

REVIEW ARTICLE OPEN

The role of integrins in inflammation and angiogenesis

Olachi J. Mezu-Ndubuisi¹ and Akhil Maheshwari²

Integrins are heterodimeric transmembrane cell adhesion molecules made up of alpha (α) and beta (β) subunits arranged in numerous dimeric pairings. These complexes have varying affinities to extracellular ligands. Integrins regulate cellular growth, proliferation, migration, signaling, and cytokine activation and release and thereby play important roles in cell proliferation and migration, apoptosis, tissue repair, as well as in all processes critical to inflammation, infection, and angiogenesis. This review presents current evidence from human and animal studies on integrin structure and molecular signaling, with particular emphasis on signal transduction in infants. We have included evidence from our own laboratory studies and from an extensive literature search in databases PubMed, EMBASE, Scopus, and the electronic archives of abstracts presented at the annual meetings of the Pediatric Academic Societies. To avoid bias in identification of existing studies, key words were short-listed prior to the actual search both from anecdotal experience and from PubMed's Medical Subject Heading (MeSH) thesaurus.

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IMPACT:

- Integrins are a family of ubiquitous αβ heterodimeric receptors that interact with numerous ligands in physiology and disease. Integrins play a key role in cell proliferation, tissue repair, inflammation, infection, and angiogenesis.
- This review summarizes current evidence from human and animal studies on integrin structure and molecular signaling and promising role in diseases of inflammation, infection, and angiogenesis in infants.
- This review shows that integrin receptors and ligands are novel therapeutic targets of clinical interest and hold promise as novel therapeutic targets in the management of several neonatal diseases.

INTRODUCTION

Integrins are a family of ubiquitous $\alpha\beta$ heterodimeric receptors that exist in multiple conformations and interact with a diverse group of ligands. These molecules mediate interactions between cells and of these cells with the extracellular matrix (ECM) and thereby serve a critical role in signaling and homeostasis. By facilitating dynamic linkages between the intracellular actin cytoskeleton and the ECM, integrins also transduce both external and internal mechanochemical cues and bi-directional signaling across the plasma membrane. ^{1,2} Integrins are involved in a diverse range of body processes, including cellular survival, inflammation, immunity, infection, thrombosis, angiogenesis, and malignancy. In this review, we highlight the structure and function of integrins; the mechanisms involved in integrin activation and signaling; their role in inflammation, infection, and angiogenesis; and discuss current advances in integrin-targeted therapies. Understanding the factors that regulate integrin structure, function, and signaling would enable us to identify new therapeutic targets.

STRUCTURE OF INTEGRINS

In mammals, the family of integrins is comprised of 24 $\alpha\beta$ pairs of heterodimeric transmembrane adhesion receptors and cell-surface proteins. These pairings are known to involve 18 α and 8 β subunits (Fig. 1),³ and their non-covalent associations involve an α and another β subunit (Fig. 2).⁴ The $\alpha\beta$ pairings of integrin

subunits dictate the specificity of the integrin to a particular ligand, modulate formation of intracellular adhesion complexes, and regulate downstream signaling. Six α (α_{1-6}) and seven β (β_{1-7}) subunits are known to form several unique $\alpha\beta$ subunit associations (Fig. 1). Interestingly, the earliest discovered integrins, lymphocyte function-associated antigen 1 (integrin $\alpha_L\beta_2$) and macrophage antigen 1 (integrin $\alpha_M\beta_2$), derive their specificity from specific α subunits, but these share the same β subunit.

Integrin a subunit family

The integrin α subunits carry a 200 amino acid "inserted" domain, the I-domain (α I). When present on an integrin, the α I domain is an exclusive ligand-binding site. α I integrins have 13 extracellular domains in 2 subunits, which interact with a variety of ligands. The I-domains are seen in 6 out of the 15 integrin α subunits.

Integrin **B** subunit family

In humans, integrin β subunits have a cytoplasmic tail that have <75 amino acids in length, except the β_4 tail that is about 1000 amino acids long (includes four fibronectin type III repeats). The integrin β tails have one or two NPxY/F motifs (x refers to any amino acid) that recognize protein modules, phosphotyrosine-binding domains, that are involved in several signaling and cytoskeletal proteins at the cytoplasmic face of the plasma membrane through phosphorylation of the tyrosine (Y) in the NPxY/F motif. The integrin β subunit family includes β_{1-7} , which

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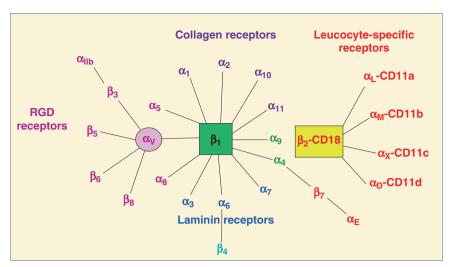


Fig. 1 Classification of integrin family. Integrin heterodimers consists of numerous combinations of α and β subunits. With respect to ligand specificity, integrins are generally classified as collagen-binding integrins ($\alpha_1\beta_1$, $\alpha_2\beta_1$, $\alpha_{10}\beta_1$, and $\alpha_{11}\beta_1$), RGD-recognizing integrins ($\alpha_5\beta_1$, $\alpha_V\beta_1$, $\alpha_V\beta_3$, $\alpha_V\beta_5$, $\alpha_V\beta_6$, $\alpha_V\beta_6$, $\alpha_V\beta_8$, and $\alpha_{IIb}\beta_3$), laminin-binding integrins ($\alpha_3\beta_1$, $\alpha_6\beta_1$, $\alpha_7\beta_1$, and $\alpha_6\beta_4$), and leukocyte integrins ($\alpha_L\beta_2$, $\alpha_M\beta_2$, $\alpha_X\beta_2$, and $\alpha_D\beta_2$). The β_2 integrin subunit (CD18) can pair with one of the four α subunits (α_L -CD11a, α_M -CD11b, α_X -CD11c, and α_D -CD11d), forming leukocyte function-associated antigen-1, Mac1/CR3 (macrophage-1 antigen, complement receptor 3), 150.95/CR4 (complement receptor 4), and CD18/CD11d, respectively. CD11a/CD18 is expressed mainly on all leukocytes, while CD11b/CD18, CD11c/CD18, and CD11d/CD18 are expressed on myeloid cells. 106,107 The $\alpha_M\beta_2$ integrin (also known as CR3, CD11b/CD18, or Mac-1) is found on phagocytic cells and implicated in the adhesion of leucocytes to endothelium and opsonization of microbes. Ligands for CR3 include the complement component iC3b, the intercellular adhesion molecule (1CAM-1), and coagulation factors like fibrinogen and factor X.

