



Parasite Fauna of the White-Streaked Grouper (*Epinephelus ongus*) from the Thousand Islands, Java, Indonesia

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Abstract

Purpose Fish parasites can cause diseases in humans and lead to commercial losses in fisheries and aquaculture. The objectives of this study were to analyze *E. ongus*'s parasite fauna regarding food safety and parasite transmission risk between *Epinephelus* species and test whether *E. ongus* populations can be distinguished by their parasite community.

Methods We studied the metazoan parasite fauna of 30 white-streaked groupers *Epinephelus ongus* from the Thousand Islands, Java Sea, Indonesia, and compared the parasite community with specimens from Karimunjawa archipelago, Java Sea, from a former study. We used common fish parasitological methods for fish examination and parasite calculations.

Results We found 12 metazoan parasite species, establishing five new host and five new locality records, increasing the known parasite fauna of *E. ongus* by 21%. No anisakid worms infected *E. ongus*. All but one (trematode *Gyuliauchen* cf. *nahaensis*) species have been previously reported from *Epinephelus*. Parasite abundance of *E. ongus* differed significantly between the two regions.

Conclusions Due to a certain degree of host specificity to groupers, there is potential risk of parasite transmission from *E. ongus* into groupers in mariculture or surrounding fishes, which increases (sea) food security related health risks from zoonotic parasites and calls for better monitoring and management plans for *E. ongus*. The regional separation of the Thousand Islands and Karimunjawa with different food availability and fish ecology causes different parasite abundances, distinguishing two separate *E. ongus* populations by their parasite fauna.

Keywords Fish stock separation · Parasite diversity · *Gyuliauchen* cf. *nahaensis* · Grouper fisheries management · Seafood health risks · Food safety of fisheries products

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Introduction

With more than 17,000 islands and 80,791 km coastline, Indonesia is the biggest archipelago in the world [1]. Consequently, fisheries and aquaculture play an important role for the national economy [2]. In 2016, behind China, Indonesia ranked second in marine capture production with over 6 million tons [3].

Grouper (Serranidae: Epinephelinae) fisheries is of high economic value for food supply and the live reef fish trade in Indonesia, where they are mainly exported to China, Japan and Singapore [4]. Serranids also contribute to stable livelihoods in developing countries and are important for reef ecosystems as (top) predators [5, 6]. A rising demand for serranids increases fishing effort which drives their risk of extinction [7]. It is estimated that 19 grouper species are threatened and especially the live reef food fish trade remains largely unmanaged [7, 8]. Further knowledge on the ecology and life history of these commercially important species is needed to predict future implications of increasingly high fishing pressure.

Parasites are one of the most successful life forms on Earth [9]. They can impair their fish hosts and also human consumers when raw or undercooked fish is consumed, causing zoonosis as a disease transmitted from animals to humans [10]. The most well-known marine waterborne disease is anisakiasis [11] caused by nematodes of the family Anisakidae. Monitoring parasites of food fishes is important to continuously assess sea food health risks. Previous studies have suggested a low risk of anisakiasis in Indonesian waters [12–14], but climate change may shift marine boundaries causing different food web compositions in the future [15, 16] and there are also other marine helminths with zoonotic potential, e.g., *Hysterothylacium* or *Pseudoterranova* [13]. Diverse ecosystems such as Indonesia have huge potential for parasitological research [14]. Parasites provide new insights into fish diet, habitat

range and trophic position [17, 18] and inform about host ecology due to different infection pathways or accumulation of pollutants [19, 20]. Consequently, commercially important and often aquaculture species such as the groupers *Epinephelus coioides* and *E. fuscoguttatus* have been subject to a number of disease, parasite and environmental studies [19–26].

Today, fishing pressure is shifting toward smaller species such as *E. ongus*, which is now targeted as food fish in some regions, e.g., Japan [27, 28]. Demand for *E. ongus* has increased steadily [28, 29], also based on landings of *E. ongus* in Indonesia (Karimunjawa) [28, 30]. With only seven previous studies on the parasite fauna of *E. ongus* in South-East Asia and Australia [28, 31–36], we herewith sample this fish species from the Thousand Islands archipelago in close proximity to the heavily polluted Jakarta Bay. Our objectives for this study are to assess the transmission risk of parasites from *E. ongus* to congeners and vice versa and to determine parasite-borne health risks of *E. ongus* to consumers in terms of food safety and to mariculture fish. Additionally, we aim to test whether *E. ongus* populations can be distinguished by their parasite community. For that we compare our data to a former study where the parasite fauna of white-streaked groupers (similar in size and weight) was analyzed in Karimunjawa, an archipelago off the coast of Java, Indonesia, in 2013 by Neubert et al. [28] and discuss infection patterns, parasite biodiversity and fish ecology data.

Materials and Methods

Sample Collection

From April to June 2018, a total of 30 spearfished *Epinephelus ongus* were obtained from fishermen in the Thousand Islands archipelago (5°48′07.7″S 106°30′38.4″E), north coast of West Java, Indonesia (Fig. 1; Table 1). Fish were

Fig. 1 Map of study areas on Java, Indonesia. *TI* Thousand Islands, *KJ* Karimunjawa (samples taken in 2013 by Neubert et al. [28])



Table 1 Fish biological parameters of *Epinephelus ongus* from the Thousand Islands and Karimunjawa

Location	TL [cm]	SL [cm]	TW [g]	SW [g]	m	f	n.i
Thousand Islands	24.0 ± 2.8 (20.2–30.8)	20.5 ± 2.4 (16.9–26.6)	259.7 ± 99.5 (137.1–525.8)	241.6 ± 92.8 (127.3–494.2)	6	24	–
Karimunjawa ^a	25.5 ± 2.5 (20.5–30.4)	21.8 ± 2.3 (17.0–26.2)	266.3 ± 90.1 (108.5–498.0)	238.0 ± 80.5 (99.8–463.6)	23	7	5

SL standard length, SW slaughter weight, TL total length, TW total weight, $\bar{x} \pm SD$ mean \pm standard deviation, range in parentheses, f female, m male, n.i. not identified

^aData published by Neubert et al. [28] (sampled in May 2013, using the same methods as described for this study)

individually frozen in plastic bags, usually for 2–5 days, until examination inside the laboratory. Fish were defrosted in lukewarm water and morphometric data were taken (total length, standard length, total weight and gutted weight). Fluids from inside the plastic bags were examined individually under the stereo microscope (Zeiss, Stemi DV4).

