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Preliminary Analysis of Liquid Biopsy after Hepatectomy for Colorectal Liver Metastases

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Abstract

Background: Liquid biopsies are increasingly tested in patients with colorectal cancer to assess tumor burden, response to therapy, and prognosis. The significance of liquid biopsy results after resection of colorectal liver metastases (CLM) is not well-defined.

Study Design: Sixty-three patients undergoing CLM resection between 2016–2018 had plasma drawn postoperatively for liquid biopsy evaluation. Next-generation sequencing analysis was performed to detect somatic mutations in 70 genes.

Results: Liquid biopsy after CLM resection was positive in 42 of 63 patients (67%). Eleven patients (18%) had 1 gene mutation, 14 (22%) had 2–3 mutations, and 17 (27%) had 4 mutations. The most common mutation was *APC*, detected in 32 (76%) patients, followed by *TP53* (74%) and *KRAS* (38%). Two-year OS from date of liver resection was significantly worse among patients with a positive liquid biopsy (70% versus 100%, $p = 0.005$), particularly for those with 4 gene mutations detected, whose 2-year OS was 41%. Sixteen of the 63 patients underwent serial liquid biopsies, resulting in 100 liquid biopsy with matched serum CEA and computed tomography (CT) scan results. Metastases were identified in 74 CT scans, which correlated with positive liquid biopsy in 77% of samples ($p < 0.001$) and CEA > 3 ng/ml in 45% of samples ($p < 0.22$).

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Conclusion: Liquid biopsy results provide complementary information to serum CEA and CT imaging about disease burden and prognosis. A positive liquid biopsy after CLM resection is associated with worse OS, particularly when multiple gene mutations are detected.

PRECIS

After resection of colorectal liver metastases, detection of a somatic mutation in a 70-gene liquid biopsy panel correlates with worse overall survival, particularly when multiple gene mutations are identified. A positive liquid biopsy correlates with radiologic evidence of disease recurrence.

Keywords

Liquid biopsy; liver metastases; colorectal cancer; cell-free DNA; hepatectomy; circulating tumor DNA

INTRODUCTION

Colorectal cancer is the second leading cause of cancer-related mortality in the United States, and most patients who die of colorectal cancer will have liver metastases.(1) Surgical resection of colorectal liver metastases (CLM) remains the gold standard treatment and is associated with a 5-year overall survival (OS) rate of 58%.(2) However, up to 70% of patients will suffer disease recurrence after hepatic resection.(3) Strategies to reduce recurrence rates include perioperative chemotherapy, targeted agents, and biomarkers to stratify prognosis and predict response to therapy.(4-6) Specifically, microsatellite instability and *RAS* mutation status predict response to immunotherapy and anti-epidermal growth factor receptor (EGFR) agents, respectively.(1)

Currently, biomarker detection in colorectal cancer depends upon genotyping of tissue biopsy or resection specimens. However, tissue biopsy requires a percutaneous or endoscopic procedure and only provides a single, static snapshot of a small fragment of tumor. Liquid biopsy is emerging as a promising, non-invasive alternative to tissue biopsy. (7) DNA present in the non-cellular portion of blood, or cell-free DNA (cfDNA), originates from normal tissue or tumor cells. Circulating tumor DNA (ctDNA) is cfDNA secreted into the bloodstream by tumor cells. Analysis of ctDNA, also termed liquid biopsy, has the potential to detect minimal residual disease, genotype tumors to direct targeted therapy, and track tumor evolution in real-time. In addition, liquid biopsy may provide a more comprehensive molecular profile of cancers than a single tissue biopsy.

In colorectal cancer, ctDNA detection rates have been shown to correlate with stage of disease, identified in 40% and > 90% of patients with stage I and IV disease, respectively.(8) Among patients undergoing CLM resection, preoperative detection of specific somatic gene mutations in peripheral blood is associated with worse disease-specific survival.(9) The purpose of this study was to evaluate a 70-gene liquid biopsy panel and its impact on OS after CLM resection, and correlations with postoperative computed tomography (CT) imaging and serum carcinoembryonic antigen (CEA) levels.

