



Copy neutral loss of heterozygosity (cnLOH) patterns in synchronous colorectal cancer

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Received: 10 April 2020 / Revised: 6 October 2020 / Accepted: 12 November 2020 / Published online: 2 December 2020
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Abstract

Copy neutral loss of heterozygosity (cnLOH) is a common event in several human malignancies—positing this as a mechanism of carcinogenesis. However, the role of cnLOH in synchronous colorectal cancer (SCRC), a unique CRC subtype, is not well understood. The aim of this study was to establish a cnLOH profile of SCRC using a single-nucleotide polymorphism array (SNP-A), and to explore associations between cnLOH and the genomic landscape of frequently mutated genes in SCRC. Among 74 paired SCRC cases, the most frequently altered regions were 16p11.2–p11.1 (59.5%) and 11p11.2–p11.12 (28.4%). Notably, the 6q11.21–q11.22 region altered by cnLOH was uniquely associated with polyclonal SCRCs ($p = 0.038$). Together, our analysis suggests that inactivation of tumor suppressor genes and cnLOH are rare events among SCRC cases. This study defines distinct patterns of cnLOH in SCRC, and provides initial evidence of a role for cnLOH in SCRC etiology.

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Supplementary information The online version of this article (<https://doi.org/10.1038/s41431-020-00774-w>) contains supplementary material, which is available to authorized users.

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Introduction

Colorectal cancer (CRC) is one of the most common malignant tumors worldwide [1]. Patients with CRC have an increased risk for developing multiple primary CRCs, which encompasses synchronous CRC (SCRC) and metachronous CRC [2]. SCRC cases provide a good model to evaluate a possible field effect—neoplasms that develop in a background of common etiologic factors within an individual, and which may also share molecular features indicating a likely clonal origin [3].

Copy-number alterations (CNAs) are recurrent molecular changes in human cancers and show a tumor-specific landscape of DNA gains and losses [4]. Several studies have posited a possible clonal origin in tumor subsets, including bilateral breast cancer or SCRC cases, based on the concordance of CNA profiles or mutational status [5, 6]. As such, we recently defined a SCRC classification based upon paired-tumor clonality via single-nucleotide polymorphism arrays (SNP-As), which allowed us to characterize SCRC genomic profiles by identifying DNA gains and losses [6]. SCRC cases were classified as monoclonal when paired tumors shared similar patterns of somatic changes indicative that could be derived from a single cell or polyclonal if paired tumors had independent molecular profiles. Combination of clonality analysis and tumor

location also led to SCRC categorization as monoclonal monosegmental (MM), monoclonal pancolonic (MP), polyclonal monosegmental (PM), and polyclonal pancolonic (PP) [6].

The use of SNP-A allows for identification of loss of heterozygosity (LOH) and copy neutral LOH (cnLOH) [7]. cnLOH leads to LOH by duplication of one chromosome (or chromosomal region) and concurrent loss of the other allele [8]. According to the two-hit-hypothesis by Knudson, regions with LOH may contain tumor suppressor genes (TSGs), which could be inactivated by either a non-synonymous variant or a genomic loss. In this context, cnLOH could arise as an alternative mechanism to inactivate TSGs as well as activate oncogenes in CRC. Indeed, several studies have described gains of function of homozygous variants in oncogenes and have found that homozygously mutated genes also are localized in cnLOH regions [9].

In this study, we examined regions with cnLOH using SNP-A in a cohort of SCRC patients, and association of cnLOH by clonality and tumor location [6]. We also integrated the mutational profile of frequently mutated genes in SCRCs with the presence of cnLOH in the regions where these genes are encoded in order to explore underlying SCRC etiologies.

Materials and methods

See Supplementary Material Boxes 1 and 2.

