cutoff to classify pre-PML patients at 2 years and patients from the replication cohort, with a sensitivity and specificity of 67% and 76%, respectively (figure 2C).

Serum NFL Levels Distinguish PML From MS Relapses

Serum NFL levels in the relapsing cohort did not significantly differ between untreated and interferon-beta-treated patients, and hence, this cohort was first analyzed as a whole. Comparison of serum NFL levels between PML and MS relapses revealed significantly higher NFL levels in patients during the PML condition ($p = 3 \times 10^{-6}$; figure 3A), which represents a 8-fold increase in serum NFL levels compared with the relapsing cohort (mean levels: 163.6 ± 153.8 pg/mL vs 20.8 ± 28.0 pg/mL, respectively). Performance of serum NFL levels to differentiate between PML and MS relapses showed an AUC of 91% ($p = 2 \times 10^{-5}$), with an NFL value of 52.7 pg/mL as the best cutoff to classify PML and MS relapses (figure 3B). Sensitivity and specificity associated with this cutoff were 85% and 93%, respectively.

Figure 1 Increased NFL Levels in Serum After 2 Years of NTZ Treatment in Pre-PML Patients Compared With NTZ-ctr



Graphs comparing serum NFL levels between pre-PML and NTZ-ctr patients at baseline (T0; n = 16 for pre-PML and n = 36 for NTZ-ctr), after 1 (T1; n = 12 for pre-PML and n = 36 for NTZ-ctr) and 2 years (T2; n = 12 for pre-PML and n = 34 for NTZ-ctr) of treatment and during PML (n = 13). Each symbol represents an individual, and horizontal bars indicate the median values and interquartile ranges. A y-axis segmentation was performed to represent better high and low serum NFL levels. *Refers to *p* values < 0.05. **Refers to *p* values < 0.01. ***Refers to *p* values < 0.001 in Mann-Whitney *U* tests (unpaired data) and Wilcoxon matched-paired test (paired data). NFL = neurofilament light chain; NTZ = natalizumab; PML = progressive multifocal leukoencephalopathy.

A subanalysis in the untreated relapsing patients revealed similar results to the whole cohort, with an AUC of 90% ($p = 9 \times 10^{-5}$) and the same NFL value of 52.7 pg/mL as the best cutoff to classify patients. Sensitivity and specificity were 85% and 91%, respectively.

Discussion

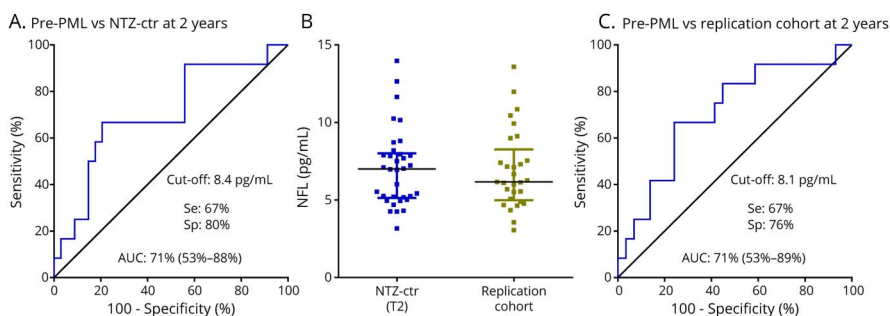
Few molecular biomarker studies have aimed to identify patients with MS at increased risk for PML under NTZ treatment.⁸⁻¹⁰ However, none of the proposed biomarkers are

at present routinely measured in clinical practice to estimate PML risk in patients with RRMS receiving NTZ. In a recent study, serum NFL levels measured with an electrochemiluminescence assay were found to be 10-fold higher at PML onset compared with the pre-PML condition.⁶ Furthermore, serum NFL levels also demonstrated high performance to discriminate between patients with MS at PML onset and NTZ-treated patients who did not develop PML and treated patients with clinical or neuroradiologic evidence of disease activity 4 weeks before sample collection.⁶