bind the a subunits in different combinations. The most frequently seen β subunit integrin heterodimers is β_1 . Although β_2 integrins show functional overlap, the corresponding α subunit defines its individual functional properties.⁶ The β_2 /CD18 chain has also received attention because of its involvement in several inflammatory receptors such as CD11a/CD18, $\alpha_L\beta_2$, lymphocyte functionassociated antigen-1 (LFA-1); CD11b/CD18, $\alpha_M\beta_2$, Mac-1, complement receptor 3 (CR3); CD11c/CD18 ($\alpha_X\beta_2$, p150.95, CR4); and CD11d/CD18, $\alpha_D\beta_2$; Fig. 1). In these β_2 integrins, the α subunits bind specific ligands such as the intercellular adhesion molecules (ICAMs). The non-I-domain α subunits in other integrins, such as the laminin-binding α_3 , α_6 , and α_7 , and others that recognize the arginine (R), glycine (G), aspartic acid (D) (RGD) motif (α_V , α_8 , α_5 , and α_{IIb}), are also closely related to each other. The α subunit of each integrin is the primary determinant of its extracellular ligand specificity. The β chain binds acidic residues in ICAMs and in cytoplasmic adapters such as paxillin, talins, and kindlins to facilitate cellular adhesions with the ECM. Integrins interact with the actin cytoskeleton through the talin- and kindlin-binding motifs present in the cytoplasmic domains of their β subunits.⁸

Characteristics of specific integrin heterodimers

Integrin aß heterodimers are divided into four classes (leukocyte, collagen-binding, Arg-Gly-Asp (RGD)-binding, and laminin-binding integrins; Fig. 1) based on evolutionary associations, ligand specificity, and restricted expression on white blood cells (β₂ and β_7 integrins). Leucocyte integrins have a common β_2 chain that is linked to CD-18 and bind receptors such as ICAM and plasma proteins such as complement components C3b and C4b.9 Collagen-binding integrins have a common β_1 chain that binds various α chains in integrins $\alpha_1\beta_1,\,\alpha_2\beta_1,\,\alpha_{10}\beta_1,$ and $\alpha_{11}\beta_1.$ The $\alpha_2\beta_1$ integrin binds its primary ligand, collagen, 10 and chondroadherin, a matrix protein.¹¹ The RGD-binding integrins have a common α_V chain or β₁ chain. The RGD peptide motif was first discovered in fibronectin¹² but was later found in several other ECM proteins, such as fibronectin, osteopontin, vitronectin, von Willebrand factor (VWF), 15 and laminin. 16 Among the 24 human integrin subtypes known to date, eight integrin dimers recognize the tripeptide RGD motif within ECM proteins, namely: $\alpha_V\beta_1,\,\alpha_V\beta_3,\,\alpha_V\beta_5,\,\alpha_V\beta_6,\,\alpha_V\beta_8,\,\alpha_5\beta_1,\,\alpha_8\beta_1,\,\text{and}\,\,\alpha_{IIb}\beta_3.$ Laminin-binding integrins $(\alpha_3\beta_1,\,\alpha_6\beta_1,\,\alpha_7\beta_1,\,\text{and}\,\,\alpha_6\beta_4)$ mediate the adhesion of cells to basement membranes in various tissues. The $\alpha_4\beta_1,\,\alpha_9\beta_1,\,\text{and}\,\,\alpha_4\beta_7$ integrin family binds fibronectin in a RGD-independent manner.

INTEGRIN-LIGAND BINDING AND CONSEQUENT ACTIVATION

The structure and function of integrins are complex. Integrins bind numerous extracellular ligands, intracellular signaling molecules, and the cytoskeleton in a bivalent-cation-dependent manner with varying specificities. Integrins also have many states with multiple conformations and affinities.

Mechanism of integrin ligand binding and conformational states Integrins bind cell-surface ligands to promote cellular interactions with the ECM and with other cells in the transduction of complex signals that modulate many cellular processes, such as adhesion, migration, and differentiation. These soluble, ECM, or cell surface-bound ligands may include growth factors, structural constituents of the ECM, proteases, cytokines, plasma proteins, microbial pathogens, or receptors specific to immune cells. The affinity and avidity of a ligand may change actively by inside-out signaling in specific pathways. Ligand affinity may vary with the strength of interaction and dissociation of a monovalent protein and its ligand, where ligand avidity refers to its ability to form multiple combinations of bonds.¹⁷

Integrins exist primarily in three conformational states: bent–closed (inactive; the predominant state), extended–closed (active; low affinity or intermediate state), and the extended–open (active; high affinity). The affinity of integrins to various inhibitory and stimulatory ligands is modulated by bivalent cations, which induce a range of conformational changes in integrins ranging from a folded, inactive, and low-affinity state to a high-affinity conformation (Fig. 2). These conformational changes in the extracellular domains of integrins modulate both ligand binding and downstream cellular signaling.

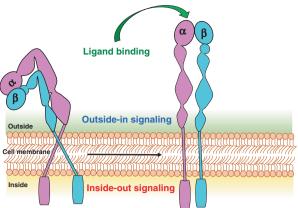


Fig. 2 Schematic of integrin structure and activation. Structurally, the $\alpha\beta$ integrin subunits are type 1 transmembrane proteins. Each subunit consists of one large multi-domain extracellular segment, one transmembrane helix, and a short cytoplasmic tail. The extracellular region interacts with ECM ligands and is composed of about 1104 (700–1100) residues in the α subunit and 778 residues in the β subunits³² and shorter cytoplasmic domains with 30–50 residues.¹⁰⁸ The short cytoplasmic tails are composed of 20–70 amino acids and mediate interactions with intracellular cytoskeletal and signaling proteins. In response to intracellular or extracellular stimuli, integrin activation occurs by ligand binding or by the changes on the cytoplasmic domains, resulting in elongation and separation of the legs. Integrins appear in a closed or "bent" conformation on resting cells and display a low binding affinity for ligand rendering them inactive to ligand binding or signal transduction; while once activated, the integrin shape extends to an open conformation leading to a high affinity.1 In a closed conformation, integrins show low ligand-binding affinity, partly due to the bend in the center of the α and β subunits, which brings the ligand-binding site within 5 nm of the cell surface.1 However, when the conformation is open, the two subunits straighten with increased integrin affinity for the ligand. 111 The initial binding of extracellular ligand effects separation of the cytoplasmic domains, allowing interaction with signal transduction and cytoskeletal molecules during outside-in signaling, while separation of the cytoplasmic domains by talin and other activators activates the head to enable ligand binding during inside-out signaling.

Integrin activation

The activation of integrins increases the affinity of these molecules to extracellular ligands. Integrin tail domains play a critical role in these steps, and any genetic mutations in these parts of integrins can disrupt downstream intracellular signaling. Integrinmediated signaling across cell membranes is typically bidirectional and termed "outside-in" and "inside-out" signaling. When integrins interact with ECM ligands, a conformational change allows adherence to downstream adaptor molecules in the cell-membrane plane. Once clustered, integrins are able to recruit and activate kinases such as Src family kinases, focal adhesion and scaffold molecules such as the adaptor protein p130CRK-associated substrate/breast cancer anti-estrogen resistance 1 (p130CAS/BCAR1). These integrin-associated complexes include discrete active and inactive integrin organizations, which can activate unique signaling pathways. One of these molecules are described in the extractivate unique signaling pathways.

