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Full Paper

Computational study of the therapeutic potentials of a new series of imidazole derivatives against SARS-CoV-2



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ABSTRACT

Owing to the urgent need for therapeutic interventions against the SARS-coronavirus 2 (SARS-CoV-2) pandemic, we employed an in silico approach to evaluate the SARS-CoV-2 inhibitory potential of newly synthesized imidazoles. The inhibitory potentials of the compounds against SARS-CoV-2 drug targets - main protease (Mpro), spike protein (Spro) and RNA-dependent RNA polymerase (RdRp) were investigated through molecular docking analysis. The binding free energy of the protein-ligand complexes were estimated, pharmacophore models were generated and the absorption, distribution, metabolism, excretion and toxicity (ADMET) properties of the compounds were determined. The compounds displayed various levels of binding affinities for the SARS-CoV-2 drug targets. Bisimidazole C2 scored highest against all the targets, with its aromatic rings including the two imidazole groups contributing to the binding. Among the phenyl-substituted 1H-imidazoles, C9 scored highest against all targets. C11 scored highest against Spro and C12 against Mpro and RdRp among the thiophene-imidazoles. The compounds interacted with HIS 41 - CYS 145 and GLU 288 – ASP 289 – GLU 290 of Mpro, ASN 501 of Spro receptor binding motif and some active site amino acids of RdRp. These novel imidazole compounds could be further developed as drug candidates against SARS-CoV-2 following lead optimization and experimental studies.

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1. Introduction

SARS-CoV-2 (severe acute respiratory syndrome - coronavirus 2) has since been declared a pandemic by the World Health Organization (WHO) and it requires immediate health intervention.^{1,2}

Presently, there are limited effective treatment options and targeted therapeutics approved for the clinical management of SARS-CoV-2.^{3,4} SARS-CoV-2 is a member of Beta-coronaviruses which is similar to Severe Acute Respiratory Syndrome Human coronavirus (SARS-CoV-1) and the Middle-East Respiratory Syndrome Human coronavirus (MERS CoV).⁵ Coronavirus is a positive-sense, single-stranded RNA virus comprising of over 30,000 nucleotides. The replication and transcription of the viral genome require the replicase gene responsible for encoding overlapping polyproteins, pp1a, and pp1ab.⁶ SARS-CoV-2 attacks the respiratory tract and has become a lethal viral respiratory disease with symptoms ranging

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from breathing difficulties, sore throat, high fever, diarrhea, cough to multiple organ failure and ultimately death.⁷

The infection occurs by the binding of SARS-CoV-2 spike protein (Spro) to the Angiotensin-Converting Enzyme-2 (ACE-2) in the alveoli of the host.⁸ Upon entry, viral RNA translation results in the synthesis of polyproteins responsible for the production of new virions from single-stranded RNA with the aid of RNA-dependent RNA polymerase (RdRp). These proteins are believed to represent primary druggable targets to contrast SARS-CoV-2 growth and replication.^{9–11} Analysis of several viruses have also shown that the viral protease is a common target for antiviral drug development,^{12–14} hence, the main protease (Mpro) of coronavirus could also be targeted due to its role in viral replication.^{15,16} Development of small molecule inhibitors of these protein target could therefore offer an effective treatment options against SARS-CoV-2.

Traditional methods of developing and discovering new therapeutic agents require rigorous scientific procedures, which consume time and cost. Computational tools, however, offer considerable promise for determining the affinities of small drug-like molecules for protein targets.¹⁷ This approach is a viable option for the designing and development of new drugs of biomedical interest.

Imidazoles are heterocyclic compounds with biological potency and proven pharmacological properties that include anti-parasitic,^{18–20} anti-inflammatory,²¹ anticancer,²² antifungal,²³ antibacterial,²⁴ antimalarial,²⁵ antitubercular,²⁶ and antiviral.²⁷ They are also reported to be carboxypeptidase inhibitors, β -lactamase inhibitors, heme oxygenase inhibitors and 20-HETE synthase inhibitors.^{18,19,28,29} The Imidazole ring is a nitrogen-containing heterocyclic aromatic molecule with different synthetic strategies and pharmaceutical importance.^{30,31} Thus, the incorporation of imidazole nucleus and electron-rich imidazole ring have become essential in the design, formulation, and development of imidazole-based drugs by pharmaceutical industries. The antiproliferative properties of imidazole derivatives have encouraged their use and synthesis for antiviral research.²² Their ability to attack enzymes and/or proteins critical to the viral life cycle is also a major contributing factor to their antiviral properties.^{32,33}

In a previous study, we reported the synthesis and *in vitro* anti-parasitic activities of a series of imidazole derivatives.¹⁹ Owing to the urgency for therapeutic intervention against the coronavirus, we employed the computational approach for evaluating the therapeutic potential of these imidazole compounds against SARS-CoV-2.

2. Materials and methods

2.1. Imidazole derivatives

The test compounds which are mainly imidazole derivatives (Fig. 1) were synthesized and characterized as previously described.^{19,34,35} Compounds C1 to C5 are bisimidazoles, C6 to C10 are phenyl-substituted 1H-imidazoles and C11 to C14 are thiophene-imidazoles.

2.2. Ligand preparation

The canonical SMILES of compounds C1 to C14 were converted to PDB format using Chimera 1.14 while the structure data file (SDF) format of standard ligands: Benzyl (Z, 4S)-4-[[[(2S)-4-methyl-2-[[[(2S)-3-methyl-2-[[[(2S)-2-[(5-methyl-1,2-oxazole-3-carbonyl)amino]propanoyl]amino]butanoyl]amino]pentanoyl]amino]-5-[(3S)-2-oxopyrrolidin-3-yl]pent-2-enoate (inhibitor N3), Pravastatin and remdesivir were obtained from PubChem database. The SDF format of compounds and standard ligands were uploaded to PyRx software and converted to PDBQT format using the OpenBabel

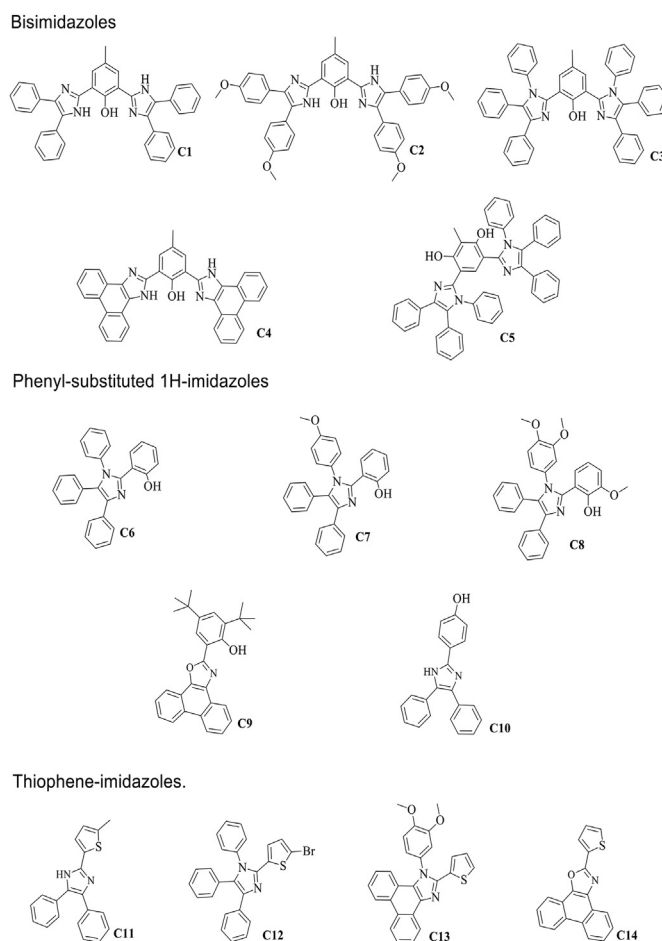


Fig. 1. Structures of imidazole derivatives.

plugin. The output files were minimized to obtain the minimum energy for the ligand docking.

