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## **Age-Associated Proinflammatory Elastic Fiber Remodeling in Large Arteries**

#### **Soo Hyuk Kim**, **Robert E. Monticone**, **Kimberly R. McGraw**, **Mingyi Wang**

Laboratory of Cardiovascular Science, Intramural Research Program, National Institution on Aging, National Institutes of Health, Biomedical Research Center (BRC), 251 Bayview Boulevard, Baltimore, MD 21224, USA.

## **Abstract**

Elastic fibers are the main components of the extracellular matrix of the large arterial wall. Elastic fiber remodeling is an intricate process of synthesis and degradation of the core elastin protein and microfibrils accompanied by the assembly and disassembly of accessory proteins. Age-related morphological, structural, and functional proinflammatory remodeling within the elastic fiber has a profound effect upon the integrity, elasticity, calcification, amyloidosis, and stiffness of the large arterial wall. An age-associated increase in arterial stiffness is a major risk factor for the pathogenesis of diseases of the large arteries such as hypertensive and atherosclerotic vasculopathy. This mini review is an update on the key molecular, cellular, functional, and structural mechanisms of elastic fiber proinflammatory remodeling in large arteries with aging. Targeting structural and functional integrity of the elastic fiber may be an effective approach to impede proinflammatory arterial remodeling with advancing age.

#### **Keywords**

Aging; Large artery; Elastic fibers; Proinflammation

## **1. Introduction**

Aging is a major risk factor for the development of cardiovascular disease due mainly to age-associated adverse arterial remodeling through proinflammation and stiffening (Gavish and Izzo, 2016; Kohn et al., 2015; Villa-Bellosta, 2020; Wang et al., 2014a; Wang et al., 2020). With advancing age, extensive extracellular matrix (ECM) remodeling, especially elastic fiber degradation and collagen deposition, are commonly observed in the large arterial wall (Wang et al., 2014a; Wang et al., 2015; Wang et al., 2020). In the arterial wall,

<sup>\*</sup>Correspondence to Mingyi Wang, MD, PhD, FAHA, Laboratory of Cardiovascular Science, Intramural Research Program, National Institution on Aging, National Institutes of Health, Biomedical Research Center (BRC), 251 Bayview Boulevard, Baltimore, MD 21224, USA, mingyiw@grc.nia.nih.gov, Tel: (+1) 410-454-8579.

Conflicts of interest

None

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Elastic fiber degradation causes a decrease in or loss of arterial elasticity (Fhayli et al., 2019a; Rafuse et al., 2019). The degraded elastic fibers are susceptible to being oxidized, calcified, and can even become amyloidogenic (Basalyga et al., 2004; Davies et al., 2019; Davies et al., 2015; Gourgas et al., 2019; Ngai et al., 2018; Pai et al., 2011). Modified and degraded elastic fibers further enhance stiffness and fragility, significantly influencing the behavior of vascular smooth muscle cells (VSMCs) and endothelial cells (ECs) either directly or indirectly (Fhayli et al., 2019a; Karimi and Milewicz, 2016; Maurice et al., 2013; Pai et al., 2011; Qin, 2015; Rafuse et al., 2019).

Notably, elastic degradation in the arterial wall leads to insufficiently conferring the Windkessel effect thereby buffering the stroke volume ejected by the heart during systole and forwarding blood flow to the peripheral circulation during diastole (Humphrey et al., 2016). A common oversimplification is comparing the complicated functional and structural insufficiency of the degraded elastic fibers in the aging large arteries to a repeatedly stretched rubber band, which unavoidably loses its original recoil or elasticity over time but ignores the resultant proinflammation (Gundiah et al., 2009; Rauscher and Pomes, 2012). Notably, arterial stiffening is alleviated through the repair and prevention of the degradation of the elastic fiber with aging (Fhayli et al., 2020; Fhayli et al., 2019b; Wang et al., 2018b).

Arterial stiffening with aging plays a precipitating role in the initiation and progression of hypertension and atherosclerosis (Gavish and Izzo, 2016; Kohn et al., 2015; Villa-Bellosta, 2020; Wang et al., 2014a; Wang et al., 2015). In this mini review, we provide an update on the molecular mechanisms of the age-associated functional and structural elastic fiber remodeling and diseases such as hypertension, atherosclerosis, and aneurysm in large arteries.

#### **2. Elastogenesis in the large arteries**

Wang et al., 2018a).

The elastic fiber network consists of  $\sim$ 90% elastin (ELN) and  $\sim$ 10% fibrillin rich microfibrils (Fhayli et al., 2020). ELN is insoluble and is composed of polymerized tropoelastin. Tropoelastin, a soluble protein, with a recyclable ELN binding protein (EBP), is secreted from vascular elastogenic cells, mainly VSMCs (Lannoy et al., 2014; Yanagisawa and Davis, 2010). EBP, a cargo/chaperone protein, has the capacity to bind to the hydrophobic repeats of tropoelastin. The release of tropoelastin from EBP due to competition by unmasking galactosugars on the microfibrils via a series of catalysis of serine carboxypeptidase A on the cell surface (Pshezhetsky and Hinek, 2009). Once separated, EBP is recycled back into the cell; and tropoelastin is then aligned, coacervated, and crosslinked through desmosine and isodesmosine by lysyl oxidases (LOX) catalysis forming polymerized tropoelastin on the microfibrils (Pshezhetsky and Hinek, 2009). Polymerized tropoelastin becomes mature insoluble ELN, which then is embedded on the

microfibrils forming elastic fibers (Kothapalli and Ramamurthi, 2009; Yanagisawa and Davis, 2010). Thus, elastic fibers are composite structures, composed of a core ELN protein and an outer layer of fibrillin rich microfibrils (Fhayli et al., 2019a; Lannoy et al., 2014; Wagenseil and Mecham, 2007). In addition, a recent review suggests that potential alternative pathways exist and tropoelastin could be partially crosslinked on the cell surface or outside the cell before interacting with microfibrils and other fiber building components ELN globular protein (Kozel and Mecham, 2019).

