

Research Paper

Silencing of METTL3 effectively hinders invasion and metastasis of prostate cancer cells

Yabing Chen^{1,2#}, Chun Pan^{1,2#}, Xiaotong Wang³, Dihui Xu^{1,2}, Yuhan Ma^{1,2}, Jianhang Hu^{1,2}, Peilin Chen^{1,2}, Zou Xiang⁴, Qiu Rao^{3✉}, Xiaodong Han^{1,2✉}

1. Immunology and Reproduction Biology Laboratory & State Key Laboratory of Analytical Chemistry for Life Science, Medical School, Nanjing University, Nanjing 210093, China.
2. Jiangsu Key Laboratory of Molecular Medicine, Nanjing University, Nanjing 210093, China.
3. Department of Pathology, Jinling Hospital, Nanjing University School of Medicine, Nanjing 210002, China.
4. Department of Health Technology and Informatics, Faculty of Health and Social Sciences, The Hong Kong Polytechnic University, Hung Hom, Kowloon, Hong Kong China.

#These authors contributed equally to this study.

✉ Corresponding authors: Qiu Rao, E-mail: raoqiu1103@126.com, Department of Pathology, Jinling Hospital, Nanjing 210002, China; Xiaodong Han, E-mail: hanxd@nju.edu.cn, Immunology and Reproduction Biology Laboratory, Medical School, Nanjing University, Nanjing 210093, China, Tel/fax: +86 25 83686497.

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Abstract

Background: Since primary prostate cancer (PCa) can advance to the life-threatening metastatic PCa, exploring the molecular mechanisms underlying PCa metastasis is crucial for developing the novel targeted preventive strategies for decreasing the mortality of PCa. RNA N⁶-methyladenosine (m⁶A) is an emerging regulatory mechanism for gene expression and its specific roles in PCa progression remains elusive.

Methods: Western blotting, quantitative real-time PCR and immunohistochemical analyses were used to detect target gene expression in PCa cells *in vitro* and prostate tissues from patients. RNA immunoprecipitation was conducted to analyze the specific binding of mRNA to the target protein. Migration and invasion assays were used to assess the migratory capacities of cancer cells. The correlation between target gene expression and survival rate of PCa patients was analyzed based the TCGA database.

Results: We found that total RNA N⁶-methyladenosine (m⁶A) modification levels were markedly upregulated in human PCa tissues due to increased expression of methyltransferase like 3 (METTL3). Further studies revealed that the migratory and invasive capacities of PCa cells were markedly suppressed upon METTL3 knockdown. Mechanistically, METTL3 mediates m⁶A modification of USP4 mRNA at A2696, and m⁶A reader protein YTHDF2 binds to and induces degradation of USP4 mRNA by recruiting RNA-binding protein HNRNPD to the mRNA. Decrease of USP4 fails to remove the ubiquitin group from ELAVL1 protein, resulting in a reduction of ELAVL1 protein. Lastly, downregulation of ELAVL1 in turn increases ARHGDI1 expression, promoting migration and invasion of PCa cells.

Conclusions: Our findings highlight the role of METTL3 in modulating invasion and metastasis of PCa cells, providing insight into promising therapeutic strategies for hindering PCa progressing to deadly metastases.

Key words: METTL3, m⁶A, prostate cancer, metastasis, therapeutic strategies

Introduction

Prostate cancer (PCa) is the most frequently diagnosed cancer and the second leading cause of cancer-related death among males [1, 2]. Given the importance of androgen receptor in prostate carcinogenesis, those patients with low-grade localized tumors are sensitive to androgen-deprivation therapies and are generally curable by surgery, chemotherapy and radiotherapy [3].

However, once primary PCa advances to a deadly metastatic PCa and acquire resistance to androgen-deprivation therapies, the 5-year survival for patients drop to 30% [4, 5]. Thus, hindering PCa metastasis can improve the prognoses of patients. Considering cancer metastasis is a highly complex process facilitated by a series of key events, and thus understanding the underlying molecular mechanism

associated with these events will be beneficial to clinical management [2, 6, 7]. Moreover, identifying and targeting those vital genes involved in PCa metastasis will be essential for developing future therapeutic strategies to control PCa metastatic spread.

N⁶-methyladenosine (m⁶A), the most abundant internal modification in eukaryotic mRNA, mediates gene expression by regulating RNA stability, subcellular localization, translation efficiency, and alternative splicing [8, 9]. m⁶A modification is usually embedded within the consensus motif DRACH (D = A, G, U; R = A, G; H = A, C, U) and usually happens around stop codons [9]. Moreover, m⁶A modification is a dynamic and reversible process, during which the methyltransferase complex consisting of methyltransferase-like 3 (METTL3) and METTL14 catalyze m⁶A modification; however, alkB homolog 5 (ALKBH5) and fat mass and obesity-associated protein (FTO) reverse the m⁶A modifications [9]. Moreover, the prerequisite for m⁶A exerting its effects is recruiting m⁶A-binding proteins to m⁶A sites, among of which YTHDF2 binds to and induce degradation of m⁶A-modified mRNAs by recruiting some RNA-binding proteins to the mRNAs [10, 11].

Recently, growing evidences have supported that m⁶A modification regulates tumor progression including gastric cancer [12], endometrial cancer [13], liver cancer [14], and lung cancer [15]. Thus, we wonder whether m⁶A also plays a key role in modulating PCa progression. In the present study, we observe that METTL3 is upregulated in human androgen-independent PC3 and DU145 cells as well as androgen-sensitive LNCaP cells, accompanied by increased cellular m⁶A levels. We also find that testosterone effectively induces METTL3 expression and m⁶A modification in LNCaP cells but not PC3 and DU145 cells. Further studies demonstrate that m⁶A facilitates PCa metastasis by modulating ARHGDI1 expression via the METTL3-USP4-ELAVL1 cascade.

Materials and Methods

Main reagents and cell culture

RPMI 1640 medium, fetal bovine serum (FBS), and penicillin-streptomycin were purchased from Gibco (Grand Island, NY). The protein synthesis inhibitor cycloheximide (HY-12320), proteasome inhibitor MG132 (HY-13259), DNA transcription inhibitor actinomycin D (HY-17559), and Bobcat339 (HY-111558A) were obtained from Medchemexpress (Monmouth Junction, NJ). The detailed information about antibodies used in this study was shown in **Table S1**. PC3, DU145, and LNCaP cells were obtained from ATCC, and were cultured in RPMI

1640 medium supplemented with 10% FBS and 1% penicillin-streptomycin.

Quantitative real-time PCR (qRT-PCR) and RNA stability analyses

RNA was isolated using Trizol reagent (15596018) (Invitrogen, Carlsbad, CA) following the manufacturer's instructions, and cDNA was generated with iScript cDNA Synthesis Kit (170-8890) (BioRad, Hercules, CA). The qRT-PCR assay was performed using ChamQ SYBR Master Mix (Q311-02) (Vazyme, Nanjing, China) on ViiA 7 Q-PCR System (Applied Biosystems, Waltham, MA). All primers used in the present study were listed in **Table S2**, and *GAPDH* was used as an internal control to measure the relative mRNA levels of targeted genes. RNA stability assay were performed as described previously [16].

Western blotting and coimmunoprecipitation (Co-IP) analyses

Total protein lysates were isolated with RIPA buffer (P0013C) (Beyotime, Shanghai, China), and the concentration of protein was determined with BCA Protein Quantification Kit (Vazyme, E112). Western blotting was performed as described previously [16], and the intensity of the western blotting bands was quantified using Image J software. Furthermore, *GAPDH* was chosen as marker protein in this study. For protein stability assay, cells were treated with cycloheximide at 100 µg/mL for indicated times, after which protein levels were determined by western blotting. Moreover, Co-IP was performed as described previously [14].

m⁶A RIP-qRT-PCR analyses and measurement of cellular m⁶A levels

To assess the m⁶A modification levels of *USP4* mRNA, m⁶A RIP was performed using Magna MeRIP™ m⁶A kit (17-10499) (Millipore Sigma, Billerica, MA) according to manufacturer's instructions with a slight modification. Briefly, the isolated RNAs were fragmented with RNA fragmentation buffer. After saving one tenth of the total RNA as input, the remaining RNAs were immunoprecipitated with m⁶A antibody-conjugated magnetic beads. m⁶A-modified RNAs were washed with immunoprecipitation buffer for three times and then eluted with elution buffer. Total RNAs from elution buffer were recovered with Trizol reagent and then subjected to qRT-PCR assays. The specific primer information about *USP4* was listed in **Table S3**. The relative m⁶A modification levels of *USP4* at different m⁶A modification sites were normalized to input. Moreover, EpiQuik m⁶A RNA Methylation

Quantification Kit (P-9005-96) (Epigentek, Farmingdale, NY) was chosen to measure total cellular m⁶A modification levels according to manufacturer's instructions.

RNA-immunoprecipitation (RIP)-qRT-PCR analyses

RIP analyses were performed as previously established protocols [16]. Briefly, cells were firstly lysed with RIP lysis buffer containing protease and RNase inhibitor, after which cell lysate supernatant was incubated with magnetic beads coated with antibodies against rabbit immunoglobulin G, YTHDF2, HNRNPD, or ELAVL1 overnight at 4 °C. The beads were then washed with IP buffer for three times, followed by being treated with proteinase K (Millipore Sigma, 107393) at 65 °C for 0.5 h. Total RNA from the supernatant was recovered with Trizol reagent. The association between *USP4* transcript and target proteins were measured by qRT-PCR assay, and the data were normalized to input. Specific primer information was listed in **Table S4**.

Human PCa tissue specimens

In this study, a total of 25 pairs of PCa tissues and adjacent normal tissues were collected from department of pathology at Jinling hospital (Nanjing, China) with appropriate informed consent from patients. Clinical information about these patients was provided in **Table S5**.

Immunohistochemical analyses

The human and mouse sections need to be dewaxed and rehydrated, followed by antigen retrieval using 10 mM citrate buffer. After being treated with 3% H₂O₂, the sections were immersed with primary antibodies overnight at 4 °C and then incubated with HRP-conjugated secondary antibody for 1 h at room temperature. The immune complexes were examined using the diaminobenzidine (G1212-200T) (Servicebio, Wuhan, China) according to manufacturer's instructions. The Image-Pro Plus software (Media Cybernetics, MD) was chosen to quantify the protein levels by calculating the integrated optical density per stained area (IOD/area).

Migration and invasion assays

PC3, DU145, and LNCaP cells were plated onto 24-well transwell chambers with polycarbonate membranes (353097) (Corning Life Science, Acton, MA) to assess cell migration and invasion capacities with or without Matrigel (Corning Life Science, 354234). The outer chamber was filled with the complete medium and the upper chamber was seeded with cells at a density of 5 × 10⁵ cells/well in

serum-free medium. After being cultured for 12 h, cells adhering to the underside of the chamber were fixed with 4% paraformaldehyde for 5 min, and then incubated with 100% methanol for 20 min. Finally, migrating cells on the lower surface of the membrane were stained with crystal violet for 15 min, and the cells were counted under an optical microscope.