Parasitological Examination

The body cavity was opened and internal organs were removed for subsequent endoparasite examination under the stereo microscope. Fish fillets were examined with translucent light under the stereo microscope. All detected parasites were cleaned from host tissue and collected in 70% ethanol. Selected specimens were transferred into glycerin on microscopic slides following Riemann [37]. Parasites were identified according to keys and original descriptions. After identification, the parasite specimens were deposited at LIPI, Science Centre, Jakarta-Bogor, Indonesia (Accession numbers: 45, 173–181, 542, 1482–1484).

Processing of Parasitological Data

Parasitological parameters prevalence (P), intensity (I), mean intensity (mI) and mean abundance (mA) followed Bush et al. [38]. The Shannon–Wiener diversity index [39] and Pielou index of evenness [40] were used to estimate the diversity of the metazoan parasite fauna, as well as the Berger–Parker index of dominance [41]. These indices were calculated for ecto- and endoparasites pooled, and endoparasites exclusively [22, 25, 26]. Ecto- to endoparasite ratio was calculated following Rückert et al. [42].

Statistical Analyses

All data, including the raw data by Neubert et al. [28], were analyzed using PRIMER software (version 7.0) to compare the parasite fauna of *E. ongus* sampled in this study in the Thousand Islands and sampled by Neubert et al. [28] in Karimunjawa. A total of 65 fish were compared and a similarity matrix was calculated by applying a Bray–Curtis similarity measure. The data for the Bray–Curtis index were transformed (fourth root = $\sqrt[4]{}$) prior to analysis. Fish

without parasites were omitted from the analysis. A multi-variant scaling plot (MDS) was used to display the relation between samples based on similarity matrices. The analysis of similarities (ANOSIM) was used to calculate the level of significance between both data sets. The differences between or within the data groups were given by the *R* value that ranged from – 1 to 1. An *R* value of 0 indicated no differences. A value of – 1 meant that the identical data point was located outside of the group, whereas with an *R* value of + 1 they were located within the group. The significance level was stated to $p < 0.05$. To identify which parasite taxa were responsible for the differences between the examined groups, SIMPER was applied. Additionally, a one-way ANOVA with Student's *T* test (SigmaPlot Version 11.0) was conducted to determine statistical differences in size of the fish between Karimunjawa [28] and the Thousand Islands.

Results

Parasite Community

A total of 12 parasite taxa infested *E. ongus* from the Thousand Islands. Four Crustacea taxa [*Alcirona* sp. (Hansen, 1890), *Caligus acanthopagri* (Lin, Ho and Chen, 1994), Gnathiidae indet. (Leach, 1814), Isopoda indet. (Latreille, 1817)], four Nematoda taxa [*Hysterothylacium* sp. (Ward and Magath, 1917), *Philometra epinepheli* (Dewi and Palm, 2013), *Philometra* sp. (Costa, 1845), Nematoda indet.], three Digenea taxa (*Cainocreadium epinepheli* (Durio and Menter, 1968), *Gyliuchen* cf. *nahaensis* [Ozaki, 1937, *Proso-rhynchus* sp. (Odhner, 1907)] and one Monogenea taxon [*Pseudorhabdosynochus* sp. (Yamaguti, 1958)] were found.

Ectoparasites represented the most predominant parasite group in *E. ongus*. The isopods *Alcirona* sp. and Gnathiidae indet. had the highest prevalences of 50.0% and 43.0%. Albeit a lower prevalence (23.3%), the Monogenea *Pseudorhabdosynochus* sp. had the highest mean abundance (5.4) and mean intensity (23.0) (Table 2). Fish which served as a host for this monogenean were highly infested (up to 83 worms per fish, Table 2).

With regard to endoparasites, the nematode *Hysterothylacium* sp. was the most common taxon with a prevalence

Table 2 Parasites from 30 specimens of *E. ongus* from the Thousand Islands sampled in 2018, including prevalence (P (%)), intensity (I), mean intensity (mI), mean abundance (mA), and diversity indices

Parasite/parasitological index	Stage	P (%)	mI	(I)	mA
Ectoparasites					
<i>Pseudorhabdosynochus</i> sp. (M)***	Adult	23.3	23.0	2–83	5.4
<i>Alcirona</i> sp. (Cr)***	Adult	50.0	2.1	1–7	1.0
<i>Caligus acanthopagri</i> (Cr)*	Adult	16.7	1.0	2	0.2
Isopoda indet. (Cr)	Adult	13.3	1.0	1–3	0.1
Gnathiidae indet. (Cr)***	Larval	43.3	1.4	1–3	0.6
Endoparasites					
<i>Cainocreadium epinepheli</i> (D)*	Adult	10.0	4.3	1–8	8.0
<i>Gyuliauchen</i> cf. <i>nahaensis</i> (D)**	Adult	16.7	1.6	1–4	0.3
<i>Prosorhynchus</i> sp. (D)*	Larval	6.7	1.0	1	0.1
<i>Hysterothylacium</i> sp. (N)***	Larval	20.0	1.5	1–3	0.3
<i>Philometra epinepheli</i> (N)***	Adult	3.3	1.0	1	0.0
<i>Philometra</i> sp. (N)*	Adult	10.0	1.3	1–2	0.1
Nematoda indet. (N)***	Larval	3.3	2.0	1	0.1
Parasitological indices					
Shannon–Wiener index of species diversity (total)		1.41			
Shannon–Wiener index of species diversity (endoparasites)		0.54			
Berger–Parker index of dominance (total)		0.62			
Berger–Parker index of dominance (endoparasites)		0.33			
Pielou index of evenness (total)		0.57			
Pielou index of evenness (endoparasites)		0.28			
Ecto- to endoparasite ratio		0.71			

Cr Crustacea, D Digenea, M Monogenea, N Nematoda

*New host records for *E. ongus*

**New host record for epinephelids

***Also found in Karimunjawa (Neubert et al. [28])

of 20%, followed by the digeneans *Gyuliauchen* cf. *nahaensis* and *Cainocreadium epinepheli* with prevalences of 16% and 10%, respectively. The digenean *Ca. epinepheli* was the most abundant taxon in the present study (see Table 1). *Gyuliauchen* cf. *nahaensis* (Table 3; Fig. 2), usually host specific to herbivorous fish (43), is herewith reported for the first time from an *Epinephelus* species.

