



# Near-Complete Genome Sequences of Five Siciniviruses from North America

 Iryna V. Goraichuk,<sup>a,b</sup> James F. Davis,<sup>c</sup>  D. Joshua Parris,<sup>a</sup>  Henry M. Kariithi,<sup>a,d</sup>  Claudio L. Afonso,<sup>a</sup>  David L. Suarez<sup>a</sup>

<sup>a</sup>Exotic and Emerging Avian Viral Disease Research Unit, Southeast Poultry Research Laboratory, U.S. National Poultry Research Center, ARS, USDA, Athens, Georgia, USA

<sup>b</sup>National Scientific Center Institute of Experimental and Clinical Veterinary Medicine, Kharkiv, Ukraine

<sup>c</sup>Georgia Poultry Laboratory Network, Gainesville, Georgia, USA

<sup>d</sup>Biotechnology Research Institute, Kenya Agricultural and Livestock Research Organization, Loresho, Nairobi, Kenya

**ABSTRACT** Here, we report near-complete genome sequences of sicinivirus from U.S. poultry flocks in 2003 to 2005 and Mexico in 2019. They show highest nucleotide identity (84.5 to 85.5%) with other members of the *Sicinivirus* genus. These sequences update knowledge on diversity and contribute to a better understanding of the molecular epidemiology of sicinivirus.

The *Picornaviridae* are a family of small, icosahedral viruses with single-stranded, positive-sense RNA genomes (1). The family comprises 47 genera, with 13 of these identified from avian species (1–3). Avian picornaviruses of the *Sicinivirus* genus have been identified in both healthy and diseased chickens, and the viruses' role in disease remains unclear (4–9). Here, we report the sequences of five near-complete genomes of sicinivirus from North America.

Fecal samples from broiler chickens with severe hypoglycemia were collected at commercial farms in the state of Georgia in 2003 to 2005, and bursal swab samples (clinical signs were not provided) were collected from a commercial farm in Mexico in 2019 and preserved on Flinders Technology Associates (FTA) cards (Whatman, USA) (Table 1). Feces were first diluted 3:7 in sterile phosphate-buffered saline and then centrifuged for 10 min at 3,200 rpm. The supernatants were passed sequentially through a 1.2- $\mu$ m- and a 0.45- $\mu$ m-pore-size filter (Merck Millipore, USA) to remove bacteria and large-cell-size particles and were DNase treated using the TURBO DNA-free kit (Ambion, USA) to remove host DNA. Total nucleic acids were isolated using the DNeasy blood and tissue kit (Qiagen, Germany). For the FTA card bursal swabs, first, nucleic acids were eluted from 24 3-mm discs punched out from the card by incubation in Tris-EDTA (TE) buffer, and then total RNA was extracted from the TE eluate using the MagMAX total RNA isolation kit (Thermo Fisher, USA). Sequence-independent single-primer amplification (10) was used to produce random amplicons. Briefly, first-strand cDNA was synthesized using the random octamer primer tagged with a fixed-sequence K-8N and SuperScript IV reverse transcriptase (Invitrogen, USA), followed by second-strand synthesis using Klenow polymerase (New England Biolabs, USA). Finally, random primer amplification was conducted using Phusion DNA polymerase (New England Biolabs) and the primer consisting of the fixed portion of the random K-8N primer. DNA libraries were subsequently prepared with the Nextera XT DNA library preparation kit (Illumina, USA). Paired-end (2  $\times$  150-bp) sequencing was performed on an Illumina MiSeq instrument. A customized workflow on the Galaxy platform (11) was used to perform preprocessing and assembly of the raw sequencing reads, as described previously (12, 13). Briefly, raw read quality was assessed using FastQC v0.63 (14), and residual adapter sequences were trimmed using Cutadapt v1.6 (15). Sequence data were assembled *de novo* utilizing MIRA3 v0.0.1 (16). Default parameters were used for all software. The MiSeq runs generated 1,096,430 to 2,446,408

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Address correspondence to David L. Suarez, david.suarez@usda.gov.