The extracellular domains of integrins are known to undergo a diverse range of conformational changes that alter the ligand-binding domains. In the cytoplasmic tails of integrins, α -helices are seen as heterodimers, 25 and the β -strands often bind intracellular proteins, such as talin or filamin. 26,27 The cytoplasmic tail may undergo several specific conformational changes to bind a range of other signal transducers. 28,29

Integrin bi-directional inside-out and outside-in signaling Mechanical stress³⁰ and extracellular chemicals³¹ can induce rapid conformational changes to cause inside-out activation of integrins.³² Integrins display bi-directional signaling across the plasma membrane. Ligand binding induces extracellular-to-cytoplasm signal transduction, and inside-out signaling or priming regulates integrin-ligand binding conformations (Fig. 2). During integrin activation and signaling, the cytoplasmic tail acts as both a receptor and transmitter of signals. Specifically, during inside-out signaling, the activating signals make an impression on the cytoplasmic tail to induce large conformational changes to the extracellular domain, thereby transforming the integrin from a resting to an active state.³³ During outside-in signaling, the binding of a ligand to the extracellular domain of active integrin transmits a conformational change to the cytoplasmic tail, which leads to the activation of kinases and adaptor molecules in the cytosol. In contrast, talin and kindlin interaction with the βcytoplasmic tail can trigger inside-out signaling, leading to integrin activation, clustering, and recruitment of intercellular adaptor proteins to strengthen cellular adhesion. Talin is a large dimeric actin-binding protein and a major regulator of integrin activation, and the regulation of talin-integrin interactions is important in the control of integrin activation and signaling pathways.³³ Direct interactions between the talin head and the short cytoplasmic tails of β integrin subunits disrupt inhibitory interactions between α and β integrin subunits.³³ This leads to conformational changes in the integrin extracellular domains and consequent increase in their ligand affinity. The role of kindlins are not clearly defined, but they are structurally related to the talin head. The synergistic binding of talin and kindlin to β integrin cytoplasmic tails induces integrin activation by disrupting the α - β interactions at the transmembrane and the cytoplasmic domains. 33,34

INTEGRINS IN INFLAMMATION AND INFECTION

In the resting state, β_2 integrins are expressed specifically on leucocyte receptors. During inflammation, the inflammatory cytokines activate these integrins and promote cellular adherence to the counter-receptors such as ICAMs and promote phagocytosis and cytotoxic killing. Integrin receptors on leukocytes, such as the macrophage-1 antigen (Mac-1, also known as CR3, $\alpha_M\beta_2$, CD11b/CD18) interact with platelet antigens such as the glycoprotein Iba (GPIba) during inflammation. Integrins bind to the pro-domain of transforming growth factor (TGF)- β_1 to activate it and promote its secretion. The pro-TGF- β_3 are biosynthesized and stored in tissues in latent forms, and integrins $\alpha_V\beta_6$ and $\alpha_V\beta_8$ can uniquely bind and activate pro-TGF- β_1 and pro-TGF- β_3 . The $\alpha_V\beta_6$ integrin is known to specifically bind the RGDLXXL/I motif in TGF- β_1 and TGF- β_3 .

 β_2 integrins promote recruitment of leukocytes to the sites of inflammation by promoting the adhesion of circulating leukocytes to vascular endothelium, transendothelial migration, 36,3 formation of immunological synapses in leucocytes, 38 and inflammatory signaling in involved cells. 39 Activated β_2 integrins on dendritic cells (DCs) may act as negative regulators of DC migration in certain conditions and may also regulate T cell activation. 40,41 β integrins on the leukocyte surface are also involved in the tethering, rolling, and adhesion of leukocytes to activated endothelial cells.⁴² β_2 integrins can also initiate intracellular signaling pathways in macrophages and neutrophils and stimulate cytokine secretion from these cells either directly or in synergy with Toll-like receptors (TLRs).⁴³ Integrins may also integrate the impact of the epidermal growth factor receptor, platelet-derived growth factor receptor, insulin receptor, met receptor superfamily (hepatocyte growth factor receptor), and the vascular endothelial growth factor receptor (VEGFR) in inflammatory cells.44

Table 1.	Integrin-targeted murine models and the effect of their signal modulation.		
Integrin	Tissue target	Effect of signal modulation	Mouse model
$\alpha_3\beta_1$	Endothelial cells	Inhibition of angiogenesis	Endothelial cells $\alpha 3-/-$ knockout mice
$\alpha_2\beta_1$	Retinal Muller cells	Reduced neovascularization	$\alpha_2\beta_1$ integrin deficient mice ⁸⁸
$\alpha_2\beta_1$	Mast cells	Cytokine release following Listeria infection	$\alpha_2\beta_1$ knockout mouse model of Listeria infection 62
$\alpha_V \beta_6$	Epithelial cells of the lung	Activates transforming growth factor beta (TGF- β) to regulate pulmonary fibrosis and inflammation	Genetic knockdown ¹¹⁶
α_{V}	Intestinal Th17 cells, colon	Decreased regulatory T (Treg) cells in the colon, leading to severe colitis, autoimmunity, and cancer	α_{v} -deficient mice ¹¹⁷
β_1	Fibroblasts	Delayed cutaneous wound closure and reduced formation of granulation tissue and reduced ECM production	$\beta_1\text{-deficient fibroblast-specific knockout mice}^{118}$
β ₃	Fibroblasts, epithelial cells	Accelerated re-epithelialization, enhanced TGF- $\!\beta$ signaling, dermal fibroblast infiltration	β_3 -deficient mice (genetic knockdown) ¹¹⁹

 β_2 integrins are important regulators of adhesion, leukocyte recruitment, and immunological signaling. These integrins mediate adhesive interactions between myeloid cells, endothelial cells, antigen-presenting cells, T cells and the ECM. L-selectin, the CCR7 chemokine receptor, interacts with specific carbohydrate epitopes on the endothelium and promotes leukocyte rolling and transmigration through the vascular endothelium. Leukocyte rolling induces a rapid, although transient, increase in the affinity of the $\beta 1$ and $\beta 2$ integrins to the endothelial ligands. Conformational changes in the structure of the inserted (I) domain of the α_L subunit of LFA-1 49 enhance firm leukocyte adhesion under shear flow. 31,49

ROLE OF INTEGRINS IN NEONATAL ORGANS DURING NORMAL DEVELOPMENT AND INFLAMMATION

Integrins in the lung during normal development and in inflammation

Integrins and receptor tyrosine kinases act with cytokine and growth factors to modulate the extracellular signal-regulated kinase and phosphatidylinositol 3-kinase (PI3K)-AKT signaling pathways during regeneration, inflammation, developmental, and pathological processes in the developing lung. 2,44,50,51 The ECM in the lung contains collagen, fibronectin, laminin, and entactin,⁵² and alterations in the formation and structure of the ECM during normal development, healing from injury, or in chronic lung disease could lead to profound alterations in the lung structure. 53,54 For instance, fibronectin in the ECM promotes integrin-mediated cellular migration and differentiation of cells during lung development. $^{55}\,\beta_1$ integrin activates several signaling pathways, particularly the PI3K/AKT pathway activated during wound healing in the presence of collage VI in the lung ECM.⁵⁶ β_1 integrins play a critical role in alveolar homeostasis, as seen in chronic lung disease depicted in β_1 integrin-deficient mice.⁵⁷ In addition, β₁ integrin-deficient alveolar epithelial cells produce excessive monocyte chemoattractant protein 1 and reactive oxygen species, suggesting that β_1 integrins may be involved in alveolar homeostasis.⁵⁸ In murine models of bronchopulmonary dysplasia, perinatal exposure to lipopolysaccharide and increased expression of interleukin-33 may activate neutrophils and promote fibronectin degradation in alveolar epithelial cells.5 Other studies have noted increased expression of integrin $\alpha_2\beta_1$ on mast cells and activation/release of inflammatory cytokines. 60,61 Similar findings have been noted in murine models with Listeria monocytogenes infections.⁶² Mice deficient in integrin α_2^{63} and integrin a_{llb}^{64} show defective platelet interaction with collagen. $\alpha_2\beta_1$ integrin-null mice have normal angiogenesis but may have altered angiogenic responses during injury repair.65 In contrast, integrin β₁ knockout mice may have altered development and are not viable, indicating an essential role of β_1 during development. Table 1 outlines murine models of integrins, their target tissues, and signaling.