Elastogenesis is a very complicated series of molecular events, that still not fully understood. Herein, a stepwise micro-macro-assembly of tropoelastin, microfibrils, and ELN into elastic fibers is briefly illustrated in Figure 1, based on previous reports (NivisoŽ Smith and Weiss, 2011; Pshezhetsky and Hinek, 2009; Schmelzer et al., 2020), as followings: (1) Tropoelastin is transcribed and translated from the ELN gene and is soluble and fragile. (2) Tropoelastin binds to EBP, limiting self-aggregation and degradation, and is secreted from vascular elastogenic cells; (3) Tropoelastin is released from EBP at the cell surface through competition by galactosugars in the microfibrils. Released tropoelastin is aligned, coacervated, and bound to fibulin on the microfibrils while released EBP is eventually recycled back into the cell. (4) Fibulin binds tropoelastin aggregates and stabilize them. (5) LOX on the microfibrils further processes aligned, coacervated the fibulin-stabilized tropoelastin in the extracellular space, which polymerizes tropoelastin to form mature insoluble ELN through desmosine and isodesmosine cross-links. (6) Finally, the elastic fiber is assembled, which consists of a core of ELN surrounded by microfibrils. Notably, aging increases total cell surface glycosaminoglycans (GAGs) including heparan sulfate (HS), chondroitin sulfate, and hyaluronic acid during the first 40 years of life and then decreases in the human aortic wall (Tovar et al., 1998); and tropoelastin interacts with GAG via its cterminal-domain, i.e., HS is present inside elastic fibers and plays a role in the assembly and stability of elastin coacervates (Annovi et al., 2012; Broekelmann et al., 2005; Pshezhetsky and Hinek, 2009). In addition, EBP with protective protein/cathepsin A and neuraminidase-1 (Neu1) forms a cell surface-targeted complex, known as elastin receptor complex. Notably, Neu1 can desialylate neighboring microfibrillar glycoproteins and facilitate the deposition of insoluble elastin, which also contributes to the maintenance of neighboring cellular quiescence (Hinek et al., 2008).

The elastic fiber is composed of ELN and microfibrils. ELN appears as slender bundles of polymerized tropoelastin, storing energy, and providing, strength, elasticity, and resilience of the arterial wall. Secreted fibrillin from VSMCs is incorporated into insoluble microfibrils, under the catalysis of transglutaminase 2 (TG2), providing a scaffold for deposition of ELN (Davis and Summers, 2012). Microfibrils appear as a complex 56 nm "beads-on-a-string" structure, which is directed by the fibronectin network (Kielty et al., 2005). Microfibrils not only direct/guide elastogenesis but also provide vascular elasticity and signal molecules such as transforming growth factor-beta 1 (TGF-β1) via a series of cleavage latent TGF binding protein-1 (LTBP-1) as well as cellular signaling via integrin receptors (Davis and Summers, 2012; Halper and Kjaer, 2014; Lannoy et al., 2014).

## **3. Elastic fiber remodeling in the large arterial wall**

Figure 2 shows that the large arterial elastic laminae (a sheet of concentrated elastic fibers) remodeling and associated adverse cell-matrix events with advancing age. The lumen becomes dilated, the tunica intima, the tunica media, and the tunica adventitia become thickened or enlarged. The elastic laminae become thinned, eroded, fragmented, calcified, and even amyloidogenic (Atanasova et al., 2010; Wang et al., 2014a; Wang et al., 2010; Wang et al., 2015; Wang et al., 2018a; Wang et al., 2012b; Wang et al., 2018b). The VSMCs embedded in the degenerated elastic fiber networks become labile, invasive, and migratory, penetrating the subendothelial space. The invaded intimal cells proliferate and secrete large amounts of ECM and inflammatory factors such as elastase, matrix metalloproteinases, milk fat globule-epidermal growth factor-VIII (MFG-E8), leading to intimal cellularity, thickening, inflammation, stiffening, and promoting endothelial permeability to circulating plasma proteins, such as oxidized low density lipoprotein (oxLDL) and circulating monocytes (Jaffe et al., 2012; Karimi and Milewicz, 2016; Lannoy et al., 2014; Pai et al., 2011; Pustlauk et al., 2020; Rensen et al., 2007; Spofford and Chilian, 2001; Wang et al., 2014a; Wang et al., 2010; Wang et al., 2015; Wang et al., 2018a; Wang et al., 2012b; Wang et al., 2018b). This molecular, cellular, and elastic fiber remodeling is the fertile soil for the development of hypertension, atherosclerosis, and aneurysm (Ailawadi et al., 2009; Chan et al., 2017; Di Gregoli et al., 2017; Harvey et al., 2016; Jacob et al., 2001; Karimi and Milewicz, 2016; Kunecki and Nawrocka, 2001; Mackey et al., 2007; Maurice et al., 2013; Rabkin, 2017; Toma and McCaffrey, 2012; Wagenseil and Mecham, 2012; Wang et al., 2015; Zampetaki et al., 2014). Growing evidence indicates that the disassembly of elastic fibers and the degradation of ELN release elastokines, known as ELN-derived peptides (EDPs), which play a precipitating role in age-associated arterial adverse remodeling such as endothelial dysfunction, fibrosis, calcification, and amyloidosis (Antonicelli et al., 2007; Fhayli et al., 2019a; Fhayli et al., 2020; Fhayli et al., 2019b; Greenwald, 2007; Hirai et al., 2007; Robert, 1998).

## **3.1 Tropoelastin**

Age related disruption of the elastic fiber laminae is predominantly associated with the suppressed tropoelastin expression and the degradation of ELN (Fhayli et al., 2019a; Fhayli et al., 2020; Fhayli et al., 2019b; Hirai et al., 2007; Robert, 1998; Rosenbloom, 1984; Wang et al., 2018b). Both ECs and VSMCs produce tropoelastin, forming ELN. In large arteries, ELN of the elastic laminae is mainly derived from VSMC production; in small arterioles such as muscular and resistance arteries, ELN of the internal elastic lamina is mainly derived from ECs (Lin et al., 2019). Notably, once ELN has been deposited, ELN synthesis ceases and turnover is close to nil. The half-life of vascular elastin, is  $\sim$ 25 years in mice and  $\sim$ 50 years in human (Arribas et al., 2006; Davis, 1993; Sherratt, 2009).However, a program of neosynthesis of ELN can be activated under certain conditions (Fhayli et al., 2020; Fhayli et al., 2019b).