Plasmid construction and transfection

Lentiviral vectors expressing *METTL3*-specific, *ELAVL1*-specific, or *USP4*-specific shRNA were constructed by OBiO (Shanghai, China). The target sequences of shRNA are as following: sh-*METTL3*#1: 5'-CTGCAAGTATGTTCACTATGA-3'; sh-*METTL3*#2: 5'-CGTCAGTATCTTGGGCAA GTT-3'; sh-*ELAVL1*: 5'-GAGAACGAATTTGATCGTCAACT-3'; sh-*USP4*: 5'-GCCCAGAATGTGCTAAGGTTTCT-3'; sh-control: TTCTCCGAACGIGTCACTG. The *METTL3* expression plasmid was purchased from GENECHM (Shanghai, China). HA-Ubiquitin plasmid was obtained from Addgene (18712) (Watertown, MA). The full-length human *ARHGDI*A cDNA was cloned into the pcDNA3.1 vector (VT1001) (YouBio, Changsha, China) to generate expression plasmid. The *USP4* expression plasmid was generated by cloning the full-length *USP4* cDNA into pcDNA3.1 vector with HA tag (YouBio, VT8001). Wild-type or mutated *USP4*-CDS sequence (2600 ~ 2900) was cloned into pmirGLO luciferase vector (YouBio, VT1439). For sg*METTL3* expressing cells, the target sequences (5'-AGAGTCCAGCTGCTTCTTGT-3') were cloned into pLentiCRISPR v2 [17]. PCa cells were transfected with indicated plasmids. After 48 h of transfection, PC3, DU145, and LNCaP cells were selected with puromycin at 6 µg/ml, 2 µg/ml, and 2 µg/ml, respectively. For the *METTL3* rescue experiment, the *METTL3* expression plasmid was transfected into those PCa cells with *METTL3* knockout.

Methylation-specific PCR

DNA from RWPE-1 cells, PC3 cells, DU145 cells, and LNCaP cells were extracted using DNA extraction kit (DP304) (TIANGEN, Beijing, China). The sodium bisulfite DNA treatments were conducted using a DNA bisulfite conversion kit (TIANGEN, DP215) according to the manufacturer's instructions. PCR amplifications were performed using Phanta® UC Super-Fidelity DNA Polymerase (Vazyme, P507-01) according to the manufacturer's instructions. The PCR primers for methylation-specific PCR were listed in **Table S6**.

Luciferase reporter assay

Cells seeded in 12-well plates were transfected with the pmirGLO luciferase reporter vector fused

with or without the wild-type or mutated USP4-CDS. The firefly luciferase and Renilla luciferase activity in each well were measured by a dual-luciferase reporter assay system (E1910) (Promega, Madison, WI) and the relative luciferase activity was further normalized to Renilla luciferase activity.

Animal experiments

Male BALB/c nude mice and male SCID mice aged 4-5 weeks were purchased from the Model Animal Research Center of Nanjing University, Nanjing, China and were maintained in a pathogen-free animal facility at least 1 week before use. A total of 1×10^6 PC3 cells or DU145 cells suspended in a mixture of 100 μ L PBS and Matrigel were subcutaneously injected into BALB/c nude mice. Tumor weight were measured 2 months after the engraftment. To evaluate the role of METTL3 in tumor metastasis, PC3 cells with or without knockdown of METTL3 were injected into SCID mice through the tail vein (1×10^6 cells per mouse). After eight weeks, mice were sacrificed and their lung tissues were collected for subsequent analyses. The whole experiments on mice were approved by the Animal Care and Use Committee of Nanjing University under the animal protocol number SYXK (Su) 2009-0017.

Flow cytometric analyses

Cell cycle distribution was determined by flow cytometric analyses to investigate whether METTL3 knockdown affect the proliferation of PCa cells. Briefly, PC3 cells were incubated with 500 μ l PI (550825) (BD Pharmingen, San Jose, CA) for 30 min, after which cellular PI absorbance was measured by a FACSCalibur flow cytometer (BD Biosciences, San Jose, CA).

Statistical analyses

The data were analyzed for statistical significance with SPSS 18.0 (SPSS, Chicago, IL). All statistical tests were two-sided, and $p < 0.05$ was considered statistically significant. Differences between groups were tested by One-way analysis of variance (ANOVA) followed by Duncan's *post hoc* test. Data violated the assumption of homogeneity of variances, an alternative Welch's ANOVA with Dunnett's T3 test was carried out.

Results

PCa is associated with upregulated METTL3 expression

In this study, we measured an increase of m⁶A levels in human androgen-independent PC3 and

DU145 PCa cells as well as androgen-sensitive LNCaP cells in comparison to RWPE-1 cells, a normal adult epithelial prostate cell line (Figure 1A-C). We next examined upregulated expression of METTL3 in PCa cells, and no significant changes about METTL14, FTO, and ALKBH5 were observed (Figure 1D-E). Moreover, in three PCa cell lines, relatively lower METTL3 and m⁶A levels were examined in androgen-sensitive LNCaP cells (Figure 1D-E). It is the nuclear but not cytoplasmic-localized METTL3 mediates m⁶A deposition on nuclear RNA [9]. As predicted, we examined marked enrichment of METTL3 but not METTL14, FTO, and ALKBH5 in the nuclei of PCa cells relative to RWPE-1 cells (Figure S1).

Additionally, we found that METTL3 mRNA levels were increased in primary prostate tumor and metastatic tumor tissues in comparison to normal prostate tissues (Figure 1F-I) by analyzing several publicly available Genomic Spatial Event (GSE) databases [18-20]. Through analyzing The Cancer Genome Atlas (TCGA) dataset (<http://ualcan.path.uab.edu/index.html>) [21], we further confirmed that METTL3 mRNA levels were significantly increased in PCa tissues (Figure 1J). Survival curve analyses (gepia.cancer-pku.cn) revealed patients with higher METTL3 mRNA levels exhibited much shorter relapse-free survival rate (Figure 1K) [22]; however, no significant correlation between METTL14, FTO, or ALKBH5 expression and survival rate was observed (Figure 1L-N).

Moreover, we observed upregulated m⁶A levels in human PCa tissues relative to normal tissues, accompanied by a significant increase of METTL3 protein levels (Figure 1O-R); however, no remarkable changes about METTL14, ALKBH5, and FTO were observed (Figure S2).

METTL3 is required for invasion and migration of PCa cells

In this study, we established stable METTL3-knockdown PCa cells with two independent shRNA sequences (sh-METTL3#1 and sh-METTL3#2) (Figure 2A-B and Figure S3A-B). As predicted, knockdown of METTL3 significantly reduced m⁶A modification levels in PCa cells (Figure 2C and Figure S3C). Considering that sh-METTL3#1 exhibited higher efficiency than sh-METTL3#2 in reducing METTL3 protein levels, and thus we chosen sh-METTL3#1 for further analyses. Moreover, we observed that METTL3 knockdown did not affect FTO, ALKBH5, and METTL4 levels in PCa cells (Figure S3D-E).