Integrins in intestinal inflammation and in necrotizing enterocolitis (NEC)

The regulation of intestinal leukocyte responses is vital to maintaining immune homeostasis and prevention of intestinal inflammatory conditions. Integrin $\alpha_v\beta_5$ is expressed on neonatal intestinal macrophages; the expression is developmentally regulated and is not dependent on microbial colonization. These integrins bind different ECM components, such as laminins, collagens, and fibronectin, and are known to coordinate epithelial cell adhesion and movement. These integrins recognize the RGD tri-peptide sequence present in ECM proteins, such as fibronectin and vitronectin. The integrin $\alpha_v\beta_5$ can be found in both focal adhesions and in clathrin-coated membrane domains. The integrin $\alpha_v\beta_5$ can be found in both focal adhesions and in clathrin-coated membrane domains.

Integrin $\alpha_v \beta_8$ plays an important role in epithelial homeostasis and is a major activator of TGF- β expression.⁷¹ α_3 and β_1 integrins, which are known to increase epithelial migration, are upregulated by bacterial products during NEC.^{72,73} TLR4 signaling on enterocytes promotes the efflux of β_1 integrins from the cytoplasm toward the cell membrane and enhances cell-matrix contacts that limit cellular movement.⁷⁴ In other studies, Besner and colleagues have examined the role of E-cadherin and integrins in NEC and showed that the growth factor, heparinbound epidermal growth factor, can promote intestinal restitution in NEC through its effects on integrin-ECM interaction and intercellular adhesions.⁷⁵ Intestinal epithelial cells also express $\alpha_3\beta_1$, another set of integrins of translational importance. In NEC. increased epithelial expression of $\alpha_3\beta_1$ may impair the migration of epithelial cells needed for mucosal wound healing.⁷⁴ However, the same α_3 integrins are also required for morphologic differentiation of the intestinal epithelium in the developing intestine. The physiological needs of the β_1 integrins, therapeutic targeting of these molecules may still be possible with information on the best timing and the possibility of regionally focused intervention.

Integrins in the developing eye and in retinopathy of prematurity In the developing eye, disruption of the oxygen supply to the retina can disrupt neuronal dysfunction needed for transduction and transmission of photosensitive visual signals to the occipital lobe and other cognitive centers. Integrin $\alpha_2\beta_1$ and VEGF interact closely in several intracellular angiogenic signaling (Fig. 3).^{77,78} Cyclic peptides selectively inhibit $\alpha_V\beta_3$ and $\alpha_V\beta_5$, and are potent inhibitors of endothelial cell invasion and differentiation induced by VEGF-A or fibroblast growth factor-2.⁷⁸ Integrin $\alpha_V\beta_3$ works

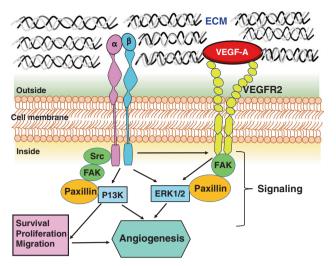


Fig. 3 Schematic of integrin regulation of angiogenic signaling. The schematic shows the interaction between the signaling pathways regulated by $\alpha\beta$ integrins and the VEGF receptor. VEGF-A promotes angiogenesis through VEGF receptor-2 (VEGFR2), a tyrosine kinase receptor expressed by endothelial cells.¹¹² When VEGF-A binds to VEGFR2, numerous intracellular signaling pathways are activated, such as phosphatidylinositol 3-kinase (PI3K), extracellular signal-regulated kinase (Erk), focal adhesion kinase (FAK), c-Src family, and paxillin, a signal transduction adaptor protein associated with focal adhesion. 113,114 Specifically, FAK phosphorylates its substrate, paxillin, which activates ERK signaling. 114 When integrins activate the tyrosine phosphorylation of FAK, it binds to signaling structural proteins, PI3K, and paxillin.¹¹⁵ Src family kinases (SFKs) play a critical role in cell adhesion, survival, and angiogenesis, interact with VEGF receptor, regulate gene expression of angiogenic growth factors, modulate cell proliferation via the mitogen-activated protein kinases (MAPK)-ERK pathway, and interact with integrins to regulate cell adhesion and migration. ECM extracellular matrix, VEGF vascular endothelial growth factor.

synergistically with VEGF to activate angiogenesis in endothelial cells via VEGFR-2 phosphorylation. Pendothelial cells are the primary cells expressing both VEGFR-2 and $\alpha_2\beta_1$ integrin. Proteoglycans such as decorin and perlecan in the ECM of the eye can modulate $\alpha_2\beta_1$ and play a vital role in angiogenesis. The Cterminal fragment of perlecan, known as endorepellin, has an opposite effect and blocks angiogenesis through antagonism of VEGFR-2 and $\alpha_2\beta_1$ integrin on endothelial cells. Retinal pigment epithelial cells express beta-8 integrin at the surface, and the knockdown of beta-8 integrin significantly decreased retinal pigment epithelial cell migration in wound-healing assays. 81

The retinal tissue has one of the body's highest metabolic demands, placing it at risk of injury from oxidative stress, metabolic derangements, and consequent pathologic neovascularization seen in retinopathy of prematurity (ROP) and other proliferative retinal vitreoretinopathies. ROP is a bi-phasic disease of retinal vascular development due to dysregulation of VEGF. 82,83 In phase 1, VEGF is downregulated during exposure to hyperoxia, while in phase 2, VEGF is upregulated in relative/true hypoxia. VEGF is known to have several isoforms; VEGFA165 is the predominant isoform in the eye with multiple pro- and antiangiogenic splice variants.84 In a newborn mouse model of oxygen-induced retinopathy (OIR), oxidative stress from fluctuating hyperoxia and hypoxia leads to altered vascular development with tortuous arteries, dilated veins, and capillary attrition, akin to human ROP.82,85 These changes persist in adult mice with longterm abnormalities in vascularization, structure, and function both in vivo and histologically.86,8

Integrin-targeted therapy holds promise in ROP. Targeting $\alpha_2\beta_1$ integrin expression on endothelial cells mitigates OIR, 88 and the

administration of 3-[3-(6-guanidino-1-oxoisoindolin-2-yl) propanamido]-3-(pyridin-3-yl) propanoic acid dihydrochloride, a novel non-peptide $\alpha\nu\beta3$ antagonist, can inhibit retinal neovascularization. ⁸⁹ There are exciting possibilities that endothelial $\alpha_2\beta_1$ may be therapeutic target in pathological angiogenesis.

