Stereoselective synthesis and transformation of pinane-based 2-amino-1,3-diols

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Abstract

A library of pinane-based 2-amino-1,3-diols was synthesised in a stereoselective manner. Isopinocarveol prepared from (−)-α-pinene was converted into condensed oxazolidin-2-one in two steps by carbamate formation followed by a stereoselective aminohydroxylation process. The relative stereochemistry of the pinane-fused oxazolidin-2-one was determined by 2D NMR and X-ray spectroscopic techniques. The regioisomeric spiro-oxazolidin-2-one was prepared in a similar way starting from the commercially available (1R)-(−)-myrtenol (10). The reduction or alkaline hydrolysis of the oxazolidines, followed by reductive alkylation resulted in primary and secondary 2-amino-1,3-diols, which underwent a regioselective ring closure with formaldehyde or benzaldehyde delivering pinane-condensed oxazolidines. During the preparation of 2-phenyliminooxazolidine, an interesting ring–ring tautomerism was observed in CDCl₃.

Introduction

The best known 2-amino-1,3-diol derivative sphingosine (1) plays a crucial role in intracellular signaling as second messenger, and its derivatives called sphingolipids are also critical for cell growth, cell differentiation, cell recognition, and apoptosis [1-7]. Due to its involvement in a wide range of cellular processes, significant efforts have been made in the last two decades targeting sphingosine analogues signalling as a therapeutic strategy. For instance, FTY720-P (3), the phosphate of FTY720 (2, fingolimod), proved to be a very good agonist for the S1P1 receptor (Figure 1). Sphingosine 1-phosphate (S1P, 4), in turn, performed critical regulator functions in many physiological and pathological treatments, such as Alzheimer's disease
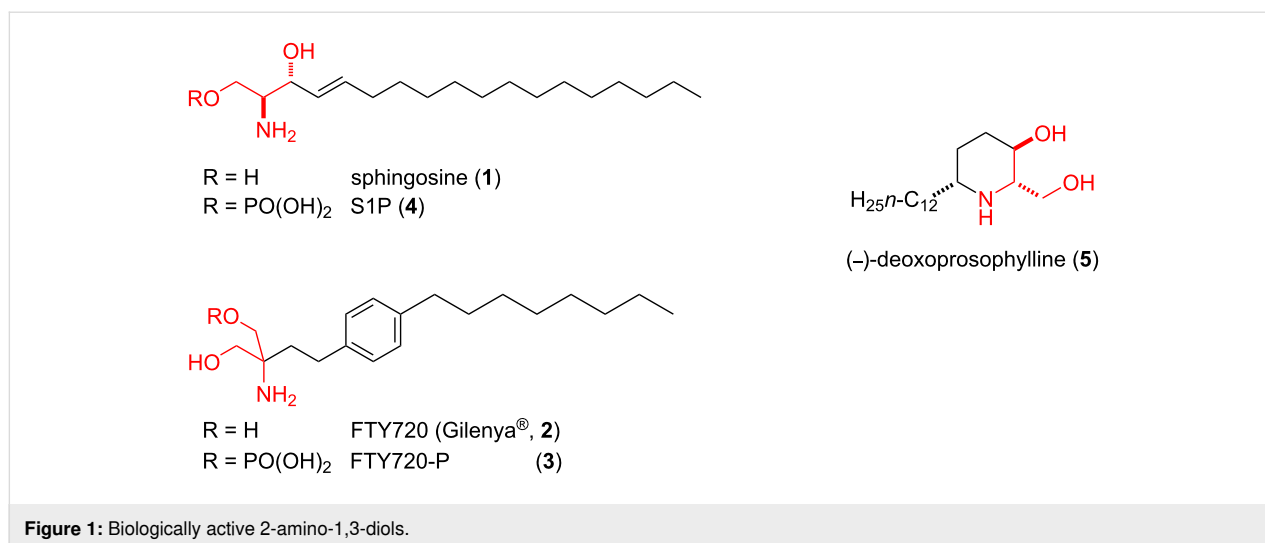
Figure 1: Biologically active 2-amino-1,3-diols.

[8,9], cancer [10-13], multiple sclerosis [14], and inflammation [15].

Due to the lack of a readily available natural sources and the high biological importance of sphingolipid analogues, their synthesis has been the subject of numerous studies [16]. The key step for the synthesis is the stereoselective construction of the 2-amino-1,3-diol moiety of the molecules. Generally, two main synthetic strategies are used to prepare these analogues. One requires the insertion of the alcohol and amino groups in the α,β position with the correct stereochemistry [17-20]. The second strategy involves a bond formation between two chiral centers to produce the targeted 2-amino-1,3-diol [21,22]. For instance, deoxoprosophylline (**5**) as a cyclic 2-amino-1,3-diol target molecule was prepared by Kokatla et al. in an 8 step synthesis starting from Perlin aldehydes, via Pd(OH)$_2$-catalyzed reductive azidoketon cyclisation [23]. Another synthetic pathway involves a stereoselective aminohydroxylation process starting from allylic carbamates usually carried out in the presence of potassium osmate [24-28].