Results

cnLOH profiling in SCRC

Overall, the majority of SCRC cases showed important proportions of regions affected by cnLOH—with a median of 7.5 events per sample. By categorization, we observed a median of 7, 4.5, 8, and 8 cnLOH events for MM, MP, PM, and PP, respectively (Fig. 1). Overlap between CNAs and regions with LOH were illustrated for each SCRC subtype (Supplementary Material Box 3). Recurrent chromosome

regions in our SCRC samples affected by cnLOH are listed in Table 1. Notably, the most commonly altered region across all SCRC subtypes was 16p11.2–p11.1 (59.5%), followed by 11p11.2–p11.12 (28.4%) (Supplementary Material Box 4). Across monoclonal groups, a total of 20 tumors showed cnLOH in 16p11.2–p11.1, including 14 paired tumors from the same patients (70.0%). Twenty tumors also showed the alteration within polyclonal groups, of which only 10 were paired tumors (50.0%). While SCRC groups presented with similar alteration patterns, region 6p22.1–p21.32 affected by cnLOH was uniquely linked to polyclonal groups ($p = 0.038$) (Table 1B).

cnLOH in CRC-related genes

Recent studies have proposed cnLOH as a “second hit” to inactivate TSGs as well as to activate oncogenes [7, 10]. In order to study cnLOH in SCRC, we explored associations between the presence of cnLOH and genomic landscape of commonly altered SCRC genes. We evaluated DNA sequencing data (where only protein truncating and damaging missense variants were considered) and examined regions where these genes were located to integrate mutation status with cnLOH events in SCRC. In our cohort, 33 (44.6%) tumors presented with *APC* variants—of which 9 cases (27.3%) also had cnLOH detected. Approximately 37.5% of monoclonal groups presented with both *APC* variants and cnLOH events in this region, compared with only 18.7% of polyclonal cases. *KRAS* variants were also observed in 32 (43.2%) neoplasms, irrespective of cnLOH events. Thirteen (17.6%) tumors had detected variants in *TP53*, with corresponding cnLOH observed in only one (7.7%) of these tumors. Seven (9.5%) tumors presented with *FBXW7* variants, although only one (14.3%) variant was associated with cnLOH. Although *SMAD4* variants were associated with the MM subgroup ($p = 0.049$), no tumors presented with concurrent cnLOH (Table 2).

Discussion

SCRC is a heterogeneous disease that remains poorly understood. Consequently, defining genetic patterns specific

Fig. 1 Quantification of cnLOH events in SCRC groups. **A** Four SCRC groups. **B** Two SCRC groups according to clonality. Blue line indicates the median of cnLOH in the whole SCRC cohort.

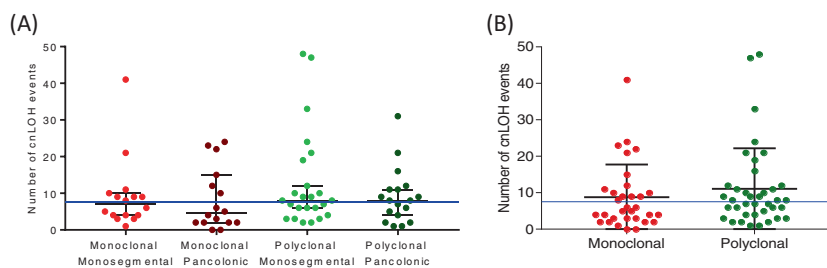


Table 1 Frequent regions affected by cnLOH in SCRC cases: (A) Four SCRC groups and (B) Two SCRC groups.