**3.1.1 Transcription factors—**Transcription factor specificity protein 1 (SP1) regulates the expression of the ELN gene in VSMCs (Jensen et al., 1995). SP1 contains a zinc finger that binds to the GC-rich motifs of the ELN promoter (Sen et al., 2011; Yeh et al., 1989).

ELN is a target of TGF-β1 transcription and an integrated downstream molecule of its SMAD (an acronym from the fusion of Caenorhabditis elegans Sma genes and the Drosophila Mad, Mothers against decapentaplegic proteins) signaling pathway (Davidson, 2002; Jaffe et al., 2012; Sproul and Argraves, 2013). The TGF-β1 downstream SMAD molecule activates SP1, and subsequently activates the ELN gene promoter and or stabilizes ELN mRNA, leading to increases in tropoelastin and ELN formations in both VSMCs and ECs (Ellenrieder, 2008; Sproul and Argraves, 2013). Insulin-like growth factor 1 (IGF-1), like TGF-β1, increases tropoelastin production and ELN formation in elastogenic cells through the activation of SP1 (Davie et al., 2008; Li et al., 2004; Sproul and Argraves, 2013). Notably, with advancing age, intimal TGF-β1 increases in the thickened intima while medial TGF-β1 decreases in the arterial wall (Sauvage et al., 1998; Wang et al., 2006); and aging also decreases IGF-1 abundance, which is closely associated with the downregulation of ELN mRNA expression in the arterial medial wall (Foster et al., 1990).

Fos-related antigen 1 (FRA1) is a leucine zipper protein and dimerizes with oncogene c-Jun forming the transcription factor complex, AP-1. Basic fibroblast growth factor (bFGF) inhibits ELN gene expression through an enhancement of an AP-1/cAMP-response element hybrid site in the distal promoter in VSMCs (Carreras et al., 2002; Rich et al., 1999; Sproul and Argraves, 2013). Tumor necrosis factor-alpha (TNF-α) enhances AP-1 suppression of the ELN promoter and inhibits its activity, leading to decreased tropoelastin expression and ELN formation in VSMCs (Kahari et al., 1992; Spofford and Chilian, 2001).

ELN is a critical regulatory molecule that regulates the phenotypic modulation, proliferation, and migration of VSMCs. ELN expression and VSMCs proliferation are coupled inversely: potent activators such as platelet-derived growth factor-BB (PDGF-BB) of cell proliferation may potentially inhibit ELN expression and potent inhibitors of cell proliferation can stimulate ELN production (Seyama and Wachi, 2004). ELN is expressed maximally at the G0 and minimally at the G2/M phase during the cell cycle, revealing that its expression is tightly regulated by the cell growth rate (Seyama and Wachi, 2004). Notably, retinoblastoma protein (Rb) modulates the inverse relationship between cellular proliferation and elastogenesis, i.e., PDGF-BB increases cellular proliferation but decreases elastic fiber production via mitigation of phosphorylating Rb on Thr-821; IGF decreases cellular proliferation but increases ELN production via enhancement of phosphorylating RB on Thr-821 (Sen et al., 2011).

**3.1.2 Post transcriptional/translational modifications—**Post-transcriptional/ translational modifications are predominant factors for the reduction of ELN levels in the aging arterial wall. Micro-RNA (miR), elastase, matrix metalloproteinases (MMPs) and calpain-1 either directly or indirectly modify intact ELN abundance at the posttranscriptional or post-translational levels.

**3.1.2.1 miRs:** A miR is a small noncoding RNA molecule that functions in RNA silencing and post-transcriptional regulation of gene expression. miRs effect tropoelastin expression and ELN formation in VSMCs and the arterial wall (Chan et al., 2017; Di Gregoli et al., 2017; Zampetaki et al., 2014). miR-29 downregulates ELN expression, prevents elastic laminae breaks, and VSMC osteoblastic differentiation in the arterial wall (Sudo et al.,

2015). miR-181b decreases the expression of ELN expression and promotes elastic fiber destruction in the arterial wall (Di Gregoli et al., 2017). miR-195 reduces ELN expression in VSMCs in vitro while antagomiR-195 upregulates aortic ELN expression in vivo, and silenced miR-195 inactivates MMP-2/9 and subsequently diminishes ELN degradation (Zampetaki et al., 2014). miR-516a-5p regulates MMP-2 and TIMP-1 expressions in human VSMCs, possibly promoting the proteolytic degradation of elastin in the arterial wall (Chan et al., 2017).

**3.1.2.2. Elastolysis:** Proteases can degrade ELN or elastic fibers, known as elastases. The following elastases, including serine proteases leukocyte and pancreatic elastases, metallopeptidases MMP-2/−7/−9/−12–14, and cysteine proteases cathepsins and calpain-1, directly or indirectly break down elastic fibers and degrade ELN through a process known as elastolysis, that with collagen deposition, predominantly determine the mechanical molecular stiffness properties of the arterial wall with advancing age(Heinz, 2020; Raffetto and Khalil, 2008; Wang et al., 2015). Elastolysis is a very slow enzymatic catalytic process progressing over decades in the aging arterial wall (Heinz, 2020; Wang et al., 2015).

*3.1.2.2.1 Serine Proteases:* Elastase is a serine endopeptidase, which effectively cleaves tropoelastin, ELN, and elastic fibers in the arterial wall. Leucocyte elastase protein levels increase in aging arterial walls and serum leukocyte elastase activity is increased in old rats (Fornieri et al., 1999; Paczek et al., 2009). Leukocyte and pancreatic elastases cleave the elastic fiber/network and degrades ELN with aging (Antonicelli et al., 2007; Barolet et al., 2001; Fornieri et al., 1999; Paczek et al., 2009; Wang et al., 2015). Notably, calciumphosphorus products accumulate and bind to degraded elastic fibers, increasing exposed ELN susceptibility to be cleaved in the adverse remodeled arterial wall (Seyama and Wachi, 2004). Importantly, EDPs further enhances the degradation of ELN and elastic fibers via the increase of elastase activity in a feed-forward manner (Hornebeck and Emonard, 2011; Maurice et al., 2013).