In our study, serum NFL levels measured in a prospective cohort of NTZ-treated patients with the more sensitive Simoa assay¹¹ were not predictive of PML development at baseline or after 1 year of treatment. However, despite a general significant decrease by the effect of treatment, serum NFL levels at 2 years were significantly increased in patients who will develop PML compared with NTZ-ctr patients, and NFL levels had good potential to discriminate between these 2 groups of patients in terms of PML development. Of interest, performance of serum NFL levels to predict PML in an independent cohort of NTZ-treated patients for 2 years was remarkably similar to the original cohort, results that support the use of similar cutoff values between MS centers to estimate PML risk in different cohorts of patients with MS after 2 years of NTZ treatment.

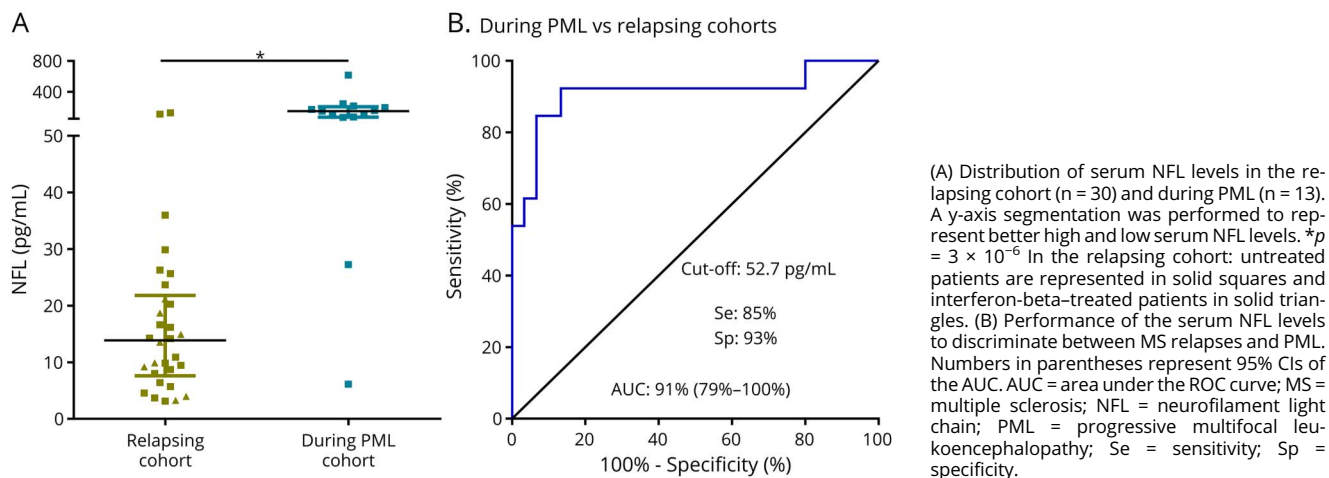
In agreement with Dalla Costa et al.,⁶ serum NFL levels were far more elevated during PML compared with earlier stages of the disease. In our study, NFL levels were 16-fold higher during PML compared with pre-PML patients at 2 years of NTZ treatment and 8-fold higher compared with a relapsing cohort. The latter may have implications in clinical practice to set a cutoff value of serum NFL levels that distinguish the PML condition from clinical relapses in patients receiving NTZ treatment. A limitation in our study was the inclusion of a relapsing cohort either untreated or receiving interferon-beta, a treatment that was not associated with significant reductions in serum NFL levels. In this context, the inclusion of a relapsing cohort of NTZ-treated patients would probably have been associated with greater differences in serum NFL levels between relapsing and PML patients, considering the

Figure 2 Serum NFL Levels Differentiate Between Pre-PML and NTZ-ctr Patients After 2 Years of NTZ Treatment



(A) Performance of serum NFL levels to discriminate between pre-PML and NTZ-ctr patients after 2 years of treatment. (B) Distribution of serum NFL levels in the NTZ-ctr cohort at 2 years of treatment (n = 34), and in an independent cohort of patients with MS treated with NTZ for 2 years (replication cohort; n = 29). (C) Performance of the serum NFL levels to discriminate between pre-PML patients at 2 years and patients from the replication cohort after 2 years of treatment. NFL = neurofilament light chain; NTZ = natalizumab; PML = progressive multifocal leukoencephalopathy.

