



Vitreoscilla massiliensis sp. nov., Isolated From the Stool of an Amazonian Patient

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Abstract

Strain SN6^T is a non-motile and non-spore-forming gram-negative bacterium which was isolated from the stool sample of an Amazonian patient. The optimum growth was observed at 37 °C, pH 7, and 0–5 g/l of NaCl. Based on the 16S rRNA gene sequence similarity, the strain SN6^T exhibited 97.5% identity with *Vitreoscilla stercoraria* strain ATCC_15218 (L06174), the phylogenetically closest species with standing in nomenclature. The predominant fatty acid was hexadecenoic acid (31%). The genomic DNA G + C content of the strain SN6^T was 49.4 mol %. After analysis of taxonogenomic data, phenotypic and biochemical characteristics, we concluded that strain SN6^T represents a new species of the genus *Vitreoscilla* for which the name *Vitreoscilla massiliensis* sp.nov is proposed. The type strain is SN6^T (=CSUR P2036 = LN870312 = DSM 100958).

Abbreviations

| | |
|----------|--|
| CSUR | Collection de Souches de l'Unité des Rickettsies |
| DSM | Deutsche Sammlung von Mikroorganismen |
| FAME | Fatty acid methyl ester |
| GC/SM | Gaz chromatography/mass spectrometry |
| OrthoANI | Orthologous average nucleotide identity |
| COGs | Clusters of orthologous groups |
| NR BLAST | Non-redundant protein sequence basic local alignment search tool |
| ORF | Open reading frame |
| GGDC | Genome-to-genome distance calculator |

Introduction

This strain was isolated from the stool specimen of an obese Amazonian patient as part of the culturomics study [1] to search for microaerophilic bacteria from human gut. The genus *Vitreoscilla* was first described by Pringsheim in 1951, after having proposed the family *Vitreoscillaceae* in 1949. In 1986, Strohl et al. proposed three new species with validated names (*Vitreoscilla stercoraria*, *Vitreoscilla beggiatoides* and *Vitreoscilla filiformis*) of this genus [2]. In 2013, through the use of new-generation sequencing tools, the genus *Vitreoscilla* was placed in the *Neisseriaceae* family on the basis of its branching in the 16S rRNA gene tree [3]. Within its clade, members of the genus were the only ones capable of evolving in different habitats.

Since the use of the culturomic concept, the repertoire of bacteria isolated from the human digestive microbiota [4] has expanded considerably. The characterization of these new species is based on a ribosomal RNA sequencing coupled with a taxonogenomics description, a strategy combining a comparison of genomic analysis and phenotypic characteristics, including the matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF) spectrum. In the present study, we used this approach to facilitate the identification and the description of this novel species named *Vitreoscilla massiliensis* sp.nov.

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Materials and Methods

Vitreoscilla massiliensis SN6^T was isolated by cultivation on 5% sheep blood agar under microaerophilic conditions at 37 °C for 48 h and the strain could not be identified by Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry.

(MALDI-TOF MS). The bacterial spectrum obtained was incremented in our database and its comparison with those of BioTyper database spectra and our own collection did not allow for its identification. Sequencing of 16S rRNA gene of the strain SN6^T showed a nucleotide sequence similarity of 97.5% with *V. stercoraria* strain (ATCC 15218) and *V. stercoraria* strain Göttingen 1488-6 (NR_025894.1), the phylogenetically closest species with standing in nomenclature (Fig. 1).

Optimal Growth

Growth at various temperatures (28 °C, 37 °C, 42 °C, 45 °C) in different atmospheres (aerobic, microaerophilic using CampyGen from Thermo Scientific and anaerobic using AnaeroGenTM from bioMérieux) was tested by culture on Columbia agar (bioMérieux) after 48 h of incubation. The salinity acceptance limit of SN6^T strain was investigated by culture on a home-made culture medium consisting of a Columbia agar culture medium (Sigma-Aldrich,

Saint-Quentin Fallavier, France) modified by adding (per liter) 5 g MgCl₂ 6H₂O, 5 g MgSO₄ 7H₂O, 2 g KCl, 1 g CaCl₂ 2H₂O; 0.5 g NaBr, 0.5 g NaHCO₃, and 2 g glucose with various NaCl concentrations 0, 5, 10, 25, 50, and 75 g/L. The pH range (6; 6.5; 7; 8.5) for growth was also determined and pH was adjusted by addition of HCl or NaOH.

