

# **HHS Public Access**

Author manuscript *Ultrasound Med Biol.* Author manuscript; available in PMC 2022 July 01.

Published in final edited form as:

Ultrasound Med Biol. 2021 July ; 47(7): 1920–1930. doi:10.1016/j.ultrasmedbio.2021.03.011.

# Sonographic features of abscess maturation in a porcine model

Daniel F. Leotta<sup>a</sup>, Matthew Bruce<sup>a</sup>, Yak-Nam Wang<sup>a</sup>, John Kucewicz<sup>a</sup>, Tatiana Khokhlova<sup>b</sup>, Keith Chan<sup>c</sup>, Wayne Monsky<sup>c</sup>, Thomas J. Matula<sup>a</sup>

<sup>a</sup>Applied Physics Laboratory, University of Washington, Seattle, WA 98105, USA

<sup>b</sup>Department of Gastroenterology, University of Washington, Seattle, WA 98105, USA

<sup>c</sup>Department of Radiology, University of Washington, Seattle, WA 98105, USA

## Abstract

Abscesses are walled-off collections of infected fluids that often develop as complications in the setting of surgery and trauma. Treatment is usually limited to percutaneous catheterization with a course of antibiotics. As an alternative to current treatment strategies, a histotripsy approach was developed and tested in a novel porcine animal model. The goal of the current work is to use advanced ultrasound imaging modes to extract sonographic features associated with the progression of abscess development in a porcine model. Intramuscular or subcutaneous injections of a bimicrobial bacteria mixture plus dextran particles as an irritant led to identifiable abscesses over a 2- to 3-week period. Selected abscesses were imaged at least weekly with B-mode, 3D B-mode, shear-wave elastography (SWE) and plane-wave Doppler imaging. Mature abscesses were characterized by a well-defined core of varying echogenicity surrounded by a hypoechoic capsule that was highly vascularized on Doppler imaging. 3D imaging demonstrates the natural history of abscess morphology, with the abscess becoming less complex in shape and increasing in volume. Furthermore, SWE demonstrated variations in stiffness as phlegmon becomes abscess and then liquifies, over time. These ultrasound features potentially provide biomarkers to aid selection of treatment strategies for abscesses.

## Keywords

Abscess; Ultrasound; Three-Dimensional; Curvature; Shear Wave Elastography; Doppler Imaging; Plane-Wave Doppler

## INTRODUCTION

Infected abscesses are walled-off collections of inflammatory cells, cellular debris, and bacteria. They often develop as complications in the setting of surgery, trauma, systemic infections, and other disease states. Clinical management of an abscess depends on size,

Corresponding Author: Daniel F. Leotta, PhD, University of Washington, Applied Physics Laboratory, Box 355640, Seattle, WA 98105, USA, Phone (206) 616-6787, leotta@uw.edu.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

location, composition, complexity, and patient comorbidities. Abscesses can result in significant morbidity, overwhelming sepsis, and death if inadequately treated. For deeper abscesses, current treatment is typically limited to antibiotics with long-term catheter drainage, or surgical wash-out when inaccessible to percutaneous drainage or unresponsive to initial care efforts. Ultrasound imaging can assist in reliable identification of abscesses so that appropriate treatment is applied (O'Rourke et al. 2015). For example, ultrasound can be used to distinguish abscesses from other soft-tissue pathologies such as phlegmon, cysts, hematomas, and cellulitis. It can also be useful in planning therapy strategies by establishing extent, maturity, viscosity of the fluid, and structural features such as loculations (internal chambers).

Variations in echogenicity observed within abscesses have been noted in B-mode imaging studies, which can make definitive diagnosis challenging (Kuligowska et al. 1982). Contrast ultrasound studies have been used to distinguish between inflammatory tissue that may represent early infections (phlegmons), and fully developed mature abscesses by measuring enhancement within the infected tissue (Ripollés et al. 2011, 2013). Full enhancement within the tissue without liquefied components signifies a phlegmon, while lack of internal enhancement is a sign of a mature abscess containing potentially-drainable fluid. Blood flow surrounding the core of an abscess has also been reported with power Doppler imaging (Arlsan et al. 1998) and with contrast-enhanced ultrasound (Liu et al. 2008; Lu et al. 2019). Elastography has been described as a potential ultrasound modality to diagnose and characterize abscesses (Gaspari et al. 2009).

Both B-mode and contrast imaging have been used to classify abscess maturity by comparing imaging features observed in abscesses across different patients (Kunze et al. 2015). However, progression of abscess development as observed by ultrasound has yet to be described. The objective of this study was to use the latest advances in ultrasound technologies to extract abscess features during maturation in a novel porcine animal model (Wang et al. 2020). This animal model was developed to test a histotripsy approach to treating abscesses as an alternative to current treatment strategies (Matula et al. 2020). In conjunction with this histotripsy study, ultrasound imaging was performed at least weekly to evaluate abscess maturation following intramuscular injection of bacteria. The controlled induction of the abscess allowed for imaging at definitive time points in abscess development. Abscesses were imaged during the maturation process up to 4 weeks post-injection using B-mode, 3D B-mode, elastography and Doppler ultrasound, to evaluate changes in echogenicity, size, shape, stiffness, and local vasculature.

Longitudinal imaging was performed to identify sonographic features of 14 abscesses during the maturation process, and detailed image analysis was performed on two representative abscess cases. The first case was followed over a 3-week period and imaged at maturity in the third week with B-mode and Doppler. The second case was followed over a 4-week period using B-mode, 3D B-mode, shear-wave elastography, and plane wave Doppler. This is the first example in which multiple ultrasound modalities have been used together to longitudinally extract features from a maturing abscess.

