

Multiple Stepwise Gene Knockout Using CRISPR/Cas9 in *Escherichia coli*

Enrico König[#], Francesca Zerbini[#], Ilaria Zanella[#], Davide Fraccascia and Guido Grandi^{*}

Synthetic and Structural Vaccinology Unit, CIBIO, University of Trento, Via Sommarive, 9, Trento, Italy

*For correspondence: guido.grandi@unitn.it

[#]These authors contributed equally to this work

[Abstract] With the recent implementation of the CRISPR/Cas9 technology as a standard tool for genome editing, laboratories all over the world are undergoing one of the biggest advancements in molecular biology since PCR. The key advantage of this method is its simplicity and universal applicability for species of any phylum. Of particular interest is the extensively studied Gram-negative bacterium *Escherichia coli*, as it is considered as the workhorse for both research and industrial purposes. Here, we present a simple, robust and effective protocol using the CRISPR/Cas9 system in combination with the λ Red machinery for gene knockout in *E. coli*. Crucial in our procedure is the use of a double-stranded donor DNA and a curing strategy for removal of the guide RNA encoding plasmid that allows starting a new mutation after only two working days. Our protocol allows multiple, stepwise gene knockout strains with high mutagenesis efficiencies applicable for high-throughput approaches.

Keywords: Biotechnology, Synthetic biology, High-throughput, Genome editing, Double-stranded donor DNA

[Background] The Gram-negative bacterium *Escherichia coli* is one of the most important organisms for biotechnological engineering. It has been successfully implemented in a wide range of processes in different sectors of industry, such as energy, agriculture, food production, biotechnology, and medicine. The biotechnology sector is improving fast due to a constant technological progress. In particular, the CRISPR/Cas9 technology is probably the biggest revolution in (molecular) biology since PCR (Ledford, 2015). Briefly, CRISPR/Cas9 protects bacteria from invasive genetic elements such as plasmids and viruses (Marraffini, 2015). Taking advantage of this kind of acquired immune system from prokaryotes, a remarkably powerful tool for genome editing based on the CRISPR/Cas9 systems has been developed (Jinek *et al.*, 2012).

The CRISPR/Cas9 system consists of a guide RNA (gRNA) and the endonuclease Cas9 both forming a complex in a way that directs the enzyme to the target site that is complementary to the gRNA promoting site-specific cleavage (Sander and Joung, 2014). The enzymatically generated double strand break (DSB) is being subsequently repaired by the cell through either homologous recombination (HR) or non-homologous end joining (NHEJ) (Iyama and Wilson III, 2012). However, NHEJ works poorly in *E. coli* (Shuman and Glickman, 2007; Wright *et al.*, 2016) and thus genome editing protocols cannot rely on such a mechanism for this Gram-negative bacterium. Therefore, most approaches aiming to modify chromosomal DNA in *E. coli* exploit the phage recombinase-mediated homologous recombination (recombineering) using either the λ Red prophage system (Zhang *et al.*, 1998; Datta *et al.*, 2006) or the

three bacteriophage λ Red proteins Exo, Beta, and Gam (Murphy, 1998; Muyrers *et al.*, 1999; Ellis *et al.*, 2001). In fact, the presence of the λ Red machinery considerably increases mutagenesis efficiency as demonstrated in a recent study (Pyne *et al.*, 2015), especially when combined with the CRISPR/Cas9 technology (Jiang *et al.*, 2013).

Our laboratory established a robust, simple and rapid protocol for multiple, stepwise gene knockout in *E. coli* using the CRISPR/Cas9 technology in combination with the λ Red machinery (Zerbini *et al.*, 2017). Here, we want to provide our genome editing protocol in detail, which results in high mutagenesis efficiencies and has the potential to be applied in high-throughput approaches.

Materials and Reagents

1. Pipette tips: GPR-10G (Mettler-Toledo, Rainin, catalog number: 17001862), GPR-250 (Mettler-Toledo, Rainin, catalog number: 17001861), GPR-1000 (Mettler-Toledo, Rainin, catalog number: 17001859)
2. 50 ml tube (Corning, Falcon®, catalog number: 352070)
3. Primo® Vacuum Filter Systems, 250 ml, 0.22 μ m, PES (Euroclone, catalog number: EPVPE22250)
4. Syringe filters PES 0.22 μ m filters (Euroclone, catalog number: EPSPE2230)
5. Petri dishes (Corning, Falcon®, catalog number: 351007)
6. MAX efficiency™ DH5 α ™ competent cells (Thermo Fisher Scientific, Invitrogen™, catalog number: 18258012)
Note: You can also use homemade competent cells of E. coli DH5 α (see Step E1).
7. pCasRed plasmid (Zerbini *et al.*, 2017), modified pCas9 plasmid from Addgene (Addgene, catalog number: 42876) available upon request
8. pCRISPR-SacB plasmid (Zerbini *et al.*, 2017), modified pCRISPR plasmid from Addgene (Addgene, catalog number: 42875), available upon request
9. T4 Polynucleotide Kinase (PNK) (New England Biolabs, catalog number: M0201S)
10. T4 Polynucleotide Kinase (PNK) Reaction Buffer (New England Biolabs, catalog number: B0201S)
11. Oligonucleotide 1 or gDNA forward (Sigma-Aldrich; desalted, synthesis scale 0.025 μ M) for cloning of the mutagenic pCRISPR-SacB-gDNA (see Procedure B)
12. Oligonucleotide 2 or gDNA reverse (Sigma-Aldrich; desalted, synthesis scale 0.025 μ M) for cloning of the mutagenic pCRISPR-SacB-gDNA (see Procedure B)
13. T4 ligase (New England Biolabs, catalog number: M0202S)
14. 10x T4 ligase buffer (New England Biolabs, catalog number: B0202S)
15. 1 M NaCl (Sigma-Aldrich, catalog number: S7653-1KG)
16. *Bsa*I enzyme (New England Biolabs, catalog number: R0535L)
17. *Dpn*I enzyme (New England Biolabs, catalog number: R0176S)
18. Alkaline phosphatase, Calf Intestinal (CIP, New England Biolabs, catalog number: M0290S)

