Review Article Hippo-YAP signaling in digestive system tumors

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Abstract: The Hippo pathway is an evolutionally conserved pathway and plays an important role in regulating tissue hemostasis and organ size control. Deregulation of the Hippo pathway is implicated in various human digestive system tumors. The past two decades have witnessed the discovery and elucidation of key signaling components and molecular mechanisms of the Hippo pathway. Among these, the signaling transducers YAP/TAZ are in the center of this complex network to sense and respond to extracellular cues such as cell contact, matrix stiffness and growth factors. In this review, we summarize the biological and clinical significance of Hippo-YAP signaling in the digestive system tumors, and explore the novel therapeutic strategies for targeting Hippo-YAP signaling.

Keywords: Hippo pathway, YAP, esophagus, stomach, colon, pancreas, liver, tumorigenesis

Introduction

Originally discovered through Drosophila genetic screen for molecules regulating organ size, the Hippo-YAP pathway has emerged as an evolutionally conserved signaling pathway to regulate various biological functions including tissue homeostasis, regeneration and tumorigenesis [1, 2]. The first Hippo pathway gene identified was the NDR family protein kinase Warts (homologous to mammalian gene Large tumor suppressor kinase 1/2 [Lats1/2]) [3, 4]. Inactivation of Warts gene leads to dramatic tissue overgrowth in Drosophila. Subsequently, additional Hippo pathway components, including Salvador (homologous to mammalian gene Sav1, also known as WW45) [5, 6], Ste20-like protein kinase Hippo (homologous to mammalian gene MST1/2) [7-11], and Mats (homologous to mammalian gene Mob1) [12], were identified through similar genetic approaches. The identification of Yki (homologous to mammalian gene Yes-Associated Protein [YAP] and Transcriptional Co-activator with PDZ-binding Motif [TAZ]), published in a seminal article in 2005 [13], completed the missing link in the Hippo signaling kinase cascade. In mammals, the classical pathway proceeds as follows:

Mst1/2 kinase binds its cofactor Sav1 (also known as WW domain containing adaptor 45, or WW45), and this complex phosphorylates and activates Lats1/2 kinase. Lats1/2 kinase then binds its cofactor Mob1, which phosphorylates and inactivates transcription coactivators YAP and TAZ. In the setting of signaling deregulation, YAP is dephosphorylated and translocate to nucleus. In conjunction with its transcription factor TEA domain family members (TEAD 1-4) [14-16], nuclear YAP functions as a potent transcription coactivator to promote target gene transcription (Figure 1). Besides transcription factor TEAD, the nuclear events for Hippo-YAP signaling also involve Vestigial-like family transcriptional coactivator VGLL 1-4. In the Hippo pathway, VGLL could compete with YAP/TAZ for TEAD binding [17]. The dramatic tissue overgrowth phonotype observed in Hippo pathway inactivation in Drosophila has been attributed to the ability of activated YAP to induce transcription of genes that promote cell growth and proliferation, as well as inhibition of apoptosis [7].

Multiple membrane-associated cytoskeletal proteins, including Merlin (homologous to mammalian gene NF2) and Expanded [18], Kibra



Figure 1. Schematic of Hippo-YAP signaling. When Hippo signaling is on, NF2 recruits Lats1/2 kinase to the membrane. Mst1/2 kinase will then phosphorylate Lats1/2 kinase, which leads to phosphorylation, cytoplasmic retention, and subsequent ubiquitination and degradation of YAP. When Hippo signaling is off, YAP translocates to the nucleus and competes with VGLL for TEAD binding to regulate target gene expression with resultant anti-apoptosis and pro-proliferation effects.

[19-21], and Fat [22-25], have been identified as upstream components and regulators of Hippo-YAP signaling pathway. These upstream components provide important clues to answering the ultimate question: how is Hippo-YAP signaling regulated by environmental cues for tumor initiation, progression and metastasis?

The first clue for this question comes from cellcell contact inhibition, a well-known phenomenon in which cells cease proliferation once they contact another cell [26]. It is a common belief that cancer cells have the ability to escape from this barrier to achieve uncontrolled growth [27]. Phosphorylation and inactivation of YAP through the Hippo pathway plays an important role to control cell growth and proliferation upon cell contact [28]. At low cell density, YAP is active and predominantly localized in the nuclei to promote target gene transcription. High cell density, on the other hand, leads to Hippo pathway-dependent YAP phosphorylation and cytoplasmic translocation for its deactivation [28]. The interaction between YAP and alpha-catenin, a critical cell-cell junction component and cell density sensor in the skin, has been demonstrated in this process to control epidermal stem cell proliferation and tumorigenesis [29]. Cell density and NF2 could also control transcription factor TEAD activity through regulation of its palmitoylation status [30].

The Hippo-YAP signaling also plays an important role in matrix stiffening-associated mechanotransduction. Frequently, tumors are stiffer than the surrounding benign tissue, largely due to increased fibrosis, interstitial pressure and angiogenic blood flow shear stress [31, 32]. Mechanotransduction is the concept that the cancer and stromal cells can sense matrix stiffening and transduce this mechanical force to promote tumorigenic biochemical pathways for cell cycle progression, epithelial-mesenchymal transition, cell motility and metastasis. YAP is essential for tissue tension and morphogenesis during development [33]. Activation of the YAP is also a signature feature of cancer-associated fibroblasts (CAFs), and YAP-dependent matrix remodeling is required for CAFs to promote cancer cell invasion and angiogenesis [34].

In the past decade, the combined efforts of genetics and biochemistry have furthered our understanding of this important signaling pathway. With the advances in sequencing technology, it has become evident that the Hippo-YAP signaling pathway is frequently deregulated in different human cancers. In this review, we will focus on its role in gastrointestinal, pancreatic and hepatic tumorigenesis.

Hippo-YAP signaling in esophageal cancer

Esophageal cancer is the seventh most common cancer worldwide. It is also the sixth most common cause for cancer-associated death [35]. Esophageal squamous cell carcinoma is the most prevalent esophageal cancer worldwide, with a high incidence in developing countries. In the United States, even with decreasing incidence of esophageal squamous cell carcinoma, the prevalence, incidence and associated mortality of esophageal cancer (adenocarcinoma) have been increasing over the past several decades, largely due to the pandemic of obesity and associated acid reflux [36].

Frequent inactivating mutations of the Hippo-YAP signaling, including mutations in AJUBA, FAT1-4, STK3, LATS1 and YAP, have been identified and validated in roughly half of the esophageal squamous cell carcinoma cases [37, 38], supporting the critical role of the Hippo-YAP signaling in the tumorigenesis of esophageal squamous cell carcinoma. The Cancer Genome Atlas (TCGA) Research Network detected higher rates of YAP (11q22.1) amplification and deletion of VGLL4, a negative regulator of Hippo-YAP signaling, in esophageal squamous cell carcinoma [39]. This study also demonstrated the strong resemblance of esophageal adenocarcinoma with the chromosomally unstable variant of gastric adenocarcinoma [39].

Unlike esophageal squamous cell carcinoma, the involvement of the Hippo-YAP pathway in the development of esophageal adenocarcinoma is less clear. One study reported increased cytoplasmic and nuclear YAP expression in esophageal high-grade dysplasia and adenocarcinoma [40]. Preliminary data from our investigations also suggests that Hippo signaling effector YAP plays critical roles in the progression of neoplasia associated with Barrett's esophagus (BE), a precursor lesion of esophageal adenocarcinoma that is more commonly seen in patients with longstanding gastroesophageal reflux disease. YAP expression is absent on the surface of benign esophageal columnar epithelium (Figure 2A), and is only weakly expressed in the deep crypts of benign esophageal mucus glands and non-dysplastic BE glands. In contrast, BE with high-grade dysplasia (Figure 2B) and esophageal adenocarcinoma show significantly increased YAP expression (Figure 2C). These data support a potential role of Hippo-YAP signaling in the tumorigenesis of esophageal adenocarcinoma.