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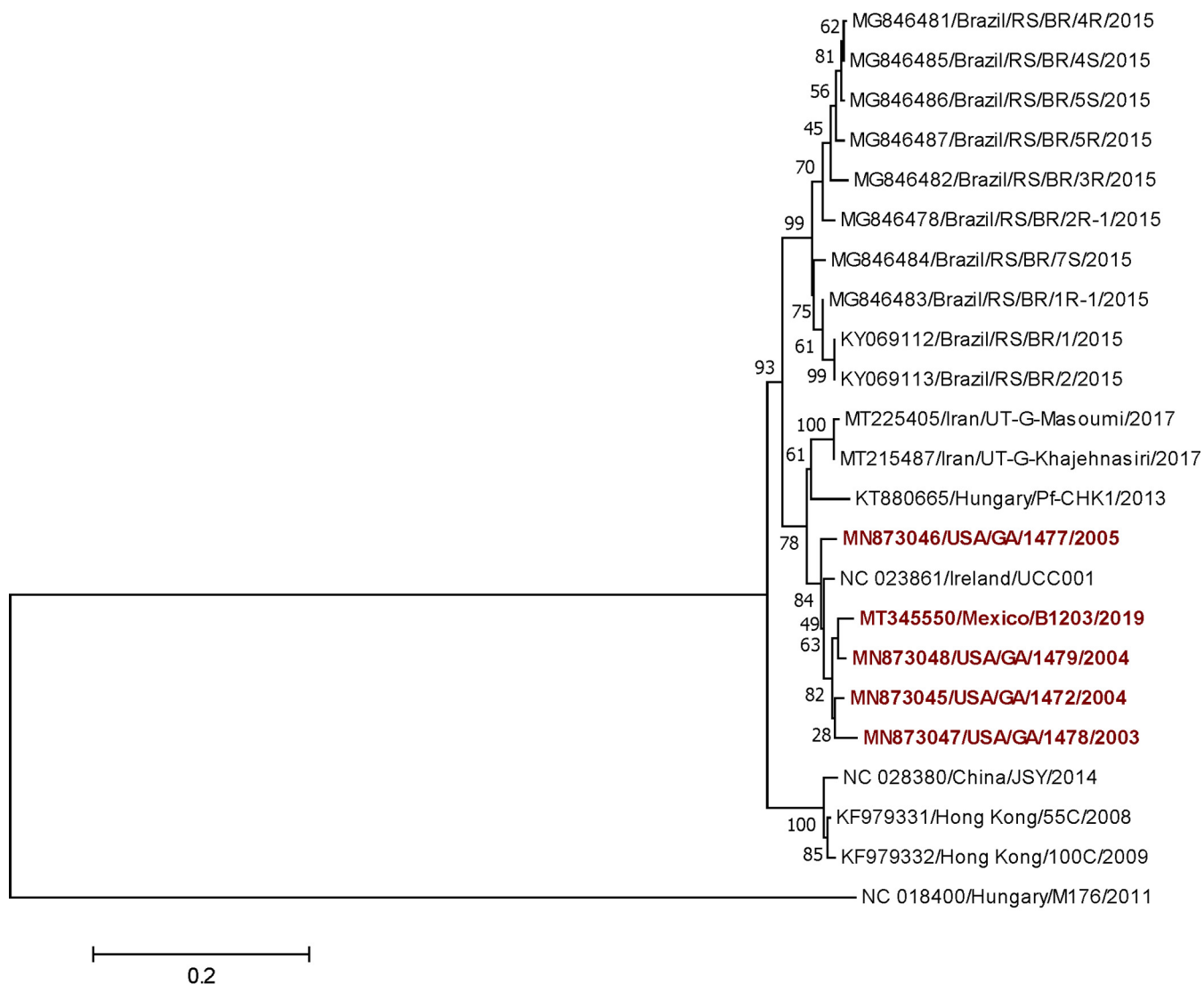
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**TABLE 1** Samples, sampling locations, dates, sequencing metrics, and accession numbers of the sicinivirus genomes and contigs in this report

| Isolate name   | Collection date (mo/day/yr) | Host               | Total no. of raw read pairs | No. of mapped reads | Median coverage depth (reads) | Mean read length (nt) <sup>a</sup> | Consensus length (bp) | GC content (%) | GenBank accession no. | SRA accession no. |
|----------------|-----------------------------|--------------------|-----------------------------|---------------------|-------------------------------|------------------------------------|-----------------------|----------------|-----------------------|-------------------|
| GA/1472/2004   | 10/5/2004                   | 22-day-old broiler | 2,088,901                   | 45,973              | 434                           | 138                                | 9,775                 | 53.9           | MN873045              | SRR10500281       |
| GA/1477/2005   | 9/29/2005                   | 16-day-old broiler | 2,446,408                   | 45,379              | 399                           | 121                                | 9,687                 | 54.5           | MN873046              | SRR10566436       |
| GA/1478/2003   | 4/28/2003                   | 18-day-old broiler | 1,303,453                   | 132,008             | 2,040                         | 127                                | 9,670                 | 54.1           | MN873047              | SRR10566435       |
| GA/1479/2004   | 6/8/2004                    | 15-day-old broiler | 2,212,775                   | 8,649               | 76                            | 157                                | 9,806                 | 54.1           | MN873048              | SRR10586503       |
| MEX/B1203/2019 | 6/11/2019                   | Chicken bursa      | 1,096,430                   | 22,464              | 366                           | 131                                | 9,706                 | 54.9           | MT345550              | SRR11542284       |

