

Biomarkers in Rheumatoid Arthritis

Samantha C. Shapiro¹

1. Rheumatology, University of Texas at Austin, Dell Medical School, Austin, USA

Corresponding author: Samantha C. Shapiro, samantha.shapiro@austin.utexas.edu

Abstract

The utilization and identification of biomarkers in rheumatoid arthritis (RA) to facilitate timely diagnosis and the optimal management of the disease is an area of active investigation. This review focuses on biomarkers available for routine clinical use, details potential investigational biomarkers, and raises outstanding clinical questions.

Categories: Rheumatology

Keywords: rheumatoid arthritis, biomarker, acpa, ccp, rf, esr, crp

Introduction And Background

Rheumatoid arthritis (RA) is a chronic and common systemic inflammatory disease that results in joint deformity and functional disability when not properly managed. The early diagnosis and treatment of RA are imperative for optimal disease control, greater chances of remission, and prevention of permanent clinical and radiographic damage. RA remains a clinical diagnosis although the use and discovery of biomarkers to assist with these goals remain a focus of ongoing research. The 2010 RA Classification Criteria developed by the American College of Rheumatology and European League Against Rheumatism (ACR/EULAR) define a scoring system that includes elements of history, physical exam, and biomarkers that identify patients with RA for the purpose of clinical trial standardization. The criteria have a sensitivity of 84% and specificity of 60% for classification as RA [1-2]. In clinical practice, rheumatologists frequently use these criteria to defend the diagnosis of RA. This review focuses on the four biomarkers included in these criteria that are available for routine clinical use: rheumatoid factor, autoantibodies against citrullinated proteins, erythrocyte sedimentation rate, and C-reactive protein; the multi-biomarker disease activity test is also discussed. A short discussion of investigational biomarkers and outstanding clinical questions follows.

Review

The current state of the utilization of biomarkers in the diagnosis, prognosis, and management of RA

Rheumatoid Factor (RF)

RFs are autoantibodies directed against the Fc portion of immunoglobulin (Ig) G. In clinical practice, IgM RF is most commonly measured although IgA and IgG RF also exist. RF are found in up to 80% of RA patients but can occur in a myriad of other inflammatory conditions that trigger chronic antigenic stimulation, limiting its specificity. These include but are not limited to other rheumatologic conditions (eg. systemic lupus erythematosus, Sjogren's syndrome), infectious diseases (eg. hepatitis C virus, subacute bacterial endocarditis, Epstein-Barr virus), malignancy (eg. B-cell neoplasms), and healthy individuals [3]. Smoking has also been associated with an increased prevalence of RF [4]. Approximately 30% to 45% of patients with early RA do not have RF, though some may develop RF later in the course of disease [5].

As with any diagnostic test, the positive predictive value of RF increases when utilized in patients with a high pre-test probability of disease (eg. those with inflammatory arthritis). Testing patients with nonspecific arthralgia, myalgia, or osteoarthritis is not recommended [6]. RF positivity increases the risk of developing RA, with higher titers conferring higher risk [7-8]. RF titers may fall with effective RA therapy but a fluctuation in RF titer does not correlate reliably with disease activity [9]. Serial monitoring of RF levels is not recommended [10-11]. In regards to the selection of RA therapy, RF positivity may increase the chance of response to B-cell depleting monoclonal antibodies (eg. rituximab) [12].

Autoantibodies Against Citrullinated Proteins (ACPA)

Autoantibodies to citrullinated protein epitopes have been a focus of biomarker research in RA for many years. Citrullination is a post-translational modification of proteins that can generate new epitopes to which the immune system is not tolerant, leading to the generation of new autoantibodies [13]. Amongst ACPAs, the assay for anti-cyclic citrullinated peptide (anti-CCP2) is widely clinically available and has excellent diagnostic and prognostic value.

Review began 04/19/2021

Review ended 05/10/2021

Published 05/16/2021

© Copyright 2021

Shapiro. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Both RF and anti-CCP2 have similar sensitivities for the diagnosis of RA but anti-CCP2 is more specific [14]. Anti-CCP2 is positive in 20%-30% of RA patients who are negative for RF [15]. A systemic review and meta-analysis that included 37 studies of anti-CCP2 positive patients and 50 studies of RF positive patients showed the pooled sensitivities of RF and anti-CCP2 to be 69% and 67%, and 85% and 95%, respectively [16]. This being said, anti-CCP2 positivity may be found in other rheumatologic diseases (eg. myositis, Sjogren's Syndrome), especially in the setting of erosive inflammatory arthritis [17]. Anti-CCP2 positivity may also occur with active pulmonary tuberculosis albeit with minimal rheumatologic symptoms [18]. High titer RF and anti-CCP2 antibodies are both associated with an increased risk of erosive joint damage; anti-CCP2 antibodies may confer a higher risk than RF [19-21]. High titer anti-CCP2 is associated with better clinical response to certain biologics (rituximab, abatacept) and thus may aid clinicians in personalizing therapy for the greatest chance of response [22-23].