Hippo-YAP signaling in gastric cancer

Gastric cancer is the fifth most common cancer and the fourth most common cause of cancerassociated death worldwide [35]. With adenocarcinoma being the most common cancer type, gastric cancer has a clear relationship with long-standing chronic inflammation [41]. The common environmental factors include infectious agents such as Helicobacter pylori (HP) and Epstein-Barr virus (EBV), tobacco use, and dietary factors. HP eradication could reduce the risk of gastric adenocarcinoma [42]. Based on TCGA database, Hippo-YAP pathway is positively associated with HP infection [43]. Infection with cytotoxin-associated gene A (CagA) positive HP is the strongest risk factor for the development of gastric adenocarcinoma [44, 45]. CagA+ HP infection significantly increased YAP expression in gastric epithelial cells, as compared with CagA- HP strain. Furthermore, CagA promotes epithelial mesenchymal transition in gastric carcinogenesis through Hippo-YAP pathway [46].

Multiple studies also confirm that the expression of YAP is significantly increased in gastric adenocarcinoma [47-49] (**Figure 2D**), with higher levels of cytoplasmic YAP in the early tumor stages and higher levels of nuclear YAP in the advanced tumor stages. Nuclear YAP accumulation is associated with poor survival, especially in patients with early-stage gastric cancer



Figure 2. YAP expression in various digestive system tumors. (A-C) YAP expression in Barrett's esophagus and esophageal adenocarcinoma. A. Negative YAP expression in benign esophageal columnar epithelium (×200); B. YAP expression in Barrett's esophagus with high-grade dysplasia, note the increased YAP expression extending to surface epithelium (×200); C. YAP expression in esophageal adenocarcinoma, note the sharp transition between non-dysplastic mucosa and carcinoma (×200); D. YAP expression in gastric adenocarcinoma (×100); E. YAP expression in colorectal adenocarcinoma (×100); F. YAP expression in pancreatic ductal adenocarcinoma (×100).

[50]. YAP mediates peritoneal carcinomatosis in advanced gastric adenocarcinoma patients [51]. Oncoprotein Myc has been identified as a key downstream target of YAP to initiate gastric tumorigenesis [52]. Up-regulation of YAP significantly correlates with HP infection status and gastric cancer progression including tumor size and tumor stage. On the contrary, the transcription levels of VGLL4, a negative regulator of Hippo signaling, were downregulated in 61% of gastric cancer samples, with inverse correlation with gastric cancer progression (tumor size, tumor stage and lymph node metastasis) [53]. The YAP/VGLL4 ratio has the potential to serve as a biomarker for gastric cancer.

In a study of 86 patients with advanced gastric cancer who received first-line chemotherapy, a significant association between nuclear TAZ expression and Wnt mutations was noted. The patients with tumors harboring a signature of combined nuclear TAZ expression and Wnt mutations had significantly increased risk of disease progression and poor overall survival [54]. Interestingly, the patients with tumors harboring a different signature (combined YAP expression and *TP53* mutations) had favorable survival outcomes that is linked to the administration of first-line chemotherapy [55].

Hippo-YAP signaling in colorectal cancer

Colorectal cancer is the third most common cancer and the second leading cause of cancer

death in the United States [35]. Colorectal cancers can be divided into 3 major categories based on their unique molecular signatures: chromosomal instability pathway (84%) involving molecular alterations of APC, TP53, KRAS, SMAD and PIK3CA; microsatellite instability (MSI) hypermutant pathway (13%) involving the mismatch repair genes MLH1 and MSH2; ultramutant pathway (3%) involving POLE gene [56].

Adenomatous polyposis coli (APC) alterations are the most common molecular events for the colorectal cancer, mostly through conventional adenoma-carcinoma sequence [57]. APC is one of the key players in the Wnt pathway, a critical signaling pathway in regulating intestinal stem cell proliferation, differentiation and cell-fate decisions. Loss-of-function APC gene mutations lead to activation of Wnt pathway through accumulation of nuclear β-catenin. The cross-talk between Hippo and Wnt pathway plays important roles during intestinal regeneration, homeostasis and tumorigenesis [58]. The interactions of these two important pathways appear complex, multi-layered, and context-dependent. In the nucleus, a β-catenin YAP complex is essential to the transformation and survival of β-catenin-dependent colon cancers [59]. However, in the cytoplasm, YAP/TAZ can function as negative regulators of Wnt/βcatenin signaling [60-62]. On the other hand, Wnt-APC pathway negatively regulates YAP function through β-catenin destruction complex [61]. Additionally, APC can function as a scaffold protein to promote Hippo signaling and to restrict YAP activity [63]. Upon APC deficiency, YAP/TAZ and β -catenin are required for intestinal crypt overgrowth and adenoma formation [61, 63].

In the normal colon tissue, YAP is predominantly expressed in the basal crypt zones where the intestinal progenitor cells are located. YAP is critical for intestinal regeneration and tumorigenesis [64, 65]. Colonic adenocarcinoma tissues have significantly elevated YAP protein expression in both the cytoplasm and nucleus [66] (Figure 2E). Elevated protein expression levels of YAP and TAZ are independent predictors of poor prognosis and are positively correlated with tumor stage, nodal status, and metastasis [66-69]. Elevated YAP protein expression, together with high plasma carcinoembryonic antigen (CEA) level, are potential prognostic biomarkers in patients with earlystage colorectal cancer [68]. Consistently, positive expression of VGLL4, combined with low YAP expression, is an independent good prognostic factor in colorectal cancers [70].

Although YAP has well been accepted as a potent oncoprotein, dual roles of YAP in colorectal cancer have been proposed, and there are multiple lines of evidence that YAP may also function as tumor suppressor, presumably in a context-dependent manner [71]. One example is its negative regulation of Wnt signaling. During intestinal regeneration, loss of YAP causes Wnt pathway hyperactivation, resulting in intestinal stem cell expansion, as well as ectopic crypt and microadenoma formation [62]. Alternatively, YAP is able to induce DNA damage-associated apoptosis through p73 transcription factor [72-74]. Complete loss of YAP expression was correlated with aggressive clinical behaviors, including larger tumor size, higher tumor stage, and worse overall survival, and therefore represented an aggressive subtype of colorectal cancer [75]. The proposed tumor suppressor function of YAP might also play an important role in chronic colitis-associated colorectal cancer (CAC). In a recent study, our group has identified a subgroup of CAC with double negative expression of YAP and CDX2 that is more frequently seen in younger patients (mean age of 45.3 years) with higher pathological tumor stage and nodal metastasis [76].

Hippo-YAP signaling in pancreatic cancer

Pancreatic ductal adenocarcinoma (PDAC), the most common form of pancreatic cancer, is one of the most aggressive cancers and ranks the seventh of all cancer-associated death [35]. Through next-generation sequencing, recurrent molecular alterations have been found in PDAC, with the oncogenic mutations of KRAS and loss-of-function mutation/deletion of tumor suppressor genes TP53, SMAD4 and CDKN2A (P16) being the most common [77]. Oncogenic mutations of KRAS are present in the majority of human PDAC cases, and targeting KRAS pharmacologically has a great clinical potential for the treatment of pancreatic cancer. Two elegant studies have independently identified YAP as a central driver to compensate for the loss of KRAS signaling and to sustain tumor maintenance in an advanced PDAC mouse model [78, 79]. In a KRAS-mutant mouse model. YAP is also required for pancreatitis-induced acinar-to-ductal metaplasia, a precursor of pancreatic intraepithelial neoplasia that can progress to PDAC [80].

