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LRP10 variants in progressive supranuclear palsy

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Abstract

The aim of this study was to explore whether variants in *LRP10*, recently associated with Parkinson's disease and dementia with Lewy bodies, are observed in 2 large cohorts (discovery and validation cohort) of patients with progressive supranuclear palsy (PSP). A total of 950 patients with PSP were enrolled: 246 patients with PSP (n = 85 possible (35%), n = 128 probable (52%)), n

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Disclosure statement

The authors declare that there are no conflicts of interest associated with this work.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <https://doi.org/10.1016/j.neurobiolaging.2020.04.016>.

= 33 definite (13%)) in the discovery cohort and 704 patients with definite PSP in the validation cohort. Sanger sequencing of all *LRP10* exons and exon-intron boundaries was performed in the discovery cohort, and whole-exome sequencing was performed in the validation cohort. Two patients from the discovery cohort and 8 patients from the validation cohort carried a rare, heterozygous, and possibly pathogenic *LRP10* variant (p.Gly326Asp, p.Asp389Asn, and p.Arg158His, p.Cys220Tyr, p.Thr278Ala, p.Gly306Asp, p.Glu486Asp, p.Arg554*, p.Arg661Cys). In conclusion, possibly pathogenic *LRP10* variants occur in a small fraction of patients with PSP and may be overrepresented in these patients compared with controls. This suggests that possibly pathogenic *LRP10* variants may play a role in the development of PSP.

Keywords

Genetics; *LRP10*; Rare variants; Progressive supranuclear palsy

1. Introduction

Progressive supranuclear palsy (PSP) is an adult-onset, progressive neurodegenerative disorder clinically characterized by parkinsonism, vertical supranuclear gaze palsy, and postural instability with falls. Other PSP features include frontal lobe and bulbar dysfunction, and cognitive decline (Litvan et al., 1996b; Respondek et al., 2013). The clinical presentation of PSP is heterogeneous, and 10 different clinical phenotypes have been described in patients with PSP neuropathology (Höglinger et al., 2017). PSP brain pathology includes neurofibrillary tangles, neutrophil threads, tufted astrocytes, neuronal loss, and gliosis in multiple subcortical areas and other regions (Hauw et al., 1994).

PSP is usually considered a sporadic tauopathy of unknown etiology (Im et al., 2015). However, rare familial forms have been reported (Donker Kaat et al., 2009; Fujioka et al., 2014). Mutations in the *microtubule-associated protein tau (MAPT)* gene have been reported as the likely disease cause in a few pathologically confirmed patients with PSP (Fujioka et al., 2015; Poorkaj et al., 2002). Furthermore, genome-wide association studies have shown associations between PSP and the *MAPT*, *syntaxin-6 (STX6)*, *myelin-associated oligodendrocyte basic protein (MOBP)*, and *eukaryotic translation initiation factor 2-alpha kinase (EIF2AK)* genes, modulating the risk of developing PSP (Chen et al., 2018; Sanchez-Contreras et al., 2018). Mutations in the *leucine-rich repeat kinase 2 (LRRK2)* gene have also been implicated in a small number of PSP cases (Ross et al., 2006; Sanchez-Contreras et al., 2017) and are the most common genetic cause of Parkinson's disease (PD) (Healy et al., 2008). Interestingly, pleomorphic neuropathology has been observed in PD patients with *LRRK2* mutations, ranging from typical alpha-synuclein-positive pathology (seen in most cases) to PSP-like pathology (Zimprich et al., 2004). Indeed in the original Japanese family, which nominated the linkage region for *LRRK2*, the affected members presented with tauopathy (Ujiiie et al., 2012). Because of these genetic and pathological overlaps between PD and PSP, we hypothesized that variants in the *low-density lipoprotein receptor related protein 10 (LRP10)* gene, recently associated with PD and dementia with Lewy bodies (Quadri et al., 2018; Vergouw et al., 2019), might also be implicated in PSP. LRP10 is a surface protein, whose function is largely elusive. However, some studies have suggested a

role of LRP10 in ligand trafficking between the trans-Golgi network, endosomes, and the plasma membrane. Furthermore, LRP10 has been linked to the metabolism of amyloid- β and α -synuclein (Brodeur et al., 2012; von Einem et al., 2015, 2017).