Biochemical and Chemotaxonomic Analysis

The abilities of the strain SN6^T to use various substrates as sole carbon sources were evaluated using the API 20NE and API 50CH (bioMérieux) and the presence of some enzyme activities using APIZYM following the manufacturer's instructions. All tests were performed in duplicate. Susceptibility to antimicrobial agents was determined by the disk (i2a, Montpellier, France) diffusion method [5] on Mueller–Hinton agar in a Petri dish (BioMérieux) after 48 h of incubation at 37 °C under aerobic conditions. The interpretation of inhibition diameters to the manual measurement using a ruler was done using a Sirscan system[©] (i2a, Montpellier, France) according to the criteria proposed by the Comité de l'Antibiogramme of the French Society for Microbiology [6]. Cellular fatty acid methyl ester (FAME) analysis was performed by GC/MS. Two samples were prepared with approximately 65 mg of bacterial biomass per tube harvested from several culture plates. Fatty acid methyl esters were prepared as previously described by Sasser [7].

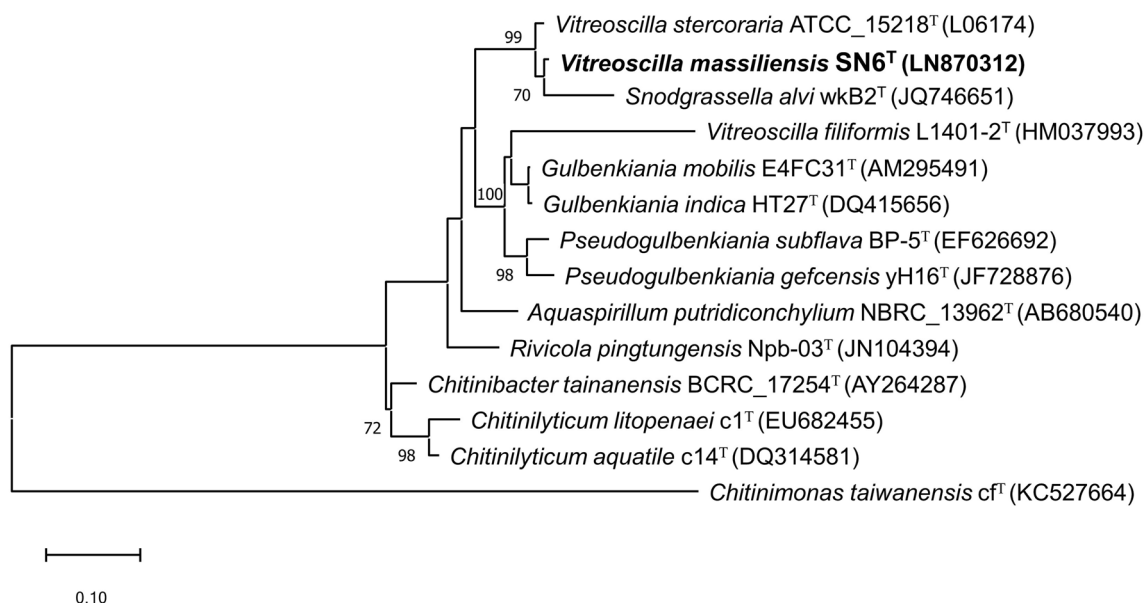


Fig. 1 Phylogenetic tree showing the position of *Vitreoscilla massiliensis* SN6^T relative to other phylogenetically close neighbors. Sequences were aligned using CLUSTALW, and phylogenetic inferences are obtained with kimura two-parameter models using the maximum-likelihood method within the MEGA software. Numbers

at nodes are percentages of bootstrap values obtained by repeating analysis 1,000 times to generate majority consensus tree. Scale bar indicates 1% nucleotide sequence divergence. The scale bar represents 500 nm

GC/MS analyses were carried out as previously described by Dione et al. [8]. Briefly, fatty acid methyl esters were separated using an Elite 5-MS column and monitored by mass spectrometry (Clarus 500—SQ 8 S, Perkin Elmer, Courtaboeuf, France). Spectral database search was performed using MS Search 2.0 operated with the Standard Reference Database 1A (NIST, Gaithersburg, USA) and the FAMES mass spectral database (Wiley, Chichester, UK).

Genome Sequencing and Assembly

DNA of strain SN6^T was extracted on the EZ1 biorobot (Qiagen) with a EZ1 DNA tissues kit after pretreatment by a lysozyme incubation at 37 °C, as previously described [9]. Genomic DNA (gDNA) was quantified by a Qubit assay with the high-sensitivity kit (Life technologies, Carlsbad, CA, USA) and sequenced on the MiSeq Technology (IlluminaInc, San Diego, CA, USA) with the mate pair strategy, as previously described [9]. The gDNA was barcoded in order to be mixed with 11 other projects with the Nextera Mate Pair sample prep kit (Illumina). The assembly of the genome was carried out with the help of a pipeline that allowed the creation of an assembly with different softwares (Velvet [10], Spades [11] and Soap Denovo [12], on trimmed (MiSeq and Trimmomatic [13] softwares) or untrimmed data (only MiSeq software). For each of the six assemblies performed, GapCloser [12] was used to reduce gaps. Then, contamination with Phage Phix was identified (blastn against Phage Phix174 DNA sequence) and eliminated. Finally, scaffolds whose size was less than 800 bp were removed and scaffolds whose depth value was lower than 25% of the mean depth were removed (identified as possible contaminants). The best assembly was selected using different criteria (number of scaffolds, N50 and number of N). Spades gave the best assembly of this strain, with a depth coverage of 98.

