MOLECULAR
METABOLISM

Metabolic drivers of non-alcoholic fatty liver disease

Kendra K. Bence*, Morris J. Birnbaum

ABSTRACT

Background: The incidence of non-alcoholic fatty liver disease (NAFLD) is rapidly increasing worldwide parallel to the global obesity epidemic. NAFLD encompasses a range of liver pathologies and most often originates from metabolically driven accumulation of fat in the liver, or non-alcoholic fatty liver (NAFL). In a subset of NAFL patients, the disease can progress to non-alcoholic steatohepatitis (NASH), which is a more severe form of liver disease characterized by hepatocyte injury, inflammation, and fibrosis. Significant progress has been made over the past decade in our understanding of NASH pathogenesis, but gaps remain in our mechanistic knowledge of the precise metabolic triggers for disease worsening.

Scope of review: The transition from NAFL to NASH likely involves a complex constellation of multiple factors intrinsic and extrinsic to the liver. This review focuses on early metabolic events in the establishment of NAFL and initial stages of NASH. We discuss the association of NAFL with obesity as well as the role of adipose tissue in disease progression and highlight early metabolic drivers implicated in the pathological transition from hepatic fat accumulation to steatohepatitis.

Major conclusions: The close association of NAFL with features of metabolic syndrome highlight plausible mechanistic roles for adipose tissue health and the release of lipotoxic lipids, hepatic de novo lipogenesis (DNL), and disruption of the intestinal barrier in not only the initial establishment of hepatic steatosis, but also in mediating disease progression. Human genetic variants linked to NASH risk to date are heavily biased toward genes involved in the regulation of lipid metabolism, providing compelling support for the hypothesis that NASH is fundamentally a metabolic disease.

© 2020 The Authors. Published by Elsevier GmbH. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. INTRODUCTION

NAFLD comprises a continuum of liver pathologies ranging from NAFL, defined by the presence of fat accumulation in ≥5% of hepatocytes by histological assessment or non-invasive imaging [1], to a more serious and progressive form of the disease, NASH. The definitive diagnosis of NASH is established by liver biopsy and histologically characterized by the presence of hepatocyte injury and inflammation and typically pericellular fibrosis [2]. Patients with NASH are at a substantially greater risk of progression to cirrhosis and hepatocellular carcinoma (HCC) than those with hepatic steatosis alone and at a greater risk of mortality, with fibrosis being the most predictive histological feature [3,4]. A scarcity of robust longitudinal data in NASH patients and a lack of stage-specific circulating biomarkers continues to create impediments for understanding who might be at risk for disease progression. Moreover, reliance on a liver biopsy as the gold standard for NASH diagnosis creates challenges as accuracy and reproducibility can be diminished by sampling variability and subjectivity of pathological grading.

The prevalence of NAFLD is escalating globally at an unprecedented rate, paralleling the increase in obesity and features of metabolic syndrome. In the US alone, the incidence of NAFLD is estimated at ∼25% of the population and the prevalence of NASH is estimated to be 3−4% [5], with a projected economic burden of more than $100 billion in NAFLD-related US health care costs [6]. Notably, NASH is associated with an increased risk of cardiovascular disease (CVD), a leading cause of mortality in patients with the disease [7,8]. Despite these staggering numbers and a renewed focus on the etiology of this liver disease, gaps remain in our collective understanding of the specific molecular and biochemical events underlying the transition from fatty liver to NASH.

The journey along the disease spectrum from NAFL to NASH is heterogenous and a complex collective integration of metabolic comorbidities, genetic variations, age, ethnicity, and diet [9]. At a high level, the disease trajectory of NAFLD can be described in three phenotypic stages: NAFL, NASH (with or without fibrosis), and ultimately cirrhosis and/or HCC. Early metabolic features driving NAFL include overnutrition, insulin resistance, and obesity. Multiple insults have been posited to contribute to worsening of the disease subsequent or in parallel to excess hepatic fat accumulation, including lipotoxicity, increased oxidative stress, inflammatory cytokine release, and intestinal barrier disruption (Figure 1). In later disease stages,

Internal Medicine Research Unit, Pfizer Worldwide Research, Development, and Medical, Cambridge, MA, USA

*Corresponding author. Pfizer, Inc., Internal Medicine Research Unit, 1 Portland Street, 4th floor, Cambridge, MA, 02139, USA. E-mail: kendrakathleen.bence@pfizer.com (K.K. Bence).

pathogenic factors promote inflammation and fibrosis in the liver via activation of resident tissue Kupffer cells, recruitment of circulating immune cells, and subsequent activation of hepatic stellate cells (HSCs) ultimately leading to fibrogenesis. In concordance, later stages of NASH are associated with significant histological evidence of inflammation and fibrosis, highlighting an important role of inflammatory cells and activation of HSCs later in the course of disease (reviewed extensively elsewhere, including in [10—13]). Understanding
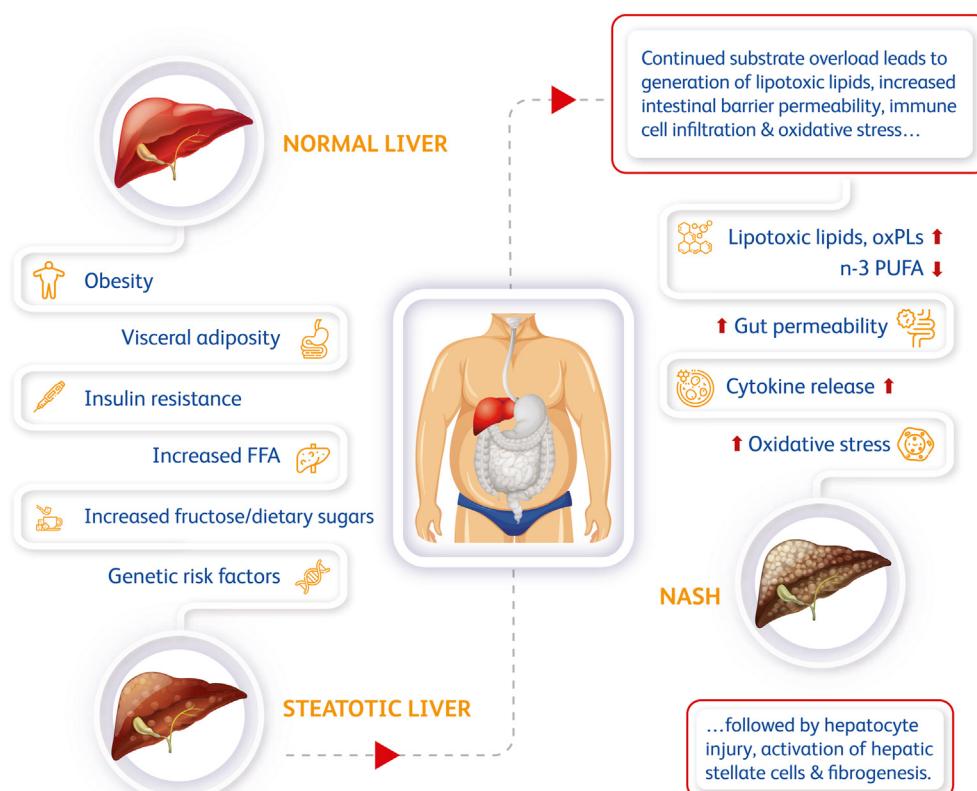
Figure 1: Risk factors associated with progression from fatty liver to NASH. Obesity, insulin resistance, and excess visceral adiposity are associated with the development of NAFL. Hepatic steatosis can be further exacerbated by overnutrition and consumption of dietary sugars and may also be influenced by genetic risk. FFA flux to the liver combined with increased DNL results in substrate overload and an inability of the liver to properly metabolize lipids. Many factors are hypothesized to play a role in the transition to NASH, including a build-up of lipotoxic or oxidized lipids, disrupted intestinal barrier function, increased immune cell activation, secretion of inflammatory cytokines, and elevated hepatic oxidative stress.

the molecular basis for progression from hepatic steatosis to NASH, a critical first step for effective diagnosis and treatment of this disease, has proven challenging given the complex and heterogeneous disease trajectory. In this review, we focus on the metabolic mechanisms that contribute to the initial deposition of fat in the liver and metabolic insults that may subsequently influence progression from NAFL to early stages of NASH.

2. HEPATIC STEATOSIS AND ADIPOSE TISSUE INSULIN RESISTANCE

Fat can accumulate in the liver via several mechanisms; the major sources in humans are flux of free fatty acids (FFAs) from adipose tissue due to increased lipolysis, increased DNL, and dietary fat intake [14,15]. Human isotope-labeling studies demonstrate that (1) DNL is significantly upregulated in NAFLD patients and (2) the major component of hepatic fat in NAFLD comes from excess circulating FFAs [16—19]. Under normal conditions, plasma FFA concentrations rise during fasting to supply substrate for tissues in the body and decline in response to feeding due to insulin-induced suppression of lipolysis. In the context of insulin resistance, however, unrestrained lipolysis leads to increased levels of circulating FFAs and deposition of fat in the liver, highlighting adipocyte insulin resistance as a major contributor to NAFLD [20]. Consistent with this hypothesis, plasma FFA levels were suppressed and liver triglyceride (TG) levels were significantly decreased in high-fat diet (HFD)-fed mice upon chronic suppression of lipolysis via activation of adipocyte Gi signaling [21]. Recent studies also pointed to an important link between a failure of insulin to suppress lipolysis in adipose and the regulation of hepatic glucose production (HGP). Using in vivo metabolomics in rats, Perry et al. showed that the ability of insulin to suppress lipolysis is critical for suppression of HGP via reducing hepatic acetyl CoA levels and pyruvate carboxylase activity, which regulates the first step of gluconeogenesis in the liver [22]. Titchenell et al. utilized mouse loss-of-function models to demonstrate that excess circulating FFAs, rather than direct hepatocyte mediators of insulin signaling, are the major factor promoting HGP in the context of obesity and insulin resistance [23]. These mechanistic studies are consistent with the fact that most significant comorbid conditions associated with NAFLD are prominent features of insulin resistance, including hypertriglyceridemia, type 2 diabetes (T2D), and obesity [24].

3. OBESITY AND NAFLD

Epidemiological studies convincingly show that obesity is a major risk factor for developing NAFLD [5,25]. In adults, overweight is defined as a body mass index (BMI) between 25.0 and 29.9 kg/m^2 and obesity is defined as a BMI $\geq$ 30 kg/m^2. Recent data showed that more than 100 million people in the US and $\sim$1.5 billion people worldwide are overweight or obese, with this trend continuing to rise [26,27]. Given the widespread global epidemic of obesity and associated metabolic dysfunction, the burden of NAFLD on the health care system continues to grow [29]. Obesity also continues to affect the health of children and adolescents [30], putting them at increased risk of developing NAFLD [31—33]. A recent meta-analysis confirmed that overweight or obese children have greater odds of developing NAFLD as adults than normal weight children [34]. The risk of having NAFLD and elevated alanine aminotransferase levels was mitigated in subjects who were obese in childhood but non-obese as adults, supporting the idea that at least some features of NAFLD may be reversible with normalization of body weight [35]. Indeed, compelling data in support of weight gain and

insulin resistance as major etiological factors in the development of NASH as well as attractive therapeutic targets lies in bariatric surgery outcomes [36,37]. Bariatric surgery leads to significant weight loss and improvements in metabolic regulation resulting in reduced hepatic steatosis and in many cases, resolution of NASH. A meta-analysis of bariatric surgery studies demonstrated improvements in hepatic steatosis in over 90% of patients and improved steatohepatitis in more than 80% of patients with NASH [38]. The efficacy of weight loss in the reversal of NASH is independent of the means of weight reduction, as intensive nutritional interventions also resulted in reduced steatosis and improvements in NASH [39—41].