Two taxa of *Philometra* were isolated from *E. ongus*, *Philometra epinepheli* (from the operculum) and *Philometra* sp. 1 (subcutaneous inside the mouth). The latter might represent a new species. It differed from already recorded *Philometra* species from groupers in Indonesia with two lip-like structures at the mouth, a distinctively shaped swollen esophagus, a different body length and site of infection. So far, only three *Philometra* species have been recorded from *E. ongus*: *P. epinepheli*, *P. cf. lateolabracis* and *P. ocularis* [28]. *Philometra* sp. 1 differs from *P. epinepheli* by larger gravid females (280 mm vs. 70–190 mm) and a different infection site (subcutaneous at mouth vs. subcutaneous at operculum) [22, 28, 44]. *Philometra* sp. 1 differs from *P. lateolabracis* in morphology (shorter uterus, ovaries do not reach up to the nerve ring) and in infection site (subcutaneous at mouth vs. gonads) [28, 45–47]. *Philometra* sp.

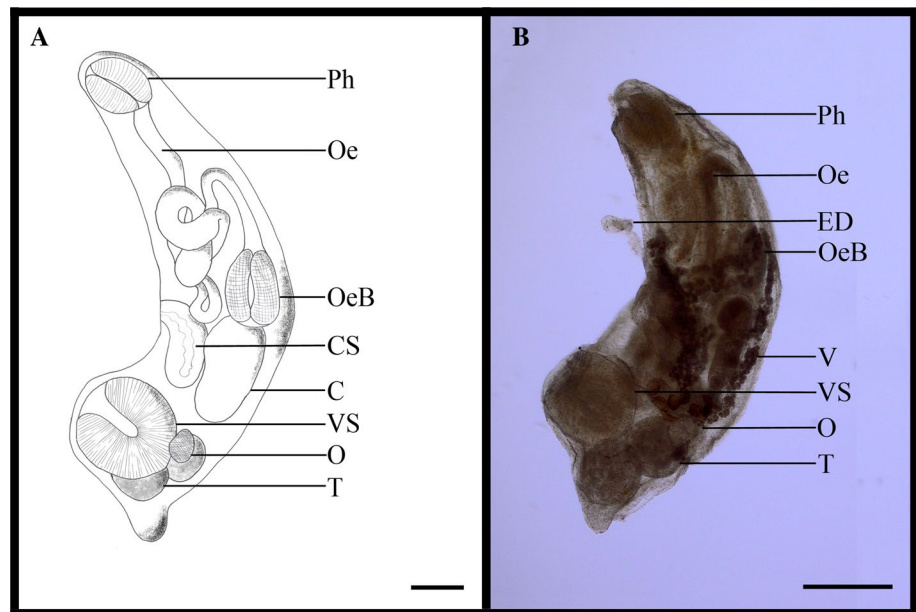
1 differs from *P. ocularis* by the lack of four large fleshy anterior papillae, a round mouth and in infection site (subcutaneous at mouth vs. eye cavity) [28, 48]. The herewith recorded *Philometra* sp. 1 shares most morphological characters (ovaries not reaching up to nerve ring, similar body length of 240 mm, comparable swollen esophagus) with *Philometra* sp. found by Kleinertz [22] in *E. coioides* and *E. areolatus*, and by Rückert [21] in *E. coioides* and *E. fuscoguttatus*. In Kleinertz [22] *Philometra* sp. 2 infested the gonads, gills, gill cavity, mesentery, mouth cavity, nose cavity and opercula, whereas in Rückert [21] *Philometra* sp. 3 infested the gills, gill cavity, mouth cavity and nose cavity. In both previous studies, *Philometra* sp. 2 and 3 were not found subcutaneous at the mouth.

The Berger–Parker index of dominance reached 0.62 when considering the total parasite fauna, but decreased to 0.3 for endoparasites only. The total parasite fauna had a high Shannon–Wiener index of diversity of 1.41. For only endoparasites, the Shannon–Wiener index decreased to 0.54. This trend goes along with a Pielou index of evenness, which was 0.57 including the ectoparasites and decreased to 0.28 for only endoparasites.

Table 3 Measurements of *Gyuliauchen* cf. *nahaensis* from *Epinephelus ongus* from the Thousand Islands compared to data published by Yamaguti [55]^a and Nahhas and Wetzel [59]^b

Physiological feature	<i>Gyuliauchen</i> cf. <i>nahaensis</i>		<i>Gyuliauchen nahaensis</i> ^a		<i>Gyuliauchen nahaensis</i> ^b	
	mm	Percent of body length	mm	Percent of body length	mm	Percent of body length
Host	<i>Epinephelus ongus</i>		<i>Acanthurus</i> sp.		<i>Siganus punctatus</i>	
Site of infection	Gills		Intestine		Intestine	
Locality	Thousand Islands, Indonesia		Makassar, Indonesia		Suva, Fiji	
Body L	0.94–1.67		1.65–2.40		1.30–2.45	
Body W	0.28–0.56	30.3	0.58–0.75	32.8	0.70–1.13	48.8
Oral sucker L	0.14–0.24	14.5	0.13–0.19	7.9	0.20–0.28	12.8
Oral sucker W	0.11–0.18	10.6	0.10–0.15	6.2	0.17–0.24	10.9
Pharynx L	0.14–0.22	13.6	0.15–0.25	9.9	0.22–0.39	16.3
Pharynx W	0.11–0.18	10.5	0.13–0.30	10.6	0.18–0.28	12.3
Ventral sucker L	0.18–0.32	18.5	0.25–0.36	15.1	0.27–0.43	18.7
Ventral sucker W	0.17–0.32	18.8	–	–	0.26–0.45	18.9
Ovary L	0.07	5.7	0.06–0.11	4.2	0.08–0.20	7.5
Ovary W	0.06	4.4	0.06–0.10	4.0	0.07–0.15	5.9
Eggs L	0.06–0.07	5.2	0.08	4.0	0.06–0.08	3.7
Eggs W	0.03–0.04	2.7	0.04–0.05	2.2	0.04–0.06	2.7
Testes L	0.21–0.23	16.9	0.11–0.23	8.4	0.18–0.33	13.6
Testes W	0.21–0.21	16.3	0.10–0.20	7.4	0.13–0.33	12.3