19. Wizard® SV Gel and PCR Clean-Up System (Promega, catalog number: A9282)
20. QIAprep Spin Miniprep Kit (QIAGEN, catalog number: 27106)
21. ATLAS ClearSight DNA Stain (BioAtlas, catalog number: BH40501)
22. Magnesium chloride (MgCl₂) (Sigma-Aldrich, catalog number: M8266-1KG)
Note: See Recipes for preparation of the 100 mM working solution.
23. Calcium chloride dihydrate (CaCl₂·2H₂O) (Sigma-Aldrich, catalog number: C3306-250G)
Note: See Recipes for preparation of the 100 mM working solution in 15% glycerol.
24. GoTaq Green Master Mix, 2x (Promega, catalog number: M7123)
25. LB broth Miller (Sigma-Aldrich, catalog number: L3522-1KG)
Note: See Recipes for preparation of LB medium.
26. LB agar (Sigma-Aldrich, catalog number: L2897-1KG)
Note: See Recipes for preparation of Petri dishes with LB-agar.
27. Kanamycin (Thermo Fisher Scientific, catalog number: 11815024)
Note: Prepare a stock in a concentration of 50 mg/ml (see Recipes).
28. Chloramphenicol (Sigma-Aldrich, catalog number: C0378-25G)
Note: Prepare a stock in a concentration of 25 mg/ml (see Recipes).
29. L-(+)-Arabinose (Sigma-Aldrich, catalog number: A3256-100G)
Note: Prepare a stock in a concentration of 20% L-(+)-arabinose (see Recipes).
30. Sucrose (Sigma-Aldrich, catalog number: S5016-2.5KG)
Note: Prepare a stock in a concentration of 50% sucrose (see Recipes).
31. > 99% Glycerol (Acros Organics, catalog number: 158920010)
Note: Prepare a 75% stock (see Recipes).
32. LB medium (liquid) (see Recipes)
33. LB-agar (for Petri dishes) (see Recipes)
34. Antibiotics (see Recipes)
35. 20% L-(+)-arabinose (see Recipes)
36. 50% sucrose (see Recipes)
37. 75% glycerol solutions (see Recipes)
38. Sterile 100 mM MgCl₂ (see Recipes)
39. Sterile 100 mM CaCl₂ in 15% glycerol (see Recipes)

Equipment

1. Pipettes: Pipetman P2L metal ejector (Gilson, catalog number: FA10001M), Pipetman P10L metal ejector (Gilson, catalog number: FA10002M), Pipetman P20L metal ejector (Gilson, catalog number: FA10003M), Pipetman P200L metal ejector (Gilson, catalog number: FA10005M), Pipetman P1000L metal ejector (Gilson, catalog number: FA10006M)
2. pH meter (e.g., Fisher Scientific, model: Fisher Scientific™ accumet™ AB150, catalog number: 13-636-AB150)

3. -80 °C freezer for storage of strains (any commercially available freezer is suitable)
4. -20 °C freezer (any commercially available freezer is suitable)
5. Autoclave (e.g., Tuttnauer, model: 5075 EL, 160 L)
6. Centrifuge, low temperature, up to 6,000 rpm (e.g., Thermo Fisher Scientific, Thermo Scientific™, model: Sorvall™ ST16)
7. DNA sequencer (or DNA sequencing service EUROFINS)
8. Gene Pulser®/MicroPulser™ Electroporation Cuvettes, 0.1 cm gap (Bio-Rad Laboratories, catalog number: 1652089)
9. MicroPulser™ Electroporator (Bio-Rad Laboratories, catalog number: 1652100)
10. Incubator, temperature scale at least 30-37 °C, shaking range 0-300 r.p.m. (e.g., Eppendorf, New Brunswick™, model: Innova® 44)
11. MilliQ water system (e.g., Elga Veolia, model: PURELAB® flex 3)
12. Spectrophotometer (e.g., Biochrom, model: WPA CO7500 Colorimeter)
13. Thermocell mixing block (e.g., BionovaTec, model: MB 102)
14. Thermocycler for PCR (e.g., Eppendorf, model: Mastercycler® nexus)
15. Transilluminator documentation system (e.g., UviTEC Cambridge, Laboratorium Life Science, model: UVIdoc HD5)

Procedure

Generally, our CRISPR/Cas9 knockout protocol for *Escherichia coli* employs three main components:

- 1) pCasRed plasmid (Procedure A)
- 2) pCRISPR-SacB-gDNA plasmid (Procedure B)
- 3) The mutagenic double-stranded donor DNA oligonucleotide (Procedure C).

First, we transform with the pCasRed plasmid (1), which encodes not only the Cas9 endonuclease but also the λ Red machinery (see Procedure A for details). In a subsequent step, we then co-transform with the pCRISPR-SacB-gDNA plasmid (2) and the mutagenic double-stranded donor DNA (3).

Our protocol results in the inactivation of a target gene by directing the Cas9 endonuclease to the 5'-end of the target locus through a specifically designed guide RNA, encoded and previously cloned into the pCRSIPR-SacB-gDNA plasmid (see Procedure B for details). Upon DNA cleavage at the target locus, the mutagenic double-stranded donor DNA (ds-dDNA) allows for λ Red-assisted repair of the double strand break, thereby introducing an in-frame stop codon and removing the original *protospacer adjacent motif* (PAM) region (Figure 1). A workflow is shown in Figure 2.