## MATERIALS AND METHODS

#### Animal Model

All procedures were approved by the Institutional Animal Care and Use Committee (IACUC) at R&R Rabbitry (Stanwood, WA). Two female domestic swine (Yorkshire/ Hampshire cross, age 3–6 months, weight 45–65 kg) were used in this study. In a recent paper (Wang et al. 2020) we described the development of a porcine animal model in which multiple large multiloculated bimicrobial abscesses can be formed at distinct sites in the same animal. These abscesses satisfy the true definition of an abscess and are formed within a short period. Briefly, on the day of inoculation, the bacterial mix, consisting of E. coli (strain ATCC 25922, American Type and Culture Collection, MD, USA), B. fragilis strain (ATCC 23745, American Type and Culture Collection, MD, USA) and sterile dextran microcarrier bead suspension in a ratio of 1:1:2 was prepared so that the final inoculum contained approximately 106 cfu/ml E. coli and  $10^6$  cfu/ml B. fragilis. Bacterial concentration was determined using spectrophotometric absorbance readings and confirmed using Compact  $Dry^{TM}$  EC100 and TC assay plates (Hardy Diagnostics, Santa Maria, CA USA).

Animals were sedated (Telazol/Ketamine/Xylazine) and maintained under a surgical plane of anesthesia with isoflurane. All animals were instrumented to monitor heart rate, ECG and blood oxygen saturation and temperature. The haunches, abdomen and in one animal the neck were depilated, cleaned and sterilized with consecutive rounds of isopropyl alcohol. Injections were performed under ultrasound guidance in up to 8 separate sites in a single animal: 1.5 cm deep into the left and right biceps femoris and brachiocephalicus, and subcutaneously on either side of the midline. Before the intramuscular injections, the tip of the needle was moved back and forth approximately 1 cm fifty times, in a fanning motion, to create a small pocket of injury before injecting. For each animal 10 ml of the inoculate were injected at each site and the location was superficially marked with tissue-marking dye. After the inoculations, the animals were recovered and monitored every day for signs of pain or distress, reduction in appetite, loss of weight and mobility. Abscesses were allowed to form over 3–4 weeks.

#### Ultrasound imaging of abscesses

Growth and development of a total of 14 abscesses in the two animals were monitored at least weekly with ultrasound imaging using an Aixplorer (Supersonic Imagine, Aix-en-Provence, France) with the SL15–4 linear probe. Abscesses grew to different sizes over the course of 3 to 4 weeks, with the largest abscess measuring  $125 \times 71 \times 43$  mm. As the study progressed, detailed imaging was performed only on larger abscesses that were identified as the most clinically-relevant for histotripsy studies (greater than 3 cm in at least one dimension). Plane-wave Doppler imaging (Angio PLUS) was used to follow changes in abscess vascularity and provided enhanced sensitivity to low-velocity blood flow over conventional color-flow Doppler imaging (Bercoff et al. 2011; Liu et al. 2019). Shear wave elastography (SWE) (Sigrist et al. 2017) was used to follow changes in elasticity during abscess evolution. In some cases, palpation with the ultrasound probe demonstrated movement of the abscess contents as a gross indication of the liquidity of the pus. For one

animal three-dimensional B-mode imaging was performed by tracking the ultrasound scanhead tracked with a magnetic tracking system (trakSTAR, NDI, Waterloo, ON, Canada). Custom software recorded the 3D location and orientation of a series of 2D image planes acquired while scanning the abscess. The image-capture software was programmed using MATLAB (MathWorks, Natick, MA, USA), which synchronously recorded the 3D tracker readings and the ultrasound video output through a frame-grabber (USB3HDCAP, StarTech.com Ltd., Groveport, OH, USA) at a rate of 60 frames per second.

#### Image analysis

Detailed image analysis was performed on two representative abscess cases, to first illustrate the general characteristics of a mature abscess, and then to quantify the changes in size, shape and composition during maturation. The first case is an intramuscular injection that progressed to a compact abscess over the course of 3 weeks. The second case is an intramuscular injection in a second animal that progressed to an elongated abscess over the course of 4 weeks.

The first case (ABS1) was scanned with B-mode and Doppler imaging up to 3 weeks (23 days) post-injection. A 3D reconstruction of the plane-wave directional color power Doppler images at 20 days post-injection was generated from a slow untracked sweep across the full extent of the abscess, assuming a linear scan at constant speed. A total of 605 2D images were acquired during this sweep. This abscess was chosen for 3D Doppler reconstruction because its intra-muscular location kept it free of respiratory flash artifact during image acquisition. Since the full abscess could be visualized in a single 2D field of view for this smaller abscess, the sweep distance was determined from size measurements derived from 2D images. The color pixels on each image were automatically segmented from the background so that the 3D reconstruction represents the Doppler component only.