Weight loss in the $\geq$10% range is recognized as the most compelling treatment modality to date for meaningful histological improvements in most NASH patients [24,41,42]. These data have spurred the evaluation of the glucagon-like peptide-1 (GLP-1) class of obesity and T2D drugs for treating NASH. In recently reported results of a 72-week, randomized, double-blind placebo-controlled phase 2 study examining the treatment of NASH patients with the GLP-1 receptor agonist semaglutide (NCT02970942), 59% of patients who received a 0.4 mg daily dose of subcutaneous semaglutide had NASH resolution without worsening of fibrosis compared to 17% of subjects on placebo [43]. While it remains to be seen whether this mechanism will lead to improvements in fibrosis in larger biopsy studies, these results are consistent with the promise of GLP-1 therapies for preventing disease worsening and potentially reversing the course of the disease. It is likely that the predominant mechanism by which GLP-1 improves NASH is via weight loss and long-term reduction in insulin resistance, although direct effects on the liver have also been proposed [44].

4. VISCERAL ADIPOSITY AND ADIPOSE TISSUE DYSFUNCTION

In the context of obesity, regional adiposity influences the risk of metabolic complications. Nielson et al. utilized elegant tracer and computational methods in humans to demonstrate that the delivery of FFAs derived from visceral fat to the portal circulation increases coordinately with the elevated visceral fat mass seen in obesity [45,46]. Expansion of visceral adipose in particular is linked to systemic inflammation [47—49]. Visceral obesity is associated with increased production of many proinflammatory cytokines, including interleukin 6, tumor necrosis factor alpha, and C-reactive protein, and a reduction in protective adipokines such as adiponectin and is thus postulated to have a more toxic profile in the context of metabolic dysfunction compared to subcutaneous fat [48,50,51]. In addition to being more metabolically active in secreting cytokines, visceral fat is more insulin resistant and lipolytic than subcutaneous fat [20,52].

Although it is widely reported that increased visceral adiposity is a risk factor for NAFLD (see Table 1 in Reference [53]), is associated with liver inflammation and fibrosis independent of hepatic steatosis [54], and is predictive of NASH with significant fibrosis [55], the specific contribution of visceral fat to the establishment of NAFL vs progression to NASH remains controversial. Fabbrini et al. suggested that intrahepatic triglyceride content, rather than visceral adipose tissue, may be a better indicator of metabolic complications associated with obesity [56]. Using waist circumference (WC) as a surrogate marker of visceral adiposity, Francanzani et al. reported that patients with normal WC can have NASH and are at a risk of progression to fibrosis, suggesting that visceral adiposity may be less important in the progression to severe liver disease [57]. It is likely that some of these discrepancies in the literature are a result of different measures used to assess visceral fat, including WC, waist-hip ratio, visceral adiposity index, and

Gene	Protein name	Principal SNP and variant	NAFLD/NASH risk	Biochemical function	Reference
PNPLA3	Patatin-like phospholipase domain-containing protein 3 or adiponutrin	rs738409; p.I148M	Increased	Lipid droplet protein with triglyceride hydrolase activity and retinyl-palmitate lipase activity; remodeling of hepatic fatty acids	[185]
TM6SF2	Transmembrane 6 superfamily member 2	rs58542926; p.E167K	Increased	Involved in hepatic VLDL secretion and lipoprotein metabolism	[186]
MBOAT7	Membrane-bound O-acyltransferase domain-containing protein 7	rs641738; p.G17E	Increased	Phospholipid remodeling; acyltransferase that catalyzes the acylation of lysophosphatidylinositol with arachidonoyl-CoA	[187]
GCKR	Glucokinase regulatory protein	rs1260326; p.P446L	Increased	Inhibitor of glucokinase activity; regulator of hepatic de novo lipogenesis	[188]
HSD17β13	Hydroxysteroid 17-beta dehydrogenase 13	rs72613567:TA splice variant	Decreased	Hepatic LD protein with retinol dehydrogenase activity; other family members linked to steroid and fatty acid metabolism	[192,193]
MARC1	Mitochondrial amidoxime-reducing component 1	rs2642438; missense p.A165T	Decreased	Molybdenum-containing enzyme capable of reducing N-hydroxylated compounds; associated with the outer mitochondrial membrane	[194]

Table 1 — Human genes associated with NAFLD and NASH identified from exome- and/or genome-wide association studies [181—184,206] and links to metabolism.

imaging, some of which cannot distinguish between visceral and subcutaneous fat compartments [58,59]. Additionally, it may be the "health" of the visceral adipose compartment that is relevant to NAFLD pathogenesis rather than the fat mass per se.

5. LIPOTOXICITY: ARE ALL LIPIDS CREATED EQUAL?

Adipose tissue acts as the body's main reservoir for lipid storage. When this storage capacity is overloaded in the context of obesity and overnutrition, ectopic fat accumulates in the liver and organs that usually only store relatively small amounts of fat, including the skeletal muscle and heart, inducing lipotoxicity. Lipotoxicity is characterized by the accumulation of harmful lipid species, leading to chronic inflammation and progression from hepatic steatosis to NASH [60]. TG is the predominant lipid that accumulates in the liver in NAFL. However, studies suggest that increased TG, due to either increased de novo synthesis [61] or reduced export [62], is not necessarily pathogenic in isolation and may even be protective against liver injury [63]; rather, it is likely that the delivery of excess FFA to the liver or generation of other lipotoxic species within the liver leads to lipotoxicity and subsequent hepatocyte dysfunction [64,65]. Remarkably, under some experimental conditions, DNL can actually be preventative against lipotoxicity [66]. Fatty acids taken up by hepatocytes serve as substrates for lipid synthesis or mitochondrial beta-oxidation [15]. Some lipid classes may be protective in the context of hepatic injury, including n-3 polyunsaturated fatty acids (PUFAs). Levels of n-3 PUFAs are reportedly decreased in NASH patients [67] and an elevated n-6:n-3 PUFA ratio has been associated with the severity of NAFLD [68]. Consistent with the role of n-3 PUFAs in reducing hepatic lipid, meta-analyses demonstrated consistent reductions in steatosis with n-3 supplementation but no effect on NASH activity scores or fibrosis, suggesting the beneficial effects of n-3 PUFAs may be limited to reductions in steatosis or possibly that more long-term benefits exist but were not captured in the endpoints of these studies [69,70].

The specific lipid species responsible for promoting hepatocyte injury and NASH pathogenesis is of considerable debate [71]. FFAs have most commonly been implicated as possible drivers of hepatic lipotoxicity. Some studies highlighted a more severe pathogenic role for saturated fatty acids (FAs), such as palmitic acid, compared to unsaturated FAs; however, most of these studies were performed in vitro and/or in rodent NASH models, and there is less evidence pointing to a definitive role of saturated FAs in driving NASH progression in NAFLD patients ([67]; reviewed in [60,71,72]). The lack of an animal model that faithfully mimics the histological progression of NASH in humans has severely hampered such mechanistic studies. In the context of metabolically driven NAFLD, the lipid composition of the liver is predominated by saturated and mono-unsaturated TGs and ceramides [73,74]. In contrast, however, in carriers of the patatin-like phospholipase domain-containing protein 3 (PNPLA3)-148M NASH risk allele, hepatic TGs are enriched in PUFAs compared to PNPLA3-148I carriers [75,76], demonstrating that there is much more to learn about liver TG remodeling, the metabolism of PUFAs in the liver, and implications int the risk of progression to NASH. A lipidomic analysis of human liver biopsies comparing normal, NAFLD, and NASH livers identified a lipid signature that could discriminate NASH livers: NASH livers were found to have a lipidomic profile interpreted as indicative of decreased of fatty acid desaturase 1 and elongation of very long-chain fatty acid 6 activity, leading to an increase in long-chain fatty acids (LCFA) [77], oxidation of which can lead to elevated reactive oxygen species (ROS) in the liver [78]. Other candidate lipotoxic species implicated in NASH progression include ceramides, diacylglycerols, cholesterol, and lysophosphatidylcholine (reviewed in detail elsewhere [65,71,74,79—84]); however, whether these lipid intermediates actually contribute to severity of NASH in humans remains uncertain. Adding to the complexity is the uncertainty of how well the circulating lipidome reflects the hepatic milieu.

6. HEPATIC LIPID METABOLISM AND NAFLD

The endogenous synthesis of lipids from dietary substrates, or DNL, is driven by increases in consumption of a high-carbohydrate diet. Elegant tracer studies have shown that hepatic DNL is increased in individuals with NAFLD and is a significant contributor to elevation in liver fat [17—19]. Collectively these studies were very well aligned and

showed that the contribution of DNL to liver TG synthesis in the fed state is in the range of ∼25—38% in obese NAFLD subjects. Increased DNL in the context of NAFLD may be at least partially explained by dysregulated transcriptional regulation of lipogenesis in the liver. Two key transcription factors regulate lipogenesis in the liver: sterol regulatory element-binding protein 1c (SREBP1c), which is activated by insulin and the liver X receptor, and carbohydrate regulatory element-binding protein (ChREBP), which is activated by products of carbohydrate metabolism [85—88]. SREBP1c activity and, as a consequence, DNL is increased in the liver in the context of NAFLD due to signaling driven by hyperinsulinemia, despite sustained HGP, a metabolic state that has been called selective insulin resistance [89—91]. SREBP1c also potently induces the transcription of PNPLA3, further exacerbating hepatic steatosis [92]. Similarly, hyperglycemia provides a stimulus for increased transcriptional activity of ChREBP, which, in cooperation with SREBP1c, leads to increased expression of lipogenic genes that include acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS). ACC catalyzes the first committed step in fatty acid synthesis, the production of malonyl-CoA, which is an allosteric inhibitor of carnitine palmitoyltransferase and reduces fatty acid oxidation [93], further exacerbating hepatic steatosis. As expected, inhibition of ACC results in potent reduction in DNL and increased fatty acid oxidation, reversing steatosis and resolving multiple features of NASH in rodents and humans. ACC inhibition also results in an adaptive increase in SREBP1 activity, however, and a subsequent elevation in circulating TGs [94—101].

Although increased DNL clearly contributes to hepatic steatosis, whether it specifically drives progression to NASH is unclear. DNL is associated with elevated levels of saturated fatty acids and ceramides, which can lead to increased hepatocyte injury and inflammation [8,102]. In parallel, defects in hepatic mitochondrial fatty acid beta-oxidation have been suggested to contribute to the development of liver steatosis and NASH progression. The data are mixed, however, on whether fatty acid oxidation is altered in NAFLD and NASH patients. Recent measurements of hepatic fluxes using stable isotope tracers in individuals with NAFLD revealed increases in acetyl-CoA oxidation in the tricarboxylic acid cycle despite reductions in ketogenesis [103]. These data indicate that the fate of oxidized fatty acids is important to driving increased glucose production during insulin-resistant states. Further, partitioning of LCFA to peroxisomes for oxidation increases levels of ROS and toxic lipid species. As outlined herein, many lipid species are altered in the context of NAFLD, thus it is likely that the overall composition of the hepatic fatty acid "pool" ultimately determines the tipping point from steatosis to more severe liver injury.