Erythrocyte Sedimentation Rate (ESR) and C-reactive Protein (CRP)

The ESR, the rate at which erythrocytes fall through plasma when suspended in a vertical tube, is an indirect measure of the levels of acute-phase reactants (mainly fibrinogen). ESR levels are influenced by several factors, such as the size, shape, and number of red blood cells, as well as other plasma constituents like immunoglobulins. Elevated ESR levels may be caused by systemic or local inflammatory processes, infection, malignancy, tissue injury, end-stage renal disease, nephrotic syndrome, and obesity. ESR values increase with age and are slightly higher in women than men. Furthermore, many factors may contribute to spuriously low ESR values, like abnormal erythrocyte shape, extreme leukocytosis, heart failure, and cachexia [24]. Not surprisingly, the ESR is not a specific marker of inflammation.

CRP is an acute-phase reactant in the pentraxin protein family, which comprises pattern recognition molecules involved in the innate immune response [25]. CRP occurs in both acute and chronic inflammatory states, infectious and noninfectious. Low-grade CRP elevation is associated with various metabolic stressors, including but not limited to atherosclerosis, obesity, type 2 diabetes, sedentary lifestyles, unhealthy diet, and even being unmarried [26-27]. CRP levels vary with age, sex, and race, though less so than ESR levels [28]. Furthermore, there is no standardized reference range or unit of measure for CRP values; these vary between laboratories [29]. In the RA synovium, there is an overabundance of pro-inflammatory cytokines that stimulate the production of CRP by the liver, thus making it an attractive candidate as a disease activity biomarker [30]. However, CRP measurement in RA is not foolproof. For example, elevated CRP levels have been independently associated with truncal adiposity in women with RA, regardless of articular involvement or the use of biologic agents [31].

Although ESR and CRP measurements are imperfect, both continue to play a role in the diagnosis and management of RA. Elevated ESR and CRP levels are included in the 2010 ACR/EULAR Classification Criteria for RA [2]. CRP values of less than or equal to 1 mg/dL are included in the 2011 ACR/EULAR definition of RA remission used in clinical trials [32]. The ACR has endorsed six RA disease activity measures for use in clinical practice, two of which include ESR or CRP measurement: the Disease Activity Score 28-ESR or CRP (DAS28-ESR or DAS28-CRP) and the Simplified Disease Activity Index (SDAI) [33]. The 2015 ACR Guideline for the Treatment of RA, widely used in clinical practice, encourages the use of these disease activity measures, though does not specify a preference for measures that include laboratory values over those that do not. The guidelines also do not specifically recommend routine monitoring of ESR and CRP in all RA patients [34]. An update to these treatment guidelines is currently in progress, anticipated fall 2021.

Multiple studies have shown a correlation between ESR and CRP elevation, and radiographic and functional outcomes in patients with RA [30,35]. Elevated ESR is thought to be a better predictor of these outcomes in early RA, whereas CRP may be superior in later stages of the disease given less susceptibility to other factors like immunoglobulin levels and anemia [30]. This being said, ESR and CRP are normal in about 40% of patients with RA [36-37]. Furthermore, even in those patients with baseline elevation, values may remain stable despite clinical improvement with treatment [38]. Interestingly, ESR and CRP values may also be discrepant [24]. A large observational study that included over 9,000 patients from a practice-based registry noted discordant ESR and CRP values in 26% of patients, despite active RA as measured by joint counts and global assessments [39]. When results are discordant, they may no longer predict the progression of radiographic joint damage [40]. Lastly, biologic therapies like tocilizumab, a humanized monoclonal antibody against the interleukin-6 receptor, will normalize CRP values, eliminating utility as a trackable disease activity biomarker.

Multi-Biomarker Disease Activity (MBDA) Test

The MBDA test is a commercially available assay that measures 12 serum protein biomarkers and applies an algorithm to summarize the information into a single score that indicates the level of "RA inflammation." The following biomarkers are tested: vascular cell adhesion molecule-1 (VCAM-1), epidermal growth factor (EGF), vascular endothelial growth factor A (VEGF-A), interleukin 6 (IL-6), tumor necrosis factor receptor type 1 (TNF-R1), matrix metalloproteinase-1 (MMP-1), matrix metalloproteinase-3 (MMP-3), human cartilage glycoprotein 39 (YKL-40), leptin, resistin, serum amyloid (SAA), and CRP [41]. A 2019 systematic review and meta-analysis identified eight studies that reported correlations between the MBDA and RA