In recent years, we have extensively studied the stereoselective synthesis, as well as catalytic and pharmacological applications of monoterpene-based 3-amino-1,2-diols, which are the regioisomers of potential monoterpenic 2-amino-1,3-diols [29-33]. These trifunctionalized terpenoids may also possess diverse biological activities and could successfully applied as chiral catalysts in enantioselective transformations [34]. In the present study, our aim was to synthesize novel, cyclic potentially analogues of sphingosine, incorporating a lipophilic natural pinane skeleton, starting from commercially available monoterpene-based allylic alcohols via a stereoselective hydroxyamination in the presence of a potassium osmate(VI) catalyst. We also planned to explore the regioselectivity of the ring closure of the resulting 2-amino-1,3-diols to obtain promising 1,3-heterocycles. To reach our goal, (1*S*)-(−)-α-pinene (**6**) and (1*R*)-(−)-myrtenol (**10**), two naturally occurring monoterpenoids were selected as precursors, as both are commercially available, cheap starting materials.
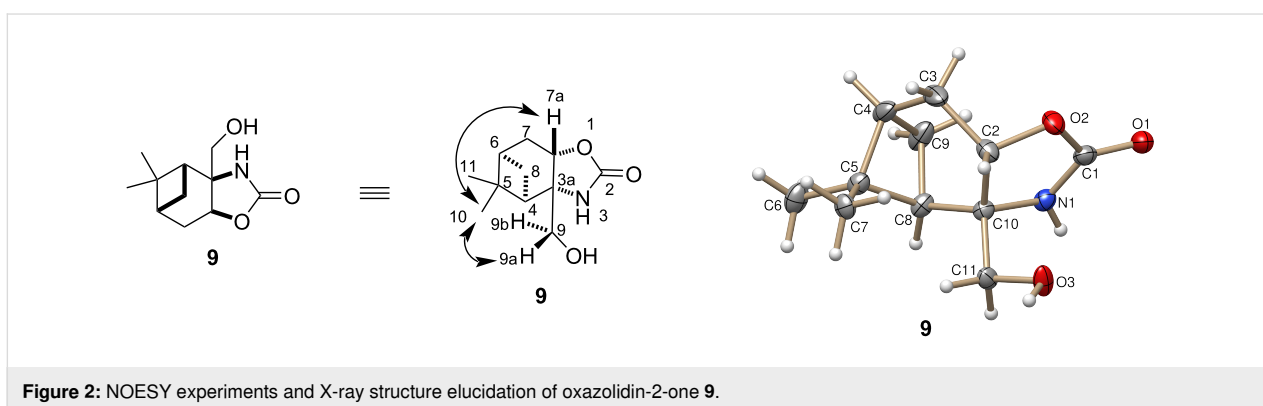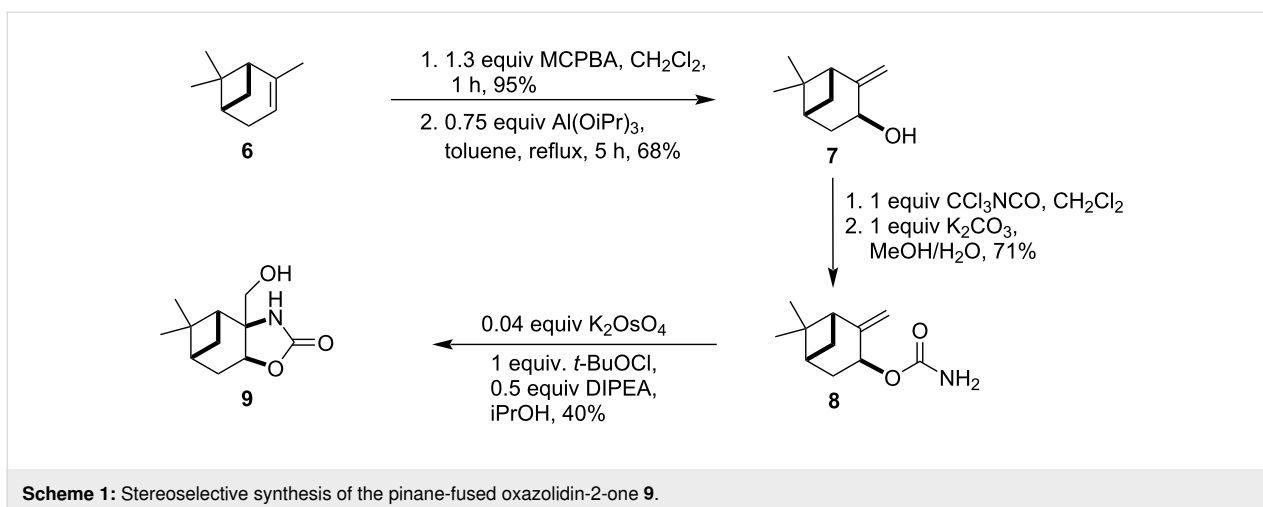
Results and Discussion
Synthesis of regioisomeric oxazolidinones from (1*S*)-(−)-α-pinene (**6**) and (1*R*)-myrtenol (**10**)

The synthesis of isopinocarveol (**7**), the key intermediate allylic alcohol, was performed according to a literature procedure in good yield [35]. The first step was the stereoselective epoxidation of (−)-α-pinene (**6**), carried out with *meta*-chloroperoxybenzoic acid (MCPBA), followed by a base-catalyzed allylic rearrangement mediated by aluminium isopropoxide (Al(O*i*Pr)$_3$). The resulting allylic alcohol **7** was reacted with trichloroacetyl isocyanate, followed by alkaline treatment, delivering carbamate **8** in good yield [27,28,36]. In the next step, the aminohydroxylation was accomplished by potassium osmate(VI) as the catalyst and *t*-BuOCl in the presence of DIPEA affording oxazolidine-2-one **9** [27]. The reaction was found to be highly stereoselective, giving exclusively the *diexo*-fused tricyclic **9** ring system (Scheme 1).