2.3. Protein preparation

The crystal structures of the SARS-CoV-2 target proteins were obtained from the RCSB protein data bank (PDB). Main protease (Mpro: 6LU7) was in complex with inhibitor N3, obtained through X-RAY diffraction method, with a resolution 2.16 Å, R-Value free 0.235, R-Value work 0.202 and R-Value observed 0.204.³⁶ Spike receptor-binding domain in complex with its receptor ACE2 (Spro: 6LZG) was through X-RAY diffraction, resolution 2.50 Å, R-Value free 0.216, R-Value work 0.188 and R-Value observed 0.190.³⁷ RNA-dependent RNA polymerase in complex with cofactors (RdRp: 6M71) was obtained through electron microscopy with a Resolution of 2.90 Å.³⁸

The PDB format of the structures were uploaded to Chimera 1.14 workspace and the non-standard residues including ions, water and bounded ligands were first removed. The proteins were structurally minimized at 100 steepest descent steps, 0.02 steepest descent steps size (Å), 10 conjugate gradient steps, 0.02 conjugate gradient steps size (Å), and 10 update intervals, using the structure editing wizard Chimera 1.14. Furthermore, solvents were removed, hydrogen bonds were added, charges were assigned using Gas-teiger force field and histidine was set for the protonation state. Every available selenomethionine (MSE) were changed to methionine (MET), bromo-UMP (5BU) to UMP (U), methylselenyl-dUMP (UMS)

to UMP (U) and methylselenyl-dCMP (CSL) to CMP (C). The prepared proteins were uploaded to the PyRx software for molecular docking analysis.

2.4. Molecular docking

Molecular docking of the prepared ligands and proteins were performed using AutoDock vina in the PyRx workspace. Grid space was set by targeting important amino acid residues selected through literature³⁹ and from UniProtKB. Grid box size $x = 52.07 \text{ \AA}$, $y = 65.24 \text{ \AA}$ and $z = 58.07 \text{ \AA}$ and grid centre dimensions $x = -22.94$, $y = 14.30$, $z = 58.65$ were set for Mpro: 6LU7; grid box size $x = 43.86 \text{ \AA}$, $y = 46.19 \text{ \AA}$ and $z = 58.59 \text{ \AA}$ and grid center dimensions $x = -32.42$, $y = 30.30$, $z = 22.14$ for Spro: 6LZG; and $x = 78.79 \text{ \AA}$, $y = 83.87 \text{ \AA}$, $z = 84.28 \text{ \AA}$ and $x = 121.71$, $y = 122.39$, $z = 113.69$ respectively for RdRp: 6M71. The output files were uploaded to Chimera 1.14 workspace for post docking analysis and preparation of the 3D views of the protein-ligand complex. The 2D views of the molecular interactions were generated using UCSF Chimera 1.14 and Discovery Studio 2020.

2.5. Binding free energy calculation

The binding free energy of the protein-ligand complexes was employed to determine the stability of their complexes via Prime MM-GBSA program (Schrödinger suite version 20,018–4). Beforehand, the imidazole derivatives were prepared by ligprep, while the respective proteins were prepared using the protein preparation wizard, methods as previously described.⁴⁰ The active sites of the proteins were predicted by sitemap. Subsequently, the compounds were docked with proteins using glide extra precision (XP) docking. The Prime MM-GBSA panel was used to calculate binding free energy for ligand–protein complexes using the MM-GBSA technology available with Prime.⁴¹ OPLS3 force field was selected and VSGB was used as the continuum solvent model. Other options were set as default.

2.6. Receptor-ligand complex pharmacophore modelling

The highest-ranking compound based on binding affinity against the target proteins was selected to develop a receptor-ligand complex pharmacophore model using the PHASE module of Schrödinger suite. The auto (E-pharmacophore) method was used to generate ligand-based pharmacophore hypotheses. The maximum number of features to be generated was set at 7, minimum feature–feature distance was at 2.00, minimum feature–feature distance for feature of the same type at 4.00 and donors as vectors.

2.7. ADMET predictions

The absorption, distribution, metabolism, excretion and toxicity (ADMET) properties (lipophilicity ($\log P_{o/w}$), water solubility ($\log S$), druglikeness, bioavailability score, pharmacokinetics and toxicity profile) of the test compounds were determined using in silico integrative model predictions at the SwissADME, ADMETSar and ProTox-II online servers. Lipophilicity was measured using the partition coefficient between n-octanol and water ($\log P_{o/w}$) according to XLOGP, WLOGP, MLOGP, iLOGP and SILICOS-IT predictive models. The arithmetic mean of the values predicted by the five models is the consensus $\log P$.⁴² Water solubility was estimated as the logarithm of the molar solubility in water ($\log S$) using the SILICOS-IT predictive model.⁴² Druglikeness was according to the rule-based filters namely – Lipinski and Verber.^{43,44} Pharmacokinetic properties predicted include: skin permeation, gastrointestinal (GI) absorption permeation, blood–brain (BBB) permeation,

substrate and inhibitor of permeability glycoprotein (P-gp) and cytochrome p450 (CYP) respectively. Toxicity properties considered are carcinogenicity, eye corrosion, eye irritation, Ames mutagenesis and hepatotoxicity.

3. Results

3.1. Binding affinities and stability of test compounds with SARS-CoV-2 drug targets

The compounds exhibited various levels of binding affinities with Gibbs free energy (ΔG kcal/mol) ranging from -10.8 to -6.5 for Mpro (6LU7), -10.2 to -6.6 for Spro (6LZG) and -11.4 to -6.3 for RdRp (6M71). Among the three classes of imidazole derivatives, the bisimidazole compound C2 exhibits the highest binding affinity for all the drug targets (-10.8 , -10.2 , -11.4 kcal/mol for Mpro, Spro and RdRp respectively). Among the phenyl-substituted 1H-imidazoles, C9 has the highest binding affinity for Mpro (-8.0 kcal/mol), Spro (-7.4 kcal/mol) and RdRp (-7.6 kcal/mol). For the thiophene-imidazoles, C11 exhibits the highest binding affinity for Spro (-7.1 kcal/mol) and C12 for Mpro (-7.7 kcal/mol) and RdRp (-7.8 kcal/mol). Many of the test compounds including the 5 bisimidazoles gave binding affinities higher than those of the standard inhibitors (Table 1). The estimated binding free energy of the imidazole derivatives with the protein targets are shown in Table 2. The major energy contributors to the compounds binding were non-polar salvation terms (ΔG_{lip}), van der Waals (ΔG_{vdW}), and covalent energy ($\Delta G_{\text{covalent}}$).

3.2. Pharmacophore modelling

The pharmacophore models of the highest affinity compound (bisimidazole C2) are shown on Fig. 2. The models showed that the two imidazole rings of the compound contributed to its binding affinity for the protein targets. Apart from forming two aromatic interactions with the three target proteins, one of the imidazole rings of the compound acts as a hydrogen bond donor to Spro and

Table 1
Binding affinities (ΔG in kcal/mol) of test compounds for SARS-CoV-2 drug targets.