YAP is highly expressed in PDAC [79, 81-83] (Figure 2F). In fact, YAP is the top upregulated protein in PDAC samples on mass spectrometry-based proteomics [84]. Expression level of YAP is an independent predictor of poor overall survival (OS), disease-free survival (DFS) and liver metastasis [83-85], partially through its ability to promote transcription of target genes involved in the remodeling of pancreatic tumor microenvironment. YAP can also modulate the tumor immune microenvironment in PDAC by promoting the expression of cytokines/chemokines that recruit and differentiate myeloidderived suppressor cells [86]. Therefore, YAP/ TAZ is at the center of a signaling network that is crucial for the development and progression of PDAC [87].

Hippo-YAP signaling in liver cancer

Liver cancer is the sixth most common cancer and the third leading cause of cancer-related death worldwide [35]. Hepatocellular carcinoma (HCC) is the most common primary liver cancer, and its major risk factors include viral infection (hepatitis B or C virus), alcohol abuse, obesity and iron overload, among others [88]. Due to its high degree of heterogeneity, HCC remains a challenge for actionable molecular subtyping.

During the past decade, Hippo-YAP signaling has emerged as a key pathway for the normal liver homeostasis maintenance, as well as the initiation and progression of liver cancer. In mouse model, liver-specific deletion of Hippo pathway tumor suppressor components NF2 [89], Mst1/2 [90-92], Sav1 [92, 93], Mob1 [94] and Lats1/2 [95], or conditional overexpression of YAP in the liver [96] all leads to hepatomegaly and liver cancer formation. The liver tumors developed in these mouse model include hepatic adenoma, HCC and intrahepatic cholangiocarcinoma (iCC). Interesting, heterozygosity of YAP is sufficient to suppress hepatomegaly and HCC formation in those mouse models, supporting YAP/TAZ as potential therapeutic targets for HCC. Besides its function to regulate cellular proliferation and organ size, Hippo-YAP signaling also plays a critical role by influencing cell fates in the liver. YAP could induce a ductal fate in hepatocytes by dedifferentiating hepatocytes into progenitor cells [97]. These findings lay the groundwork for regenerative medicine of liver disease and liver cancer through the manipulation of Hippo-YAP signaling.

The role of Hippo-YAP signaling in liver tumorigenesis is quite complex, even with multiple lines of evidence demonstrating the potent oncogenic activity of YAP/TAZ. In a recent Science article, Moya et al. have revealed an unexpected non-autonomous tumor suppressor function of YAP/TAZ in liver cancer [98]. Overexpression of YAP in peritumoral hepatocytes triggered regression of primary liver tumors and melanoma-derived liver metastases, and deletion of YAP/TAZ in peritumoral hepatocytes accelerated liver tumor growth. These interesting findings support a "cell competition" theory originally proposed in a Drosophila study [99], that during interaction between tumor cells and their surrounding tissue, the cell with low YAP/TAZ level will become the "loser" and be eliminated through cell-cell competition by the "winner", the cell with high YAP/TAZ level.

In human HCC, YAP overexpression is significantly associated with poor tumor differentiation and high serum alpha-fetoprotein (AFP) level and is an independent predictor for HCC-

specific OS and DFS [100, 101]. The serum level of Amphiregulin, a member of the epidermal growth factor family and one of the direct target genes of YAP/TAZ, is frequently elevated in HCC patients and may be used as a serological biomarker of HCC [100]. Besides Amphiregulin, it is now evident that YAP overexpression can induce the expression and secretion of many paracrine-acting factors including plasminogen activator inhibitor-1 (PAI-1), C-X-C motif chemokine ligand 13 (CXCL13), CXCL16 during liver tumorigenesis. Amphiregulin and PAI-1 may play unique roles in liver tumorigenesis and therefore could serve as novel targets in HCC therapy [102-105]. Sohn et al. further extended these findings and identified a unique subgroup of HCC with silence of Hippo (SOH) signature. The SOH subgroup consists approximately 21% in the National Cancer Institute cohort, has a significantly poorer prognosis than non-SOH HCC patients. The SOH signature was an independent predictor of DFS on multivariate analysis [106].

YAP/TAZ has also played a critical role for epithelioid hemangioendothelioma (EHE), a rare form of primary liver sarcoma. The chromosomal translocations involving 1p36.3 and 3q25, with formation of TAZ (WWTR1) - CAMTA1 (calmodulin-binding transcription activator 1) gene fusion, has been identified in about 90% of EHE cases [107, 108]. The remaining 10% of EHE cases harbor YAP-TFE3 gene fusion [109]. CAMTA1 nuclear expression by immunohistochemistry has been proven to be highly sensitive and specific for the diagnosis of hepatic EHE, and could be used to differentiate hepatic EHE from its close mimic angiosarcoma [110]. In two recent studies, TAZ-CAMTA1 gene fusion is sufficient to drive EHE tumorigenesis by dysregulating YAP/TAZ signaling in a mouse model [111, 112].

Therapeutic perspective

Given its important role in tumorigenesis, targeting Hippo-YAP signaling is a novel and attractive approach for cancer therapy. As the signaling transducer and nexus of Hippo pathway, YAP/TAZ has been in the center for such development. Few experimental prototype molecules have been discovered due to their ability to suppress YAP/TAZ activity. Verteporfin, the first photosensitizer approved for the treatment of age-related macular degeneration, was identi-

fied as a potent inhibitor of the physical interaction between YAP with transcription factor TEAD. In a mouse model, Verteporfin significantly suppressed YAP-induced hepatomegaly [113]. Song et al. identified CA3 through a small molecular screen that could potently inhibit YAP/TEAD transcriptional activity. CA3 inhibits esophageal adenocarcinoma cell growth both in vitro and in vivo [114]. Jiao et al. designed a VGLL4-mimicking peptide that could suppress gastric cancer growth through competing with YAP for TEAD interaction [53]. Drugs targeting the cellular receptors including G-proteincoupled receptor (GPCR) [115, 116], epidermal growth factor receptor (EGFR) [117, 118] and vascular endothelial growth factor receptor (VEGFR) [119, 120] could also inhibit YAP/TAZ activity through an indirect manner.

Moroishi et al. revealed an unexpected function of Hippo pathway in modulating tumor immunogenicity. In tumor cells, deletion of LATS1/2 leads to enhanced tumor-specific humoral and cellular immune responses, resulting in immune-mediated tumor cell elimination. Deletion of LATS1/2 also enhances tumor vaccine efficacy through adaptive immunity. These interesting findings raise the possibility of targeting LATS1/2 kinase in cancer immunotherapy [121]. Meanwhile, pharmacological targeting of MST1/2 kinase has resulted in the discovery of XMU-MP-1, a potent and selective inhibitor of MST1/2 kinase. XMU-MP-1 has demonstrated its in vivo efficacy to promotes liver and intestinal regeneration and to attenuate acetaminophen-induced liver injury [122]. Searching for potent and specific small molecule kinase inhibitor targeting Mst1/2 and Lats1/2 kinases is therefore of great clinical relevance.

