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The rheumatoid arthritis gene expression signature among women who improve or worsen during pregnancy – a pilot study

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Abstract

Objective: To assess whether gene expression signatures associated with rheumatoid arthritis (RA) before pregnancy differ between women who improve or worsen during pregnancy, and determine whether these expression signatures are altered during pregnancy when RA improves or worsens.

Methods: Clinical data and blood samples were collected before pregnancy (T0) and at the third trimester (T3) from 11 RA and 5 healthy women. RA disease activity was assessed using the Clinical Disease Activity Index (CDAI). At each time-point, RA-associated gene expression signatures were identified using differential expression analysis of RNA sequencing profiles between RA and healthy women.

Results: Of the women with RA, 6 improved by T3 ($RA_{improved}$), 3 worsened ($RA_{worsened}$) and 2 were excluded. At T0, mean CDAI scores were similar in both groups ($RA_{improved}$: 11.2±9.8; $RA_{worsened}$: 13.8±6.7; Wilcoxon-rank test: p=0.6). In the $RA_{improved}$ group, 89 genes were differentially expressed at T0 (q<0.05 and fold-change (FC) 2) compared to healthy women. When RA improved at T3, 65 of 89 (73%) of these no longer displayed RA-associated expression.

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In the $RA_{worsened}$ group, a largely different RA gene expression signature (429 genes) was identified at T0. When RA disease activity worsened at T3, 207 of 429 (48%) lost their differential expression, while an additional 151 genes became newly differentially expressed.

Conclusion: In our pilot dataset, pre-pregnancy RA expression signatures differed between women who subsequently improved or worsened during pregnancy, suggesting that inherent genomic differences perhaps influence how pregnancy impacts disease activity. Further, these RA signatures were altered during pregnancy, as disease activity changed.

Keywords

Rheumatoid arthritis; Gene expression signature; Pregnancy; RNA-seq

INTRODUCTION

Rheumatoid arthritis (RA) is a systemic inflammatory disease that leads to significant disability resulting from pain and swelling of inflamed joints and from joint destruction. To date, there is no cure. Pregnancy is known to have disease-modifying properties (1–4) on RA, with a significant proportion of women experiencing an improvement in disease activity during pregnancy, while in others, the disease may remain unchanged or may even worsen. Even though there are medications, including some traditional and biologic disease modifying anti-rheumatic drugs (DMARDs) that are considered safe for use in pregnancy (5, 6), many women with RA prefer to stop taking medications during pregnancy. However, because there are no known biomarkers at present to predict who will likely improve or worsen or whose RA will remain unchanged during pregnancy, these women are hesitant to plan a pregnancy because they do not know whether their disease will worsen if they stop taking medications in order to try to conceive.

Several case-control studies, based on gene expression data from microarrays (7–12) or RNA sequencing (RNA-seq), have been conducted to investigate gene expression signatures associated with RA (13). However, gene expression studies that have been conducted in the context of RA pregnancy did not examine RA-associated expression signatures in the non-pregnant state due to pre-pregnancy samples not being available (14–17). It is thus not known whether the pre-pregnancy RA expression signature can be used to predict whether RA will subsequently improve or worsen during pregnancy. Further, given that gene expression is a dynamic process, it is possible that the RA-associated gene expression signature may be altered during pregnancy. Genes modulating disease activity during pregnancy may show altered expression when disease activity changes over time, i.e. their expression may either no longer be associated with RA when disease activity is low or in remission during pregnancy, or additional genes may show RA-associated expression when RA worsens during pregnancy. However, the influence of pregnancy on the RA gene expression signature, if any, has not been investigated.

In the present study, we have used our unique prospective pilot pregnancy cohort of RA and healthy women that includes a pre-pregnancy time-point (18, 19) as a case-control dataset to examine gene expression signatures associated with RA at the pre-pregnancy baseline. We hypothesized that the baseline RA-associated gene expression signature among women who

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subsequently improved during pregnancy differs from that among women who worsened during pregnancy. We also evaluated a second hypothesis that the gene expression signature associated with RA at the pre-pregnancy baseline is altered during pregnancy, when disease activity improves or worsens.

SUBJECTS AND METHODS

Study subjects

Healthy women and women with RA of Danish descent who were planning a pregnancy were recruited and enrolled in our pregnancy cohort in Denmark and were prospectively followed, as previously described (18). A subset of 11 RA and 5 healthy women from this cohort, on whom we reported longitudinal changes in expression (19), was included in the present study. The women with RA fulfilled the 1987 revised American College of Rheumatology criteria for the disease (20). The study was approved by the Ethics Committee for Region Hovedstaden (Denmark), the Danish Data Protection Agency, and the Children's Hospital Oakland Research Institute Institutional Review Board (IRB number: 2009–073). All subjects provided written informed consent prior to enrollment. Data and samples were collected as previously described (19).

Assessment of RA disease activity

RA disease activity was assessed using the Clinical Disease Activity Index (CDAI) (21), because it does not include acute phase reactants such as C-reactive protein (CRP) whose levels are known to fluctuate during pregnancy (22, 23); further, acute phase reactants do not contribute much additional information on top of what is provided by the CDAI (24). The change in CDAI (CDAI) from before pregnancy (T0) to the third trimester (T3) was used to determine whether disease activity improved or worsened. Patients were categorized as having improved by T3 ($RA_{improved}$), if their CDAI fit the criteria for a minimum clinically important difference (MCID) based on baseline (T0) disease activity; CDAI values of 12, 6 and 1 were used as threshold when disease activity at T0 was high, moderate or low, respectively (25). Those women with an increase in CDAI from T0 to T3, satisfying the MCID criteria for worsening of disease activity, were included in the "worsened" subset, referred to as $RA_{worsened}$.