INTEGRINS IN THROMBOSIS AND FIBROSIS

Platelet adhesion and signaling play key roles in hemostasis and thrombosis. Two platelet receptors, integrin $\alpha_{llb}\beta_3$ and GPlba, mediate the early and mid-stages of platelet adhesion in the vascular environment. 90 GPlba is a key part of the receptor for VWF, and its binding to VWF enables platelet rolling during the formation of thrombotic plugs at the sites of vascular injury. $^{91-93}$ allbβ3 is expressed on both platelets and the endothelium, and upon activation, it promotes platelet adhesion and aggregation by cross-linking with soluble fibrinogen, fibronectin, and VWF.

In alloimmune thrombocytopenia, autoantibodies are frequently seen against integrin β_3 and GPIba. 94,95 Intracranial hemorrhages may be seen more frequently in infants with anti- β_3 integrin antibodies than in those with antibodies against GPIba. 96 Existing in vitro and in vivo data suggest that the β_3 integrin may bind a wider range of ligands, including fibrinogen and VWF, and autoantibodies that block its function may induce a deeper functional deficit than the anti-GPIba antibodies. 97

INTEGRIN-TARGETED THERAPIES

Integrin dysregulation is implicated in the pathogenesis of numerous diseases with altered angiogenesis, inflammation, or in infectious diseases. In these conditions, therapeutic strategies may either directly target integrins or their ligands. Out of the 24 known human integrins, many have already been identified as therapeutic targets for monoclonal antibodies, peptides, and/or small molecules. In adult subjects, efforts are ongoing to target platelet integrin $\alpha_{IIb}\beta_3$ to prevent thrombotic complications after percutaneous vascular interventions, lymphocyte $\alpha_4\beta_1$ and $\alpha_4\beta_7$ integrins in the treatment of multiple sclerosis, and β_7 integrins $(\alpha_4\beta_7)$ and $\alpha_E\beta_7$ integrins) in inflammatory bowel disease. 98 Specifically, a humanized anti-α4 antibody (Natalizumab) works in reduction of inflammation in multiple sclerosis by blocking the $\alpha_4\beta_1\text{--}vascular$ cell adhesion molecule interaction or the $\alpha_4\beta_7$ -mucosal addressin cell adhesion molecule interaction on mucosal endothelium and blocking leukocyte trafficking across the blood-brain barrier. 99 In another study, a micellar delivery vehicle decorated with an anti-angiogenic peptide has been shown to inhibit $\alpha_V \beta_3$ -mediated neovascularization in endothelial cells.¹⁰⁰ Several anticancer drugs have also been developed against integrin ligands or by using integrin-targeted encapsulated nanoparticles as vehicles to unload drugs into the vasculature of several tumors. 101 In a mouse model of hepatic fibrosis, cyclic peptide-guided liposomes preferentially targeted the activated hepatic stellate cells (not quiescent ones) to treat the fibrotic phenotype. 102 $\alpha_V \beta_3$ antagonists are being tried for the inhibition of retinal neovascularization and may have therapeutic value in ROP.⁸⁹ In a mouse model of laser-induced choroidal neovascularization, intravenous injection of irradiated nanoparticles loaded with doxorubicin allowed nanoparticle accumulation in the neovascular lesions and reduced the size of neovascular lesions.¹⁰³ Integrin antagonists may also be used in fibrotic diseases; IDL-2965 is being studied as a selective, highly potent, anti-fibrotic integrin antagonist in idiopathic pulmonary fibrosis. 104 Small molecule pure antagonists, TDI-4161 and TDI-3761, have been designed to inhibit $\alpha_V \beta_3$ -mediated cell adhesion to $\alpha_V \beta_3$ ligands. Further studies are needed to improve the specificity of anti-integrin drugs to improve both the safety profile and therapeutic success of these agents.

CONCLUSIONS

Enhanced understanding of integrin ligand interactions will enable development of therapies targeting specific receptors in order to modulate angiogenic, thrombotic, infections, and inflammatory disorders. Although numerous animal studies have shown promise in the clinical use of integrins as therapeutic targets, there is a need for clinical studies to confirm efficacy and safety in neonates and young infants. In this review, we have summarized and outlined the roles of integrins in inflammation, angiogenesis, and infectious conditions. Therapies could be targeted specifically to alpha subunits, but their overlapping roles are a critical factor to be considered. Further studies are needed both on molecular signaling and regulatory mechanisms of integrin function and the safety and efficacy in clinical settings.

AUTHOR CONTRIBUTIONS

Substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; drafting the article or revising it critically for important intellectual content; and final approval of the version to be published: both authors.

ADDITIONAL INFORMATION

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REFERENCES

- Morse, E. M., Brahme, N. N. & Calderwood, D. A. Integrin cytoplasmic tail interactions. *Biochemistry* 53, 810–820 (2014).
- Cooper, J. & Giancotti, F. G. Integrin signaling in cancer: mechanotransduction, stemness, epithelial plasticity, and therapeutic resistance. *Cancer Cell* 35, 347–367 (2019).
- 3. Takada, Y., Ye, X. & Simon, S. The integrins. Genome Biol. 8, 215 (2007).
- 4. Hynes, R. O. Integrins: bidirectional, allosteric signaling machines. *Cell* **110**, 673–687 (2002)
- Kurzinger, K., Ho, M. K. & Springer, T. A. Structural homology of a macrophage differentiation antigen and an antigen involved in T-cell-mediated killing. *Nature* 296, 668–670 (1982).
- 6. Bednarczyk, M., Stege, H., Grabbe, S. & Bros, M. β2 Integrins—multi-functional leukocyte receptors in health and disease. *Int. J. Mol. Sci.* **21**, 1402 (2020).
- Lee, J. O., Bankston, L. A., Arnaout, M. A. & Liddington, R. C. Two conformations of the integrin A-domain (I-domain): a pathway for activation? *Structure* 3, 1333–1340 (1995)
- 8. McCarty, J. H., Cook, A. A. & Hynes, R. O. An interaction between ανβ8 integrin and Band 4.1B via a highly conserved region of the band 4.1 C-terminal domain. *Proc. Natl Acad. Sci. USA* **102**, 13479–13483 (2005).
- Koivisto, L., Heino, J., Häkkinen, L. & Larjava, H. Integrins in wound healing. Adv. Wound Care 3, 762–783 (2014).
- Emsley, J., Knight, C. G., Farndale, R. W., Barnes, M. J. & Liddington, R. C. Structural basis of collagen recognition by integrin alpha2beta1. *Cell* 101, 47–56 (2000)
- Haglund, L. et al. Identification and characterization of the integrin alpha2beta1 binding motif in chondroadherin mediating cell attachment. J. Biol. Chem. 286, 3925–3934 (2011).
- Pierschbacher, M. D. & Ruoslahti, E. Cell attachment activity of fibronectin can be duplicated by small synthetic fragments of the molecule. *Nature* 309, 30–33 (1984).
- Oldberg, A., Franzen, A. & Heinegard, D. Cloning and sequence analysis of rat bone sialoprotein (osteopontin) cDNA reveals an Arg-Gly-Asp cell-binding sequence. Proc. Natl Acad. Sci. USA 83, 8819–8823 (1986).
- Suzuki, S., Oldberg, A., Hayman, E. G., Pierschbacher, M. D. & Ruoslahti, E. Complete amino acid sequence of human vitronectin deduced from cDNA. Similarity of cell attachment sites in vitronectin and fibronectin. *EMBO J.* 4, 2519–2524 (1985)
- Plow, E. F., Pierschbacher, M. D., Ruoslahti, E., Marguerie, G. A. & Ginsberg, M. H.
 The effect of Arg-Gly-Asp-containing peptides on fibrinogen and von Will-ebrand factor binding to platelets. Proc. Natl Acad. Sci. USA 82, 8057–8061 (1985).