Fig. 2 Habitus of two specimens of *Gyuliauchen* cf. *nahaensis*. Scale bars **a** 0.2 mm, **b** 0.25 mm. *C* caeca, *CS* cirrus sack, *ED* ejaculatory duct, *Oe* esophagus, *OeB* esophagus bulb, *O* ovary, *Ph* pharynx, *T* testes, *V* vitellarium, *VS* ventral sucker

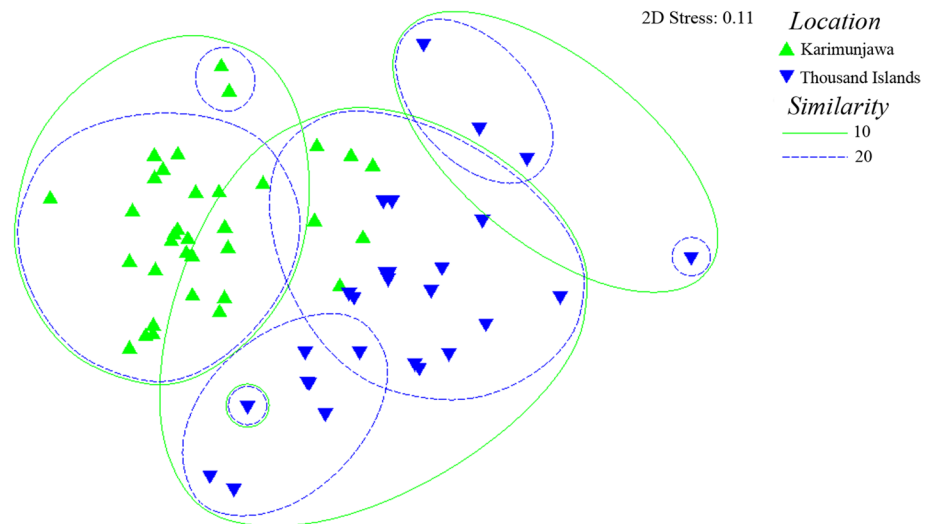


Comparison of *Epinephelus ongus* from the Thousand Islands and Karimunjawa

A combination of both data sets resulted in a total of 23 parasite taxa (17 in Karimunjawa and 12 in Thousand Islands) for *E. ongus* from Indonesia, 6 of them recorded for both locations (Table 2). The MDS plot based on parasite abundances shows that the infection patterns at both localities

were distinctively grouped (999 permutations) (Fig. 3). The data support a significant difference between both sampling sites (ANOSIM: $R=0.527$, $p < 0.05$), although some samples of Karimunjawa and the Thousand Islands overlap. The major taxa contributing to the separation of the two localities were the gill monogeneans *Pseudorhabdosynochus quadratus* and the crustaceans *Caligus* sp. found in Karimunjawa (SIMPER Analysis).

Fig. 3 MDS plot of the parasite fauna from *Epinephelus ongus* in Karimunjawa and the Thousand Islands based on parasite abundances. Data from Karimunjawa according to Neubert et al. [28]



Discussion

Parasite Records

The present study extends our knowledge on the parasite fauna of *E. ongus* by 21%. Five new host and five new locality records were established, with the digeneans *Cainocreadium epinepheli* and *Proisorhynchus* sp., the nematode *Philometra* sp. and the copepod *Caligus acanthopagri* reported for the first time from *E. ongus* and the digenean *Gyuliauchen* cf. *nahaensis* for the first time in *Epinephelus*. Additionally, the Thousand Islands represent a new locality record for *G.* cf. *nahaensis*, *Philometra epinepheli*, *Philometra* sp., *C. acanthopagri* and *Ca. epinepheli*. To date, seven earlier studies investigated the parasite fauna of the white-streaked grouper, two without any parasite records [32, 36], three reporting one parasite species [31, 33, 35] and two from Indonesia with 17 and eight taxa, respectively [28, 34]. Though Stolz [34] focused on ectoparasites only, the latter findings suggest that the parasite load in white-streaked groupers from Indonesia underlies considerable variability in the number of isolated parasite species.

Ectoparasites predominated the parasite fauna of *E. ongus* and the most abundant and prevalent one was the monogenean *Pseudorhabdosynochus* sp. Parasites of this genus are usually host specific and very common in groupers [49] with *P. quadratus* having been described from *E. ongus* from New Caledonia in the past [33]. The high infestation of these diplectanid monogeneans coincides with findings of other studies [28, 33, 34]. Another generalist was the ectoparasite *Alcirona* sp. which has already been recorded from, e.g., *E. areolatus*, *E. coioides*, *E. fuscoguttatus*, *E. quoyanus* and *Variola albimarginata*. *Caligus* spp. also infected *E. areolatus*, *E. coioides*, *E. corallicola*, *E. quoyanus* and *V. albimarginata* in Indonesia [28, 34].

Most of the isolated endoparasites have already been found in other serranids in Indonesian waters, e.g., *Ca. epinepheli*, *Philometra* spp., *Hysterothylacium* sp. or *Proisorhynchus* sp. [19–22, 25, 26, 34]. With three *Philometra* species so far isolated from *E. ongus*, this genus is also present in *E. areolatus*, *E. coioides* and *E. fuscoguttatus* and commonly infects groupers [28, 50]. In addition, the record of *Philometra* sp. that is suggested to represent a new species and awaits description adds to this biodiversity in groupers [21, 22, 44]. New specimen collections from *E. ongus* are required to confirm its real identity.

Here, similar to earlier studies of *E. ongus*, we did not find any anisakid nematodes and only Neubert et al. [28] reported *Anisakis typica* at a low prevalence of 2.9%. No nematodes infected the musculature, where the possible parasite transmission of zoonotic helminths to the consumer takes place. However, post-mortem migration of helminth larvae into the fillet can occur and *Hysterothylacium* sp. has caused zoonoses in the past, even though it originally infected the intestinal tract of host fishes [51]. With *Hysterothylacium* sp. larvae present in the studied white-streaked groupers, parasite-borne health risks should not be underestimated, but further studies are needed to better understand and evaluate the infection patterns of zoonotic parasites of *E. ongus*, which is a species of rising commercial interest.