The second case (ABS2) was imaged at four time points corresponding to weeks 1 through 4 post-injection (days 8, 15, 20 and 29 post-injection). This abscess was scanned in B-mode at weeks 2, 3 and 4 post-injection with the 3D image capture system. The total number of images collected were 1212, 1783, and 341 for the scans at weeks 2 through 4, respectively. These tracked scans were used to create grayscale volume reconstructions by compiling the series of 2D images in a regular 3D grid (Leotta and Martin 2000). Manual image segmentation for a series of axial planes in the reconstructed volume (Leotta et al. 2018) was performed with custom software (Legget et al. 1998) to construct surface mesh models of the outer capsule and the inner core (Leotta et al. 2001). Manual segmentation was performed for every 20<sup>th</sup> axial plane of the volume reconstructions (spatial step sizes of 0.83 mm, 1.56 mm and 1.61 mm for weeks 2, 3 and 4 respectively). The surface models were analyzed with the MeshLab open-source analysis package (Cignoni et al. 2008) to quantify the dimensions (length, width, height) and volume of both the total abscess size (outer extent of the capsule) and the core. As a measure of the surface complexity, MeshLab was also used to calculate the mean curvature of the surface reconstruction of the abscess core at each time point. For each surface model, the local curvature was calculated at each mesh vertex using the method described by Meyer et al. (2003). Curvature is zero for a flat surface,

positive for a convex surface, and negative for a concave surface; increasing magnitude signifies greater deviation from a plane.

For case ABS2, shear wave elastography was performed at weeks 1 through 4. At weeks 2 and 3, slow freehand sweeps at constant speed were acquired along the length of the abscess, imaged in cross-section. The 3D tracking system was not used during these scans. Instead, 3D reconstructions were approximated by compiling the image series into a 3D volume assuming a linear scan at a constant sweep speed. The scan distance was determined from the 3D reconstructions of the tracked B-mode scans. The total number of images compiled for each scan was 862 and 1584 for weeks 2 and 3 respectively. SWE values for the abscess core were extracted by 1) isolating the core in the 3D volume by manual segmentation, and 2) converting the color pixels within the core to stiffness values in kPa according to the SWE color map.

For case ABS2 it was possible to use probe palpation to visualize the motion of lowerviscosity liquid components under B-mode when the abscess was pushed on by the ultrasound probe. A video clip acquired at week 3 post-injection showing this effect is presented as supplemental data (Video 1). From a portion of the video, particle tracking was used to measure the motion of the liquid pus within the core. For this analysis, particle displacement direction and magnitude were tracked over 13 sequential video frames using optical flow analysis (Fleet and Weiss 2006) as the probe pushed on the abscess. Optical flow is a video processing method for tracking movement within a video sequence based on spatial and temporal image gradients. The Lucas-Kanade optical flow method was used, which uses least-squares regression to solve the optical flow equation for regions of pixels rather than individual pixels (Lucas and Kanade 1981).

Doppler images were also acquired each week for case ABS2. Doppler imaging modes included standard color flow, directional color power, and plane-wave directional color power.

## Histopathology

Histological analysis of all abscesses was performed at the end of the study period. After the animals were euthanized, the abscesses were carefully removed *en bloc* and fixed in 10% neutral buffered formalin for histological evaluation. The fixed lesion was grossed and processed to maintain the lesion as whole cross-sections as much as possible. Serial sections were stained with Hematoxylin and Eosin (H&E) for general tissue morphology, Masson's trichrome stain to visualize the presence of a fibrous capsule, Gram stain to evaluate the presence of bacteria, or labeled with anti-CD31 antibodies to visualize blood vessels. The lesions were evaluated for the presence of a connective tissue capsule; the extent of the capsule; the presence of infected material and cell debris; and blood vessels.

## RESULTS

#### Abscess maturity

For all 14 abscesses imaged in this study the initial bacteria inoculation led to a pocket with an identifiable capsule within 2–3 weeks. Two-dimensional B-mode and Doppler images are

shown in Figure 1 for abscess case ABS1 at 3 weeks (23 days) post-injection. The B-mode image shows the abscess core surrounded by a fibrous capsule. Doppler imaging shows a dense network of blood vessels within the capsule but not in the abscess core. The 3D distribution of the blood vessels in the capsule is shown in Figure 2 at 21 days post-injection. A video clip showing a rotational view of this 3D vascular reconstruction is presented as supplemental data (Video 2). This abscess is representative of the general features that were visualized at maturation for all 14 abscesses: a well-circumscribed, avascular, mixed echogenic core surrounded by a vascularized hypoechoic capsule.

#### Longitudinal study

The series of cross-sectional B-mode images in Figure 3 show the maturation of case ABS2 over a 4-week period. The abscess evolves from diffuse inflammatory tissue (phlegmon) at 1-week post-injection into an organized fluid collection with a well-defined capsule by week 3. Volume renderings of the 3D grayscale reconstructions (Figure 4) illustrate the morphological development over time.

Size and volume changes over time are quantified from the 3D surface reconstructions of the outer extent of the capsule and of the inner core (Figure 5). Table 1 shows the dimensions (Length  $\times$  Width  $\times$  Height) and the volume for both the entire abscess and for the abscess core over time. The length of the abscess decreased over time (102 mm at week 2 to 68 mm at week 4) as the volume increased (20.4 ml at week 2 to 37.2 ml at week 4).

Curvature measurements of the core surface over time show a change from an irregular border to a rounded and more well-defined border (Figure 6). Histograms of the curvature measurements at each vertex of the surface mesh (Figure 6) show that the spread in the curvature measurements decreases over time (standard deviation 0.37/mm at week 2 to 0.14/mm at week 4, Table 1). The curvature mean is approximately zero at each time point, indicating a nearly equal distribution of convex and concave curvatures.

