AMERICAN SOCIETY FOR MICROBIOLOGY



The Physiological TMPRSS2 Inhibitor HAI-2 Alleviates SARS-CoV-2 Infection

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KEYWORDS SARS-CoV-2, HAI-2, TMPRSS2

he largest disease pandemic in modern human history, caused by severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2), is still ongoing. The disease severity varies greatly (from asymptomatic to death) among individuals and by age (1, 2), but the mechanisms for the variation remain unclear. SARS-CoV-2 uses host proteases to activate its spike protein (3). The type II transmembrane serine protease TMPRSS2 plays an important role for spike protein activation (3-5). A recent study demonstrated that hepatocyte growth factor activator inhibitor 2 (HAI-2) is a cognate inhibitor of TMPRSS2 (6). Albeit less efficiently, HAI-1 also inhibits TMPRSS2 (6). In this study, the potential role of HAI-2 in inhibiting SARS-CoV-2 infection was analyzed. The His-tagged full-length ectodomain of HAI-2 was expressed in HEK293S cells lacking *N*-acetylglucosaminyltransferase I (293S GnTI⁻ cells) and purified by Ni²⁺-NTA affinity column and Superdex 200 GL 10/300 gel filtration chromatography, as reported previously (7). Individual fractions were then tested for trypsin inhibition activity using a Pierce Fluorescent protease assay kit, and the fraction (fraction 18-20) that contained HAI-2 proteins with an expected size (\sim 27 kDa) and showing the highest activity was used for subsequent assays. SARS-CoV-2 may use TMPRSS2 at the plasma membrane (3–5) or lysosomal cathepsin L (8) for spike protein activation. Nafamostat and E-64d inhibit TMPRSS2- and cathepsin L-mediated coronavirus entry, respectively (9, 10). VeroE6 and VeroE6/TMPRSS2 cells (4) were infected with the SARS-CoV-2 Wk521 strain (4) in the presence or absence of inhibitors (10 μ M nafamostat, 10 μ M E-64d, or $20 \,\mu$ g/ml HAI-2), and the viral RNA levels at 6 h postinfection were quantified by realtime RT-qPCR as reported previously (4). In VeroE6/TMPRSS2 cells both TMPRSS2- and cathepsin L-mediated entry pathways are used, while in VeroE6 cells only the cathepsin L-mediated pathway is available. As expected, E-64d, but not nafamostat, blocked SARS-CoV-2 infection of VeroE6 cells, while neither nafamostat nor E-64d alone blocked SARS-CoV-2 infection of VeroE6/TMPRSS2 cells (Fig. 1A). The combined use of nafamostat and E-64d efficiently blocked SARS-CoV-2 infection of VeroE6/TMPRSS2 cells (viral RNA level reduced by \sim 100-fold). In this experimental condition, HAI-2 showed comparable inhibitory ability to nafamostat (Fig. 1A). Dose-dependent inhibition of SARS-CoV-2 infection in VeroE6/TMPRSS2 cells by HAI-2 in the presence of 10 µM E-64d was also observed (Fig. 1B). HAI-2 is endogenously expressed in many cell types (11) and may thus inherently inhibit or alleviate SARS-CoV-2 infection. Expression of HAI-2 in human lung adenocarcinoma Calu-3 cells was knocked down (KD) by small interfering RNA (siRNA) transfection (Fig. 1C), and virus infection assays were performed. In a previous study using Middle East respiratory syndrome coronavirus, it was

Citation Tomita Y, Matsuyama S, Fukuhara H, Maenaka K, Kataoka H, Hashiguchi T, Takeda M. 2021. The physiological TMPRSS2 inhibitor HAI-2 alleviates SARS-CoV-2 infection. J Virol 95: e00434-21. https://doi.org/10.1128/JVI.00434-21.

Editor Tom Gallagher, Loyola University Chicago

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Accepted manuscript posted online 31 March 2021

Published 24 May 2021



FIG 1 SARS-CoV-2 infection inhibition assay results. (A and B) VeroE6/TMPRSS2 or VeroE6 cells treated with protease inhibitors were infected with SARS-CoV-2 at an MOI of 0.1 and the viral RNA levels at 6 h postinfection were quantified by real-time RT-PCR. (A) Black and white bars indicate data with VeroE6/TMPRSS2 and VeroE6, respectively. The asterisk and double asterisk indicate *P* < 0.001 and *P* < 0.01, respectively. (B) Dose-dependent inhibition of SARS-CoV-2 infection in VeroE6TMPRSS2 cells by HAI-2 in the presence of 10 μ M E-64d. (C) Effect of HAI-2 KD by siRNA transfection in Calu-3 cells. (D) Virus infection assay results in HAI-2 KD and control cells. Calu-3 cells transfected with HAI-2-specific siRNA or control siRNA were infected with SARS-CoV-2 at an MOI of 10 and the viral RNA levels at 3 days postinfection were quantified by real-time RT-PCR.

demonstrated that TMPRSS2 is mainly utilized during virus entry into Calu-3 cells and perhaps into the lung as well (12). The level of viral RNA in HAI-2 KD cells was ~40 times greater than that in control siRNA-transfected cells (Fig. 1D). These data indicated that the endogenous level of HAI-2 in Calu-3 cells alleviated SARS-CoV-2 infection. The present study provides two key messages. First, the expression level of HAI-2 modulates the infection level of SARS-CoV-2. Because SARS-CoV-2 spreads systemically *in vivo* (3), even a small imbalance or change in TMPRSS2 and HAI-2 expression may modulate tissue tropism or disease severity caused by SARS-CoV-2. Second, studies on HAI-2 may open a way to develop protein-based therapeutics against SARS-CoV-2, as already suggested for the treatment of TMPRSS2-mediated malignant tumor invasions (6, 11).