METHODS

Study population and follow-up

A prospectively maintained database was queried to identify 63 consecutive patients who underwent CLM resection at The University of Texas MD Anderson Cancer Center between January 2016 and November 2018, and had plasma drawn postoperatively for liquid biopsy analysis (Fig. 1).

Major hepatectomy was defined as resection of ≥ 3 contiguous Couinaud segments.(10) Synchronous liver metastases were defined as metastases diagnosed within 6 months of primary tumor diagnosis. R1 resection margin was defined as tumor cells at the line of transection. Patients undergoing 1st-stage hepatectomy without completing the 2nd-stage were classified as having R2 resections. Number and size of liver metastases were recorded from surgical pathology reports or CT imaging.

Postoperatively, patients were followed with history and physical examination, serum CEA evaluation, and CT scans of the chest, abdomen, and pelvis every 3 months for the first 2 years after CLM resection. Sixteen patients underwent serial liquid biopsies, resulting in 100 samples with matched CEA and CT results. The study was approved by the Institutional Review Board.

Genetic analyses

A next-generation sequencing analysis to detect somatic mutations in 70 cancer-related genes was performed on ctDNA isolated from plasma in a Clinical Laboratory Improvement Amendments (CLIA)-certified molecular diagnostics laboratory, based on Guardant360[®]CDx technology (Guardant Health, Inc., Redwood City, CA).(11) The genes and exons tested are listed in eTable 1. The lower limit of detection was 0.3% mutation allelic frequency for 30 ng cfDNA and 1% for 5 ng cfDNA. A positive liquid biopsy was defined as one or more gene mutations in plasma.

Next-generation sequencing was performed on tissue from primary tumors and/or liver metastases to detect somatic mutations in coding sequences of 128 cancer-related genes from DNA extracted from formalin-fixed paraffin-embedded tumor samples, as previously described. (12) Fourteen patients underwent analysis of 50 genes only, and 5 patients did not undergo tissue multigene panel testing.

Statistical analyses

Group comparisons were performed using chi-square tests for categorical variables and Mann–Whitney U test for continuous variables. OS was estimated using the Kaplan–Meier method and compared with the log-rank test. Factors significant on univariable analysis were entered into a multivariable Cox analysis of OS. All p values were 2-sided, and $p < 0.05$ was considered statistically significant. All analyses were performed with SPSS Statistics 23.0 (IBM Corp., Chicago, IL).

RESULTS

Clinicopathologic characteristics of the 63 patients are listed in Table 1. Median follow-up was 30 months (range, 9–53 months). Median time interval between hepatectomy and postoperative blood draw for liquid biopsy analysis was 13 months (range, 1–45 months) (Fig. 2). Most liquid biopsies were ordered by medical oncologists to detect minimal residual disease (21 patients), to guide systemic therapy (n = 19), including detection of resistance mutations to anti-EGFR therapy; for clinical trial screening (n = 13), and as baseline evaluation before surgery or ablation for lung or recurrent liver metastases (n = 10).

Among the 21 patients with extrahepatic metastases, 12 patients underwent synchronous hepatectomy and resection of disease in the portal or retroperitoneal lymph nodes (n = 6), lung (n = 2), peritoneum (n = 2), ovary (n = 1), or adrenal (n = 1). Unresected extrahepatic metastases included low-volume disease responding to chemotherapy in the lungs (n = 3) or retroperitoneal lymph nodes (n = 2); bone metastasis treated with radiation (n = 1), and 3 patients who underwent 1st-stage hepatectomy only.

First postoperative liquid biopsy and correlation with tumor tissue

The number of gene mutations detected in each plasma sample was no mutation in 21 patients (33%), one mutation in 11 patients (18%), 2–3 mutations in 14 patients (22%), and 4 mutations in 17 patients (27%). Among the 42 patients with positive liquid biopsy, the most common gene mutation was *APC*, detected in 32 patients (76%), followed by *TP53* (74%) and *KRAS* (38%).