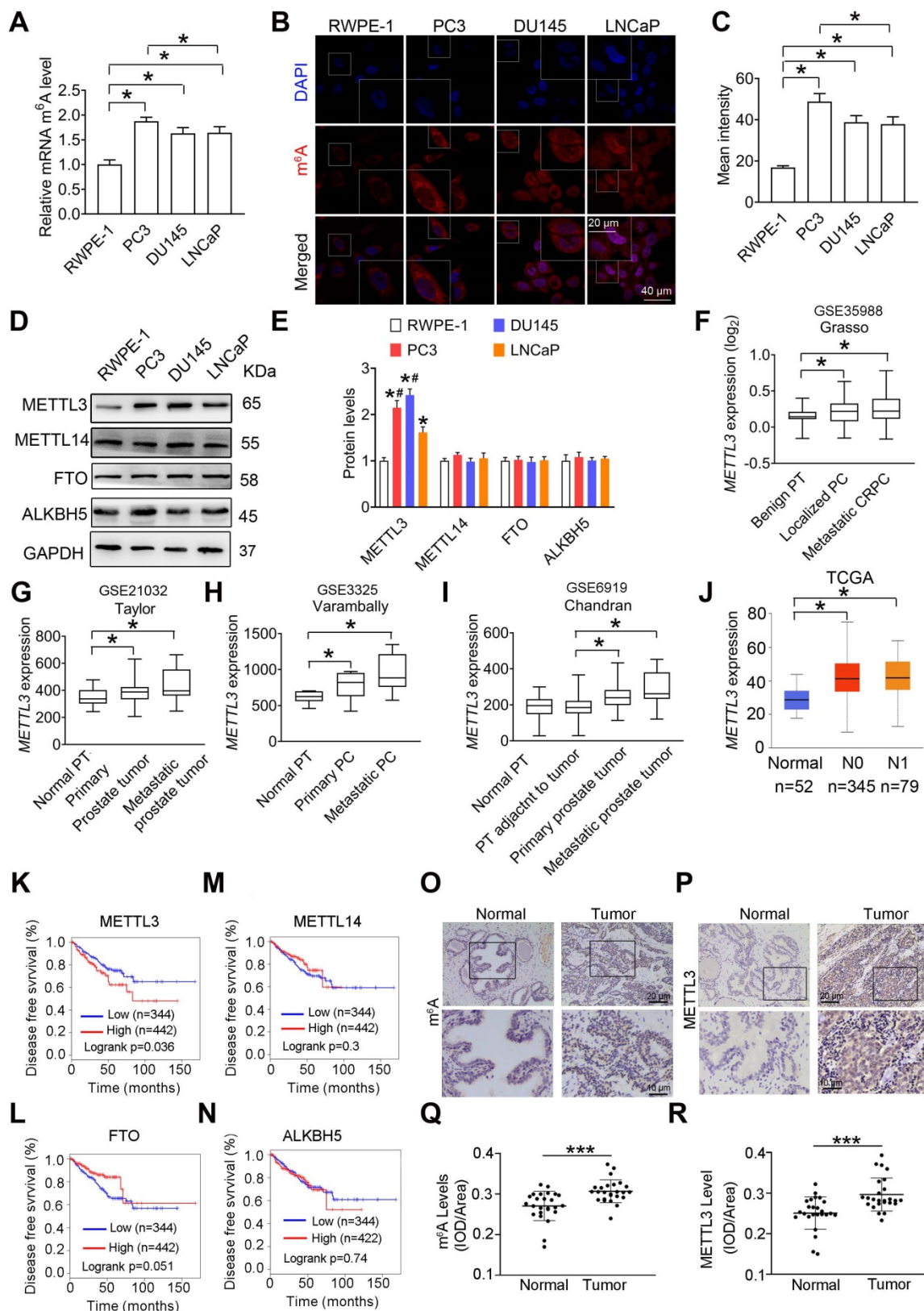


Figure 1. The m⁶A mRNA methylation mediates prostate cancer (PCa) progression. **A**, Relative m⁶A levels in RWPE-1, PC3, DU145, and LNCaP cells were measured by ELISA-based m⁶A quantitative analyses. Data were presented as means ± SEM (n = 3), *p < 0.05. **B-C**, Cellular m⁶A levels were assessed by immunofluorescence staining, and fluorescence intensity was presented as means ± SEM (n = 3), *p < 0.05. **D-E**, Protein levels of m⁶A regulatory enzymes in indicated cells were determined by western blotting and quantitatively analyzed. Data were presented as means ± SEM (n = 3), *p < 0.05 vs. the RWPE-1 cells. **F-J**, The METTL3 mRNA levels were analyzed based on 4 different GSE datasets and the TCGA database (*p < 0.05). PT: prostate tissue; PC: prostate cancer; CRPC: castration-resistant prostate cancer. **K-N**, Association of METTL3, FTO, METTL14, and ALKBH5 mRNA levels with survival of PCa patients was performed using Kaplan-Meier survival curve analysis methods based on the TCGA database. **O-R**, Immunohistochemical staining was performed to evaluate the m⁶A levels and expression of METTL3 in 25 paired human PCa tissues and their adjacent normal prostate tissues. The m⁶A levels (**Q**) and METTL3 protein levels (**R**) were analyzed by calculating the integrated optical density per area (IOD/area). Data were presented as means ± SEM (n = 25), *** p < 0.001.

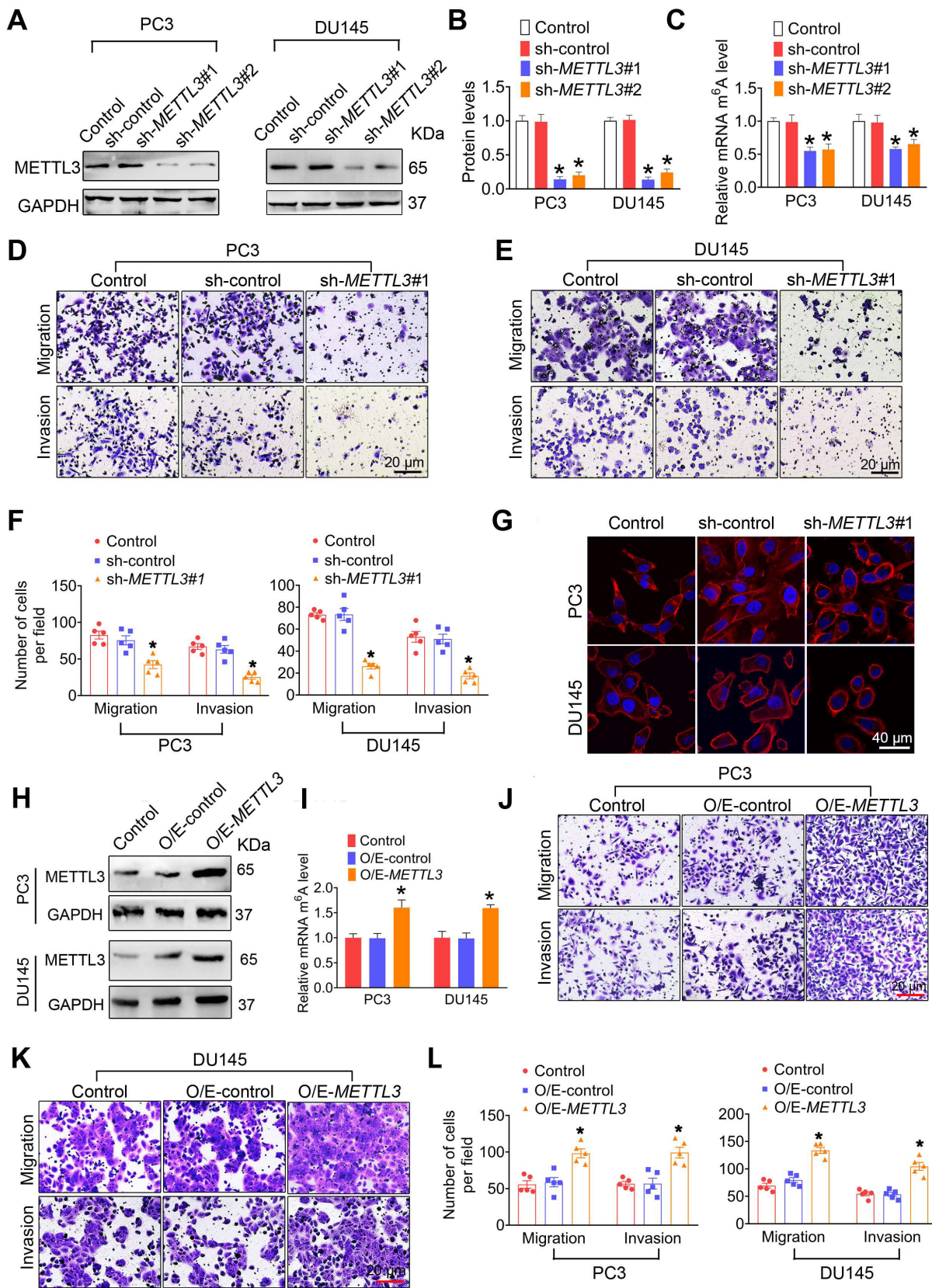


Figure 2. METTL3 promotes prostate cancer (PCa) metastasis. A-C, Two independent shRNA sequences targeting *METTL3* (sh-METTL3#1 and sh-METTL3#2) were separately transfected into PC3 or DU145 cells. Protein levels of *METTL3* were determined by western blotting and quantitatively analyzed (A, B), and cellular m⁶A levels were measured by ELISA-based m⁶A quantitative analyses (C). D-F, The migration and invasion abilities of indicated cells were evaluated. Representative images (D, E) and quantification (F) of the cell migration and invasion assay results were shown. G, PCa cells were stained with rhodamine phalloidin (red) and DAPI (blue). H-L, Overexpression constructs of *METTL3* (O/E-METTL3) were stably transfected into PC3 and DU145 cells, respectively. Protein levels of *METTL3* were determined by western blotting (H), and cellular m⁶A levels were measured by ELISA-based m⁶A quantitative analyses (I). The migration and invasion abilities of indicated cells were assessed, and the representative images of the cell migration and invasion assay results were shown (J-L). Data were presented as means ± SEM where relevant, *p < 0.05 vs. the control cells.

We next found that METTL3 knockdown markedly inhibited migration and invasion of PC3 [42.2 ± 10.89 vs. 82.6 ± 10.69 for migration assay, 25.2 ± 6.05 vs. 67.0 ± 7.97 for invasion assay], DU145 [26.20 ± 4.50 vs. 73.2 ± 3.54 for migration assay, 17.4 ± 5.31 vs. 53.0 ± 9.76 for invasion assay], and LNCaP cells [15.6 ± 2.96 vs. 33.6 ± 6.83 for migration assay, 12.2 ± 3.49 vs. 35.8 ± 5.71 for invasion assay] (**Figure 2D-F and Figure S3F-G**). Consistently, phalloidin-staining assay revealed that lamellipodia were formed depending on METTL3 expression in PCa cells (**Figure 2G**). Moreover, we observed that METTL3 overexpression brought about increased m⁶A modification levels (**Figure 2H-I and Figure S3H-J**), and concurrent enhanced migratory and invasive capacities in PC3 [98.2 ± 12.12 vs. 55.6 ± 10.23 for migration assay, 99.2 ± 14.52 vs. 56.6 ± 6.28 for invasion assay], DU145 [133.6 ± 10.89 vs. 69.4 ± 8.71 for migration assay, 104.6 ± 14.22 vs. 54.6 ± 6.97 for invasion assay], and LNCaP cells [44.2 ± 5.08 vs. 31.2 ± 6.24 for migration assay, 38.6 ± 4.54 vs. 28.8 ± 6.43 for invasion assay] (**Figure 2J-L and Figure S3K-L**). To further confirm the role of METTL3 in mediating the PCa metastasis, we transfected the *METTL3* expression plasmid into those PCa cells with knockout of METTL3, and we found that *METTL3* overexpression effectively rescued the *METTL3* deficiency-induced suppressed migratory and invasive capacities of PCa cells (**Figure S4**).

In this study, we found that testosterone robustly induced METTL3 expression and upregulation of cellular m⁶A levels in androgen-sensitive LNCaP cells but not androgen-independent PC3 and DU145 cells (**Figure S5A-C**). Moreover, we identified that testosterone markedly upregulated migratory and invasive abilities of LNCaP but not PC3 and DU145 cells (**Figure S5D-H**).

ARHGDI A is an METTL3 downstream effector for PCa metastasis

We assessed the effects of METTL3 knockdown on the expression of invasion- and migration-related proteins, including Rho GDP dissociation inhibitor α (ARHGDI A), Rho GDP dissociation inhibitor β (ARHGDI B), ras-related C3 botulinum toxin substrate 1, 2, 3 (RAC1,2,3), matrix metalloproteinase-9 (MMP-9), and ras homolog gene family, member A (RHOA) [23-27]. The results demonstrated that METTL3 knockdown markedly reduced expression of ARHGDI A (**Figure 3A**). Moreover, METTL3 knockdown did not affect expression of epithelial-mesenchymal transition (EMT)-related genes, including E-cadherin, vimentin, Zinc finger E-box binding homeobox 1 (ZEB1), and ZEB2 (**Figure 3B**).

We found that ARHGDI A protein levels were increased in human PCa tissues relative to adjacent normal tissues (**Figure 3C-D**). Additionally, ARHGDI A mRNA levels were also markedly upregulated in PCa tissues (**Figure 3E-G**). Survival curve analyses revealed PCa patients with higher ARHGDI A expression exhibited much shorter relapse-free survival (**Figure 3H**). To further verify the biological contribution of ARHGDI A in METTL3-mediated invasion of PCa cells, the constructs that express ARHGDI A were stably transfected into PCa cells (**Figure 3I-J**). We observed that ectopic expression of ARHGDI A effectively reversed METTL3 knockdown-induced reduced migratory and invasive capacities in PC3 and DU145 cells (**Figure 3K-N**).