Figure 3 NFL Levels in Serum Identify PML From MS Relapses



effect of NTZ reducing significantly serum NFL levels, as shown in our study.

Based on our findings, in patients receiving NTZ treatment, we recommend to measure NFL levels longitudinally, and those patients having protein levels above the cut-offs calculated in the study after 2 years of treatment should be monitored more closely for neurologic symptoms with additional NFL and MRI measures to rule out PML.

In conclusion, our results support the use of serum NFL levels in clinical practice to identify patients with RRMS at higher risk for PML based on protein levels at 2 years of NTZ treatment and to differentiate PML from clinical relapses in patients receiving NTZ.

Study Funding

No targeted funding reported.

Disclosure

The authors report no disclosures relevant to the manuscript. Go to Neurology.org/NN for full disclosures.

Publication History

Received by *Neurology: Neuroimmunology & Neuroinflammation* November 23, 2020. Accepted in final form March 3, 2021.

Appendix Authors

Name	Location	Contribution
Nicolás Fissolo, PhD	Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Institut de Recerca Vall d'Hebron (VHIR), Hospital Universitari Vall d'Hebron, Universitat Autònoma de Barcelona, Spain	Designed and conceptualized the study; acquisition and analysis of the data; and drafted the manuscript for intellectual content

Appendix (continued)

Name	Location	Contribution
Beatrice Pignolet, PhD	CRC-SEP Neurosciences Centre Hospitalier Universitaire Toulouse, CPTP INSERM UMR 1043 CNRS UMR 5282 et Université de Toulouse III, UPS, France	Acquisition and analysis of the data and revised the manuscript for intellectual content
Jordi Rio, MD, PhD	Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Hospital Universitari Vall d'Hebron, Universitat Autònoma de Barcelona, Spain	Analysis of the data and revised the manuscript for intellectual content
Patrick Vermersch, MD, PhD	Univ. Lille, Inserm U1172, CHU Lille, FHU Imminent, France	Acquisition of the data and revised the manuscript for intellectual content
Auréli Ruet, MD, PhD	Université, Bordeaux, France, CHU de Bordeaux, INSERM-CHU CIC-P 0005, & Services de Neurologie, France, and Neurocentre Magendie, INSERM U1215 Bordeaux, France	Acquisition of the data and revised the manuscript for intellectual content
Jerome deSèze, MD, PhD	Department of Neurology, Hôpital Civil, Strasbourg, France	Acquisition of the data and revised the manuscript for intellectual content
Pierre Labauge, MD, PhD	Department of Neurology, CHU Montpellier, France	Acquisition of the data and revised the manuscript for intellectual content
Sandra Vukusic, MD, PhD	Department of Neurology CHU Lyon, Bron, France	Acquisition of the data and revised the manuscript for intellectual content
Caroline Papeix, MD	Department of neurology, Hôpital de la Salpêtrière, Paris, France	Acquisition of the data and revised the manuscript for intellectual content

Continued

Appendix (continued)