The absolute configuration of compound **9** was determined by 2D NMR spectroscopic techniques. Clear NOE signals were observed between the H-7a and Me-10 as well as the H$_a$-9 and Me-10 protons. Beside NOESY experiments, the structure was also elucidated by X-ray crystallography (Figure 2).
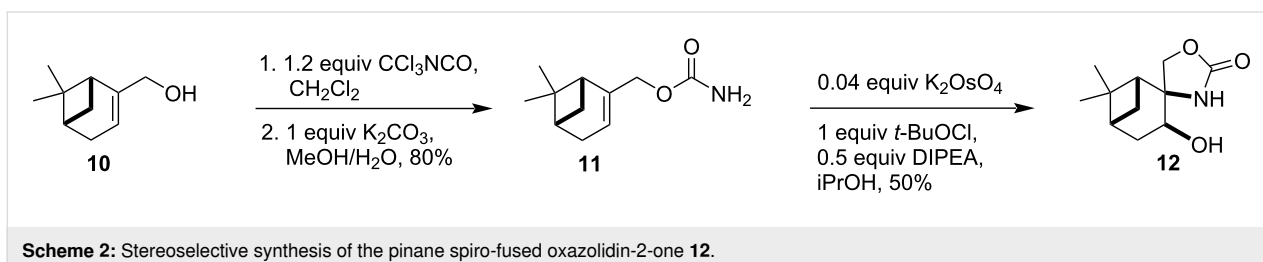
To synthesize the regioisomeric spiro-oxazolidinone derivative **12**, (1*R*)-(−)-myrtenol (**10**) was chosen as starting material

Scheme 1: Stereoselective synthesis of the pinane-fused oxazolidin-2-one **9**.

Figure 2: NOESY experiments and X-ray structure elucidation of oxazolidin-2-one **9**.
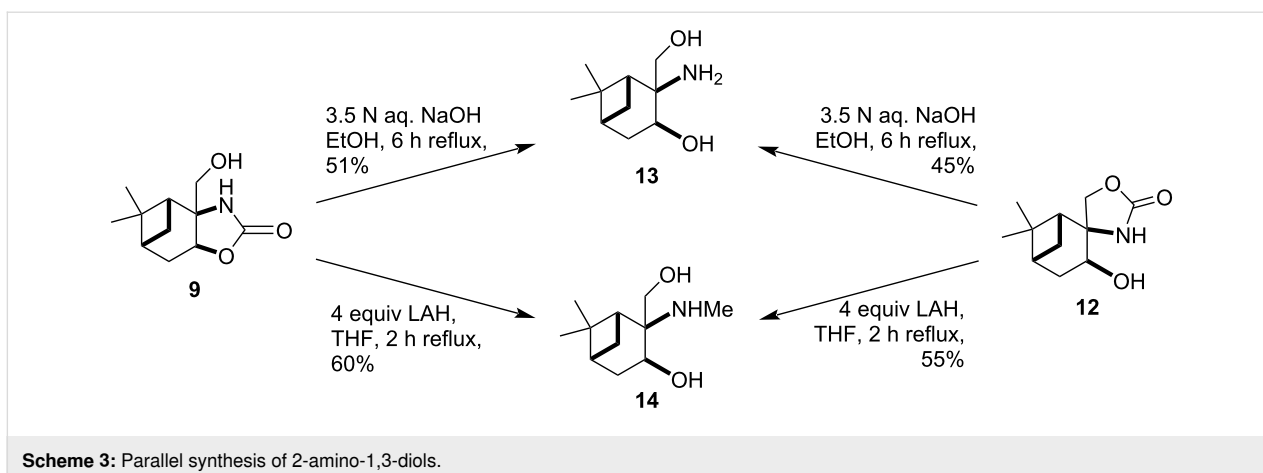
(Scheme 2). The synthetic method was similar to that mentioned above for (−)-isopinocarveol. In the first step, carbamate **11** was prepared [37], then the aminohydroxylation was carried out catalyzed by potassium osmate(VI), which led to the formation of the spiro-oxazolidine-2-one **12** in a highly regio- and stereoselective manner. Based on the NMR spectroscopic measurements of the crude product, the spiro derivative **12** was obtained exclusively with the relative configuration depicted in Scheme 2. Beside 2D NMR spectroscopic studies, the absolute configuration of compound **12** was determined by its transformation into the corresponding aminodiols **13** and **14**, comparing the products with those obtained from the regioisomer **9** (discussed in Scheme 3).

## Synthesis and transformations of pinane-based 2-amino-1,3-diols

To obtain a library of pinane-based 2-amino-1,3-diols, the oxazolidine-2-ones **9** and **12** were applied as starting materials. The alkaline hydrolysis of both **9** and **12** resulted in the same primary aminodiol **13** [38]. According to the NMR spectra and other physical and chemical properties, there was no difference between the products of the two reactions. Since the relative configuration of compound **9** was clarified by NMR spectroscopy and X-ray crystallographic results, we were able to assign the stereochemistry of spiro-derivate **12**, too. In a similar manner, the LiAlH$_4$ (LAH) reduction of both **9** and **12** gave the same *N*-methylaminodiol **14** with modest yield (Scheme 3).