With many parasite species having high host specificity to groupers, but a low host specificity within epinephelids, there is a risk of parasite transmission between *E. ongus* and its congeners. This is problematic in and around mariculture facilities, especially under consideration of increasing mariculture activities throughout the region. Consequently, the recorded parasite taxa in this non-cultivated grouper species might still lead to future economic losses [52, 53] in maricultured fish and also impacting wild fish stocks.

Ecological Parameters

The parasite fauna was predominated by the ectoparasite *Pseudorhabdosynochus* sp., implied by a Berger–Parker index of 0.62 for the total parasite fauna and a lower index of 0.3 for only endoparasites. Evenness and diversity indices were higher in the total parasite fauna compared with only endoparasites. Consequently, the taxa are more evenly distributed even when ectoparasites (including the dominant *Pseudorhabdosynochus* sp.) are considered. This can be explained from a relatively low amount of isolated endoparasites in *E. ongus* resulting in a very low Shannon–Wiener index of 0.54, compared to other grouper species (*E. coioides* up to 1.84, 20, *E. fuscoguttatus*). Both larger sized species have been sampled in more detail with up to 420 specimens per study e.g., [21, 22].

Differences in diet and habitat most likely result in the observed less diverse parasite fauna of *E. ongus* compared to other species [20, 24, 26]. Though diet and behavior of adult *E. coioides*, *E. fuscoguttatus* and *E. ongus* are approximately the same [54], the parasite composition is different [23, 24, 28]. Sampled specimens of *E. coioides* and *E. fuscoguttatus* often were juveniles, whereas our *E. ongus* were adults. Juvenile *E. coioides* and *E. fuscoguttatus* prefer more shallow waters and live over muddy bottoms, fine sediments or in seagrass beds [54, 55], while adult *E. ongus* live in deeper coral reef habitats and rocky bottoms, often in caves in up to 60 m depth [27, 55]. The different ecology, size and age of juvenile and adult *E. coioides* and *E. fuscoguttatus* favor a broad accumulation of parasites, which leads to a diverse and species-rich parasite fauna. Until 2016, 57 different parasite species have been recorded in, e.g., *E. coioides* [22, 26], and even from highly polluted regions around Jakarta Bay up to 31 parasite species were found [20]. This differs from *E. ongus*, with only 30 recorded parasite species so far. Neubert et al. [28] isolated only 17 species in *E. ongus* in Karimunjawa, a region where the biodiversity is believed to be particularly high [28, 29], suggesting that the observed differences in parasite diversity are real and not caused by biased sampling.

An interesting finding was the digenean *Gyuliauchen* cf. *nahaensis* in *E. ongus* (Gyuliauchenidae, Fig. 2; Table 2). This genus has been recorded in Indonesia only three times, in Sulawesi and Bali [14, 56] in herbivorous reef fish. Gyuliauchenid trematodes infect the intestine of their hosts in the Indo-West Pacific [57] and have almost exclusively been recorded from fishes from the herbivorous families Acanthuridae, Chaetodontidae, Pomacanthidae, Scaridae, Siganidae and Zanclidae [43, 58]. However, Srivastava [59] described the species *G. ozakii* from the intestine of the carnivorous predator *Harpodon nehereus* in the Arabian Sea [60]. Lopez [61] isolated *Gyuliauchen* sp. from the carnivorous grouper *Plectropomus leopardus* in the Philippines. Here, the finding

of *G. cf. nahaensis* in the gills of carnivorous *E. ongus* likely depicts an accidental infection, because this genus is very host specific and highly adapted to herbivorous fish [43]. Nevertheless, *G. cf. nahaensis* is a rare parasite in Indonesian waters and our findings reflect parasitological research potential in this region.

Regional Differences

The herewith analyzed fish specimens from the two archipelagos were similar in size and weight (Table 2). Size is an important factor for comparability because larger fish often have more endoparasites [26]. The fish from the two locations differed in sex ratio (Table 2, more males in Karimunjawa, more females in the Thousand Islands), but there is no consistency in the literature how or if sex affects parasite abundance in groupers. Samples were obtained 5 years apart: Karimunjawa in 2013 [28] and Thousand Islands in 2018. While pollution and anthropogenic effects can have an impact on the fish parasite fauna of groupers [20], previous studies in Indonesia have shown that seasonal change and natural inter-annual variability seem to have low impact on the metazoan fish parasite fauna [23, 26]. Therefore, we conclude that the samples were comparable and we suggest that the significant differences in the parasite abundances of *E. ongus* seen in the MDS plot (Fig. 3) are mainly caused by different biotic and abiotic conditions at both sites and are not biased by annual change or host factors.

While the MDS plot shows a clear separation of samples, some fish from Karimunjawa and the Thousand Islands are grouped together. Most likely this is because these samples harbored identical parasite species, as the locations share six common parasite species. This minor overlap in parasite abundance is not very surprising because we sampled the same species in a quite similar habitat—but it is all the more interesting to observe a significant difference in infection patterns in white-streaked groupers based on the sample location. The ectoparasites *Pseudorhabdosynochus quadratus* and *Caligus* sp. from Karimunjawa were the main factors for the discrepancies. Some gill parasites are thought to accumulate over time, depending on the age of the fish [28]. However, no significant differences in the total fish length between both sites could be detected (Student's *T* test: $p=0.719$). Despite high monogenean infestation, Karimunjawa is considered a healthy marine ecosystem due to high endoparasite diversity, high diversity indices and low ecto- to endoparasite ratios [28, 42]. While both localities are important spawning and nursery grounds for groupers [62, 63], Karimunjawa has established management plans, fishing guidelines and protected zones for grouper [29]. Grouper abundance might be higher there which might result in higher ectoparasite transmission and lead to higher ectoparasite infestation in this region.

E. ongus from Karimunjawa harbored one single Digenea species, whereas *E. ongus* from the Thousand Islands harbored three. The first intermediate hosts of this parasite group are gastropods. Yulianto [64] reported a sharp decrease of clams in Karimunjawa between 2005 and 2009, possibly linked to increased water pollution from coastal development [65] which may have also affected gastropod abundance. In Jakarta Bay, mollusk abundance decreased over seven decades due to anthropogenic pollution [66]. Finding three digenean taxa in the Thousand Islands suggests that pollution diluted from Jakarta Bay toward the more remote Thousand Islands [66], resulting in a sufficient gastropod abundance for the parasites to complete their life cycles.