disease activity measures currently used in clinical trials and clinical practice. There was a modest correlation between MBDA, DAS28-CRP, and DAS28-ESR, with weaker correlations observed with SDAI, Clinical Disease Activity Index (CDAI), and Routine Assessment of Patient Index Data (RAPID3) [42]. However, subsequent post hoc analysis of data from the AMPLE trial (abatacept versus adalimumab for RA) showed disagreement between the MBDA test score and these measures [43-44]. One trial showed that the MBDA test may be useful in deciding whether or not to continue biologic therapy in the setting of clinical remission [45], and post hoc analyses of data have shown that a high baseline MBDA score is a strong independent predictor of radiographic progression at one year [46-49]. Further study is needed in both regards. At this time, the use of this test is not included in the 2015 ACR Guideline for the Treatment of RA [34]. The cost-effectiveness and role of this test in routine clinical practice remain controversial.

Table 1 summarizes the biomarkers discussed thus far.

	Diagnostic Biomarkers (2010 ACR/EULAR Classification Criteria for RA)	Disease Activity Monitoring Biomarkers
RF	x	-
anti-CCP2	x	-
ESR	x	DAS28-ESR
CRP	x	DAS28-CRP, SDAI
MBDA	-	Role unclear

TABLE 1: RA biomarkers used in routine clinical practice

ACR/EULAR, American College of Rheumatology/European League Against Rheumatism; RF, rheumatoid factor; anti-CCP2, anti-cyclic citrullinated peptide2; ESR, erythrocyte sedimentation rate; CRP, C-reactive protein; MBDA, multi-biomarker disease activity test; DAS28, Disease Activity Score 28; SDAI, Simplified Disease Activity Index

Investigational biomarkers in RA

Investigational Biomarkers for Diagnosis

Several biomarkers are currently under study in hopes of improving the accuracy and timeliness of the diagnosis of RA. Approximately 20% to 25% of patients are classified as seronegative RA (negative RF and anti-CCP2 testing). About half of patients are seronegative early in the disease course but eventually become seropositive [14]. Seronegative RA patients experience a delay in diagnosis and delay in initiation of therapy. Hence, they are less likely to attain remission and more likely to suffer joint damage and disability. This suggests a missed “window of opportunity” for intervention (within the first three to six months of illness) [50]. It is unclear whether these patients are truly seronegative, or whether they simply possess RA antibodies that are yet to be identified.

Anti-mutated citrullinated vimentin (anti-MCV), an antibody in the ACPA family, has a similar specificity for RA as anti-CCP2. However, systematic review and meta-analysis of the literature did not reveal superior diagnostic accuracy to anti-CCP2, ultimately limiting the adaptation of anti-MCV testing in routine clinical practice [51-53]. No study has specifically addressed whether adding anti-MCV testing to RF and anti-CCP2 testing would improve overall diagnostic accuracy for RA.

Serum 14-3-3eta, an intracellular chaperonin protein, has been studied as a diagnostic biomarker in RA, but data to date have not been robust enough to defend its routine clinical use. When tested in addition to RF and anti-CCP2, testing may minimally improve rates of diagnosis (from 72% to 78%) or reclassify individuals previously deemed seronegative [54-57]. Further study is needed in this regard.

Antibodies to carbamylated proteins (anti-CarP) have been found in the serum of RA patients. Similar to the other novel biomarkers discussed, studies have not shown increased sensitivity or specificity when compared to the RF and anti-CCP2 testing currently used in clinical practice [58].

Investigational Biomarkers for Disease Activity Monitoring

Given the limitations of ESR and CRP testing as described above, the search for a clinically useful biomarker for disease activity monitoring persists. A biomarker that accurately identifies subclinical disease activity could help guide management decisions and lead to better patient outcomes.

Multiple types of biomarkers are being investigated for the purpose of RA disease activity monitoring: serum

acute phase reactants, genetic factors, and tissue-specific markers from cartilage, bone, and synovium. IL-6, a prominent acute phase reactant in RA, remains under investigation but unfortunately has not been found to correlate with the radiographic progression of the disease [59]. Genetic testing may ultimately play a role in the prognosis and selection of therapy given the well-known association of RA with certain human leukocyte antigen (HLA)-DR alleles. Synovium-specific markers of interest include serum hyaluronan, MMP-1, and MMP-3; these have been shown to correlate with radiographic progression [60-61]. Cartilage and bone-specific markers under investigation include serum cartilage oligomeric matrix protein (COMP) and urine C-terminal crosslinked peptides from type I and type II collagen (CTX-I and CTX-II), among others [62-63]. Serum VEGF, a vascular marker, is elevated in RA patients and correlates with radiographic progression [64]. Synovial fluid biomarkers have also been identified, but the clinical utility of these would be limited given the requirement for arthrocentesis to perform testing. Ultimately, the combined use of multiple biomarkers may prove to be a more effective measure of disease activity. Future studies may follow in this regard.