*3.1.2.2.2 Metallopeptidases:* Matrix metalloproteinase types 2/7/9/12/14 (MMP2/9/12) are extracellular proteinases and metalloelastases. They are able to bind to elastic fibers, and in turn degrade the elastic network, and subsequently increase the bioactivity and bioavailability of latent TGF binding proteins (LTBPs), latent associated protein (LAP), and TGF-β1 (Basalyga et al., 2004; Skjot-Arkil et al., 2012; Van Doren, 2015; Wang et al., 2006). Activated MMPs and its activator membrane-type of MMP (MT-MMP, MMP14) effectively cleave elastic fibers and produce EDPs (Robinet et al., 2005; Szychowski et al., 2019). Tissue inhibitors  $1/2/3/4$  of matrix metalloproteinase (TIMP $1/2/3/4$ ) are endogenous inhibitors of MMP activation (Szychowski et al., 2019). Thus, a balance of the MMP/TIMP ratio plays an important role in maintaining the integrity of the elastic laminae or network in the aging arterial wall (Wang et al., 2014a; Wang et al., 2014b; Wang and Khalil, 2018).

*3.1.2.2.3 Cysteine proteinases:* Cathepsins are cysteine proteases. Normal arteries contained little or no cathepsin K or S, in contrast, intimal smooth muscle cells (SMC), especially those appearing to traverse the internal elastic laminae and macrophages in the atheroma contained abundant immunoreactive cathepsins K and S. These enzymes to

effectively degrade elastin and play an elastolytic role during arterial wall remodeling and in the stability of atherosclerotic plaques(Sukhova et al., 1998).

Calpain-1, an intracellular cysteine protease, is increased with age in VSMCs and the arterial wall (Jiang et al., 2008; Jiang et al., 2012). Calpain-1 effectively activates MMP-2 in VSMCs with advancing age (Jiang et al., 2012). The activated MMP-2 degrades elastic fibers (Jiang et al., 2012; Wang et al., 2010). In addition, calpain-1 increases alkaline phosphatase (ALP) levels in cultured VSMCs while decreases the calcification inhibitors, osteopontin and osteonectin, eventually facilitating the calcification of aging VSMCs and the arterial wall (Blewniewski et al., 2011; Jiang et al., 2012). Thus, inhibiting calpain-1 potentially delays the age-related elastic fiber damage and calcification of the arterial wall (Jiang et al., 2012).

**3.1.2.3. Lysyl oxidase (LOX):** LOX is an extracellular copper-dependent enzyme that catalyzes formation of aldehydes from lysine residues in tropoelastin. The aldehydes are highly reactive and undergo spontaneous chemical reactions with other LOX-derived aldehyde residues, resulting in cross-linking tropoelastin by desmosine or isodesmosine (Gourgas et al., 2019; Kothapalli and Ramamurthi, 2009). LOX deficiency causes a defect in tropoelastin crosslinking and disrupts the elastic fibers or network assembly (Maki et al., 2002). LOX, together with tropoelastin, decreases in the aging arterial wall (Fhayli et al., 2020). Overexpressing or preserving LOX increases the yield of tropoelastin and ELN formation in VSMCs and in the arterial wall with aging (Fhayli et al., 2020; Kothapalli and Ramamurthi, 2009; Oleggini et al., 2007).

**3.1.2.4 Glycosylation and Carbamylation:** Aging is a progressive chronic process determined by genetic, epigenetic, and acquired factors. The chemical reactions referred to as nonenzymatic posttranslational modifications (NEPTMs) are acquired factors, such as glycosylation and carbamylation, which are responsible for molecular aging. Glycation is associated with aging and has been shown to increase arterial stiffness. Glycation directly effects the mechanical response of intact arteries and the mechanical response and structure of ELN isolated from the arteries (Stephen et al., 2014). Glycation leads to the thinning of elastic fibers and the weakening of the ELN of arteries, contributing to vascular defects in aging (Stephen et al., 2014). Thus, prevention of glycation reactions may be an important consideration for vascular health later in life.

Carbamylation is a more recently described NEPTM that is caused by the nonenzymatic binding of isocyanate derived from urea dissociation or myeloperoxidase-mediated catabolism of thiocyanate to free amino groups of proteins. This modification is considered an adverse reaction because it induces alterations of protein and cell properties. Protein carbamylation may be considered a hallmark of aging in mammalian species that may significantly contribute to the structural and functional adverse remodeling that occurs during aging (Gorisse et al., 2016). Cyanate carbamylation at the N-terminal of amino acids is observed in aortic ELN (Lefevre and Rucker, 1983). Age-related modifications including glycation and carbamylation greatly impact on ECM bioroles and their mechanical properties (Birch, 2018).

#### **3.2 Microfibrils**

Microfibrils are composed of fibrillin and are functionally affected by other accessary proteins, including fibulin and emilin. In general, these accessory proteins modify the size and configuration of ELN aggregates on the microfibrils (Yanagisawa and Davis, 2010). The aggregated ELN interacts with the microfibril framework on the cell surface through integrins, inducing cellular remodeling such as the VSMC phenotypic switch (Lannoy et al., 2014; Yanagisawa and Davis, 2010).

**3.2.1 Fibrillin—**Fibrillin is an essential glycoprotein for the formation of elastic fibers (Davis and Summers, 2012). Secreted fibrillin from VSMCs is incorporated into insoluble microfibrils, providing a scaffold for the deposition of aggregated ELN (Davis and Summers, 2012). Microfibrils not only fulfill the assembly of the elastic fiber but also provide tissue elasticity (Halper and Kjaer, 2014; Lannoy et al., 2014). Fibrillin includes three isoforms: fibrillin-1, −2 and −3. Fibrillin-1 and fibrillin-2 enhances the stability of elastic fibers (Davis and Summers, 2012). Fibrillin-1 also sequesters LTBP and controls the bioavailability of active TGF-β1 (Davis and Summers, 2012). Fibrillin-2 is expressed earlier and is involved in elastic fiber formation in fetal development. Fibrillin-3 is mainly expressed in the brain (Kielty et al., 2005; Kielty et al., 2002). Notably, fibrillin-1 deficient mice have substantial fragmentation and decreased number of elastic lamellae in the aortic wall with aging, which were correlated with an increase in aortic stiffness, a decrease in vasoreactivity, an altered expression of ELN, and a decrease in the relative ratio between tissue ELN and collagen (Mariko et al., 2011).