Conclusions

As an evolutionally conserved pathway, Hippo-YAP signaling has played an important role in regulating cell proliferation, apoptosis and differentiation from *Drosophila* to human. Hippo-YAP signaling has been shown to extensively involve in intestinal stem cell reprogramming, intestinal and liver regeneration, and digestive system tumorigenesis. YAP/TAZ may function either as oncogene or tumor suppressor depending on the cellular context. Current studies have begun to reveal its complex multifaceted nature, as well as its complicated interplay with other conserved signaling pathways, such as the Wnt/ β -catenin/APC pathway. It is of great clinical interest to develop pharmacological molecules targeting Hippo-YAP signaling for personalized medicine on digestive system tumors.

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Disclosure of conflict of interest

None.

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References

- [1] Zheng Y and Pan D. The hippo signaling pathway in development and disease. Dev Cell 2019; 50: 264-282.
- [2] Yu FX, Zhao B and Guan KL. Hippo pathway in organ size control, tissue homeostasis, and cancer. Cell 2015; 163: 811-828.
- [3] Justice RW, Zilian O, Woods DF, Noll M and Bryant PJ. The Drosophila tumor suppressor gene warts encodes a homolog of human myotonic dystrophy kinase and is required for the control of cell shape and proliferation. Genes Dev 1995; 9: 534-546.
- [4] Xu T, Wang W, Zhang S, Stewart RA and Yu W. Identifying tumor suppressors in genetic mosaics: the Drosophila lats gene encodes a putative protein kinase. Development 1995; 121: 1053-1063.
- [5] Tapon N, Harvey KF, Bell DW, Wahrer DC, Schiripo TA, Haber D and Hariharan IK. salvador Promotes both cell cycle exit and apoptosis in Drosophila and is mutated in human cancer cell lines. Cell 2002; 110: 467-478.
- [6] Kango-Singh M, Nolo R, Tao C, Verstreken P, Hiesinger PR, Bellen HJ and Halder G. Shar-pei mediates cell proliferation arrest during imaginal disc growth in Drosophila. Development 2002; 129: 5719-5730.
- [7] Wu S, Huang J, Dong J and Pan D. Hippo encodes a Ste-20 family protein kinase that restricts cell proliferation and promotes apoptosis in conjunction with salvador and warts. Cell 2003; 114: 445-456.

- [8] Harvey KF, Pfleger CM and Hariharan IK. The Drosophila Mst ortholog, hippo, restricts growth and cell proliferation and promotes apoptosis. Cell 2003; 114: 457-467.
- [9] Udan RS, Kango-Singh M, Nolo R, Tao C and Halder G. Hippo promotes proliferation arrest and apoptosis in the Salvador/Warts pathway. Nat Cell Biol 2003; 5: 914-920.
- [10] Pantalacci S, Tapon N and Léopold P. The Salvador partner Hippo promotes apoptosis and cell-cycle exit in Drosophila. Nat Cell Biol 2003; 5: 921-927.
- [11] Jia J, Zhang W, Wang B, Trinko R and Jiang J. The Drosophila Ste20 family kinase dMST functions as a tumor suppressor by restricting cell proliferation and promoting apoptosis. Genes Dev 2003; 17: 2514-2519.
- [12] Lai ZC, Wei X, Shimizu T, Ramos E, Rohrbaugh M, Nikolaidis N, Ho LL and Li Y. Control of cell proliferation and apoptosis by mob as tumor suppressor, mats. Cell 2005; 120: 675-685.
- [13] Huang J, Wu S, Barrera J, Matthews K and Pan D. The Hippo signaling pathway coordinately regulates cell proliferation and apoptosis by inactivating Yorkie, the Drosophila Homolog of YAP. Cell 2005; 122: 421-434.
- [14] Zhang L, Ren F, Zhang Q, Chen Y, Wang B and Jiang J. The TEAD/TEF family of transcription factor Scalloped mediates Hippo signaling in organ size control. Dev Cell 2008; 14: 377-387.
- [15] Goulev Y, Fauny JD, Gonzalez-Marti B, Flagiello D, Silber J and Zider A. SCALLOPED interacts with YORKIE, the nuclear effector of the hippo tumor-suppressor pathway in Drosophila. Curr Biol 2008; 18: 435-441.
- [16] Wu S, Liu Y, Zheng Y, Dong J and Pan D. The TEAD/TEF family protein Scalloped mediates transcriptional output of the Hippo growth-regulatory pathway. Dev Cell 2008; 14: 388-398.
- [17] Koontz LM, Liu-Chittenden Y, Yin F, Zheng Y, Yu J, Huang B, Chen Q, Wu S and Pan D. The Hippo effector Yorkie controls normal tissue growth by antagonizing scalloped-mediated default repression. Dev Cell 2013; 25: 388-401.
- [18] Hamaratoglu F, Willecke M, Kango-Singh M, Nolo R, Hyun E, Tao C, Jafar-Nejad H and Halder G. The tumour-suppressor genes NF2/ Merlin and Expanded act through Hippo signalling to regulate cell proliferation and apoptosis. Nat Cell Biol 2006; 8: 27-36.
- [19] Yu J, Zheng Y, Dong J, Klusza S, Deng WM and Pan D. Kibra functions as a tumor suppressor protein that regulates Hippo signaling in conjunction with Merlin and expanded. Dev Cell 2010; 18: 288-299.
- [20] Baumgartner R, Poernbacher I, Buser N, Hafen E and Stocker H. The WW domain protein Kibra

acts upstream of Hippo in Drosophila. Dev Cell 2010; 18: 309-316.

- [21] Genevet A, Wehr MC, Brain R, Thompson BJ and Tapon N. Kibra is a regulator of the Salvador/Warts/Hippo signaling network. Dev Cell 2010; 18: 300-308.
- [22] Bennett FC and Harvey KF. Fat cadherin modulates organ size in Drosophila via the Salvador/Warts/Hippo signaling pathway. Curr Biol 2006; 16: 2101-2110.
- [23] Silva E, Tsatskis Y, Gardano L, Tapon N and Mc-Neill H. The tumor-suppressor gene fat controls tissue growth upstream of expanded in the hippo signaling pathway. Curr Biol 2006; 16: 2081-2089.
- [24] Willecke M, Hamaratoglu F, Kango-Singh M, Udan R, Chen CL, Tao C, Zhang X and Halder G. The fat cadherin acts through the hippo tumorsuppressor pathway to regulate tissue size. Curr Biol 2006; 16: 2090-2100.
- [25] Cho E, Feng Y, Rauskolb C, Maitra S, Fehon R and Irvine KD. Delineation of a Fat tumor suppressor pathway. Nat Genet 2006; 38: 1142-1150.
- [26] Eagle H and Levine EM. Growth regulatory effects of cellular interaction. Nature 1967; 213: 1102-1106.
- [27] Hanahan D and Weinberg RA. The hallmarks of cancer. Cell 2000; 100: 57-70.
- [28] Zhao B, Wei X, Li W, Udan RS, Yang Q, Kim J, Xie J, Ikenoue T, Yu J, Li L, Zheng P, Ye K, Chinnaiyan A, Halder G, Lai ZC and Guan KL. Inactivation of YAP oncoprotein by the Hippo pathway is involved in cell contact inhibition and tissue growth control. Genes Dev 2007; 21: 2747-2761.
- [29] Schlegelmilch K, Mohseni M, Kirak O, Pruszak J, Rodriguez JR, Zhou D, Kreger BT, Vasioukhin V, Avruch J, Brummelkamp TR and Camargo FD. Yap1 acts downstream of α -catenin to control epidermal proliferation. Cell 2011; 144: 782-795.
- [30] Kim NG and Gumbiner BM. Cell contact and Nf2/Merlin-dependent regulation of TEAD palmitoylation and activity. Proc Natl Acad Sci U S A 2019; 116: 9877-9882.
- [31] Broders-Bondon F, Nguyen Ho-Bouldoires TH, Fernandez-Sanchez ME and Farge E. Mechanotransduction in tumor progression: the dark side of the force. J Cell Biol 2018; 217: 1571-1587.
- [32] Chin L, Xia Y, Discher DE and Janmey PA. Mechanotransduction in cancer. Curr Opin Chem Eng 2016; 11: 77-84.
- [33] Porazinski S, Wang H, Asaoka Y, Behrndt M, Miyamoto T, Morita H, Hata S, Sasaki T, Krens SFG, Osada Y, Asaka S, Momoi A, Linton S, Miesfeld JB, Link BA, Senga T, Shimizu N, Nagase H, Matsuura S, Bagby S, Kondoh H, Nishi-

na H, Heisenberg CP and Furutani-Seiki M. YAP is essential for tissue tension to ensure vertebrate 3D body shape. Nature 2015; 521: 217-221.