Table 2 summarizes the biomarkers discussed thus far.

Potential Diagnostic Biomarkers	Potential Disease Activity Monitoring Biomarkers
anti-MCV	IL-6
serum-14-3-3eta anti-CarP	serum VEGF
	serum COMP
	urine CTX-I and CTX-II
	serum hyaluronan
	serum MMP-1 and MMP-3
	synovial fluid biomarkers

TABLE 2: Investigational biomarkers in RA for diagnosis and disease activity monitoring

Anti-MCV, anti-mutated citrullinated vimentin; anti-CarP, anti-carbamylated proteins; IL-6, interleukin-6; VEGF, vascular endothelial growth factor; COMP, cartilage oligomeric matrix protein; CTX-I and CTX-II, C-terminal crosslinked peptides from type I and type II collagen; MMP-1, matrix metalloproteinase-1; MMP-3, matrix metalloproteinase-3

Conclusions

To conclude, the biomarkers currently available for the diagnosis, prognosis, and management of RA have several limitations. RF lacks specificity, as any condition that triggers chronic antigenic stimulation may result in positive RF testing. Anti-CCP2 is more specific, but both tests fail to identify 20%-25% of patients with seronegative RA. Disease activity monitoring remains clinical due to the lack of adequate biomarkers for this purpose. ESR and CRP are nonspecific acute phase reactants that may be elevated for a myriad of reasons, and the role of the commercially available MBDA test remains unclear. Novel RA biomarkers of many types (eg. serum, tissue-specific, genetic factors) are under active investigation for both diagnosis and disease activity monitoring. The rheumatology community eagerly awaits data in this regard.

Many outstanding clinical questions remain that better biomarker identification could help answer. Does seronegative RA truly exist, or have we simply not yet identified the antibodies this subset of patients makes? Can we identify a biomarker that permits earlier RA diagnosis, widening the golden three- to six-month “window of opportunity” to therapeutically intervene? Can a universal biomarker be found that accurately identifies ongoing subclinical disease activity, permitting better titration of RA therapies? Can we identify biomarkers that allow for the personalized selection of RA therapies, permitting more rapid and effective disease control? Future work should pursue answers to these questions. Better biomarkers could lead to earlier diagnosis, treatment, and outcomes. Once a better biomarker is identified, the cost and feasibility of testing will need to be considered in order to ensure clinical utility on a worldwide scale.

Additional Information

Disclosures

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no

other relationships or activities that could appear to have influenced the submitted work.