Compounds	ΔG Energy (Kcal/mol)		
	Mpro (6LU7)	Spro (6LZG)	RdRp (6M71)
Standard ligands			
N3	-7.4		
Pravastatin		6.3	
Remdesivir			-7.4
Bisimidazoles			
C1	-8.6	-8.6	-9.1
C2	-10.8	-10.2	-11.4
C3	-7.6	-7.9	-9
C4	-10.3	-10	-8.4
C5	-7.9	-6.7	-8.2
Phenyl-substituted 1H-imidazoles			
C6	-7.4	-7.2	-7.2
C7	-7.1	-6.9	-6.8
C8	-6.9	-6.6	-6.9
C9	-8	-7.4	-7.6
C10	-7.8	-7.2	-7.3
Thiophene-imidazoles			
C11	-6.8	-7.1	-6.8
C12	-7.7	-7.0	-7.8
C13	-6.5	-6.8	-7
C14	-6.9	-6.9	-6.3

Table 2
Binding free energy of the imidazole derivatives bound to proteins as calculated by MM-GBSA.

Compounds	ΔG_{bind} Energy (Kcal/mol)		
	Mpro (6LU7)	Spro (6LZG)	RdRp (6M71)
C1	-53.26	-31.18	-76.23
C2	-64.23	-35.30	-45.43
C3	-41.05	-37.42	-27.45
C4	-33.56	-29.43	-14.30
C5	-26.56	-37.36	-45.76
C6	-42.42	-17.04	-33.47
C7	-36.17	-44.98	-21.54
C8	-53.29	-29.70	-17.25
C9	-41.07	-28.97	-57.22
C10	-36.17	-58.63	-56.55
C11	-44.88	-50.772	-25.32
C12	-38.07	-29.270	-67.25
C13	-39.78	-28.097	-53.56
C14	-36.58	-32.47	-38.63

RdRp. The remaining five aromatic rings of Mpro and four of Spro and RdRp are all involved in the receptor-ligand binding.

3.3. ADMET profile

Table 3 shows the SwissADME predicted lipophilicity, water solubility, druglikeness and bioavailability scores of the compounds. The bisimidazole compounds have Log P values as high as 9.03 (C3) and 8.19 (C5), the least value being 4.37 (C2). The Log P values of the phenyl-substituted 1H-imidazoles ranged between 4.3 and 5.87 while those of the thiophene-imidazoles ranged between 4.02 and 6.15. The bisimidazoles C2 and C4 are moderately soluble while the remaining three are poorly soluble. The other two classes of imidazole compounds are either moderately soluble or soluble. For the druglikeness prediction, apart from C2 which violates only one rule, each of the bisimidazoles violates two Lipinski rules, while the remaining two classes of compounds violate only one rule. Besides, all the compounds pass the Veber's rule. For the bioavailability prediction, the bisimidazoles apart from C2 have a score of 0.17 while the remaining compounds score 0.55.

Table 4 shows the result of pharmacokinetics prediction of the test compounds. As highlighted in the table, skin permeation values (log Kp in cm/s) of the test compounds ranged from -6.8 (least permeant) to -1.02 (most permeant). Bisimidazoles C3 and C5 are the most skin permeant of all the compounds. All the test compounds apart from C3 and C5 possess high GI absorption potential and 8 of the compounds (1 bisimidazole, 4 phenyl-substituted 1H-

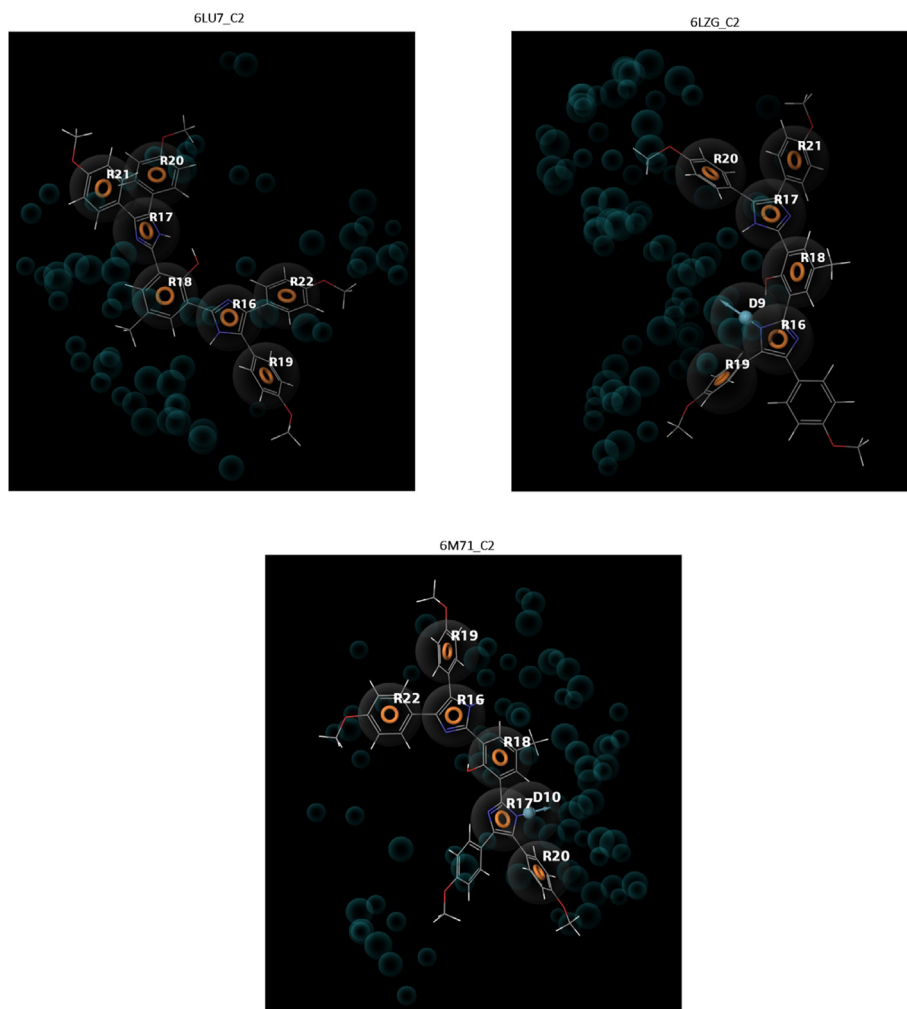


Fig. 2. Pharmacophore models of Bisimidazole (C2) on SARS-COV-2 main protease (6LU7), Spike protein (6LZG) and RNA-dependent RNA polymerase (6M71). R = Aromatic ring, D = Hydrogen bond donor.

Table 3
Predicted Lipophilicity (Log P), Water solubility (Log Sw), Druglikeness and Bioavailability scores of test compounds.

Compounds	Molecular Weight	Consensus Log P	Log Sw (Silicos-IT)	Solubility Class	Lipinski #violations	Veber #violations	Bioavailability Score
C1	582.95	6.23	-6.39	Poorly soluble	2	0	0.17
C2	703.05	4.37	-5.81	Moderately soluble	1	0	0.55
C3	747.23	9.06	-7.59	Poorly soluble	2	0	0.17
C4	578.91	5.5	-5.46	Moderately soluble	2	0	0.17
C5	763.23	8.19	-6.76	Poorly soluble	2	0	0.17
C6	416.68	5.34	-4.25	Moderately soluble	1	0	0.55
C7	446.71	5.01	-4.12	Moderately soluble	1	0	0.55
C8	506.76	4.3	-3.85	Soluble	1	0	0.55
C9	389.61	4.8	-3.76	Soluble	1	0	0.55
C10	445.72	5.87	-4.68	Moderately soluble	1	0	0.55
C11	336.58	4.43	-4.11	Moderately soluble	1	0	0.55
C12	483.59	6.15	-5.17	Moderately soluble	1	0	0.55
C13	462.73	4.51	-3.89	Soluble	1	0	0.55
C14	321.52	4.02	-3.18	Soluble	1	0	0.55

Table 4
Pharmacokinetics prediction output of test compounds.