When required, plasmid curing can be executed through sucrose selection, following Hale *et al.* (2010) (for other references see Gay *et al.*, 1983; Steinmetz *et al.*, 1983; Gay *et al.*, 1985; Murphy *et al.*, 2000). Briefly, the *SacB* gene encodes for levansucrase from *Bacillus subtilis*, which metabolizes sucrose into a toxic product if *E.coli* grows in the presence of this disaccharide. Therefore, cells will only grow if they have lost the pCRISPR-SacB-gDNA plasmid.

Concerning the length of deletion and the length of the ds-dDNA you may adapt this protocol for your own needs. Here, we generally describe 30 nucleotide deletions, corresponding to the target site using ds-dDNA of 70 base pairs. However, in our laboratory, we observed that longer deletions of up to 2,200 nucleotides work better with ds-dDNA of 120 base pairs (Zerbini *et al.*, 2017).

Before you get started, you should start with the design of guides (Procedure B, Figure 3), donors and primers (Procedure C, Figure 4 and Figure 5, respectively). We recommend ordering them before moving to the wet laboratory procedures. We also recommend referring to the oligonucleotide sequences (target sites, donor DNAs, primers for colony PCR) published in our original research paper (Zerbini *et al.*, 2017) if you want to include any positive control in your own experiments.

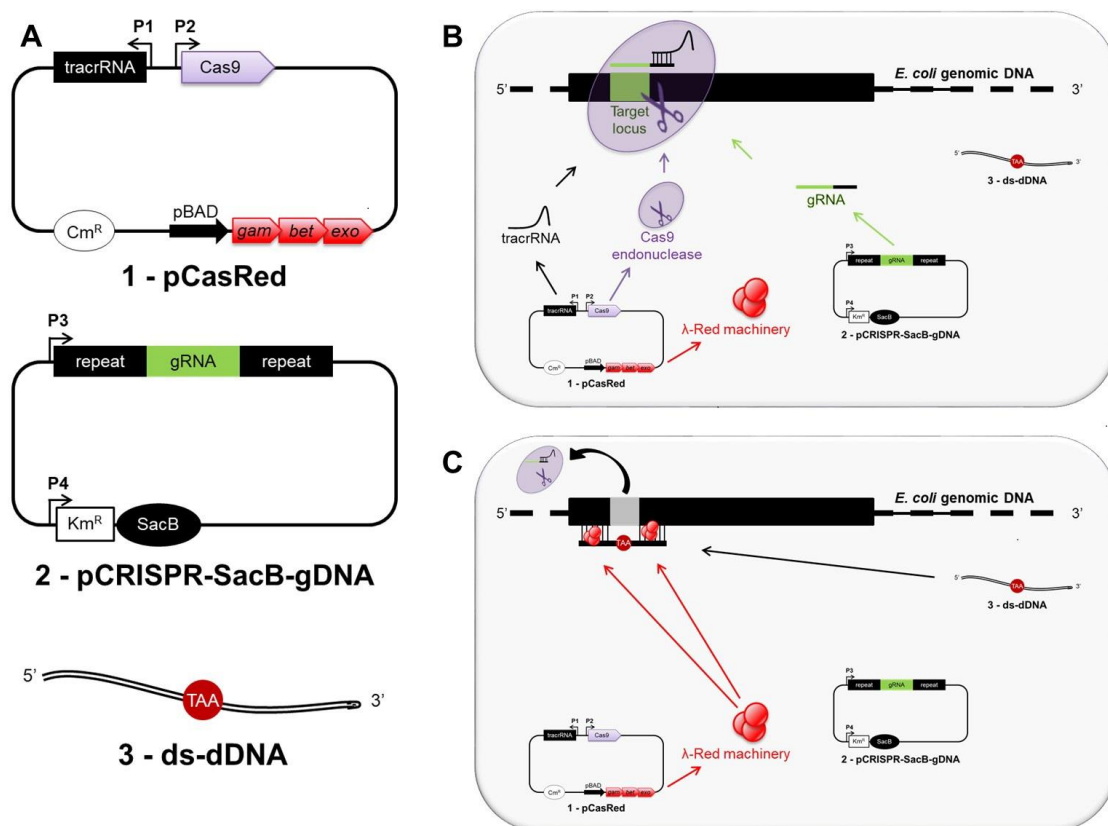


Figure 1. Overview of CRISPR/Cas9 genome editing strategy in *Escherichia coli*. A shows all three main components required for genome editing in *E. coli*: 1) the pCasRed plasmid expressing the λ Red (Exo, Beta, Gam) machinery, the Cas9 endonuclease, and the tracrRNA; 2) the pCRISPR-SacB-gDNA plasmid encoding the gRNA and the *SacB* gene; 3) a synthetic, double-stranded mutagenic oligonucleotide (ds-dDNA). After transformation, the tracrRNA anneals to the gRNA, which specifies the site of cleavage for Cas9 resulting in a three-component complex at the target locus, where the endonuclease activity mediates a chromosomal DNA double strand break (B). The double strand break is subsequently repaired by λ Red-mediated homologous recombination taking place between the extremities of the cleaved chromosomal DNA and the ds-dDNA (C).