Genome Annotation and Comparison

We used Prodigal as predicting tool of open reading frames (ORFs) [14] with default parameters. The predicted ORFs were excluded if they spanned a sequencing gap region (contained N). Using BLASTP, predicted bacterial protein sequences were blasted against GenBank and clusters of orthologous groups (COG) databases, DNA G+C content was identified by The RAST Server [15], and the tRNAs and rRNAs were predicted using the tRNAScanSE [16] and RNAmmer tools [17], respectively. SignalP was used for Signal peptides prediction [18], the number of transmembrane helices was predicted using TMHMM [19], ORFans were identified if their BLASTP E-value was lower than 1e03 for alignment length greater than 80 amino acids. If alignment lengths were smaller than 80 amino acids, we used an E-value of 1e-05. Artemis

[20] and DNA Plotter [21] were used for data management and visualization of genomic features, respectively. The Average Nucleotide identity at the genome level between *V. massiliensis* SN6 CZPV00000000.1, *V. stercorearia* ATCC_15218 (ARNN00000000.1), *V. filiformis* ATCC_43190 (CP022423.1), *Gulbenkiania mobilis* E4FC31 (LIVN00000000.1), *Chitinilyticum litopenaei* DSM_21440 (ATZJ00000000.1), *Chitinilyticum aquatile* c14 (AUMS00000000.1), *Chitinibacter tainanensis* BCRC_17254 (AUCN00000000.1), and *Snodgrassella alvi* wkB2_wkB2 (CP007446.1) was estimated using Orthologous Average nucleotide identity tool (OAT) [22].

Results and Discussion

Based on the sequence similarity threshold values of the 16S rRNA gene that delineate a new species according to the recommendations of Stackebrandt and Ebers [23], the strain SN6^T can, therefore, be classified as a new species of the genus *Vitreoscilla* and was accordingly named *V. massiliensis* SN6^T [24].

Biochemical and Chemotaxonomic Analyses

API ZYM tests show positive reactions for esterase, esterase lipase, leucine arylamidase, acid phosphatase, and naphthol-AS-BI-phosphohydrolase. In API 50CH, no substrate fermentation was observed and in API 20NE assimilation of substrates was observed for L-arginine dihydrolase and potassium gluconate. Some phenotypic characteristics of SN6^T with those of closely related species are presented in Table 1. The most abundant fatty acid is hexadecenoic acid (31%). Several hydroxyl fatty acids such as C_{12:0} 3-OH (4.5 ± 1.0) and C_{14:0} 3-OH (2.9 ± 0.1) are described. Other fatty acids such as 9-Hexadecenoic acid (22.0 ± 0.5), Dodecanoic acid (10.2 ± 0.6), 2-hexyl-cyclopropanoic acid (8.8 ± 0.4), Octadecenoic acid (8.0 ± 0.2), Pentadecanoic acid (5.5 ± 0.2), Tetradecanoic acid (3.3 ± 0.3), and Heptadecanoic acid (1.1 ± 0.1) were detected. The strain SN6^T was resistant to Oxacillin and Metronidazole, but susceptible to other antibiotics tested.

Genome Properties

The genome is 3,716,289 bp long with 49.4% GC content (Fig. 2, Table 2). It is composed of 10 scaffolds (composed of 13 contigs). Of the 3 716 predicted genes, 3 627 were protein-coding genes and 89 were RNAs (5 genes are 5S rRNA, 5 genes are 16S rRNA, 5 genes are 23S rRNA and 74 genes are TRNA genes). A total of 2,263 genes (62.3%) were assigned with putative function (by cogs or by NR blast). 475 genes were identified as ORFans (13.1%). The

Table 1 Differential characteristics of *Vitreoscilla massiliensis* SN6^T, *Vitreoscilla stercoraria* ATCC 15218, *Vitreoscilla filiformis* ATCC 15551, *Vitreoscilla beggiatoides* B23SS, *Gulbenkiania mobilis* E4FC31, *Chitinibacter tainanensis* BCRC 17254, *Chitinilyticum litopenaei* DSM_21440_c1, and *Snodgrassella alvi* wkB2 wkB2