Scheme 2: Stereoselective synthesis of the pinane spiro-fused oxazolidin-2-one **12**.

Scheme 3: Parallel synthesis of 2-amino-1,3-diols.

Subsequently, compound **13** was reacted with benzaldehyde. In this process, the Schiff' base **15A** was generated in situ. Our efforts to reduce it with sodium borohydride failed, since we did not observe the formation of the expected *N*-benzylaminodiol either at room temperature or under reflux conditions, probably due to the strong steric hindrance of the bicyclic system and the hydroxymethyl group. The ¹H NMR spectroscopic measurements in CDCl₃ clearly showed that the crude product was a five-component tautomeric mixture containing condensed oxazolidine **15E** as the main component. Additional minor components included the other condensed oxazolidine (**15D**),

spiro compounds **15B** and **15C** as well as the Schiff' base **15A** existing in a ratio of **15A/15B/15C/15D/15E** = 4:<1:4:12:79 (Scheme 4) [39,40]. The structures of the five components **15A–E** were determined by 2D NMR spectroscopic techniques (NOESY and HMBC). Since this finding is quite unusual in the case of Schiff' bases, we decided to study the ring/chain tautomeric mixture (**15A–E**) in the reaction of **13** with benzaldehyde by ¹H NMR spectroscopy. When a time-dependent ¹H NMR spectroscopic measurement was accomplished, we observed that the equilibrium composition was established rapidly, without any significant change in the ratio of the

15A/15B/15C/15D/15E = 4:<1:4:12:79
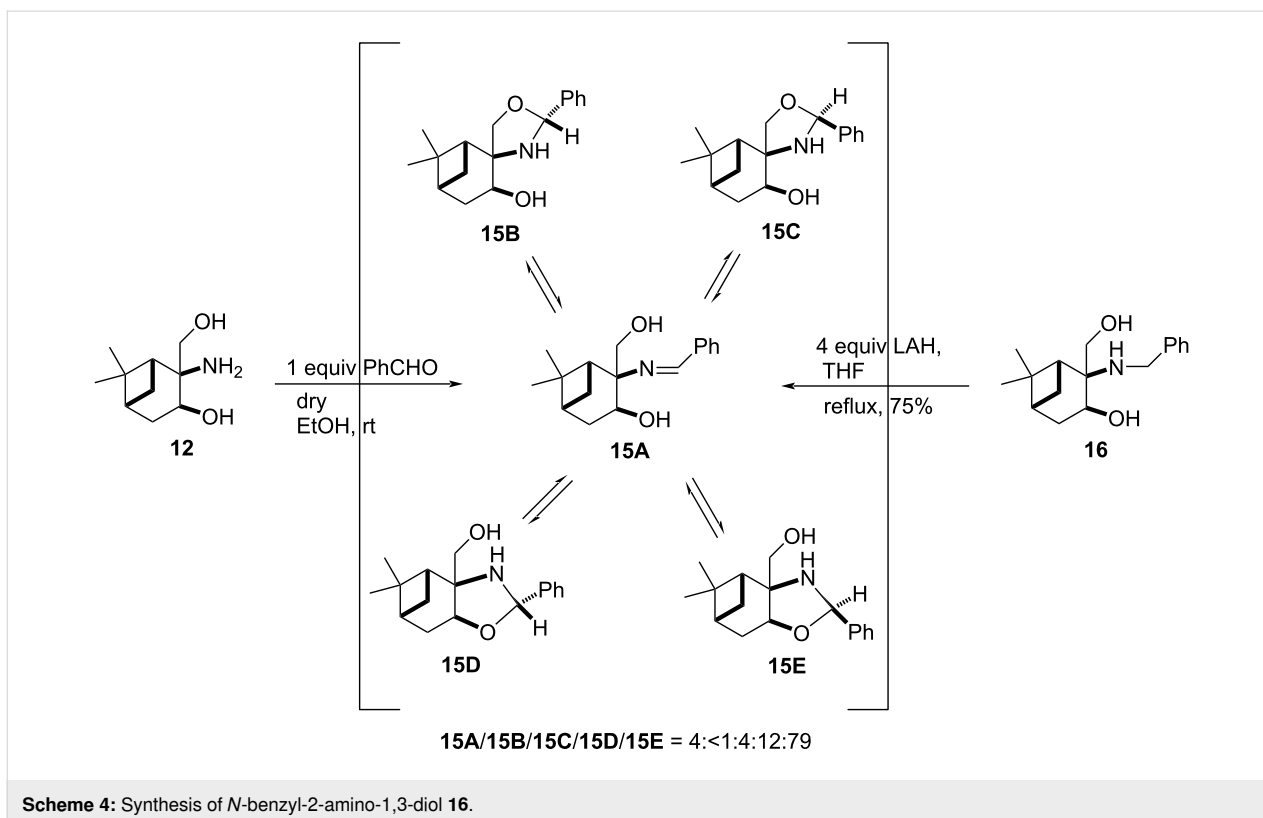
Scheme 4: Synthesis of *N*-benzyl-2-amino-1,3-diol **16**.