The aim of this study is to explore whether possibly pathogenic variants in *LRP10* are observed in a large Dutch cohort of patients with PSP (discovery cohort) using Sanger sequencing. In addition, we try to validate our findings in a large cohort of patients with PSP (validation cohort) from the United States and Europe using whole-exome sequencing (WES).

2. Methods

2.1. Subjects

Patients with PSP from 2 large cohorts were enrolled in this study. PSP was diagnosed according to the criteria of the National Institute for Neurological Disorders and Stroke/Society for PSP (NINDS-SPSP) (Litvan et al., 1996a). The discovery cohort consisted of 246 patients with PSP enrolled from a large Dutch cohort (Donker Kaat et al., 2007), consecutively collected between 2003 and 2012 (Table 1). Patients were ascertained at the outpatient clinic of the Erasmus Medical Center Rotterdam, at home or at nursing homes. At inclusion, information about patients' medical and family history and current medical status was collected. Furthermore, neurological examination was performed, and a blood sample was collected. The validation cohort consisted of 704 neuropathologically confirmed patients with PSP enrolled from a large cohort of patients from the United States and Europe (Höglinger et al., 2011) (Table 1). These patients were identified from brain banks, research hospitals, and neuropathologists. The study was approved by the relevant Institutional Ethical Authorities, and all participants or legal representatives signed informed consent.

2.2. Genetic analyses

2.2.1 . Genetic analyses in the 2 cohorts—Genomic DNA was isolated from blood in the discovery cohort and from brain tissue in the validation cohort using standard methods. Sanger sequencing was performed for the entire open reading frame and exon-intron boundaries of *LRP10* in the discovery cohort (protocol reported by Vergouw et al., 2019). WES was performed in the validation cohort (Supplementary Information). Possibly pathogenic *LRP10* variants identified by WES in the validation cohort were validated by Sanger sequencing (Supplementary Information). We considered variants as possibly pathogenic according to the following criteria: (1) heterozygous state; (2) rarity, defined as a frequency <0.1% in the Genome Aggregation Database (GnomAD v2.1); (3) exonic location and non-synonymous, or predicted to affect splicing; and (4) predicted as pathogenic by at least 5 of 11 in silico programs (Supplementary Information).

2.2.2. Additional genetic analyses in possibly pathogenic LRP10 variant carriers in the discovery cohort

WES, multiple ligation-dependent probe amplification (P051-Parkinson mix 1), and *C9orf72* repeat expansion analysis were performed in patients who carried possibly pathogenic *LRP10* variants to exclude possibly pathogenic variants in other known genes causing

parkinsonism or dementia (Supplementary Table 1). The presence of possibly pathogenic variants in known genes causing parkinsonism or dementia in possibly pathogenic *LRP10* variant carriers decreases the chance of the *LRP10* variant to be truly pathogenic. WES and multiple ligation-dependent probe amplification were performed as reported previously by Vergouw et al. (2019). Details of the methods of the *C9orf72* repeat expansion analysis can be found in the Supplementary Information.

3. Results

3.1. Demographic and clinical characteristics of the 2 cohorts

The discovery cohort consisted of 85 (35%) patients with possible PSP, 128 (52%) with probable PSP (52%), and 33 (13%) with definite PSP. The mean disease onset age in this cohort was 65.8 ± 7.5 years and 52% of patients were male; 29% of patients had at least one first-degree relative and 9% had at least one second-degree relative with a neurodegenerative disease. The validation cohort consisted of 704 patients with definite PSP. The mean disease-onset age in this cohort was 68.1 ± 8.4 years (data only available in $n = 476$), and 54% of patients were male (Table 1).