- [34] Calvo F, Ege N, Grande-Garcia A, Hooper S, Jenkins RP, Chaudhry SI, Harrington K, Williamson P, Moeendarbary E, Charras G and Sahai E. Mechanotransduction and YAP-dependent matrix remodelling is required for the generation and maintenance of cancer-associated fibroblasts. Nat Cell Biol 2013; 15: 637-646.
- [35] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A and Bray F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021; 71: 209-249.
- [36] Patel N and Benipal B. Incidence of esophageal cancer in the United States from 2001-2015: a United States Cancer Statistics Analysis of 50 States. Cureus 2018; 10: e3709.
- [37] Gao YB, Chen ZL, Li JG, Hu XD, Shi XJ, Sun ZM, Zhang F, Zhao ZR, Li ZT, Liu ZY, Zhao YD, Sun J, Zhou CC, Yao R, Wang SY, Wang P, Sun N, Zhang BH, Dong JS, Yu Y, Luo M, Feng XL, Shi SS, Zhou F, Tan FW, Qiu B, Li N, Shao K, Zhang LJ, Zhang LJ, Xue Q, Gao SG and He J. Genetic landscape of esophageal squamous cell carcinoma. Nat Genet 2014; 46: 1097-1102.
- [38] Sawada G, Niida A, Uchi R, Hirata H, Shimamura T, Suzuki Y, Shiraishi Y, Chiba K, Imoto S, Takahashi Y, Iwaya T, Sudo T, Hayashi T, Takai H, Kawasaki Y, Matsukawa T, Eguchi H, Sugimachi K, Tanaka F, Suzuki H, Yamamoto K, Ishii H, Shimizu M, Yamazaki H, Yamazaki M, Tachimori Y, Kajiyama Y, Natsugoe S, Fujita H, Mafune K, Tanaka Y, Kelsell DP, Scott CA, Tsuji S, Yachida S, Shibata T, Sugano S, Doki Y, Akiyama T, Aburatani H, Ogawa S, Miyano S, Mori M and Mimori K. Genomic landscape of esophageal squamous cell carcinoma in a Japanese population. Gastroenterology 2016; 150: 1171-1182.
- [39] The Cancer Genome Atlas Research Network. Integrated genomic characterization of oesophageal carcinoma. Nature 2017; 541: 169-175.
- [40] Lam-Himlin DM, Daniels JA, Gayyed MF, Dong J, Maitra A, Pan D, Montgomery EA and Anders RA. The hippo pathway in human upper gastrointestinal dysplasia and carcinoma: a novel oncogenic pathway. Int J Gastrointest Cancer 2006; 37: 103-109.
- [41] Fox JG and Wang TC. Inflammation, atrophy, and gastric cancer. J Clin Invest 2007; 117: 60-69.
- [42] Toh JWT and Wilson RB. Pathways of gastric carcinogenesis, helicobacter pylori virulence

and interactions with antioxidant systems, vitamin C and phytochemicals. Int J Mol Sci 2020; 21: 6451.

- [43] Hu Y, He C, Liu JP, Li NS, Peng C, Yang-Ou YB, Yang XY, Lu NH and Zhu Y. Analysis of key genes and signaling pathways involved in Helicobacter pylori-associated gastric cancer based on The Cancer Genome Atlas database and RNA sequencing data. Helicobacter 2018; 23: e12530.
- [44] Higashi H, Tsutsumi R, Muto S, Sugiyama T, Azuma T, Asaka M and Hatakeyama M. SHP-2 tyrosine phosphatase as an intracellular target of Helicobacter pylori CagA protein. Science 2002; 295: 683-686.
- [45] Miftahussurur M, Yamaoka Y and Graham DY. Helicobacter pylori as an oncogenic pathogen, revisited. Expert Rev Mol Med 2017; 19: e4.
- [46] Li N, Feng Y, Hu Y, He C, Xie C, Ouyang Y, Artim SC, Huang D, Zhu Y, Luo Z, Ge Z and Lu N. Helicobacter pylori CagA promotes epithelial mesenchymal transition in gastric carcinogenesis via triggering oncogenic YAP pathway. J Exp Clin Cancer Res 2018; 37: 280.
- [47] Hu X, Xin Y, Xiao Y and Zhao J. Overexpression of YAP1 is correlated with progression, metastasis and poor prognosis in patients with gastric carcinoma. Pathol Oncol Res 2014; 20: 805-811.
- [48] Da CL, Xin Y, Zhao J and Luo XD. Significance and relationship between Yes-associated protein and survivin expression in gastric carcinoma and precancerous lesions. World J Gastroenterol 2009; 15: 4055-4061.
- [49] Kim E, Ahn B, Oh H, Lee YJ, Lee JH, Lee Y, Kim CH, Chae YS and Kim JY. High Yes-associated protein 1 with concomitant negative LATS1/2 expression is associated with poor prognosis of advanced gastric cancer. Pathology 2019; 51: 261-267.
- [50] Kang W, Tong JH, Chan AW, Lee TL, Lung RW, Leung PP, So KK, Wu K, Fan D, Yu J, Sung JJ and To KF. Yes-associated protein 1 exhibits oncogenic property in gastric cancer and its nuclear accumulation associates with poor prognosis. Clin Cancer Res 2011; 17: 2130-2139.
- [51] Ajani JA, Xu Y, Huo L, Wang R, Li Y, Wang Y, Pizzi MP, Scott A, Harada K, Ma L, Yao X, Jin J, Zhao W, Dong X, Badgwell BD, Shanbhag N, Tatlonghari G, Estrella JS, Roy-Chowdhuri S, Kobayashi M, Vykoukal JV, Hanash SM, Calin GA, Peng G, Lee JS, Johnson RL, Wang Z, Wang L and Song S. YAP1 mediates gastric adenocarcinoma peritoneal metastases that are attenuated by YAP1 inhibition. Gut 2021; 70: 55-66.
- [52] Choi W, Kim J, Park J, Lee DH, Hwang D, Kim JH, Ashktorab H, Smoot D, Kim SY, Choi C, Koh

GY and Lim DS. YAP/TAZ Initiates Gastric Tumorigenesis via Upregulation of MYC. Cancer Res 2018; 78: 3306-3320.