tautomers. The equilibrium shifting strongly to product **15E** can account of the difficulty of the reduction process and the necessity to use a stronger reducing agent and more severe conditions. The reduction step, therefore, was performed by applying LAH, a stronger reducing agent, and longer reflux, resulting in *N*-benzyl-2-amino-1,3-diol **16** (Scheme 4).
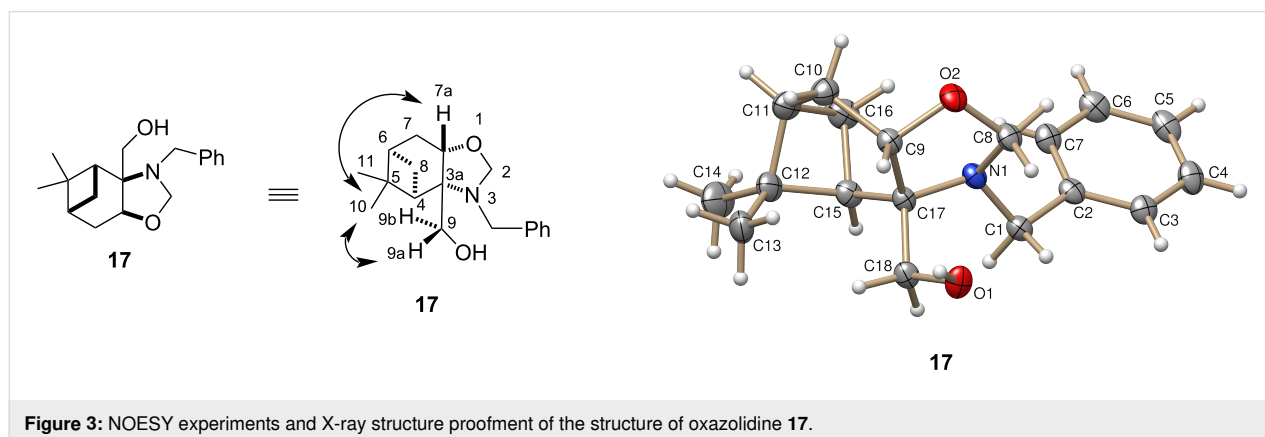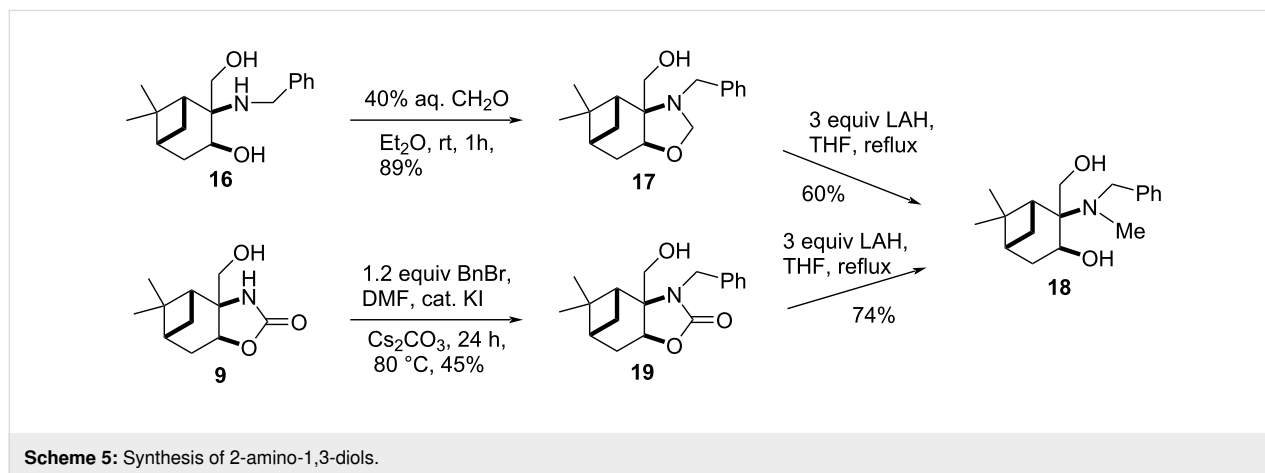
When compound **16** was treated with formaldehyde at room temperature, pinane-fused oxazolidine **17** was obtained regioselectively (Scheme 5), as it was indicated by clear HMBC correlations between the CH$_2$ of the oxazolidine ring and the anellation carbons, in contrast to the results observed in the case of the regioisomeric 3-amino-1,2-diols, where spiro-oxazolidines formed exclusively [41]. The configuration of oxazolidine **17** was determined by 2D NMR spectroscopic techniques. Clear NOE signals were observed between the H-7a and Me-10 as well as the H$_a$-9 and Me-10 protons. In addition to NOESY experiments, the structure was also elucidated by X-ray crystallography (Figure 3).
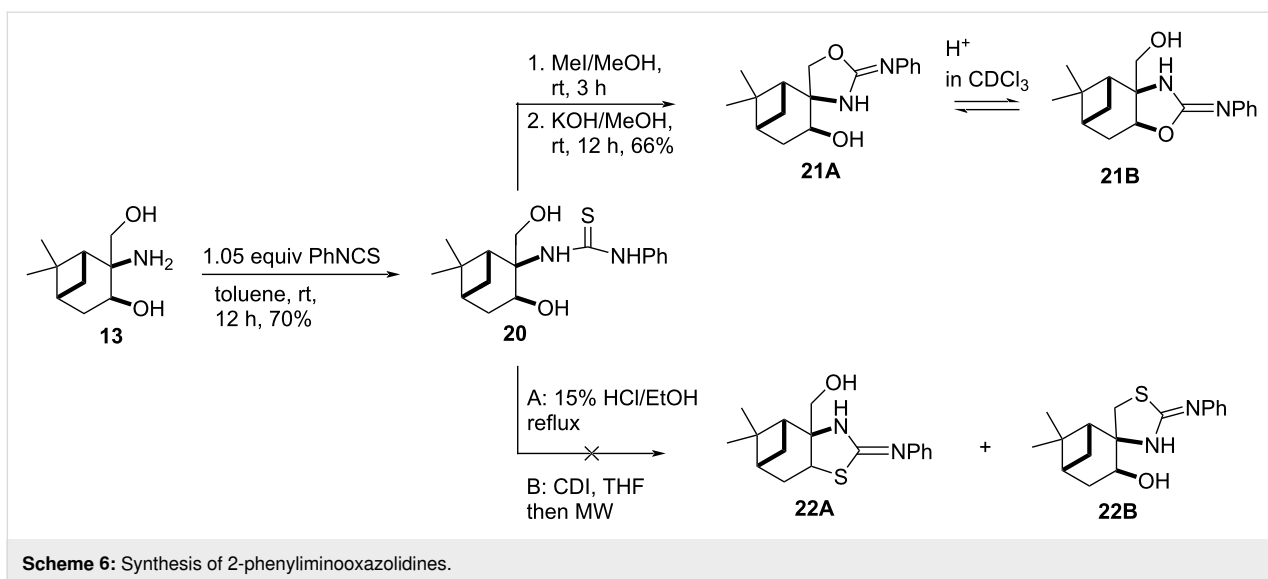
The LAH reduction of oxazolidine **17** gave *N*-benzyl-*N*-methyl analogue **18** which, alternatively, was prepared directly from 2-oxazolidinone **9** via *N*-benzylation followed by LAH reduction in 2 steps.