3.2. Genetic findings

Two possibly pathogenic *LRP10* variants were detected in the discovery cohort, each in single patients (p.Gly326Asp and p.Asp389Asn). In the validation cohort, 7 possibly pathogenic *LRP10* variants were detected in 8 patients (p.Arg158His, p.Cys220Tyr, p.Thr278Ala, p.Gly306Asp, p.Glu486Asp, p.Arg554*, and p.Arg661Cys; see Table 2 and Supplementary Table S2 for specifications). Supplementary Figure S1A shows the *LRP10* gene structure with the location of the identified variants, and Supplementary Figure S1B shows the *LRP10* protein structure with the location of the amino acid changes. Other variants in *LRP10* which did not fulfill the criteria for possible pathogenicity, as described in Section 2.2.1., are depicted in Supplementary Table S3. Additional WES analysis (average depth of $>170\times$ with 99% of the target region covered $>20\times$) in the possibly pathogenic *LRP10* variant carriers from the discovery cohort revealed a heterozygous *VPS13C* variant (p.Gln2546*, absent in GnomAD v2.1) in 1 patient (Supplementary Table S4). No other mutations in genes causing parkinsonism or dementia were found.

3.3. Clinical information of possibly pathogenic *LRP10* variant carriers

An overview of the clinical information of the possibly pathogenic *LRP10* variant carriers is shown in Table 2. Patient 1 from the discovery cohort (*LRP10* p.Asp389Asn variant) experienced falls from the age of 61 years, followed by swallowing problems. At the age of 63 years, a mild downward vertical supranuclear gaze palsy, dysarthric speech, reduced arm swing, palatal tremor, and impaired balance, but no clear ataxia, were observed. He had a favorable response to levodopa. At neuropsychological examination, deficits were observed in attention, concentration, and executive functioning. Furthermore, mild memory and naming problems were seen. Brain MRI showed mild parieto-occipital and cerebellar atrophy and hypertrophy of the olivary nuclei. The patient died at the age of 66 years. Family history was negative for parkinsonism, dementia, or motor neuron disease. Brain autopsy was not performed. This patient was diagnosed with probable PSP during life

according to the NINDS-SPSP criteria (Litvan et al., 1996a) and can retrospectively be classified as probable PSP with Richardson's syndrome according to the MDS criteria (Höglinger et al., 2017). Patient 2 from the discovery cohort (*LRP10* p.Gly326Asp variant) experienced tremor of the right leg from the age of 55 years, followed by falls, rigidity, swallowing, speech, and memory problems from the age of 60 years. At the age of 64 years, vertical supranuclear gaze palsy, bradykinesia, intermittent rest tremor of arms and legs, and balance problems were observed. She had a favorable response to levodopa. Neuropsychological examination showed severe deficits, especially with frontal subcortical and language problems. Brain MRI was unremarkable. The patient died at the age of 65 years. Family history was negative for parkinsonism, dementia, or motor neuron disease. Brain autopsy was not performed. This patient was diagnosed with possible PSP during life according to the NINDS-SPSP criteria (Litvan et al., 1996a) and can retrospectively be classified as probable PSP with predominant parkinsonism according to the MDS criteria (Höglinger et al., 2017).

4. Discussion

In this study, we explored the presence of *LRP10* variants in 2 cohorts with a total of 950 PSP patients (discovery cohort n = 246, validation cohort n = 704). The PSP diagnosis was pathologically confirmed in 78% of these patients. Two possibly pathogenic *LRP10* variants (p.Gly326Asp and p.Asp389Asn) were identified in 2 patients from the discovery cohort, and 7 possibly pathogenic *LRP10* variants (p.Arg158His, p.Cys220Tyr, p.Thr278Ala, p.Gly306Asp, p.Glu486Asp, p.Arg554*, and p.Arg661Cys) were identified in 8 patients from the validation cohort. These variants are very rare, are predicted to be pathogenic by 5 in silico programs, and are mostly located in *LRP10* exon 5, where other probably pathogenic variants were previously found (Quadri et al., 2018). Interestingly, the frequency of possibly pathogenic *LRP10* variants is significantly higher in the validation cohort (8/1408 alleles = 0.6%) compared with a previous published control cohort of patients with abdominal aneurysms (Quadri et al., 2018) (1/1248 alleles = 0.08%; Fisher's exact test p-value 0.04). In addition, the p.Gly306Asp variant has been identified previously in 2 of 2835 patients with PD and 1 of 5343 controls (Kia et al., 2018), the p.Gly326Asp variant in 1 of 264 patients with multiple system atrophy and in no controls (Pihlström et al., 2018), and the p.Glu486Asp variant in 3 of 2835 patients with PD and 1 of 111 patients with dementia with Lewy bodies compared with none in 5343 and 233 controls, respectively (Kia et al., 2018). The p.Arg158His, p.Arg554*, and p.Arg661Cys variants have previously been identified in single controls (Kia et al., 2018; Guerreiro et al., 2018; Pihlström et al., 2018; Supplementary Table S5).