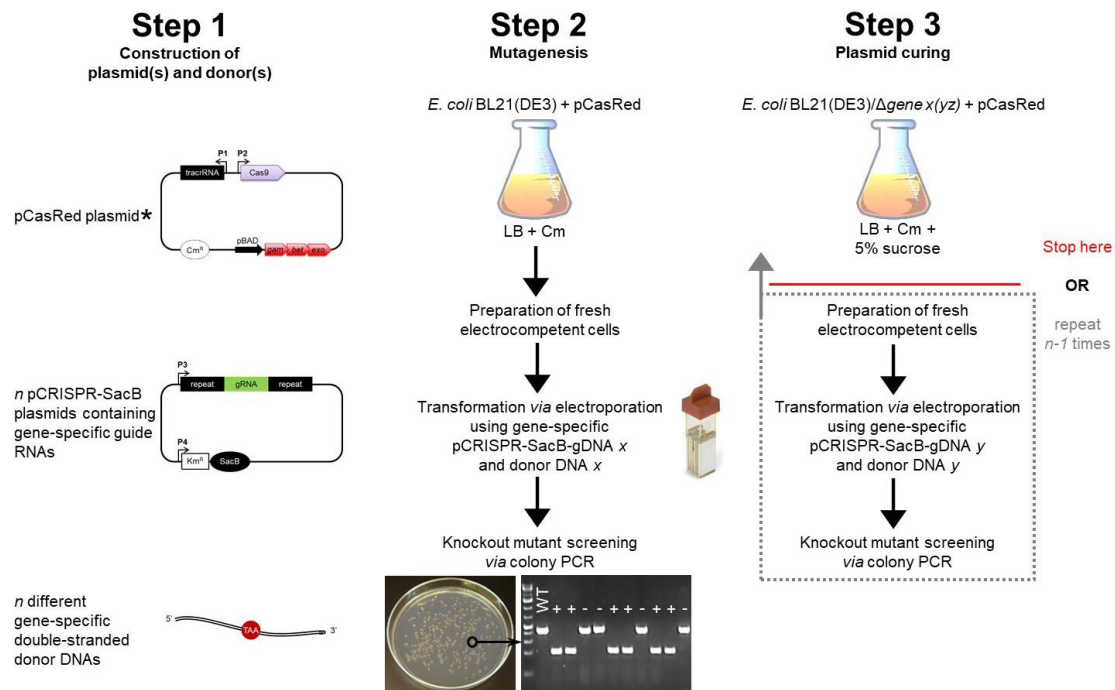


Figure 2. Schematic workflow for gene knockout in *E. coli* using the CRISPR/Cas9 technology.

Step 1 includes the construction of the three main components needed for this protocol (see Figure 1). The *E. coli* strain you want to gene-edit (e.g., *E. coli* BL21(DE3)) has to be transformed with the pCasRed plasmid first (note asterisk, see Step E4a.i). The *E. coli* + pCasRed strain is the starting point for mutagenesis (Step 2), in which fresh electrocompetent cells are prepared (Step E3) and directly used for transformation (Step E4). The next day, a colony PCR is performed to identify positive knockout mutants (Steps E5 and E6), which are subsequently inoculated in liquid LB medium containing 5% sucrose for curing of the pCRISPR-SacB-gDNA used before (Step 3). Here, the protocol can be stopped or you continue with another round of mutagenesis following only the procedures from Step 2 highlighted in the dotted box. Legend: chloramphenicol (Cm), wild type (WT), positive mutant (+), escaper (-).

A. Required plasmids

1. The **pCasRed plasmid** (Zerbini *et al.*, 2017) derives from the pCas9 plasmid from Addgene (#42876) and is available from our laboratory upon request.

Briefly, pCasRed encodes the Cas9 nuclease, the λ Red (Exo, Beta, Gam) cassette, the trans-activating CRISPR RNA (tracrRNA), and contains a chloramphenicol resistance gene. The Cas9 gene and the tracrRNA coding sequence are under the control of constitutive promoters while the λ Red gene cassette is transcribed using an arabinose-inducible promoter (pBAD).

2. The **pCRISPR-SacB plasmid** (Zerbini *et al.*, 2017) is available from our laboratory upon request. It derives from the pCRISPR plasmid LB and the pKM154 plasmid from Addgene (#42875 and #13036, respectively).

Briefly, pCRISPR-SacB contains a cloning site, which allows insertion of a self-designed (see Procedure B), synthetic DNA fragment coding for the desired guide RNA, between two repeat

the gDNA is the reverse complement of the protospacer + PAM forward sequence with the addition of five base pairs (in red) as done for the forward oligonucleotide.

4. The synthetic DNA fragments have to be cloned into the pCRISPR-SacB as follows:
 - a. Use T4 PNK to phosphorylate the two complementary single-stranded synthetic oligonucleotides encoding the gDNA.
 - 1 μ l oligonucleotide 1 (100 μ M)
 - 1 μ l oligonucleotide 2 (100 μ M)
 - 5 μ l 10x T4 ligase buffer
 - 1 μ l T4 PNK
 - 42 μ l MilliQ water (final volume 50 μ l)

Note: If you have oligonucleotides diluted to concentrations other than 100 μ M (e.g., 50 μ M), make sure you adjust the volume of oligonucleotide 1 and 2 (e.g., 2 μ l) and MilliQ water (e.g., 40 μ l) to get a final volume of 50 μ l.
 - b. Incubate for 30 min at 37 °C.
 - c. For annealing, add 2.5 μ l of 1 M NaCl to the reaction mix and heat to 95 °C for 5 min.
 - d. Allow to cool down slowly to room temperature by simply switching off the thermoblock (ca. 2 h). In the meanwhile, proceed with Steps B4e-B4h.
 - e. Digest the pCRISPR-SacB with *Bsa*I.
 - 10 μ l pCRISPR-SacB (100 ng/ μ l, total amount ca. 1-2 μ g)
 - 1 μ l *Bsa*I
 - 5 μ l 10x NEB Buffer
 - 34 μ l MilliQ water (final volume 50 μ l)