(A)								
Chromosome	Cytoband start	Cytoband end	SCRC	MM SCRC	MP SCRC	PM SCRC	PP SCRC	<i>p</i> value ^a
16	p11.2	p11.1	44 (59.5)	10 (62.5)	12 (75.0)	12 (50.0)	10 (55.6)	NS
11	p11.2	p11.12	21 (28.4)	5 (31.3)	3 (18.8)	5 (20.8)	8 (44.4)	NS
2	q11.1	q11.2	11 (14.9)	1 (6.3)	3 (18.8)	2 (8.3)	5 (27.8)	NS
10	q22.1	q22.2	11 (14.9)	4 (25.0)	1 (6.3)	4 (16.7)	2 (11.1)	NS
8	q11.1	q23.3	10 (13.5)	2 (12.5)	5 (31.3)	3 (12.5)	–	NS
6	p22.1	p21.32	9 (12.2)	–	1 (6.25)	6 (25.0)	2 (11.1)	NS
7	q11.21	q11.22	9 (12.2)	4 (25.0)	–	3 (12.5)	2 (11.1)	NS
4	q24	q35.2	7 (9.5)	2 (12.5)	–	3 (12.5)	2 (11.1)	NS
5	q34	q35.3	7 (9.5)	1 (6.3)	3 (18.8)	2 (8.3)	1 (5.6)	NS
8	p23.1	p11.1	7 (9.5)	2 (15.5)	3 (18.8)	2 (8.3)	–	NS
10	p12.31	p12.1	7 (9.5)	2 (12.5)	3 (18.8)	1 (4.2)	1 (5.6)	NS
(B)								
Chromosome	Cytoband start	Cytoband end	SCRC	Monoclonal	Polyclonal	<i>p</i> value ^a		
16	p11.2	p11.1	44 (59.5)	22 (68.8)	22 (52.4)	NS		
11	p11.2	p11.12	21 (28.4)	8 (25.0)	13 (31.0)	NS		
2	q11.1	q11.2	11 (14.9)	4 (12.5)	7 (16.7)	NS		
10	q22.1	q22.2	11 (14.9)	5 (15.6)	6 (14.3)	NS		
8	q11.1	q23.3	10 (13.5)	7 (21.9)	3 (7.1)	NS		
6	p22.1	p21.32	9 (12.2)	1 (3.1)	8 (19.0)	0.038		
7	q11.21	q11.22	9 (12.2)	4 (12.5)	5 (11.9)	NS		
4	q24	q35.2	7 (9.5)	2 (6.3)	5 (11.9)	NS		
5	q34	q35.3	7 (9.5)	4 (12.5)	3 (7.1)	NS		
8	p23.1	p11.1	7 (9.5)	5 (15.6)	2 (4.8)	NS		
10	p12.31	p12.1	7 (9.5)	5 (15.6)	2 (4.8)	NS		

^aStatistical analysis was performed by Pearson's Chi square (χ^2) test. Parentheses refer to percentage numbers. *cnLOH* copy-number-neutral loss of heterozygosity, *SCRC* synchronous colorectal cancer, *MM* monoclonal monosegmental, *MP* monoclonal pancolonial, *PM* polyclonal monosegmental, *PP* polyclonal pancolonial.

Table 2 The most frequently mutated genes accompanied by cnLOH in SCRC.

	SCRC	MM SCRC	MP SCRC	PM SCRC	PP SCRC	<i>p</i> value ^a
<i>APC</i>						
“1 st hit” mutation	33 (44.6)	10 (62.5)	6 (37.5)	9 (37.5)	7 (38.9)	NS
“2 nd hit” cnLOH	9 (27.3)	4 (40.0)	2 (33.3)	2 (22.2)	1 (14.3)	NS
<i>KRAS</i>						
“1 st hit” mutation	32 (43.2)	5 (31.3)	7 (43.8)	11 (45.8)	9 (50.0)	NS
“2 nd hit” cnLOH	–	–	–	–	–	–
<i>TP53</i>						
“1 st hit” mutation	13 (17.6)	4 (25.0)	–	4 (16.7)	5 (27.8)	NS
“2 nd hit” cnLOH	1 (7.7)	1 (25.0)	–	1 (25.0)	–	–
<i>FBXW7</i>						
“1 st hit” mutation	7 (9.5)	3 (18.8)	1 (6.3)	1 (4.2)	2 (27.8)	NS
“2 nd hit” cnLOH	1 (14.3)	–	–	–	1 (50.0)	–
<i>SMAD4</i>						
“1 st hit” mutation	4 (5.4)	3 (18.8)	–	–	1 (5.6)	0.049
“2 nd hit” cnLOH	–	–	–	–	–	–

^aStatistical analysis was performed by Pearson's chi square (χ^2) test. Parentheses refer to percentage numbers. *cnLOH* copy neutral loss of heterozygosity, *SCRC* synchronous colorectal cancer, *MM* monoclonal monosegmental, *MP* monoclonal pancolonial, *PM* polyclonal monosegmental, *PP* polyclonal pancolonial, *NS* not significant.

to SCRC cases may provide critical insight into possible clonal origins of these tumors. Previous work by our group explored CNAs and tumor location for paired SCRCs, and defined a unique classification system based on clonality and tumor site [6]. In this study, we employed SNP-A to assess cnLOH profiles in SCRC among all cases and by subgroup classification in order to better understand etiologies underlying sporadic colorectal carcinogenesis.