Both patients from the discovery cohort displayed uncommon PSP clinical features. Patient 1 had a palatal tremor, inferior olivary hypertrophy, and cerebellar atrophy. Inferior olivary hypertrophy is observed in 1.5% of pathologically confirmed patients with PSP (Katsuse and Dickson, 2004), but associated palatal tremor is very rare in PSP (Katsuse and Dickson, 2004; Suyama et al., 1997). The syndrome of progressive ataxia and palatal tremor (Mongin et al., 2016) may retrospectively also be considered in patient 1, yet the clinical phenotype is most consistent with PSP. An uncommon feature in patient 2 was the presence of an isolated tremor of the right leg in the first 5 years of the disease. Unfortunately, autopsy studies were

not performed in these patients, and therefore, the diagnosis could not be verified at the pathological level. The absence of a family history of PSP or other neurodegenerative disorders in these 2 patients would be compatible with an incomplete penetrance or a de novo occurrence of the *LRP10* variants.

Of note, a *VPS13C* variant (p.Gln2546*) was observed in one *LRP10* variant carrier. Mutations in *VPS13C* are associated with autosomal recessive forms of early-onset parkinsonism (Lesage et al., 2016). In our patient, the variant was found in the heterozygous state and is therefore most likely an incidental finding.

Strengths of this study are the large sample size of the 2 PSP cohorts, the validation of our findings in an independent cohort, and the high percentage of neuropathologically confirmed patients with PSP. Limitations are the lack of screening for *LRP10* genomic deletions of multiplications (not detectable by Sanger methods).

In conclusion, this is the first study of *LRP10* in 2 large PSP cohorts. We showed that rare, possibly pathogenic *LRP10* variants occur in a small but substantial fraction of patients with PSP. Furthermore, possibly pathogenic *LRP10* variants may be overrepresented in patients with PSP compared with controls and may therefore play a role in disease pathogenesis. Further studies are warranted to replicate our findings and to study which molecular mechanisms underlie the possible association between *LRP10* and PSP.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

- Brodeur J, Theriault C, Lessard-Beaudoin M, Marcil A, Dahan S, Lavoie C, 2012. LDLR-related protein 10 (LRP10) regulates amyloid precursor protein (APP) trafficking and processing: evidence for a role in Alzheimer's disease. *Mol. Neurodegener.* 7, 31. [PubMed: 22734645]
- Chen JA, Chen Z, Won H, Huang AY, Lowe JK, Wojta K, Yokoyama JS, Bensimon G, Leigh PN, Payan C, Shatunov A, Jones AR, Lewis CM, Deloukas P, Amouyel P, Tzourio C, Dartigues JF, Ludolph A, Boxer AL, Bronstein JM, Al-Chalabi A, Geschwind DH, Coppola G, 2018. Joint genome-wide association study of progressive supranuclear palsy identifies novel susceptibility loci and genetic correlation to neurodegenerative diseases. *Mol. Neurodegener.* 13, 41. [PubMed: 30089514]
- Donker Kaat L, Boon AJ, Kamphorst W, Ravid R, Duivenvoorden HJ, van Swieten JC, 2007. Frontal presentation in progressive supranuclear palsy. *Neurology* 69, 723–729. [PubMed: 17709703]