References

- van der Linden MP, Knevel R, Huizinga TW, van der Helm-van Mil AH: Classification of rheumatoid arthritis: comparison of the 1987 American College of Rheumatology criteria and the 2010 American College of Rheumatology/European League Against Rheumatism criteria. *Arthritis Rheum*. 2011, 63:37-42. [10.1002/art.30100](https://doi.org/10.1002/art.30100)
- Aletaha D, Neogi T, Silman AJ, et al.: 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum*. 2010, 62:2569-81. [10.1002/art.27584](https://doi.org/10.1002/art.27584)
- Westwood OM, Nelson PN, Hay FC: Rheumatoid factors: what's new?. *Rheumatology (Oxford)*. 2006, 45:379-85. [10.1093/rheumatology/kei228](https://doi.org/10.1093/rheumatology/kei228)
- Padyukov L, Silva C, Stolt P, Alfredsson L, Klareskog L: A gene-environment interaction between smoking and shared epitope genes in HLA-DR provides a high risk of seropositive rheumatoid arthritis. *Arthritis Rheum*. 2004, 50:3085-92. [10.1002/art.20553](https://doi.org/10.1002/art.20553)
- Pincus T: Advantages and limitations of quantitative measures to assess rheumatoid arthritis: joint counts, radiographs, laboratory tests, and patient questionnaires. *Bull NYU Hosp Jt Dis*. 2006, 64:32-9.
- Aggarwal A: Role of autoantibody testing. *Best Pract Res Clin Rheumatol*. 2014, 28:907-20. [10.1016/j.berh.2015.04.010](https://doi.org/10.1016/j.berh.2015.04.010)
- Halldórsdóttir HD, Jónsson T, Thorsteinnsson J, Valdimarsson H: A prospective study on the incidence of rheumatoid arthritis among people with persistent increase of rheumatoid factor. *Ann Rheum Dis*. 2000, 59:149-51. [10.1136/ard.59.2.149](https://doi.org/10.1136/ard.59.2.149)
- Nielsen SF, Bojesen SE, Schnohr P, Nordestgaard BG: Elevated rheumatoid factor and long term risk of rheumatoid arthritis: a prospective cohort study. *BMJ*. 2012, 345:e5244. [10.1136/bmj.e5244](https://doi.org/10.1136/bmj.e5244)
- Bruns A, Nicaise-Roland P, Hayem G, et al.: Prospective cohort study of effects of infliximab on rheumatoid factor, anti-cyclic citrullinated peptide antibodies and antinuclear antibodies in patients with long-standing rheumatoid arthritis. *Joint Bone Spine*. 2009, 76:248-53. [10.1016/j.jbspin.2008.09.010](https://doi.org/10.1016/j.jbspin.2008.09.010)
- Barra L, Pope J, Bessette L, Haraoui B, Bykerk V: Lack of seroconversion of rheumatoid factor and anti-cyclic citrullinated peptide in patients with early inflammatory arthritis: a systematic literature review. *Rheumatology (Oxford)*. 2011, 50:311-6. [10.1093/rheumatology/keq190](https://doi.org/10.1093/rheumatology/keq190)
- De Rycke L, Verhelst X, Kruihof E, Van den Bosch F, Hoffman IE, Veys EM, De Keyser F: Rheumatoid factor, but not anti-cyclic citrullinated peptide antibodies, is modulated by infliximab treatment in rheumatoid arthritis. *Ann Rheum Dis*. 2005, 64:299-302. [10.1136/ard.2004.023523](https://doi.org/10.1136/ard.2004.023523)
- Edwards JC, Szczepanski L, Szechinski J, et al.: Efficacy of B-cell-targeted therapy with rituximab in patients with rheumatoid arthritis. *N Engl J Med*. 2004, 350:2572-81. [10.1056/NEJMoa032534](https://doi.org/10.1056/NEJMoa032534)
- Valesini G, Gerardi MC, Iannuccelli C, Pacucci VA, Pendolino M, Shoenfeld Y: Citrullination and autoimmunity. *Autoimmun Rev*. 2015, 14:490-7. [10.1016/j.autrev.2015.01.013](https://doi.org/10.1016/j.autrev.2015.01.013)
- Whiting PF, Smidt N, Sterne JA, et al.: Systematic review: accuracy of anti-citrullinated peptide antibodies for diagnosing rheumatoid arthritis. *Ann Intern Med*. 2010, 152:456-64; W155-66. [10.7326/0003-4819-152-7-201004060-00010](https://doi.org/10.7326/0003-4819-152-7-201004060-00010)
- Gilliam BE, Moore TL: The role of anti-cyclic citrullinated peptide (CCP) antibodies in early detection of rheumatoid arthritis: an overview of the INOVA Diagnostics, Inc. QUANTA Lite CCP assays. *Expert Opin Med Diagn*. 2012, 6:359-69. [10.1517/17530059.2012.694423](https://doi.org/10.1517/17530059.2012.694423)
- Nishimura K, Sugiyama D, Kogata Y, et al.: Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody and rheumatoid factor for rheumatoid arthritis. *Ann Intern Med*. 2007, 146:797-808. [10.7326/0003-4819-146-11-200706050-00008](https://doi.org/10.7326/0003-4819-146-11-200706050-00008)
- Fabien N, Olsson NO, Goetz J, et al.: Prevalence of autoantibodies to cyclic citrullinated peptide in patients with rheumatic diseases other than rheumatoid arthritis: a French multicenter study. *Clin Rev Allergy Immunol*. 2008, 34:40-4. [10.1007/s12016-008-8073-2](https://doi.org/10.1007/s12016-008-8073-2)
- Elkayam O, Segal R, Lidgi M, Caspi D: Positive anti-cyclic citrullinated proteins and rheumatoid factor during active lung tuberculosis. *Ann Rheum Dis*. 2006, 65:1110-2. [10.1136/ard.2005.045229](https://doi.org/10.1136/ard.2005.045229)
- Meyer O, Labarre C, Dougados M, et al.: Anticitrullinated protein/peptide antibody assays in early rheumatoid arthritis for predicting five year radiographic damage. *Ann Rheum Dis*. 2003, 62:120-6. [10.1136/ard.62.2.120](https://doi.org/10.1136/ard.62.2.120)
- Rönnelid J, Wick MC, Lampa J, Lindblad S, Nordmark B, Klareskog L, van Vollenhoven RF: Longitudinal analysis of citrullinated protein/peptide antibodies (anti-CP) during 5 year follow up in early rheumatoid arthritis: anti-CP status predicts worse disease activity and greater radiological progression. *Ann Rheum Dis*. 2005, 64:1744-9. [10.1136/ard.2004.033571](https://doi.org/10.1136/ard.2004.033571)
- Carpenter L, Norton S, Nikiphorou E, et al.: Reductions in radiographic progression in early rheumatoid arthritis over twenty-five years: changing contribution from rheumatoid factor in two multicenter UK inception cohorts. *Arthritis Care Res (Hoboken)*. 2017, 69:1809-17. [10.1002/acr.23217](https://doi.org/10.1002/acr.23217)
- Gardette A, Ottaviani S, Tubach F, et al.: High anti-CCP antibody titres predict good response to rituximab in patients with active rheumatoid arthritis. *Joint Bone Spine*. 2014, 81:416-20. [10.1016/j.jbspin.2014.06.001](https://doi.org/10.1016/j.jbspin.2014.06.001)
- Gottenberg JE, Ravaut P, Cantagrel A, et al.: Positivity for anti-cyclic citrullinated peptide is associated with a better response to abatacept: data from the 'Orencia and Rheumatoid Arthritis' registry. *Ann Rheum Dis*. 2012, 71:1815-9. [10.1136/annrheumdis-2011-201109](https://doi.org/10.1136/annrheumdis-2011-201109)
- Gabay C, Kushner I: Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med*. 1999, 340:448-54. [10.1056/NEJM199902113400607](https://doi.org/10.1056/NEJM199902113400607)
- Black S, Kushner I, Samols D: C-reactive protein*. *J Biol Chem*. 2004, 279:48487-90. [10.1074/jbc.R400025200](https://doi.org/10.1074/jbc.R400025200)
- Kushner I, Rzewnicki D, Samols D: What does minor elevation of C-reactive protein signify?. *Am J Med*. 2006, 119:166.e17-28. [10.1016/j.amjmed.2005.06.057](https://doi.org/10.1016/j.amjmed.2005.06.057)
- Antonelli M, Kushner I: It's time to redefine inflammation. *FASEB J*. 2017, 31:1787-91.