<sup>a</sup>nt, nucleotides.



**FIG 1** Phylogenetic analysis of sicinivirus sequences based on the complete amino acid sequence of the 3D<sup>pol</sup> protein. The evolutionary history was inferred using the maximum likelihood method based on the Jones-Taylor-Thornton matrix-based model in MEGA v7.0.26. The tree with the highest log likelihood (-3,357.96) is shown. The percentage of trees in which the associated taxa clustered together in the bootstrap test (1,000 replicates) is shown next to the branches. The initial tree(s) for the heuristic search was obtained automatically by applying the neighbor-join and BioNJ algorithms to a matrix of pairwise distances estimated using a JTT model and then selecting the topology with the superior log likelihood value. The tree is drawn to scale, with branch lengths measured in the number of substitutions per site. The analysis involved 23 amino acid sequences (the sequence of *Gallivirus* isolate [NC\\_018400](#) is included as an outgroup). All positions containing gaps and missing data were eliminated. There was a total of 472 positions in the final data set. The samples used in this study are shown in red.

total paired-end reads per sample (Table 1). All final consensus were called from the raw reads aligned to the *de novo*-generated contigs using the Burrows-Wheeler Aligner MEM algorithm (BWA-MEM) (17). Open reading frames were identified using Geneious v11.1.5 and confirmed by alignment with published sicinivirus genomes using MEGA v7.0.26. The genomes of all clinical samples had 100% complete coding sequences and showed the typical genetic structure of siciniviruses, with a single polyprotein cleaved into smaller nonstructural and structural capsid proteins, flanked by untranslated regions at both termini (1). Phylogeny based on 3D<sup>pol</sup> amino acid sequences confirmed that all viruses clustered together with other members of the *Sicinivirus* genus (Fig. 1). The highest BLASTp homology score for the 3D<sup>pol</sup> protein of all viruses was 96.4% to 97.7% amino acid sequence identity to the Ireland/UCC001 strain (NCBI RefSeq accession number [NC\\_023861.1](#)) (5). All viruses also possessed conserved amino acid motifs in 2C<sup>hel</sup> (GPPGCGKS, DDVGQ) and 3D<sup>pol</sup> (KDELK, GGNSPG, YGDD, and FLKR) proteins. The

Georgian viruses additionally demonstrated the conserved motifs in 3C<sup>Pro</sup> (QFKDL, GLCG) (5, 18, 19). Across the entire polyprotein gene, the strains designated GA/1472/2004, GA/1477/2005, GA/1478/2003, GA/1479/2004, and MEX/B1203/2019 showed the highest (85.5%, 85.3%, 84.9%, 84.5%, and 84.6%, respectively) nucleotide identity to isolate UCC001, from broiler chickens in Ireland (GenBank accession number [KF741227.1](https://doi.org/10.1009/jgv.0.000911)) (5). These near-complete genome sequences of sicinivirus collected from North America provide molecular epidemiological data needed to explore the evolution, epidemiology, and detailed pathogenesis of chicken picornaviruses globally.

**Data availability.** The complete coding sequences of all four Georgian sicinivirus samples have been deposited in GenBank under the accession numbers [MN873045](https://doi.org/10.1009/jgv.0.000911) to [MN873048](https://doi.org/10.1009/jgv.0.000911). The raw sequence data were deposited in the NCBI Sequence Read Archive (SRA) under BioProject number [PRJNA590745](https://doi.org/10.1009/jgv.0.000911). The Mexican sequence was deposited in GenBank under accession number [MT345550](https://doi.org/10.1009/jgv.0.000911). The raw sequence data were deposited in the SRA under BioProject number [PRJNA625289](https://doi.org/10.1009/jgv.0.000911).

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