Note: If your pCRISPR-SacB plasmid is diluted to concentrations other than 100 ng/ μ l (e.g., 50 ng/ μ l), make sure you adjust the volume of the plasmid (e.g., 20 μ l) and MilliQ water (e.g., 24 μ l) to get a final volume of 50 μ l.
 - f. Incubate for 1 h at 37 °C.
 - g. Dephosphorylate the digested pCRISPR-SacB plasmid using Calf Intestinal Alkaline Phosphatase (CIP).
 - 5 μ l 10x CutSmart Buffer
 - 1 μ l CIP
 - 25 μ l pCRISPR-SacB plasmid cleaned in a PCR cleaning column (20 ng/ μ l, total amount of 500 ng plasmid)
 - 19 μ l MilliQ water

Note: If the purified pCRISPR-SacB plasmid is diluted to concentrations other than 20 ng/ μ l (e.g., 15 ng/ μ l), make sure you adjust the volume of the plasmid (e.g., 33.3 μ l) and MilliQ water (e.g., 10.7 μ l) to get a final volume of 50 μ l. The concentration of your plasmid after purification should therefore not be lower than 11.4 ng/ μ l!
 - h. Incubate for 2 h at 37 °C.

- i. Purify the plasmid directly from the reaction mix using the Wizard® SV Gel and PCR Clean-Up System.
- j. To generate the pCRISPR-SacB-gDNA plasmid perform a final ligation step as follows:
 - 3 µl digested pCRISPR-SacB (10 ng/µl, total amount of ca. 30 ng plasmid)
 - 2 µl phosphorylated, double-stranded gDNA (1 µM)
 - 1 µl T4 ligase
 - 2 µl T4 ligase buffer
 - 12 µl MilliQ water

Note: You may use different volumes, but make sure the concentrations remain the same in a final volume of 20 µl.
- k. Incubate for 2 h at room temperature.

Note: Alternatively, if your time does not allow proceeding the same day, you may also incubate overnight at 16 °C.
- l. The ligation reaction (10 µl) is used to transform *E. coli* DH5α chemically competent cells to obtain the pCRISPR-SacB-gDNA plasmid (see Step E2 for transformation via heat-shock).

Note: If you do not want to use commercial E. coli DH5α cells, you can also prepare your own batch of chemically competent cells (Step E1).
- m. Incubate at 37 °C with vigorous shaking (200 rpm) for 1 h.
- n. Plate the transformation mix on LB-agar (see Recipes) containing kanamycin (50 µg/ml).

Note: As transformation efficiency should be high at this stage, do not plate the entire transformation mix but rather 100 µl or 200 µl only.
- o. Incubate the LB-agar plate at 37 °C overnight.
- p. Inoculate 5 ml LB containing kanamycin (50 µg/ml) from a single colony and incubate at 37 °C overnight.
- q. Isolate the plasmid from the overnight culture using a QIAprep Spin Miniprep Kit.
- r. Analyze the plasmid by DNA sequencing using a primer (primer sequence: 5'-GCCAGTCATAGCCGAATAG-3') for the plasmid annealing downstream of the inserted fragment encoding the guide. The primer can be used for all pCRISPR-SacB-gDNAs and is therefore universal for every cloning of different guides.

If positive, your pCRISPR-SacB-gDNA plasmid is ready for the following mutagenesis step (Step E4a.ii).

C. Design of mutagenic donor DNA and primers for PCR screening

1. We recommend designing your mutagenic donor DNA (dDNA) in a way that will result in the deletion of 30 nucleotides ($\Delta 30$) within your target gene through the removal of the protospacer and the PAM region (to avoid post-mutation Cas9 cleavage). At the same time, an in-frame stop codon has to be inserted close to the 5'-end of the target gene, thereby inactivating it. Note that the trinucleotide encoding the stop codon has no complementary sequence on the target strand (Figure 4). In our hands, this strategy works well to achieve gene knockout. For a schematic

example, refer to Figure 4. Note that the length of the deletion can be adapted to your needs. However, in case of longer deletions you may have to increase also the length of the dDNA.

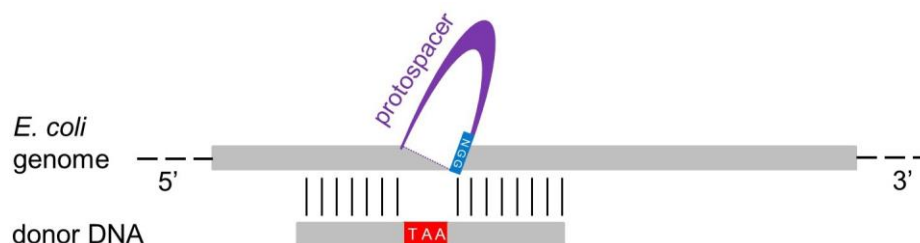


Figure 4. Design of the dDNA. The donor DNA (dDNA) can vary in length (e.g., 70 or 120 nucleotides, see text). To delete the protospacer + PAM region identified for Cas9 cleavage (Figure 3), the 5' moiety of the dDNA (35 or 60 nucleotides) has to be complementary to the sequence upstream of the protospacer region, while the 3' moiety of the dDNA (35 or 60 nucleotides) has to be complementary to the sequence downstream of the protospacer. To inactivate the transcript, an in-frame stop codon (TAA) is inserted, and the PAM region (NGG) is also eliminated to prevent further cleavage through Cas9. Finally, a reverse complement oligonucleotide is designed to get a double-stranded dDNA.