METTL3 stabilizes ARHGDI A mRNA by regulating ELAVL1 expression

We found that ARHGDI A mRNA levels were markedly reduced in response to METTL3 knockdown (**Figure 4A and Figure S6A**). Moreover, comparable ARHGDI A promoter transcriptional activity between METTL3 knockdown cells and control cells was examined, excluding the possibility that METTL3 downregulation inhibited ARHGDI A transcription (**Figure 4B and Figure S6B**). We then investigated the possibility that METTL3 knockdown impaired the stability of ARHGDI A mRNA. As expected, we observed elevated decay rates of ARHGDI A mRNA upon the knockdown of METTL3 (**Figure 4C and Figure S6C**).

To determine the upstream regulators mediating ARHGDI A mRNA stabilization by METTL3, we examined expression of heterogeneous nuclear ribonucleoprotein D (HNRNP D), ELAV like RNA-binding protein 1 (ELAV1), and nucleolin (NCL), which were identified to bind to mRNA and regulate mRNA stability. The results showed that ELAVL1 was upregulated in PCa cells after knockdown of METTL3 (**Figure 4D-E and Figure S6D-E**). Accordingly, we determined reduced ELAVL1 expression in human PCa tissues compared to normal prostate tissues (**Figure 4F-G**). It was noted that no significant correlation of ELAVL1 mRNA levels with survival of PCa patients was observed (**Figure 4H**).

We observed that ELAVL1 knockdown markedly increased ARHGDI A expression and promoted migration and invasion of PCa cells (**Figure 4I-K and Figure S6F-H**). Additionally, the knockdown of ELAVL1 effectively attenuated METTL3 knockdown-induced ARHGDI A reduction and concurrently impaired invasion ability (**Figure 4L-O and Figure S6I-L**). Consistently, ELAVL1 knockdown markedly slowed down the decay rates of

*ARHGDI*A mRNA (Figure 4P and Figure S6M). As expected, *METTL3* knockdown significantly enhanced the interaction between *ELAVL1* and

*ARHGDI*A mRNA, and the interaction was significantly inhibited upon knock down of *ELAVL1* (Figure 4Q and Figure S6N).

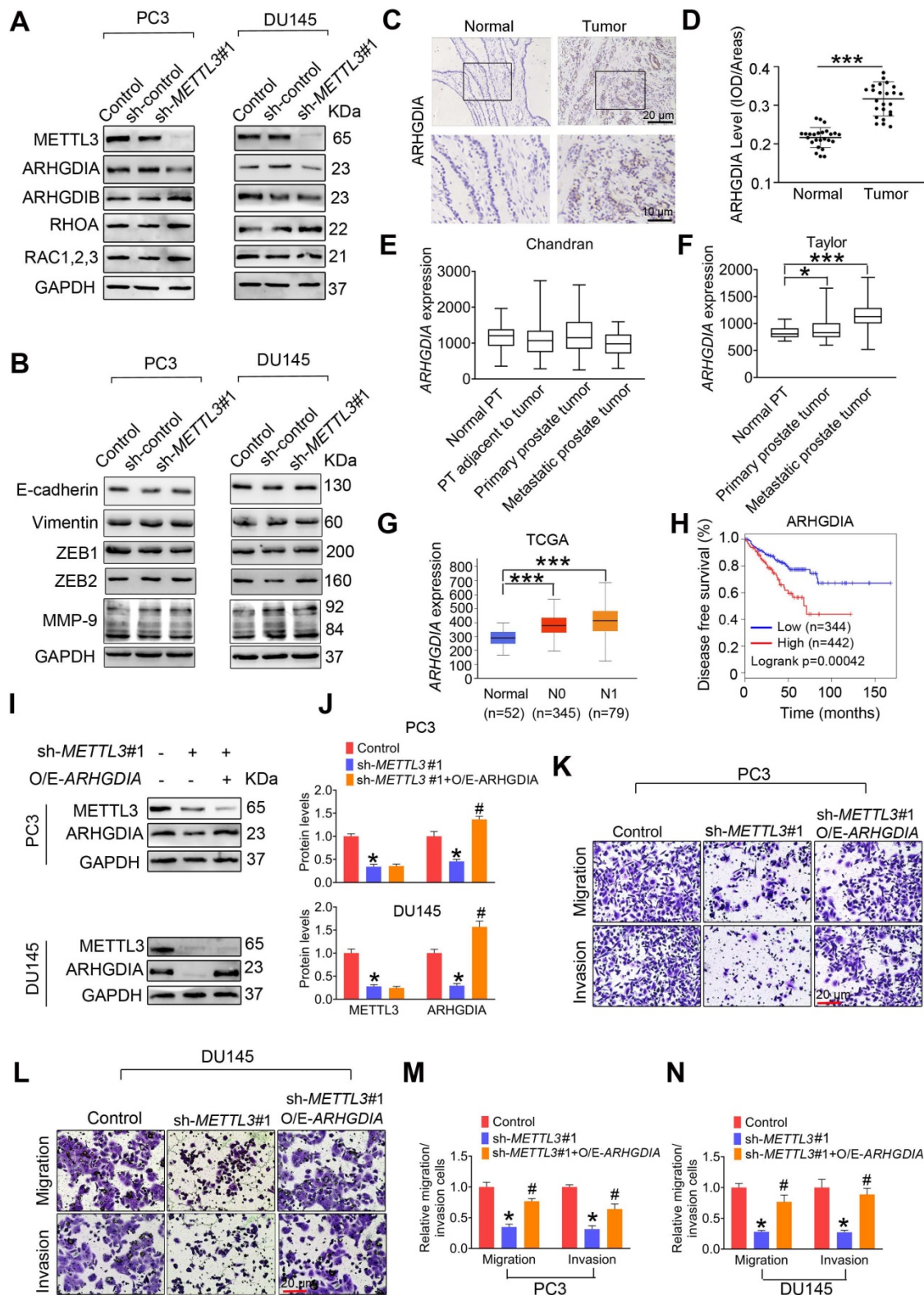


Figure 3. ARHGDI A mediates METTL3 promotion of prostate cancer (PCa) metastasis. A-B, PC3 and DU145 cells were transfected with shRNA targeting *METTL3* (sh-METTL3), respectively. Western blotting was used to examine protein expression of invasion- and EMT-related genes. C-D, Immunohistochemical staining was performed to evaluate the expression of ARHGDI A in 25 paired human PCa tissues and their adjacent normal prostate tissues, and ARHGDI A protein levels were analyzed by calculating the integrated optical density per area (IOD/area). Data were presented as means ± SEM (n = 25), *** p < 0.001. E-G, The ARHGDI A mRNA levels were analyzed in 2 different GSE datasets and the TCGA database, *p < 0.05. PT: prostate tissue. H, Correlation between ARHGDI A mRNA expression and survival of PCa patients was performed

using Kaplan-Meier survival curve analysis methods based on the TCGA database. **I-N**, PCa cells were transfected with sh-METTL3#1 before transfection with pcDNA3.1-ARHGDI (O/E-ARHGDI). Protein levels of ARHGDI were determined by western blotting and quantitatively analyzed (**I, J**). The migration and invasion abilities of indicated cells were evaluated. Representative images (**K, L**) and quantification (**M, N**) of the cell migration and invasion assay results were shown. Data were presented as means \pm SEM (n = 3), * p < 0.05 vs. the control cells, # p < 0.05 vs. the sh-METTL3-treated cells.

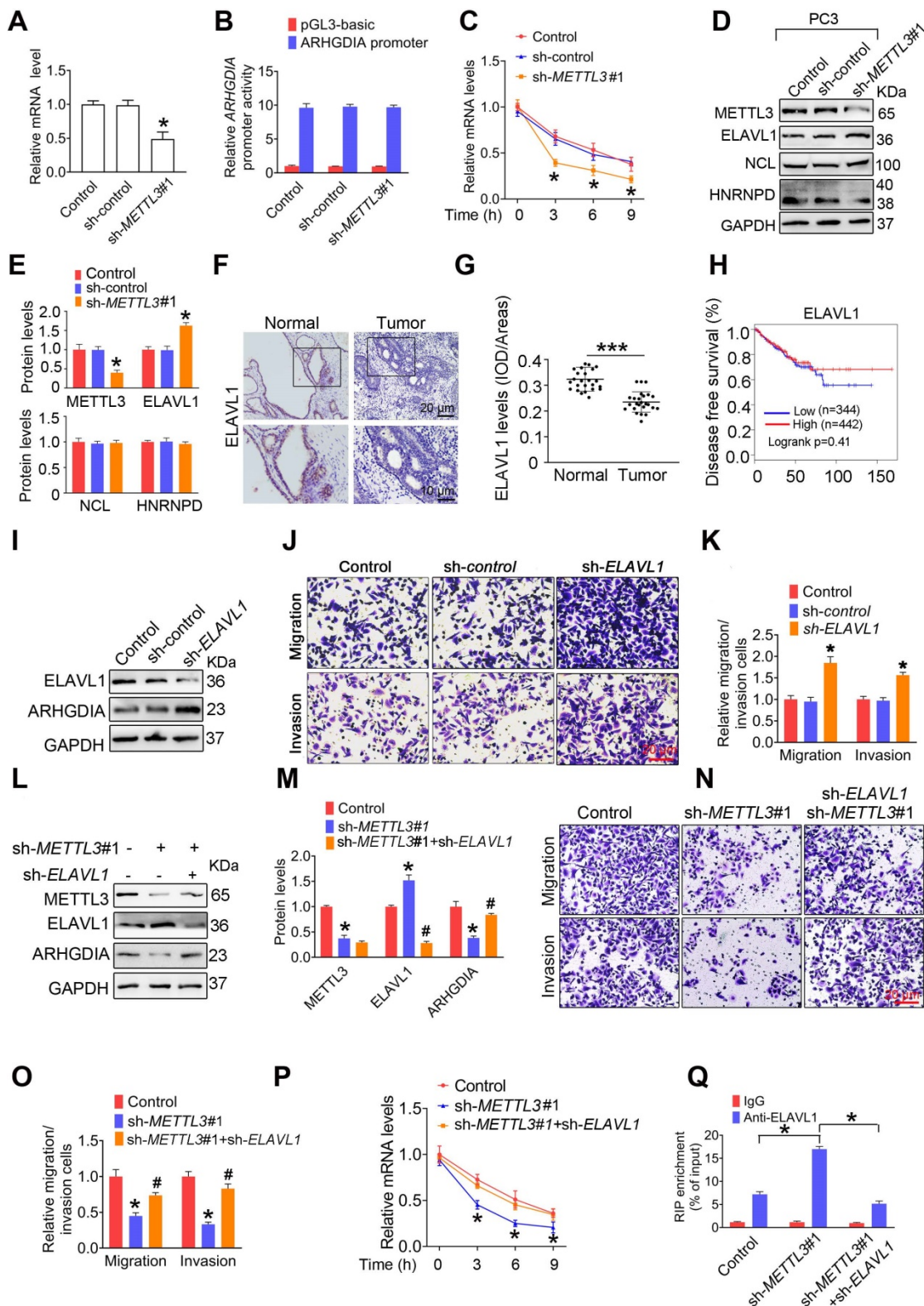


Figure 4. ELAVL1 reduces ARHGDI mRNA stability and prostate cancer (PCa) metastasis. **A-E**, PC3 cells were transfected with shRNA targeting METTL3 (sh-METTL3). The ARHGDI mRNA levels in indicated cells were determined by qRT-PCR assay (**A**). The ARHGDI promoter constructs were transfected into PC3 cells, and luciferase activity was measured (**B**). PC3 cells were treated with actinomycin D (5 μ g/mL) for 2 h, followed by measurement of ARHGDI mRNA levels at indicated times (**C**). Protein levels of ELAVL1, NCL, and HNRNP D were determined by western blotting and quantitatively analyzed (**D, E**). Data were presented as means \pm SEM (n = 3), * p < 0.05 vs. the control cells. **F-G**, Immunohistochemical staining was performed to evaluate the expression of ELAVL1 in 25 paired human PCa tissues and their adjacent normal prostate tissues (**F**), and ELAVL1 protein levels (**G**) were analyzed by calculating the integrated optical density per area (IOD/area). Data were presented as means \pm SEM (n = 25), *** p