Multigene panel testing on the primary tumor and/or liver metastases was performed in 58 of the 63 patients. Figure 3 shows the distribution of somatic mutations identified in liquid biopsy only, tissue only, and both liquid biopsy and tissue. The most commonly mutated genes, *TP53*, *APC*, and *KRAS*, were identified in both liquid biopsy and tissue among 38%, 25%, and 16% of patients, respectively. A *KRAS* mutation was detected in tissue but not liquid biopsy in 2 patients. Conversely, *KRAS* or *NRAS* mutation was identified in liquid biopsy but not tissue in 8 patients. In these 8 patients, ctDNA analysis was performed > 1 year after tissue genotyping, and anti-EGFR therapy was administered in the interim. Four patients had a single gene mutation in liquid biopsy samples not identified in tissue; the 4 single gene mutations were *RET*, *MTOR*, *EGFR*, and *GNAS*.

Association between liquid biopsy and survival

Two-year OS from date of liver resection was significantly lower among patients with a positive liquid biopsy (70% versus 100%, $p = 0.005$) (Fig. 4A). When further stratified by number of gene mutations, the presence of 4 mutations was associated with significantly lower 2-year OS of 41% ($p < 0.001$) (Fig. 4B). Results of univariable and multivariable analyses of the predictors of survival are shown in Table 2. Factors significantly associated with worse survival were resection margin and 4 gene mutations on liquid biopsy. Size of liver metastasis was marginally significant ($p = 0.06$). On multivariable analysis, resection margin and 4 gene mutations on liquid biopsy remained independent predictors of OS.

Associations between first postoperative liquid biopsy and clinicopathologic factors

A positive postoperative liquid biopsy was not significantly associated with the following clinicopathologic factors: node-positive primary, number and size of liver metastases, CEA > 200 ng/ml, disease-free interval < 12 months between diagnosis of primary tumor and CLM, and extrahepatic disease. All 8 patients who underwent R2 resection had a positive liquid biopsy, including 6 with 4 gene mutations. Resection margin was significantly associated with the presence of 4 gene mutations on liquid biopsy, detected in 21% (8/39), 19% (3/16), and 75% (6/8) of patients with R0, R1, and R2 margins, respectively ($p = .005$).

First postoperative liquid biopsy and recurrence after CLM resection

Liquid biopsy results were not significantly associated with disease recurrence in the liver, lungs, or peritoneum. Among 11 patients with recurrence in abdominal and/or retroperitoneal lymph nodes, 8 patients had 4 gene mutations on liquid biopsy, compared with 3 patients with 1–3 gene mutations ($p = .041$).

Correlations between liquid biopsy, CEA, and CT imaging

Sixteen of the 63 patients underwent serial liquid biopsies, resulting in an additional 37 liquid biopsy samples. For all 63 patients, there were 100 liquid biopsy with matched serum CEA and CT results. Metastatic disease was identified in 74 of the 100 CT scans, either at the initial postoperative restaging scan or subsequent follow-up scans. CT evidence of disease correlated with positive liquid biopsy in 77% of samples ($p < 0.001$) and CEA > 3 ng/ml in 45% of samples ($p = 0.22$).

DISCUSSION

To our knowledge, the current study is the first to analyze the impact of a 70-gene liquid biopsy panel on patient survival after CLM resection and associations with CT imaging and serum CEA. We report that patients with a positive liquid biopsy after CLM resection had significantly worse 2-year OS than those with a negative liquid biopsy. The number of gene mutations detected in liquid biopsy samples correlated with survival, with lower OS observed with 4 mutations, compared with a single gene mutation. A positive liquid biopsy, but not elevated serum CEA, was significantly associated with CT evidence of metastatic disease.

Our study results showing worse survival with positive liquid biopsy after CLM resection are consistent with data from non-metastatic patients undergoing resection of stages I-III colorectal cancer, in whom ctDNA-positivity on postoperative day 30 conferred a 7-fold higher rate of disease relapse.(8) After CLM resection, Scholer and colleagues observed a 100% relapse rate for patients with positive liquid biopsy 3 months postoperatively, compared with 50% relapse with negative liquid biopsy.(13) In the current study, resection margin after CLM resection was significantly associated with 4 gene mutations on liquid biopsy, observed in 75% of patients with R2 margin, compared with 21% of patients with R0 margin. Taken together, these data suggest that after resection of localized and metastatic colorectal cancer, a positive liquid biopsy correlates with the presence of residual disease.