2. We explicitly recommend the use of double-stranded mutagenic oligonucleotides (ds-dDNAs), preferably in HPLC purification grade, as this considerably increases mutagenesis efficiency (Zerbini *et al.*, 2017). Thus, design both forward and reverse oligonucleotides. For $\Delta 30$ deletions, we recommend dDNA of 70 nucleotides in length.
3. Finally, the ds-dDNA is generated by annealing 10 μg of each complementary oligonucleotide pair in a total volume of 20 μl at 95 $^{\circ}\text{C}$ for 5 min and allowing the reaction mix to slowly cool down to room temperature by switching off the thermoblock. The annealing reaction is verified by loading 500 ng of each single stranded oligonucleotides and 1 μg of total DNA in the annealing reaction and by visualizing the bands using ATLAS ClearSight DNA Stain. The band corresponding to the single stranded oligonucleotides must disappear in the annealing reaction sample.
4. The primers necessary for colony PCR screening following the mutagenesis step (Step E4a.ii) should be designed in order to get an amplified DNA fragment of ca. 200 bp, a fragment length that is ideal to distinguish the difference of a 30 bp deletion between the wild-type colony and a mutated colony. For a schematic example, refer to Figure 5.

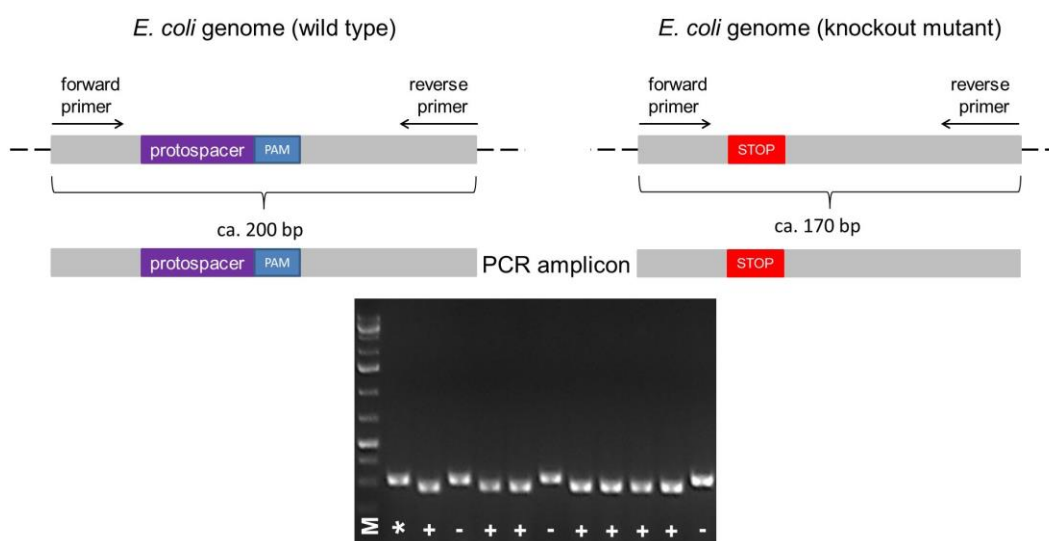


Figure 5. Illustration of the colony PCR. The forward and reverse primers for PCR are designed to anneal upstream and downstream of the protospacer + PAM sequence, respectively, resulting in the amplification of a ~200 bp fragment in the wild type. According to the size of deletion (e.g., $\Delta 30$), a knockout mutant can be identified by its shorter PCR fragment (e.g., ~170 bp) compared to the wild type (A). The PCR fragments different in size can be identified by running a 2% agarose gel electrophoresis (B). Legend: marker (M), wild type control (*), mutant (+), wild type 'escaper' (-).

D. Bacterial strains and culture conditions

Note: Make sure you use the correct antibiotic required for selection of your E. coli strain. We routinely use the DH5 α and BL21(DE3) strain.

1. For any experimental step, grow the *Escherichia coli* strain in LB broth (Sigma-Aldrich) at 37 °C with vigorous shaking (200 rpm), if not stated otherwise.
2. Prepare stocks of your strains in LB + glycerol (15-25% final concentration) and store them at -80 °C.

Example: For a stock of 1 ml prepare 200 μ l of 75% glycerol and add 800 μ l of your strain after overnight growth.

3. Start from an overnight culture taken from a frozen/glycerol stock before each bacterial manipulation step, if not stated otherwise.

E. Gene knockout using CRISPR/Cas9

For preparation of the pCRISPR-SacB-gDNA plasmids (see Step B4) we recommend using *E. coli* DH5 α cells, which have to be made chemically competent and transformed via heat-shock (Steps E1 and E2). Instead, the strain you want to use for all mutagenesis experiments should be prepared for electroporation (Steps E3 and E4) to increase transformation efficiency.

The strain should be first transformed via electroporation (Step E4) with the pCasRed plasmid (from Step A1) before transforming via electroporation with the pCRISPR-SacB-gDNA plasmid (generated in Step B4).

1. Preparation of chemically competent cells

- a. Streak *E. coli* DH5 α on an LB-agar plate and incubate at 37 °C overnight.
- b. Inoculate a pre-culture of 5 ml LB medium (see Recipes) from a single colony and incubate at 37 °C with vigorous shaking (200 rpm) overnight.
- c. Dilute the overnight culture to an OD_{600 nm} of 0.1 in LB (e.g., 50 ml) and incubate at 37 °C with vigorous shaking (200 rpm) until an OD_{600 nm} of 0.4-0.6 is reached (ca. 2 h).
- d. Transfer the culture to a 50 ml tube and harvest the cells by centrifugation at 4 °C, 2,500 x *g* for 20 min.
- e. Discard the supernatant and resuspend the pellet in 5 ml of ice-cold, sterile 100 mM MgCl₂.
- f. Keep the suspension on ice for 30 min.
- g. Harvest the cells by centrifugation at 2,500 x *g* for 20 min.
- h. Discard the supernatant and gently resuspend the cell pellet in 1 ml of ice-cold, sterile 100 mM CaCl₂ in 15% glycerol and prepare aliquots of 110 μ l.

Note: If you started with a higher culture volume keep a ratio of 10 ml 100 mM CaCl₂ in 15% glycerol/50 ml starting culture for resuspension.

