



The genetics of recurrent hydatidiform moles in Mexico: further evidence of a strong founder effect for one mutation in *NLRP7* and its widespread

Mónica Aguinaga¹ · Maryam Rezaei² · Irma Monroy¹ · Nawel Mechtouf² · Javier Pérez¹ · Elsa Moreno³ · Yolotzin Valdespino³ · Carolina Galaz⁴ · Guadalupe Razo¹ · Daniela Medina¹ · Raúl Piña⁵ · Rima Slim⁶

Received: 1 January 2021 / Accepted: 24 February 2021 / Published online: 22 March 2021

© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2021

Abstract

Purpose To investigate the frequency of a founder mutation in *NLRP7*, L750V, in independent cohorts of Mexican patients with recurrent hydatidiform moles (RHMs).

Methods Mutation analysis was performed by Sanger sequencing on DNA from 44 unrelated Mexican patients with RHMs and seven molar tissues from seven additional unrelated patients.

Results L750V was present in homozygous or heterozygous state in 37 (86%) patients and was transmitted on the same haplotype to patients from different states of Mexico. We also identified a second founder mutation, c.2810+2T>G in eight (18.1%) patients, and a novel premature stop-codon mutation W653*.

Conclusion Our data confirm the strong founder effect for L750V, which appears to be the most common mutation in *NLRP7*. We also report on six healthy live births to five patients with biallelic *NLRP7* mutations, two from spontaneous conceptions and four from donated ovum and discuss our recommendations for DNA testing and genetic counseling.

Keywords Recurrent hydatidiform moles · Founder mutation

Introduction

Hydatidiform mole (HM) is an aberrant human pregnancy characterized by abnormal embryonic development and excessive proliferation of the trophoblast. Common HM is sporadic and affects 1 in every 600 pregnancies [1]. At the histopathological level, HM is classified as complete or partial. Complete hydatidiform moles (CHMs) are characterized by the absence of embryo and excessive proliferation of the trophoblast. Partial hydatidiform moles (PHMs) have moderate focal trophoblastic proliferation and may contain embryonic tissues. CHMs are androgenetic while PHMs are triploid dispermic [2]. Recurrent hydatidiform moles (RHMs) are defined by the occurrence of at least two molar pregnancies in the same patient and affect approximately 1–9.4% of women with a prior HM, depending on studies and populations [3–7]. Based on morphological analysis, RHMs may be classified as CHM or PHM.

Biallelic *NLRP7* mutations are the major cause for RHMs (OMIM 231090) [8] and explain the genetic etiology of 55% of patients [9]. A second gene responsible for RHMs, *KHDC3L*, was identified in 2011 [10] and its biallelic

✉ Mónica Aguinaga
aguinagamonica09@gmail.com

¹ Department of Genetics and Human Genomics, Instituto Nacional de Perinatología, Montes Urales 800 Col. Lomas Virreyes, 11000 Mexico City, Mexico

² Department of Human Genetics, McGill University, Montreal, Quebec, Canada

³ Department of Pathology, Instituto Nacional de Perinatología, Mexico City, Mexico

⁴ Académico tiempo completo en la Escuela de Ciencias de la Salud, Universidad del Valle de México, Campus Noroeste, Mexico City, Mexico

⁵ Instituto de Ciencias en Reproducción Humana (Instituto Vida), León, Guanajuato, Mexico

⁶ Departments of Human Genetics and Obstetrics and Gynecology, Research Institute of the McGill University Health Centre, Montreal, Quebec, Canada

mutations explain the etiology of 5% of patients with RHMs (hydatidiform mole, recurrent type 2 (OMIM 611687)) [9, 11]. Molar tissues from patients with mutations in *NLRP7* or *KHDC3L* are diploid biparental. Both genes are components of the subcortical maternal complex, which is essential for epigenetic reprogramming of the oocyte genome and the activation of the embryonic genome [12–14]. Recently, biallelic mutations in three other genes, *MEI1*, *TOP6BL* (C11orf80), and *REC114*, with roles in meiotic double-strand break formation have been identified in patients with recurrent androgenetic complete hydatidiform moles, miscarriages, and infertility [15].

In 2013, our group analyzed *NLRP7* mutations in 20 Mexican patients with RHMs and found that 17 of them have biallelic mutations in *NLRP7* [16] and all the 17 patients had at least one copy of a previously reported mutation, c.2248C>G, p.Leu750Val (L750V) in two Mexican patients [17]. Furthermore, of the 17 patients, 12 were homozygous for L750V. These 12 patients were born in different parts of Mexico and all denied consanguinity between their parents. In addition, the L750V was found in a heterozygous state in 5% of control subjects from the general Mexican population [16]. These data suggested a strong founder effect for L750V in the Mexican population.