Compounds	GI absorption	Blood–brain permeant	Pgp substrate	CYP1A2 inhibitor	CYP2C19 inhibitor	CYP2C9 inhibitor	CYP2D6 inhibitor	CYP3A4 inhibitor	Skin permeation log Kp (cm/s)
C1	High	No	No	No	No	No	No	No	-3.25
C2	High	No	No	No	No	No	No	No	-6.8
C3	Low	No	Yes	No	No	No	No	Yes	-1.02
C4	High	Yes	No	No	No	No	No	No	-3.83
C5	Low	No	Yes	No	No	No	No	Yes	-1.8
C6	High	Yes	No	No	No	No	No	No	-3.53
C7	High	Yes	No	No	No	No	No	No	-4.15
C8	High	Yes	No	No	No	No	No	No	-5.27
C9	High	Yes	No	No	No	No	No	No	-3.93
C10	High	No	No	No	No	No	No	No	-3.25
C11	High	Yes	No	No	No	Yes	No	No	-4.23
C12	High	No	No	No	No	No	No	No	-3.16
C13	High	Yes	Yes	No	No	No	No	No	-4.74
C14	High	Yes	No	Yes	Yes	Yes	No	No	-4.54

imidazoles and 3 thiophene-imidazoles) displayed the ability to penetrate the blood–brain barrier. Compounds C3, C5 and C13 are substrates of Pgp. C3 and C5 were predicted to be inhibitors of CYP3A4, C11 an inhibitor of CYP2C9 and C14 an inhibitor of CYP1A2, CYP2C19, and CYP2C9. As shown on Table 5, none of the test compounds has the tendency for carcinogenicity, mutagenesis, hepatotoxicity, eye corrosion, eye irritation and human either-a-go-go (hERG) inhibition.

Table 5
Toxicity profiles of imidazole molecules.

Compounds	Carcinogenicity	Eye corrosion	Eye irritation	Ames mutagenesis	human either-a-go-go inhibition	Hepatotoxicity
C1	–	–	–	–	–	–
C2	–	–	–	–	–	–
C3	–	–	–	–	–	–
C4	–	–	–	–	–	–
C5	–	–	–	–	–	–
C6	–	–	–	–	–	–
C7	–	–	–	–	–	–
C8	–	–	–	–	–	–
C9	–	–	–	–	–	–
C10	–	–	–	–	–	–
C11	–	–	–	–	–	–
C12	–	–	–	–	–	–
C13	–	–	–	–	–	–
C14	–	–	–	–	–	–

(–) = Inactive.

3.4. Post docking analysis of selected compounds

Figs. 3–5 show the three-dimensional (3D) and two-dimensional (2D) structures of the SARS-CoV-2 target proteins in complex with the standard inhibitors and the highest affinity compounds from each of the classes (bisimidazole C2 and phenyl-substituted 1H-imidazole C9 for Mpro, Spro and RdRp; thiophene-imidazole C11 for Spro and C12 with Mpro and RdRp).

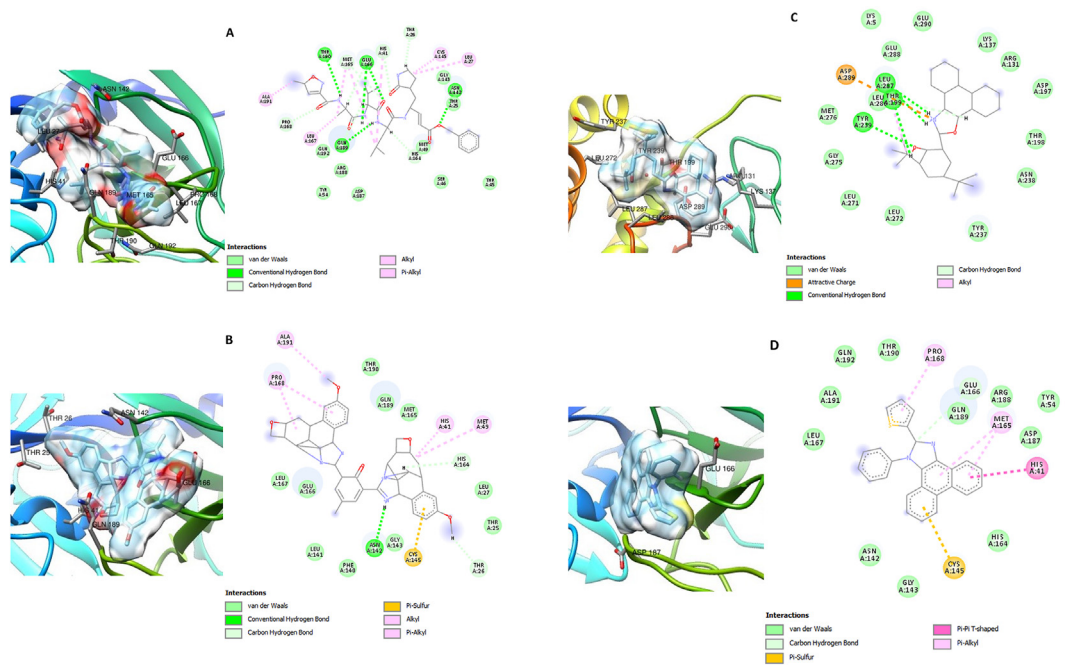


Fig. 3. 3D (left) and 2D (right) views of the molecular interactions of amino-acid residues of Mpro (6LU7) with (A) N3 (B) C2 (C) C9 (D) C12.

C2 and C12 interacted with ASN 142, GLU 166, CYS 145, HIS 41 and other amino acid residues at the inhibitor (N3) binding site of SARS-CoV-2 Mpro. C9 interacted with residues like GLU 288, ASP 289 and GLU 290 at a different binding pocket on Mpro (Fig. 3). The SARS-CoV-2 Spro formed a complex with C2, C9, C11 and Pravastatin through various types of interactions with some amino acids residues of its receptor binding domain (RBD) (Fig. 4). ASN 501 of Spro formed van der Waals interaction with C2 and C9 (Fig. 3 B and C) and hydrogen bond interaction with Pravastatin and C11 (Fig. 3 A and D). Furthermore, C2, C9, C12 and remdesivir interacted with some amino acid residues at the binding pocket of SARS-CoV-2

RdRp including: ASP 618, TYR 619, ASP 760, ASP 761, TRP 800, GLU 811, CYS 813, and SER 814 in (Fig. 5).