< 0.001. **H**, Correlation between *ELAVL1* mRNA expression and survival of PCa patients was analyzed using Kaplan-Meier survival curve analysis methods based on the TCGA database. **I-K**, PC3 cells were transfected with sh-*ELAVL1*. Protein levels of *ELAVL1* and *ARHGDI1* were determined by western blotting (**I**). The migration and invasion abilities of indicated cells were evaluated. Representative images (**J**) and quantification of the cell migration and invasion assay results were shown (**K**). Data were presented as means \pm SEM ($n = 5$), * $p < 0.05$ vs. the control cells. **L-P**, PCa cells were transfected with sh-*METTL3* before transfection with sh-*ELAVL1*. Protein levels of *METTL3*, *ELAVL1*, and *ARHGDI1* were determined by western blotting and quantitatively analyzed (**L, M**). The migration and invasion abilities of indicated cells were evaluated. Representative images and quantification of the cell migration and invasion assay results were shown (**N, O**). PC3 cells were treated with actinomycin D (5 μ g/mL) for 2 h, followed by measurement of *ARHGDI1* mRNA levels at indicated times (**P**). Data were presented as means \pm SEM ($n = 3$), * $p < 0.05$ vs. the control cells, # $p < 0.05$ vs. the sh-*METTL3*-treated cells. **Q**, *ELAVL1* was immunoprecipitated, followed by qRT-PCR assay to evaluate the association of the *ARHGDI1* transcripts with *ELAVL1* protein. Data were presented as means \pm SEM ($n = 3$), * $p < 0.05$.

METTL3 induces degradation of ELAVL1 protein by reducing USP4 protein levels

In this study, we examined that *METTL3* knockdown did not affect *ELAVL1* mRNA levels in PCa cells (**Figure 5A and Figure S7A**), consistent with the survival analyses results that no significant association of *ELAVL1* mRNA levels with survival rate of PCa patients were examined (**Figure 4H**). Thus, we speculated that the increase of protein stability or enhancement of translation efficiency might upregulate *ELAVL1* protein levels in PCa cells upon *METTL3* knockdown. Moreover, we observed that degradation rate of *ELAVL1* was markedly attenuated in PCa cells in response to *METTL3* knockdown (**Figure 5B-C and Figure S7B-C**), while no significant changes of synthesis rate of *ELAVL1* was observed (**Figure 5D-E and Figure S7D-E**). Taken together, these results lead us to believe that *METTL3* might reduce *ELAVL1* protein levels by inducing its degradation.

To investigate whether ubiquitin-proteasome pathway participates in *ELAVL1* degradation, we firstly assessed the effect of *METTL3* knockdown on ubiquitination levels of *ELAVL1*. We found that knocking down *METTL3* remarkably reduced the ubiquitination level of *ELAVL1* (**Figure 5F and Figure S7F**). Given that protein deubiquitination is regulated by deubiquitinating enzymes (DUBs) and ubiquitin-specific protease (USP) family is the largest family of DUBs [28, 29], we then compared the mRNA levels of a total of 42 USP members by analyzing the TCGA data (**Table S7**). We found that *USP4*, *USP8*, *USP11*, *USP25* and *USP28* mRNA levels were markedly decreased in PCa tissues in comparison to normal prostate tissues, while *USP3*, *USP7*, *USP22* and *USP40* mRNA levels were increased. Based on the fact that *METTL3* knockdown decrease ubiquitination level of *ELAVL1* and *METTL3* protein levels were upregulated in PCa tissue, we speculate that expression of the target USP members involving *ELAVL1* regulation might be decreased in PCa tissues. And thus, we chose *USP4*, *USP8*, *USP11*, *USP25* and *USP28* for further analyses, and western blotting revealed that *METTL3* knockdown markedly increased *USP4* protein levels (**Figure 5G-H and Figure S7G-H**). Moreover, we determined reduced *USP4* levels in human PCa tissues relative to normal

prostate tissues (**Figure 5I-J**). Consistently, a poorer survival rate was observed to be associated with lower *USP4* mRNA levels (**Figure 5K-L**).

Further studies identified that the *USP4* knockdown effectively attenuated *METTL3* knockdown-induced *ELAVL1* increase, *ARHGDI1* reduction, and concurrent inhibited migratory and invasive capacities of PCa cells (**Figure 5M-P and Figure S7I-K**). Consistently, *USP4* knockdown markedly reduced protein stability of *ELAVL1* (**Figure 5Q-R and Figure S7L-M**).

To understand the molecular mechanism accounting for the regulation of *ELAVL1* by *USP4*, we firstly assessed the interaction between *USP4* and *ELAVL1* in PCa cells. The results showed that *METTL3* knockdown markedly enhanced the interaction between *USP4* and *ELAVL1* (**Figure 5S and Figure S7N**). Additionally, the overexpression of *USP4* effectively decreased the ubiquitination levels of *ELAVL1* (**Figure 5T and Figure S7O**). Taken together, our results demonstrated that *USP4* upregulates *ELAVL1* expression by removing the ubiquitin group from *ELAVL1* protein.

m⁶A methylation reduces USP4 protein levels by affecting USP4 mRNA stability

We next found that *METTL3* knockdown significantly upregulated *USP4* mRNA levels in PCa cells (**Figure 6A**). Moreover, *USP4* promoter transcriptional activity was not affected upon *METTL3* knockdown (**Figure 6B**), excluding the possibility that *METTL3* downregulation inhibited *USP4* transcription. By contrast, we observed that knock down of *METTL3* slowed down degradation rate of *USP4* mRNA (**Figure 6C**). Given the key role of m⁶A mRNA methylation in affecting mRNA stability, it was reasonable to speculate that *METTL3* knockdown reduces the m⁶A modification levels in *USP4* mRNA and thus increase *USP4* mRNA levels by attenuating the inhibitory effect of m⁶A modification on *USP4* mRNA stability. Consistent with this hypothesis, it was knockdown of *YTHDF2* but not *YTHDF1* remarkably upregulated *USP4* protein levels to a similar extent as *METTL3* knockdown in PCa cells (**Figure 6D-E**). Furthermore, *YTHDF2* enrichment at *USP4* transcripts was markedly reduced in PC3 cells upon knockdown of *METTL3* (**Figure 6F and Figure S8A**).

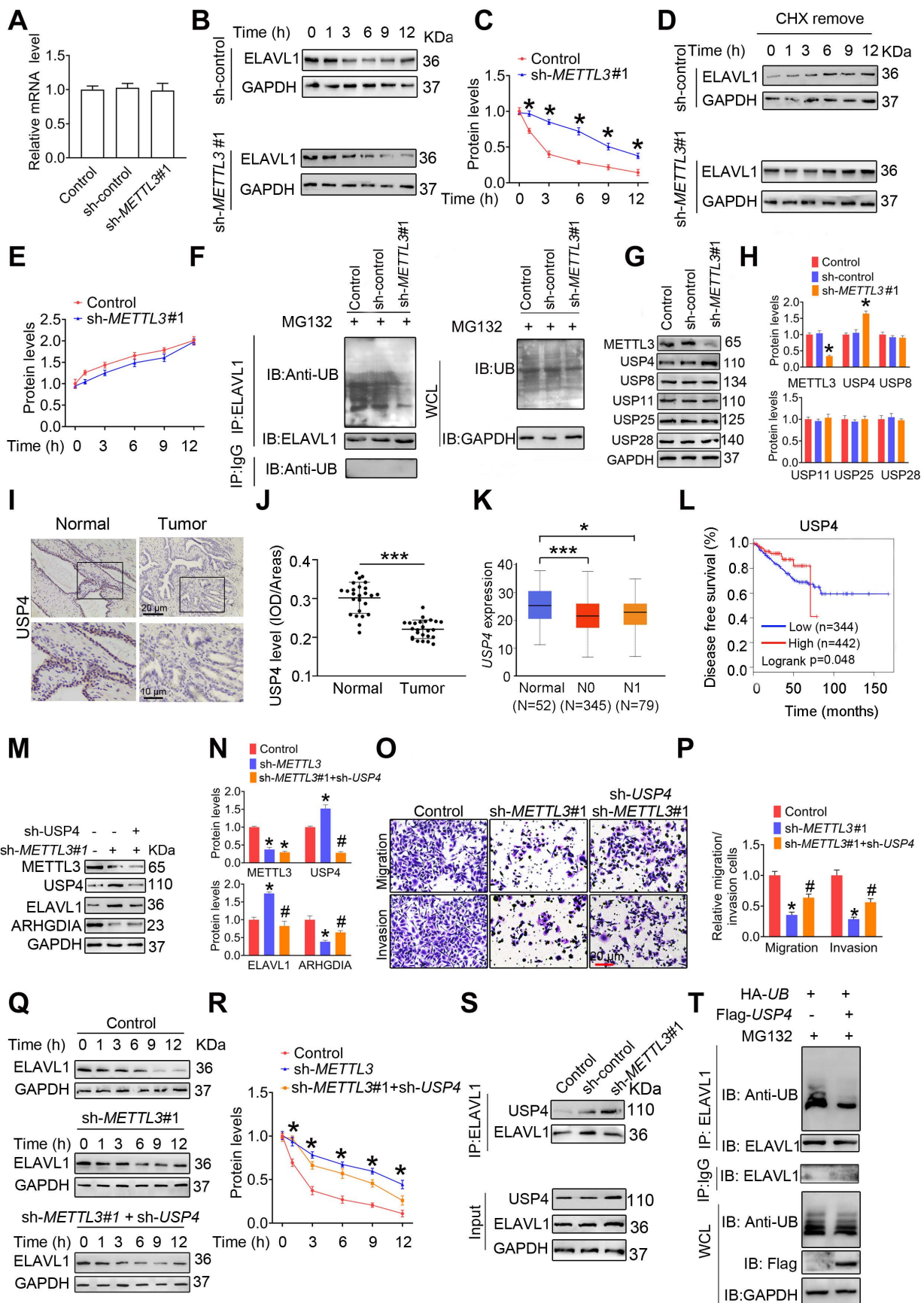


Figure 5. USP4 is an METTL3 downstream effector and mediates ELAVL1 protein stability. **A**, The *ELAVL1* mRNA levels in indicated cells were determined by qRT-PCR assay. **B-C**, The indicated PC3 cells were pretreated with cycloheximide (CHX, 10 μg/mL) for 3 h, followed by measurement of *ELAVL1* protein levels at indicated times. **D-E**, PC3 cells were treated with cycloheximide (CHX) for 12 h. After washing out CHX, cells were cultured for the indicated times. *ELAVL1* synthesis levels were determined by western blotting and quantitatively analyzed. **F**, PC3 cells were treated with MG132 for 6 h. Lysates from the indicated cells were subjected to