7. LIPID METABOLISM AND OXIDATIVE STRESS

In addition to hepatic steatosis and aberrant lipid metabolism, oxidative stress is a prominent feature associated with progression to NASH. The generation of ROS is a major source of hepatocyte and liver injury and is closely linked to the accumulation of hepatic FFA [104,105]. Excess delivery of FFA can upregulate beta-oxidation and lead to elevation in mitochondrial ROS production, particularly in the context of an environment with impaired anti-oxidant function as occurs in NAFLD [106]. ROS act as normal signaling mediators within cells as well as damaging oxidizing agents for DNA, proteins, and lipids. The intersection of lipid metabolism and oxidative stress has led to a focus on a potential role for oxidized lipids in promoting liver inflammation and worsening of NASH. Oxidized LDL (oxLDL), for example, is a well-known atherogenic factor that has also been linked to inflammation in rodent NASH [107—109]. The uptake of oxLDL by macrophages is

pro-inflammatory and promotes the formation of foam cells in the context of atherosclerosis. Similarly, it has been hypothesized that hepatic inflammation in NASH is triggered by the recognition of oxLDL by Kupffer cells. In support, HFD-fed mice administered oxLDL rapidly develop significant hepatic inflammation and cellular injury [110], and strategies to neutralize oxLDL in a rodent model of NASH blunts immune cell infiltration and hepatic inflammation [111]. Oxidized phospholipids (oxPLs) form by lipid peroxidation or chemical modification of phospholipids. OxPLs accumulate in oxLDL and lipoprotein(a), plasma lipoproteins that are linked to cardiovascular disease, and are released by injured or apoptotic cells [112—114]. Recent work by Witztum et al. highlighted the potential relevance of oxPLs in promoting oxidative stress and inflammation in the context of atherosclerosis and NASH. Mice engineered to produce an antibody that binds to oxPL and prevents oxPL-containing LDL particles from being taken up by macrophages displayed reduced systemic inflammation and less atherosclerosis [115]. Similarly, neutralization of oxPL in $Ldlr^{-/-}$ mice fed a NASH-promoting diet enriched in trans-fat, fructose, and cholesterol significantly reduces liver TG, markers of inflammation and fibrosis, and restores mitochondrial function in the liver, although it is important to note that body weight is also reduced in these mice [116]. Given the links between disordered lipid metabolism, NASH, and CVD [8], it is intriguing to consider elevation of oxidized lipids and lipoproteins as a possible causative trigger in both NAFL progression to NASH and CVD in some people. More studies are needed to demonstrate whether neutralization of these inflammatory oxidized lipid species can reverse and/or prevent progression of the disease. Further, although some studies have reported evidence to support altered oxidized lipids or lipoproteins in NAFLD or NASH patients [116—120], circulating and liver levels of oxidized lipids and lipoproteins have not been fully evaluated in large cohorts of NASH patients by disease stage, making even correlative assessments difficult.

8. FRUCTOSE AND NAFLD

Overconsumption of sugar-sweetened beverages containing sucrose (which is broken down into fructose and glucose) or high-fructose corn syrup (a mixture of fructose and glucose) is linked to increased visceral adiposity, hepatic steatosis, and NASH [121—128]. Excess intake of calories in the form of dietary sugars, particularly simple sugars, potently promotes fatty acid synthesis in humans [129,130]. The ability of moderate amounts of fructose ingestion to promote metabolic dysfunction in people is a point of controversy. Some studies either found no associations with fructose and metabolic dysfunction in cross-sectional studies or attributed any deleterious effects of fructose simply to excess caloric intake [131—133]. In contrast, fructose has been shown to be particularly detrimental in those with even acute fructose consumption, leading to rapid elevations in DNL, very low-density lipoprotein (VLDL) TG secretion, and hepatic steatosis [122,123,134—136].

Unique features associated with fructose metabolism may help explain why it appears to be particularly metabolically harmful compared to glucose [126,137—139]. While glucose metabolism in the liver is tightly regulated, fructose is extracted from the portal circulation and delivered to the liver where it undergoes rapid and uninhibited phosphorylation by ketohexokinase (KHK) to form fructose-1-phosphate, generating substrates for fatty acid synthesis [140—142]. In response to a high fructose load, the metabolism of fructose can deplete ATP in the liver, increasing uric acid production and hepatocyte stress [143]. However, the major mechanism by which fructose promotes DNL is by increasing the intracellular concentration of fructose-1-phosphate, a

powerful allosteric activator of glucokinase. In cooperation with increased concentrations of circulating glucose, enhanced glycolytic flux activates ChREBP, which as previously described promotes the transcription of genes that encode lipogenic enzymes [126,138,144]. To better understand the metabolic fate of fructose, Jang et al. utilized isotope tracers and mass spectrometry to demonstrate that when physiological levels of glucose and fructose are delivered in a 1:1 ratio in mice, ~90% of fructose is metabolized in the small intestine [145]. Administration of higher doses of fructose results in saturation of small intestine fructose clearance and a significant amount of fructose reaching the liver. These studies provide the framework that fructose metabolism by the small intestine serves to protect the liver from the deleterious effects of fructose. Consistently, deletion of *Khk-C* in the intestine in mice results in increased delivery of fructose to the liver and enhanced hepatic steatosis [146]. In mice fed a HFD, consumption of fructose-sweetened water was associated with more pronounced weight gain, increased expression of hepatic *Srebp1c* and *Chrebp-β*, insulin resistance, and augmented fatty acid synthesis compared to mice fed a HFD and given glucose-sweetened water [137]. Suppression of KHK in the liver improved hepatic steatosis and glucose tolerance in the fructose-supplemented mice. Consistent with this study, mice with genetic deletion of *Khk-A/C* were protected from fructose-induced metabolic dysfunction [147] and mice with liver-selective *Khk-A/C*-deficiency were protected from fructose-mediated hepatic steatosis and insulin resistance [141]. The relevance of these preclinical studies to human is an active area of investigation (reviewed in [148]); however, in totality the evidence supports the concept that strategies aimed at limiting fructose metabolism (such as KHK inhibition [149,150]) or reducing fructose in the diet ([121,151,152]) may represent attractive therapeutic approaches for NAFLD and NASH. Indeed, a recent randomized, double-blind placebo-controlled, Phase 2a study (NCT03256526) assessed the effects of the reversible KHK inhibitor PF-06835919 on metabolic parameters in participants with NAFLD. Participants receiving a once-daily oral 300 mg dose of PF-06835919 for 6 weeks displayed a significant reduction in whole liver fat by magnetic resonance imaging-proton density fat fraction (difference of −18.73%; p = 0.04 compared to placebo; no differences were observed at a lower 75 mg dose) [207].

9. GUT MICROBIOME AND BARRIER FUNCTION IN NAFLD

There is a growing body of literature on the "gut—liver axis" highlighting a possible role for the contribution of the gut microbiome to the progression of NAFLD. NAFLD has been linked to intestinal bacterial overgrowth and dysbiosis [153—155], consistent with higher endotoxin levels in the serum of NAFLD patients compared to controls [156]. Numerous studies have reported altered composition of the bacterial microbiome in NAFLD subjects (reviewed in [157]). Collectively, however, cross-sectional studies have not convincingly identified a consistent microbial signature associated with NAFLD in humans. The reported data are often correlative and inconsistent (reviewed in [158,159]), which is not surprising given that the gut microbiome is dynamic, responsive to diet and environment, and influenced by age, sex, bile acids, use of medications, and other factors, making it particularly challenging to study [154]. Further, it is often difficult to determine whether associations between altered gut microbiome and NAFLD are primarily obesity-driven rather than specific to NAFLD. It is currently unclear whether a particular gut microbiome composition can worsen NASH in humans; however, in mice it was reported that co-housing of NASH-susceptible inflammasome-deficient mice with wild-type mice could exacerbate

features of NASH in wild-type cage mates on a methionine-choline deficient diet, suggesting the possibility of an autonomous contribution of the gut microbiome to the induction of metabolic dysfunction and liver disease [160].

While challenges remain for studying the precise microbiota composition in NAFLD, more compelling support is emerging for the concept of increased gut permeability influencing progression of liver disease. The gut barrier, consisting of a mucus layer, intestinal epithelial cells (IECs), and the underlying lamina propria, separates the contents of the gut lumen from the circulation. IECs play an active role in responding to gut luminal signals by selective uptake of nutrients and responding to numerous metabolic signals, but they also play an important role in regulating the passive permeability of luminal contents through the expression of tight junction proteins (TJPs) between adjacent cells [161]. Dysbiosis-induced impairment of TJPs leads to increased gut permeability and subsequent transport of bacterial endotoxins to the portal circulation, exposing the liver to mediators of hepatic inflammation [162]. In a meta-analysis, NAFLD and NASH patients had increased rates of intestinal permeability compared to healthy subjects [163].

Macronutrients in obesogenic diets can also induce gut barrier dysfunction. Fructose ingestion promotes bacterial overgrowth in the intestine and induces disruption of the intestinal barrier in rodents, non-human primates, and people, leading to elevations in gut-derived endotoxins entering the portal circulation [164—167], providing a possible mechanistic link between fructose and worsening of liver disease. In a recent report, Todoric et al. described a distinct pathway whereby fructose-induced endotoxemia and gut barrier disruption leads to activated Toll-like receptor signaling in liver macrophages and downstream induction of ACC1, FAS, and hepatic steatosis [168]. This study suggested that inflammation associated with fructose-induced intestinal barrier disruption contributes to fructose-induced DNL in the liver and provides a rationale to improve intestinal barrier function as a part of NASH therapy. Supporting this concept, daily treatment of NAFLD subjects for 12 weeks with lubiprostone, a bicyclic fatty acid compound derivative previously shown to maintain tight junction barrier function, resulted in a modest reduction in liver enzyme levels and liver fat compared to placebo [169,170]. In summary, more studies are needed to understand the timing of altered gut permeability in the course of disease progression from steatosis to NASH to understand if it is causative or correlative and the specific mechanistic pathways by which improving inflammation in the gut and liver leads to a reduction in hepatic steatosis.

10. LEAN NAFLD

While most patients with fatty liver are obese, it is important to note that NAFLD can be present in lean individuals, leading to the terms "lean NAFLD" or "non-obese NAFLD" [171,172]. Lean NAFLD was originally described in Asian countries where obesity rates and average BMI are lower, but has since been documented in other populations [5,173,174]. However, population studies such as these are subject to misclassification artifacts introduced by ethnic variations in body fat distribution and definitions of "obesity" based on the Western, Caucasian body habitus [175]. Some studies suggest that lean NAFLD patients are at risk for more severe and progressive liver disease and metabolic abnormalities compared to patients with NAFLD and higher BMI [176—178] and reviewed in [171,172], whereas another study recently showed lower rates of cirrhosis and cardiovascular disease in a lean NAFLD cohort compared to non-lean study participants [179]. Although there are many non-metabolic

factors that may contribute to NAFLD in the absence of having a BMI that is classified as overweight/obese, including lipodystrophy, certain genetic mutations, hepatitis C infection, inflammatory disease of the gut, and drug-induced liver toxicity, the most common comorbidities with lean NAFLD are insulin resistance and increased visceral adiposity [172,178]. Indeed, many patients identified with lean NAFLD have often increased waist circumferences and one or more features of metabolic syndrome, raising the possibility that this subset of NAFLD patients has metabolically driven ectopic fat deposition despite normal BMI [180]. Thus, it is likely that adipose tissue dysfunction, particularly in the visceral adipose compartment, contributes to disease pathogenesis in this context.

Despite the classical paradigm described in this review, it is important to consider that there is not always evidence of elevated liver fat preceding NASH. Indeed, in a smaller subset of individuals, NASH can develop independently of hepatic steatosis [171,174]. The origins of NASH in these subjects is not well-characterized, likely to vary, and may include underreported alcohol or medication use, infections, genetic mutations, and altered bile acid profiles and gut microbiome [171,174,180]. Although the histological disease end points may be similar in NASH patients without a prior history of NAFL, it is likely that the disease etiology and trajectory are mechanistically distinct. More longitudinal studies are needed on this subset of NASH patients to understand the risk factors underlying the phenotype of NASH independent of NAFL.