- Grant, D. S. et al. Two different laminin domains mediate the differentiation of human endothelial cells into capillary-like structures in vitro. Cell 58, 933–943 (1989).
- Abram, C. L. & Lowell, C. A. The ins and outs of leukocyte integrin signaling. Annu. Rev. Immunol. 27, 339–362 (2009).
- Nishida, N. et al. Activation of leukocyte β2 integrins by conversion from bent to extended conformations. *Immunity* 25, 583–594 (2006).
- Mould, A. P., Garratt, A. N., Puzon-McLaughlin, W., Takada, Y. & Humphries, M. J. Regulation of integrin function: evidence that bivalent-cation-induced conformational changes lead to the unmasking of ligand-binding sites within integrin alpha5 beta1. *Biochem. J.* 331(Pt 3), 821–828 (1998).
- 20. Calderwood, D. A. Integrin activation. J. Cell Sci. 117, 657-666 (2004).
- Van Agthoven, J. F. et al. Structural basis for pure antagonism of integrin alphaVbeta3 by a high-affinity form of fibronectin. *Nat. Struct. Mol. Biol.* 21, 383–388 (2014).
- Desgrosellier, J. S. & Cheresh, D. A. Integrins in cancer: biological implications and therapeutic opportunities. *Nat. Rev. Cancer* 10, 9 (2010).
- Li, J. & Springer, T. A. Integrin extension enables ultrasensitive regulation by cytoskeletal force. Proc. Natl Acad. Sci. USA 114, 4685–4690 (2017).
- De Mets, R. et al. Cellular tension encodes local Src-dependent differential beta1 and beta3 integrin mobility. Mol. Biol. Cell 30, 181–190 (2019).
- Bhunia, A., Tang, X. Y., Mohanram, H., Tan, S. M. & Bhattacharjya, S. NMR solution conformations and interactions of integrin alphaLbeta2 cytoplasmic tails. *J. Biol. Chem.* 284, 3873–3884 (2009).
- Kiema, T. et al. The molecular basis of filamin binding to integrins and competition with talin. Mol. Cell 21, 337–347 (2006).
- Ellis, S. J. et al. The talin head domain reinforces integrin-mediated adhesion by promoting adhesion complex stability and clustering. *PLoS Genet.* 10, e1004756 (2014).
- Beglova, N., Blacklow, S. C., Takagi, J. & Springer, T. A. Cysteine-rich module structure reveals a fulcrum for integrin rearrangement upon activation. *Nat. Struct. Biol.* 9, 282–287 (2002).
- Armulik, A., Nilsson, I., von Heijne, G. & Johansson, S. Determination of the border between the transmembrane and cytoplasmic domains of human integrin subunits. J. Biol. Chem. 274, 37030–37034 (1999).
- 30. Zwartz, G. J. et al. Real-time analysis of very late antigen-4 affinity modulation by shear. J. Biol. Chem. 279, 38277–38286 (2004).
- Constantin, G. et al. Chemokines trigger immediate β2 integrin affinity and mobility changes: differential regulation and roles in lymphocyte arrest under flow. *Immunity* 13, 759–769 (2000).
- 32. Arnaout, M. A., Mahalingam, B. & Xiong, J.-P. Integrin structure, allostery, and bidirectional signaling. *Annu. Rev. Cell Dev. Biol.* **21**, 381–410 (2005).
- Calderwood, D. A., Campbell, I. D. & Critchley, D. R. Talins and kindlins: partners in integrin-mediated adhesion. *Nat. Rev. Mol. Cell Biol.* 14, 503–517 (2013).
- Kim, C., Ye, F. & Ginsberg, M. H. Regulation of integrin activation. Annu. Rev. Cell Dev. Biol. 27, 321–345 (2011).
- Dong, X., Hudson, N. E., Lu, C. & Springer, T. A. Structural determinants of integrin beta-subunit specificity for latent TGF-beta. *Nat. Struct. Mol. Biol.* 21, 1091–1096 (2014).
- von Andrian, U. H. et al. Two-step model of leukocyte-endothelial cell interaction in inflammation: distinct roles for LECAM-1 and the leukocyte beta 2 integrins in vivo. Proc. Natl Acad. Sci. USA 88, 7538–7542 (1991).
- Lammermann, T. et al. Rapid leukocyte migration by integrin-independent flowing and squeezing. *Nature* 453, 51–55 (2008).
- Monks, C. R., Freiberg, B. A., Kupfer, H., Sciaky, N. & Kupfer, A. Three-dimensional segregation of supramolecular activation clusters in T cells. *Nature* 395, 82–86 (1998).
- Szukiewicz, D. et al. Chorioamnionitis (ChA) modifies CX3CL1 (fractalkine) production by human amniotic epithelial cells (HAEC) under normoxic and hypoxic conditions. J. Inflamm. 11, 12 (2014).
- Varga, G. et al. Active MAC-1 (CD11b/CD18) on DCs inhibits full T-cell activation. Blood 109, 661–669 (2007).
- Schittenhelm, L., Hilkens, C. M. & Morrison, V. L. β2 integrins as regulators of dendritic cell, monocyte, and macrophage function. *Front. Immunol.* 8, 1866 (2017).
- Herter, J. & Zarbock, A. Integrin regulation during leukocyte recruitment. J. Immunol. 190, 4451–4457 (2013).
- Wolf, D. et al. A ligand-specific blockade of the integrin Mac-1 selectively targets pathologic inflammation while maintaining protective host-defense. *Nat. Commun.* 9, 1–11 (2018).
- Giancotti, F. G. & Tarone, G. Positional control of cell fate through joint integrin/ receptor protein kinase signaling. Annu. Rev. Cell Dev. Biol. 19, 173–206 (2003).
- Shimizu, Y., Rose, D. M. & Ginsberg, M. H. Integrins in the immune system. Adv. Immunol. 72, 325–380 (1999).