| Properties | <i>V.massiliensis</i> | <i>V.stercoraria</i> ^a | <i>V.filiformis</i> ^b | <i>V.beggiatoides</i> ^c | <i>G.mobilis</i> ^d | <i>C.litopenaei</i> ^e | <i>C.tainanensis</i> ^f | <i>S.alvi</i> ^g |
|------------------------------------|-----------------------------|-----------------------------------|----------------------------------|------------------------------------|-------------------------------|----------------------------------|-----------------------------------|----------------------------|
| Cell diameter (µm) | 0.5 | 1.0 | 1.0–1.5 | 2.5–3 | 0.2–0.4 | 0.3–0.5 | 0.5–0.9 | 0.4 |
| Oxygen requirement | Aerobic/ Microaerophilic | Aerobic | Aerobic/ Microaerophilic | Aerobic/ Microaerophilic | Aerobic | Aerobic/ Anaerobic | Aerobic | Microaerophilic |
| Motility | – | + | + | + | + | + | + | – |
| Endospore formation | – | Na | Na | Na | – | – | Na | Na |
| pH | 7–7.5 | 7.5–7.7 | 7.5 | 7.5 | 5.5–9.0 | 7–11 | 5.5–9 | 6.0–6.5 |
| NaCl % (w/v) | 0–0.5 | Na | 0 | 0.5 | 1.0 | 0–0.75 | Na | Na |
| Indole | – | Na | Na | Na | – | – | – | – |
| Production of Alkaline phosphatase | – | Na | Na | Na | Na | + | Na | Na |
| Catalase | – | – | – | – | + | + | + | + |
| Oxidase | – | – | + | + | + | – | + | – |
| Nitrate reductase | – | – | + | + | – | + | – | + |
| Urease | – | Na | Na | Na | – | – | – | + |
| β-galactosidase | – | Na | Na | Na | – | – | – | – |
| N-acetylglucosamine | – | Na | Na | Na | – | + | + | Na |
| Acid from L-Arabinose | + | – | – | – | – | – | – | – |
| Trehalose | – | – | – | – | – | + | – | Na |
| D-mannose | + | – | – | – | – | + | – | – |
| Mannitol | + | – | – | – | – | – | – | – |
| D-glucose | + | – | + | – | – | + | – | – |
| D-fructose | – | – | – | – | – | + | – | – |
| Maltose | + | – | – | – | – | + | – | – |
| D-lactose | – | – | – | – | – | – | Na | Na |
| D-raffinose | – | Na | – | – | – | – | Na | Na |
| Habitat | Human gut | Dung of Cow | Freshwater sediments | Sandy sediments | Wastewater | Freshwater pond | Soil | Gut of Bees |

Na not available

^aData from Mayfield et al. [25]

^{b, c}Data from Strohl et al. [2]

^dData from Vaz-Moreira et al. [26]

^eData from Chang et al. [27]

^fData from Chern et al. [28]

^gData from Kwong et al. [29]

remaining genes were described as hypothetical proteins (744 genes ≥ 20.5%). A summary of the distribution of *V. massiliensis* genes into the different COGs categories is presented in Table 3.

Genome Comparison

The draft genome sequence and the G+C content of *V. massiliensis* (3.71 MB and 49.4%) is larger than that of

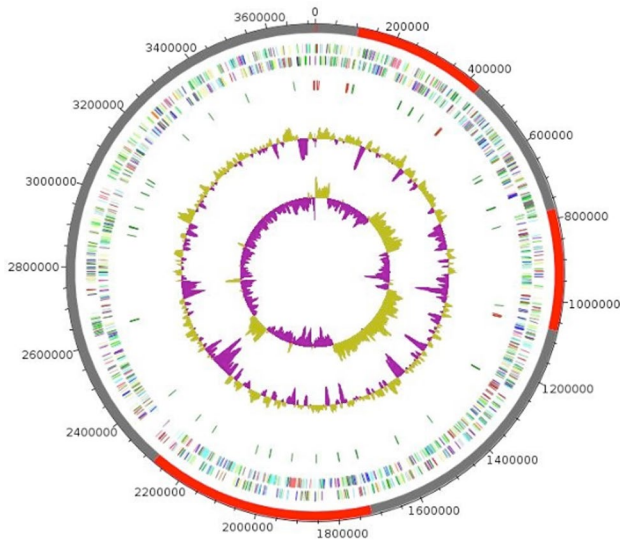


Fig. 2 Graphical circular map of the genome of *Vitreoscilla massiliensis* strain SN6^T from outside to the center: genes on the forward strand colored by COG categories (only genes assigned to COG), genes on the reverse strand colored by COG categories (only gene assigned to COG), RNA genes (tRNAs green, rRNAs red), GC content, and GC skew (Color figure online)

Table 2 Nucleotide content and gene count levels of the genome

| Attribute | Genome (total) | |
|--------------------------------------|----------------|-------------------------|
| | Value | % of total ^a |
| Size (bp) | 3,716,289 | 100 |
| G+C content (%) | 1,836,063 | 49.42 |
| Coding region (bp) | 3,249,937 | 87.45 |
| Total genes | 3,716 | 100 |
| RNA genes | 89 | 2.39 |
| Protein-coding genes | 3,627 | 100 |
| Genes with function prediction | 2,263 | 62.39 |
| Genes assigned to COGs | 2,184 | 60.21 |
| Genes with peptide signals | 677 | 18.66 |
| Genes with transmembrane helices | 774 | 21.33 |
| Genes associated to virulence | 715 | 19.71 |
| ORFn genes | 475 | 13.09 |
| Genes associated with PKS or NRPS | 20 | 0.55 |
| Genes associated to toxin/antitoxine | 115 | 3.17 |