4. Discussion

The novel coronavirus is a global health challenge that calls for emergency and urgent antiviral strategies for the treatment and prevention of the infection. Some antiviral agents including remdesivir were currently repurposed against COVID-19 to allow rapid availability of treatments options^{3,4} as the search for new anti-SARS-CoV-2 drugs continues. The use of heterocyclic

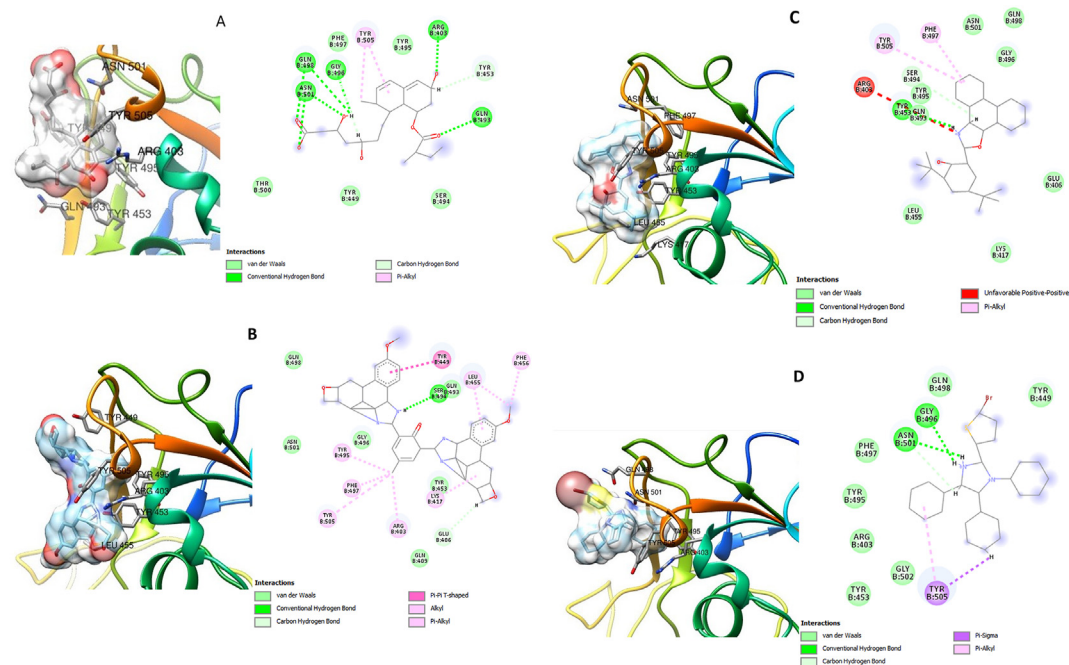


Fig. 4. 3D (left) and 2D (right) views of the molecular interactions of amino-acid residues of Spro (6LZG) with (A) Pravastatin (B) C2 (C) C9 (D) C11.

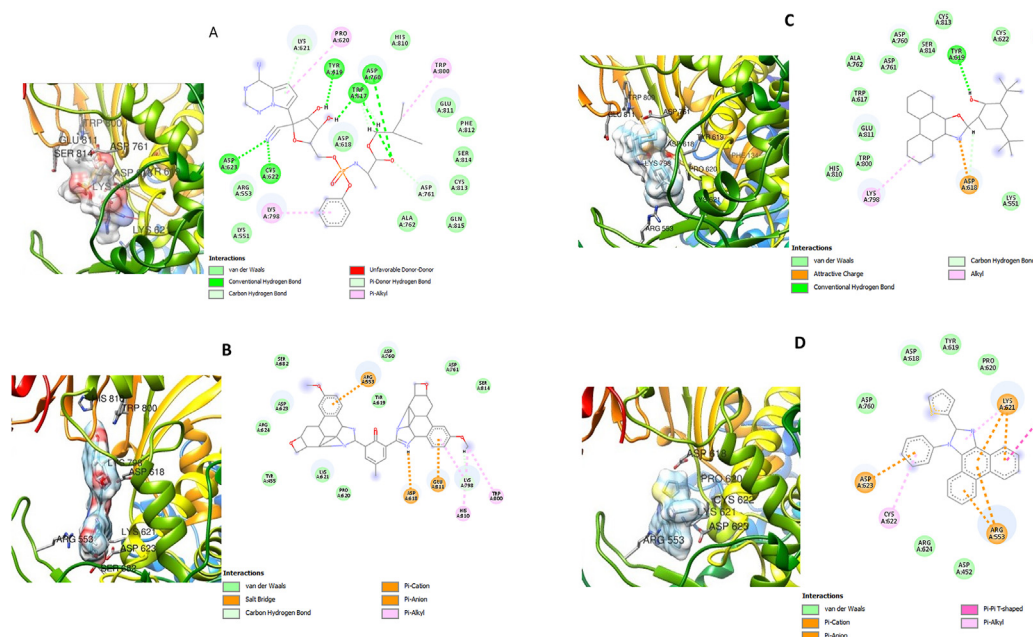


Fig. 5. 3D (left) and 2D (right) views of the molecular interactions of amino-acid residues of RdRp (6M71) with (A) Remdesivir (B) C2 (C) C19 (D) C12.

compounds as templates for antiviral drug development has become very beneficial as they have been reported to be potent against several viral infections including hepatitis C (HCV), hepatitis B (HBV), human immunodeficiency virus (HIV), herpes simplex (HSV), rotavirus, adenoviruses and coxsackie.^{27,32,33} Antiviral imidazole compounds are said to exhibit their anti-viral activity by targeting viral proteases and other critical enzymes/proteins essential to the viral life cycle.^{32,33}

The high binding affinities exhibited by the imidazole compounds for SARS-CoV-2 target proteins in this study is an indication of the inhibitory potential of these compounds against these biomolecules and their possible roles as therapeutic agents against SARS-CoV-2. Due to structural similarity with the amino acid histidine, an imidazole can easily interact with important protein molecules thereby modifying their functions.^{32,33} In this study, it was observed that the bisimidazoles possess higher binding affinity for all the target proteins than the other two classes of imidazoles. This could be linked to the two imidazole rings and several other aromatic rings contained in the structure of these compounds. From the result of the pharmacophore modelling of C2 (the highest affinity compound), it was observed that the two imidazole rings together with the other aromatic groups contributed to the binding of this compound to the targets. Interactions involving aromatic rings are very essential to biological recognition including protein-ligand interaction as about 20% of amino acids are aromatic in nature. Aromatic interactions are known to be of paramount importance in drug design for improved efficacy and lead optimization.⁴⁵

The binding free energy (Table 2) of the complexes agrees with the docking score shown on Table 1. Determination of protein-ligand stability is regarded as a very reliable method for validating docking score^{9,41} and the free energy of favorable reaction is always negative. The results obtained in this study showed that the imidazole derivatives formed a stable complex with the drug targets upon binding, affirming the robustness of the docking results.

In addition to the inhibitory potential demonstrated by the imidazole compounds, quite a number of them possessed moderate ADMET properties. However, some of the compounds may require lead optimization to improve these properties and still maintain

the binding affinities. ADMET analysis is a procedure used for defining whether the compounds can be easily absorbed, transported to their target site of action, metabolized in a way that does not instantly remove the activity and easily eliminated from the body while preventing toxic effects. These properties are collectively referred to as ADMET (absorption, distribution, metabolism, elimination and toxicity).⁴⁶ In silico prediction of ADMET properties is a cheap and time saving alternative to standard experimental approaches^{47,48} and its inclusion at earlier stages of drug discovery programs^{49,50} has become essential^{51–53} for reducing the rate of pharmacokinetics-related failure of drugs in the clinical phases.

Lipophilicity and water solubility are critical physicochemical properties that determines the ADMET behaviors of a drug. An orally administered drug should be sufficiently lipophilic to pass through the intestinal lining, penetrate the membrane of target cells and sufficiently hydrophilic to travel in the aqueous blood. The higher the log P value of a compound, the higher its lipophilicity and the lower its water solubility.⁴¹ Hence, the compounds with Log P values between 4.02 and 6.15 (all the phenyl substituted imidazoles, the thiophene imidazoles and bisimidazoles C2 and C4) came out to be either soluble or moderately soluble while the bisimidazoles C1, C3 and C5 with higher log P values were predicted to be poorly soluble (Table 3). The presence of more aromatic rings and the higher molecular weights of the bisimidazoles may have contributed to the poor solubility of these compounds, however the ones possessing additional polar side chains (C2 and C4) are moderately soluble.