**3.2.2 Fibulin—**The fibulin family is a group of glycoproteins involved in elastic fiber formation (Yanagisawa and Davis, 2010). The fibulin family is composed of long fibulins (fibulin-1,  $-2$ ,  $-6$ ) and short fibulins (fibulin-3,  $-4$ ,  $-5$ ,  $-7$ ). Fibulin-1 and  $-2$  are mainly produced by VSMCs involved in formation of elastic fibers within the medial layer of the artery (de Vega et al., 2009). Fibulin-4 and −5 participate in elastic fiber assembly and affect the deposition of ELN during elastic fiber generation (Choudhury et al., 2009; Hirai et al., 2007; Yamauchi et al., 2010). Notably, fibulin-5 deficient mice display early vascular aging appearing as marked elastopathy: an increased stiffness of the proximal central arteries due largely to increased wall thickness and collagen–ELN ratio (Cuomo et al., 2019).

**3.2.3 Emilin-1—ELN** microfibril interface-located protein 1 (Emilin-1) is a glycoprotein that is able to anchor VSMCs to elastic fibers, maintain the quiescence of VSMCs in the normal adult arterial wall and controls the release of the cytokine TGF-β1 (Randell and Daneshtalab, 2017; Zanetti et al., 2004). Emilin-1 is able to bind to ELN and to fibulin-5 stabilizing elastic fibers (Angel et al., 2017; Zanetti et al., 2004). The decrease of Emilin-1 increases TGF-β1 fibrogenesis and inflammation in VSMCs and in the arterial wall with aging (Angel et al., 2017; Randell and Daneshtalab, 2017). Importantly, Emilin-1 deficiency induced the defects of arterial elastogenesis such as elastic fibers fragmentation (Angel et al., 2017).

## **4 Elastic fiber remodeling associated with other ECM restructuring**

Elastic fiber remodeling occurs with the restructuring of other ECM components, such as collagen, fibronectin (FN), laminin (LM), and milk fat globule-epidermal growth factor 8 (MFG-E8) in the arterial wall with aging.

#### **4.1 Collagen**

ELN degradation and elastic fiber degeneration dramatically increase activated TGF-β1 bioavailability and release EDPs (Lannoy et al., 2014; Oleggini et al., 2007; Simionescu et al., 2005; Yamashiro and Yanagisawa, 2018) which are powerful peptides that activate TGFβ1 secreted from VSMCs (Simionescu et al., 2005). TGF-β1 signaling potentially increases collagen types I and III production in VSMCs (Wang et al., 2006). Collagen type I is the most abundant collagen in vascular walls and is associated with tensile strength, stiffening, and calcification while collagen type III has a long, inflexible, triple-helical domain, contributing to the elasticity of the vascular wall (Kohn et al., 2015; Wagenseil and Mecham, 2012). Increases in the ratios of collagen/ELN and collagen type I to III in the arterial wall dramatically increase with aging (Kochova et al., 2012; Wang and Lakatta, 2002). Notably, elastolysis often accompanies the degradation of collagen, promoting proinflammation and dramatically increases stiffness in the arterial wall (Mozos et al., 2017; Wang et al., 2020).

#### **4.2 Fibronectin**

Fibronectin (FN) is a "master organizer" in matrix assembly and is a downstream molecule of TGF-β1 signaling, which exerts pivotal roles in directing VSMC-mediating remodeling of scaffolds toward the production of a physiological-like, elastin-containing ECM with excellent mechanical properties (Pezzoli et al., 2018; Wang et al., 2006). FN connects with elastic fibers, influencing elastic fiber assembly, deposition, and integrity via the assembly of microfibrils (Pezzoli et al., 2018; Xu and Shi, 2014). With advancing age, FN expression and deposition increases while ELN content declines, elastic fibers degrades, and vascular calcification increases (Zhang et al., 2020). In addition, the functional interplay of EBP and interleukin-1 receptor regulates the production of FN in VSMCs (Hinek et al., 1996).

#### **4.3 Laminin**

Activated TGF-β1 triggers the production of laminin (LM) from VSMCs (Basson et al., 1992). LM is a major component of the basal lamina of cells. LM effects on the biosynthesis of ELN and interacts with ELN or EDPs through the transmembrane ELN–LM receptor (ELR) complex, affecting the properties of VSMCs such as proinflammation and proliferation (Kunecki and Nawrocka, 2001; Robert, 1998; Spofford and Chilian, 2001).

#### **4.4 MFG-E8**

MFG-E8 is a secreted glycoprotein, which functions as a bridge molecule to connect VSMCs to elastic fibers in the arterial wall(Wang et al., 2012a; Wang et al., 2013). The MFG-E8 fragment, medin, is an amyloidogenic protein and also binds to elastic fibers and VSMCs in the arterial wall (Degenhardt et al., 2020; Migrino et al., 2020; Wang et al., 2012a; Wang et al., 2013; Younger et al., 2020). Aging increases both MFG-E8 and medin in the arterial wall (Wang et al., 2012a; Wang et al., 2013). Both MFG-E8 and medin signaling

influence the VSMC phenotypic switch and vessel function (Chiang et al., 2019; Degenhardt et al., 2020; Fu et al., 2009; Migrino et al., 2020; Younger et al., 2020). MFG-E8 increases the invasion, proliferation, and migration of VSMCs; and medin increases the necrosis and MMP-2 activation of VSMCs (Chiang et al., 2019; Fu et al., 2009; Mackey et al., 2007; Wang et al., 2012a; Wang et al., 2013). Notably, MFG-E8, medin fragments, and EDPs all are elements of aortic medial amyloidosis (Boraldi et al., 2018; Davies et al., 2019; Davies et al., 2015; Larsson et al., 2006; Peng et al., 2007).