Name	Location	Contribution
Laurent Martinez-Almoyna, MD	Chi Aix en Provence, France	Acquisition of the data and revised the manuscript for intellectual content
Ayman Tourbah, MD, PhD	Department of Neurology and Faculté de Médecine de Reims, CHU de Reims, URCA, France; LPN EA2027 Université Paris VIII, Saint-Denis, Paris, France	Acquisition of the data and revised the manuscript for intellectual content
Pierre Clavelou, MD, PhD	Department of Neurology, CHRU Clermont Ferrand, France	Acquisition of the data and revised the manuscript for intellectual content
Thibault Moreau, MD, PhD	Department of Neurology, CHU Dijon, France	Acquisition of the data and revised the manuscript for intellectual content
Jean Pelletier, MD, PhD	Aix-Marseille Univ, APHM, Hôpital de la Timone, Pôle de Neurosciences Cliniques, Service de Neurologie, CNRS, CRMBM UMR 7339, Marseille, France	Acquisition of the data and revised the manuscript for intellectual content
Christine Lebrun-Frenay, MD, PhD	Service de Neurologie, CHU de Nice Pasteur2, Université Nice Côte d'Azur UR2CA URRIS, France	Acquisition of the data and revised the manuscript for intellectual content
Bertrand Bourre, MD	Neurologie, CHU Rouen, France	Acquisition of the data and revised the manuscript for intellectual content
Gilles Defer, MD, PhD	Neurologie, CHU Caen, France	Acquisition of the data and revised the manuscript for intellectual content
Xavier Montalban, MD	Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Hospital Universitari Vall d'Hebron, Universitat Autònoma de Barcelona, Spain	Revised the manuscript for intellectual content

Appendix (continued)

Name	Location	Contribution
David Brassat, MD, PhD	CRC-SEP Neurosciences Centre Hospitalier Universitaire Toulouse, CPTP INSERM UMR 1043 CNRS UMR 5282 et Université de Toulouse III, UPS, France	Acquisition of the data and revised the manuscript for intellectual content
Manuel Comabella, MD, PhD	Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya (Cemcat), Institut de Recerca Vall d'Hebron (VHIR), Hospital Universitari Vall d'Hebron, Universitat Autònoma de Barcelona, Spain	Designed and conceptualized the study; analyzed the data; and drafted the manuscript for intellectual content

References

1. Polman CH, O'Connor PW, Havrdova E, et al. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* 2006;354(9):899–910.
2. Bloomgren G, Richman S, Hotermans C, et al. Risk of natalizumab-associated progressive multifocal leukoencephalopathy. *N Engl J Med* 2012;366(20):1870–1880.
3. Kappos L, Bates D, Edan G, et al. Natalizumab treatment for multiple sclerosis: updated recommendations for patient selection and monitoring. *Lancet Neurol* 2011;10:745–758.
4. Khalil M, Teunissen CE, Otto M, et al. Neurofilaments as biomarkers in neurological disorders. *Nat Rev Neurol* 2018;14(8):577–589.
5. Kapoor R, Smith KE, Allegretta M, et al. Serum neurofilament light as a biomarker in progressive multiple sclerosis. *Neurology* 2020;95(10):436–444.
6. Dalla Costa G, Martinelli V, Moiola L, et al. Serum neurofilaments increase at progressive multifocal leukoencephalopathy onset in natalizumab-treated multiple sclerosis patients. *Ann Neurol* 2019;85(4):606–610.
7. Outterryck O, Ongagna JC, Brochet B, et al. A prospective observational post-marketing study of natalizumab-treated multiple sclerosis patients: clinical, radiological and biological features and adverse events. The BIONAT cohort. *Eur J Neurol* 2014;21(1):40–48.
8. Schwab N, Schneider-Hohendorf T, Posevitz V, et al. L-selectin is a possible biomarker for individual PML risk in natalizumab-treated MS patients. *Neurology* 2013;81(10):865–871.
9. Pignolet B, Schwab N, Schneider-Hohendorf T, et al. CD62L test at 2 years of Natalizumab predicts progressive multifocal leukoencephalopathy. *Neurology* 2016;87(20):2491–2494.
10. Fissolo N, Pignolet B, Matute-Blanch C, et al. Matrix metalloproteinase 9 is decreased in natalizumab-treated multiple sclerosis patients at risk for progressive multifocal leukoencephalopathy. *Ann Neurol* 2017;82(2):186–195.
11. Kuhle J, Barro C, Andreasson U, et al. Comparison of three analytical platforms for quantification of the neurofilament light chain in blood samples: ELISA, electrochemiluminescence immunoassay and Simoa. *Clin Chem Lab Med* 2016;54(10):1655–1661.