^aThe total is based on either the size of the genome in base pairs or the total number of protein-coding genes in the annotated genome

V. stercoraria (2.58 MB and 43.9% respectively). Also, the gene content of *V. massiliensis* is larger than that of (3,627 and 2,440 respectively). The distribution of genes into COG categories was similar in all 7 compared genomes (Fig. 3). All genomes were compared with *V. massiliensis* using Orthologous average nucleotide identity. The OrthoANI analysis showed that identity nucleotide value is

76.4% with *V. stercoraria* which is lower than 95% (Fig. 4). Likewise, we obtained similar results for the analysis of the digital DNA-DNA hybridization (dDDH) with 31.60% between *V. massiliensis* and *V. stercoraria* (Table 4).

Conclusion

Based on the phenotypic characteristics, and phylogenetic and genomic analyses of strain SN6^T, we suggest the creation of a new species within the *Vitreoscilla* genus, for which the name *V. massiliensis* sp. nov., is proposed.

Description of *Vitreoscilla massiliensis* sp. nov.

Vitreoscilla massiliensis (mas.si.li.en'sis. L. fem. adj. *massiliensis*, of Massilia, the Latin name of Marseille where strain SN6^T was first isolated).

Cells are Gram-negative (0.5 × 1.5–2 μm), non-motile, non-spore-forming, and often occur in a long chain under electron microscopy. *V. massiliensis* SN6^T grows at 28–37 °C and pH 7–7.5 and does not grow above 0.5% salinity. *V. massiliensis* SN6^T grows under microaerophilic atmosphere and a lower growth was observed under anaerobic conditions. On agar plates, colonies were gray, smooth, and hemolytic with 0.5 to 1 mm in diameter after 48 h of incubation under aerobic conditions. They are catalase and oxidase negative. Tests were negative for urease, the reduction of nitrates, indole production, and fermentation of β-galactosidase. API 50CH shows that the carbohydrates provided by this panel were not used. *V. massiliensis* SN6^T is susceptible to Vancomycin (0.5), Cefotaxime (0.94), Tobamycin (0.38 μg), Fosfomycin (16 μg), Teicoplanin (1.5 μg), Rifampicin (0.29 μg), Colistin (0.32 μg), Imipenem (0.23 μg), Erythromycin (0.25 μg), Ceftriaxone (0.32 μg), and resistant to Oxacillin and Metronidazole. Major fatty acids are hexadecanoic acid (C_{16:0}), 9-hexadecenoic acid (C_{16:1n7}), and an unusual cylo fatty acid named 2-hexyl-cyclopropaneoctanoic acid (C_{16:0} 9,10-methylene). The DNA G + C content is about 49.4%.

The type strain is SN6^T (=CSUR P2036=LN870312=DSM 100958) and was isolated from the stool specimen of an obese Amazonian patient.

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Table 3 Number of genes associated with the 25 general COG functional categories

| Code | Value | % of total | Description |
|------|-------|-------------|---|
| [J] | 213 | 5.872622 | Translation |
| [A] | 1 | 0.027570996 | Rna processing and modification |
| [K] | 121 | 3.3360906 | Transcription |
| [L] | 88 | 2.4262476 | Replication, recombination and repair |
| [B] | 4 | 0.110283986 | Chromatin structure and dynamics |
| [D] | 32 | 0.8822719 | Cell cycle control, mitosis and meiosis |
| [Y] | 0 | 0 | Nuclear structure |
| [V] | 44 | 1.2131238 | Defense mechanisms |
| [T] | 85 | 2.3435347 | Signal transduction mechanisms |
| [M] | 140 | 3.8599393 | Cell wall/membrane biogenesis |
| [N] | 29 | 0.7995589 | Cell motility |
| [Z] | 0 | 0 | Cytoskeleton |
| [W] | 19 | 0.52384895 | Extracellular structures |
| [U] | 27 | 0.7444169 | Intracellular trafficking and secretion |
| [O] | 97 | 2.6743865 | Post-translational modification, protein turnover, chaperones |
| [X] | 77 | 2.1229665 | Mobilome, prophages, transposons |
| [C] | 177 | 4.8800664 | Energy production and conversion |
| [G] | 125 | 3.4463744 | Carbohydrate transport and metabolism |
| [E] | 287 | 7.9128757 | Amino acid transport and metabolism |
| [F] | 64 | 1.7645438 | Nucleotide transport and metabolism |
| [H] | 119 | 3.2809484 | Coenzyme transport and metabolism |
| [I] | 133 | 3.6669421 | Lipid transport and metabolism |
| [P] | 136 | 3.7496552 | Inorganic ion transport and metabolism |
| [Q] | 90 | 2.4813895 | Secondary metabolites biosynthesis, transport and catabolism |
| [R] | 234 | 6.4516125 | General function prediction only |
| [S] | 145 | 3.9977942 | Function unknown |
| – | 1443 | 39.784946 | Not in COGs |