- Donker Kaat L, Boon AJ, Azmani A, Kamphorst W, Breteler MM, Anar B, Heutink P, van Swieten JC, 2009. Familial aggregation of parkinsonism in progressive supranuclear palsy. *Neurology* 73, 98–105. [PubMed: 19458322]
- Fujioka S, Van Gerpen JA, Uitti RJ, Dickson DW, Wszolek ZK, 2014. Familial progressive supranuclear palsy: a literature review. *Neurodegener. Dis.* 13, 180–182. [PubMed: 24080486]
- Fujioka S, Sanchez Contreras MY, Strongosky AJ, Ogaki K, Whaley NR, Tacik PM, van Gerpen JA, Uitti RJ, Ross OA, Wszolek ZK, Rademakers R, Dickson DW, 2015. Three sib-pairs of autopsy-confirmed progressive supranuclear palsy. *Parkinsonism Relat. Disord.* 21, 101–105. [PubMed: 25443551]
- Guerreiro R, Orme T, Neto JL, Bras J, International DLBGC, 2018. LRP10 in alpha-synucleinopathies. *Lancet Neurol.* 17, 1032–1033.
- Hauw JJ, Daniel SE, Dickson D, Horoupian DS, Jellinger K, Lantos PL, McKee A, Tabaton M, Litvan I, 1994. Preliminary NINDS neuropathologic criteria for Steele-Richardson-Olszewski syndrome (progressive supranuclear palsy). *Neurology* 44, 2015–2019. [PubMed: 7969952]
- Healy DG, Falchi M, O'Sullivan SS, Bonifati V, Durr A, Bressman S, Brice A, Aasly J, Zabetian CP, Goldwurm S, Ferreira JJ, Tolosa E, Kay DM, Klein C, Williams DR, Marras C, Lang AE, Wszolek ZK, Berciano J, Schapira AH, Lynch T, Bhatia KP, Gasser T, Lees AJ, Wood NW, International LRRK2 Consortium, 2008. Phenotype, genotype, and worldwide genetic penetrance of LRRK2-associated Parkinson's disease: a case-control study. *Lancet Neurol.* 7, 583–590. [PubMed: 18539534]
- Höglinger GU, Melhem NM, Dickson DW, Sleiman PM, Wang LS, Klei L, Rademakers R, de Silva R, Litvan I, Riley DE, van Swieten JC, Heutink P, Wszolek ZK, Uitti RJ, Vandrovцова J, Hurtig HI, Gross RG, Maetzler W, Goldwurm S, Tolosa E, Borroni B, Pastor P, Group PSPGS, Cantwell LB, Han MR, Dillman A, van der Brug MP, Gibbs JR, Cookson MR, Hernandez DG, Singleton AB, Farrer MJ, Yu CE, Golbe LI, Revesz T, Hardy J, Lees AJ, Devlin B, Hakonarson H, Muller U, Schellenberg GD, 2011. Identification of common variants influencing risk of the tauopathy progressive supranuclear palsy. *Nat. Genet.* 43, 699–705. [PubMed: 21685912]