We observed that the presence of 4 gene mutations, occurring in 40% of patients with a positive liquid biopsy, was associated with significantly worse OS compared with a solitary gene mutation. The lower survival with multiple mutations in liquid biopsy samples is congruent with emerging data on the importance of co-mutations in tissue samples after CLM resection, particularly the co-occurrence of *RAS*, *TP53*, and *SMAD4* mutations.(5, 14) Similarly, in a study of 298 patients with metastatic cancer, including liver, pancreas, and colorectal cancer, the presence of more than one nonsynonymous mutation in ctDNA correlated with significantly worse OS.(11) The inferior prognosis with multiple mutations is attributed to increased tumor heterogeneity and the emergence of subclones resistant to systemic therapy.

Our data demonstrate a significant correlation between CT evidence of metastatic disease and a positive liquid biopsy but not serum CEA levels. In a study of patients undergoing surgery for stages I–III colorectal cancer, serial liquid biopsies revealed disease recurrence up to 16.5 months earlier than standard-of-care radiologic imaging.(8) Given the small number of patients who underwent serial liquid biopsy in our study, the ability of liquid biopsy to detect early recurrence was not analyzed.

Until recently, the use of liquid biopsy was limited by the need to sequence known individual gene mutations present in tissue. In colorectal cancer, several genes have been identified that are recurrently somatically mutated, including *TP53*, *RAS*, and *APC*. In this analysis, we used a ctDNA detection technology that analyzes mutations in 70 genes. We observed 83% concordance in *RAS* mutation status between liquid biopsy and tissue. Prior reports have shown concordance rates in *RAS* mutation status between liquid biopsy and tissue ranging between 71% to 100%.(15, 16) In this study, 8 patients had *RAS* mutations detected in ctDNA but not tumor tissue. All 8 patients underwent liquid biopsy testing > 1 year after tissue genotyping and received anti-EGFR therapy in the interim. The emergence of *RAS* mutant subclones or new *RAS* mutations arising from ongoing mutagenesis are increasingly recognized as important causes of acquired resistance to anti-EGFR therapy.(17) Other causes of discordance between mutations detected in liquid biopsy versus tissue include tumor location, sample volume, amount of ctDNA, and assay sensitivity.(7) In our study, discordant results between liquid biopsy and tissue are partly attributable to varying levels of tumor burden among patients who underwent ctDNA analysis at different time intervals after hepatectomy.

In the current report, 4 patients had liquid biopsy samples with a single gene mutation that was not identified in tumor tissue. False-positive liquid biopsy results can result from a phenomenon known as clonal hematopoiesis of indeterminate potential (CHIP), which results in somatic mutations released by hematopoietic stem cells related to aging, and not colorectal cancer.(7) Thus, liquid biopsy results with a single gene mutation not identified in tissue should be interpreted with caution.

According to our study results, liquid biopsies are currently used to detect minimal residual disease, screen for clinical trials, and guide systemic therapy, including detection of *RAS* mutant clones driving acquired resistance to anti-EGFR therapy. Further research is needed to standardize ctDNA assays across institutions and to validate interpretation of results,

particularly false-positive mutations. Future uses of ctDNA include counseling patients on adjuvant therapy after CLM resection and determining the extent and frequency of postoperative surveillance. In patients with negative radiologic findings, a positive liquid biopsy may signal the need for additional imaging or closer follow-up.