- Lasky, L. A. et al. An endothelial ligand for L-selectin is a novel mucin-like molecule. Cell 69, 927–938 (1992).
- Grabovsky, V. et al. Subsecond induction of alpha4 integrin clustering by immobilized chemokines stimulates leukocyte tethering and rolling on endothelial vascular cell adhesion molecule 1 under flow conditions. J. Exp. Med. 192, 495–506 (2000).
- Chen, C. et al. High affinity very late antigen-4 subsets expressed on T cells are mandatory for spontaneous adhesion strengthening but not for rolling on VCAM-1 in shear flow. J. Immunol. 162, 1084–1095 (1999).
- Salas, A., Shimaoka, M., Chen, S., Carman, C. V. & Springer, T. Transition from rolling to firm adhesion is regulated by the conformation of the I domain of the integrin lymphocyte function-associated antigen-1. *J. Biol. Chem.* 277, 50255–50262 (2002).
- Danen, E. H. & Yamada, K. M. Fibronectin, integrins, and growth control. J. Cell. Physiol. 189, 1–13 (2001).
- Mižíková, I. & Morty, R. E. The extracellular matrix in bronchopulmonary dysplasia: target and source. Front. Med. 2, 91 (2015).
- Kahsai, T. Z. et al. Seminiferous tubule basement membrane. Composition and organization of type IV collagen chains, and the linkage of alpha3(IV) and alpha5 (IV) chains. J. Biol. Chem. 272, 17023–17032 (1997).
- Vlahovic, G., Russell, M. L., Mercer, R. R. & Crapo, J. D. Cellular and connective tissue changes in alveolar septal walls in emphysema. *Am. J. Respir. Crit. Care Med.* 160, 2086–2092 (1999).
- Burgstaller, G. et al. The instructive extracellular matrix of the lung: basic composition and alterations in chronic lung disease. Eur. Respir. J. 50, 1601805 (2017)
- Roy, D. C. & Hocking, D. C. Recombinant fibronectin matrix mimetics specify integrin adhesion and extracellular matrix assembly. *Tissue Eng. Part A* 19, 558–570 (2013).
- Mereness, J. A. et al. Type VI collagen promotes lung epithelial cell spreading and wound-closure. PLoS ONE 13, e0209095 (2018).
- 57. Plosa, E. J. et al. β1 integrin regulates adult lung alveolar epithelial cell inflammation. *JCI Insight* **5**, e129259 (2020).
- Plosa, E. J. et al. Epithelial β1 integrin is required for lung branching morphogenesis and alveolarization. *Development* 141, 4751–4762 (2014).
- Jin, R. et al. IL-33-induced neutrophil extracellular traps degrade fibronectin in a murine model of bronchopulmonary dysplasia. *Cell Death Discov.* 6, 33 (2020).
- Ringel-Scaia, V. M., Powell, M. D., Read, K. A., Allen, I. C. & Oestreich, K. J. Systemic Listeria monocytogenes infection as a model to study T helper cell immune responses. Methods Mol. Biol. 1960, 149–160 (2019).
- McCall-Culbreath, K. D., Li, Z. & Zutter, M. M. Crosstalk between the α2β1 integrin and c-met/HGF-R regulates innate immunity. *Blood* 111, 3562–3570 (2008).
- Edelson, B. T., Li, Z., Pappan, L. K. & Zutter, M. M. Mast cell-mediated inflammatory responses require the α2β1 integrin. Blood 103, 2214–2220 (2004).
- Holtkotter, O. et al. Integrin alpha 2-deficient mice develop normally, are fertile, but display partially defective platelet interaction with collagen. *J. Biol. Chem.* 277, 10789–10794 (2002).
- Bouvard, D. et al. Functional consequences of integrin gene mutations in mice. Circ. Res. 89, 211–223 (2001).
- Chen, J., Diacovo, T. G., Grenache, D. G., Santoro, S. A. & Zutter, M. M. The alpha (2) integrin subunit-deficient mouse: a multifaceted phenotype including defects of branching morphogenesis and hemostasis. *Am. J. Pathol.* 161, 337–344 (2002).
- Barczyk, M., Carracedo, S. & Gullberg, D. Integrins. Cell Tissue Res. 339, 269 (2010).
- Bodary, S. & McLean, J. W. The integrin beta 1 subunit associates with the vitronectin receptor alpha v subunit to form a novel vitronectin receptor in a human embryonic kidney cell line. J. Biol. Chem. 265, 5938–5941 (1990).
- Charo, I. F., Nannizzi, L., Smith, J. W. & Cheresh, D. A. The vitronectin receptor avb3 binds fibronectin and acts in concert with a5b1 in promoting cellular attachment and spreading on fibronectin. J. Cell Biol. 111, 2795–2800 (1990).
- Grove, J. et al. Flat clathrin lattices: stable features of the plasma membrane. Mol. Biol. Cell 25, 3581–3594 (2014).
- Lampe, M., Vassilopoulos, S. & Merrifield, C. Clathrin coated pits, plaques and adhesion. J. Struct. Biol. 196, 48–56 (2016).
- Sherman, M. P. New concepts of microbial translocation in the neonatal intestine: mechanisms and prevention. Clin. Perinatol. 37, 565–579 (2010).
- Neu J., Pammi M. Pathogenesis of NEC: impact of an altered intestinal microbiome. Semin. Perinatol. 41, 29–35 (2017).
- Fundora, J. B., Guha, P., Shores, D. R., Pammi, M. & Maheshwari, A. Intestinal dysbiosis and necrotizing enterocolitis: assessment for causality using Bradford Hill criteria. *Pediatr. Res.* 87, 235–248 (2019).