- [53] Jiao S, Wang H, Shi Z, Dong A, Zhang W, Song X, He F, Wang Y, Zhang Z, Wang W, Wang X, Guo T, Li P, Zhao Y, Ji H, Zhang L and Zhou Z. A peptide mimicking VGLL4 function acts as a YAP antagonist therapy against gastric cancer. Cancer Cell 2014; 25: 166-180.
- [54] Melucci E, Casini B, Ronchetti L, Pizzuti L, Sperati F, Pallocca M, De Nicola F, Goeman F, Gallo E, Amoreo CA, Sergi D, Terrenato I, Vici P, Di Lauro L, Diodoro MG, Pescarmona E, Barba M, Mazzotta M, Mottolese M, Fanciulli M, Ciliberto G, De Maria R, Buglioni S and Maugeri-Saccà M. Expression of the Hippo transducer TAZ in association with WNT pathway mutations impacts survival outcomes in advanced gastric cancer patients treated with first-line chemotherapy. J Transl Med 2018; 16: 22.
- [55] Pallocca M, Goeman F, De Nicola F, Melucci E, Sperati F, Terrenato I, Pizzuti L, Casini B, Gallo E, Amoreo CA, Vici P, Di Lauro L, Buglioni S, Diodoro MG, Pescarmona E, Mazzotta M, Barba M, Fanciulli M, De Maria R, Ciliberto G and Maugeri-Saccà M. Coexisting YAP expression and TP53 missense mutations delineates a molecular scenario unexpectedly associated with better survival outcomes in advanced gastric cancer. J Transl Med 2018; 16: 247.
- [56] Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer. Nature 2012; 487: 330-337.
- [57] Fodde R, Smits R and Clevers H. APC, signal transduction and genetic instability in colorectal cancer. Nat Rev Cancer 2001; 1: 55-67.
- [58] Li N, Lu N and Xie C. The Hippo and Wnt signalling pathways: crosstalk during neoplastic progression in gastrointestinal tissue. FEBS J 2019; 286: 3745-3756.
- [59] Rosenbluh J, Nijhawan D, Cox AG, Li X, Neal JT, Schafer EJ, Zack TI, Wang X, Tsherniak A, Schinzel AC, Shao DD, Schumacher SE, Weir BA, Vazquez F, Cowley GS, Root DE, Mesirov JP, Beroukhim R, Kuo CJ, Goessling W and Hahn WC. β-Catenin-driven cancers require a YAP1 transcriptional complex for survival and tumorigenesis. Cell 2012; 151: 1457-1473.
- [60] Varelas X, Miller BW, Sopko R, Song S, Gregorieff A, Fellouse FA, Sakuma R, Pawson T, Hunziker W, McNeill H, Wrana JL and Attisano L. The Hippo pathway regulates Wnt/betacatenin signaling. Dev Cell 2010; 18: 579-591.
- [61] Azzolin L, Panciera T, Soligo S, Enzo E, Bicciato S, Dupont S, Bresolin S, Frasson C, Basso G, Guzzardo V, Fassina A, Cordenonsi M and Piccolo S. YAP/TAZ incorporation in the β -catenin destruction complex orchestrates the Wnt response. Cell 2014; 158: 157-170.

- [62] Barry ER, Morikawa T, Butler BL, Shrestha K, de la Rosa R, Yan KS, Fuchs CS, Magness ST, Smits R, Ogino S, Kuo CJ and Camargo FD. Restriction of intestinal stem cell expansion and the regenerative response by YAP. Nature 2013; 493: 106-110.
- [64] Gregorieff A, Liu Y, Inanlou MR, Khomchuk Y and Wrana JL. Yap-dependent reprogramming of Lgr5(+) stem cells drives intestinal regeneration and cancer. Nature 2015; 526: 715-718.
- [65] Cai J, Zhang N, Zheng Y, de Wilde RF, Maitra A and Pan D. The Hippo signaling pathway restricts the oncogenic potential of an intestinal regeneration program. Genes Dev 2010; 24: 2383-2388.
- [66] Steinhardt AA, Gayyed MF, Klein AP, Dong J, Maitra A, Pan D, Montgomery EA and Anders RA. Expression of Yes-associated protein in common solid tumors. Hum Pathol 2008; 39: 1582-1589.
- [67] Wang Y, Xie C, Li Q, Xu K and Wang E. Clinical and prognostic significance of Yes-associated protein in colorectal cancer. Tumour Biol 2013; 34: 2169-2174.
- [68] Xu Z, Wang H, Gao L, Zhang H and Wang X. YAP levels combined with plasma CEA levels are prognostic biomarkers for early-clinical-stage patients of colorectal cancer. Biomed Res Int 2019; 2019: 2170830.
- [69] Song R, Gu D, Zhang L, Zhang X, Yu B, Liu B and Xie J. Functional significance of Hippo/YAP signaling for drug resistance in colorectal cancer. Mol Carcinog 2018; 57: 1608-1615.
- [70] Kim JY, Kim EK, Lee WM, Hong YO and Lee H. VGLL4 with low YAP expression is associated with favorable prognosis in colorectal cancer. Apmis 2020; 128: 543-551.
- [71] Ou C, Sun Z, Li S, Li G, Li X and Ma J. Dual roles of yes-associated protein (YAP) in colorectal cancer. Oncotarget 2017; 8: 75727-75741.
- [72] Lapi E, Di Agostino S, Donzelli S, Gal H, Domany E, Rechavi G, Pandolfi PP, Givol D, Strano S, Lu X and Blandino G. PML, YAP, and p73 are components of a proapoptotic autoregulatory feedback loop. Mol Cell 2008; 32: 803-814.
- [73] Yuan M, Tomlinson V, Lara R, Holliday D, Chelala C, Harada T, Gangeswaran R, Manson-Bishop C, Smith P, Danovi SA, Pardo O, Crook T, Mein CA, Lemoine NR, Jones LJ and Basu S. Yes-associated protein (YAP) functions as a tumor suppressor in breast. Cell Death Differ 2008; 15: 1752-1759.
- [74] Strano S, Monti O, Pediconi N, Baccarini A, Fontemaggi G, Lapi E, Mantovani F, Damalas A, Citro G, Sacchi A, Del Sal G, Levrero M and

Blandino G. The transcriptional coactivator Yes-associated protein drives p73 gene-target specificity in response to DNA Damage. Mol Cell 2005; 18: 447-459.

- [75] Zhang S, Wei Q, Yang Y, Qin H, Li X, Cai S and Ma Y. Loss of Yes-associated Protein Represents an Aggressive Subtype of Colorectal Cancer. J Cancer 2019; 10: 689-696.
- [76] Yin F, Xie H, Lai J, Chen Y, Dong J, Zhang X and Liu X. Double negativity for expression of YAP1 and CDX2 defines an aggressive type of colitisassociated cancer. Anticancer Res 2020; 40: 5411-5416.
- [77] Pelosi E, Castelli G and Testa U. Pancreatic cancer: molecular characterization, clonal evolution and cancer stem cells. Biomedicines 2017; 5: 65.
- [78] Shao DD, Xue W, Krall EB, Bhutkar A, Piccioni F, Wang X, Schinzel AC, Sood S, Rosenbluh J, Kim JW, Zwang Y, Roberts TM, Root DE, Jacks T and Hahn WC. KRAS and YAP1 converge to regulate EMT and tumor survival. Cell 2014; 158: 171-184.
- [79] Kapoor A, Yao W, Ying H, Hua S, Liewen A, Wang Q, Zhong Y, Wu CJ, Sadanandam A, Hu B, Chang Q, Chu GC, Al-Khalil R, Jiang S, Xia H, Fletcher-Sananikone E, Lim C, Horwitz GI, Viale A, Pettazzoni P, Sanchez N, Wang H, Protopopov A, Zhang J, Heffernan T, Johnson RL, Chin L, Wang YA, Draetta G and DePinho RA. Yap1 activation enables bypass of oncogenic Kras addiction in pancreatic cancer. Cell 2014; 158: 185-197.
- [80] Gruber R, Panayiotou R, Nye E, Spencer-Dene B, Stamp G and Behrens A. YAP1 and TAZ control pancreatic cancer initiation in mice by direct up-regulation of JAK-STAT3 signaling. Gastroenterology 2016; 151: 526-539.
- [81] Yang S, Zhang L, Purohit V, Shukla SK, Chen X, Yu F, Fu K, Chen Y, Solheim J, Singh PK, Song W and Dong J. Active YAP promotes pancreatic cancer cell motility, invasion and tumorigenesis in a mitotic phosphorylation-dependent manner through LPAR3. Oncotarget 2015; 6: 36019-36031.
- [82] Diep CH, Zucker KM, Hostetter G, Watanabe A, Hu C, Munoz RM, Von Hoff DD and Han H. Down-regulation of Yes Associated Protein 1 expression reduces cell proliferation and clonogenicity of pancreatic cancer cells. PLoS One 2012; 7: e32783.
- [83] Salcedo Allende MT, Zeron-Medina J, Hernandez J, Macarulla T, Balsells J, Merino X, Allende H, Tabernero J and Ramon Y Cajal S. Overexpression of yes associated protein 1, an independent prognostic marker in patients with pancreatic ductal adenocarcinoma, correlated with liver metastasis and poor prognosis. Pancreas 2017; 46: 913-920.