Limitations include the retrospective analysis of 63 patients who underwent postoperative ctDNA analysis, representing a small subset (17%) of the 375 patients undergoing CLM resection during the study period. Therefore, the study population was enriched for patients with suspected or radiologically-confirmed disease recurrence, and the time interval between CLM resection and liquid biopsy analysis varied widely, between 1–45 months. Prospective studies with specified time-points for blood collection are needed to assess ctDNA kinetics after CLM resection. Another limitation is analysis restricted to postoperative ctDNA detection. A prior study from our institution evaluated a 23-gene liquid biopsy panel among 54 patients treated with chemotherapy before CLM resection.(18) After neoadjuvant chemotherapy, the ctDNA mutation detection rate was 80%. In addition, a larger sample size is needed to determine associations between postoperative ctDNA and patterns of disease recurrence. Finally, we did not stratify liquid biopsies by variant allele frequency, which has been shown to correlate with tumor burden and survival.(11)

CONCLUSIONS

In this cohort of patients undergoing CLM resection, positive postoperative liquid biopsy was associated with significantly worse overall survival, particularly when 4 gene mutations were detected. A positive liquid biopsy correlated with CT evidence of disease recurrence. These data support continued investigation of liquid biopsy in CLM for non-invasive molecular profiling, detection of minimal residual disease, and prognostication.

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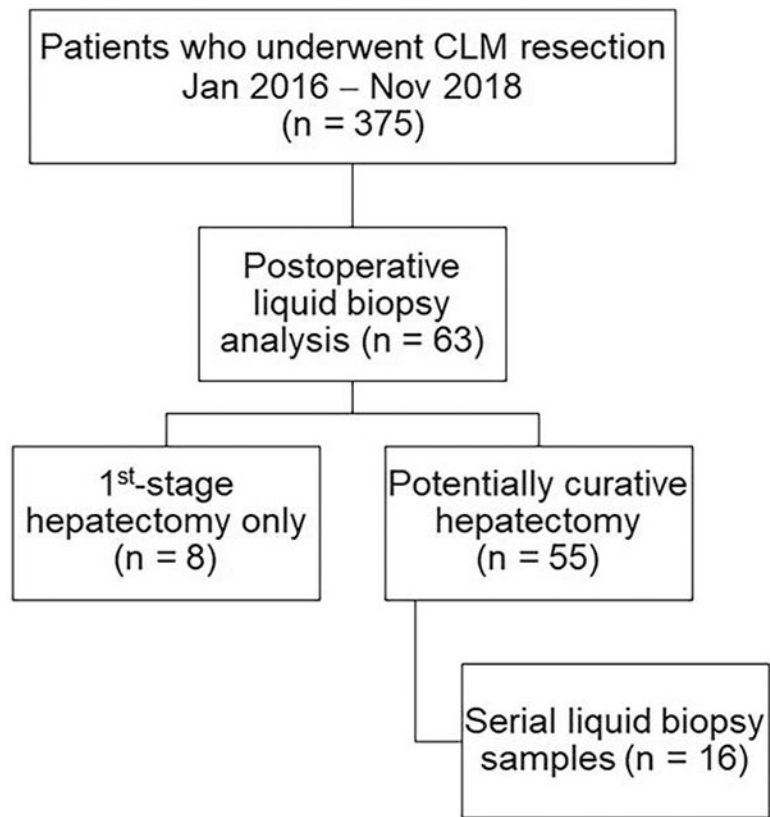


Figure 1. Selection of patients undergoing resection of colorectal liver metastases (CLM) and postoperative liquid biopsy analysis.