Drug-likeness analysis is a qualitative assessment of oral bioavailability and it is developed based on structural or physicochemical evaluation of compounds in the advanced stages of drug development.⁴² The Lipinski's Rule states that an orally active drug should have: not more than 5 hydrogen bond donors, not more than 10 hydrogen bond acceptors, a molecular weight under 500 g/mol and a log P less than 5.⁴³ A molecule would not be orally active if it violates two or more of the rules. Based on these criteria, only C2 meet the requirements for oral bioavailability among the bisimidazoles as it violates only one of the rules, but all phenyl substituted and thiophene imidazoles meet the requirements. Nonetheless, all the test compounds pass the Veber's rule comprising of only two criteria of: 10 or fewer rotatable bonds and

polar surface (TPSA) area not greater than 140 Å².⁴⁴ Furthermore, the bioavailability score which uses total charge, TPSA and the Lipinski filter⁵⁴ gave a semi-quantitative estimate of the probability of the compounds to be good oral drugs. The 0.17 bioavailability score of the bisimidazoles shows that these compounds have approximately 17% probability of at least 10% oral bioavailability in rat or measurable human colon carcinoma (Caco-2) permeability, hence these compounds apart from C2 will not be easily bioavailable orally. However, 0.55% bioavailability score of C2 and the remaining 2 classes of test compounds represents a 55% probability, hence, these compounds are likely to be good as oral drugs.

According to the pharmacokinetic predictions of the test compounds, the bisimidazoles C3 and C5 and the thiophene imidazoles C11 and C14 could possibly cause drug–drug interactions as CYP inhibitors. Cytochrome P450 (CYP) is a superfamily of isoenzymes that catalyzes many reactions in the phase I of drug metabolism.⁵⁵ It has been estimated that 50–90% of drugs are substrates of five major isoforms (CYP1A2, CYP2C19, CYP2C9, CYP2D6, and CYP3A4)⁴² and their inhibition is a major cause of pharmacokinetics-related drug–drug interactions.^{56,57} Likewise, as substrates of Pgp, compounds C3, C5 and C13 are likely to be prevented from entering their target site of action. Pgp is an important member of the ATP-binding cassette transporters and it is responsible for the active efflux of xenobiotics through biological membranes for the purpose of protecting the body from foreign chemicals. This efflux pump also contributes to drug resistance by limiting the entry of some drugs into the sensitive areas. Nevertheless, the results of the toxicity prediction showed that none of the test compounds has the tendency for any of the toxicity parameters tested, hence the compounds could be both effective and safe as potential therapeutic agents against SARS-CoV-2.

Analysis of the 3D and 2D structures of the docked SARS-CoV-2 target–imidazole complex showed that these compounds possess inhibitory potentials against the proteins. C2, C12 and the standard inhibitor N3 interacted with CYS 145 and HIS 41 at the binding pocket of the SARS-CoV-2 Mpro while C9 interacted with GLU 288, ASP 289 and Glu 290 (Fig. 3). CYS 145 and HIS 41 are very important amino acids at the catalytic site of the main protease of SARS coronavirus. The protease is a homodimer with the two subunits arranged perpendicularly to each other and each protomer have three discrete structural domains. The substrate binding site of each subunit comprises of a HIS 41–CYS 145 catalytic dyad situated at the interface between two of the structural domains. While HIS 41 acts as a general base, CYS 145 mediates an electrostatic trigger that initiates the first step of the catalytic mechanism of action of the enzyme.⁵⁸ Mutations at these catalytic structure (HIS 41 - CYS 145) have been reported to almost totally eliminate Mpro enzyme activity.^{59,60} Residues around GLU 288 – ASP 289 – GLU 290 are reported to be associated with Mpro dimerization and mutations in GLU 288 and ASP 289 gave rise to enzymes with reduced activities.⁶¹ Hence, interaction of the compounds with these amino acids suggests that these compounds could possibly interfere with the catalytic activity of SARS-CoV-2 main protease and consequently inhibit the replication of the virus.

The SARS-CoV-2 Spro interacted with C2, C9, C11 and Pravastatin through some important amino acids residues including ASN 501 of its receptor binding domain (RBD) (Fig. 4). ASN 501 has been listed as one of the key residues responsible for binding of Spro to its human host receptor (ACE2). ASN 501 of the spike receptor binding motif is reported to form hydrogen bond with Tyr 41 of ACE2. ASN 501 also interacts with ACE2 Lys353, Gly354 and Asp355.^{36,62} Binding of the test compounds to this critical amino acid residue could therefore limit the binding of SARS-CoV-2 Spro to its human host receptor thereby limiting viral entry and disease progression.

Furthermore, C2, C9 and C12 demonstrated inhibitory potentials against SARS-CoV-2 RdRp. These compounds interacted with the same amino acid residues as Remdesivir (a known RdRp inhibitor) in the binding pocket of SARS-CoV-2 RdRp (Fig. 5). The four compounds interacted with residues ASP 618, TYR 619, ASP 760 and other amino acids that have been previously identified at the enzyme's active site.⁶³ RdRp as a crucial enzyme in viral replication, catalyzes the synthesis of the RNA genome by producing a complementary RNA strand from an RNA template,⁶⁴ hence its inhibition by these imidazole compounds could offer therapeutic benefits against SARS-CoV-2.

5. Conclusions

Three classes of newly synthesized imidazole derivatives were evaluated for therapeutic potentials against SARS-CoV-2 Mpro, Spro and RdRp. The compounds showed exciting binding affinities and stability with the target proteins. Bisimidazole C2 gave the highest binding affinity for all the drug targets among the three classes of compounds. Within the phenyl-substituted 1H-imidazole class, C9 gave the highest docking score against the three targets. For the thiophene-imidazoles, C11 scored highest against Spro and C12 against Mpro and RdRp. The compounds interacted with HIS 41 - CYS 145 and GLU 288 – ASP 289 – GLU 290 of Mpro, ASN 501 of Spro receptor binding motif and some active site amino acids of RdRp. These novel imidazoles could offer therapeutic benefits against SARS-CoV-2, however, lead optimization may be required to modify the ADMET properties of some of the compounds while maintaining their biological activities and wet laboratory experiments will be necessary to further validate the results of this in silico study.

Declaration of competing interest

The authors declare no conflicts of interest.

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References

- Hsu LY, Chia PY, Lim JF. The novel coronavirus (SARS-CoV-2) pandemic. *Ann Acad Med Singapore*. 2020;49(105):105–107.
- Jebri N. World Health Organization declared a pandemic public health menace: a systematic review of the coronavirus disease 2019 “COVID-19”, up to 26th March 2020. 2020. <https://doi.org/10.2139/ssrn.3566298>. Available at: SSRN 3566298.
- Esakandari H, Nabi-Afjadi M, Fakkari-Afjadi J, Farahmandian N, Miresmaeili SM, Bahreini E. A comprehensive review of COVID-19 characteristics. *Biol Proced Online*. 2020;22:1–10. <https://doi.org/10.1186/s12575-020-00128-2>.
- Parlakpınar H, Polat S, Acet HA. Pharmacological agents under investigation in the treatment of coronavirus disease 2019 and the importance of melatonin. *Fundamental & cli. Pharm.* 2020 <https://doi.org/10.1111/fcp.12589>.
- Liya G, Yuguang W, Jian L, et al. Studies on viral pneumonia related to novel coronavirus SARS-CoV-2, SARS-CoV, and MERS-CoV: a literature review. *APMIS*. 2020;128:423–432. <https://doi.org/10.1111/apm.13047>.
- Gurung AB, Ali MA, Lee J, Farah MA, Al-Anazi KM. In silico screening of FDA approved drugs reveals ergotamine and dihydroergotamine as potential coronavirus main protease enzyme inhibitors. *Saudi J Biol Sci*. 2020;27(10):2674–2682. <https://doi.org/10.1016/j.sjbs.2020.06.005>.
- Han W, Quan B, Guo Y, et al. The course of clinical diagnosis and treatment of a case infected with coronavirus disease 2019. *J. of med. virology*. 2020;92(5):461–463. <https://doi.org/10.1002/jmv.25711>.