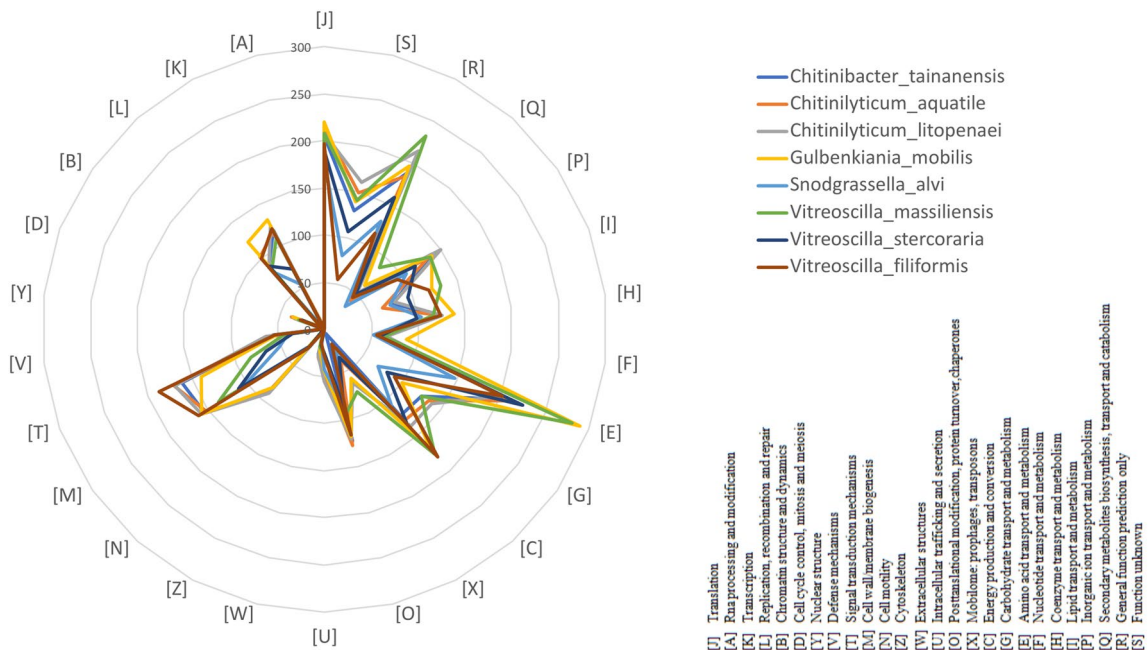
**Fig. 3** Distribution of functional classes of predicted genes according to the clusters of orthologous groups of proteins

Fig.4 Heatmap generated with OrthoANI values calculated using the OAT software between *Vitreoscilla* species and other closely related species with standing in nomenclature