Cross-sectional SWE images show a reduction in stiffness over the 4-week time period as the abscess liquefies (Figure 7). At week 4 the abscess core is generally a viscous liquid, mixed with regions of solid tissue components exhibiting increased stiffness. Transparent 3D volume renderings were produced for the shear-wave measurements in weeks 2 and 3 of the abscess core, based on the untracked cross-sectional sweeps (Figure 8). These 3D renderings show the distribution of stiffness over the entire abscess and illustrate the overall trend toward lower stiffness as the abscess matures and liquefies. This stiffness change is quantified in the histograms in Figure 9, showing the shift toward lower stiffness from week 2 to week 3 (43 kPa to 19 kPa, Table 1). The histograms in Figure 9 also show an increase in the proportion of the core for which no shear wave value was measured (values of zero on the SWE scale).

The result of optical-flow analysis of the cine-loop acquired during compression with the probe is shown in Figure 10a. The yellow arrows show the direction and relative magnitude of the measured displacements; the largest arrow corresponds to a 1 mm displacement. The regions with detected motion indicate fluid displacements around solid components. Regional differences in stiffness were also observed within the abscess core with SWE

(Figure 10b) that correlate with the stationary region around which the internal flow was measured. On histological analysis, this stationary region with higher stiffness was shown to be composed of muscle tissue (Figure 10c).

Doppler imaging for case ABS2 showed the development over time of a network of blood vessels within the hypoechoic capsule (Figure 11). At weeks 1–2 blood flow was observed throughout the abscess. At week 4 a dense network of vessels is seen within the capsule, organized around the avascular core. Immunohistochemistry for this case shows the details of the vascular network on both the deep and lateral sides of the capsule that matched the Doppler images (Figure 12). No blood vessels were observed within the abscess core.

## DISCUSSION

As an abscess matures from a region of inflammation (phlegmon) to an encapsulated organized collection of viscous pus, it consolidates and a fibrous capsule is formed around the purulent material. The maturation process includes angiogenesis of permeable vessels in the capsule, through which inflammatory cells (neutrophils, lymphocytes, macrophages) enter the infected area. Subsequently the abscess core liquefies as the inflammatory cells clear the infection, and the abscess consolidates and retracts.

For this study the evolution of a single abscess was followed from injection of an inoculate to maturity using multiple ultrasound imaging approaches. This is the first study in which an abscess has been followed longitudinally at known time points from inception, with demonstration of the changes in imaging features as the abscess evolves and liquefies. Longitudinal monitoring of induced abscesses by ultrasound to determine maturity can be useful in planning of treatment times in animal studies.

This study also demonstrates the use of complementary ultrasound modalities to characterize the maturation of a specific abscess. Standard B-mode shows how the complexity of an abscess evolves, and 3D reconstructions from B-mode scans are used to quantify the changes in morphology. SWE imaging quantifies changes in stiffness over time and Doppler shows vascular development. Clinically, abscess features relating to complexity and liquefaction may predict the success of drain placement and catheter drainage. The 14 abscesses we followed generally demonstrated the features that we describe here. However, because each abscess evolves uniquely, we illustrated the general maturation process with a detailed study of a single abscess.

In our animal model, the injection of bacteria and dextran initially resulted in the development of a phlegmon that then evolved into an organized mature abscess over 2–3 weeks. B-mode ultrasound allowed us to identify key structural characteristics, namely a well-defined hypoechoic capsule surrounding a core with mixed echogenic material, which were used as an indicator of maturity (Lu et al. 2019). Doppler imaging was used to further evaluate abscess progression to maturity. Vascularity was initially observed throughout the phlegmon/abscess, which then progressed to a vascularized capsule and an avascular core.

In general, the pig abscesses were more echogenic than often observed in humans, although echogenicity of human abscesses can vary (Kuligowska et al. 1982). One contributing factor

may be the addition of dextran particles (~150  $\mu$ m) to the inoculate, which likely increases scattering and makes the abscess appear brighter. The purpose of the dextran was to elicit a persistent inflammatory response to aid in the reliable generation of an abscess (Wang et al. 2020).

The surface reconstructions from 3D B-mode scans allow quantification of size, volume, and core curvature over time (Figures 5, 6). Surface curvature analysis of the core shows an organizational change in configuration as the abscess matures. The reduced spread of the curvature measurements over time signify a reduction in sharp bends in the surface and a general trend toward a more smooth and rounded shape. Similar curvature analysis has been used to describe changes in surface complexity in the developing fetal brain (Hu et al. 2013), and to distinguish between different kidney stone types (Duan et al. 2013). These abscess curvature changes match the development over time previously described for liver abscesses (Kunze et al. 2015). This curvature change may signify the transition from irregular inflammation to a discrete encapsulated abscess and subsequent retraction as the abscess resolves or is being cleared by inflammatory cells and macrophages. These changes are commonly seen clinically.

Shear wave elastography shows reduction in core stiffness during abscess maturation, with the largest change taking place between weeks 2 and 3 (Figure 7). The 3D volume reconstructions of SWE scans at weeks 2 and 3 quantify this stiffness change and demonstrate that the change generally occurs throughout the abscess core (Figure 8). This time course in liquefaction is consistent with what is observed clinically. Reduced stiffness corresponding to liquefaction would suggest the abscess is amenable to catheter placement and drainage.

The internal flow observed with external compression at week 3 indicates the presence of lower-viscosity liquid components in the core. Internal flow with compression has been noted as a component of the clinical ultrasound examination of abscesses that helps distinguish an abscess from other conditions, such as cellulitis and phlegmon (Adhikari and Blaivas 2012), and also helps determine if it can be adequately evacuated with drainage catheters. SWE at this site also demonstrated the heterogeneity of the abscess core. Histology confirmed that the region with higher stiffness and lack of flow with compression was muscle tissue, distinct from the general viscous content of the abscess core (Figure 10). Detecting regional variations in stiffness may have clinical implications for successful drainage by providing guidance for drain placement.