RNA sequencing and Bioinformatic analyses

RNA extractions, processing and sequencing were performed as originally described (18). Pseudoalignment of the de-multiplexed raw sequence reads (FASTQ format) to the Ensembl reference human GRCh38 transcriptome assembly (release 98) and quantification of transcript abundances were performed using kallisto (v0.43.0) (26). BioMart annotations were used to combine transcript-level counts into gene-level estimates, i.e. counts were summed by Ensembl gene ID. Gene IDs that mapped to patches or alternate haplotypes rather than to the primary reference sequence were excluded to avoid duplication. Pseudogenes, genes without annotations as well as genes with very low read counts (<1 transcript per million) in at least 25% of all samples were filtered out. Any globin and rRNA transcripts still present were also filtered out. To adjust for variable sequencing depths across samples, the gene-level counts were normalized using the Trimmed Mean of M values

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(TMM) algorithm as implemented in the edgeR package (v3.26.8) (27, 28). To assess batch effects, normalized counts from pairs of technical replicates were plotted, and outliers were filtered out to achieve a Pearson correlation of at least 95% between replicates.

Statistical analyses

Case-control differential gene expression analysis—To identify gene expression signatures associated with RA at the T0 baseline, cross-sectional differential expression analysis was performed using edgeR (v3.26.8) (27), comparing normalized T0 gene-level counts between each RA subset ($RA_{improved}$ or $RA_{worsened}$) and healthy women. In each analysis, a negative binomial distribution was used to handle the over-dispersion in RNA-seq gene counts. Differential expression was tested using generalized linear model (GLM) likelihood ratio tests; differences between RA and healthy women were assessed using the contrast argument of the *glmLRT* function in edgeR. Correction for multiple testing was performed using the False Discovery Rate (FDR) method (29). A q-value threshold of 0.05, in combination with a fold-change (FC) of at least 2, was used to assess significance. To determine whether the pre-pregnancy expression signature changed when RA improved or worsened during pregnancy, the differential expression analysis was repeated using data from the same women ($RA_{improved}$ or $RA_{worsened}$ vs. healthy) at the T3 time-point.

Functional analysis—Differentially expressed genes were analyzed for over-representation of Gene Ontology (GO) categories using a hypergeometric test implemented in the Web-based Gene Set Analysis Toolkit (WebGestalt) (30). A significance threshold of q<0.05 was used to define enrichment. Cystoscape (31) was used for functional annotations and visualization of protein interactions documented in the STRING database (32).

RESULTS

Study subjects

Of the 11 women with RA, 6 improved by T3 while 3 worsened, based on MCID thresholds. Two women were excluded because even though their disease activity improved during pregnancy, one was already in remission at T0, and the CDAI value for the other (CDAI=2.7) did not meet the MCID threshold of 6, for moderate baseline disease activity. The changes in disease activity scores from T0 to T3 were significantly correlated between the CDAI and the DAS28CRP3 (Pearson's correlation=85%, p=0.004). The average age at conception was as follows: 28.9±6.0 years for $RA_{improved}$, 33.2±1.9 years for $RA_{worsened}$, and 31.2±5.7 years for the healthy women. The women who improved had a shorter disease duration than those who worsened ($RA_{improved}$ (mean±SD): 6.5±4.2 years and $RA_{worsened}$: 8.9±1.1 years), although this difference was not statistically significant. Medications taken by the women with RA at each time-point are shown in Table 1. While mean disease activity (CDAI scores) at baseline did not differ significantly between the two RA subsets ($RA_{improved}$: 11.2±9.8 and $RA_{worsened}$: 13.8±6.7; Wilcoxon-rank test: p=0.6), the mean values at T3 differed significantly ($RA_{improved}$: 2.2±1.3 and $RA_{worsened}$: 31.7±15.1; Wilcoxon-rank test: p=0.02).

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The RA gene expression signature at the T0 (pre-pregnancy) baseline

$RA_{improved}$ vs healthy women—A total of 89 genes were differentially expressed (q<0.05; FC 2) between the 6 $RA_{improved}$ and 5 healthy women (Figure 1A and Supplementary Table S1). The genes that were over-expressed (n=44) in RA (e.g. C4BPA, CAMP, CD177, CRISP3, HLA-DQA2, MMP8, OLFM4, ORM1, S100A12) as well as those that were under-expressed (n=45) (e.g. CMPK2, HERC5, IFI44, IFI44L, IFITM3, IL1RL1, IL5RA, MX1, OAS1, OAS2, OAS3, SIGLEC1) were enriched in various immune-related gene ontology (GO) biological processes, as shown in Table 2, and in reactome pathways relating to interferon signaling (q=5.9E-06), antiviral mechanism by IFN-stimulated genes (q=7.0E-05), and p130Cas linkage to MAPK signaling for integrins (q=1.4E-02), among others. A large proportion (52%) of the 89 genes differentially expressed between $RA_{improved}$ and healthy women at the T0 baseline encode proteins that are functionally related, as shown by protein networks, based on the STRING database (32), in Cytoscape (Figure 2); most of the under-expressed genes formed a tight cluster, distinct from the over-expressed genes.