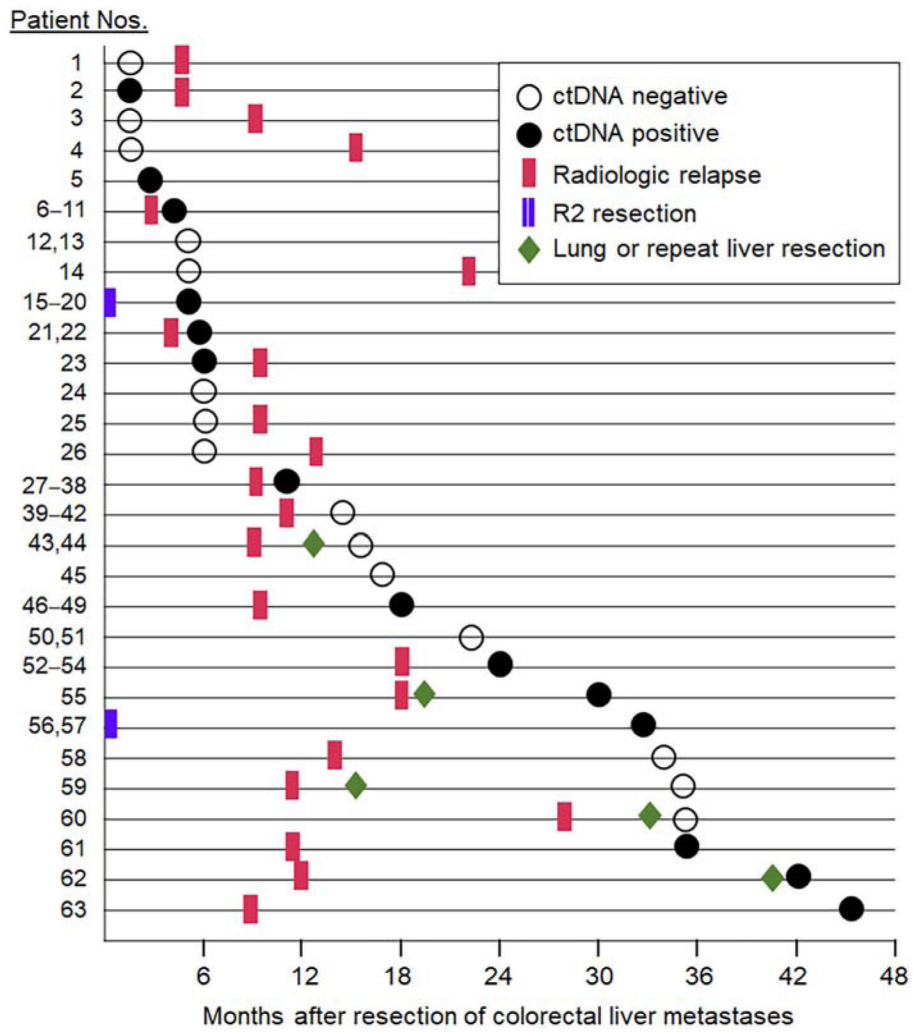


Figure 2. Timeline of first postoperative liquid biopsy and radiologic relapse after resection of colorectal liver metastases.