11. METABOLIC LINKS TO NASH: HUMAN GENETIC EVIDENCE

Perhaps the most compelling clue to the molecular underpinnings of NAFLD pathogenesis comes from human genetics. The genes identified to date from exome- and genome-wide association studies with significant odds ratios for susceptibility to or protection from NAFLD and increased (or decreased) risk of progression to NASH are largely genes encoding proteins with known or suspected roles in hepatic and/or extra-hepatic lipid metabolism. Many of these genes encode proteins with an interesting localization pattern to lipid droplets (Table 1) [9,181—184]. For example, mutations in the lipid metabolism genes PNPLA3, transmembrane 6 superfamily member 2 (TM6SF2), membrane-bound O-acyltransferase domain-containing protein 7 (MBOAT7), and glucokinase regulatory protein (GCKR) confer increased risk of developing NAFLD and NASH [185—188]. Possessing more than one risk allele may increase the severity of liver disease, as has been shown for PNPLA3 and TM6SF2 [189], leading to efforts aimed at defining a "NASH genetic risk" score [190,191]. Mutations in the hydroxysteroid 17-beta dehydrogenase 13 (HSD17b13) gene, which encodes a hepatic lipid droplet-associated protein possibly involved in steroid or fatty acid metabolism, are associated with a decreased risk of progression to NASH [192,193]. Similarly, a common missense variant in the mitochondrial amidoxime-reducing component 1 (MARC1) gene has recently been shown to confer protection from fatty liver and all-cause cirrhosis [194]. MARC1 is a molybdenum-containing enzyme capable of reducing N-hydroxylated compounds and is associated with the outer mitochondrial membrane [195]. The physiological role of MARC1 in liver disease is unclear. Carriers of the MARC1 variant (n = 53) were recently reported to have higher concentrations of hepatic polyunsaturated phosphatidylcholines than non-carriers (n = 65); however, the significance of this finding in relation to protection from NASH remains uncertain [196]. The functions of these proteins in the liver and other tissues are being actively examined, but overall we still have more to learn about how they influence NAFLD pathogenesis.

12. CONCLUSIONS

Many factors associated with dysregulated systemic metabolism have been implicated in some way with the progression from hepatic steatosis to NASH, including altered lipid metabolism, mitochondrial dysfunction, alterations in gut microbiome, oxidative stress, inflammatory cytokines, immune response, and others; however, it remains unclear which combination of these mechanisms are the key drivers of the disease. Similar to other metabolic diseases, the progression to more serious pathologic states is likely driven by several risk factors previously listed and manifests slightly differently in each individual depending on genetic risk, presence of other metabolic comorbidities (obesity and/or diabetes), lifestyle, diet, and environment.

In agreement with the importance of metabolic drivers of NAFLD pathogenesis outlined in this review, recent comprehensive surveys of NASH therapies currently in development and showing preliminary signs of efficacy reveal a bias toward targeting pathways involved in hepatic or systemic metabolism, with relatively fewer drugs directly targeting inflammation or fibrosis alone [2,197]. Additionally, many NASH monotherapies aimed at targeting inflammatory pathways, cell death pathways, or fibrosis have failed to show robust efficacy in clinical trials, including the apoptosis signal-regulating kinase 1 inhibitor selonsertib [198], lysyl-oxidase-like-2 antibody simtuzumab [199], phosphodiesterase-4 inhibitor ASP9831 [200], pan-caspase inhibitor emricasan [201], galectin-3 inhibitor belapectin [202], and dual chemokine receptor type 2/5 antagonist cenicriviroc [203].

The ability to identify a specific "trigger" that accelerates the transition from simple hepatic steatosis to NASH is understandably desirable, since this would allow the identification of individuals at risk and would enable improved monitoring for therapeutic interventions. However, the reality is that the progression from hepatic steatosis to inflammation and fibrosis is heterogeneous across the patient population, can range from relatively quick progression to decades, and as previously mentioned is likely influenced by multiple factors. Much past attention focused on a possible "two-hit" theory that suggests that insulin resistance and hepatic steatosis alone are insufficient to promote NASH, and a second insult such as oxidative stress is required to trigger disease progression [204]. As outlined herein and elsewhere, it is much more likely that the disease trajectory is continuous, defined by an accumulation of both metabolic and inflammatory insults and modified by genetic risk and diet [2,205].

A consistent theme emerging from the considerable body of research is the probable relevance of the substrate overload model of NAFLD, whereby continuous exposure of the liver to excess glucose, fructose, and fatty acids results in partitioning of metabolites and fatty acids into pathways that promote lipogenesis and liver injury. Further, as we learn more about the importance of interorgan communication in driving NASH pathogenesis, it is clear that this is a disease of altered systemic metabolism rather than a liver disease in isolation. In obesity-driven NAFLD, the health of the adipose tissue appears to play an important role in determining the hepatic lipid environment, while in lean NAFLD, the gut microbiome and barrier integrity may play a prominent role. Indeed, examination of intestinal permeability in lean NAFLD provides support for the notion that altered gut microbiome and intestinal permeability may play a pathogenic role in NAFLD independent of obesity; a recent study of lean NAFLD subjects reported an altered gut microbiota profile in addition to higher serum bile acid levels compared to non-lean NAFLD subjects [171]. Learning more about the role of

Review

adipose-to-liver communication and the gut—liver axis should shed light on the intersection of these organs in liver disease [10]. Additionally, as we learn more about the function of the metabolic proteins encoded by NASH risk and protective alleles, we should gain critical insight into additional cellular mechanisms and pathways influencing progression to NASH.

AUTHOR CONTRIBUTIONS

KKB and MJB contributed to the review article's overall concept, reviewed the literature, and drafted, edited, and approved the final manuscript.

ACKNOWLEDGMENTS

CONFLICT OF INTEREST

The authors are employees and shareholders of Pfizer, Inc.

REFERENCES

[1] Reeder, S.B., Sirlin, C.B., 2010. Quantification of liver fat with magnetic resonance imaging. Magnetic Resonance Imaging Clinics of North America 18(3):337—357 [ix].

[2] Friedman, S.L., Neuschwander-Tetri, B.A., Rinella, M., Sanyal, A.J., 2018. Mechanisms of NAFLD development and therapeutic strategies. Nature Medicine 24(7):908—922.

[3] Anstee, Q.M., Targher, G., Day, C.P., 2013. Progression of NAFLD to diabetes mellitus, cardiovascular disease or cirrhosis. Nature Reviews Gastroenterology & Hepatology 10(6):330—344.

[4] Simon, T.G., Roelstraete, B., Khalili, H., Hagstrom, H., Ludvigsson, J.F., 2020. Mortality in biopsy-confirmed nonalcoholic fatty liver disease: results from a nationwide cohort. Gut. https://doi.org/10.1136/gutjnl-2020-322786. Epub ahead of print.

[5] Younossi, Z., Anstee, Q.M., Marietti, M., Hardy, T., Henry, L., Eslam, M., et al., 2018. Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention. Nature Reviews Gastroenterology & Hepatology 15(1): 11—20.

[6] Younossi, Z.M., Blissett, D., Blissett, R., Henry, L., Stepanova, M., Younossi, Y., et al., 2016. The economic and clinical burden of nonalcoholic fatty liver disease in the United States and Europe. Hepatology 64(5):1577—1586.

[7] Mantovani, A., Scorletti, E., Mosca, A., Alisi, A., Byrne, C.D., Targher, G., 2020. Complications, morbidity and mortality of nonalcoholic fatty liver disease. Metabolism 111S:154170.

[8] Deprince, A., Haas, J.T., Staels, B., 2020. Dysregulated lipid metabolism links NAFLD to cardiovascular disease. Molecular Metabolism 42:101092. Epub ahead of print.

[9] Eslam, M., George, J., 2020. Genetic contributions to NAFLD: leveraging shared genetics to uncover systems biology. Nature Reviews Gastroenterology & Hepatology 17(1):40—52.

[10] Hart, K.M., Fabre, T., Sciurba, J.C., Gieseck 3rd, R.L., Borthwick, L.A., Vannella, K.M., et al., 2017. Type 2 immunity is protective in metabolic disease but exacerbates NAFLD collaboratively with TGF-beta. Science Translational Medicine 9(396).

[11] Heymann, F., Tacke, F., 2016. Immunology in the liver—from homeostasis to disease. Nature Reviews Gastroenterology & Hepatology 13(2): 88—110.

[12] Kazankov, K., Jorgensen, S.M.D., Thomsen, K.L., Moller, H.J., Vilstrup, H., George, J., et al., 2019. The role of macrophages in nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. Nature Reviews Gastroenterology & Hepatology 16(3):145—159.

[13] Tsuchida, T., Friedman, S.L., 2017. Mechanisms of hepatic stellate cell activation. Nature Reviews Gastroenterology & Hepatology 14(7):397—411.

[14] Canbay, A., Bechmann, L., Gerken, G., 2007. Lipid metabolism in the liver. Zeitschrift für Gastroenterologie 45(1):35—41.

[15] Alves-Bezerra, M., Cohen, D.E., 2017. Triglyceride metabolism in the liver. Comparative Physiology 8(1):1—8.

[16] Jacome-Sosa, M.M., Parks, E.J., 2014. Fatty acid sources and their fluxes as they contribute to plasma triglyceride concentrations and fatty liver in humans. Current Opinion in Lipidology 25(3):213—220.

[17] Lambert, J.E., Ramos-Roman, M.A., Browning, J.D., Parks, E.J., 2014. Increased de novo lipogenesis is a distinct characteristic of individuals with nonalcoholic fatty liver disease. Gastroenterology 146(3):726—735.

[18] Donnelly, K.L., Smith, C.I., Schwarzenberg, S.J., Jessurun, J., Boldt, M.D., Parks, E.J., 2005. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. Journal of Clinical Investigation 115(5):1343—1351.

[19] Smith, G.I., Shankaran, M., Yoshino, M., Schweitzer, G.G., Chondronikola, M., Beals, J.W., et al., 2020. Insulin resistance drives hepatic de novo lipogenesis in nonalcoholic fatty liver disease. Journal of Clinical Investigation 130(3): 1453—1460.

[20] Guilherme, A., Virbasius, J.V., Puri, V., Czech, M.P., 2008. Adipocyte dysfunctions linking obesity to insulin resistance and type 2 diabetes. Nature Reviews Molecular Cell Biology 9(5):367—377.

[21] Wang, L., Pydi, S.P., Zhu, L., Barella, L.F., Cui, Y., Gavrilova, O., et al., 2020. Adipocyte Gi signaling is essential for maintaining whole-body glucose homeostasis and insulin sensitivity. Nature Communications 11(1):2995.

[22] Perry, R.J., Camporez, J.G., Kursawe, R., Titchenell, P.M., Zhang, D., Perry, C.J., et al., 2015. Hepatic acetyl CoA links adipose tissue inflammation to hepatic insulin resistance and type 2 diabetes. Cell 160(4):745—758.

[23] Titchenell, P.M., Chu, Q., Monks, B.R., Birnbaum, M.J., 2015. Hepatic insulin signalling is dispensable for suppression of glucose output by insulin in vivo. Nature Communications 6:7078.

[24] Sheka, A.C., Adeyi, O., Thompson, J., Hameed, B., Crawford, P.A., Ikramuddin, S., 2020. Nonalcoholic steatohepatitis: a review. Journal of the American Medical Association 323(12):1175—1183.

[25] Loomis, A.K., Kabadi, S., Preiss, D., Hyde, C., Bonato, V., St Louis, M., et al., 2016. Body mass index and risk of nonalcoholic fatty liver disease: two electronic health record prospective studies. Journal of Clinical Endocrinology & Metabolism 101(3):945—952.