$RA_{worsened}$ vs healthy women—A total of 429 genes were differentially expressed (FC 2; q<0.05) between $RA_{worsened}$ and healthy women at T0 (Figure 3A). This gene expression signature largely differed from the one identified above ($RA_{improved}$ vs healthy); only 19 of the 429 genes overlapped with those differentially expressed between $RA_{improved}$ and healthy women at T0, with the majority demonstrating similar expression patterns in both RA sub-groups compared to healthy women, i.e. over-expressed: OLFM4, UBB, ORM1, SEPTIN3, KRT1, TUBB2A or under-expressed: IL1RL1, IGLC3, IGLV2–14, PF4V1, FADS2, NKX3–1 (Supplementary Table S2). HLA-DRQA2, on the other hand, was 3-fold over-expressed among the $RA_{improved}$, and 3-fold under-expressed among the $RA_{worsened}$ women, compared to the healthy women.

The RA gene expression signature is altered when RA improves or worsens during pregnancy

$RA_{improved}$ vs healthy women at T3—When disease activity improved by T3, most of the baseline RA signature genes identified among $RA_{improved}$ women (65 of 89, i.e. 73%) were no longer differentially expressed between the $RA_{improved}$ and healthy women (Figure 1B and Supplementary Table S1). Of note, there were 24 genes that remained differentially expressed between $RA_{improved}$ and healthy women at T3 (e.g. C4BPA, HLA-DQA2, IGLC3, IL5RA, MAOA, OLIG2, PTGDR2, TUBB2A), and an additional few (n=27) became newly differentially expressed (e.g. ADORA3, CYP27A1, DSC1, RAP1GAP, SCL29A1).

$RA_{worsened}$ vs healthy women at T3—When disease activity worsened by T3, 207 of the 429 genes (48%) differentially expressed at the T0 baseline lost their differential expression, while an additional 151 genes became newly differentially expressed (Figure 3B).

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DISCUSSION

In the present study, our goal was to examine whether the pre-pregnancy RA gene expression signature differs between women who subsequently improved during pregnancy and those who worsened during pregnancy. We also examined whether the pre-pregnancy RA gene expression signature was altered in any way during pregnancy, when disease activity improved or worsened.

In our pilot dataset, even though mean disease activity was similar between the $RA_{improved}$ and $RA_{worsened}$ groups at the pre-pregnancy (non-pregnant) baseline, there was very little overlap in the sets of genes showing RA-associated expression within each of the two groups. The few genes that overlapped between the two RA expression signatures included some that have previously been implicated in RA such as IL1RL1 (33), ORM1 (34), KRT1 (35), and HLA-DQA2 (36). Although HLA-DQA2 expression was associated with RA in both subsets, this gene demonstrated contrasting expression patterns in the two RA subsets; it was 3-fold over-expressed in the $RA_{improved}$ group and 3-fold under-expressed in the $RA_{worsened}$ group, both compared to the same set of healthy women. While increased HLA-DQA2 expression has been reported in RA (37), a negative correlation has also been found between expression levels in synovial tissue fibroblast cells and Health Assessment Questionnaire (HAQ) scores (38). Nonetheless, the significance of these contrasting expression patterns in the two groups of women with RA is not entirely clear. The RA expression signature identified among the $RA_{improved}$ women included many additional genes whose expression and/or methylation patterns have previously been associated with RA, such as S100A12 (39), CRISP3 (40), MMP8 (41), and CAMP (12). Of interest, the IFN-inducible genes IFI44, IFI44L, CMPK2, HERC5, MX1, SIGLEC1, OAS1, OAS2 and OAS3 were also part of the baseline RA expression signature among the $RA_{improved}$ women. Compared to healthy women, these genes were under-expressed in this RA subset at baseline as we recently reported (19), in contrast to other studies of RA and other autoimmune conditions (9, 42, 43). While our pre-pregnancy RA signatures also included many genes that did not overlap with RA signatures from previous case-control studies, results were inconsistent even across those previous studies. This could be attributed to a number of factors including: the source of RNA [whole blood (our study) vs. peripheral blood mononuclear cells (PBMCs)] (9, 11, 14–17), synovial fibroblasts (13), or neutrophils (44); differences in gene expression technology used [RNA-seq (our study) vs. microarrays] (9, 11, 14–17); patient sample and sex ratio (only women, most of whom experienced improvement of RA disease activity during pregnancy (our study) vs. women and men) (9, 11, 13, 44).

In the present study, we also examined the influence of pregnancy on RA-associated gene expression signatures. This had not previously been reported since pre-pregnancy samples were not available in prior studies (14–17). We observed a dilution of the baseline RA signature during pregnancy, with the majority (73%) of signature genes showing similar expression in both RA and healthy women by the third trimester, when RA improved. These results are consistent with those of a previous study demonstrating minimal differences in PBMC expression profiles between RA and healthy women at the third trimester (15). While our study design does not allow us to determine whether the loss of association with RA at

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the 3rd trimester is specific to pregnancy or not, it is plausible that as the expression profiles of the women with RA undergo pregnancy-induced changes, they start to resemble those of healthy women, as we observed, and these changes are accompanied by an improvement of the disease. On the other hand, when RA worsened during pregnancy, many genes (n=151) demonstrated new expression patterns that became associated with RA, as would be expected when the “case” and “control” groups become phenotypically more different from each other.