- i. Use the aliquots freshly for the subsequent transformation via heat-shock (see Step E2)

Note: If some aliquots remain, store them at -80 °C.

2. Transformation via heat-shock

- a. Mix 50 μ l of chemically competent cells with 10 μ l of ligation mix (see Step B4i).
- b. Keep the reaction mix on ice for 30 min.
- c. Transfer the cells to a thermoblock set at 42 °C for 45 sec.
- d. Keep the cells on ice for 2 min.
- e. Add 940 μ l LB and incubate the cells at 37 °C with vigorous shaking (200 rpm) for 1 h.
- f. Plate the cells on LB-agar plates containing the appropriate antibiotic (chloramphenicol 25 μ g/ml for transformation with pCasRed plasmid or kanamycin 50 μ g/ml for transformation with pCRISPR-SacB-gDNA)

Note: We recommend plating 100 μ l and 900 μ l of the transformation mix separately, but if you prefer you may choose different volumes (e.g., 500 μ l or 250 μ l).

- g. Incubate at 37 °C overnight.
- h. Count the colonies to determine the transformation efficiency of your chemically competent cells.

3. Preparation of electrocompetent cells

- a. Streak your *E. coli* strain on an LB-agar plate and incubate at 37 °C overnight.
- b. Inoculate a pre-culture of 5 ml LB medium (see Recipes) from a single colony and incubate at 37 °C with vigorous shaking (200 rpm, overnight)

Note: If you already performed a mutation round, use the overnight culture from the plasmid curing step (see Step E7).

- c. Dilute the overnight culture to an OD_{600 nm} of 0.1 in LB (e.g., 50 ml) and incubate at 37 °C with vigorous shaking (200 rpm) until an OD_{600 nm} of 0.6-0.8 is reached (ca. 2-3 h).

Note: If you prepare competent cells from a strain previously transformed with the pCasRed plasmid, make sure your media contains chloramphenicol (25 µg/ml) and add L-(+)-arabinose (final concentration of 0.2%, see Recipes) at an OD_{600 nm} of 0.2 to the cells for the induction of λ Red genes.

- d. Transfer the culture into a 50 ml tube and harvest the cells by centrifugation at 4 °C, 4,100 x g for 20-30 min.
- e. Discard the supernatant and resuspend the pellet in 50 ml ice-cold, sterile MilliQ water. Centrifuge again at 4 °C, 4,100 x g for 10 min. Repeat this wash step three times.
- f. Finally, wash the pellet with ice-cold, sterile 10% glycerol and centrifuge again at 4 °C, 4,100 x g for 10 min.
- g. Discard as much supernatant as possible. Immediately resuspend the cell pellet in 250 µl ice-cold, sterile 10% glycerol and prepare 110 µl aliquots.

Note: If you started with a higher culture volume, keep a ratio of 250 µl 10% glycerol/50 ml starting culture for resuspension.

- h. Use the aliquots freshly for the subsequent transformation via electroporation (see Step E4).
- Note: If some aliquots remain, store them at -80 °C.*

4. Transformation via electroporation

- a. Electroporate 50 µl of competent cells using a 1 mm Gene Pulser cuvette at 1.8 kV with one of the following reaction mixes:

1 ng of pCasRed plasmid (see Step A1)

Mutagenesis step:

100 ng pCRISPR-SacB-gDNA (prepared in Step B4)

10 µg of the corresponding ds-dDNA (designed in Procedure C).

Note: Your control should consist in a transformation using 100 ng of the 'empty' pCRISPR-SacB (see Step B2) without adding any ds-dDNA.

- b. Add 950 µl LB and incubate the cells at 30 °C with vigorous shaking (200 rpm) for 3 h.
- c. Plate the cells on LB-agar plates containing chloramphenicol 25 µg/ml (and kanamycin 50 µg/ml, when performing the mutagenesis step).

Note: We recommend plating the entire transformation mix of the mutagenesis step (1 ml), but if you prefer you may choose different volumes (e.g., 500 µl or 250 µl).

- d. Incubate at 37 °C overnight.
 - e. Count the colonies to determine the transformation efficiency of your electrocompetent cells.
- Note: The control of your mutagenesis step should count ca. 10⁶ CFU/ml/µg. A drop in transformation efficiency of at least three orders of magnitudes is indicative for the quality of the gDNA (Zerbini et al., 2017).*

If you transformed with pCasRed plasmid start return to Step E3.

5. Colony PCR
 - a. Prepare a master mix for colony PCR using the GoTaq Green Master Mix.

Note: In our hands, screening of ten colonies is sufficient (Zerbini et al., 2017). Thus, prepare the master mix for 12 reactions (10 samples + 1 control, see Note from Step E4a.ii).

 - 11 x 1 µl primer, forward (10 µM)
 - 11 x 1 µl primer, reverse (10 µM)
 - 11 x 12.5 µl GoTaq
 - 11 x 10.5 µl MilliQ water
 - b. Pick individual colonies using a pipette tip and transfer directly to the PCR reaction mix. Make sure you will have enough cells from the colony for further analysis (see Step E7 and/or E8) by inoculating onto a transfer plate before dipping into the PCR reaction tube.
 - c. Start PCR with the following settings:
 - i. Initial denaturation 95 °C for 5 min
 - ii. 30 cycles of:
 - Denaturation 95 °C for 30 sec
 - Annealing 58-63 °C for 30 sec
 - Elongation 72 °C for 30 sec
 - iii. Final elongation 72 °C for 3 min

Note: Annealing temperature depends on the primer sequence. We designed them to use a range between 58-63 °C.
6. Verify the mutants by loading 4-8 µl of each PCR on a 2% agarose gel and run electrophoresis. Purify positive PCR amplicons ($\Delta 30$ between control and sample) using the Wizard® SV Gel and PCR Clean-Up System. Confirm the mutation by DNA sequencing using one of the primers you used for colony PCR above.
7. Curing of pCRISPR-SacB-gDNA plasmid

Note: If you are going to perform more than one knockout round, this step is mandatory. However, at the end of your genome editing experiment(s), removal of the pCRISPR-SacB-gDNA plasmid is optional.