- [10.1096/fj.201601326R](https://doi.org/10.1096/fj.201601326R)
28. Woloshin S, Schwartz LM: Distribution of C-reactive protein values in the United States . *N Engl J Med*. 2005, 352:1611-3. [10.1056/NEJM200504143521525](https://doi.org/10.1056/NEJM200504143521525)
 29. Kushner I, Antonelli MJ: What should we regard as an "elevated" C-reactive protein level? . *Ann Intern Med*. 2015, 163:326. [10.7326/L15-5126](https://doi.org/10.7326/L15-5126)
 30. Emery P, Gabay C, Kraan M, Gomez-Reino J: Evidence-based review of biologic markers as indicators of disease progression and remission in rheumatoid arthritis. *Rheumatol Int*. 2007, 27:793-806. [10.1007/s00296-007-0357-y](https://doi.org/10.1007/s00296-007-0357-y)
 31. Giles JT, Bartlett SJ, Andersen R, Thompson R, Fontaine KR, Bathon JM: Association of body fat with C-reactive protein in rheumatoid arthritis. *Arthritis Rheum*. 2008, 58:2632-41. [10.1002/art.23766](https://doi.org/10.1002/art.23766)
 32. Felson DT, Smolen JS, Wells G, et al.: American College of Rheumatology/European League Against Rheumatism provisional definition of remission in rheumatoid arthritis for clinical trials. *Arthritis Rheum*. 2011, 63:573-86. [10.1002/art.30129](https://doi.org/10.1002/art.30129)
 33. Anderson J, Caplan L, Yazdany J, et al.: Rheumatoid arthritis disease activity measures: American College of Rheumatology recommendations for use in clinical practice. *Arthritis Care Res (Hoboken)*. 2012, 64:640-7. [10.1002/acr.21649](https://doi.org/10.1002/acr.21649)
 34. Singh JA, Saag KG, Bridges SL Jr, et al.: 2015 American College of Rheumatology Guideline for the treatment of rheumatoid arthritis. *Arthritis Care Res (Hoboken)*. 2016, 68:1-25. [10.1002/acr.22783](https://doi.org/10.1002/acr.22783)
 35. Dixey J, Solymosy C, Young A: Is it possible to predict radiological damage in early rheumatoid arthritis (RA)? A report on the occurrence, progression, and prognostic factors of radiological erosions over the first 3 years in 866 patients from the Early RA Study (ERAS). *J Rheumatol Suppl*. 2004, 69:48-54.
 36. Sokka T, Pincus T: Erythrocyte sedimentation rate, C-reactive protein, or rheumatoid factor are normal at presentation in 35%-45% of patients with rheumatoid arthritis seen between 1980 and 2004: analyses from Finland and the United States. *J Rheumatol*. 2009, 36:1387-90. [10.3899/jrheum.080770](https://doi.org/10.3899/jrheum.080770)
 37. Wolfe F, Michaud K: The clinical and research significance of the erythrocyte sedimentation rate . *J Rheumatol*. 1994, 21:1227-37.
 38. Wolfe F, Pincus T: The level of inflammation in rheumatoid arthritis is determined early and remains stable over the longterm course of the illness. *J Rheumatol*. 2001, 28:1817-24.
 39. Kay J, Morgacheva O, Messing SP, et al.: Clinical disease activity and acute phase reactant levels are discordant among patients with active rheumatoid arthritis: acute phase reactant levels contribute separately to predicting outcome at one year. *Arthritis Res Ther*. 2014, 16:R40. [10.1186/ar4469](https://doi.org/10.1186/ar4469)
 40. Amos RS, Constable TJ, Crockson RA, Crockson AP, McConkey B: Rheumatoid arthritis: relation of serum C-reactive protein and erythrocyte sedimentation rates to radiographic changes. *Br Med J*. 1977, 1:195-7. [10.1136/bmj.1.6055.195](https://doi.org/10.1136/bmj.1.6055.195)
 41. Vectra. Accessed: April 14, 2021: <https://vectrascore.com/>.
 42. Johnson TM, Register KA, Schmidt CM, O'Dell JR, Mikuls TR, Michaud K, England BR: Correlation of the multi-biomarker disease activity score with rheumatoid arthritis disease activity measures: a systematic review and meta-analysis. *Arthritis Care Res (Hoboken)*. 2019, 71:1459-72. [10.1002/acr.23785](https://doi.org/10.1002/acr.23785)