- Höglinger GU, Respondek G, Stamelou M, Kurz C, Josephs KA, Lang AE, Mollenhauer B, Muller U, Nilsson C, Whitwell JL, Arzberger T, Englund E, Gelpi E, Giese A, Irwin DJ, Meissner WG, Pantelyat A, Rajput A, van Swieten JC, Troakes C, Antonini A, Bhatia KP, Bordelon Y, Compta Y, Corvol JC, Colosimo C, Dickson DW, Dodel R, Ferguson L, Grossman M, Kassubek J, Krismer F, Levin J, Lorenzl S, Morris HR, Nestor P, Oertel WH, Poewe W, Rabinovici G, Rowe JB, Schellenberg GD, Seppi K, van Eimeren T, Wenning GK, Boxer AL, Golbe LI, Litvan I, Movement Disorder Society-endorsed PSP Study Group, 2017. Clinical diagnosis of progressive supranuclear palsy: the movement disorder society criteria. *Mov. Disord.* 32, 853–864. [PubMed: 28467028]
- Im SY, Kim YE, Kim YJ, 2015. Genetics of progressive supranuclear palsy. *J. Mov. Disord.* 8, 122–129. [PubMed: 26413239]
- Katsuse O, Dickson DW, 2004. Inferior olivary hypertrophy is uncommon in progressive supranuclear palsy. *Acta Neuropathol.* 108, 143–146. [PubMed: 15235807]
- Kia DA, Sabir MS, Ahmed S, Trinh J, Bandres-Ciga S, International Parkinson's Disease Genomics, C., 2018. In alpha-synucleinopathies. *Lancet Neurol.* 17, 1032.
- Lesage S, Drouet V, Majounie E, Deramecourt V, Jacoupy M, Nicolas A, Cormier-Dequaire F, Hassoun SM, Pujol C, Ciura S, Erpapazoglou Z, Usenko T, Maurage CA, Sahbatou M, Liebau S, Ding J, Bilgic B, Emre M, Erginel-Unaltuna N, Guven G, Tison F, Tranchant C, Vidailhet M, Corvol JC, Krack P, Leutenegger AL, Nalls MA, Hernandez DG, Heutink P, Gibbs JR, Hardy J, Wood NW, Gasser T, Durr A, Deleuze JF, Tazir M, Destee A, Lohmann E, Kabashi E, Singleton A, Corti O, Brice A, French Parkinson's Disease Genetics Study, International Parkinson's Disease Genomics, Consortium, 2016. Loss of VPS13C function in autosomal-recessive parkinsonism causes mitochondrial dysfunction and increases PINK1/parkin-dependent mitophagy. *Am. J. Hum. Genet.* 98, 500–513. [PubMed: 26942284]
- Litvan I, Agid Y, Calne D, Campbell G, Dubois B, Duvoisin RC, Goetz CG, Golbe LI, Grafman J, Growdon JH, Hallett M, Jankovic J, Quinn NP, Tolosa E, Zee DS, 1996a. Clinical research criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome): report of the NINDS-SPSP international workshop. *Neurology* 47, 1–9. [PubMed: 8710059]