- 74. Qureshi, F. G. et al. Increased expression and function of integrins in enterocytes by endotoxin impairs epithelial restitution. *Gastroenterology* **128**, 1012–1022 (2005)
- Su, Y., Yang, J. & Besner, G. E. HB-EGF promotes intestinal restitution by affecting integrin–extracellular matrix interactions and intercellular adhesions. *Growth Factors* 31, 39–55 (2013).
- Zhang, X., Cromwell, J. W., Kunjummen, B. D., Yee, D. & Garcia-Aguilar, J. The alpha2 and alpha3 integrins are required for morphologic differentiation of an intestinal epithelial cell line. Surgery 133, 429–437 (2003).
- Campochiaro, P. A., Aiello, L. P. & Rosenfeld, P. J. Anti-vascular endothelial growth factor agents in the treatment of retinal disease: from bench to bedside. Ophthalmology 123, S78–S88 (2016).
- Nisato, R. E., Tille, J.-C., Jonczyk, A., Goodman, S. L. & Pepper, M. S. ανβ3 and ανβ5 integrin antagonists inhibit angiogenesis in vitro. *Angiogenesis* 6, 105–119 (2003).
- 79. Soldi, R. et al. Role of $\alpha v \beta 3$ integrin in the activation of vascular endothelial growth factor receptor-2. *EMBO J.* **18**, 882–892 (1999).
- Douglass, S., Goyal, A. & lozzo, R. V. The role of perlecan and endorepellin in the control of tumor angiogenesis and endothelial cell autophagy. *Connect. Tissue Res.* 56. 381–391 (2015).
- Caceres, P. S., Hanke-Gogokhia, C., Ridano, M. E., Zobel, G. & Rodriguez-Boulan, E. Cell-cell and cell-extracellular matrix communication pathways identified in the polarized surface proteome of retinal pigment epithelial cells. FASEB J. 34, 1–1 (2020)
- Mezu-Ndubuisi, O. J. In vivo angiography quantifies oxygen-induced retinopathy vascular recovery. Optom. Vis. Sci. 93, 1268–1279 (2016).
- Mezu-Ndubuisi, O. J. et al. Intravitreal delivery of VEGF-A165-loaded PLGA microparticles reduces retinal vaso-obliteration in an in vivo mouse model of retinopathy of prematurity. Curr. Eye Res. 44, 275–286 (2019).
- Zhao, M. et al. Expression of pro-and anti-angiogenic isoforms of VEGF in the mouse model of oxygen-induced retinopathy. Exp. Eye Res. 93, 921–926 (2011).
- Mezu-Ndubuisi, O. J. et al. In vivo retinal vascular oxygen tension imaging and fluorescein angiography in the mouse model of oxygen-induced retinopathy. *Investiq. Ophthalmol. Vis. Sci.* 54, 6968–6972 (2013).
- Mezu-Ndubuisi, O. J., Adams, T., Taylor, L. K., Nwaba, A. & Eickhoff, J. Simultaneous assessment of aberrant retinal vascularization, thickness, and function in an in vivo mouse oxygen-induced retinopathy model. *Eye* 33, 363–373 (2019).
- Mezu-Ndubuisi, O. J. et al. Long-term evaluation of retinal morphology and function in a mouse model of oxygen-induced retinopathy. *Mol. Vis.* 26, 257–276 (2020).
- Madamanchi, A. et al. Mitigation of oxygen-induced retinopathy in α2β1 integrin-deficient mice. *Investig. Ophthalmol. Vis. Sci.* 55, 4338–4347 (2014).
- Li, Y.-J. et al. Therapeutic efficacy of a novel non-peptide αvβ3 integrin antagonist for pathological retinal angiogenesis in mice. Exp. Eye Res. 129, 119–126 (2014).
- Monti, M. et al. Integrin-dependent cell adhesion to neutrophil extracellular traps through engagement of fibronectin in neutrophil-like cells. PLoS ONE 12, e0171362 (2017).
- 91. Ruggeri, Z. M. Mechanisms initiating platelet thrombus formation. *Thrombosis Haemost.* **78**, 611–616 (1997).
- Savage, B., Almus-Jacobs, F. & Ruggeri, Z. M. Specific synergy of multiple substrate-receptor interactions in platelet thrombus formation under flow. *Cell* 94, 657–666 (1998).
- Tsuji, S. et al. Real-time analysis of mural thrombus formation in various platelet aggregation disorders: distinct shear-dependent roles of platelet receptors and adhesive proteins under flow. Blood. J. Am. Soc. Hematol. 94, 968–975 (1999).
- 94. Brooks, P. C., Clark, R. A. & Cheresh, D. A. Requirement of vascular integrin alpha v beta 3 for angiogenesis. *Science* **264**, 569–571 (1994).
- 95. Di, Q. et al. Impaired cross-activation of $\beta 3$ integrin and VEGFR-2 on endothelial progenitor cells with aging decreases angiogenesis in response to hypoxia. *Int. J. Cardiol.* **168**, 2167–2176 (2013).
- 96. Yougbaré, I. et al. Maternal anti-platelet β3 integrins impair angiogenesis and cause intracranial hemorrhage. *J. Clin. Investig.* **125**, 1545–1556 (2015).
- 97. Yang, H. et al. Fibrinogen and von Willebrand factor-independent platelet aggregation in vitro and in vivo. *J. Thrombosis Haemost.* **4**, 2230–2237 (2006).
- Ley, K., Rivera-Nieves, J., Sandborn, W. J. & Shattil, S. Integrin-based therapeutics: biological basis, clinical use and new drugs. *Nat. Rev. Drug Discov.* 15, 173 (2016).
- 99. O'Connor, P. Natalizumab and the role of α 4-integrin antagonism in the treatment of multiple sclerosis. *Expert Opin. Biol. Ther.* **7**, 123–136 (2007).
- 100. Nagaraj, R. et al. High density display of an anti-angiogenic peptide on micelle surfaces enhances their inhibition of $\alpha v \beta 3$ integrin-mediated neovascularization in vitro. *Nanomaterials* **10**, 581 (2020).

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- Majumder, P. Integrin-mediated delivery of drugs and nucleic acids for antiangiogenic cancer therapy: current landscape and remaining challenges. *Bioengineering* 5, 76 (2018).
- 102. Li, Y. et al. An integrin-based nanoparticle that targets activated hepatic stellate cells and alleviates liver fibrosis. *J. Controlled Release* **303**, 77–90 (2019).
- Wang, Y. et al. Intravenous treatment of choroidal neovascularization by phototargeted nanoparticles. Nat. Commun. 10, 1–9 (2019).
- Kossen, K. et al. IDL-2965: a selective, highly-potent, oral Integrin antagonist for IPF. Eur. Respir. J. 54, PA5374 (2019).
- 105. Li, J. et al. Novel pure αVβ3 integrin antagonists that do not induce receptor extension, prime the receptor, or enhance angiogenesis at low concentrations. ACS Pharmacol. Transl. Sci. 2, 387–401 (2019).
- 106. Tan, S.-M. The leucocyte β2 (CD18) integrins: the structure, functional regulation and signalling properties. Biosci. Rep. 32, 241–269 (2012).
- Fagerholm, S. C., Guenther, C., Asens, M. L., Savinko, T. & Uotila, L. M. Beta2integrins and interacting proteins in leukocyte trafficking, immune suppression, and immunodeficiency disease. *Front. Immunol.* 10, 254 (2019).
- 108. Humphries, M. J. Integrin structure. Biochem. Soc. Trans. 28, 311-339 (2000).
- Luo, B.-H., Carman, C. V. & Springer, T. A. Structural basis of integrin regulation and signaling. *Annu. Rev. Immunol.* 25, 619–647 (2007).
- 110. Xiong, J. P. et al. Crystal structure of the extracellular segment of integrin alpha Vbeta3 in complex with an Arg-Gly-Asp ligand. Science 296, 151–155 (2002).
- Takagi, J., Petre, B. M., Walz, T. & Springer, T. A. Global conformational rearrangements in integrin extracellular domains in outside-in and inside-out signaling. Cell 110, 599–511 (2002).
- 112. Ferrara, N., Gerber, H.-P. & LeCouter, J. The biology of VEGF and its receptors. *Nat. Med.* **9**, 669–676 (2003).
- Abhinand, C. S., Raju, R., Soumya, S. J., Arya, P. S. & Sudhakaran, P. R. VEGF-A/ VEGFR2 signaling network in endothelial cells relevant to angiogenesis. *J. Cell Commun. Signal.* 10, 347–354 (2016).

- 114. Yang, W. J. et al. Paxillin regulates vascular endothelial growth factor A-induced in vitro angiogenesis of human umbilical vein endothelial cells. *Mol. Med. Rep.* 11, 1784–1792 (2015).
- 115. Sin, C. C. Effect of PI3K δ Isoform on the Expression of Integrin Beta 3 and p130cas in Glioblastoma Multiforme. MSc thesis, Hong Kong Polytechnic Univ. (2016).
- 116. Munger J. et al. The integrin alpha v beta 6 binds and activates latent TGF beta
 1: A mechanism for regulating pulmonary inflammation and fibrosis. Cell. 96, 319–328 (1999).
- 117. Acharya M. et al. av Integrin expression by DCs is required for Th17 cell differentiation and development of experimental autoimmune encephalomyelitis in mice. *J Clin Invest.* **120**, 4445–4452 (2010).
- 118. Liu S. et al. Expression of integrin β1 by fibroblasts is required for tissue repair in vivo. *J Cell Sci* **123**, 3674–3682 (2010).
- 119. Reynolds L. et al. Accelerated re-epithelialization in β 3-integrin-deficient-mice is associated with enhanced TGF-β1 signaling. *Nat Med.* **11**, 167–174 (2005).

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