Previous reports have analyzed cnLOH profiles across diverse gastrointestinal tumors and revealed the lowest level of regions with cnLOH (3 per sample) for colon cancers by tumor type [10]. Nevertheless, compared with previous results, our specific interrogation of SCRCs revealed a higher frequency of regions altered by cnLOH (median of 7.5 events per sample). We also observed that SCRCs showed a distinct pattern of cnLOH events compared with cases with one primary invasive CRC [10]. It is important to note that while previous studies have leveraged similar techniques and cnLOH criteria, the biospecimen type (e.g. fresh tissue vs FFPE) across studies may also lend to some of these noted differences. Our results also pointed out that the generation of regions with cnLOH in SCRC occurred mainly in the genomic region 16p11.2–p11.1 (59.5%). In contrast, several studies have reported high frequencies of 3p, 8q, 13q, or 20q [11], as well as 5q12.1–q35.3 [10] regions in CRC. Consequently, our findings emphasize the importance of considering CRC heterogeneity in clinical settings and support additional large-scale studies to validate these results. In addition, comparison of cnLOH profiles by clinicopathological features and between SCRC and MCRC cases would provide additional insight into distinct etiologies of colorectal carcinogenesis.

Our discovery of LOH in region 16p11.2–11.1 was largely attributable to cnLOH as it is not a region affected by genomic alterations. Previous studies have reported homologous segmental duplications in this region [12] and, importantly, this region has been associated with early-onset obesity [13] (a well-known risk factor for CRC). Additional studies have revealed that *SN2B1* is encoded within this region, and is a gene associated with cancer progression [14]. *MAZ*—a transcription factor upregulated in several human cancers—is also located in this region [15, 16]. Indeed, several studies have suggested an important role for *MAZ* in CRC progression by inducing inflammation through the regulation of *STAT3* signaling [17]. Additionally, our analysis revealed differential regions altered by cnLOH by SCRC classification, as the 6p22.1–p21.32 genomic region was uniquely associated with the polyclonal tumor subgroup. This finding, together with the more common cnLOH in the 16p11.2–p11.1 region, or the connection of *APC* variants and cnLOH events in the corresponding region, support our idea that

altered chromosomal segments linked with the SCRC carcinogenesis may provide more detailed insight into SCRC clonality [6, 18, 19].

cnLOH events could also act as a “second hit” to achieve biallelic inactivation of TSGs—a potential mechanism for oncogene activation [20]. In this study, we examined the mutational status of *APC* recurrently inactivated in CRC and its association with cnLOH. Our results showed the possibility that, among SCRC cases, cnLOH could contribute to the functional impairment of this gene, more frequently in monoclonal tumors. However, as no previous associations have been observed between *TP53*, *FBXW7*, and *SMAD4*, with cnLOH, these results suggest that cnLOH events may not play a main role in SCRC. Moreover, cnLOH may not be involved in oncogenic activation of *KRAS* among these cases—however, prospective studies are warranted to confirm these findings.

In summary, SCRC cases harbor distinct molecular patterns, including the number of cnLOH per tumor as well as regions frequently altered by cnLOH, when compared with single primary invasive CRC profiles. Furthermore, inactivation of some TSGs via damaging variants and cnLOH were rare events among SCRC cases. Additional studies are needed to further explore the role of cnLOH in SCRC etiology.

Acknowledgements We thank the Tumor Registry of the Pathology Department of the 12 University Hospital, Madrid.

Funding This work was funded by Projects PI16/01650 to JP, and PI16/01920 to RG-S, from the Spanish Ministry of Health and Consumer Affairs and FEDER, by Project 2012-0036 from the Mutua Madrileña Foundation. ANH was supported by the National Institutes of Health K12 HD043483 from the Eunice Kennedy Shriver National Institute of Child Health and Human Development.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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