- Litvan I, Agid Y, Jankovic J, Goetz C, Brandel JP, Lai EC, Wenning G, D'Olhaberriague L, Verny M, Chaudhuri KR, McKee A, Jellinger K, Bartko JJ, Mangone CA, Pearce RK, 1996b. Accuracy of clinical criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome). *Neurology* 46, 922–930. [PubMed: 8780065]
- Mongin M, Delorme C, Lenglet T, Jardel C, Vignal C, Roze E, 2016. Progressive ataxia and palatal tremor: think about POLG mutations. *Tremor Other Hyperkinet Mov* 6, 1–2.
- Pihlström L, Schottlaender L, Chelban V, Houlden H, Consortium MSAE, 2018. LRP10 in alpha-synucleinopathies. *Lancet Neurol.* 17, 1033–1034. [PubMed: 30507385]
- Poorkaj P, Muma NA, Zhukareva V, Cochran EJ, Shannon KM, Hurtig H, Koller WC, Bird TD, Trojanowski JQ, Lee VM, Schellenberg GD, 2002. An R5L tau mutation in a subject with a progressive supranuclear palsy phenotype. *Ann. Neurol.* 52, 511–516. [PubMed: 12325083]
- Quadri M, Mandemakers W, Grochowska MM, Masius R, Geut H, Fabrizio E, Breedveld GJ, Kuipers D, Minneboo M, Vergouw LJM, Carreras Mascaro A, Yonova-Doing E, Simons E, Zhao T, Di Fonzo AB, Chang HC, Parchi P, Melis M, Correia Guedes L, Criscuolo C, Thomas A, Brouwer RWW, Heijnsman D, Ingrassia AMT, Calandra Buonauro G, Rood JP, Capellari S, Rozemuller AJ, Sarchioto M, Fen Chien H, Vanacore N, Olgiatei S, Wu-Chou YH, Yeh TH, Boon AJW, Hoogers SE, Ghazvini M, AS IJ, van IWFJ, Onofrij M, Barone P, Nicholl DJ, Puschmann A, De Mari M, Kievit AJ, Barbosa E, De Michele G, Majoor-Krakauer D, van Swieten JC, de Jong FJ, Ferreira JJ, Cossu G, Lu CS, Meco G, Cortelli P, van de Berg WDJ, Bonifati V, International Parkinsonism Genetics Network, 2018. LRP10 genetic variants in familial Parkinson's disease and dementia with Lewy bodies: a genome-wide linkage and sequencing study. *Lancet Neurol.* 17, 597–608. [PubMed: 29887161]
- Respondek G, Roeber S, Kretschmar H, Troakes C, Al-Sarraj S, Gelpi E, Gaig C, Chiu WZ, van Swieten JC, Oertel WH, Hoglinger GU, 2013. Accuracy of the National Institute for Neurological Disorders and Stroke/Society for Progressive Supranuclear Palsy and neuroprotection and natural history in Parkinson plus syndromes criteria for the diagnosis of progressive supranuclear palsy. *Mov. Disord.* 28, 504–509. [PubMed: 23436751]
- Ross OA, Whittle AJ, Cobb SA, Hulihan MM, Lincoln SJ, Toft M, Farrer MJ, Dickson DW, 2006. Lrrk2 R1441 substitution and progressive supranuclear palsy. *Neuropathol. Appl. Neurobiol.* 32, 23–25. [PubMed: 16409550]
- Sanchez-Contreras M, Heckman MG, Tacik P, Diehl N, Brown PH, Soto-Ortolaza AI, Christopher EA, Walton RL, Ross OA, Golbe LI, Graff-Radford N, Wszolek ZK, Dickson DW, Rademakers R, 2017. Study of LRRK2 variation in tauopathy: progressive supranuclear palsy and corticobasal degeneration. *Mov. Disord.* 32, 115–123. [PubMed: 27709685]
- Sanchez-Contreras MY, Kouri N, Cook CN, Serie DJ, Heckman MG, Finch NA, Caselli RJ, Uitti RJ, Wszolek ZK, Graff-Radford N, Petrucelli L, Wang LS, Schellenberg GD, Dickson DW, Rademakers R, Ross OA, 2018. Replication of progressive supranuclear palsy genome-wide association study identifies SLCO1A2 and DUSP10 as new susceptibility loci. *Mol. Neurodegener.* 13, 37. [PubMed: 29986742]
- Suyama N, Kobayashi S, Isino H, Iijima M, Imaoka K, 1997. Progressive supranuclear palsy with palatal myoclonus. *Acta Neuropathol.* 94, 290–293. [PubMed: 9292700]
- Ujiiie S, Hatano T, Kubo S, Imai S, Sato S, Uchihara T, Yagishita S, Hasegawa K, Kowa H, Sakai F, Hattori N, 2012. LRRK2 I2020T mutation is associated with tau pathology. *Parkinsonism Relat. Disord.* 18, 819–823. [PubMed: 22525366]
- Vergouw LJM, Ruitenberga A, Wong TH, Melhem S, Breedveld GJ, Criscuolo C, De Michele G, de Jong FJ, Bonifati V, van Swieten JC, Quadri M, 2019. LRP10 variants in Parkinson's disease and dementia with Lewy bodies in the South-West of The Netherlands. *Parkinsonism Relat. Disord.* 65, 243–247. [PubMed: 31147221]
- von Einem B, Wahler A, Schips T, Serrano-Pozo A, Proepper C, Boeckers TM, Rueck A, Wirth T, Hyman BT, Danzer KM, Thal DR, von Arnim CA, 2015. The golgi-localized gamma-ear-containing ARF-binding (GGA) proteins alter amyloid-beta precursor protein (APP) processing through interaction of their GAE domain with the beta-site APP cleaving enzyme 1 (BACE1). *PLoS One* 10, e0129047. [PubMed: 26053850]
- von Einem B, Eschbach J, Kiechle M, Wahler A, Thal DR, McLean PJ, Weishaupt JH, Ludolph AC, von Arnim CAF, Danzer KM, 2017. The Golgi-localized, gamma ear-containing, ARF-binding