## **5 Elastic fiber remodeling effects on VSMC phenotypes in the arterial wall**

#### **5.1 Proinflammation**

The age-associated inflammation in the arterial wall or VSMCs is a low grade of sterile inflammation where cytokines are secreted mainly by vascular cells rather than traditional immune cells, known as proinflammation. Maintaining stable functional ELN-contractile units is a key to warranting the structural and functional integrity and health of the arterial wall (Karimi and Milewicz, 2016). The integrity of the elastic network has significant effects on VSMC behaviors (Kinnear et al., 2020; Lannoy et al., 2014; Lin et al., 2019; Pai et al., 2011; Sudo et al., 2015). In the arterial wall, VSMCs are generally categorized into contractile and synthetic phenotypes (Rensen et al., 2007). In healthy adult vascular walls, the vast majority of VSMCs are in a quiescent non-proliferative phenotype, known as contractile, while the phenotype of proliferating, invading/migrating, proinflammatory secreting VSMCs in old arterial wall have been widely referred to as synthetic (Wang et al., 2014a). Adverse ELN remodeling, such as releasing EDPs or fragment fibers, plays a permissive role in the phenotypic switching of the contractile into synthetic VSMCs (Kinnear et al., 2020). Synthetic VSMCs exert a proinflammatory role in arterial intimal medial thickening and stiffening with aging (Laurent et al., 2005; Wang et al., 2014a).

Contractile VSMCs are enriched in smooth muscle specific 22 kDa protein (SM22α) and alpha-smooth muscle actin (α-SMA). These cellular markers increase in young arterial walls while decrease in the old (Ailawadi et al., 2009; Martin-Pardillos and Sorribas, 2015). Contractile VSMCs are converted to the synthetic phenotype and are usually accompanied by a decrease in the expression of these contractile proteins and a reduction in ELN production (Ailawadi et al., 2009; Jaffe et al., 2012; Martin-Pardillos and Sorribas, 2015; Rensen et al., 2007). Collapse of the arterial elastic network facilitates the shift from contractile to synthetic phenotypes of VSMCs (Karimi and Milewicz, 2016; Lannoy et al., 2014; Wagenseil and Mecham, 2007; Yamashiro and Yanagisawa, 2018). ELN degradation is associated with synthetic VSMCs, and the increase of ELN-contractile unit damage and modulus, contributes to intimal thickening and arterial stiffening (Barolet et al., 2001; Karimi and Milewicz, 2016). Low levels of elastic fiber deposition around VSMCs coincide with an increase in the proliferation rate, which is reversed by the addition of exogenous ELN (Urban et al., 2002). VSMCs lacking ELN proliferate at a rate greater than that of the wild type cells, fail to form a mature contractile phenotype, and migrate more aggressively to an exogenous chemoattractant than that of the wild type cells. Importantly, these lacking ELN-associated phenotypic shifts in VSMCs are rescued in vitro by the addition of recombinant tropoelastin (Karnik et al., 2003; Li et al., 1998; Seyama and Wachi, 2004). The

small, infiltrated synthetic VSMCs have been found in the aged, thickened intimae contain an abundance of angiotensin II and activated MMP-2 which accompanies the destruction of the elastic network, which has evolved into a fertile proinflammatory field (Wang et al., 2003).

Notably, deteriorated elastic fibers release elastokines, or EDPs, that bind to the ELN receptor and other receptors, stimulate the migration and proliferation of VSMCs, and damage ECs (Dale et al., 2016; Kawecki et al., 2014; Maurice et al., 2013; Robinet et al., 2005; Simionescu et al., 2005). In addition, EDPs are involved in thrombosis via increases in platelet aggregation and clotting signaling (Kawecki et al., 2014). These detrimental effects can be largely inhibited by genetic or pharmacological blockades of the ELN receptor complex or through the neutralization of an antibody against EDPs (Qin, 2015). EDPs are chemoattractant proteins and regulates invasion/migration and matrix metalloproteinase expression. EDPs are chemotactic for inflammatory cells such as monocytes, VSMCs, ECs, and myofibroblasts (Annovi et al., 2012; Duca et al., 2016).

#### **5.2 Procalcification**

Age-associated arterial proinflammation and profibrosis coupling is the foundation of arterial calcification (Wang et al., 2020). Ectopic micro-and macro-calcification can be detected in either the intimal or medial layers of the arterial wall (Gourgas et al., 2019; Ngai et al., 2018; Pai et al., 2011). Medial calcification appears as linear deposits of calcium along the degraded elastic fibers (Basalyga et al., 2004; Gourgas et al., 2019). The increase in elastic fiber degradation is closely associated with the accumulation of calcification (Basalyga et al., 2004; Maurice et al., 2013). Remarkably, synthetic VSMCs are susceptible to calcification, and are associated with the downregulation of ELN expression and degraded elastic fibers (Maurice et al., 2013; Sudo et al., 2015). Calcium absorbs on ELN-like peptide filaments or fibers, and then evolve upon nucleation, and finally calcium spread out to ELPlike globular structure forming calcification (Gourgas et al., 2019).

Fetuin-A, also known as alpha-2-HS (Heremans-Schmid)-glycoprotein, is a mineral carrier protein and a systemic inhibitor of pathological mineralization (Jahnen-Dechent et al., 2011; Robinson and Teran-Garcia, 2016; Villa-Bellosta, 2020). It carries insoluble calcium phosphate forming soluble complexes that inhibit calcium deposition (Villa-Bellosta, 2020). Loss of fetuin-A increases systemic soft tissue calcification in mice (Jahnen-Dechent et al., 2011; Westenfeld et al., 2009). Fetuin-A decreases with advancing age and thus promotes an age-associated increase in ectopic calcification (Jahnen-Dechent et al., 2011; Robinson and Teran-Garcia, 2016). A significant negative correlation is also observed between plasma fetuin-A concentrations and arterial calcification in humans (Westenfeld et al., 2009).