When compound **13** was reacted with phenylisothiocyanate, thiourea **20** was obtained, which underwent a regioselective ring closure resulting in **21A**. The structure of **21A** was determined by ^{1}H (whereas the CH-*OH* gave a dublet in DMSO-d_6 while the CH$_2$-*OH* of **21B** could be detected as triplet) and 2D NMR spectroscopic techniques (HMBC). It is important to mention that this regioselectivity is the opposite to that observed in the reaction of aminodiols **13** and **16** with aldehydes (see Scheme 4 and Scheme 5), but it is similar to that observed in our earlier study with pinane-based 3-amino-1,2-diols [41]. During the NMR spectroscopic study of **21A** in CDCl$_3$ for 30 days, an unknown slow ring–ring tautomerization was observed, forming a 1:1 mixture of the two regioisomers **21A** and **21B**. Compound **21B** could be isolated from the mixture by column chromatography in pure form.

The synthesis of the heteroanalogue 2-phenyliminothiazolidines **22A** and **22B** failed, even when the reaction was attempted under acidic or even milder conditions (Scheme 6).

Scheme 5: Synthesis of 2-amino-1,3-diols.

Figure 3: NOESY experiments and X-ray structure proofment of the structure of oxazolidine **17**.

Scheme 6: Synthesis of 2-phenyliminooxazolidines.

The proposed reaction pathway for the ring–ring tautomerism of **21A** and **21B** is presented in Figure 4 and it explains why the acidic environment (present generally in CDCl₃ solution) is necessary. In a similar manner, an oxazolidine–1,3-oxazine tautomerism of pulegone-based 3-amino-1,2-diols was recently reported [31]. When compound **21A** or **21B** were treated in less protic solvents such as DMSO-d_6 or CD₃OD, tautomerization was not observed.

Conclusion

A small library of pinane-based 2-amino-1,3-diols was synthesized in a stereoselective manner starting from (1R)-(−)-myrtenol and isopinocarveol prepared from α-pinene. Pinane-condensed or spiro-oxazolidin-2-ones were formed in three steps by a stereoselective hydroxyamination process. The relative stereochemistry of new compounds was determined by 2D NMR spectroscopic and X-ray techniques. The resulting primary and secondary 2-amino-1,3-diols underwent a regioselec-
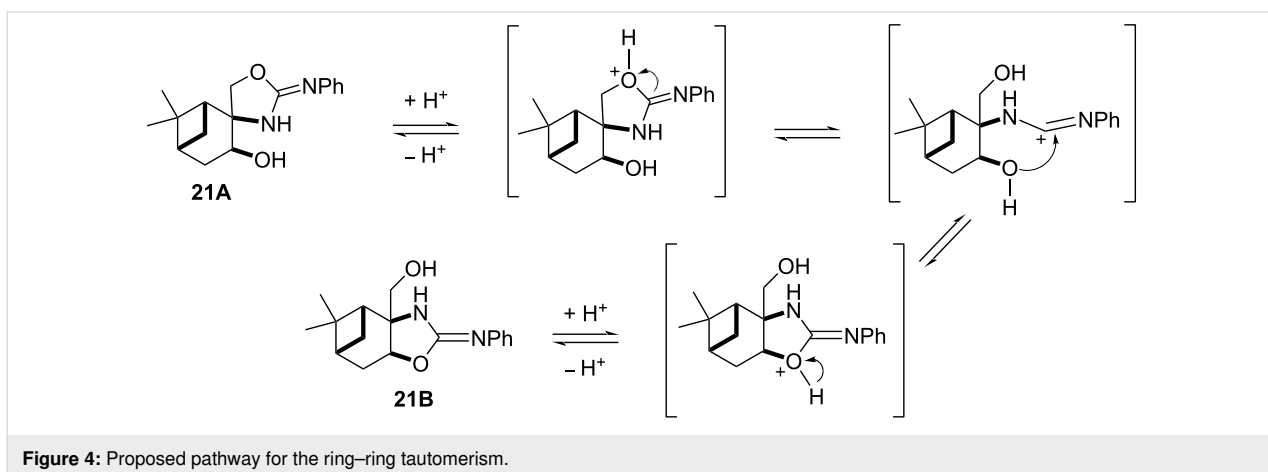
tive ring closure with formaldehyde and benzaldehyde producing pinane-condensed oxazolidines. In the case of 2-phenylimi-nooxazolidine, an interesting ring–ring tautomerism was observed in CDCl₃. The prepared trifunctional compounds may serve as chiral catalysts in enantioselective transformations, while the 2-phenyliminooxazolidines could be interesting in the field of antiproliferative or antioxidants studies based on our former studies on 2-imino-1,3-heterocycles [42,43].