(GGA) protein family alters alpha synuclein (alpha-syn) oligomerization and secretion. *Aging (Albany NY)* 9,1677–1697. [PubMed: 28722658]

Zimprich A, Biskup S, Leitner P, Lichtner P, Farrer M, Lincoln S, Kachergus J, Hulihan M, Uitti RJ, Calne DB, Stoessl AJ, Pfeiffer RF, Patenge N, Carbajal IC, Vieregge P, Asmus F, Muller-Myhsok B, Dickson DW, Meitinger T, Strom TM, Wszolek ZK, Gasser T, 2004. Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron* 44, 601–607. [PubMed: 15541309]

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Table 1

Demographic and clinical characteristics

	n = 246	n = 704
Sex, male	127 (52%)	377 (54%)
Diagnosis (NINDS-SPSP)		
Possible PSP	85 (35%)	0 (0%)
Probable PSP	128 (52%)	0 (0%)
Definite PSP	33 (13%)	704 (100%)
Age at disease onset, y (n = 246;476)	65.8 (7.5)	68.1 (8.4)
Family history of neurodegenerative diseases (n = 244;0)		
1st degree	71 (29%)	NA
2nd degree	21 (9%)	NA
No	152 (62%)	NA
Deceased (n = 244;704)	242 (98%)	704 (100%)
Age at death, y (n = 241;698)	73.7 (7.3)	75.3 (8.2)

Values are presented as n (%) or mean (SD).

Key: NINDS-SPSP, National Institute for Neurological Disorders and Stroke/Society for PSP; PSP, progressive supranuclear palsy; NA, not available.

Table 2

Possibly pathogenic *LRP10* variants

Genetic information										Clinical information		
Genomic position	Nucleotide change	Amino acid change	Exon	Coding effect	dbSNP 142 accession number	Allele frequency GnomAD (alleles)	Functional predictions: Pathogenic (total)	Splicing predictions: Deleterious (total)	Patient	NINDS-SFSP criteria	Age at onset (y)	Age at death (y)
Discovery cohort												
14:23344522	c.1165 G>A	p.Asp389Asn	5	Missense	rs754181235	0.01% (37)	6/11	n.a.	1	Probable PSP	61	66
14:233445134	c.977 G>A	p.Gly326Asp	5	Missense	rs547591765	0.006% (14)	6/11	n.a.	2	Possible PSP	55	65
Validation cohort												
14:23344630	c.473 G>A	p.Arg158His	5	Missense	rs764424911	0.005% (13)	6/11	n.a.	1	Definite PSP	68	75
14:23344816	c.659 G>A	p.Cys220Tyr	5	Missense	rs867533372	.	10/11	n.a.	2	Definite PSP	70	74
14:23344989	c.832 A>G	p.Thr278Ala	5	Missense	.	.	5/11	n.a.	3	Definite PSP	74	87
14:23345074	c.917 G>A	p.Gly306Asp	5	Missense	rs375748692	0.007% (21)	9/11	n.a.	4	Definite PSP	NA	66
14:23345931 ^a	c.1458 G>C	p.Glu486Asp	6	Missense	rs142130715	0.01% (32)	11/11	0/4	5	Definite PSP	74	80
14:23346254	c.1660 C>T	p.Arg554*	7	Stop gain	rs201213246	0.01% (28)	n.a.	n.a.	6	Definite PSP	78	83
14:23346575	c.1981 C>T	p.Arg661Cys	7	Missense	rs771796662	0.004% (10)	8/11	n.a.	7	Definite PSP	74	84
									8	Definite PSP	NA	84

The Genome Reference Consortium Human Build 37 (hg19) and NM_014045-4 LRP10 transcript were used.

Splicing prediction programs: SSF, MaxEnt, NNSPLICE, GeneSplicer.

Key: GnomAD, Genome Aggregation Database; NINDS-SFSP, National Institute for Neurological Disorders and Stroke/Society for progressive supranuclear palsy; n.a., not applicable; NA, not available.

^aThis variant was not validated by Sanger sequencing because no additional DNA was available.