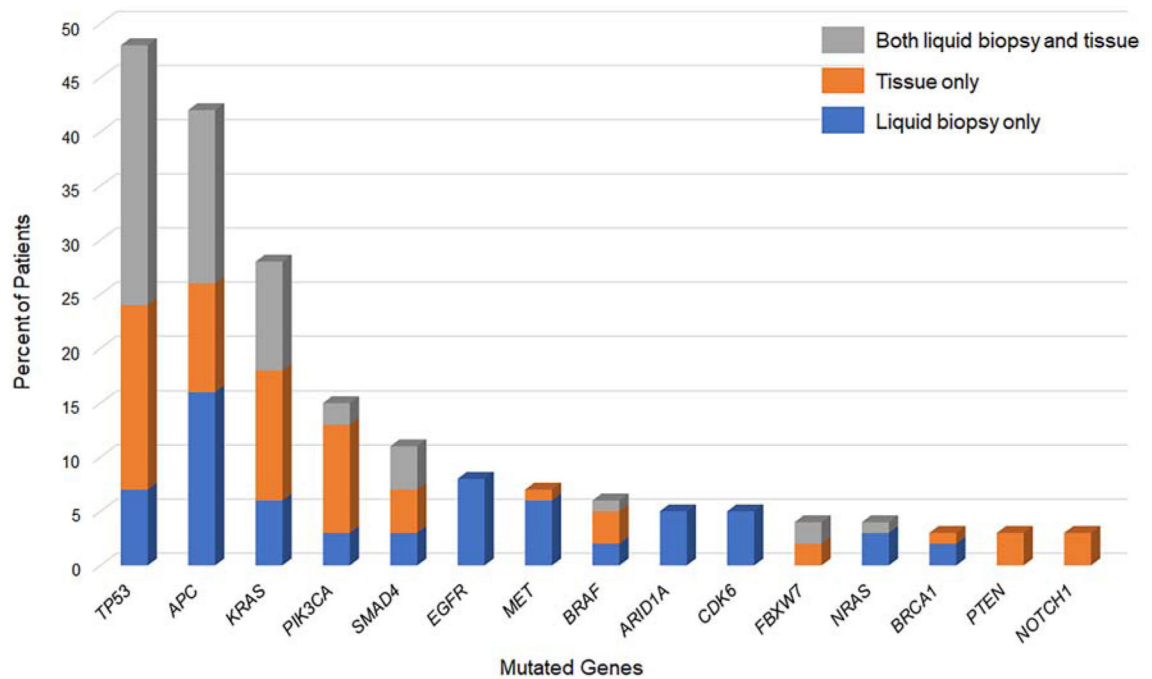


Figure 3. Prevalence of gene mutations in liquid biopsy and tumor tissue among 63 patients undergoing resection of colorectal liver metastases. Rare gene mutations, identified in < 5% of patients, not shown.

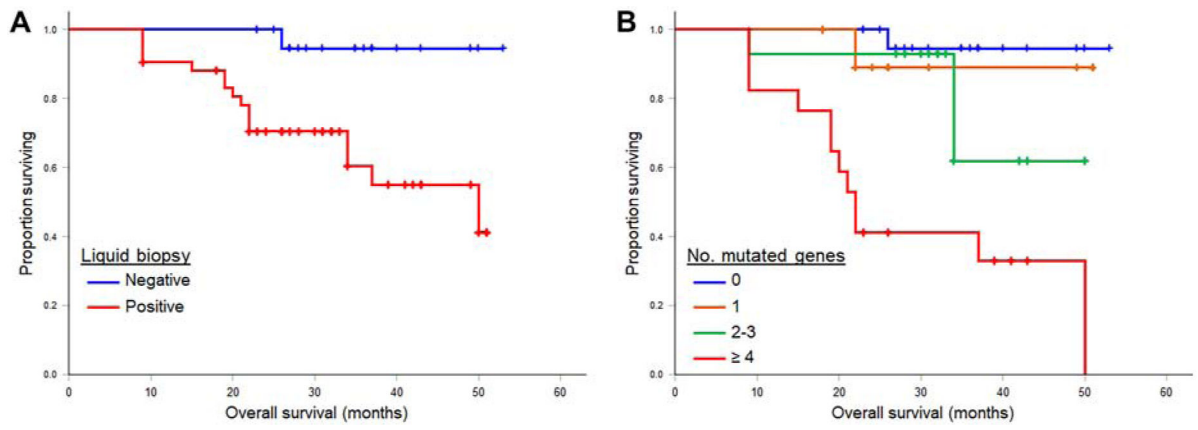


Figure 4. Overall survival of 63 patient after resection of colorectal liver metastases, by (A) postoperative liquid biopsy status and (B) number of mutated genes in liquid biopsy samples.

Table 1.

Patient and Tumor Characteristics (n = 63)

Characteristic	N	%
Age, y, median (range)	55 (30-82)	
Sex, n (%)		
Male	32 (51)	
Female	31(49)	
Node-positive primary tumor, n (%)	54 (86)	
Synchronous diagnosis of liver metastases, n (%)	46 (73)	
Chemotherapy before liver resection, n (%)	55 (87)	
Chemotherapy after liver resection, n (%)	37 (59)	
Preoperative CEA, ng/mL, median (range)	4.1 (0-1639)	
Extrahepatic metastases, n (%)	21 (33)	
No. of liver metastases, median (range)	3 (1-13)	
Size of largest liver metastasis, cm, median (range)	2.5 (0.7-8.2)	
Surgical resection margin, n (%)		
R0	39 (62)	
R1	16 (25)	
R2 (1 st stage hepatectomy only)	8 (13)	
Major hepatectomy, n (%)	23 (37)	
Completed two-stage hepatectomy, n (%)	6 (10)	
Disease recurrence, n (%)	56 (89)	

CEA, carcinoembryonic antigen

Table 2.