coimmunoprecipitation (Co-IP) assay with anti-ELAVL1 antibody, and the blots were then probed with anti-ubiquitin (UB) antibody for detection of ubiquitination of ELAVL1. **G-H**, Protein levels of ELAVL1, NCL, and HNRNP D in indicated cells were determined by western blotting and quantitatively analyzed. Data were presented as means \pm SEM (n = 25), * p < 0.001 vs. the control cells. **I-J**, Immunohistochemical staining was performed to evaluate the expression of USP4 in 25 paired human PCa tissues and their adjacent normal prostate tissues (**I**), and USP4 protein levels (**J**) were analyzed by calculating the integrated optical density per area (IOD/area). Data were presented as means \pm SEM (n = 25), *** p < 0.001. **K**, The USP4 mRNA levels were analyzed in the TCGA database. **L**, Correlation between USP4 mRNA expression and survival of PCa patients was analyzed using Kaplan-Meier survival curve analysis methods based on the TCGA database. **M-R**, PC3 cells were transfected with sh-METTL3 before transfection with sh-ELAVL1. Protein levels of METTL3, USP4, ELAVL1, and ARHGDI A were determined by western blotting and quantitatively analyzed. The migration and invasion abilities of indicated cells were evaluated. Representative images and quantification of the cell migration and invasion assay results were shown. The indicated cells were pretreated with CHX for 3 h, followed by measurement of ELAVL1 protein levels at indicated times. **S**, Lysates from the indicated cells were subjected to Co-IP with anti-ELAVL1, and the blots were probed with anti-USP4 antibody. **T**, PC3 cells were transfected with indicated plasmids, and ubiquitination of ELAVL1 was measured by Co-IP assay. Data were presented as means \pm SEM (n = 3), * p < 0.05 vs. the control cells, # p < 0.05 vs. the sh-METTL3-treated cells. WCL: whole cell lysate.

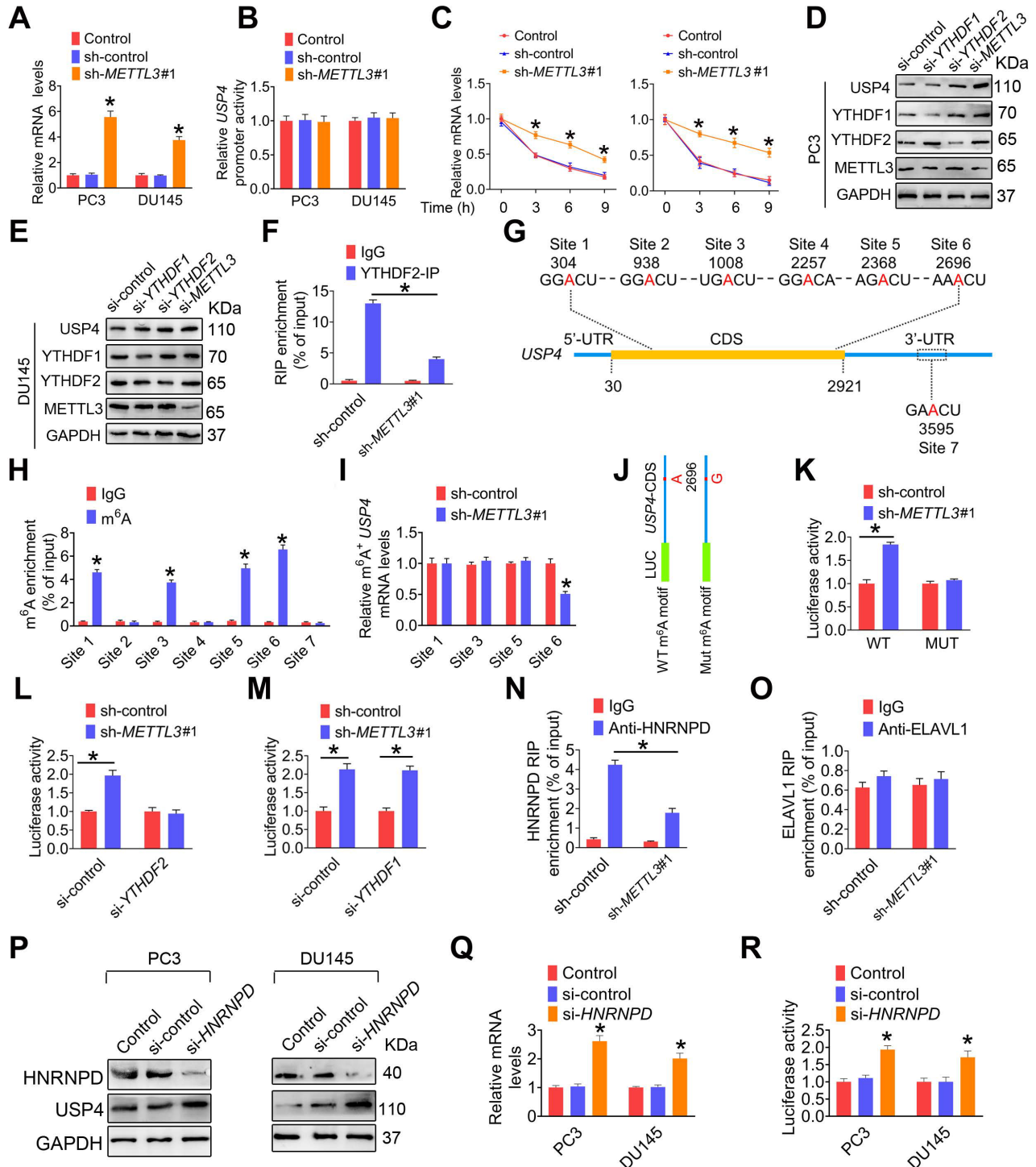


Figure 6. m6A induces decay of the USP4 transcript in prostate cancer (PCa) cell. **A-C**, PC3 and DU145 cells were transfected with shRNA targeting METTL3 (sh-METTL3), respectively. The USP4 mRNA levels in indicated cells were determined by qRT-PCR assay (**A**). The USP4 promoter constructs were transfected into indicated cells, and luciferase activity was measured (**B**). PCa cells were treated with actinomycin D (5 μ g/mL) for 2 h, followed by measurement of USP4 mRNA levels at indicated times (**C**). Data were presented as means \pm SEM (n = 3), * p < 0.05 vs. the control cells. **D-E**, PC3 cells (**D**) and DU145 cells (**E**) were treated with indicated siRNAs, and cell lysates were subject to western blotting. **F**, Lysates from the PC3 cells were subjected to immunoprecipitation with anti-YTHDF2, and the association of the USP4 transcript with

YTHDF2 was determined by qRT-PCR. Data were presented as means \pm SEM (n = 3), * p < 0.05. **G**, Schematic representation of the position of m⁶A motifs within *USP4* transcript. **H**, Abundance of *USP4* transcript among mRNA immunoprecipitated with anti-m⁶A antibody was measured by qRT-PCR and normalized to input. Data are presented as means \pm SEM (n = 3). * p < 0.05 vs. the IgG group. **I**, Abundance of *USP4* transcript among mRNA immunoprecipitated with anti-m⁶A antibody was measured by qRT-PCR. Data are presented as means \pm SEM (n = 3). * p < 0.05 vs. the control cells. **J**, *USP4*-CDS of the wild-type or mutant (A to G) was fused with a luciferase reporter. **K-M**, Luciferase activity of *USP4*-CDS was measured and normalized to Renilla luciferase activity. Data are presented as means \pm SEM (n = 3). * p < 0.05. **N-O**, Lysates from the indicated cells were subjected to immunoprecipitation with anti-HNRNP (N) or anti-ELAVL1 (O), and the association of the *USP4* transcript with each protein was determined by qRT-PCR. Data were presented as means \pm SEM (n = 3), * p < 0.05. **P-R**, PC3 cells and DU145 cells were treated with indicated siRNAs, and cell lysates were subject to western blotting (P). The *USP4* mRNA levels in indicated cells were determined by qRT-PCR (Q). Luciferase activity of *USP4*-CDS in indicated cells was measured and normalized to Renilla luciferase activity (R). Data are presented as means \pm SEM (n = 3), * p < 0.05 vs. the control cells.

We next performed sequence analysis of *USP4* transcript and found six m⁶A modification sites within the CDS region and one m⁶A site in the 3'-UTR (Figure 6G). m⁶A RIP analyses demonstrated that m⁶A was markedly enriched at site 1, site 3, site 5, and site 6 (Figure 6H). Moreover, m⁶A enrichment levels at site 6 were significantly reduced upon knockdown of METTL3 (Figure 6I). We generated a luciferase reporter construct containing a firefly luciferase placed before the *USP4*-CDS (Figure 6J), and subsequent luciferase reporter assays demonstrated that METTL3 knockdown significantly increased the reporter luciferase activity (Figure 6K); however, knock down of YTHDF2 but not YTHDF1 effectively aborted this increase (Figure 6L-M). Additionally, we mutated the potential m⁶A motif AAACU to AAGCU, and METTL3 knockdown cannot upregulate the luciferase activity of the reporter bearing mutated *USP4*-CDS (Figure 6K).

It has been reported that HNRNP and ELAVL1 could bind to m⁶A-modified transcript to regulate pre-mRNA splicing or affect mRNA stability [10, 30]. In this study, we examined that METTL3 knockdown significantly decreased the binding of HNRNP to *USP4* pre-mRNA (Figure 6N and Figure S8B); however, the interaction between ELAVL1 and *USP4* pre-mRNA were not significantly changed (Figure 6O and Figure S8C). Moreover, HNRNP deficiency significantly increased *USP4* expression at mRNA and protein levels, accompanied by a increase of *USP4*-CDS luciferase activity (Figure 6P-R). Taken together, these results suggested m⁶A methylation reduces *USP4* mRNA stability by promoting the binding of HNRNP to *USP4* mRNA.