[26] Flegal, K.M., Kruszon-Moran, D., Carroll, M.D., Fryar, C.D., Ogden, C.L., 2016. Trends in obesity among adults in the United States, 2005 to 2014. Journal of the American Medical Association 315(21):2284—2291.

[27] Bray, G.A., Heisel, W.E., Afshin, A., Jensen, M.D., Dietz, W.H., Long, M., et al., 2018. The science of obesity management: an endocrine society scientific statement. Endocrine Reviews 39(2):79—132.

[29] Younossi, Z.M., Koenig, A.B., Abdelatif, D., Fazel, Y., Henry, L., Wymer, M., 2016. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology 64(1): 73—84.

[30] Ogden, C.L., Carroll, M.D., Lawman, H.G., Fryar, C.D., Kruszon-Moran, D., Kit, B.K., et al., 2016. Trends in obesity prevalence among children and adolescents in the United States, 1988-1994 through 2013-2014. Journal of the American Medical Association vol. 315(21):2292—2299.

[31] Ahima, R.S., 2008. The natural history of nonalcoholic fatty liver disease: insights from children and mice. Gastroenterology 135(6):1860—1862.

[32] Mann, J.P., Valenti, L., Scorletti, E., Byrne, C.D., Nobili, V., 2018. Nonalcoholic fatty liver disease in children. Seminars in Liver Disease 38(1):1—13.

[33] Nobili, V., Alisi, A., Valenti, L., Miele, L., Feldstein, A.E., Alkhouri, N., 2019. NAFLD in children: new genes, new diagnostic modalities and new drugs. Nature Reviews Gastroenterology & Hepatology 16(9):517—530.

[34] Anderson, E.L., Howe, L.D., Jones, H.E., Higgins, J.P., Lawlor, D.A., Fraser, A., 2015. The prevalence of non-alcoholic fatty liver disease in children and adolescents: a systematic review and meta-analysis. PLoS One 10(10):e0140908.

[35] Yan, Y., Hou, D., Zhao, X., Liu, J., Cheng, H., Wang, Y., et al., 2017. Childhood adiposity and nonalcoholic fatty liver disease in adulthood. Pediatrics 139(4).

[36] Klebanoff, M.J., Corey, K.E., Chhatwal, J., Kaplan, L.M., Chung, R.T., Hur, C., 2017. Bariatric surgery for nonalcoholic steatohepatitis: a clinical and cost-effectiveness analysis. Hepatology 65(4):1156—1164.

[37] Hannah Jr., W.N., Harrison, S.A., 2016. Effect of weight loss, diet, exercise, and bariatric surgery on nonalcoholic fatty liver disease. Clinics in Liver Disease 20(2):339—350.

[38] Mummadi, R.R., Kasturi, K.S., Chennareddygari, S., Sood, G.K., 2008. Effect of bariatric surgery on nonalcoholic fatty liver disease: systematic review and meta-analysis. Clinical Gastroenterology and Hepatology 6(12): 1396—1402.

[39] Huang, M.A., Greenson, J.K., Chao, C., Anderson, L., Peterman, D., Jacobson, J., et al., 2005. One-year intense nutritional counseling results in histological improvement in patients with non-alcoholic steatohepatitis: a pilot study. American Journal of Gastroenterology 100(5):1072—1081.

[40] Hsu, C.C., Ness, E., Kowdley, K.V., 2017. Nutritional approaches to achieve weight loss in nonalcoholic fatty liver disease. Advance Nutrition 8(2):253—265.

[41] Vilar-Gomez, E., Martinez-Perez, Y., Calzadilla-Bertot, L., Torres-Gonzalez, A., Gra-Oramas, B., Gonzalez-Fabian, L., et al., 2015. Weight loss through lifestyle modification significantly reduces features of nonalcoholic steatohepatitis. Gastroenterology 149(2):367—378 e365; quiz e314-365.

[42] Hydes, T.J., Ravi, S., Loomba, R., M, E.G., 2020. Evidence-based clinical advice for nutrition and dietary weight loss strategies for the management of NAFLD and NASH. Clinical and Molecular Hepatology 26(4):383—400.

[43] Newsome, P.N., Buchholtz, K., Cusi, K., Linder, M., Okanoue, T., Ratziu, V., et al., 2020. A placebo-controlled trial of subcutaneous semaglutide in nonalcoholic steatohepatitis. New England Journal of Medicine, 13. https:// doi.org/10.1056/NEJMoa2028395. Epub ahead of print.

[44] Gupta, N.A., Mells, J., Dunham, R.M., Grakoui, A., Handy, J., Saxena, N.K., et al., 2010. Glucagon-like peptide-1 receptor is present on human hepa-tocytes and has a direct role in decreasing hepatic steatosis in vitro by modulating elements of the insulin signaling pathway. Hepatology 51(5): 1584—1592.

[45] Nielsen, S., Guo, Z., Johnson, C.M., Hensrud, D.D., Jensen, M.D., 2004. Splanchnic lipolysis in human obesity. Journal of Clinical Investigation 113(11):1582—1588.

[46] Klein, S., 2004. The case of visceral fat: argument for the defense. Journal of Clinical Investigation 113(11):1530—1532.

[47] Hotamisligil, G.S., 2006. Inflammation and metabolic disorders. Nature 444(7121):860—867.

[48] Fontana, L., Eagon, J.C., Trujillo, M.E., Scherer, P.E., Klein, S., 2007. Visceral fat adipokine secretion is associated with systemic inflammation in obese humans. Diabetes 56(4):1010—1013.

[49] Li, C., Spallanzani, R.G., Mathis, D., 2020. Visceral adipose tissue Tregs and the cells that nurture them. Immunological Reviews 295(1):114—125.

[50] Wajchenberg, B.L., Giannella-Neto, D., da Silva, M.E., Santos, R.F., 2002. Depot-specific hormonal characteristics of subcutaneous and visceral adi-pose tissue and their relation to the metabolic syndrome. Hormone and Metabolic Research 34(11—12):616—621.

[51] Mirza, M.S., 2011. Obesity, visceral fat, and NAFLD: querying the role of adipokines in the progression of nonalcoholic fatty liver disease. ISRN Gastroenterology 2011. Article ID: 592404.

[52] Arner, P., 2005. Human fat cell lipolysis: biochemistry, regulation and clinical role. Best Practice & Research Clinical Endocrinology & Metabolism 19(4): 471—482.

[53] Machado, M.V., Cortez-Pinto, H., 2011. No need for a large belly to have NASH. Journal of Hepatology 54(6):1090—1093.

[54] van der Poorten, D., Milner, K.L., Hui, J., Hodge, A., Trenell, M.I., Kench, J.G., et al., 2008. Visceral fat: a key mediator of steatohepatitis in metabolic liver disease. Hepatology 48(2):449—457.

[55] Yu, S.J., Kim, W., Kim, D., Yoon, J.H., Lee, K., Kim, J.H., et al., 2015. Visceral obesity predicts significant fibrosis in patients with nonalcoholic fatty liver disease. Medicine (Baltimore) 94(48):e2159.

[56] Fabbrini, E., Magkos, F., Mohammed, B.S., Pietka, T., Abumrad, N.A., Patterson, B.W., et al., 2009. Intrahepatic fat, not visceral fat, is linked with metabolic complications of obesity. Proceedings of the National Academy of Sciences of the U S A 106(36):15430—15435.

[57] Fracanzani, A.L., Valenti, L., Bugianesi, E., Vanni, E., Grieco, A., Miele, L., et al., 2011. Risk of nonalcoholic steatohepatitis and fibrosis in patients with nonalcoholic fatty liver disease and low visceral adiposity. Journal of Hep-atology 54(6):1244—1249.

[58] Almeida, N.S., Rocha, R., Cotrim, H.P., Daltro, C., 2018. Anthropometric indicators of visceral adiposity as predictors of non-alcoholic fatty liver dis-ease: a review. World Journal of Hepatology 10(10):695—701.

[59] Vongsuvanh, R., George, J., McLeod, D., van der Poorten, D., 2012. Visceral adiposity index is not a predictor of liver histology in patients with non-alcoholic fatty liver disease. Journal of Hepatology 57(2):392—398.

[60] Neuschwander-Tetri, B.A., 2010. Hepatic lipotoxicity and the pathogenesis of nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. Hepatology 52(2):774—788.

[61] Monetti, M., Levin, M.C., Watt, M.J., Sajan, M.P., Marmor, S., Hubbard, B.K., et al., 2007. Dissociation of hepatic steatosis and insulin resistance in mice overexpressing DGAT in the liver. Cell Metabolism 6(1):69—78.

[62] Liao, W., Hui, T.Y., Young, S.G., Davis, R.A., 2003. Blocking microsomal triglyceride transfer protein interferes with apoB secretion without causing retention or stress in the ER. The Journal of Lipid Research 44(5):978—985.

[63] Alkhouri, N., Dixon, L.J., Feldstein, A.E., 2009. Lipotoxicity in nonalcoholic fatty liver disease: not all lipids are created equal. Expert Review of Gastroenterology & Hepatology 3(4):445—451.

[64] Sharma, M., Mitnala, S., Vishnubhotla, R.K., Mukherjee, R., Reddy, D.N., Rao, P.N., 2015. The riddle of nonalcoholic fatty liver disease: progression from nonalcoholic fatty liver to nonalcoholic steatohepatitis. Journal of Clinical Experiment Hepatology 5(2):147—158.

[65] Marra, F., Svegliati-Baroni, G., 2018. Lipotoxicity and the gut-liver axis in NASH pathogenesis. Journal of Hepatology 68(2):280—295.

[66] Papazyan, R., Sun, Z., Kim, Y.H., Titchenell, P.M., Hill, D.A., Lu, W., et al., 2016. Physiological suppression of lipotoxic liver damage by complementary actions of HDAC3 and SCAP/SREBP. Cell Metabolism 24(6):863—874.

[67] Puri, P., Baillie, R.A., Wiest, M.M., Mirshahi, F., Choudhury, J., Cheung, O., et al., 2007. A lipidomic analysis of nonalcoholic fatty liver disease. Hep-atology 46(4):1081—1090.

[68] Araya, J., Rodrigo, R., Videla, L.A., Thielemann, L., Orellana, M., Pettinelli, P., et al., 2004. Increase in long-chain polyunsaturated fatty acid n - 6/n - 3 ratio in relation to hepatic steatosis in patients with non-alcoholic fatty liver dis-ease. Clinical Science 106(6):635—643.

[69] Parker, H.M., Johnson, N.A., Burdon, C.A., Cohn, J.S., O'Connor, H.T., George, J., 2012. Omega-3 supplementation and non-alcoholic fatty liver disease: a systematic review and meta-analysis. Journal of Hepatology 56(4): 944—951.

[70] Sanyal, A.J., Abdelmalek, M.F., Suzuki, A., Cummings, O.W., Chojkier, M., Group, E.-A.S., 2014. No significant effects of ethyl-eicosapentanoic acid on histologic features of nonalcoholic steatohepatitis in a phase 2 trial. Gastroenterology 147(2):377—384 e371.

[71] Farrell, G.C., Haczeyni, F., Chitturi, S., 2018. Pathogenesis of NASH: how metabolic complications of overnutrition favour lipotoxicity and pro-inflammatory fatty liver disease. Advances in Experimental Medicine & Biology 1061:19—44.

[72] Pierantonelli, I., Svegliati-Baroni, G., 2019. Nonalcoholic fatty liver disease: basic pathogenetic mechanisms in the progression from NAFLD to NASH. Transplantation 103(1):e1—e13.