Supporting Information

Supporting Information File 1

Experimental part, analytical data, NMR spectra and X-ray data of the prepared compounds.
[https://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-17-80-S1.pdf]

Figure 4: Proposed pathway for the ring–ring tautomerism.

Funding

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References

1. Posse de Chaves, E.; Sipione, S. *FEBS Lett.* **2010,** *584,* 1748–1759. doi:10.1016/j.febslet.2009.12.010
2. Hannun, Y. A.; Obeid, L. M. *J. Biol. Chem.* **2002,** *277,* 25847–25850. doi:10.1074/jbc.r200008200
3. Kågedal, K.; Zhao, M.; Svensson, I.; Brunk, U. T. *Biochem. J.* **2001,** *359,* 335. doi:10.1042/0264-6021:3590335
4. Kolesnick, R. N.; Goñi, F. M.; Alonso, A. *J. Cell. Physiol.* **2000,** *184,* 285–300. doi:10.1002/1097-4652(200009)184:3<285::aid-jcp2>3.0.co;2-3
5. Perry, D. K.; Hannun, Y. A. *Biochim. Biophys. Acta, Mol. Cell Biol. Lipids* **1998,** *1436,* 233–243. doi:10.1016/s0005-2760(98)00145-3
6. Igarashi, Y. *J. Biochem.* **1997,** *122,* 1080–1087. doi:10.1093/oxfordjournals.jbchem.a021865
7. Spiegel, S.; Foster, D.; Kolesnick, R. *Curr. Opin. Cell Biol.* **1996,** *8,* 159–167. doi:10.1016/s0955-0674(96)80061-5
8. Takasugi, N.; Sasaki, T.; Suzuki, K.; Osawa, S.; Isshiki, H.; Hori, Y.; Shimada, N.; Higo, T.; Yokoshima, S.; Fukuyama, T.; Lee, V. M.- Y.; Trojanowski, J. Q.; Tomita, T.; Iwatsubo, T. *J. Neurosci.* **2011,** *31,* 6850–6857. doi:10.1523/jneurosci.6467-10.2011
9. Prager, B.; Spampinato, S. F.; Ransohoff, R. M. *Trends Mol. Med.* **2015,** *21,* 354–363. doi:10.1016/j.molmed.2015.03.006
10. Heffernan-Stroud, L. A.; Obeid, L. M. Sphingosine Kinase 1 in Cancer. *Advances in Cancer Research;* Elsevier, 2013; Vol. 117, pp 201–235. doi:10.1016/b978-0-12-394274-6.00007-8
11. Pyne, N. J.; Tonelli, F.; Lim, K. G.; Long, J. S.; Edwards, J.; Pyne, S. *Biochem. Soc. Trans.* **2012,** *40,* 94–100. doi:10.1042/bst20110602
12. Plano, D.; Amin, S.; Sharma, A. K. *J. Med. Chem.* **2014,** *57,* 5509–5524. doi:10.1021/jm4011687
13. Pyne, N. J.; Pyne, S. *Nat. Rev. Cancer* **2010,** *10,* 489–503. doi:10.1038/nrc2875
14. Chi, H. *Trends Pharmacol. Sci.* **2011,** *32,* 16–24. doi:10.1016/j.tips.2010.11.002
15. Maceyka, M.; Spiegel, S. *Nature* **2014,** *510,* 58–67. doi:10.1038/nature13475
16. Howell, A. R.; So, R. C.; Richardson, S. K. *Tetrahedron* **2004,** *60,* 11327–11347. doi:10.1016/j.tet.2004.09.056
17. Cardillo, G.; Orena, M.; Porzi, G.; Sandri, S. *J. Chem. Soc., Chem. Commun.* **1981,** 465. doi:10.1039/c39810000465
18. Azuma, H.; Takao, R.; Niiro, H.; Shikata, K.; Tamagaki, S.; Tachibana, T.; Ogino, K. *J. Org. Chem.* **2003,** *68,* 2790–2797. doi:10.1021/jo0206824
19. He, L.; Byun, H.-S.; Bittman, R. *J. Org. Chem.* **2000,** *65,* 7627–7633. doi:10.1021/jo001226n
20. Sasai, H.; Tokunaga, T.; Watanabe, S.; Suzuki, T.; Itoh, N.; Shibasaki, M. *J. Org. Chem.* **1995,** *60,* 7388–7389. doi:10.1021/jo00128a005
21. Masui, M.; Shioiri, T. *Tetrahedron Lett.* **1998,** *39,* 5199–5200. doi:10.1016/s0040-4039(98)01020-x