Dysregulation of METTL3-USP4-ELAVL1-ARHGDI regulatory axis hinders PCa metastasis

To further confirm the role of METTL3-ELAVL1-ARHGDI regulatory axis in regulating PCa metastasis, we examined expression of downstream effector of METTL3 including *USP4*, *ELAVL1*, and *ARHGDI* in those PCa cells with *METTL3* overexpression. As predicted, we found that *METTL3* overexpression induced *ARHGDI* expression, but downregulated *ELAVL1* and *USP4* (Figure S9). We noted that *METTL3* knockdown markedly suppressed xenograft tumor formation and reduced the tumor

weight (Figure 7A-C), consistent with the cell cycle assay results demonstrating that *METTL3* knockdown exerted inhibitory effects on cell proliferation of PC3 cells *in vitro* (Figure S10). Moreover, we observed that *USP4* knockdown effectively alleviated *METTL3* knockdown-induced inhibited xenograft tumor growth (Figure 7A-C). To further confirm the role of *METTL3*-*ELAVL1*-*ARHGDI* regulatory axis in regulating PCa metastasis, we also examined expression of *USP4*, *ELAVL1* and *ARHGDI* in xenograft model with *METTL3*-deficient PCa cells. We observed reduced *METTL3*, m⁶A, and *ARHGDI* levels, but increased *USP4* and *ELAVL1* levels in *in vivo* xenograft nude mice injected with those PC3 cells with *METTL3* knock down (Figure S11); however, the knockdown of *USP4* effectively attenuated *METTL3* deficiency-induced these changes (Figure S11).

Moreover, *METTL3* silencing dramatically suppressed lung metastatic abilities of PC3 cells, as evidenced by the number of lung metastatic nodules (Figure 7D-F); however, the knockdown of *USP4* effectively attenuated *METTL3* deficiency-induced decreased lung metastatic capacity of PC3 cells (Figure 7D-F). As expected, we observed a reduction in *ARHGDI* protein levels, and *USP4* and *ELAVL1* increase in lung metastatic lesions of sh-*METTL3* cells-injected mice, accompanied with *METTL3* and m⁶A decrease (Figure 7G-L), and *USP4* knock down effectively reversed *METTL3* deficiency-induced these changes.

Reduced promoter methylation is associated with increased expression of METTL3

We found that methylation levels in the *METTL3* promoter were significantly reduced in PCa tissues compared to the adjoining normal prostate tissues by analyzing the TCGA database (Figure 8A). We next bioinformatically analyzed the potential CpG island of human *METTL3* promoter, and a total of three CpG island signals were predicted in the *METTL3* promoter region (Figure 8B). Additionally, we found that the methylation levels at site 1 and site 2 were significantly reduced in PCa cells relative to the RWPE-1 cells, accompanied by increased *METTL3* mRNA levels in PCa cells (Figure 8C-F).

It is noted that DNA methylation is dynamically regulated by methyl writing enzymes and methyl erasing enzymes. In this study, we examined that

METTL3 mRNA and protein levels were significantly downregulated in PCa cells following exposure to Bobcat339, the inhibitor of methyl erasing enzymes

(Figure 8G-I). Accordingly, Bobcat339 effectively decreased migratory and invasive capacities of PCa cells (Figure 8J-L).

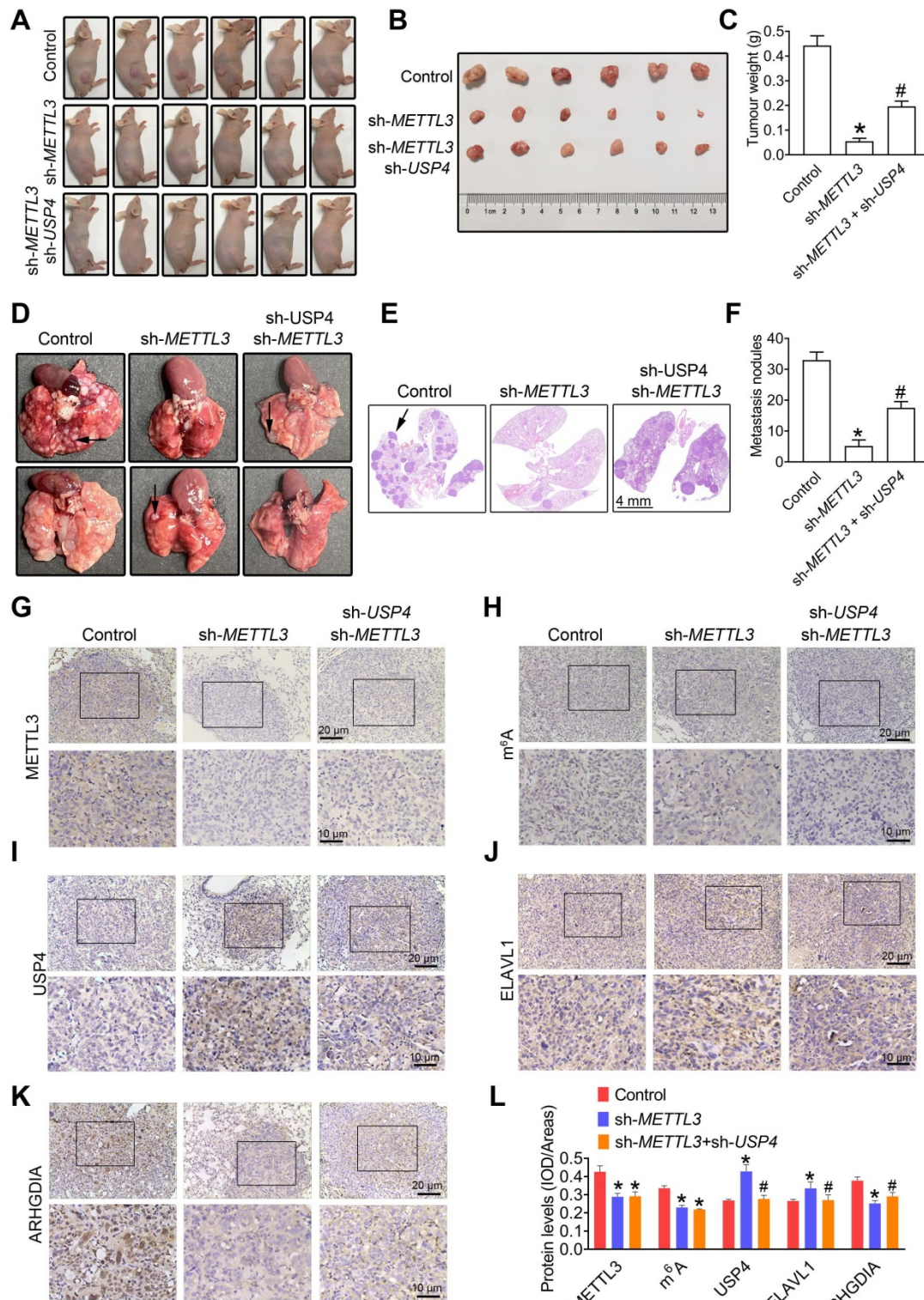


Figure 7. METTL3-USP4-ELAVL1-ARHGDI2 regulation axis promotes prostate cancer (PCa) metastasis. PC3 cells were transfected with shRNA targeting *METTL3* (sh-METTL3) before transfection with sh-USP4. **A-C**, Athymic nude mice were subcutaneously injected into the right axillary region of each mouse with indicated cells. Eight weeks after cell injection, the mice were sacrificed and the tumors were surgically removed and photographed (**A**, **B**), as well as weighed (**C**). **D-F**, The PC3 cells were injected into the SCID mice by tail vein injection. Representative images of metastatic nodules in the lung (**D**) and the H&E staining results were shown (**E**), and the number of metastatic nodules were quantitatively analyzed (**F**). **G-K**, The lung tissues obtained from SCID mice were subjected to immunohistochemical staining assay for evaluating *METTL3* (**G**), m⁶A modification (**H**), USP4 (**I**), ELAVL1 (**J**), and ARHGDI2 (**K**) levels. **L**, The m⁶A levels and the expression levels of the target proteins were analyzed by calculating the integrated optical density per area (IOD/area). Data were presented as means ± SEM (n = 6), * p < 0.05 vs. the control cells, # p < 0.05 vs. the sh-METTL3-treated cells.