Doppler imaging shows development of a vascularized capsule with an avascular core (Figure 11), which were confirmed by histology (Figure 12). Abscess rim enhancement has also been observed with contrast-enhanced ultrasound (Kunze et al. 2015; Liu et al. 2008). This is the equivalent of the enhancing rim or capsule seen with contrast enhanced CT or MRI (Mavilia et al. 2016), which is used clinically to determine if a fluid collection is an abscess or sterile fluid such as a seroma, ascites or effusion.

## CONCLUSION

Controlled induction of abscesses in a large animal model allowed for tracking of abscess development by ultrasound imaging at definitive time points in the maturation process. Serial imaging demonstrated the progression from a site of diffuse inflammation to an organized, avascular core of varying echogenicity, surrounded by a hypoechoic, highly vascularized capsule. Stiffness measurements also quantified the trend toward liquefication during the maturation process. Identification of abscess maturity is of clinical relevance because the core must be relatively liquefied in order for drainage to be successful. The changes in abscess morphology, stiffness and vascularity over time demonstrated in this study represent ultrasound imaging features that can potentially identify abscess maturity and guide treatment strategies.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

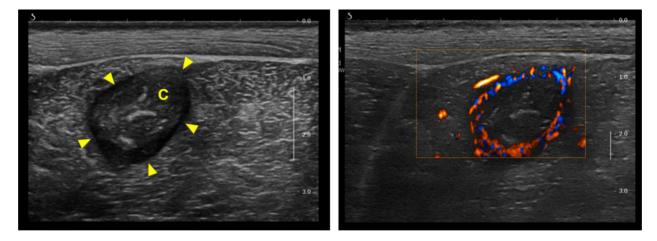
Work supported in part by NIH grants 5R01EB019365, R01GM122859 and K01 DK104854.

## REFERENCES

- Adhikari S, Blaivas M. Sonography first for subcutaneous abscess and cellulitis evaluation. J Ultrasound Med 2012;31:1509–12. [PubMed: 23011612]
- Arslan H, Sakarya ME, Bozkurt M, Unal O, Dilek ON, Harman M. The role of power Doppler sonography in the evaluation of superficial soft tissue abscesses. Eur J Ultrasound 1998;8:101–6. [PubMed: 9845787]
- Bercoff J, Montaldo G, Loupas T, Savery D, Mézière F, Fink M, Tanter M. Ultrafast compound Doppler imaging: providing full blood flow characterization. IEEE Trans Ultrason Ferroelectr Freq Control 2011;58:134–47. [PubMed: 21244981]
- Cignoni P, Callieri M, Corsini M, Dellepiane M, Ganovelli F, Ranzuglia G. 2008 MeshLab: an opensource mesh processing tool, In: Scarano V, De Chiara R, Erra U, eds. Sixth Eurographics Italian Chapter Conference. Salerno, It.: The Eurographics Association, 2008. pp. 129–36.
- Duan X, Qu M, Wang J, Trevathan J, Vrtiska T, Williams JC Jr, Krambeck A, Lieske J, McCollough C. Differentiation of calcium oxalate monohydrate and calcium oxalate dihydrate stones using quantitative morphological information from micro-computerized and clinical computerized tomography. J Urol 2013;189:2350–6. [PubMed: 23142201]
- Fleet DJ, Weiss Y. Optical flow estimation, In: Paragios N, Chen Y, and Faugeras OD, eds. Handbook of Mathematical Models in Computer Vision. New York: Springer, 2006. pp. 237–57.
- Gaspari R, Blehar D, Mendoza M, Montoya A, Moon C, Polan D. Use of ultrasound elastography for skin and subcutaneous abscesses. J Ultrasound Med 2009;28:855–60. [PubMed: 19546327]
- Hu HH, Chen HY, Hung CI, Guo WY, Wu YT. Shape and curvedness analysis of brain morphology using human fetal magnetic resonance images in utero. Brain Struct Funct 2013;218:1451–62. [PubMed: 23135358]
- Kuligowska E, Connors SK, Shapiro JH. Liver abscess: sonography in diagnosis and treatment. AJR Am J Roentgenol 1982;138:253–7. [PubMed: 6976725]
- Kunze G, Staritz M, Köhler M. Contrast-enhanced ultrasound in different stages of pyogenic liver abscess. Ultrasound Med Biol 2015;.41:952–9. [PubMed: 25701525]
- Legget ME, Leotta DF, Bolson EL, McDonald JA, Martin RW, Li XN, Otto CM, Sheehan FH. System for quantitative three-dimensional echocardiography of the left ventricle based on a magnetic-field

position and orientation sensing system. IEEE Trans Biomed Eng 1998;45:494–504. [PubMed: 9556966]