Conclusion

The present study suggests that the parasite fauna of the small-sized white-streaked grouper has less species diversity compared to its larger sized congeners. Its parasite community differed between the Thousand Islands and Karimunjawa archipelagos shown in a comparison between samples from 2013 by Neubert et al. [28] and 2018. This is due to the differences in habitats and regional differences, leading to different ecological needs of the fishes which implies that the parasite composition of *E. ongus* can indicate different populations at the two sites. Fishing efforts on *E. ongus* is steadily increasing and threatens their occurrence throughout the Indonesian archipelago in recent years. This study suggests that the fish parasite community can indicate different origin of groupers and possibly distinct populations, supporting the development of better monitoring and management plans for this species in future. Finding numerous new host and locality records and parasites with a certain affinity to groupers, we suggest potential risks of parasite transmission within different grouper species, for example those cultivated in mariculture facilities or free living groupers. While this poses food safety concerns to consumers of fisheries and aquaculture products in Indonesia, increasing fishing efforts on *E. ongus* call for more research on this grouper species and better monitoring and management plans are needed.

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Availability of Data and Materials If wanted, we can share our raw data.

Code Availability Not applicable.

Compliance with Ethical Standards

Conflict of Interest On behalf of all authors, the corresponding author states that there is no conflict of interest or competing interests.

Ethics Approval The research was conducted under collaboration between IPB University (Bogor Agricultural University) and University of Rostock (under existing MoU).

Consent to Participate Not applicable.

Consent for Publication Not applicable.

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References

- Harris E (2001) Status of Indonesian fisheries today and the research needed. In Proceedings of the JSPS-DGHE International Symposium on Fisheries Science in tropical area. Faculty of Fisheries and Marine Science-IPB Bogor-Indonesia 62–66
- Tran N, Rodriguez UP, Chan CY, Phillips MJ, Mohan CV, Henriksson PJG, Koeshendrajana S, Suri S, Hall S (2017) Indonesian aquaculture futures: an analysis of fish supply and demand in Indonesia to 2030 and role of aquaculture using the AsiaFish model. *Mar Policy* 79:25–32. <https://doi.org/10.1016/j.marpol.2017.02.002>
- World Health Organization (2018) The state of food security and nutrition in the world 2018: building climate resilience for food security and nutrition. Food and Agriculture Organization