ACKNOWLEDGMENTS

This study was supported by Grants-in-Aid from AMED (grant number 19fk0108111j [to T.H. and M.T.]) and JSPS (grant numbers 18H02665 [to M.T.] and 20H05773 [to T.H.]).

REFERENCES

- Long QX, Tang XJ, Shi QL, Li Q, Deng HJ, Yuan J, Hu JL, Xu W, Zhang Y, Lv FJ, Su K, Zhang F, Gong J, Wu B, Liu XM, Li JJ, Qiu JF, Chen J, Huang AL. 2020. Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections. Nat Med 26:1200–1204. https://doi.org/10.1038/s41591 -020-0965-6.
- Wu Z, McGoogan JM. 2020. Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China: summary of a report of 72314 cases from the Chinese Center for Disease Control and Prevention. JAMA 323:1239–1242. https://doi.org/10.1001/jama.2020.2648.
- Fuentes-Prior P. 2020. Priming of SARS-CoV-2 S protein by several membrane-bound serine proteinases could explain enhanced viral infectivity and systemic COVID-19 infection. J Biol Chem 296:100135. https://doi .org/10.1074/jbc.REV120.015980.
- Matsuyama S, Nao N, Shirato K, Kawase M, Saito S, Takayama I, Nagata N, Sekizuka T, Katoh H, Kato F, Sakata M, Tahara M, Kutsuna S, Ohmagari N, Kuroda M, Suzuki T, Kageyama T, Takeda M. 2020. Enhanced isolation of SARS-CoV-2 by TMPRSS2-expressing cells. Proc Natl Acad Sci U S A 117:7001–7003. https://doi.org/10.1073/pnas.2002589117.

- Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu NH, Nitsche A, Muller MA, Drosten C, Pohlmann S. 2020. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 181:271–280. https://doi.org/10.1016/j.cell.2020.02.052.
- Ko CJ, Hsu TW, Wu SR, Lan SW, Hsiao TF, Lin HY, Lin HH, Tu HF, Lee CF, Huang CC, Chen MM, Hsiao PW, Huang HP, Lee MS. 2020. Inhibition of TMPRSS2 by HAI-2 reduces prostate cancer cell invasion and metastasis. Oncogene 39:5950–5963. https://doi.org/10.1038/s41388-020-01413-w.
- Hashiguchi T, Ose T, Kubota M, Maita N, Kamishikiryo J, Maenaka K, Yanagi Y. 2011. Structure of the measles virus hemagglutinin bound to its cellular receptor SLAM. Nat Struct Mol Biol 18:135–141. https://doi.org/10 .1038/nsmb.1969.
- Ou X, Liu Y, Lei X, Li P, Mi D, Ren L, Guo L, Guo R, Chen T, Hu J, Xiang Z, Mu Z, Chen X, Chen J, Hu K, Jin Q, Wang J, Qian Z. 2020. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune crossreactivity with SARS-CoV. Nat Commun 11:1620. https://doi.org/10.1038/ s41467-020-15562-9.

- Matsuyama S, Shirato K, Kawase M, Terada Y, Kawachi K, Fukushi S, Kamitani W. 2018. Middle East respiratory syndrome coronavirus spike protein is not activated directly by cellular furin during viral entry into target cells. J Virol 92:e00683-18. https://doi.org/10.1128/JVI.00683-18.
- Yamamoto M, Matsuyama S, Li X, Takeda M, Kawaguchi Y, Inoue JI, Matsuda Z. 2016. Identification of nafamostat as a potent inhibitor of Middle East respiratory syndrome coronavirus S protein-mediated membrane fusion using the split-protein-based cell-cell fusion assay. Antimicrob Agents Chemother 60:6532–6539. https://doi.org/10.1128/AAC.01043-16.
- Kataoka H, Kawaguchi M, Fukushima T, Shimomura T. 2018. Hepatocyte growth factor activator inhibitors (HAI-1 and HAI-2): emerging key players in epithelial integrity and cancer. Pathol Int 68:145–158. https://doi.org/ 10.1111/pin.12647.
- Shirato K, Kawase M, Matsuyama S. 2013. Middle East respiratory syndrome coronavirus infection mediated by the transmembrane serine protease TMPRSS2. J Virol 87:12552–12561. https://doi.org/10.1128/JVI .01890-13.