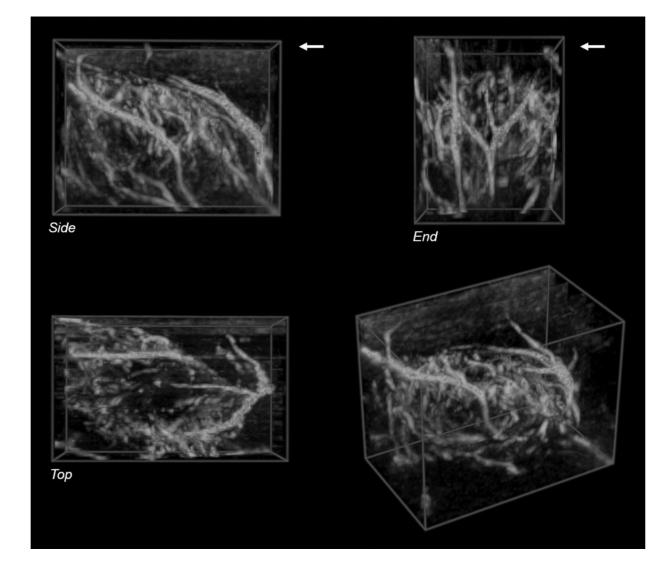
- Leotta DF, Martin RW. Three-dimensional ultrasound imaging of the rotator cuff: spatial compounding and tendon thickness measurement. Ultrasound Med Biol 2000;26:509–25. [PubMed: 10856614]
- Leotta DF, Primozich JF, Beach KW, Bergelin RO, Strandness DE Jr. Serial measurement of crosssectional area in peripheral vein grafts using three-dimensional ultrasound. Ultrasound Med Biol 2001;27:61–8. [PubMed: 11295271]
- Leotta DF, Zierler RE, Sansom K, Aliseda A, Anderson MD, Sheehan FH. Evaluation of examiner performance using a duplex ultrasound simulator. Flow velocity measurements in dialysis access fistula models. Ultrasound Med Biol 2018;44:1712–20. [PubMed: 29793851]
- Liu GJ, Lu MD, Xie XY, Xu HX, Xu ZF, Zheng YL, Liang JY, Wang W. Real-time contrast-enhanced ultrasound imaging of infected focal liver lesions. J Ultrasound Med 2008;27:657–66. [PubMed: 18359914]
- Liu H, Liao Q, Wang Y, Hu Y, Zhu Q, Wang L, Liu Q, Li J, Jiang Y. A new tool for diagnosing parathyroid lesions: angio plus ultrasound imaging. J Thorac Dis 2019;11:4829–34. [PubMed: 31903273]
- Lu C, Merrill C, Medellin A, Novak K, Wilson SR. Bowel ultrasound state of the art: grayscale and Doppler ultrasound, contrast enhancement, and elastography in Crohn disease. J Ultrasound Med 2019;38:271–88. [PubMed: 30604884]
- Lucas BD, Kanade T. An iterative image registration technique with an application to stereo vision. Proceedings of Imaging Understanding Workshop, 1981. pp. 121–30.
- Matula TJ, Wang YN, Khokhlova T, Leotta DF, Kucewicz J, Brayman AA, Bruce M, Maxwell AD, MacConaghy BE, Thomas G, Chernikov VP, Buravkov SV, Khokhlova VA, Richmond K, Chan K, Monsky W. Treating porcine abscesses with histotripsy: a pilot study. Ultrasound Med Biol. 2020 11 26:S0301–5629(20)30474–9.
- Mavilia MG, Molina M, Wu GY. The evolving nature of hepatic abscess: a review. J Clin Transl Hepatol 2016;4:158–68. [PubMed: 27350946]
- Meyer M, Desbrun M, Schröder P, Barr AH. Discrete differential-geometry operators for triangulated 2-manifolds. In: Hege HC, Polthier K, eds. Visualization and Mathematics III. Mathematics and Visualization Berlin, Heidelberg: Springer, 2003.
- O'Rourke K, Kibbee N, Stubbs A. Ultrasound for the evaluation of skin and soft tissue infections. Mo Med 2015;112:202–5. [PubMed: 26168591]
- Ripollés T, Martínez-Pérez MJ, Blanc E, Delgado F, Vizuete J, Paredes JM, Vilar J. Contrast-enhanced ultrasound (CEUS) in Crohn's disease: technique, image interpretation and clinical applications. Insights Imaging 2011;2:639–52. [PubMed: 22347983]
- Ripollés T, Martínez-Pérez MJ, Paredes JM, Vizuete J, García-Martínez E, Jiménez-Restrepo DH. Contrast-enhanced ultrasound in the differentiation between phlegmon and abscess in Crohn's disease and other abdominal conditions. Eur J Radiol 2013;82:e525–31. [PubMed: 23838329]
- Sigrist RMS, Liau J, Kaffas AE, Chammas MC, Willmann JK. Ultrasound elastography: review of techniques and clinical applications. Theranostics 2017;7:1303–29. [PubMed: 28435467]
- Wang Y-N, Brayman AA, Chan KT, Richmond K, Monsky WL, Matula TJ. A porcine bimicrobial abscess model for testing interventional treatments. bioRxiv 2020:2020.03.05.979088.



#### Figure 1.

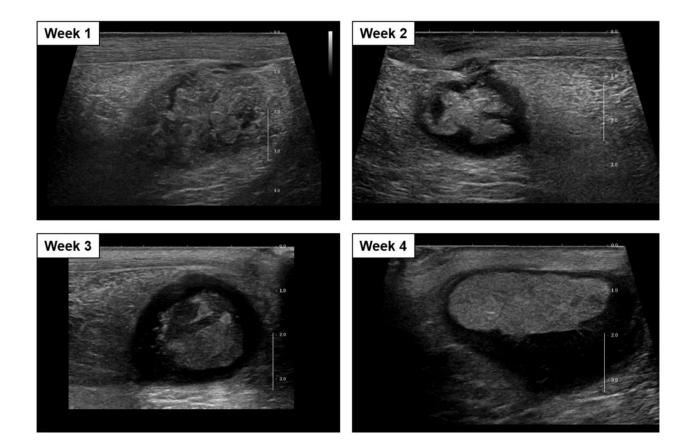
Representative B-mode (left) and color Doppler (right) images for intramuscular abscess ABS1 imaged at 3 weeks (23 days) post-injection. The center of the abscess lies approximately 2 cm below the skin surface within the biceps femoris. The B-mode image demonstrates the salient features of a mature abscess, including a well-circumscribed hypoechoic capsule (arrow heads) and a core (C) with variable echogenicity. Plane-wave directional color power Doppler shows an avascular core surrounded by a highly vascularized capsule.