4. Afero F, Miao S, Perez AA (2010) Economic analysis of tiger grouper *Epinephelus fuscoguttatus* and humpback grouper *Cromileptes altivelis* commercial cage culture in Indonesia. *Aquacult Int* 18(5):725–739. <https://doi.org/10.1007/s10499-009-9295-x>
5. Yulianto I, Hammer C, Wiryawan B, Palm HW (2019) Cost-benefits of grouper *Epinephelus fuscoguttatus* (Forsskål, 1775) stock enhancement and sea-ranching in Indonesia. *Asian Fish Sci* 32(3):124–130
6. Sadovy de Mitcheson YJ, Linardich C, Barreiros JP, Ralph GM, Aguilar-Perera A, Afonso P, Erisman BE, Pollard DA, Fennessy ST, Bertonconi AA, Nair RJ, Rhodes KL, Francour P, Brulé T, Samoilys MA, Ferreira BP, Craig MT (2020) Valuable but vulnerable: over-fishing and under-management continue to threaten groupers so what now? *Mar Policy* 116:103909. <https://doi.org/10.1016/j.marpol.2020.103909>
7. Sadovy de Mitcheson YJ, Craig MT, Bertoncini AA, Carpenter KE, Cheung WL, Choat JH, Cornish AS, Fennessy ST, Ferreira BP, Heemstra PC, Liu M, Myers RF, Pollard DA, Rhodes KL, Rocha LA, Russell BC, Samoilys MA, Sanciangco J (2013) Fishing groupers towards extinction: a global assessment of threats and extinction risks in a billion dollar fishery. *Fish Fish* 14(2):119–136. <https://doi.org/10.1111/j.1467-2979.2011.00455.x>
8. Khasanah M, Nurdin N, Sadovy de Mitcheson Y, Jompa J (2020) Management of the grouper export trade in Indonesia. *Rev Fish Sci Aquacult* 28(1):1–15. <https://doi.org/10.1080/23308249.2018.1542420>
9. Poulin R, Morand S (2000) The diversity of parasites. *Q Rev Biol* 75(3):277–293. <https://doi.org/10.1086/393500>
10. Sličko TR, Smith HV, Rose JB (2000) Emerging parasite zoonoses associated with water and food. *Int J Parasitol* 30(12–13):1379–1393. [https://doi.org/10.1016/S0020-7519\(00\)00128-4](https://doi.org/10.1016/S0020-7519(00)00128-4)
11. Klimpel S, Palm HW (2011) Anisakid nematode (Ascaridoidea) life cycles and distribution: increasing zoonotic potential in the time of climate change? In: Mehlhorn H (ed) *Progress in Parasitology*. Springer, Berlin, Heidelberg, pp 201–222
12. Kleinertz S (2017) Modern applications in terms of biological indication, health of aquatic organisms and ecosystems. Cumulative Habilitation thesis, University of Rostock
13. Palm HW, Theisen S, Damriyasa IM, Oka IBM, Suratma A, Wibowo S, Kusmintarsih ES, Setyowati EA, Kleinertz S (2017) *Anisakis* Dujardin, 1845 (Nematoda: Ascaridoidea) from Indonesia. *Dis Aquat Org* 123(2):141–157. <https://doi.org/10.3354/dao3091>
14. Theisen S (2020) Indonesian marine fish parasite biodiversity. Dissertation, University of Rostock
15. Ullah H, Nagelkerken I, Goldenberg SU, Fordham DA (2018) Climate change could drive marine food web collapse through altered trophic flows and cyanobacterial proliferation. *PLoS Biol* 16(1):e2003446. <https://doi.org/10.1371/journal.pbio.2003446>
16. Champion C, Hobday AZ, Zhang X, Pecl GT, Tracey SR (2019) Changing windows of opportunity: past and future climate-driven shifts in temporal persistence of kingfish (*Seriola lalandi*) oceanographic habitat within south-eastern Australian bioregions. *Mar Freshw Res* 70(1):33–42. <https://doi.org/10.1071/MF17387>
17. Marcogliese DJ, Scholz T (1999) Recent visitations by eels to Sable Island, Canada, confirmed by parasites. *J Fish Biol* 54(3):685–687. <https://doi.org/10.1111/j.1095-8649.1999.tb00646.x>
18. Klimpel S, Rückert S (2005) Life cycle strategy of *Hysterothylacium aduncum* to become the most abundant anisakid fish nematode in the North Sea. *Parasitol Res* 97(2):141–149. <https://doi.org/10.1007/s00436-005-1407-6>
19. Palm HW, Rückert S (2009) A new approach to visualize ecosystem health by using parasites. *Parasitol Res* 105(2):539. <https://doi.org/10.1007/s00436-009-1423-z>
20. Neubert K, Yulianto I, Theisen S, Kleinertz S, Palm HW (2016) Parasite fauna of *Epinephelus coioides* (Hamilton, 1822) (Epinephelidae) as environmental indicator under heavily polluted conditions in Jakarta Bay, Indonesia. *Mar Poll Bull* 110(2):747–756. <https://doi.org/10.1016/j.marpolbul.2016.02.07>
21. Rückert S (2006) Marine fischparasiten in Indonesien: Befalls-situation und Bedeutung für die Marikultur von Zackenbarschen. Dissertation, University of Duesseldorf
22. Kleinertz S (2010) Fish parasites as bioindicators: environmental status of coastal marine ecosystems and a grouper mariculture farm in Indonesia. Dissertation, University of Bremen
23. Rückert S, Klimpel S, Palm HW (2010) Parasites of cultured and wild brown-marbled grouper *Epinephelus fuscoguttatus* (Forsskål, 1775) in Lampung Bay, Indonesia. *Aquacult Res* 41(8):1158–1169. <https://doi.org/10.1111/j.1365-2109.2009.02403.x>
24. Palm HW (2011) Fish parasites as biological indicators in a changing world: Can we monitor environmental impact and climate change? In: Mehlhorn H (ed) *Progress in Parasitology*. Springer, Berlin, Heidelberg, pp 223–250
25. Kleinertz S, Damriyasa M, Hagen W, Theisen S, Palm HW (2014) An environmental assessment of the parasite fauna of the reef-associated grouper *Epinephelus areolatus* from Indonesian waters. *J Helminthol* 88:50–63. <https://doi.org/10.1017/S0022149X12000715>
26. Kleinertz S, Palm HW (2015) Parasites of the grouper fish *Epinephelus coioides* (Serranidae) as potential environmental indicators in Indonesian coastal ecosystems. *J Helminthol* 89(1):86–99. <https://doi.org/10.1017/S0022149X1300062X>
27. Craig MT (2007) Preliminary observations on the life history of the white-streaked grouper, *Epinephelus ongus*, from Okinawa, Japan. *Ichthyol Res* 54(1):81–88. <https://doi.org/10.1007/s10228-006-0377-8>
28. Neubert K, Yulianto I, Kleinertz S, Theisen S, Wiryawan B, Palm HW (2016) Parasite fauna of white-streaked grouper, *Epinephelus ongus* (Bloch, 1790) (Epinephelidae) from Karimunjawa, Indonesia. *Parasitol Open* 2(e12):1–11. <https://doi.org/10.1017/pao.2016.6>
29. Yulianto I, Hammer C, Wiryawan B, Palm HW (2015) Fishing-induced groupers stock dynamics in Karimunjawa, Indonesia. *Fish Sci* 81(3):417–432. <https://doi.org/10.1007/s12562-015-0863-x>
30. Neubert K (2018) Parasiten von Zackenbarschen als biologische Indikatoren in Südostasien: anthropogene Verschmutzung und Aquakulturvedfahren. Dissertation, University of Rostock
31. Bray RA, Cribb TH, Barker SC (1996) Diploproctodaeinae (Digenea: Lepocreadiidae) from the coastal fishes of Queensland, Australia, with a review of the subfamily. *J Nat Hist* 30(3):317–366. <https://doi.org/10.1080/00222939600770191>
32. Nolan MJ, Cribb TH (2004) Two new blood flukes (Digenea: Sanguinicolidae) from Epinephelinae (Perciformes: Serranidae) of the Pacific Ocean. *Parasitol Int* 53(4):327–335. <https://doi.org/10.1016/j.parint.2004.05.002>
33. Schoelinc C, Justine JL (2011) *Pseudorhabdosynochus quadratus* n. sp. (Monogenea: Diplectanidae) from the white-streaked grouper *Epinephelus ongus* (Bloch) (Perciformes: Serranidae) off New Caledonia. *Syst Parasitol* 79(1):77. <https://doi.org/10.1007/s11230-011-9295-x>
34. Stolz H (2017) Makroskopisch sichtbare Parasitenarten kommerzieller Zackenbarsche von Bali, Indonesien: Konsequenzen für Vermarktung und Aufzucht in der Aquakultur. MSc Thesis, University of Rostock
35. Shamsi S, Chen Y, Poupá A, Ghadam M, Justine JL (2018) Occurrence of anisakid parasites in marine fishes and whales off New Caledonia. *Parasitol Res* 117(10):3195–3204. <https://doi.org/10.1007/s00436-018-6018-0>
36. Santos TRN, Sikkil PC (2019) Habitat associations of fish-parasitic gnathiid isopods in a shallow reef system in the central