- [84] Zhou Q, Bauden M, Andersson R, Hu D, Marko-Varga G, Xu J, Sasor A, Dai H, Pawłowski K, Said Hilmersson K, Chen X and Ansari D. YAP1 is an independent prognostic marker in pancreatic cancer and associated with extracellular matrix remodeling. J Transl Med 2020; 18: 77.
- [85] Yoo W, Lee J, Jun E, Noh KH, Lee S, Jung D, Jung KH, Kim JS, Park YY, Kim SC and Kim S. The YAP1-NMU axis is associated with pancreatic cancer progression and poor outcome: identification of a novel diagnostic biomarker and therapeutic target. Cancers (Basel) 2019; 11: 1477.
- [86] Murakami S, Shahbazian D, Surana R, Zhang W, Chen H, Graham GT, White SM, Weiner LM and Yi C. Yes-associated protein mediates immune reprogramming in pancreatic ductal adenocarcinoma. Oncogene 2017; 36: 1232-1244.
- [87] Rozengurt E, Sinnett-Smith J and Eibl G. Yesassociated protein (YAP) in pancreatic cancer: at the epicenter of a targetable signaling network associated with patient survival. Signal Transduct Target Ther 2018; 3: 11.
- [88] Balogh J, Victor D 3rd, Asham EH, Burroughs SG, Boktour M, Saharia A, Li X, Ghobrial RM and Monsour HP Jr. Hepatocellular carcinoma: a review. J Hepatocell Carcinoma 2016; 3: 41-53.
- [89] Zhang N, Bai H, David KK, Dong J, Zheng Y, Cai J, Giovannini M, Liu P, Anders RA and Pan D. The Merlin/NF2 tumor suppressor functions through the YAP oncoprotein to regulate tissue homeostasis in mammals. Dev Cell 2010; 19: 27-38.
- [90] Song H, Mak KK, Topol L, Yun K, Hu J, Garrett L, Chen Y, Park O, Chang J, Simpson RM, Wang CY, Gao B, Jiang J and Yang Y. Mammalian Mst1 and Mst2 kinases play essential roles in organ size control and tumor suppression. Proc Natl Acad Sci U S A 2010; 107: 1431-1436.
- [91] Zhou D, Conrad C, Xia F, Park JS, Payer B, Yin Y, Lauwers GY, Thasler W, Lee JT, Avruch J and Bardeesy N. Mst1 and Mst2 maintain hepatocyte quiescence and suppress hepatocellular carcinoma development through inactivation of the Yap1 oncogene. Cancer Cell 2009; 16: 425-438.
- [92] Lu L, Li Y, Kim SM, Bossuyt W, Liu P, Qiu Q, Wang Y, Halder G, Finegold MJ, Lee JS and Johnson RL. Hippo signaling is a potent in vivo growth and tumor suppressor pathway in the mammalian liver. Proc Natl Acad Sci U S A 2010; 107: 1437-1442.
- [93] Lee KP, Lee JH, Kim TS, Kim TH, Park HD, Byun JS, Kim MC, Jeong WI, Calvisi DF, Kim JM and Lim DS. The Hippo-Salvador pathway restrains

hepatic oval cell proliferation, liver size, and liver tumorigenesis. Proc Natl Acad Sci U S A 2010; 107: 8248-8253.

- [94] Nishio M, Sugimachi K, Goto H, Wang J, Morikawa T, Miyachi Y, Takano Y, Hikasa H, Itoh T, Suzuki SO, Kurihara H, Aishima S, Leask A, Sasaki T, Nakano T, Nishina H, Nishikawa Y, Sekido Y, Nakao K, Shin-Ya K, Mimori K and Suzuki A. Dysregulated YAP1/TAZ and TGF-β signaling mediate hepatocarcinogenesis in Mob1a/1bdeficient mice. Proc Natl Acad Sci U S A 2016; 113: E71-80.
- [95] Yi J, Lu L, Yanger K, Wang W, Sohn BH, Stanger BZ, Zhang M, Martin JF, Ajani JA, Chen J, Lee JS, Song S and Johnson RL. Large tumor suppressor homologs 1 and 2 regulate mouse liver progenitor cell proliferation and maturation through antagonism of the coactivators YAP and TAZ. Hepatology 2016; 64: 1757-1772.
- [96] Dong J, Feldmann G, Huang J, Wu S, Zhang N, Comerford SA, Gayyed MF, Anders RA, Maitra A and Pan D. Elucidation of a universal size-control mechanism in Drosophila and mammals. Cell 2007; 130: 1120-1133.
- [97] Yimlamai D, Christodoulou C, Galli GG, Yanger K, Pepe-Mooney B, Gurung B, Shrestha K, Cahan P, Stanger BZ and Camargo FD. Hippo pathway activity influences liver cell fate. Cell 2014; 157: 1324-1338.
- [98] Moya IM, Castaldo SA, Van den Mooter L, Soheily S, Sansores-Garcia L, Jacobs J, Mannaerts I, Xie J, Verboven E, Hillen H, Algueró-Nadal A, Karaman R, Van Haele M, Kowalczyk W, De Waegeneer M, Verhulst S, Karras P, van Huffel L, Zender L, Marine JC, Roskams T, Johnson R, Aerts S, van Grunsven LA and Halder G. Peritumoral activation of the Hippo pathway effectors YAP and TAZ suppresses liver cancer in mice. Science 2019; 366: 1029-1034.
- [99] Chen CL, Schroeder MC, Kango-Singh M, Tao C and Halder G. Tumor suppression by cell competition through regulation of the Hippo pathway. Proc Natl Acad Sci U S A 2012; 109: 484-489.
- [100] Han SX, Bai E, Jin GH, He CC, Guo XJ, Wang LJ, Li M, Ying X and Zhu Q. Expression and clinical significance of YAP, TAZ, and AREG in hepatocellular carcinoma. J Immunol Res 2014; 2014: 261365.
- [101] Xu MZ, Yao TJ, Lee NP, Ng IO, Chan YT, Zender L, Lowe SW, Poon RT and Luk JM. Yes-associated protein is an independent prognostic marker in hepatocellular carcinoma. Cancer 2009; 115: 4576-4585.
- [102] Berasain C, Castillo J, Perugorría MJ, Prieto J and Avila MA. Amphiregulin: a new growth factor in hepatocarcinogenesis. Cancer Lett 2007; 254: 30-41.