In a previous study comparing the DAS28ESR and DAS28CRP, with and without patient global scores, during pregnancy, the DAS28CRP3 was found to perform better (45). However, since even CRP levels have been shown to fluctuate during pregnancy (22, 23), the DAS28CRP3 does not represent a gold standard for use in pregnancy. Other measures of disease activity such as the CDAI that do not include acute phase reactants have not been assessed for their performance during pregnancy. In the absence of a gold standard, we chose to use the CDAI to assess disease activity before and during pregnancy because acute phase reactants have been shown to add little information on top of the clinical variables already included in the CDAI (24). Additionally, the CDAI is more stringent than the DAS28 when assessing improvement of disease activity; it has been shown that patients can satisfy DAS28 remission criteria while still having active disease (46).

In our study, the availability of pre-pregnancy and pregnancy data from the same women who improved or worsened during pregnancy enabled us to investigate pre-pregnancy gene expression signatures between the two groups as well as the effect of pregnancy on those expression signatures. The use of RNA-seq technology to assess gene expression was another strength. Our study has some limitations. First, given that this is a pilot study, sample sizes were small. Nonetheless, patterns emerged that are supported by previous literature and thus further investigations in a larger sample are warranted. We did not examine proportions of cell types between disease states (RA vs healthy) and/or across time points because our goal was to identify overall systemic gene expression changes, resulting from altered expression of specific genes or from differences in cell proportions. Because we used total RNA from whole blood, expression profiles of neutrophils may have dominated the observed expression patterns. It is also possible that medications taken by the women with RA before pregnancy may have influenced the results. However, due to sample size limitations, we could not assess if this was the case, and we also were unable to adjust for dosage and/or specific medications.

In conclusion, we report here novel findings that women with RA who improved during pregnancy demonstrated differences in pre-pregnancy RA-associated gene expression compared to women who worsened. These differences in pre-pregnancy RA expression signatures suggest that inherent genomic differences between women with RA may influence how pregnancy alters disease activity. Our findings that RA-associated gene expression signatures are altered during pregnancy, when disease activity changes are also novel. Additional investigations in larger datasets are warranted to corroborate these preliminary findings and to identify novel drug targets and/or biomarkers of disease activity.

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Supplementary Material

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REFERENCES

1. Hench PS. The ameliorating effect of pregnancy on chronic atrophic (infectious rheumatoid) arthritis, fibrositis, and intermittent hydrarthrosis. Mayo Clin Proc 1938;13:161–7.
2. Nelson JL, Ostensen M. Pregnancy and rheumatoid arthritis. Rheum Dis Clin North Am 1997;23:195–212. [PubMed: 9031383]
3. de Man YA, Dolhain RJ, van de Geijn FE, Willemsen SP, Hazes JM. Disease activity of rheumatoid arthritis during pregnancy: Results from a nationwide prospective study. Arthritis Rheum 2008;59:1241–8. [PubMed: 18759316]
4. Barrett JH, Brennan P, Fiddler M, Silman AJ. Does rheumatoid arthritis remit during pregnancy and relapse postpartum? Results from a nationwide study in the united kingdom performed prospectively from late pregnancy. Arthritis Rheum 1999;42:1219–27. [PubMed: 10366115]
5. Gotestam Skorpen C, Hoeltzenbein M, Tincani A, Fischer-Betz R, Elefant E, Chambers C, et al. The eular points to consider for use of antirheumatic drugs before pregnancy, and during pregnancy and lactation. Ann Rheum Dis 2016;75:795–810. [PubMed: 26888948]
6. Krause ML, Amin S, Makol A. Use of dmards and biologics during pregnancy and lactation in rheumatoid arthritis: What the rheumatologist needs to know. Ther Adv Musculoskelet Dis 2014;6:169–84. [PubMed: 25342996]
7. Huber R, Hummert C, Gausmann U, Pohlers D, Koczan D, Guthke R, et al. Identification of intra-group, inter-individual, and gene-specific variances in mrna expression profiles in the rheumatoid arthritis synovial membrane. Arthritis Res Ther 2008;10:R98. [PubMed: 18721452]
8. Ungethuem U, Haeupl T, Witt H, Koczan D, Krenn V, Huber H, et al. Molecular signatures and new candidates to target the pathogenesis of rheumatoid arthritis. Physiol Genomics 2010;42A:267–82. [PubMed: 20858714]
9. van der Pouw Kraan TC, Wijbrandts CA, van Baarsen LG, Voskuyl AE, Rustenburg F, Baggen JM, et al. Rheumatoid arthritis subtypes identified by genomic profiling of peripheral blood cells: Assignment of a type i interferon signature in a subpopulation of patients. Ann Rheum Dis 2007;66:1008–14. [PubMed: 17223656]