 - a. To remove the pCRISPR-SacB-gDNA plasmid, pick a mutated colony (from Step E5b) and inoculate liquid LB medium containing 5% sucrose (see Recipes) and chloramphenicol (25 µg/ml).
 - b. Incubate at 37 °C with vigorous shaking (200 rpm) overnight.
 - c. Prepare a glycerol stock of your resultant strain for storage at -80 °C.

*Note: If you wish to **continue with a subsequent mutation round**, use the overnight culture for preparation of competent cells and **return to Step E3**.*
 - d. Control loss of the pCRISPR-SacB-gDNA plasmid by adding 10-20 µl drops on two LB-agar plates, one containing chloramphenicol (25 µg/ml) the other one containing chloramphenicol and kanamycin (25 µg/ml and 50 µg/ml, respectively). No growth should be visible on the

plate with the Cm and Km combination, indicating that all bacteria had lost the pCRISPR-SacB-gDNA plasmid.

8. Curing of pCasRed plasmid

Note: This step is optional. It is required if the presence of the pCasRed plasmid after the conclusion of your knockout experiment(s) may affect downstream experiments negatively. However, after removal of the pCasRed plasmid you may have to add a selection marker (i.e., antibiotic resistance) of your choice.

- a. To remove the pCasRed plasmid, pick a mutated colony (from Step E5b) or use a glycerol stock, inoculate in LB medium without any antibiotic, and incubate at 37 °C with vigorous shaking (200 rpm) overnight.
- b. Confirm the loss of the pCasRed plasmid by inoculating LB-agar plate containing chloramphenicol (25 µg/ml). No growth should be visible when using chloramphenicol, indicating that all bacteria have lost the pCasRed plasmid.

Data analysis

To verify if the mutation has been successfully obtained, a colony PCR must be performed (see Step E5) and subsequently analyzed via gel electrophoresis using a 2% agarose gel (see Step E6). The size shift can be observed and documented using a gel documentation system (Figure 5). If a colony results to have a fragment equal to the wild type control, it is a so-called *escaper* (Jiang *et al.*, 2013). Finally, DNA sequencing using one of the gene specific primers employed also in colony PCR (Step E5) must be performed to confirm the mutation.

Recipes

1. LB medium (liquid)
 - a. Weigh 25 g of LB broth Miller powder
 - b. Add MilliQ water to a final volume of 1 L
 - c. Stir until the solution is fully dissolved
 - d. Autoclave the medium
 - e. Store the solution at room temperature
2. LB-agar (for Petri dishes)
 - a. Weigh 20 g of LB agar powder
 - b. Add MilliQ water to a final volume of 500 ml
 - c. Stir until the solution is fully dissolved
 - d. Autoclave the medium
 - Note: If you do not pour the plates immediately, store the medium at room temperature.*
 - e. Allow the medium to cool down to ca. 45-50 °C before adding the antibiotics and mix
 - f. Pour the medium (ca. 10-15 ml) in Petri dishes

- g. Store your LB agar plates at 4 °C
3. Antibiotics
- Note: We recommend preparing aliquots of 1,000x stock solutions for each antibiotic. Store them at -20 °C.*
- a. Chloramphenicol: 25 mg/ml
- i. Weigh 250 mg of chloramphenicol powder under a safety hood
 - ii. Add MilliQ water to a final volume of 10 ml
 - iii. Mix until the solution is fully dissolved
 - iv. Sterilize the solution with a 0.22 µm syringe filter
 - v. Prepare 1 ml aliquots and store at -20 °C
- b. Kanamycin: 50 mg/ml
- i. Weigh 500 mg of kanamycin powder under a safety hood
 - ii. Add MilliQ water to a final volume of 10 ml
 - iii. Mix until the solution is fully dissolved
 - iv. Sterilize the solution with a 0.22 µm syringe filter
 - v. Prepare 1 ml aliquots and store at -20 °C
4. 20% L-(+)-arabinose
- a. Weigh 10 g of L-(+) arabinose
 - b. Add MilliQ water to a final volume of 50 ml
 - c. Mix until the solution is fully dissolved
 - d. Sterilize the solution with a 0.22 µm syringe filter
 - e. Store at room temperature
5. 50% sucrose
- a. Weigh 100 g of sucrose
 - b. Add MilliQ water to a final volume of 200 ml
 - c. Mix until the solution is fully dissolved
 - d. Sterilize the solution with a Primo® Vacuum Filter System, 0.22 µm, PES, 250 ml
 - e. Store the solution at RT
6. 75% glycerol solutions
- a. Take 75 ml of autoclaved 100% glycerol
 - b. Add 25 ml of autoclaved MilliQ water
 - c. Mix thoroughly
- Note: 100% glycerol is very viscous.*
7. 100 mM MgCl₂
- a. Weigh 95.21 mg of MgCl₂ and dissolve in 10 ml MilliQ water
 - b. Mix thoroughly
 - c. Sterilize the solution with a 0.22 µm syringe filter

8. 100 mM CaCl₂ in 15% glycerol
 - a. Weigh 147.01 mg of CaCl₂ dihydrate and dissolve in 8.5 ml MilliQ water
 - b. Add 1.5 ml of sterile 100% glycerol
 - c. Mix thoroughly
 - d. Sterilize the solution with a 0.22 µm syringe filter

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