[73] Luukkonen, P.K., Sadevirta, S., Zhou, Y., Kayser, B., Ali, A., Ahonen, L., et al., 2018. Saturated fat is more metabolically harmful for the human liver than unsaturated fat or simple sugars. Diabetes Care 41(8):1732—1739.

[74] Luukkonen, P.K., Zhou, Y., Sadevirta, S., Leivonen, M., Arola, J., Oresic, M., et al., 2016. Hepatic ceramides dissociate steatosis and insulin resistance in patients with non-alcoholic fatty liver disease. Journal of Hepatology 64(5):1167—1175.

[75] Luukkonen, P.K., Nick, A., Holtta-Vuori, M., Thiele, C., Isokuortti, E., Lallukka-Bruck, S., et al., 2019. Human PNPLA3-I148M variant increases hepatic retention of polyunsaturated fatty acids. JCI Insight 4(16).

[76] Peter, A., Kovarova, M., Nadalin, S., Cermak, T., Konigsrainer, A., Machicao, F., et al., 2014. PNPLA3 variant I148M is associated with altered hepatic lipid composition in humans. Diabetologia 57(10):2103—2107.

[77] Chiappini, F., Coilly, A., Kadar, H., Gual, P., Tran, A., Desterke, C., et al., 2017. Metabolism dysregulation induces a specific lipid signature of nonalcoholic steatohepatitis in patients. Scientific Reports 7:46658.

[78] Bradbury, M.W., Berk, P.D., 2004. Lipid metabolism in hepatic steatosis. Clinics in Liver Disease 8(3):639—671 [xi].

[79] Perry, R.J., Samuel, V.T., Petersen, K.F., Shulman, G.I., 2014. The role of hepatic lipids in hepatic insulin resistance and type 2 diabetes. Nature 510(7503):84—91.

[80] Mauer, A.S., Hirsova, P., Maiers, J.L., Shah, V.H., Malhi, H., 2017. Inhibition of sphingosine 1-phosphate signaling ameliorates murine nonalcoholic steatohepatitis. American Journal of Physiology - Gastrointestinal and Liver Physiology 312(3):G300—G313.

[81] Han, M.S., Park, S.Y., Shinzawa, K., Kim, S., Chung, K.W., Lee, J.H., et al., 2008. Lysophosphatidylcholine as a death effector in the lipoapoptosis of hepatocytes. The Journal of Lipid Research 49(1):84—97.

[82] Ioannou, G.N., 2016. The role of cholesterol in the pathogenesis of NASH. Trends in Endocrinology and Metabolism 27(2):84—95.

[83] Gorden, D.L., Myers, D.S., Ivanova, P.T., Fahy, E., Maurya, M.R., Gupta, S., et al., 2015. Biomarkers of NAFLD progression: a lipidomics approach to an epidemic. The Journal of Lipid Research 56(3):722—736.

[84] Poss, A.M., Summers, S.A., 2020. Too much of a good thing? An evolutionary theory to explain the role of ceramides in NAFLD. Frontiers in Endocrinology 11:505.

[85] Shimano, H., Sato, R., 2017. SREBP-regulated lipid metabolism: convergent physiology - divergent pathophysiology. Nature Reviews Endocrinology 13(12):710—730.

[86] Jeon, T.I., Osborne, T.F., 2012. SREBPs: metabolic integrators in physiology and metabolism. Trends in Endocrinology and Metabolism 23(2):65—72.

[87] Uyeda, K., Repa, J.J., 2006. Carbohydrate response element binding protein, ChREBP, a transcription factor coupling hepatic glucose utilization and lipid synthesis. Cell Metabolism 4(2):107—110.

[88] Horton, J.D., Goldstein, J.L., Brown, M.S., 2002. SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. Journal of Clinical Investigation 109(9):1125—1131.

[89] Kohjima, M., Higuchi, N., Kato, M., Kotoh, K., Yoshimoto, T., Fujino, T., et al., 2008. SREBP-1c, regulated by the insulin and AMPK signaling pathways, plays a role in nonalcoholic fatty liver disease. International Journal of Molecular Medicine 21(4):507—511.

[90] Brown, M.S., Goldstein, J.L., 2008. Selective versus total insulin resistance: a pathogenic paradox. Cell Metabolism 7(2):95—96.

[91] Titchenell, P.M., Lazar, M.A., Birnbaum, M.J., 2017. Unraveling the regulation of hepatic metabolism by insulin. Trends in Endocrinology and Metabolism 28(7):497—505.

[92] Huang, Y., He, S., Li, J.Z., Seo, Y.K., Osborne, T.F., Cohen, J.C., et al., 2010. A feed-forward loop amplifies nutritional regulation of PNPLA3. Proceedings of the National Academy of Sciences of the U S A 107(17):7892—7897.

[93] McGarry, J.D., Leatherman, G.F., Foster, D.W., 1978. Carnitine palmitoyl-transferase I. The site of inhibition of hepatic fatty acid oxidation by malonyl-CoA. Journal of Biological Chemistry 253(12):4128—4136.

[94] Griffith, D.A., Kung, D.W., Esler, W.P., Amor, P.A., Bagley, S.W., Beysen, C., et al., 2014. Decreasing the rate of metabolic ketone reduction in the discovery of a clinical acetyl-CoA carboxylase inhibitor for the treatment of diabetes. Journal of Medicinal Chemistry 57(24):10512—10526.

[95] Harwood Jr., H.J., Petras, S.F., Shelly, L.D., Zaccaro, L.M., Perry, D.A., Makowski, M.R., et al., 2003. Isozyme-nonselective N-substituted bipiperidylcarboxamide acetyl-CoA carboxylase inhibitors reduce tissue malonyl-CoA concentrations, inhibit fatty acid synthesis, and increase fatty acid oxidation in cultured cells and in experimental animals. Journal of Biological Chemistry 278(39):37099—37111.

[96] Kim, C.W., Addy, C., Kusunoki, J., Anderson, N.N., Deja, S., Fu, X., et al., 2017. Acetyl CoA carboxylase inhibition reduces hepatic steatosis but elevates plasma triglycerides in mice and humans: a bedside to bench investigation. Cell Metabolism 26(2):394—406 e396.

[97] Lawitz, E.J., Coste, A., Poordad, F., Alkhouri, N., Loo, N., McColgan, B.J., et al., 2018. Acetyl-CoA carboxylase inhibitor GS-0976 for 12 Weeks reduces hepatic de novo lipogenesis and steatosis in patients with nonalcoholic steatohepatitis. Clinical Gastroenterology and Hepatology 16(12):1983—1991 e1983.

[98] Savage, D.B., Choi, C.S., Samuel, V.T., Liu, Z.X., Zhang, D., Wang, A., et al., 2006. Reversal of diet-induced hepatic steatosis and hepatic insulin resistance by antisense oligonucleotide inhibitors of acetyl-CoA carboxylases 1 and 2. Journal of Clinical Investigation 116(3):817—824.

[99] Ross, T.T., Crowley, C., Kelly, K.L., Rinaldi, A., Beebe, D.A., Lech, M.P., et al., 2020. Acetyl-CoA carboxylase inhibition improves multiple dimensions of NASH pathogenesis in model systems. Cell Molecular Gastroenterology & Hepatology 10(4):829—851.

[100] Loomba, R., Kayali, Z., Noureddin, M., Ruane, P., Lawitz, E.J., Bennett, M., et al., 2018. GS-0976 reduces hepatic steatosis and fibrosis markers in patients with nonalcoholic fatty liver disease. Gastroenterology 155(5):1463—1473 e1466.

[101] Goedeke, L., Bates, J., Vatner, D.F., Perry, R.J., Wang, T., Ramirez, R., et al., 2018. Acetyl-CoA carboxylase inhibition reverses NAFLD and hepatic insulin resistance but promotes hypertriglyceridemia in rodents. Hepatology 68(6):2197—2211.

[102] Roumans, K.H.M., Lindeboom, L., Veeraiah, P., Remie, C.M.E., Phielix, E., Havekes, B., et al., 2020. Hepatic saturated fatty acid fraction is associated with de novo lipogenesis and hepatic insulin resistance. Nature Communications 11(1):1891.

[103] Fletcher, J.A., Deja, S., Satapati, S., Fu, X., Burgess, S.C., Browning, J.D., 2019. Impaired ketogenesis and increased acetyl-CoA oxidation promote hyperglycemia in human fatty liver. JCI Insight 5.

[104] Albano, E., Mottaran, E., Vidali, M., Reale, E., Saksena, S., Occhino, G., et al., 2005. Immune response towards lipid peroxidation products as a predictor of progression of non-alcoholic fatty liver disease to advanced fibrosis. Gut 54(7):987—993.

[105] Sumida, Y., Niki, E., Naito, Y., Yoshikawa, T., 2013. Involvement of free radicals and oxidative stress in NAFLD/NASH. Free Radical Research 47(11):869—880.

[106] Yesilova, Z., Yaman, H., Oktenli, C., Ozcan, A., Uygun, A., Cakir, E., et al., 2005. Systemic markers of lipid peroxidation and antioxidants in patients

with nonalcoholic Fatty liver disease. American Journal of Gastroenterology 100(4):850—855.

[107] Houben, T., Brandsma, E., Walenbergh, S.M.A., Hofker, M.H., Shiri-Sverdlov, R., 2017. Oxidized LDL at the crossroads of immunity in non-alcoholic steatohepatitis. Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids 1862(4):416—429.

[108] Bieghs, V., Rensen, P.C., Hofker, M.H., Shiri-Sverdlov, R., 2012. NASH and atherosclerosis are two aspects of a shared disease: central role for macrophages. Atherosclerosis 220(2):287—293.

[109] Walenbergh, S.M., Koek, G.H., Bieghs, V., Shiri-Sverdlov, R., 2013. Non-alcoholic steatohepatitis: the role of oxidized low-density lipoproteins. Journal of Hepatology 58(4):801—810.

[110] Yimin Furumaki, H., Matsuoka, S., Sakurai, T., Kohanawa, M., Zhao, S., et al., 2012. A novel murine model for non-alcoholic steatohepatitis developed by combination of a high-fat diet and oxidized low-density lipoprotein. Laboratory Investigation 92(2):265—281.

[111] Bieghs, V., van Gorp, P.J., Walenbergh, S.M., Gijbels, M.J., Verheyen, F., Buurman, W.A., et al., 2012. Specific immunization strategies against oxidized low-density lipoprotein: a novel way to reduce nonalcoholic steatohepatitis in mice. Hepatology 56(3):894—903.

[112] Chang, M.K., Binder, C.J., Miller, Y.I., Subbanagounder, G., Silverman, G.J., Berliner, J.A., et al., 2004. Apoptotic cells with oxidation-specific epitopes are immunogenic and proinflammatory. Journal of Experimental Medicine 200(11):1359—1370.

[113] Boffa, M.B., Koschinsky, M.L., 2019. Oxidized phospholipids as a unifying theory for lipoprotein(a) and cardiovascular disease. Nature Reviews Cardiology 16(5):305—318.

[114] Binder, C.J., Papac-Milicevic, N., Witztum, J.L., 2016. Innate sensing of oxidation-specific epitopes in health and disease. Nature Reviews Immunology 16(8):485—497.

[115] Que, X., Hung, M.Y., Yeang, C., Gonen, A., Prohaska, T.A., Sun, X., et al., 2018. Oxidized phospholipids are proinflammatory and proatherogenic in hypercholesterolaemic mice. Nature 558(7709):301—306.

[116] Sun, X., Seidman, J.S., Zhao, P., Troutman, T.D., Spann, N.J., Que, X., et al., 2020. Neutralization of oxidized phospholipids ameliorates non-alcoholic steatohepatitis. Cell Metabolism 31(1):189—206 e188.