Leotta et al.



## Figure 2.

Volume rendering of a 3D reconstruction of plane-wave directional color power Doppler images acquired during a freehand sweep across abscess case ABS1 at 20 days post-injection shows a network of blood vessels surrounding the abscess core. The color pixels in each 2D frame were segmented and reconstructed without the background gray-scale image. The skin surface is at the upper edge of the Side and End views in the top row (arrows). The bottom left shows an oblique perspective view of the reconstructed volume. Volume size: 33  $\times 20 \times 25$  mm (L  $\times$  W  $\times$  H). Voxel size: 0.31 mm.



#### Figure 3.

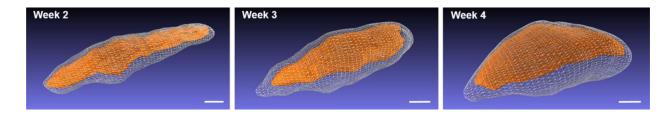
Cross-sectional B-mode images of the intramuscular abscess case ABS2 at weeks 1 through 4 (days 8, 15, 20 and 29) post-injection.

Leotta et al.



### Figure 4.

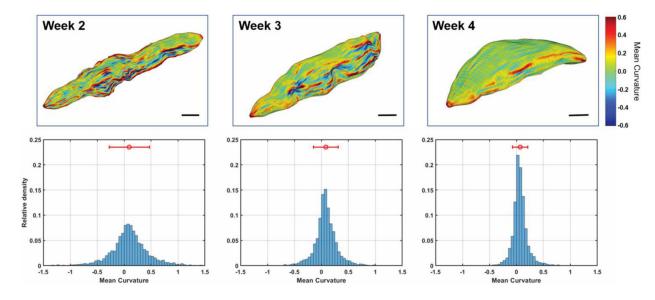
Volume rendering of the 3D volume reconstructions of case ASB2 at weeks 2 through 4 post-injection. The gray-scale volumes are truncated in the cross-sectional and longitudinal directions to visualize the internal structure of the abscess capsule and core. Displayed volume width: 30 mm Week 2, 23 mm Week 3, 32 mm Week 4. Voxel size: 0.08 mm Week 2, 0.16 mm Week 3, 0.32 mm Week 4.



## Figure 5.

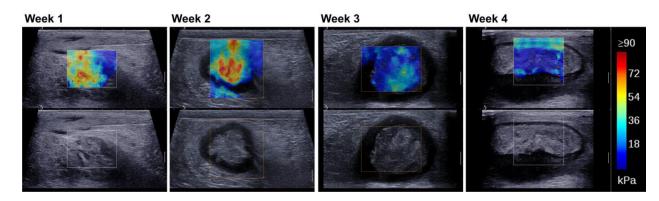
Surface reconstructions generated by manual segmentation of the outer capsule and the inner core for case ABS2 at weeks 2 through 4 post-injection. Scale bars: 1 cm.

Leotta et al.



#### Figure 6.

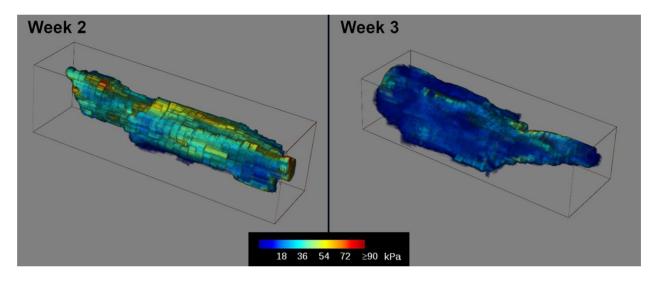
Core surface curvature over time for case ABS2 at weeks 2 through 4 post-injection. Top row: Mean curvature mapped to the surface reconstruction according to the color scale at the right. Scale bars: 1 cm. Bottom row: Histograms of the mean curvature measurements, with the mean and standard deviation shown above. Each histogram is normalized so that the bins sum to one. The curvature measurements of the core surface over time demonstrate a change from an irregular border to a rounded and more well-defined border. Circle: Mean. Bars: Standard deviation. Number of vertices compiled for each histogram: 4558 Week 2, 5144 Week 3, 4622 Week 4.



## Figure 7.

Short-axis Shear Wave Elastography images for case ABS2 at weeks 1 through 4 postinjection showing a decrease in stiffness over time (red to blue).