8. Verdecchia P, Cavallini C, Spanevello A, Angeli F. The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med.* 2020;14–20. <https://doi.org/10.1016/j.ejim.2020.04.037>.
9. Ullrich S, Nitsche C. The SARS-CoV-2 main protease as drug target. *Bioorg Med Chem Lett.* 2020;127377. <https://doi.org/10.1016/j.bmcl.2020.127377>.
10. Wu C, Liu Y, Yang Y, et al. Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods. *Acta Pharm Sin B.* 2020;10(5):766–788. <https://doi.org/10.1016/j.apsb.2020.02.008>.
11. Elekofehinti OO, Iwaloye O, Josiah SS, Lawal AO, Akinjiyan MO, Ariyo EO. Molecular docking studies, molecular dynamics and ADME/tox reveal therapeutic potentials of STOCKIN-69160 against papain-like protease of SARS-CoV-2. *Mol Divers.* 2020;1–13. <https://doi.org/10.1007/s11030-020-10151-w>.
12. Chang KO, Kim Y, Lovell S, Rathnayake AD, Groutas WC. Antiviral drug discovery: norovirus proteases and development of inhibitors. *Viruses.* 2019;11(2):197.
13. Mazzon M, Marsh M. Targeting viral entry as a strategy for broad-spectrum antivirals. *F1000Research.* 2019;8:1–10. <https://doi.org/10.12688/f1000research.19694.1>.
14. Ben-Shabat S, Yarmolinsky L, Porat D, Dahan A. Antiviral effect of phytochemicals from medicinal plants: applications and drug delivery strategies. *Drug Del. and Trans. Research.* 2020;1–14. <https://doi.org/10.1007/s13346-019-00691-6>.
15. Pillaiyar T, Meenakshisundaram S, Manickam M. Recent discovery and development of inhibitors targeting coronaviruses. *Drug Discov Today.* 2020;25(4):668–688. <https://doi.org/10.1016/j.drudis.2020.01.015>.
16. Elekofehinti OO, Iwaloye O, Famusiwa CD, Akinseye O, Rocha JB. Identification of main protease of coronavirus SARS-CoV-2 (Mpro) inhibitors from melissa officinalis". *Curr Drug Discov Technol.* 2020b;17:1. <https://doi.org/10.2174/157016381799920091810370>.
17. Hiramoto T, Nonaka Y, Inoue K, et al. Identification of endogenous surrogate ligands for human P2Y receptors through an in silico search. *J Pharmacol Sci.* 2004;95(1):81–93. <https://doi.org/10.1254/jphs.95.81>.
18. Adeyemi OS, Molina MT, Eseola AO, Fonseca-Berzal C, Gómez-Barrio A. New imidazole-based compounds active against Trypanosoma cruzi. *Comb Chem High Throughput Screen.* 2017;20(1):20–24.
19. Adeyemi OS, Eseola AO, Plass W, et al. Imidazole derivatives as antiparasitic agents and use of molecular modeling to investigate the structure–activity relationship. *Parasitol Res.* 2020a;119:1925–1941. <https://doi.org/10.1007/s00436-020-06668-6>.
20. de Araújo JS, Garcia-Rubia A, Sebastian-Perez V, et al. Imidazole derivatives as promising agents for the treatment of Chagas disease. *Antimicrob Agents Chemother.* 2019;63(4).
21. Raman N, Mitu L, Sakthivel A, Pandi MSS. Studies on DNA cleavage and antimicrobial screening of transition metal complexes of 4-aminoantipyrine derivatives of N 2 O 2 type. *J Iran Chem Soc.* 2009;6(4):738–748. <https://doi.org/10.1007/BF03246164>.
22. Sharma D, Narasimhan B, Kumar P, et al. Synthesis, antimicrobial and antiviral evaluation of substituted imidazole derivatives. *European j. of medicinal chem.* 2009;44(6):2347–2353 <https://doi.org/10.1016/j.ejmech.2008.08.010>.
23. Emami S, Foroumadi A, Falahati M, et al. 2-Hydroxyphenacyl azoles and related azolium derivatives as antifungal agents. *Bioorganic & med. chem. letters.* 2008;18(1):141–146. <https://doi.org/10.1016/j.bmcl.2007.10.111>.
24. Bhandari K, Srinivas N, Keshava GS, Shukla PK. Tetrahydronaphthyl azole oxime ethers: the conformationally rigid analogues of oxiconazole as antibacterials. *European j. of medicinal chem.* 2009;44(1):437–447 <https://doi.org/10.1016/j.ejmech.2008.01.006>.
25. Iman M, Davood A, Khamesipour A. Design of antimalarial agents based on pyrimidine derivatives as methionine aminopeptidase 1b inhibitor: molecular docking, quantitative structure activity relationships, and molecular dynamics simulation studies. *J Chin Chem Soc.* 2020;1–11. <https://doi.org/10.1002/jccs.201900165>.
26. Papadopoulou MV, Bloomer WD, Rosenzweig HS. The antitubercular activity of various nitro (triazole/imidazole)-based compounds. *Bioorg Med Chem.* 2017;25(21):6039–6048. <https://doi.org/10.1016/j.bmc.2017.09.037>.
27. Liu L, Hu Y, Shen YF, Wang GX, Zhu B. Evaluation on antiviral activity of coumarin derivatives against spring viraemia of carp virus in epithelioma papulosum cyprini cells. *Anti. Research.* 2017;144:173–185. <https://doi.org/10.1016/j.antiviral.2017.06.007>.
28. Eseola AO, Sun WH, Li W, Woods JA. Syntheses of new imidazole ligand series and evaluation of 1-, 2-and 4, 5-imidazole substituent electronic and steric effects on N-donor strengths. *J Mol Struct.* 2010;984(1–3):117–124. <https://doi.org/10.1016/j.molstruc.2010.09.015>.
29. Kumari A, Singh RK. Medicinal chemistry of indole derivatives: current to future therapeutic prospects. *Bioorg Chem.* 2019;89:103021. <https://doi.org/10.1016/j.bioorg.2019.103021>.
30. Zheng X, Ma Z, Zhang D. Synthesis of imidazole-based medicinal molecules utilizing the van leusen imidazole synthesis. *Pharmaceuticals.* 2020;13(3):37. <https://doi.org/10.3390/ph13030037>.
31. Kerru N, Gummidi L, Maddila S, Gangu KK, Jonnalagadda SB. A review on recent advances in nitrogen-containing molecules and their biological applications. *Molecules.* 2020;25(8):1909. <https://doi.org/10.3390/molecules25081909>.
32. De Clercq E. Antiviral drugs in current clinical use. *J. of clinical virology.* 2004;30(2):115–133. <https://doi.org/10.1016/j.jcv.2004.02.009>.
33. Kanwal A, Ahmad M, Aslam S, Naqvi SAR, Saif MJ. Recent advances in antiviral benzimidazole derivatives: a mini review. *Pharmaceut Chem J.* 2019;53(3):179–187. <https://doi.org/10.1007/s11094-019-01976-3>.
34. Adeyemi OS, Eseola AO, Plass W, Otuechere CA, Elebiyo TC. New imidazoles cause cellular toxicity by impairing redox balance, mitochondrial membrane potential, and modulation of HIF-1 α expression. *Biochem Biophys Res Commun.* 2020b;529(1):23–27. <https://doi.org/10.1016/j.bbrc.2020.05.059>.