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10. Bovin LF, Rieneck K, Workman C, Nielsen H, Sorensen SF, Skjodt H, et al. Blood cell gene expression profiling in rheumatoid arthritis. Discriminative genes and effect of rheumatoid factor. Immunol Lett 2004;93:217–26. [PubMed: 15158620]
11. Batliwalla FM, Baechler EC, Xiao X, Li W, Balasubramanian S, Khalili H, et al. Peripheral blood gene expression profiling in rheumatoid arthritis. Genes Immun 2005;6:388–97. [PubMed: 15973463]
12. Teixeira VH, Olaso R, Martin-Magniette ML, Lasbleiz S, Jacq L, Oliveira CR, et al. Transcriptome analysis describing new immunity and defense genes in peripheral blood mononuclear cells of rheumatoid arthritis patients. PLoS One 2009;4:e6803. [PubMed: 19710928]
13. Heruth DP, Gibson M, Grigoryev DN, Zhang LQ, Ye SQ. Rna-seq analysis of synovial fibroblasts brings new insights into rheumatoid arthritis. Cell Biosci 2012;2:43. [PubMed: 23259760]
14. Haupl T, Ostensen M, Grutzkau A, Burmester GR, Villiger PM. Interaction between rheumatoid arthritis and pregnancy: Correlation of molecular data with clinical disease activity measures. Rheumatology (Oxford) 2008;47 Suppl 3:iii19–22. [PubMed: 18504279]
15. Haupl T, Ostensen M, Grutzkau A, Radbruch A, Burmester GR, Villiger PM. Reactivation of rheumatoid arthritis after pregnancy: Increased phagocyte and recurring lymphocyte gene activity. Arthritis Rheum 2008;58:2981–92. [PubMed: 18821679]
16. Weix J, Forger F, Haupl T, Surbek D, Ostensen M, Villiger PM. Influence of pregnancy on the adipocytokine and peroxisome proliferator-activated receptor pathways in peripheral blood mononuclear cells from healthy donors and rheumatoid arthritis patients. Arthritis Rheum 2012;64:2095–103. [PubMed: 22231457]
17. Weix J, Haupl T, Raio L, Villiger PM, Forger F. The physiologic increase in expression of some type i ifn-inducible genes during pregnancy is not associated with improved disease activity in pregnant patients with rheumatoid arthritis. Transl Res 2013;161:505–12. [PubMed: 23507374]
18. Mittal A, Pachter L, Nelson JL, Kjaergaard H, Smed MK, Gildengorin VL, et al. Pregnancy-induced changes in systemic gene expression among healthy women and women with rheumatoid arthritis. PLoS One 2015;10:e0145204. [PubMed: 26683605]
19. Goin DE, Smed MK, Pachter L, Purdom E, Nelson JL, Kjaergaard H, et al. Pregnancy-induced gene expression changes in vivo among women with rheumatoid arthritis: A pilot study. Arthritis Res Ther 2017;19:104. [PubMed: 28545501]
20. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The american rheumatism association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988;31:315–24. [PubMed: 3358796]
21. Mierau M, Schoels M, Gonda G, Fuchs J, Aletaha D, Smolen JS. Assessing remission in clinical practice. Rheumatology (Oxford) 2007;46:975–9. [PubMed: 17341506]
22. Skarzynska E, Zborowska H, Jakimiuk AJ, Karlinska M, Lisowska-Myjak B. Variations in serum concentrations of c-reactive protein, ceruloplasmin, lactoferrin and myeloperoxidase and their interactions during normal human pregnancy and postpartum period. J Trace Elem Med Biol 2018;46:83–7. [PubMed: 29413114]
23. Belo L, Santos-Silva A, Rocha S, Caslake M, Cooney J, Pereira-Leite L, et al. Fluctuations in c-reactive protein concentration and neutrophil activation during normal human pregnancy. Eur J Obstet Gynecol Reprod Biol 2005;123:46–51. [PubMed: 16260340]
24. Aletaha D, Nell VP, Stamm T, Uffmann M, Pflugbeil S, Machold K, et al. Acute phase reactants add little to composite disease activity indices for rheumatoid arthritis: Validation of a clinical activity score. Arthritis Res Ther 2005;7:R796–806. [PubMed: 15987481]
25. Curtis JR, Yang S, Chen L, Pope JE, Keystone EC, Haraoui B, et al. Determining the minimally important difference in the clinical disease activity index for improvement and worsening in early rheumatoid arthritis patients. Arthritis Care Res (Hoboken) 2015;67:1345–53. [PubMed: 25988705]
26. Bray NL, Pimentel H, Melsted P, Pachter L. Near-optimal probabilistic rna-seq quantification. Nat Biotechnol 2016;34:525–7. [PubMed: 27043002]
27. Robinson MD, McCarthy DJ, Smyth GK. Edger: A bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics 2010;26:139–40. [PubMed: 19910308]