- [103] Awad AE, Ebrahim MA, Eissa LA and El-Shishtawy MM. Dickkopf-1 and amphiregulin as novel biomarkers and potential therapeutic targets in hepatocellular carcinoma. Int J Hematol Oncol Stem Cell Res 2019; 13: 153-163.
- [104] Castillo J, Goñi S, Latasa MU, Perugorría MJ, Calvo A, Muntané J, Bioulac-Sage P, Balabaud C, Prieto J, Avila MA and Berasain C. Amphiregulin induces the alternative splicing of p73 into its oncogenic isoform DeltaEx2p73 in human hepatocellular tumors. Gastroenterology 2009; 137: 1805-1815, e1801-1804.
- [105] Marquard S, Thomann S, Weiler SME, Bissinger M, Lutz T, Sticht C, Tóth M, de la Torre C, Gretz N, Straub BK, Marquardt J, Schirmacher P and Breuhahn K. Yes-associated protein (YAP) induces a secretome phenotype and transcriptionally regulates plasminogen activator Inhibitor-1 (PAI-1) expression in hepatocarcinogenesis. Cell Commun Signal 2020; 18: 166.
- [106] Sohn BH, Shim JJ, Kim SB, Jang KY, Kim SM, Kim JH, Hwang JE, Jang HJ, Lee HS, Kim SC, Jeong W, Kim SS, Park ES, Heo J, Kim YJ, Kim DG, Leem SH, Kaseb A, Hassan MM, Cha M, Chu IS, Johnson RL, Park YY and Lee JS. Inactivation of hippo pathway is significantly associated with poor prognosis in hepatocellular carcinoma. Clin Cancer Res 2016; 22: 1256-1264.
- [107] Errani C, Zhang L, Sung YS, Hajdu M, Singer S, Maki RG, Healey JH and Antonescu CR. A novel WWTR1-CAMTA1 gene fusion is a consistent abnormality in epithelioid hemangioendothelioma of different anatomic sites. Genes Chromosomes Cancer 2011; 50: 644-653.
- [108] Tanas MR, Sboner A, Oliveira AM, Erickson-Johnson MR, Hespelt J, Hanwright PJ, Flanagan J, Luo Y, Fenwick K, Natrajan R, Mitsopoulos C, Zvelebil M, Hoch BL, Weiss SW, Debiec-Rychter M, Sciot R, West RB, Lazar AJ, Ashworth A, Reis-Filho JS, Lord CJ, Gerstein MB, Rubin MA and Rubin BP. Identification of a disease-defining gene fusion in epithelioid hemangioendothelioma. Sci Transl Med 2011; 3: 98ra82.
- [109] Antonescu CR, Le Loarer F, Mosquera JM, Sboner A, Zhang L, Chen CL, Chen HW, Pathan N, Krausz T, Dickson BC, Weinreb I, Rubin MA, Hameed M and Fletcher CD. Novel YAP1-TFE3 fusion defines a distinct subset of epithelioid hemangioendothelioma. Genes Chromosomes Cancer 2013; 52: 775-784.
- [110] Doyle LA, Fletcher CD and Hornick JL. Nuclear expression of CAMTA1 distinguishes epithelioid hemangioendothelioma from histologic mimics. Am J Surg Pathol 2016; 40: 94-102.
- [111] Driskill JH, Zheng Y, Wu BK, Wang L, Cai J, Rakheja D, Dellinger M and Pan D. WWTR1(TAZ)-CAMTA1 reprograms endothelial cells to drive

epithelioid hemangioendothelioma. Genes Dev 2021; 35: 495-511.

- [112] Seavey CN, Pobbati AV, Hallett A, Ma S, Reynolds JP, Kanai R, Lamar JM and Rubin BP. WWTR1(TAZ)-CAMTA1 gene fusion is sufficient to dysregulate YAP/TAZ signaling and drive epithelioid hemangioendothelioma tumorigenesis. Genes Dev 2021; 35: 512-527.
- [113] Liu-Chittenden Y, Huang B, Shim JS, Chen Q, Lee SJ, Anders RA, Liu JO and Pan D. Genetic and pharmacological disruption of the TEAD-YAP complex suppresses the oncogenic activity of YAP. Genes Dev 2012; 26: 1300-1305.
- [114] Song S, Xie M, Scott AW, Jin J, Ma L, Dong X, Skinner HD, Johnson RL, Ding S and Ajani JA. A novel YAP1 inhibitor targets CSC-enriched radiation-resistant cells and exerts strong antitumor activity in esophageal adenocarcinoma. Mol Cancer Ther 2018; 17: 443-454.
- [115] Yu FX, Zhao B, Panupinthu N, Jewell JL, Lian I, Wang LH, Zhao J, Yuan H, Tumaneng K, Li H, Fu XD, Mills GB and Guan KL. Regulation of the Hippo-YAP pathway by G-protein-coupled receptor signaling. Cell 2012; 150: 780-791.
- [116] Tocci P, Blandino G and Bagnato A. YAP and endothelin-1 signaling: an emerging alliance in cancer. J Exp Clin Cancer Res 2021; 40: 27.
- [117] Zhang J, Ji JY, Yu M, Overholtzer M, Smolen GA, Wang R, Brugge JS, Dyson NJ and Haber DA. YAP-dependent induction of amphiregulin identifies a non-cell-autonomous component of the Hippo pathway. Nat Cell Biol 2009; 11: 1444-1450.

- [118] He C, Mao D, Hua G, Lv X, Chen X, Angeletti PC, Dong J, Remmenga SW, Rodabaugh KJ, Zhou J, Lambert PF, Yang P, Davis JS and Wang C. The Hippo/YAP pathway interacts with EGFR signaling and HPV oncoproteins to regulate cervical cancer progression. EMBO Mol Med 2015; 7: 1426-1449.
- [119] Azad T, Janse van Rensburg HJ, Lightbody ED, Neveu B, Champagne A, Ghaffari A, Kay VR, Hao Y, Shen H, Yeung B, Croy BA, Guan KL, Pouliot F, Zhang J, Nicol CJB and Yang X. A LATS biosensor screen identifies VEGFR as a regulator of the Hippo pathway in angiogenesis. Nat Commun 2018; 9: 1061.
- [120] Wang X, Freire Valls A, Schermann G, Shen Y, Moya IM, Castro L, Urban S, Solecki GM, Winkler F, Riedemann L, Jain RK, Mazzone M, Schmidt T, Fischer T, Halder G and Ruiz de Almodóvar C. YAP/TAZ orchestrate VEGF signaling during developmental angiogenesis. Dev Cell 2017; 42: 462-478, e467.
- [121] Moroishi T, Hayashi T, Pan WW, Fujita Y, Holt MV, Qin J, Carson DA and Guan KL. The hippo pathway kinases LATS1/2 suppress cancer immunity. Cell 2016; 167: 1525-1539, e1517.
- [122] Fan F, He Z, Kong LL, Chen Q, Yuan Q, Zhang S, Ye J, Liu H, Sun X, Geng J, Yuan L, Hong L, Xiao C, Zhang W, Sun X, Li Y, Wang P, Huang L, Wu X, Ji Z, Wu Q, Xia NS, Gray NS, Chen L, Yun CH, Deng X and Zhou D. Pharmacological targeting of kinases MST1 and MST2 augments tissue repair and regeneration. Sci Transl Med 2016; 8: 352ra108.