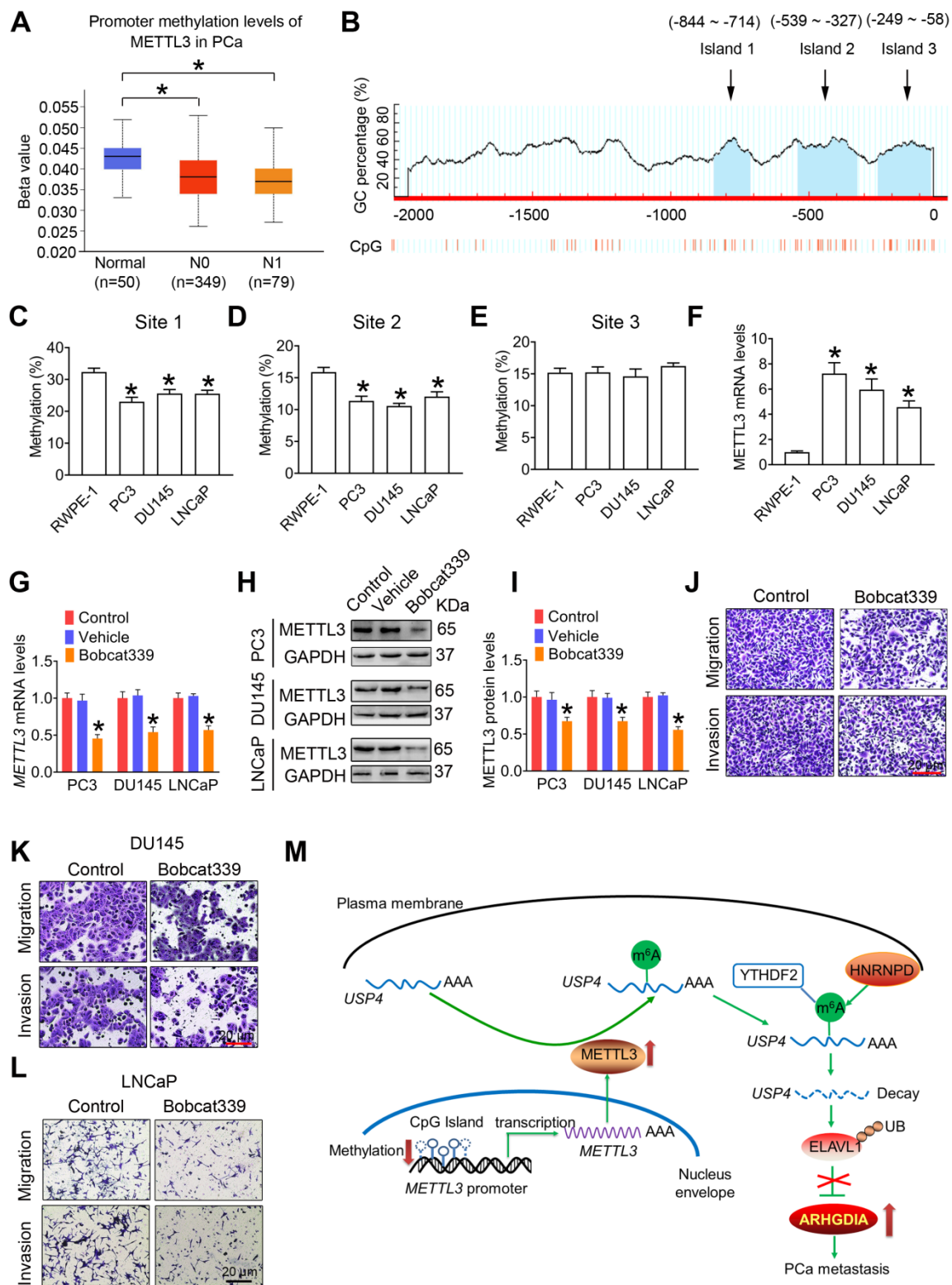


Figure 8. Enhanced METTL3 expression is associated with reduced METTL3 promoter methylation in prostate cancer (PCa) cells. **A**, The methylation levels of *METTL3* promoter were analyzed based on the TCGA database (* $p < 0.05$). **B**, Potential CpG islands in the human *METTL3* promoter were analyzed using online MethPrimer software (<http://www.urogene.org/methprimer/>), and the blue shaded regions indicate the potential CpG islands. **C-E**, The methylation status of the *USP4* promoter at different CpG islands was measured by methylation-specific PCR. Data were presented as means \pm SEM ($n = 5$), * $p < 0.05$ vs. the RWPE-1 cells. **F**, Cellular *METTL3* mRNA levels were determined by qRT-PCR. Data were presented as means \pm SEM ($n = 5$), * $p < 0.05$ vs. the RWPE-1 cells. **G-L**, PCa cells were treated with Bobcat339 at 50 μ M for 48 h. The *METTL3* mRNA levels in PCa cells were determined by qRT-PCR (**G**). The *METTL3* protein levels in PCa cells were determined by western blotting and quantitatively analyzed (**H**, **I**). The migration and invasion abilities of indicated cells were evaluated, and representative images were shown (**J-L**). Data were presented as means \pm SEM ($n = 3$), * $p < 0.05$ vs. the control cells. **M**, A model for the critical link between *METTL3* and PCa metastasis is proposed. Reduced *METTL3* promoter methylation increases *METTL3* expression by promoting its transcription. Upregulation of *METTL3* increases cellular m⁶A mRNA methylation levels, which downregulates *USP4* expression by inducing m⁶A-mediated decay of the *USP4* transcript. *USP4* reduction leads to decreased expression of *ELAVL1* by increasing ubiquitination (UB) of *ELAVL1*, contributing to upregulated expression of *ARHGDI1* by retarding *ELAVL1*-mediated decay of the *ARHGDI1* transcript.

Discussion

Growing evidences have suggested that m⁶A modifications play important roles in mediating progression of various cancers, including hepatocellular carcinoma [31], gastric cancer [12], lung cancer [15], endometrial cancer [13], nasopharyngeal cancer [32], bladder cancer [33], and acute myeloid leukaemia [34]. Moreover, METTL3 has been reported to be upregulated or downregulated in certain cancers, and its specific roles in tumorigenesis remain controversial. Recently, Lin et al. observed upregulated METTL3 levels in lung adenocarcinoma and identified that METTL3 promotes growth of lung cancer cells [15]. By contrast, Liu et al. determined decreased METTL3 expression and concurrent reduction in m⁶A methylation levels in endometrial tumors, which in turn promotes proliferation of endometrial cancer cells [13]. In this study, we determined upregulated expression of METTL3 in PCa tissues, contributing to an increase of m⁶A modification levels. Moreover, we observed that METTL3 knockdown did not change FTO, ALKBH5, and METTL4 levels, excluding the possibility that METTL3 silencing firstly affects expression of other m⁶A-modification related proteins and then reduce m⁶A modification levels of downstream targets. Further studies identified that reduced methylation levels at the CpG islands in promoter might be associated with enhanced *METTL3* transcription in PCa cells, and to confirm which DNA methyltransferase are involved in *METTL3* regulation deserves a separate study.

Accordingly, some researchers have also identified that m⁶A modification is associated with growth and migration of PCa cells [35, 36]. Cai et al. detected upregulated METTL3 levels in PCa cells, and identified that METTL3 promotes PCa growth by regulating hedgehog pathway [35]. Li et al. examined increased YTHDF2 and METTL3 in PCa cells, and proposed that YTHDF2 reduces expression of the tumor suppressors via mediating degradation of the m⁶A-modified mRNAs to induce AKT phosphorylation and subsequent tumor progression in PCa [37]. Ma et al. found that METTL3 activated the Wnt pathway, thereafter, bringing about enhanced migratory ability of the PCa cells [36]. It is noteworthy these studies are mainly concentrated on the the role of METTL3 in regulating upstream signaling pathways associated with PCa progression, and we found that METTL3 directly affect expression of migration-related key protein ARHGDI A to influence PCa migration and invasion. Based on these results, we speculate that this type of multiayed regulation of METTL3 in downstream proteins and

upstream signaling pathways might effectively mediate PCa progression; however, whether the Hedgehog, Akt, and Wnt signaling pathways are involved in METTL3-mediated ARHGDI A expression deserves a separate study. Moreover, these researchers did not validate their findings simultaneously in many PCa cell lines, subcutaneous xenografts of PCa in nude mice, and clinical samples from patients diagnosed with PCa. Compared with these researchers, we also compared the differential expression of METTL3 in androgen-dependent and androgen-independent PCa cells and confirmed the effects of androgen on the METTL3 expression in androgen-dependent LNCaP cells. In this study, we examined higher METTL3 levels in PC3 and DU145 PCa cells than LNCaP cells, which may be associated with deadly metastatic capacity of androgen-independent PCa cells. Moreover, testosterone significantly induced METTL3 expression in LNCaP cells but not PC3 and DU145 cells, indicating that androgen might modulates PCa metastasis by upregulating METTL3 expression and m⁶A levels after binding to AR in PCa cells.

EMT and migration of cancer cells are two important biological phenomenon during the tumorigenic process. In this study, we found that METTL3 knockdown did not affect expression of EMT-related genes, whereas significantly reduced migration-related ARHGDI A expression. ARHGDI A has been reported to modulate several processes during tumorigenesis, including cellular growth, cellular migration and cellular polarity. Up to date, Zhu, et al. has demonstrated that ARHGDI A upregulation inhibits growth of PCa cells, while ARHGDI A deficiency markedly promote the growth of androgen-sensitive LNCaP cells in androgen-deprived conditions [38]. Yamashita, et al. observed upregulated ARHGDI A levels in lymph node metastatic PCa patients and suggested that ARHGDI A may be useful as a diagnostic biomarker for PCa metastasis [23]. To sum up, the role of ARHGDI A in PCa progression remains elusive. In the present study, we observed an increase of ARHGDI A protein levels in human PCa tissues compared with the normal prostate tissues. Moreover, ectopic expression of ARHGDI A effectively attenuated the effect of METTL3 knockdown on invasive ability in PCa cells. Based on these results, we believe that METTL3 could promote PCa metastasis by promoting ARHGDI A expression. Further studies identified that METTL3 stablited *ARHGDI A* mRNA by modulating expression of the mRNA binding protein ELAVL1, which in turn alleviated decay of *ARHGDI A* mRNA. Interestinly, it is noted that ELAVL1 can bind to and stabilize AU-rich element-mRNAs [39], and thus we

speculate there are other factors involved in METTL3-mediated degradation of *ARHGDI1* mRNA, which deserves a follow-up study.

Subsequent studies revealed that USP4 was upregulated in PCa cells upon METTL3 knockdown. Some researchers found that USP4 was markedly increased and verified to play a tumor-promoting role in liver cancer [40, 41] and colorectal cancer [42, 43]. By contrast, USP4 was reported to play a tumor-suppressing role in breast cancer [44] and lung cancer [45]. Interestingly, USP4 may exert completely opposite effects even in the same tumor due to its different upstream or downstream effectors [46, 47]. Based on these results, we speculated that the specific roles of USP4 in modulating cancer progression are complicated and diverse, which depend on tumor types and tumor microenvironment. In this study, we provided compelling evidence to identify that METTL3 modulates PCa metastasis. Moreover, we also found that the knockdown of USP4 effectively alleviated METTL3 deficiency-induced suppressed xenograft tumor formation. These results suggested that USP4, as an downstream target of METTL3, might also regulate PCa growth, which deserves a follow-up study.

By performing m⁶A RIP-qRT-PCR assay, we found that USP4 was the key downstream target of METTL3 in PCa progression. Subsequent studies demonstrated that m⁶A could decrease USP4 expression by inducing m⁶A-dependent decay of the *USP4* transcript. Moreover, we identified that METTL3 could methylate USP4 at one m⁶A site within the CDS and around the stop codon, and the m⁶A reader protein YTHDF2 could recognize the m⁶A site and then recruit RNA-binding protein HNRNP D to the mRNA, resulting in degradation of *USP4* mRNA.

Conclusions

Collectively, we have provided compelling evidence to identify that METTL3 promotes PCa metastasis by upregulating *ARHGDI1* expression (Figure 8M). These results presented here suggested that METTL3 might be a favorable predictor for PCa, and this study also provides insight into novel therapeutic strategies by inhibiting METTL3 expression for suppressing PCa metastasis.

Abbreviations

ALKBH5: alkB homolog 5; FTO: fat mass and obesity-associated protein; m⁶A: N⁶-methyladenosine; METTL3: Methyltransferase like 3; PCa: Prostate cancer; RIP: RNA-immunoprecipitation; qRT-PCR: quantitative real-time PCR; TCGA: The Cancer Genome Atlas; WTAP: Wilms tumor 1-associating protein; YTHDF2: YTH-domain family 2.

Supplementary Material

Supplementary figures and tables.

<http://www.thno.org/v11p7640s1.pdf>

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Author Contributions

YC and CP designed the experiments. YC, CP, XW, DX, YM, JH, and PC conducted experiments. YC and CP analyzed the data. YC write the paper, and ZX helped to revise the manuscript. XH and QR supervised the study. All authors read and approved the final manuscript.

Competing Interests

The authors have declared that no competing interest exists.

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