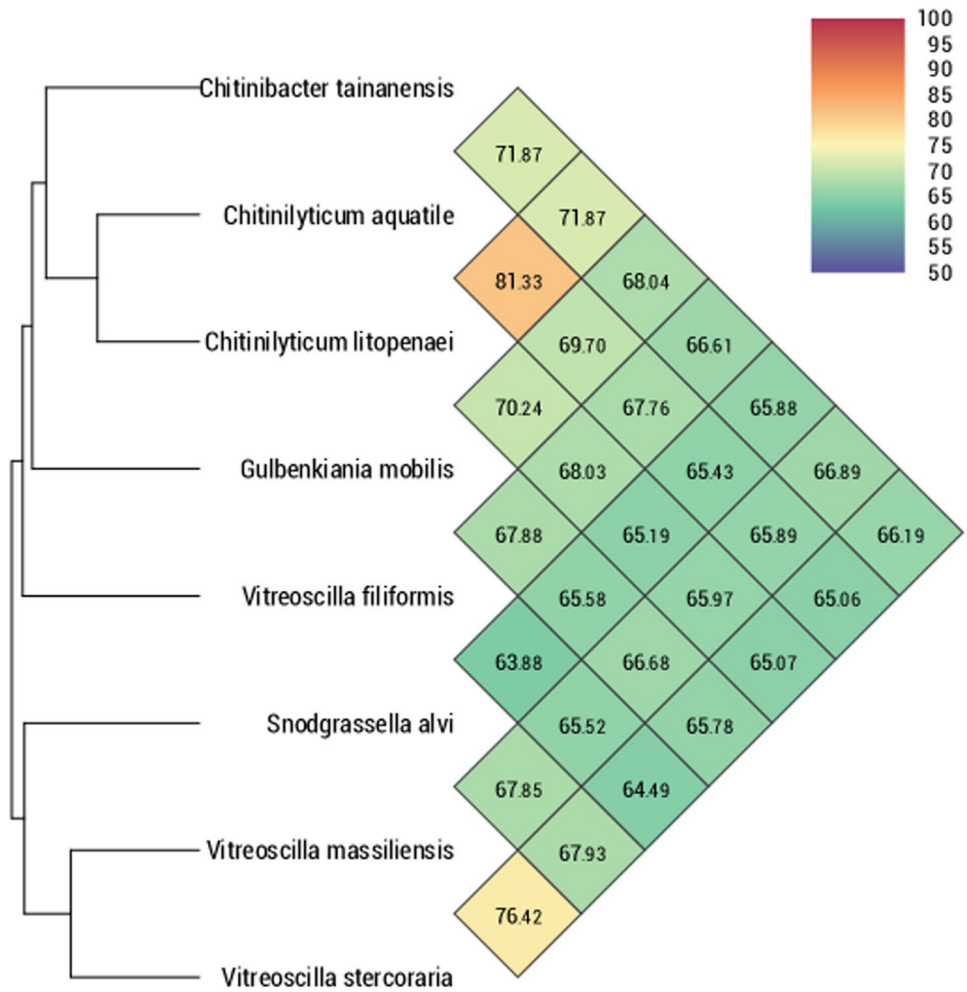


Table 4 Pairwise comparison of *Vitreoscilla massiliensis* SN6^T with other species using GGDC, formula 2(DNA-DNA hybridization estimates based on identities/HSP length)

| | <i>C. tainanensis</i> | <i>C. aquatile</i> | <i>G. mobilis</i> | <i>S. alvi</i> | <i>C. litopenaei</i> | <i>V. stercoraria</i> | <i>V. filiformis</i> |
|------------------------|-----------------------|--------------------|-------------------|-----------------|----------------------|-----------------------|----------------------|
| <i>V. massiliensis</i> | 23.20% 2.4± | 30.00% 2.45± | 29.40% 2.45± | 26.80% 2.45± | 30.80% 2.45± | 21.50% 2.35± | 28.30% 2.45± |
| <i>C. tainanensis</i> | | 18.80% 2.3± | 19.30% 2.25± | 31.10% 2.45± | 19.20% 2.3± | 19.70% 2.3± | 26.30% 2.4± |
| <i>C. aquatile</i> | | | 18.30% 2.25± | 37.20% 2.5± | 25.60% 2.45± | 28.40% 2.45± | 19.50% 2.3± |
| <i>G. mobilis</i> | | | | 29.30% 2.45± | 18.7% 2.25± | 31.20% 2.45± | 18.50% 2.3± |
| <i>S. alvi</i> | | | | | 33.8% 2.5± | 24.00% 2.35± | 29.60% 2.45± |
| <i>C. litopenaei</i> | | | | | | 32.9% 2.5± | 18.40% 2.25± |
| <i>V. stercoraria</i> | | | | | | | 31.60% 2.45± |

V. massiliensis *Vitreoscilla massiliensis* SN6^T, *G. mobilis* *Gulbenkiania mobilis* E4FC31, *C. litopenaei* *Chitinilyticum litopenaei* DSM _21440_c1, *S. alvi* *Snodgrassella alvi* wkB2 wkB2, *C. tainanensis* *Chitinibacter tainanensis* BCRC 17254, *C. aquatile* *Chitinilyticum aquatile* c14, *V. stercoraria* *Vitreoscilla stercoraria* ATCC 15218 and *V. filiformis* *Vitreoscilla filiformis* ATCC 15551

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Author Contributions SN, MM, and MR performed the experiments. SN and SK wrote the manuscript. SK, DR, RS, and PEF reviewed the drafts of the manuscript. SK, JCL, and DR are responsible for data interpretation.

Declarations

Conflict of interest The authors have no conflicts of interest to declare.