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28. Robinson MD, Oshlack A. A scaling normalization method for differential expression analysis of rna-seq data. Genome Biol 2010;11:R25. [PubMed: 20196867]
29. Benjamini Y, Hochberg Y. Controlling the false discovery rate: A practical and powerful approach to multiple testing. Journal of the Royal Statistical Society, Series B 1995;57:289–300.
30. Zhang B, Kirov S, Snoddy J. Webgestalt: An integrated system for exploring gene sets in various biological contexts. Nucleic acids research 2005;33:W741–W8. [PubMed: 15980575]
31. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: A software environment for integrated models of biomolecular interaction networks. Genome Res 2003;13:2498–504. [PubMed: 14597658]
32. von Mering C, Jensen LJ, Snel B, Hooper SD, Krupp M, Foglierini M, et al. String: Known and predicted protein-protein associations, integrated and transferred across organisms. Nucleic Acids Res 2005;33:D433–7. [PubMed: 15608232]
33. Leung BP, Xu D, Culshaw S, McInnes IB, Liew FY. A novel therapy of murine collagen-induced arthritis with soluble t1/st2. J Immunol 2004;173:145–50. [PubMed: 15210768]
34. Park YJ, Yoo SA, Hwang D, Cho CS, Kim WU. Identification of novel urinary biomarkers for assessing disease activity and prognosis of rheumatoid arthritis. Exp Mol Med 2016;48:e211. [PubMed: 26915672]
35. Bhattacharjee M, Balakrishnan L, Renuse S, Advani J, Goel R, Sathe G, et al. Synovial fluid proteome in rheumatoid arthritis. Clin Proteomics 2016;13:12. [PubMed: 27274716]
36. Andreasi RB, Khan MFJ, Galuppi E, Govoni M, Rubini M. Replication analysis of gene-gene interaction between hla-dqa2 and hla-dqb2 variants in italian rheumatoid arthritis patients. Annals of the Rheumatic Diseases 2017;76:207.
37. Niu X, Lu C, Xiao C, Ge N, Jiang M, Li L, et al. The crosstalk of pathways involved in immune response maybe the shared molecular basis of rheumatoid arthritis and type 2 diabetes. PLoS One 2015;10:e0134990. [PubMed: 26252209]
38. Galligan CL, Baig E, Bykerk V, Keystone EC, Fish EN. Distinctive gene expression signatures in rheumatoid arthritis synovial tissue fibroblast cells: Correlates with disease activity. Genes Immun 2007;8:480–91. [PubMed: 17568789]
39. Nordal HH, Brun JG, Halse AK, Jonsson R, Fagerhol MK, Hammer HB. The neutrophil protein s100a12 is associated with a comprehensive ultrasonographic synovitis score in a longitudinal study of patients with rheumatoid arthritis treated with adalimumab. BMC Musculoskelet Disord 2014;15:335. [PubMed: 25282581]
40. Chapman EA, Lyon M, Simpson D, Mason D, Beynon RJ, Moots RJ, et al. Caught in a trap? Proteomic analysis of neutrophil extracellular traps in rheumatoid arthritis and systemic lupus erythematosus. Front Immunol 2019;10:423. [PubMed: 30915077]
41. Tchetverikov I, Ronday HK, Van El B, Kiers GH, Verzijl N, TeKoppele JM, et al. Mmp profile in paired serum and synovial fluid samples of patients with rheumatoid arthritis. Ann Rheum Dis 2004;63:881–3. [PubMed: 15194590]
42. Higgs BW, Liu Z, White B, Zhu W, White WI, Morehouse C, et al. Patients with systemic lupus erythematosus, myositis, rheumatoid arthritis and scleroderma share activation of a common type i interferon pathway. Ann Rheum Dis 2011;70:2029–36. [PubMed: 21803750]
43. Toro-Dominguez D, Carmona-Saez P, Alarcon-Riquelme ME. Shared signatures between rheumatoid arthritis, systemic lupus erythematosus and sjogren's syndrome uncovered through gene expression meta-analysis. Arthritis Res Ther 2014;16:489. [PubMed: 25466291]
44. Wright HL, Thomas HB, Moots RJ, Edwards SW. Interferon gene expression signature in rheumatoid arthritis neutrophils correlates with a good response to tnfi therapy. Rheumatology (Oxford) 2015;54:188–93. [PubMed: 25125592]
45. de Man YA, Hazes JM, van de Geijn FE, Krommenhoek C, Dolhain RJ. Measuring disease activity and functionality during pregnancy in patients with rheumatoid arthritis. Arthritis Rheum 2007;57:716–22. [PubMed: 17530669]
46. Aletaha D, Smolen JS. Remission in rheumatoid arthritis: Missing objectives by using inadequate das28 targets. Nat Rev Rheumatol 2019;15:633–4. [PubMed: 31427763]

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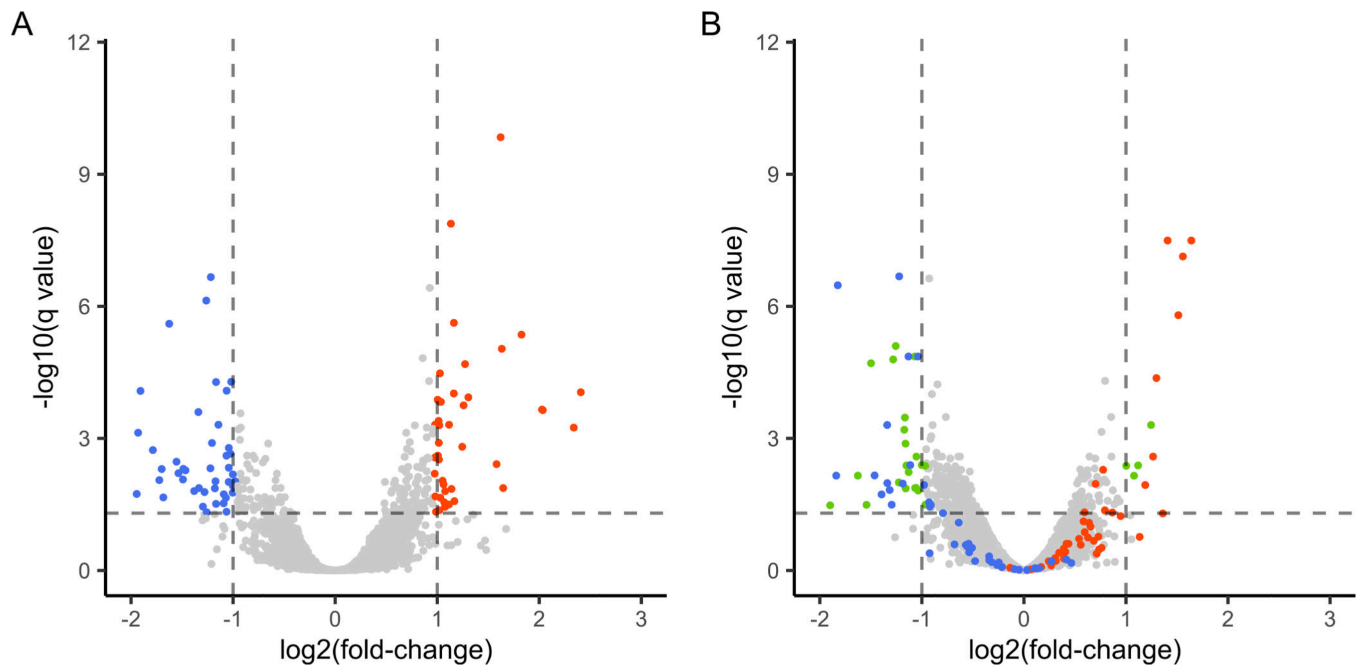