 43. Davis JM 3rd: Editorial: the multi-biomarker disease activity test for rheumatoid arthritis: is it a valid measure of disease activity?. *Arthritis Rheumatol*. 2016, 68:2061-6. [10.1002/art.39716](https://doi.org/10.1002/art.39716)
 44. Weinblatt ME, Schiff M, Valente R, et al.: Head-to-head comparison of subcutaneous abatacept versus adalimumab for rheumatoid arthritis: findings of a phase IIb, multinational, prospective, randomized study. *Arthritis Rheum*. 2013, 65:28-3. [10.1002/art.37711](https://doi.org/10.1002/art.37711)
 45. Rech J, Hueber AJ, Finzel S, et al.: Prediction of disease relapses by multibiomarker disease activity and autoantibody status in patients with rheumatoid arthritis on tapering DMARD treatment. *Ann Rheum Dis*. 2016, 75:1637-44. [10.1136/annrheumdis-2015-207900](https://doi.org/10.1136/annrheumdis-2015-207900)
 46. Li W, Sasso EH, van der Helm-van Mil AH, Huizinga TW: Relationship of multi-biomarker disease activity score and other risk factors with radiographic progression in an observational study of patients with rheumatoid arthritis. *Rheumatology (Oxford)*. 2016, 55:357-66. [10.1093/rheumatology/kev341](https://doi.org/10.1093/rheumatology/kev341)
 47. van der Helm-van Mil AH, Knevel R, Cavet G, Huizinga TW, Haney DJ: An evaluation of molecular and clinical remission in rheumatoid arthritis by assessing radiographic progression. *Rheumatology (Oxford)*. 2013, 52:839-46. [10.1093/rheumatology/kes378](https://doi.org/10.1093/rheumatology/kes378)
 48. Markuse IM, Dirven L, van den Broek M, et al.: A multibiomarker disease activity score for rheumatoid arthritis predicts radiographic joint damage in the BeSt study. *J Rheumatol*. 2014, 41:2114-9. [10.3899/jrheum.131412](https://doi.org/10.3899/jrheum.131412)
 49. Hambardzumyan K, Bolce R, Saevarsdottir S, et al.: Pretreatment multi-biomarker disease activity score and radiographic progression in early RA: results from the SWEFOT trial. *Ann Rheum Dis*. 2015, 74:1102-9. [10.1136/annrheumdis-2013-204986](https://doi.org/10.1136/annrheumdis-2013-204986)
 50. Coffey CM, Crowson CS, Myasoedova E, Matteson EL, Davis JM 3rd: Evidence of diagnostic and treatment delay in seronegative rheumatoid arthritis: missing the window of opportunity. *Mayo Clin Proc*. 2019, 94:2241-8. [10.1016/j.mayocp.2019.05.023](https://doi.org/10.1016/j.mayocp.2019.05.023)
 51. Bartoloni E, Alunno A, Bistoni O, et al.: Diagnostic value of anti-mutated citrullinated vimentin in comparison to anti-cyclic citrullinated peptide and anti-viral citrullinated peptide 2 antibodies in rheumatoid arthritis: an Italian multicentric study and review of the literature. *Autoimmun Rev*. 2012, 11:815-20. [10.1016/j.autrev.2012.02.015](https://doi.org/10.1016/j.autrev.2012.02.015)
 52. Luime JJ, Colin EM, Hazes JM, Lubberts E: Does anti-mutated citrullinated vimentin have additional value as a serological marker in the diagnostic and prognostic investigation of patients with rheumatoid arthritis? A systematic review. *Ann Rheum Dis*. 2010, 69:337-44. [10.1136/ard.2008.103283](https://doi.org/10.1136/ard.2008.103283)
 53. Lee YH, Bae SC, Song GG: Diagnostic accuracy of anti-MCV and anti-CCP antibodies in rheumatoid arthritis: A meta-analysis [Article in German]. *Z Rheumatol*. 2015, 74:911-8. [10.1007/s00393-015-1598-x](https://doi.org/10.1007/s00393-015-1598-x)
 54. Maksymowych WP, Naides SJ, Bykerk V, et al.: Serum 14-3-3 η is a novel marker that complements current serological measurements to enhance detection of patients with rheumatoid arthritis. *J Rheumatol*. 2014, 41:2104-13. [10.3899/jrheum.131446](https://doi.org/10.3899/jrheum.131446)
 55. Showman O, Gilburd B, Watad A, et al.: The diagnostic value of 14-3-3 η protein levels in patients with