[117] Ampuero, J., Ranchal, I., Gallego-Duran, R., Pareja, M.J., Del Campo, J.A., Pastor-Ramirez, H., et al., 2016. Oxidized low-density lipoprotein antibodies/high-density lipoprotein cholesterol ratio is linked to advanced non-alcoholic fatty liver disease lean patients. Journal of Gastroenterology and Hepatology 31(9):1611—1618.

[118] Ho, C.M., Ho, S.L., Jeng, Y.M., Lai, Y.S., Chen, Y.H., Lu, S.C., et al., 2019. Accumulation of free cholesterol and oxidized low-density lipoprotein is associated with portal inflammation and fibrosis in nonalcoholic fatty liver disease. Journal of Inflammation 16:7.

[119] Ikura, Y., Ohsawa, M., Suekane, T., Fukushima, H., Itabe, H., Jomura, H., et al., 2006. Localization of oxidized phosphatidylcholine in nonalcoholic fatty liver disease: impact on disease progression. Hepatology 43(3):506—514.

[120] Hendrikx, T., Watzenbock, M.L., Walenbergh, S.M., Amir, S., Gruber, S., Kozma, M.O., et al., 2016. Low levels of IgM antibodies recognizing oxidation-specific epitopes are associated with human non-alcoholic fatty liver disease. BMC Medicine 14(1):107.

[121] Stanhope, K.L., Schwarz, J.M., Keim, N.L., Griffen, S.C., Bremer, A.A., Graham, J.L., et al., 2009. Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans. Journal of Clinical Investigation 119(5):1322—1334.

[122] Schwarz, J.M., Noworolski, S.M., Erkin-Cakmak, A., Korn, N.J., Wen, M.J., Tai, V.W., et al., 2017. Effects of dietary fructose restriction on liver fat, de novo lipogenesis, and insulin kinetics in children with obesity. Gastroenterology 153(3):743—752.

[123] Softic, S., Cohen, D.E., Kahn, C.R., 2016. Role of dietary fructose and hepatic de novo lipogenesis in fatty liver disease. Digestive Diseases and Sciences 61(5):1282—1293.

[124] Ouyang, X., Cirillo, P., Sautin, Y., McCall, S., Bruchette, J.L., Diehl, A.M., et al., 2008. Fructose consumption as a risk factor for non-alcoholic fatty liver disease. Journal of Hepatology 48(6):993—999.

[125] Abid, A., Taha, O., Nseir, W., Farah, R., Grosovski, M., Assy, N., 2009. Soft drink consumption is associated with fatty liver disease independent of metabolic syndrome. Journal of Hepatology 51(5):918—924.

[126] Herman, M.A., Samuel, V.T., 2016. The sweet path to metabolic demise: fructose and lipid synthesis. Trends in Endocrinology and Metabolism 27(10):719—730.

[127] Malik, V.S., Hu, F.B., 2015. Fructose and cardiometabolic health: what the evidence from sugar-sweetened beverages tells us. Journal of the American College of Cardiology 66(14):1615—1624.

[128] Goran, M.I., 2013. Sugar-sweetened beverages, genetic risk, and obesity. New England Journal of Medicine 368(3):285—286.

[129] Parks, E.J., Skokan, L.E., Timlin, M.T., Dingfelder, C.S., 2008. Dietary sugars stimulate fatty acid synthesis in adults. Journal of Nutrition 138(6):1039—1046.

[130] Hudgins, L.C., Seidman, C.E., Diakun, J., Hirsch, J., 1998. Human fatty acid synthesis is reduced after the substitution of dietary starch for sugar. American Journal of Clinical Nutrition 67(4):631—639.

[131] Tappy, L., Le, K.A., Tran, C., Paquot, N., 2010. Fructose and metabolic diseases: new findings, new questions. Nutrition 26(11—12):1044—1049.

[132] Sievenpiper, J.L., Toronto, D.K.S., Clinical Trials, U., 2012. Fructose: where does the truth lie? Journal of the American College of Nutrition 31(3):149—151.

[133] Chiu, S., Sievenpiper, J.L., de Souza, R.J., Cozma, A.I., Mirrahimi, A., Carleton, A.J., et al., 2014. Effect of fructose on markers of non-alcoholic fatty liver disease (NAFLD): a systematic review and meta-analysis of controlled feeding trials. European Journal of Clinical Nutrition 68(4):416—423.

[134] Hudgins, L.C., Parker, T.S., Levine, D.M., Hellerstein, M.K., 2011. A dual sugar challenge test for lipogenic sensitivity to dietary fructose. Journal of Clinical Endocrinology & Metabolism 96(3):861—868.

[135] Sobrecases, H., Le, K.A., Bortolotti, M., Schneiter, P., Ith, M., Kreis, R., et al., 2010. Effects of short-term overfeeding with fructose, fat and fructose plus fat on plasma and hepatic lipids in healthy men. Diabetes & Metabolism 36(3):244—246.

[136] Merino, B., Fernandez-Diaz, C.M., Cozar-Castellano, I., Perdomo, G., 2019. Intestinal fructose and glucose metabolism in health and disease. Nutrients 12(1).

[137] Softic, S., Gupta, M.K., Wang, G.X., Fujisaka, S., O'Neill, B.T., Rao, T.N., et al., 2017. Divergent effects of glucose and fructose on hepatic lipogenesis and insulin signaling. Journal of Clinical Investigation 127(11):4059—4074.

[138] Hannou, S.A., Haslam, D.E., McKeown, N.M., Herman, M.A., 2018. Fructose metabolism and metabolic disease. Journal of Clinical Investigation 128(2):545—555.

[139] Jensen, T., Abdelmalek, M.F., Sullivan, S., Nadeau, K.J., Green, M., Roncal, C., et al., 2018. Fructose and sugar: a major mediator of non-alcoholic fatty liver disease. Journal of Hepatology 68(5):1063—1075.

[140] Tesz, G.J., Bence, K.K., 2020. Finding the sweet spot: parsing tissue-specific contributions of fructose metabolism. Cell Metabolism 32(1):6—8.

[141] Andres-Hernando, A., Orlicky, D.J., Kuwabara, M., Ishimoto, T., Nakagawa, T., Johnson, R.J., et al., 2020. Deletion of fructokinase in the liver or in the intestine reveals differential effects on sugar-induced metabolic dysfunction. Cell Metabolism 32(1):117—127 e113.

[142] Lim, J.S., Mietus-Snyder, M., Valente, A., Schwarz, J.M., Lustig, R.H., 2010. The role of fructose in the pathogenesis of NAFLD and the metabolic syndrome. Nature Reviews Gastroenterology & Hepatology 7(5):251—264.

[143] Abdelmalek, M.F., Lazo, M., Horska, A., Bonekamp, S., Lipkin, E.W., Balasubramanyam, A., et al., 2012. Higher dietary fructose is associated with impaired hepatic adenosine triphosphate homeostasis in obese individuals with type 2 diabetes. Hepatology 56(3):952—960.

[144] Kim, M.S., Krawczyk, S.A., Doridot, L., Fowler, A.J., Wang, J.X., Trauger, S.A., et al., 2016. ChREBP regulates fructose-induced glucose production independently of insulin signaling. Journal of Clinical Investigation 126(11):4372—4386.

[145] Jang, C., Hui, S., Lu, W., Cowan, A.J., Morscher, R.J., Lee, G., et al., 2018. The small intestine converts dietary fructose into glucose and organic acids. Cell Metabolism 27(2):351—361 e353.

[146] Jang, C., Wada, S., Yang, S., Gosis, B., Zeng, X., Zhang, Z., et al., 2020. The small intestine shields the liver from fructose-induced steatosis. Nature Metabolism 2(7):586—593.

[147] Ishimoto, T., Lanaspa, M.A., Le, M.T., Garcia, G.E., Diggle, C.P., Maclean, P.S., et al., 2012. Opposing effects of fructokinase C and A isoforms on fructose-induced metabolic syndrome in mice. Proceedings of the National Academy of Sciences of the U S A 109(11):4320—4325.

[148] Gonzalez, J.T., Betts, J.A., 2018. Dietary fructose metabolism by splanchnic organs: size matters. Cell Metabolism 27(3):483—485.

[149] Huard, K., Ahn, K., Amor, P., Beebe, D.A., Borzilleri, K.A., Chrunyk, B.A., et al., 2017. Discovery of fragment-derived small molecules for in vivo inhibition of ketohexokinase (KHK). Journal of Medicinal Chemistry 60(18): 7835—7849.

[150] Futatsugi, K., Smith, A.C., Tu, M., Raymer, B., Ahn, K., Coffey, S.B., et al., 2020. Discovery of PF-06835919: a potent inhibitor of ketohexokinase (KHK) for the treatment of metabolic disorders driven by the overconsumption of fructose. Journal of Medicinal Chemistry 63(22):13546—13560.

[151] Haufe, S., Engeli, S., Kast, P., Bohnke, J., Utz, W., Haas, V., et al., 2011. Randomized comparison of reduced fat and reduced carbohydrate hypocaloric diets on intrahepatic fat in overweight and obese human subjects. Hepatology 53(5):1504—1514.

[152] Abdelmalek, M.F., Suzuki, A., Guy, C., Unalp-Arida, A., Colvin, R., Johnson, R.J., et al., 2010. Increased fructose consumption is associated with fibrosis severity in patients with nonalcoholic fatty liver disease. Hepatology 51(6):1961—1971.

[153] de Faria Ghetti, F., Oliveira, D.G., de Oliveira, J.M., de Castro Ferreira, L., Cesar, D.E., Moreira, A.P.B., 2018. Influence of gut microbiota on the development and progression of nonalcoholic steatohepatitis. European Journal of Nutrition 57(3):861—876.

[154] Mehal, W.Z., 2013. The Gordian Knot of dysbiosis, obesity and NAFLD. Nature Reviews Gastroenterology & Hepatology 10(11):637—644.

[155] Hoyles, L., Fernandez-Real, J.M., Federici, M., Serino, M., Abbott, J., Charpentier, J., et al., 2018. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. Nature Medicine 24(7): 1070—1080.

[156] Harte, A.L., da Silva, N.F., Creely, S.J., McGee, K.C., Billyard, T., Youssef-Elabd, E.M., et al., 2010. Elevated endotoxin levels in non-alcoholic fatty liver disease. Journal of Inflammation 7:15.

[157] Albhaisi, S.A.M., Bajaj, J.S., Sanyal, A.J., 2020. Role of gut microbiota in liver disease. American Journal of Physiology - Gastrointestinal and Liver Physiology 318(1):G84—G98.

[158] Lang, S., Schnabl, B., 2020. Microbiota and fatty liver disease-the known, the unknown, and the future. Cell Host & Microbe 28(2):233—244.

[159] Sharpton, S.R., Yong, G.J.M., Terrault, N.A., Lynch, S.V., 2019. Gut microbial metabolism and nonalcoholic fatty liver disease. Hepatology Communications 3(1):29—43.

[160] Henao-Mejia, J., Elinav, E., Jin, C., Hao, L., Mehal, W.Z., Strowig, T., et al., 2012. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. Nature 482(7384):179—185.

[161] Vancamelbeke, M., Vermeire, S., 2017. The intestinal barrier: a fundamental role in health and disease. Expert Review of Gastroenterology & Hepatology 11(9):821—834.

[162] Ferro, D., Baratta, F., Pastori, D., Cocomello, N., Colantoni, A., Angelico, F., et al., 2020. New insights into the pathogenesis of non-alcoholic fatty liver disease: gut-derived lipopolysaccharides and oxidative stress. Nutrients 12(9).

[163] Luther, J., Garber, J.J., Khalili, H., Dave, M., Bale, S.S., Jindal, R., et al., 2015. Hepatic injury in nonalcoholic steatohepatitis contributes to altered intestinal permeability. Cell Molecular Gastroenterology & Hepatology 1(2): 222—232.