Transglutaminase-2 (TG-2) is an enzyme that in nature primarily catalyzes the formation of an isopeptide bond between  $\gamma$ -carboxamide groups of glutamine residue side chains and the ε-amino groups of lysine residue side chains and subsequently releases ammonia. TG-2 expression is increased in the aging arterial wall and is abundant in calcified arterial walls (Chabot et al., 2010; Johnson et al., 2008; Lexhaller et al., 2019). TG-2 secreted from VSMCs is a calcium-dependent enzyme that can crosslink most ECM proteins such as ELN and traps the calcium crystals, hydroxyapatites (Chabot et al., 2010; Johnson et al., 2008).

An increase in TG-2 expression enhances the chondro-osseous differentiation of VSMCs via the activation of an osseous master Runt-related transcription factor (Runx2)/ core-binding factor subunit alph-1 (Cbf-α−1) and promotes arterial calcification and stiffening (Chabot et al., 2010; Johnson et al., 2008).

Alkaline phosphatase (ALP) is a non-tissue specific enzyme that is detected in the calcified arterial wall (Pustlauk et al., 2020; Tani et al., 2020; Villa-Bellosta, 2020). ALP is a dephosphorylating compound, which converts pyrophosphate (soluble) into phosphate (insoluble), facilitating calcium deposition. A high concentration of ALP is found in the calcified arterial wall. Specifically, aging increases ALP and promotes arterial calcification (Jiang et al., 2012; Simionescu et al., 2005; Sinha and Vyavahare, 2013; Westenfeld et al., 2009).

## **6 Arterial diseases associated with elastic fiber remodeling**

Adverse age-associated elastic fiber remodeling has a significant impact on the pathogenesis of large arterial diseases (Ferrucci, 2008; Fhayli et al., 2019a; Halabi and Kozel, 2020; Robert, 1998; Xu and Shi, 2014).

#### **6.1 Hypertension**

Aging increases the prevalence of hypertension, accompanied by the fragmentation of the elastic laminae and the damage of the ELN-contractile unit that links the elastic fibers to the VSMCs in the aortic wall, contributing to an increase in blood pressure (Wagenseil and Mecham, 2012). Proinflammatory signals resulting from the breakdown of elastic fibers are important contributing factors, such as increases in bioactivity and bioavailability of TGFβ1, MFG-E8, medin, and EDPs in the arterial wall, increased vasoconstriction, and elevated progression of hypertension (Faury et al., 2003; Harvey et al., 2016; Wagenseil and Mecham, 2012; Wang et al., 2015). Activated TGF-β1 triggers MMPs activation, elastolysis, fibrosis and arterial stiffening, that promotes or maintains the state of hypertension (Greenwald, 2007; Harvey et al., 2016; Wagenseil and Mecham, 2012; Wang et al., 2011; Zacchigna et al., 2006). Indeed, ELN +/− mice have stable hypertension from birth, with a mean arterial pressure 25 to 30 mm Hg higher than their wildtype counterparts, and further physiologic studies indicated a role for the renin-angiotensin system in maintaining their hypertensive state (Faury et al., 2003). In contrast, the expression of the human ELN transgene reversed the hypertension and cardiovascular changes associated with ELN haploinsufficiency and rescued the perinatal lethality of the ELN-null phenotype (Hirano et al., 2007). In addition, serum levels of glycated ELN derived peptides (AGE-EDP) are increased in arterial hypertension (Wahart et al., 2019). Notably, elastolysis promotes arterial stiffening which precedes hypertension, thus suggesting stiffness is a cause of hypertensin(Humphrey et al., 2016). From a biomechanical perspective, arterial aging or hypertension in mice may not directly mimic human arterial aging or hypertension, but the mouse model which shares similar properties with human cells and extracellular matrix making the mouse a powerful model for studying human aging and hypertension. Analyzing a mouse model is invaluable in understanding the mechanisms of aging and hypertensin (Spronck et al., 2020).Taken together, the association of hypertension with ELN

haploinsufficiency in humans and mice strongly suggested that ELN and other proteins of the elastic fiber should be considered as causal genes for essential hypertension.

#### **6.2 Atherosclerosis**

Aging increases the arterial elastic network degradation and collapse, which is closely associated with atherogenesis (Maurice et al., 2013; Robert, 1998; Seyama and Wachi, 2004). Aging creates a notable proinflammatory environment that drastically increases elastase and MMP activity, which effectively cleaves ELN, producing elastokine EDPs (Maurice et al., 2013). EDPs also are potent proinflammatory cues to further damage elastic fibers via facilitating oxidation, oxLDL accumulation, calcification, and infiltration of lipids and immune cells into the deep aortic wall, and eventually accelerating the progress of atherosclerotic plaques (Fulop et al., 1998; Toma and McCaffrey, 2012). Fragmented elastic fibers have a high affinity to bind to calcium-phosphorus products and lipids in the arterial wall, promoting the growth and instability of the atherosclerotic plaque (Maurice et al., 2013). Indeed, mice lacking ELN died of an obstructive arterial disease that resulted from subendothelial cell proliferation and reorganization of smooth muscle, which were similar to those seen in atherosclerosis (Li et al., 1998). In a recent study of young (6–15 years-old, n=8) and old rhesus monkeys (16–28 years-old, n=8), fed a high fat and cholesterol diet for two years revealed that aging, per se, predisposes adverse cells and matrix remodeling, including ELN degradation of the arterial wall, to develop atherosclerosis (Wang and Lakatta, 2019). A high cholesterol diet for a fixed time increased atherosclerosis to a markedly greater extent in old rather than in young monkeys. Thus, the risk of atherosclerosis is attributable to age and is not due to longer exposure to dyslipidemia but rather stems from age associated adverse remodeling of the arterial wall, i.e., the soil in which atherosclerosis flourishes (Wang and Lakatta, 2019).