Figure 1. RA-associated expression among RA women who improved during pregnancy

Volcano plots showing differential expression between RA women who improved during pregnancy ($RA_{improved}$) and healthy women at two time points. (A) At pre-pregnancy (T0), 89 genes were differentially expressed (q<0.05; fold-change (FC) 2) between $RA_{improved}$ and healthy women, some being over-expressed in RA (orange dots) and others under-expressed (blue dots). (B) At the third trimester (T3), when RA improved, 65 of the 89 genes (73%) were no longer differentially expressed (orange and blue dots with −1 log_2(FC) 1 and –log_{10}(q-value)<1.3). Genes that became newly differentially expressed at T3 are shown as green dots.

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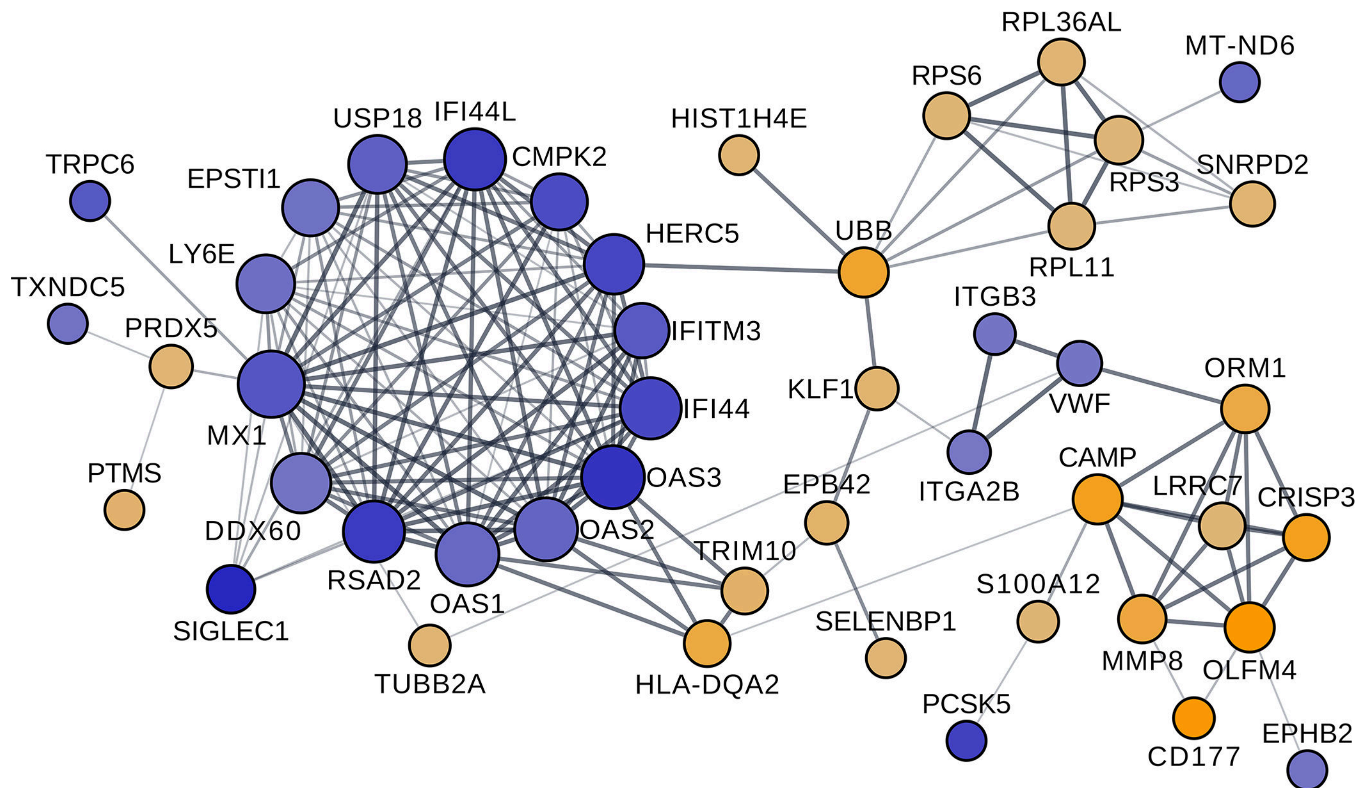

Figure 2. Protein network showing genes differentially expressed at the pre-pregnancy baseline within the same functional network

A large proportion of the 89 genes differentially expressed between $RA_{improved}$ women and healthy women at the pre-pregnancy baseline encode proteins that belong to a common functional network, based on protein interactions data from the STRING database. Most of the under-expressed genes (blue circles) formed a tight cluster, distinct from the over-expressed genes (orange circles).