Multivariable Overall Survival Analysis Of 63 Patients Undergoing Resection of Colorectal Liver Metastases

Variable	2-year overall survival, %	Univariable p value	Multivariable p value	Hazard ratio (95% CI)
Age	–	0.48		
Sex		0.88		
Male	77			
Female	84			
Primary tumor lymph node status		0.82		
Positive	81			
Negative	75			
Synchronous CLM		0.74		
Yes	80			
No	82			
Extrahepatic metastases		0.57		
Yes	83			
No	77			
Preoperative CEA	–	0.25		
Surgical margin		< 0.001		
R0	90		Ref	
R1	87		0.80	
R2	25		0.006	4.95 (1.58-15.52)
Size of largest liver metastasis	–	0.06	0.96	
Number of liver metastases		0.87		
Solitary	83			
Multiple	80			
Major hepatectomy		0.61		
Yes	88			
No	78			
4 gene mutations on liquid biopsy		< 0.001	0.001	6.60 (2.07-21.06)
Yes	41			
No	96			

CEA, carcinoembryonic antigen; CLM, colorectal liver metastasis; Ref, reference

eTable 1.

Genes and Exons Tested

Gene	Exons tested
<i>AKT1</i>	3, 6
<i>ALK</i>	18-29
<i>APC</i>	2-16
<i>AR</i>	ALL
<i>ARAF</i>	7, 14
<i>ARID1A</i>	1-3, 6-8, 11-20
<i>ATM</i>	8, 55, 63
<i>BRAF</i>	1-15, 17-21
<i>BRCA1</i>	2-23
<i>BRCA2</i>	2-11, 13-27
<i>CCND1</i>	1-2, 4-5
<i>CCND2</i>	1-2, 5
<i>CCNE1</i>	4-6, 8-12
<i>CDK4</i>	2-8
<i>CDK6</i>	2-8
<i>CDKN2A</i>	ALL
<i>CTNNB1</i>	3
<i>DDR2</i>	14-17
<i>EGFR</i>	ALL
<i>ERBB2</i>	ALL
<i>ESR1</i>	5, 6-8
<i>EZH2</i>	16
<i>FBXW7</i>	9-10, 12
<i>FGFR1</i>	2-4, 6-8, 10-13, 15, 17-18
<i>FGFR2</i>	2-13, 15-18
<i>FGFR3</i>	2-18
<i>GNA11</i>	5
<i>GNAQ</i>	5
<i>GNAS</i>	8, 9
<i>GNF1A</i>	3, 4
<i>HRAS</i>	2-5
<i>IDH1</i>	4
<i>IDH2</i>	4
<i>JAK2</i>	14
<i>JAK3</i>	13
<i>KIT</i>	ALL
<i>KRAS</i>	2-5

Gene	Exons tested
<i>MAP2K1</i>	2, 3
<i>MAP2K2</i>	2, 3
<i>MAPK1</i>	ALL
<i>MAPK3</i>	ALL
<i>MET</i>	2-9, 10-21
<i>MLH1</i>	12
<i>MPL</i>	10
<i>MTOR</i>	2-30
<i>MYC</i>	ALL
<i>NFI</i>	2-6, 8-14, 16-18, 20-21, 24-28, 30, 32-50, 52-55, 57-58
<i>NFE2L2</i>	2
<i>NOTCH1</i>	4, 8, 26, 34
<i>NOTCH2</i>	1, 34
<i>NPM1</i>	11
<i>NRAS</i>	2-5
<i>NTRK1</i>	8-10, 12, 14-15
<i>NTRK3</i>	16-17
<i>PDGFRA</i>	3-7, 9-12, 14-15, 18-20, 22-23
<i>PIK3CA</i>	2-21
<i>PTEN</i>	1-2, 4-9
<i>PTPN11</i>	3, 13
<i>RAD51</i>	2-3, 5-10
<i>RAF1</i>	2-3, 5, 7-8, 10, 15-17
<i>RB1</i>	1-4, 7-8, 10-11, 13, 16-23, 25-27
<i>RET</i>	9-12, 14-16
<i>ROS1</i>	31-32, 34-36, 38
<i>SMAD4</i>	3, 5-6, 8-12
<i>SMO</i>	5, 9
<i>STK11</i>	ALL
<i>TERT</i>	1, 5
<i>TP53</i>	2-11
<i>TSC1</i>	15, 23
<i>VHL</i>	ALL