22. Ma, N.; Ma, D. *Tetrahedron: Asymmetry* **2003,** *14,* 1403–1406. doi:10.1016/s0957-4166(03)00275-1
23. Kokatla, H. P.; Lahiri, R.; Kancharla, P. K.; Doddi, V. R.; Vankar, Y. D. *J. Org. Chem.* **2010,** *75,* 4608–4611. doi:10.1021/jo100489k
24. Han, H.; Cho, C.-W.; Janda, K. D. *Chem. – Eur. J.* **1999,** *5,* 1565–1569. doi:10.1002/(sici)1521-3765(19990503)5:5<1565::aid-chem1565>3.0.co;2-j
25. Rudolph, J.; Sennhenn, P. C.; Vlaar, C. P.; Sharpless, K. B. *Angew. Chem., Int. Ed. Engl.* **1996,** *35,* 2810–2813. doi:10.1002/anie.199628101
26. Donohoe, T. J.; Johnson, P. D.; Helliwell, M.; Keenan, M. *Chem. Commun.* **2001,** 2078–2079. doi:10.1039/b107253f
27. Donohoe, T. J.; Johnson, P. D.; Cowley, A.; Keenan, M. *J. Am. Chem. Soc.* **2002,** *124,* 12934–12935. doi:10.1021/ja0276117
28. Hovey, M. T.; Eklund, E. J.; Pike, R. D.; Mainkar, A. A.; Scheerer, J. R. *Org. Lett.* **2011,** *13,* 1246–1249. doi:10.1021/ol200155p
29. Szakonyi, Z.; Hetényi, A.; Fülöp, F. *Tetrahedron* **2008,** *64,* 1034–1039. doi:10.1016/j.tet.2007.07.065
30. Csillag, K.; Németh, L.; Martinek, T. A.; Szakonyi, Z.; Fülöp, F. *Tetrahedron: Asymmetry* **2012,** *23,* 144–150. doi:10.1016/j.tetasy.2012.01.020
31. Gonda, T.; Szakonyi, Z.; Csámpai, A.; Haukka, M.; Fülöp, F. *Tetrahedron: Asymmetry* **2016,** *27,* 480–486. doi:10.1016/j.tetasy.2016.04.009
32. Szakonyi, Z.; Csőr, Á.; Csámpai, A.; Fülöp, F. *Chem. – Eur. J.* **2016,** *22,* 7163–7173. doi:10.1002/chem.201600749
33. Le, T. M.; Csámpai, A.; Fülöp, F.; Szakonyi, Z. *Chem. – Eur. J.* **2018,** *24,* 13607–13615. doi:10.1002/chem.201802484
34. El Alami, M. S. I.; El Amrani, M. A.; Agbossou-Niedercorn, F.; Suisse, I.; Mortreux, A. *Chem. – Eur. J.* **2015,** *21,* 1398–1413. doi:10.1002/chem.201404303
35. Lavallee, P.; Bouthillier, G. *J. Org. Chem.* **1986,** *51,* 1362–1365. doi:10.1021/jo00358a041
36. Miller, K. E.; Wright, A. J.; Olesen, M. K.; Hovey, M. T.; Scheerer, J. R. *J. Org. Chem.* **2015,** *80,* 1569–1576. doi:10.1021/jo502493e
37. Kamon, T.; Shigeoka, D.; Tanaka, T.; Yoshimitsu, T. *Org. Biomol. Chem.* **2012,** *10,* 2363–2365. doi:10.1039/c2ob07190h
38. Byun, H.-S.; Bittman, R. *Chem. Phys. Lipids* **2012,** *165,* 794–801. doi:10.1016/j.chemphyslip.2012.10.002
39. Lázár, L.; Fülöp, F. *Eur. J. Org. Chem.* **2003,** 3025–3042. doi:10.1002/ejoc.200300142
40. Hetényi, A.; Szakonyi, Z.; Klika, K. D.; Pihlaja, K.; Fülöp, F. *J. Org. Chem.* **2003,** *68,* 2175–2182. doi:10.1021/jo026428t
41. Szakonyi, Z.; Hetényi, A.; Fülöp, F. *ARKIVOC* **2007,** No. iii, 33–42. doi:10.3998/ark.5550190.0009.305

42. Szakonyi, Z.; Zupkó, I.; Sillanpää, R.; Fülöp, F. *Molecules* **2014,** *19,* 15918–15937. doi:10.3390/molecules191015918

43. Firpo, G.; Ramírez, M. L.; Faillace, M. S.; Mendes de Brito, M. d. R.; Correia Lima e Silva, A. P. S.; Pereira Costa, J.; Rodríguez, M. C.; Argüello, G. A.; Szakonyi, Z.; Fülöp, F.; Peláez, W. J. *Antioxidants* **2019,** *8,* 197. doi:10.3390/antiox8060197