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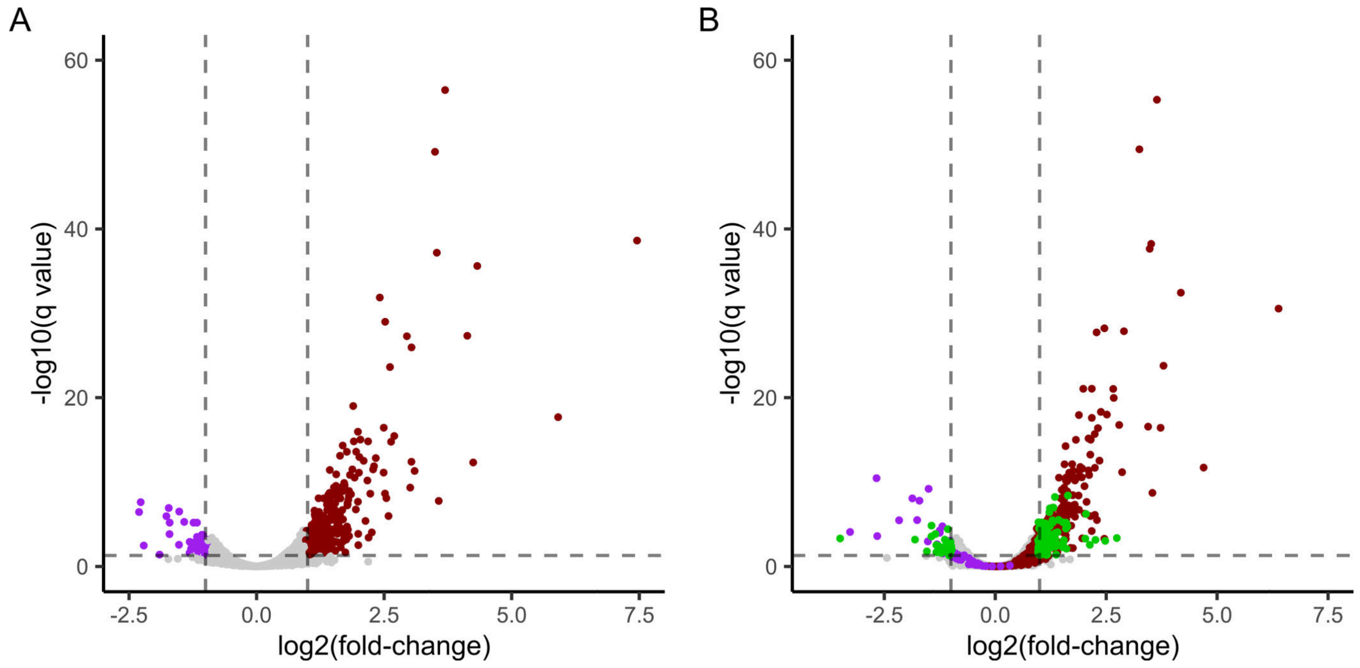

Figure 3. RA-associated expression among RA women who worsened during pregnancy

Volcano plots showing differential expression between RA women who worsened during pregnancy ($RA_{worsened}$) and healthy women before and during pregnancy. (A) At pre-pregnancy (T0), 429 genes were differentially expressed (q<0.05; fold-change (FC) 2) between $RA_{worsened}$ and healthy women, consisting of some that were over-expressed in RA (maroon dots) and some that were under-expressed (purple dots). (B) At the 3rd trimester (T3), when RA worsened, 207 of the 429 genes (48%) were no longer differentially expressed (maroon and purple dots with -1 $\log_2$(FC) 1 and $-\log_{10}$(q-value)<1.3). Numerous genes (green dots) became newly differentially expressed at T3 when disease activity worsened.

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Table 1

Medication use among the women with RA at each time-point

Patient	Pre-pregnancy	3rd trimester
Improved:		
1	None	None
2	None	None
3	None	Prednisolone + Sulfasalazine
4	Sulfasalazine	Sulfasalazine
5	Sulfasalazine	Prednisolon
6	Prednisolone + Infliximab	Prednisolone + Sulfasalazine
Worsened:		
7	Prednisolone + Sulfasalazine + Etanercept	Prednisolone + Sulfasalazine
8	Sulfasalazine + Adalimumab	Adalimumab
9	Infliximab	None

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Table 2

Gene ontology (GO) biological processes enriched in genes with RA-associated expression before pregnancy among $RA_{improved}$ women

Gene Set	Description	Enrichment ratio	q value
GO:0032069	Regulation of nuclease activity	46.6	7.6E-04
GO:0060337	Type I interferon signaling pathway	21.4	5.3E-05
GO:1903901	Negative regulation of viral life cycle	20.8	3.1E-04
GO:2001244	Positive regulation of intrinsic apoptotic signaling pathway	18.6	1.5E-02
GO:0045087	Innate immune response	5.9	1.7E-06
GO:0043312	Neutrophil degranulation	5.9	1.1E-03
GO:0051707	Response to other organism	5.1	1.7E-05
GO:0050878	Regulation of body fluid levels	4.8	2.2E-02