35. Eseola AO, Göröls H, Woods JA, Plass W. Single monodentate N-donor ligands versus multi-ligand analogues in Pd (II)-catalysed CC coupling at reduced temperatures. *Polyhedron.* 2020;114507. <https://doi.org/10.1016/j.poly.2020.114507>.
36. Jin Z, Du X, Xu Y, et al. Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. *Nature.* 2020;582:289–293. <https://doi.org/10.1038/s41586-020-2223-y>.
37. Wang Q, Zhang Y, Wu L, et al. Structural and functional basis of SARS-CoV-2 entry by using human ACE2. *Cell.* 2020;181:894–904. <https://doi.org/10.1016/j.cell.2020.03.045>. e9.
38. Gao Y, Yan L, Huang Y, et al. Structure of the RNA-dependent RNA polymerase from COVID-19 virus. *Science.* 2020;368:779–782. <https://doi.org/10.1126/science.abb7498>.
39. Chang GG. *Quaternary structure of the SARS coronavirus main protease. Molecular biology of the SARS-coronavirus.* Berlin, Heidelberg: Springer; 2010:115–128. https://doi.org/10.1007/978-3-642-03683-5_8.
40. Iwaloye O, Elekofehinti OO, Oluwarotimi EA, Babatomiwa K, Momoh I. In silico molecular studies of natural compounds as possible anti-Alzheimer's agents: ligand-based design. *Network Modeling Analysis in Health Informatics and Bioinfo.* 2020a;9:54. <https://doi.org/10.1007/s13721-020-00262-77>.
41. *Schrödinger release 2020-2: Prime.* New York, NY: Schrödinger, LLC; 2020.
42. Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Sci Rep.* 2017;7:42717. <https://doi.org/10.1038/srep42717>.
43. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev.* 1997;23(1–3):3–25. [https://doi.org/10.1016/S0169-409X\(96\)00423-1](https://doi.org/10.1016/S0169-409X(96)00423-1).
44. Veber DF, Johnson SR, Cheng HY, Smith BR, Ward KW, Kopple KD. Molecular properties that influence the oral bioavailability of drug candidates. *J. of medicinal chem.* 2002;45(12):2615–2623 <https://doi.org/10.1021/jm020017n>.
45. Singh VP. Aromatic interactions in biological systems. *Sci Technol Jpn.* 2015;3:42–48.
46. Sun H, Li Y, Shen M, et al. Assessing the performance of the MM/PBSA and MM/GBSA methods. 5, improved docking performance using high solute dielectric constant MM/GBSA and MM/PBSA rescoring. *Phys Chem Chem Phys.* 2014;16:22035–22045. <https://doi.org/10.1039/C4CP03179B>.
47. Ntie-Kang F. An in silico evaluation of the ADMET profile of the StreptomeDB database. *SpringerPlus.* 2013;2(1):353. <https://doi.org/10.1186/2193-1801-2-353>.
48. DiMasi JA, Hansen RW, Grabowski HG. The price of innovation: new estimates of drug development costs. *J. of health economics.* 2003;22(2):151–185. [https://doi.org/10.1016/S0167-6296\(02\)00126-1](https://doi.org/10.1016/S0167-6296(02)00126-1).
49. Darvas F, Keseru G, Papp A, Dorman G, Urge L, Krajcsi P. In silico and ex silico ADME approaches for drug discovery. *Current topics in med. chem.* 2002;2(12):1287–1304 <https://doi.org/10.2174/1568026023392841>.
50. Hodgson J. ADMET—turning chemicals into drugs. *Nat Biotechnol.* 2001;19(8):722–726. <https://doi.org/10.1038/90761>.
51. Navia MA, Chaturvedi PR. Design principles for orally bioavailable drugs. *Drug Discov Today.* 1996;1(5):179–189. [https://doi.org/10.1016/1359-6446\(96\)10020-9](https://doi.org/10.1016/1359-6446(96)10020-9).
52. Lombardo F, Gifford E, Shalaeva MY. In silico ADME prediction: data, models, facts and myths. *Mini Rev Med Chem.* 2003;3(8):861–875. <https://doi.org/10.2174/1389557033487629>.
53. Gleeson PM, Hersey A, Hannongbua S. In-silico ADME models: a general assessment of their utility in drug discovery applications. *Current topics in medicinal chem.* 2011;11(4):358–381. <https://doi.org/10.2174/156802611794480927>.
54. Testa B, Kramer S. The biochemistry of drug metabolism—an introduction. *Chem Biodivers.* 2009;6(5):591. <https://doi.org/10.1002/cbdv.200900022>.
55. Hollenber PF. Characteristics and common properties of inhibitors, inducers, and activators of CYP enzymes. *Drug Metab Rev.* 2002;34(1–2):17–35. <https://doi.org/10.1081/DMR-120001387>.
56. Huang SM, Strong JM, Zhang L, et al. New era in drug interaction evaluation: US Food and Drug Administration update on CYP enzymes, transporters, and the guidance process. *The J. of cli. Pharm.* 2008;48(6):662–670 <https://doi.org/10.1177/0091270007312153>.
57. Huang C, Wei P, Fan K, Liu Y, Lai L. 3C-like proteinase from SARS coronavirus catalyzes substrate hydrolysis by a general base mechanism. *Biochemistry.* 2004;43(15):4568–4574. <https://doi.org/10.1021/bi036022q>.
58. Chang HP, Chou CY, Chang GG. Reversible unfolding of the severe acute respiratory syndrome coronavirus main protease in guanidinium chloride. *Biophys J.* 2007;92(4):1374–1383. <https://doi.org/10.1529/biophysj.106.091736>.
59. Lan J, Ge J, Yu J, et al. Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. *Nature.* 2020;581(7807):215–220. <https://doi.org/10.1038/s41586-020-2180-5>.
60. Yan R, Zhang Y, Li Y, Xia L, Guo Y, Zhou Q. Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2. *Science.* 2020;367(6485):1444–1448. <https://doi.org/10.1126/science.abb2762>.

61. Novak J, Rimac H, Kandagalla S, Pathak P, Grishina M, Potemkin V. *Proposition of a new allosteric binding site for potential SARS-CoV-2 3CL protease inhibitors by utilizing molecular dynamics simulations and ensemble docking*. 2020. <https://doi.org/10.21203/rs.3.rs-34002/v1>.
62. Aftab SO, Ghouri MZ, Masood MU, et al. Analysis of SARS-CoV-2 RNA-dependent RNA polymerase as a potential therapeutic drug target using a computational approach. *J translational med.* 2020;18(1):1–15 <https://doi.org/10.1186/s12967-020-02439-0>.
63. Wang Y, Anirudhan V, Du R, Cui Q, Rong L. RNA-dependent RNA polymerase of SARS-CoV-2 as a therapeutic target. *J Med Virol.* 2020:1–40. <https://doi.org/10.1002/jmv.26264>.
64. Martin YA. Bioavailability score. *J Med Chem.* 2005;48:3164–3170. <https://doi.org/10.1021/jm0492002>.