- Philippines. *Mar Biodiv* 49(1):83–96. <https://doi.org/10.1007/s12526-017-0756-6>
37. Riemann F (1988) *Nematoda: Introduction to the study of meiofauna*. Smithsonian Institution Press, Washington DC
 38. Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997) Parasitology meets ecology on its own terms: Margolis et al. revisited. *J Parasitol* 83(4):75–583. <https://doi.org/10.2307/3284227>
 39. Shannon CE (1948) A mathematical theory of communication. *Bell Syst Tech J* 27(3):379–423. <https://doi.org/10.1002/j.1538-7305.1948.tb01338.x>
 40. Pielou DP, Matthewman WG (1966) The fauna of *Fomes fomentarius* (Linnaeus ex Fries) Kickx growing on dead birch in Gatineau Park, Quebec. *Can Entomol* 98(12):1308–1312. <https://doi.org/10.4039/Ent981308-12>
 41. Berger WH, Parker FL (1970) Diversity of planktonic foraminifera in deep-sea sediments. *Science* 168(3937):1345–1347. <https://doi.org/10.1126/science.168.3937.1345>
 42. Rückert S, Klimpel S, Al-Quraishy S, Mehlhorn H, Palm HW (2009) Transmission of fish parasites into grouper mariculture (Serranidae: *Epinephelus coioides* (Hamilton, 1822)) in Lampung Bay, Indonesia. *Parasitol Res* 104(3):523. <https://doi.org/10.1007/s00436-008-1226-7>
 43. Jones A, Bray RA, Gibson DI (2005) *Keys to the Trematoda*. Natural History Museum, London
 44. Dewi K, Palm HW (2017) Philometrid nematodes (Philometridae) of marine teleosts from Balinese waters, Indonesia, including the description of *Philometra damriyasai* sp. nov. *Zootaxa* 4341(4):577–584. <https://doi.org/10.11646/zootaxa.4341.4.10>
 45. Moravec F, Justine JL (2002) On two species of *Philometra* (Nematoda, Philometridae) from the serranid fish *Epinephelus septemfasciatus* in Japan. *Acta Parasitol* 34:40
 46. Moravec F, Glamuzina B, Marino G, Merella P, Di Cave D (2003) Occurrence of *Philometra lateolabracis* (Nematoda: Philometridae) in the gonads of marine perciform fishes in the Mediterranean region. *Dis Aquat Org* 53(3):267–269. <https://doi.org/10.3354/dao053267>
 47. Quiazon KMA, Yoshinaga T, Ogawa K (2008) Taxonomical study into two new species of *Philometra* (Nematoda: Philometridae) previously identified as *Philometra lateolabracis* (Yamaguti, 1935). *Folia Parasitol* 55(1):29–41
 48. Moravec F, Justine JL (2005) Two species of *Philometra* (Nematoda, Philometridae) from serranid fishes off New Caledonia. *Acta Parasitol* 50(4):323–331
 49. Justine JL, Sigura A (2007) Monogeneans of the malabar grouper *Epinephelus malabaricus* (Perciformes, Serranidae) off New Caledonia, with a description of six new species of *Pseudorhabdosynochus* (Monogenea: Diplectanidae). *Zootaxa* 1543:1–44
 50. Moravec F (2006) Dracunculoid and anguillicoloid nematodes parasitic in vertebrates. Academia, Prague
 51. Shamsi S, Steller E, Chen Y (2018) New and known zoonotic nematode larvae within selected fish species from Queensland waters in Australia. *Int J Food Microbiol* 272:73–82. <https://doi.org/10.1016/j.ijfoodmicro.2018.03.007>
 52. Tucker CS, Sommerville C, Wootten R (2002) Does size really matter? Effects of fish surface area on the settlement and initial survival of *Lepeophtheirus salmonis*, an ectoparasite of Atlantic salmon *Salmo salar*. *Dis Aquat Org* 49(2):145–152. <https://doi.org/10.3354/dao049145>
 53. Ogawa K (2015) Diseases of cultured marine fishes caused by Platyhelminthes (Monogenea, Digenea, Cestoda). *Parasitol* 142(1):178–195. <https://doi.org/10.1017/S0031182014000808>
 54. Heemstra PC, Randall JE (1993) *FAO Species Catalogue: Groupers of the world (family Serranidae, subfamily Epinephelinae)*. An annotated and illustrated catalogue of the grouper, rockcod, hind, coral grouper and lyretail species known to date. *FAO Fish Synop* 125(16):382
 55. Froese R, Pauly D (2020) *FishBase*. <http://www.fishbase.org>. Accessed 23 Apr 2020
 56. Yamaguti S (1953) Parasitic worms mainly from Celebes. Part 3. Digenetic trematodes of fishes 2. *Acta Med Okayama* 8(3):258–295. <https://doi.org/10.18926/AMO/31875>
 57. Al-Jahdali MO, Hassanine RES (2012) The life cycle of *Gyuliauchen volubilis* Nagaty, 1956 (Digenea: Gyuliauchenidae) from the Red Sea. *J Helminthol* 86(2):165–172. <https://doi.org/10.1017/S0022149X11000186>
 58. Bray R, Waeschenbach A, Cribb T, Weedall G, Dyal P, Littlewood D (2009) The phylogeny of the Lepocreadioidea (Platyhelminthes, Digenea) inferred from nuclear and mitochondrial genes: Implications for their systematics and evolution. *Acta Parasitol* 54(4):310–329. <https://doi.org/10.2478/s11686-009-0045-z>
 59. Srivastava HD (1938) Studies on the amphistomatous parasites of Indian food fishes. Part 2. A new trematode of the genus *Gyuliauchen* Nicoll from an Indian marine fish. *Indian J Vet Sci Anim Husband* 8(4):399–401
 60. Nahhas FM, Wetzel JA (1995) Digenetic trematodes of marine fishes from Suva, Fiji: The family Gyuliauchenidae Ozaki, 1933. *J Helminthol Soc Washington* 62(2):117–130
 61. Lopez NC (1998) Parasites of some serranid fishes from Manila Bay (Philippines). *Philipp Sci* 35:43–48
 62. Hartoko A, Chayaningrum S, Febrianti DA, Ariyanto D (2015) Carbon biomass algorithms development for mangrove vegetation in Kemujan, Parang Island Karimunjawa National Park and Demak coastal area, Indonesia. *Proc Environ Sci* 23:39–47. <https://doi.org/10.1016/j.proenv.2015.01.007>
 63. Hidayati N, Suparmoko M (2018) Fish assemblage structure in relation to seagrass bed in Tidung Kecil Island, Kepulauan Seribu. *E3S Web of Conferences* 74:02005. <https://doi.org/10.1051/e3sconf/20187402005>
 64. Yulianto I, Kartawijaya T, Susanto HA, Cambell S (2010) The effectiveness of Karimunjawa. *Proceeding of International Symposium on Small Islands and Coral Reefs*
 65. Campbell SJ, Kartawijaya T, Yulianto I, Prasetya R, Clifton J (2013) Co-management approaches and incentives improve management effectiveness in the Karimunjawa Park, Indonesia. *Mar Policy* 41:72–79. <https://doi.org/10.1016/j.marpol.2012.12.022>
 66. van der Meij SE, Moolenbeek RG, Hoeksema BW (2009) Decline of the Jakarta Bay molluscan fauna linked to human impact. *Mar Poll Bull* 59(4–7):101–107. <https://doi.org/10.1016/j.marpolbul.2009.02.021>

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