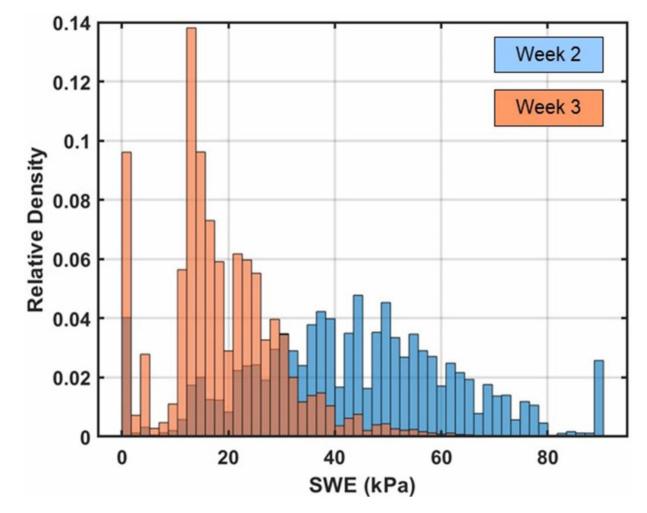
Leotta et al.



#### Figure 8.

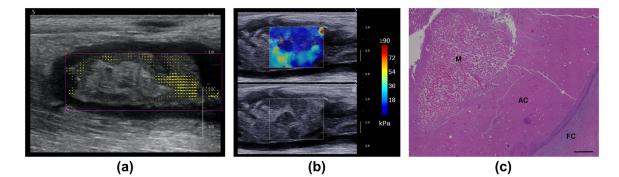
Transparent volume renderings of 3D reconstructions of the SWE measurements in the abscess core at weeks 2 and 3 post-injection for case ABS2. The largest change in abscess stiffness occurred between these two time points. The 3D volumes were generated from untracked freehand scans along the length of the abscess. Voxel colors represent the measured stiffness in kPa according to the color scale shown below. Volume size:  $89 \times 23 \times 25$  mm Week 2,  $79 \times 25 \times 19$  mm Week 2. Voxel size: 0.08 mm Week 2, 0.07 mm Week 3.

Leotta et al.



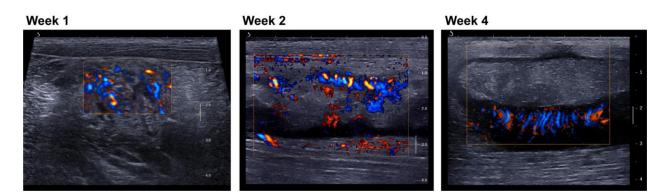
## Figure 9.

Histograms of elastography measurements compiled over the abscess core at 2 weeks and 3 weeks post-injection for case ABS2. The histograms are normalized so that the bins sum to one. The distribution shifts towards lower stiffness from week 2 to week 3. Number of voxels compiled for each histogram:  $28.3 \times 10^6$  Week 2,  $43.2 \times 10^6$  Week 3.



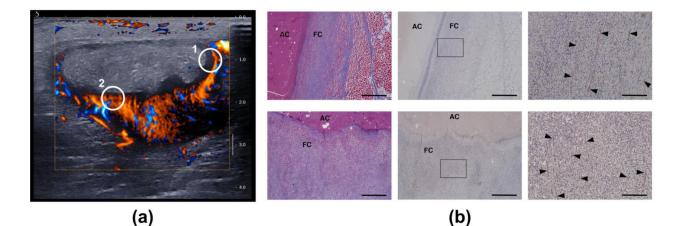
## Figure 10.

Internal flow with compression and core heterogeneity for the intramuscular abscess case ABS2. (a) Regions of flow with external compression are identified and quantified by comparing regional frame-to-frame displacements at week 3 post-injection. The yellow arrowheads show the direction and magnitude of the displacements. The largest arrows correspond to 1 mm displacements; arrows with magnitudes less than 0.35 mm are not shown. (b) Long-axis SWE image shows regions of varying stiffness in the core at week 3 post-injection. The region with higher stiffness corresponds to the stationary region in (a). (c) Histology of the abscess core with Hematoxylin and Eosin at week 4 post-injection shows a block of muscular tissue (M) within the abscess core (AC). FC: Fibrous Capsule. Scale bar: 1000 microns.



## Figure 11.

Long-axis views of abscess case ABS2 with color Doppler imaging show a network of vessels developing from a diffuse pattern throughout the injection site (week 1) to an organized network within the capsule surrounding the avascular core (week 4). Week 1: standard directional color power Doppler. Weeks 2 and 3: plane-wave directional color power Doppler.



#### Figure 12.

(a) Short-axis plane-wave directional color power Doppler image (4 weeks post-injection) of the vascularized capsule and avascular core of case ABS2. The circled regions indicate the general locations for which histology images were obtained. (b) Histology of the capsule confirms a network of vessels with circumferential orientation on the lateral side (top row, circle 1) and oriented toward the core on the deep side (bottom row, circle 2). The vessels are indicated by the black arrow heads in the high-magnification images in the right column. Left: Masson's trichrome. Middle and right: CD31 labelled. Right: magnification of the boxed regions in the middle column. FC: Fibrous Capsule. AC: Abscess Core. Scale bar: 1000 microns (left and middle)/200 microns (right).

#### Table 1.

#### 3D abscess measurements

	TOTAL		CORE			
Week	Size (mm)	Volume (ml)	Size (mm)	Volume (ml)	Curvature (1/mm)	Stiffness (kPa)
2	$102\times28\times21$	20.4	$99 \times 22 \times 19$	8.0	0.10 +/- 0.37	43.0 +/- 19.9
3	$88\times32\times29$	30.3	$80\times24\times21$	12.6	0.08 +/- 0.23	19.0 +/- 11.6
4	$68\times50\times27$	37.2	$63\times43\times17$	16.2	0.07 +/-0.14	-

Size: Length  $\times$  Width  $\times$  Height

Curvature/Stiffness: Mean +/- Standard Deviation