[164] Thuy, S., Ladurner, R., Volynets, V., Wagner, S., Strahl, S., Konigsrainer, A., et al., 2008. Nonalcoholic fatty liver disease in humans is associated with increased plasma endotoxin and plasminogen activator inhibitor 1 concentrations and with fructose intake. Journal of Nutrition 138(8):1452—1455.

[165] Spruss, A., Bergheim, I., 2009. Dietary fructose and intestinal barrier: potential risk factor in the pathogenesis of nonalcoholic fatty liver disease. The Journal of Nutritional Biochemistry 20(9):657—662.

[166] Jin, R., Willment, A., Patel, S.S., Sun, X., Song, M., Mannery, Y.O., et al., 2014. Fructose induced endotoxemia in pediatric nonalcoholic Fatty liver disease. International Journal of Hepatology, 560620, 2014.

[167] Kavanagh, K., Wylie, A.T., Tucker, K.L., Hamp, T.J., Gharaibeh, R.Z., Fodor, A.A., et al., 2013. Dietary fructose induces endotoxemia and hepatic injury in calorically controlled primates. American Journal of Clinical Nutrition 98(2):349—357.

[168] Todoric, J., Di Caro, G., Reibe, S., Henstridge, D.C., Green, C.R., Vrbanac, A., et al., 2020. Fructose stimulated de novo lipogenesis is promoted by inflammation. Nature Metabolism 2(10):1034—1045.

[169] Arakawa, K., Ishigami, T., Nakai-Sugiyama, M., Chen, L., Doi, H., Kino, T., et al., 2019. Lubiprostone as a potential therapeutic agent to improve intestinal permeability and prevent the development of atherosclerosis in apolipoprotein E-deficient mice. PLoS One 14(6):e0218096.

[170] Kessoku, T., Imajo, K., Kobayashi, T., Ozaki, A., Iwaki, M., Honda, Y., et al., 2020. Lubiprostone in patients with non-alcoholic fatty liver disease: a randomised, double-blind, placebo-controlled, phase 2a trial. Lancent Gastroenterology & Hepatology 5(11):996—1007.

[171] Chen, F., Esmaili, S., Rogers, G.B., Bugianesi, E., Petta, S., Marchesini, G., et al., 2020. Lean NAFLD: a distinct entity shaped by differential metabolic adaptation. Hepatology 71(4):1213—1227.

[172] Albhaisi, S., Chowdhury, A., Sanyal, A.J., 2019. Non-alcoholic fatty liver disease in lean individuals. JHEP Report 1(4):329—341.

[173] Das, K., Das, K., Mukherjee, P.S., Ghosh, A., Ghosh, S., Mridha, A.R., et al., 2010. Nonobese population in a developing country has a high prevalence of nonalcoholic fatty liver and significant liver disease. Hepatology 51(5):1593—1602.

[174] Younes, R., Bugianesi, E., 2019. NASH in lean individuals. Seminars in Liver Disease 39(1):86—95.

[175] Misra, A., 2015. Ethnic-specific criteria for classification of body mass index: a perspective for asian Indians and American diabetes association position statement. Diabetes Technology & Therapeutics 17(9):667—671.

[176] Hagstrom, H., Nasr, P., Ekstedt, M., Hammar, U., Stal, P., Hultcrantz, R., et al., 2018. Risk for development of severe liver disease in lean patients with nonalcoholic fatty liver disease: a long-term follow-up study. Hepatology Communications 2(1):48—57.

[177] Dela Cruz, A.C., Bugianesi, E., George, J., Day, C., Liaquat, H., Charatcharoenwitthaya, P., et al., 2014. Characteristics and long-term prognosis of lean patients with nonalcoholic fatty liver disease. Gastroenterology 146:S-909.

[178] Sinn, D.H., Kang, D., Cho, S.J., Paik, S.W., Guallar, E., Cho, J., et al., 2019. Lean non-alcoholic fatty liver disease and development of diabetes: a cohort study. European Journal of Endocrinology 181(2):185—192.

[179] Weinberg, E.M., Trinh, H.N., Firpi, R.J., Bhamidimarri, K.R., Klein, S., Durlam, J., et al., 2020. Lean Americans with nonalcoholic fatty liver disease have lower rates of cirrhosis and Co-morbid diseases. Clinical Gastroenterology and Hepatology.

[180] Verdelho Machado, M., Cortez-Pinto, H., 2012. Fatty liver in lean patients: is it a different disease? Annals of Gastroenterology 25(1):1—2.

[181] Romeo, S., Sanyal, A., Valenti, L., 2020. Leveraging human genetics to identify potential new treatments for fatty liver disease. Cell Metabolism 31(1):35—45.

[182] Tavaglione, F., Targher, G., Valenti, L., Romeo, S., 2020. Human and molecular genetics shed lights on fatty liver disease and diabetes conundrum. Endocrinology Diabetes Metabolism 3(4):e00179.

[183] Jonas, W., Schurmann, A., 2020. Genetic and epigenetic factors determining NAFLD risk. Molecular Metabolism, 101111. Epub ahead of print.

[184] Mann, J.P., Pietzner, M., Wittemans, L.B., Rolfe, E.L., Kerrison, N.D., Imamura, F., et al., 2020. Insights into genetic variants associated with NASH-fibrosis from metabolite profiling. Human Molecular Genetics 29(20): 3451—3463.

[185] Romeo, S., Kozlitina, J., Xing, C., Pertsemlidis, A., Cox, D., Pennacchio, L.A., et al., 2008. Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. Nature Genetics 40(12):1461—1465.

[186] Kozlitina, J., Smagris, E., Stender, S., Nordestgaard, B.G., Zhou, H.H., Tybjaerg-Hansen, A., et al., 2014. Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. Nature Genetics 46(4):352—356.

[187] Mancina, R.M., Dongiovanni, P., Petta, S., Pingitore, P., Meroni, M., Rametta, R., et al., 2016. The MBOAT7-TMC4 variant rs641738 increases risk of nonalcoholic fatty liver disease in individuals of European descent. Gastroenterology 150(5):1219—1230 e1216.

[188] Speliotes, E.K., Yerges-Armstrong, L.M., Wu, J., Hernaez, R., Kim, L.J., Palmer, C.D., et al., 2011. Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. PLoS Genetics 7(3):e1001324.

[189] Koo, B.K., Joo, S.K., Kim, D., Bae, J.M., Park, J.H., Kim, J.H., et al., 2018. Additive effects of PNPLA3 and TM6SF2 on the histological severity of nonalcoholic fatty liver disease. Journal of Gastroenterology and Hepatology 33(6):1277—1285.

[190] Kawaguchi, T., Shima, T., Mizuno, M., Mitsumoto, Y., Umemura, A., Kanbara, Y., et al., 2018. Risk estimation model for nonalcoholic fatty liver disease in the Japanese using multiple genetic markers. PLoS One 13(1): e0185490.

[191] Di Costanzo, A., Belardinilli, F., Bailetti, D., Sponziello, M., D'Erasmo, L., Polimeni, L., et al., 2018. Evaluation of polygenic determinants of nonalcoholic fatty liver disease (NAFLD) by a candidate genes resequencing strategy. Scientific Reports 8(1):3702.

[192] Abul-Husn, N.S., Cheng, X., Li, A.H., Xin, Y., Schurmann, C., Stevis, P., et al., 2018. A protein-truncating HSD17B13 variant and protection from chronic liver disease. New England Journal of Medicine 378(12):1096—1106.

[193] Kozlitina, J., Stender, S., Hobbs, H.H., Cohen, J.C., 2018. HSD17B13 and chronic liver disease in blacks and hispanics. New England Journal of Medicine 379(19):1876—1877.

[194] Emdin, C.A., Haas, M.E., Khera, A.V., Aragam, K., Chaffin, M., Klarin, D., et al., 2020. A missense variant in Mitochondrial Amidoxime Reducing Component 1 gene and protection against liver disease. PLoS Genetics 16(4): e1008629.

[195] Klein, J.M., Busch, J.D., Potting, C., Baker, M.J., Langer, T., Schwarz, G., 2012. The mitochondrial amidoxime-reducing component (mARC1) is a novel signal-anchored protein of the outer mitochondrial membrane. Journal of Biological Chemistry 287(51):42795—42803.

[196] Luukkonen, P.K., Juuti, A., Sammalkorpi, H., Penttila, A.K., Oresic, M., Hyotylainen, T., et al., 2020. MARC1 variant rs2642438 increases hepatic phosphatidylcholines and decreases severity of non-alcoholic fatty liver disease in humans. Journal of Hepatology 73(3):725—726.

[197] Esler, W.P., Bence, K.K., 2019. Metabolic targets in nonalcoholic fatty liver disease. Cell Molecular Gastroenterology & Hepatology 8(2):247—267.

[198] Harrison, S.A., Wong, V.W., Okanoue, T., Bzowej, N., Vuppalanchi, R., Younes, Z., et al., 2020. Selonsertib for patients with bridging fibrosis or compensated cirrhosis due to NASH: results from randomized phase III STELLAR trials. Journal of Hepatology 73(1):26—39.

[199] Harrison, S.A., Abdelmalek, M.F., Caldwell, S., Shiffman, M.L., Diehl, A.M., Ghalib, R., et al., 2018. Simtuzumab is ineffective for patients with bridging fibrosis or compensated cirrhosis caused by nonalcoholic steatohepatitis. Gastroenterology 155(4):1140—1153.

[200] Ratziu, V., Bedossa, P., Francque, S.M., Larrey, D., Aithal, G.P., Serfaty, L., et al., 2014. Lack of efficacy of an inhibitor of PDE4 in phase 1 and 2 trials of patients with nonalcoholic steatohepatitis. Clinical Gastroenterology and Hepatology 12(10):1724—1730 e1725.

[201] Frenette, C.T., Morelli, G., Shiffman, M.L., Frederick, R.T., Rubin, R.A., Fallon, M.B., et al., 2019. Emricasan improves liver function in patients with cirrhosis and high model for end-stage liver disease scores compared with placebo. Clinical Gastroenterology and Hepatology 17(4):774—783 e774.

[202] Chalasani, N., Abdelmalek, M.F., Garcia-Tsao, G., Vuppalanchi, R., Alkhouri, N., Rinella, M., et al., 2020. Effects of belapectin, an inhibitor of galectin-3, in patients with nonalcoholic steatohepatitis with cirrhosis and portal hypertension. Gastroenterology 158(5):1334—1345 e1335.

[203] Friedman, S.L., Ratziu, V., Harrison, S.A., Abdelmalek, M.F., Aithal, G.P., Caballeria, J., et al., 2018. A randomized, placebo-controlled trial of cenicriviroc for treatment of nonalcoholic steatohepatitis with fibrosis. Hepatology 67(5):1754—1767.

[204] Day, C.P., James, O.F., 1998. Steatohepatitis: a tale of two "hits"? Gastroenterology 114(4):842—845.

[205] Sanyal, A.J., 2019. Past, present and future perspectives in nonalcoholic fatty liver disease. Nature Reviews Gastroenterology & Hepatology 16(6):377—386.

[206] Eslam, M., Valenti, L., Romeo, S., 2018. Genetics and epigenetics of NAFLD and NASH: clinical impact. Journal of Hepatology 68(2):268—279.

[207] Kazierad, D., Chidsey, K., Somayaji, V., Bergman, A., Birnbaum, M.J., Calle, R., 2020. Inhibition of ketohexokinase in adults with non-alcoholic fatty liver disease reduces liver fat and inflammatory markers: A randomized Phase II trial. Medicine (N Y) in press.