- rheumatoid arthritis. *Best Pract Res Clin Rheumatol*. 2018, 32:610-7. [10.1016/j.berh.2019.01.010](https://doi.org/10.1016/j.berh.2019.01.010)
56. Guan S-Z, Yang Y-Q, Bai X, et al.: Serum 14-3-3 η could improve the diagnostic rate of rheumatoid arthritis and correlates to disease activity. *Ann Clin Lab Sci*. 2019, 49:57-62.
 57. El-Sherif WT, Nigm DA, Abd-Elsamea MH, Kassem AM: Evaluation of serum protein 14-3-3 η (eta) as a novel biomarker for rheumatoid arthritis. *Egypt J Immunol*. 2019, 26:163-75.
 58. Shi J, van Steenberg HW, van Nies JA, et al.: The specificity of anti-carbamylated protein antibodies for rheumatoid arthritis in a setting of early arthritis. *Arthritis Res Ther*. 2015, 17:339. [10.1186/s13075-015-0860-6](https://doi.org/10.1186/s13075-015-0860-6)
 59. van Leeuwen MA, Westra J, Limburg PC, van Riel PL, van Rijswijk MH: Clinical significance of interleukin-6 measurement in early rheumatoid arthritis: relation with laboratory and clinical variables and radiological progression in a three year prospective study. *Ann Rheum Dis*. 1995, 54:674-7. [10.1136/ard.54.8.674](https://doi.org/10.1136/ard.54.8.674)
 60. Paimela L, Heiskanen A, Kurki P, Helve T, Leirisalo-Repo M: Serum hyaluronate level as a predictor of radiologic progression in early rheumatoid arthritis. *Arthritis Rheum*. 1991, 34:815-21. [10.1002/art.1780340706](https://doi.org/10.1002/art.1780340706)
 61. Green MJ, Gough AK, Devlin J, Smith J, Astin P, Taylor D, Emery P: Serum MMP-3 and MMP-1 and progression of joint damage in early rheumatoid arthritis. *Rheumatology (Oxford)*. 2005, 42:83-8. [10.1093/rheumatology/keg037](https://doi.org/10.1093/rheumatology/keg037)
 62. Garnero P, Landewé R, Boers M, et al.: Association of baseline levels of markers of bone and cartilage degradation with long-term progression of joint damage in patients with early rheumatoid arthritis. The COBRA study. *Arthritis Rheum*. 2002, 46:2847-56. [10.1002/art.10616](https://doi.org/10.1002/art.10616)
 63. Forslind K, Eberhardt K, Jonsson A, Saxne T: Increased serum concentrations of cartilage oligomeric matrix protein. A prognostic marker in early rheumatoid arthritis. *Br J Rheumatol*. 1992, 31:593-8. [10.1093/rheumatology/31.9.593](https://doi.org/10.1093/rheumatology/31.9.593)
 64. Ballara S, Taylor PC, Reusch P, Marmé D, Feldmann M, Maini RN, Paleolog EM: Raised serum vascular endothelial growth factor levels are associated with destructive change in inflammatory arthritis. *Arthritis Rheum*. 2001, 44:2055-64. [10.1002/1529-0131\(200109\)44:9<2055::AID-ART355>3.0.CO;2-2](https://doi.org/10.1002/1529-0131(200109)44:9<2055::AID-ART355>3.0.CO;2-2)