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References

- Lagier J-C, Edouard S, Pagnier I et al (2015) Current and past strategies for bacterial culture in clinical microbiology. *Clin Microbiol Rev* 28:208–236
- Strohl WR, Schmidt TM, Lawry NH et al (1986) Characterization of *Vitreoscilla beggiatoides* and *Vitreoscilla filiformis* sp. nov., nom. rev., and comparison with *Vitreoscilla stercoraria* and *Beggiatoa alba*. *Int J Syst Evol Microbiol* 36:302–313
- Adeolu M, Gupta RS (2013) Phylogenomics and molecular signatures for the order *Neisseriales*: proposal for division of the order *Neisseriales* into the emended family *Neisseriaceae* and *Chromobacteriaceae* fam. nov. *Antonie Van Leeuwenhoek* 104:1–24
- Lagier J-C, Khelaifa S, Alou MT et al (2016) Culture of previously uncultured members of the human gut microbiota by culturomics. *Nat Microbiol* 1:16203
- Le Page S, van Belkum A, Fulchiron C et al (2015) Evaluation of the PREVI® Isola automated seeder system compared to reference manual inoculation for antibiotic susceptibility testing by the disk diffusion method. *Eur J Clin Microbiol Infect Dis Off Publ Eur Soc Clin Microbiol* 34:1859–1869
- The European Committee on Antimicrobial Susceptibility Testing (2016) Breakpoint tables for interpretation of MICs and zone diameters. http://www.eucast.org/clinical_breakpoints/
- Sasser M (2006) Bacterial identification by gas chromatographic analysis of fatty acid methyl esters (GC-FAME). MIDI, Technical Note, New York
- Dione N, Sankar SA, Lagier J-C et al (2016) Genome sequence and description of *Anaerosalibacter massiliensis* sp. nov. *New Microbes New Infect* 10:66–76
- Lagier J-C, Ramasamy D, Rivet R et al (2012) Non-contiguous finished genome sequence and description of *Cellulomonas massiliensis* sp. nov. *Stand Genomic Sci* 7:258–270
- Zerbino DR, Birney E (2008) Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829
- Bankevich A, Nurk S, Antipov D et al (2012) SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol J Comput Cell Biol* 19:455–477
- Luo R, Liu B, Xie Y et al (2012) SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. *GigaScience* 1:18
- Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinforma Oxf Engl* 30:2114–2120
- Hyatt D, Chen G-L, Locascio PF et al (2010) Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinform* 11:119
- Aziz RK, Bartels D, Best AA et al (2008) The RAST server: rapid annotations using subsystems technology. *BMC Genom* 9:75
- Avni D, Biberman Y, Meyuhos O (1997) The 5' terminal oligopyrimidine tract confers translational control on TOP mRNAs in a cell type- and sequence context-dependent manner. *Nucleic Acids Res* 25:995–1001
- Lagesen K, Hallin P, Rødland EA et al (2007) RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108
- Bendtsen JD, Nielsen H, von Heijne G, Brunak S (2004) Improved prediction of signal peptides: signalp 3.0. *J Mol Biol* 340:783–795
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL (2001) Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580
- Carver T, Thomson N, Bleasby A et al (2009) DNAPlotter: circular and linear interactive genome visualization. *Bioinforma Oxf Engl* 25:119–120
- Rutherford K, Parkhill J, Crook J et al (2000) Artemis: sequence visualization and annotation. *Bioinforma Oxf Engl* 16:944–945
- Lee I, Kim YO, Park S-C, Chun J (2015) OrthoANI: an improved algorithm and software for calculating average nucleotide identity. *Int J Syst Evol Microbiol* 66:1100–1103
- Stackebrandt E, Ebers J (2006) Taxonomic parameters revisited: tarnished gold standards. *Microbiol Today* 33:152–155
- Ndongo S, Lagier J-C, Raoult D, Fournier P-E (2018) *Gorillibacterium timonense* sp. nov., and *Vitreoscilla massiliensis* sp. nov., two new bacterial species isolated from stool specimens of obese Amazonian patients. *New Microbes New Infect* 23:48–51
- Mayfield DC, Kester AS (1972) Physiological studies on *Vitreoscilla stercoraria*. *J Bacteriol* 112:1052–1056
- Vaz-Moreira I, Nobre MF, Nunes OC, Manaia CM (2007) *Gulbenkiania mobilis* gen. nov., sp. nov., isolated from treated municipal wastewater. *Int J Syst Evol Microbiol* 57:1108–1112
- Chang S-C, Wu M-C, Chen W-M et al (2009) *Chitinilyticum litopenaei* sp. nov., isolated from a freshwater shrimp pond, and emended description of the genus *Chitinilyticum*. *Int J Syst Evol Microbiol* 59:2651–2655
- Chern L-L, Stackebrandt E, Lee S-F et al (2004) *Chitinibacter tainanensis* gen. nov., sp. nov., a chitin-degrading aerobe from soil in Taiwan. *Int J Syst Evol Microbiol* 54:1387–1391
- Kwong WK, Moran NA (2013) Cultivation and characterization of the gut symbionts of honey bees and bumble bees: description of *Snodgrassella alvi* gen. nov., sp. nov., a member of the family *Neisseriaceae* of the *Betaproteobacteria*, and *Gilliamella apicola* gen. nov., sp. nov., a member of *Orbaceae* fam. nov., *Orbales* ord. nov., a sister taxon to the order “*Enterobacteriales*” of the *Gammaproteobacteria*. *Int J Syst Evol Microbiol* 63:2008–2018

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