#### **6.3 Aneurysm**

With advancing age, ELN is degraded, elastic fibers are degenerated, contributing to arterial inflammation, calcification, and fragility (Kunecki and Nawrocka, 2001; Lavin et al., 2020; Maki et al., 2002). Age increases elastase and MMP activation, which degrade ELN protein and degenerate elastic fibers and the synthetic shift of VSMCs during the pathogenesis of arterial aneurysm (Ailawadi et al., 2009; Antonicelli et al., 2007; Cohen et al., 1992; Lavin et al., 2020; Rabkin, 2017). Destruction of elastic fibers at sites of abdominal aortic aneurysm (AAA) leads to the accumulation of degraded monomeric soluble tropoelastin rather than the cross-linked insoluble ELN (Lavin et al., 2020). Histologic observation indicates that in aneurysm tissue, the number of VSMCs decreases while the elastic network is destroyed, leading to weakness and microcalcification of the ELN-contractile unit (Jacob et al., 2001; Kielty et al., 2005). This weakness of the ELN-contractile unit in the arterial wall is susceptible to being expanded under mechanical and metabolic stress such as hypertension and atherosclerosis forming an opportunistic site for the initiation of arterial aneurysm (Ailawadi et al., 2009; Rabkin, 2017). The ELN associated microcalcification as a novel imaging marker successfully enables monitoring of the local ELN degradation, destruction of elastic fibers and predicts aortic adverse events in aneurysm patients (Wanga et al., 2017).

Disorders of fibrillin-1, emilin-1, fibulin-5, LOX, and ELN-laminin receptor also promote inflammation and the destruction of the ELN-contractile unit and triggers the formation of arterial aneurysm (Karimi and Milewicz, 2016; Kunecki and Nawrocka, 2001). In addition, accumulating evidence indicates that EDPs actively participate in the progression of arterial aneurysm by accelerating inflammation, LDL oxidation, ELN degeneration, calcification, and stiffening of the arterial wall (Antonicelli et al., 2007; Qin, 2015; Wen et al., 2011). EDPs also promote VSMCs proinflammation in a paracrine manner, facilitating ELN degradation and elastic fiber fragmentation and eventual elastolysis in a feed-forward manner, leading to the pathogenesis of arterial aneurysm/dissection (Antonicelli et al., 2007; Qin, 2015; Wen et al., 2011). EDPs create a proinflammatory environment in the aortic wall by inducing macrophage type 1 polarization while neutralizing the EDPs attenuated aortic dilation (Dale et al., 2016.

These pathophysiological effects are mediated by the binding of EDPs on a unique heterotrimeric receptor named ELN receptor complex (ERC) (Maurice et al., 2013).

## **7 Concluding remarks and perspectives**

ELN is a core protein of the elastic fiber. With advancing age, intact ELN protein decreases through transcription, post-transcriptional or post-translational modifications, contributing to the degradation of the arterial extracellular elastic fiber network. These defects lead to the insufficiency and collapse, accompanied by proinflammation, procalcification, amyloidosis, and eventual stiffening in the old arterial wall. Many recent studies indicate that habitual aerobic exercise, dietary calorie restriction, and a healthy lifestyle effectively preserves the integrity, elasticity, resilience of elastic fiber laminae and reduces arterial stiffening and (Alfaras et al., 2016; AlGhatrif et al., 2017; Castello et al., 2005; Seals et al., 2009; Wang et al., 2018b). Age-associated arterial structural and functional declines may not be inevitable (Fhayli et al., 2019b; Foster et al., 1990; Lefevre and Rucker, 1983; Wang et al., 2014b; Wang et al., 2012b; Wang et al., 2018b). This novel conception is robustly supported by the Baltimore Longitudinal Study of Aging (BLSA) of "successful aging in a subset of the human population" (Ferrucci, 2008). This mini review focuses on the repair/prevention of the elastic fiber, which plays a major role in age-associated adverse proinflammatory arterial remodeling, The limitation of the 'rubber band' and elastic fiber comparison (cumulative wear and tear over time) still plays an important role since the loss of elasticity not only decreases the Windkessel effect but also leads to age-associated arterial proinflammation (Avolio et al., 1998; Greenwald, 2007; O'Rourke and Hashimoto, 2007). Therefore, targeting the adverse proinflammatory molecular events of elastic fiber remodeling may be a potential fresh therapeutic approach to retard age-associated adverse structural and functional arterial remodeling and diseases such as hypertension, atherosclerosis, and aneurysm.

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## **Highlights**

- **•** Elastic laminae are the main components of the extracellular matrix of large arteries.
- **•** Elastic lamina remodeling is an intricate process of synthesis and degradation of the core tropoelastin protein accompanied by the assembly and disassembly with accessory proteins.
- **•** Age related morphological, structural, and functional proinflammatory remodeling within the elastic laminae have a profound effect upon the integrity, elasticity, calcification, amyloidosis, and stiffness of the large arterial wall.
- **•** Age-associated increase in arterial elastic fiber remodeling plays an important role in the pathogenesis of large arterial diseases, e.g., hypertension, atherosclerosis, and aneury



#### **Figure 1.**

Step by step illustration of proposed micro-and macro-assembly of elastic fibers known as elastogenesis, referred from previous reports (NivisoŽ Smith and Weiss, 2011; Pshezhetsky and Hinek, 2009; Schmelzer et al., 2020). (1) Tropoelastin is transcribed and translated from the ELN gene and is soluble and fragile. (2) Tropoelastin binds to EBP, limiting selfaggregation and degradation, and is secreted from vascular elastogenic cells; (3) Tropoelastin is released at the cell surface through competition by galactosugars in the microfibrils. Released tropoelastin is aligned and coacervated on the microfibrils while released EBP is eventually recycled back into the cell. (4) Fibulin binds tropoelastin aggregates and stabilize them. (5) LOX on the microfibrils processes tropoelastin in the extracellular space, which polymerizes tropoelastin to form mature insoluble ELN through desmosine and isodesmosine cross-links. (6) Finally, the elastic fiber is assembled, which consists of a core of ELN surrounded by microfibrils.



### **Figure 2.**

Elastic fiber remodeling and the aging vascular wall. A. Representative photomicrographs of EVG elastic staining in aortic walls from young (2mo, left) and old (30 mo, right) Fisher 344 cross-bred rats (FXBN), modified from previous finding (Wang and Lakatta, 2002). B. A list of adverse remodeling events in large aging arterial walls.