Founder mutations in *NLRP7* have been reported in other populations, including the Indian [c.2078G>C, p.(Arg693Pro) and c.2738A>G, p.(Asn913Ser)] and Egyptian [c.-39-387_2129+265dup, p.(Glu710Aspfs*7)] populations [17–19]. However, the founder effect in the Mexican population appeared stronger because the same mutation was found in all the 17 patients with biallelic mutations we reported in Estrada et al. [16]. We therefore set up to analyze another independent cohort of 44 unrelated Mexican patients with RHMs, and seven molar conceptions from unrelated patients with RHMs. Thirty-one of these patients and the seven moles were recruited or retrieved from the Instituto Nacional de Perinatología in Mexico City. We also reviewed the mutation analysis results of another cohort of 13 unrelated patients with RHMs of Mexican origin who were referred either from the USA or Mexico to the Research Institute of the McGill University Health Centre (RI-MUHC) for mutation analysis. Our data confirm our previous findings and highlight the strong founder effect for L750V in Mexico and its inheritance on the same haplotype to patients from various states. Our study also revealed a second founder mutation, c.2810+2T>G and a novel protein-truncating mutation in the Mexican population.

Material and methods

Patients with RHMs

The study was approved by the review boards of the Instituto Nacional de Perinatología (INPer), study number: 212250-

3220-11108-01-14 and McGill University (study number: A01-M07-03A). Patients with at least two HMs were referred from different hospitals in Mexico. A complete clinical evaluation including family and reproductive histories of the patients and their first-degree relatives was taken for all patients. When possible, sisters with RHMs and parents were invited to participate in the study. Written informed consents were obtained from all participants prior to obtaining venous blood samples. A total of 44 unrelated patients were included in this study, 31 were referred to INPer, and 13 were referred to the RI-MUHC. Archived formalin-fixed paraffin embedded (FFPE) molar tissues were retrieved from seven patients with RHMs from the INPer by screening the pathology department record for patients with RHMs.

DNA extraction and mutation analysis

Genomic DNA was isolated from the patient venous peripheral blood. Sequence analysis was performed at the INPer (Mexico) first for exon 6 of *NLRP7* to investigate the presence of the founder mutation L750V. Patients without biallelic mutations were screened for mutations in the other exons, 1 to 5 and 7 to 11, at the RI-MUHC (Montreal, Canada). Primer sequences and polymerase chain reaction (PCR) conditions were as previously described [17, 20] (Supplementary Table 1). PCR products were purified and directly sequenced in forward and reverse orientations using terminator dye in an ABI Prism 3130 (Applied Biosystems). All identified mutations were compared with the reference sequence NM_001127255.1 (<http://fmf.igh.cnrs.fr/ISSAID/infevers/>) and annotated according to the Human Genome Variation Society (HGVS) (<http://varnomen.hgvs.org/>). Sequence variant nomenclature is given according to the following references: NM_001127255.1 (cDNA), NG_008056.1 (genomic DNA), and NP_001120727.1 (protein). Patients who were negative for mutations in *NLRP7* were analyzed for mutations in *KHDC3L* as previously described [21].

Parental contribution to the molar tissues

Sections of FFPE molar tissues were stained with hematoxylin and eosin. Chorionic villi were separated from maternal tissues under a stereomicroscope and used to extract DNA as previously described [9, 22]. Multiplex microsatellite DNA genotyping was performed using the Powerplex 16 HS System (Promega Corporation, Fitchburg, WI, USA), and analyzed as previously described [9, 22].

Results

During the study period, a total of 31 unrelated patients with RHMs were recruited and analyzed for mutations in *NLRP7*

(Table 1). Of these 31 patients, seven had a family history of RHMs and nine (29%) patients had gestational trophoblastic disease after one of their molar pregnancies. Mutation analysis on these 31 patients revealed biallelic *NLRP7* mutations in 26 (83.8%) of them. Of these patients, seventeen were homozygous for L750V; five were compound heterozygous for L750V and c.2810+2T>G, another previously reported mutation in Mexican patients [17]; one patient was compound heterozygous for L750V and a large deletion in the promoter region, c.-6831_-39-1586del, that leads to the absence of transcripts from the allele carrying it [23]; one patient was compound heterozygous for L750V and c.2471+1G>A, p.Leu825* (L825*); one patient was compound heterozygous for L750V and a novel premature stop-codon mutation c.1959G>A, p.Trp653* (W653*); and one patient was homozygous for c.1168del p.Arg390Alafs*26 (R390Afs*26) [9]. Five patients (16.1%) did not have any pathogenic or likely pathogenic variant in *NLRP7* and were screened for *KHDC3L*, but none of them had any mutation.

In the light of the high frequency of L750V in the 31 patients, we screened the record of the Pathology Department of the Instituto Nacional de Perinatología for cases of RHMs since 2003. We found seven archived FFPE molar tissues, from seven additional unrelated patients that were available for analysis. DNA extraction from the chorionic villi of these tissues and their genotyping demonstrated that six are diploid biparental and one is diploid androgenetic monospermic. We next tested the six biparental moles for the presence of the founder L750V mutation. We found that two molar tissues were negative for L750V, three were heterozygous for L750V, and one was homozygous for L750V (Table 1). The latter observation indicates that the father of the HM carries the L750V, known to be present in 5% of control subjects from the general Mexican population [16].

We next reviewed the results of all Mexican patients with RHMs who were referred from various hospitals and medical centers from the USA or Mexico to the RI-MUHC since 2006 for *NLRP7* and *KHDC3L* mutation analyses. We found 13 unrelated patients, of them 12 had biallelic mutations in *NLRP7* (Table 1). Seven were homozygous for L750V; one was compound heterozygous for L750V and another previously reported promoter region deletion, c.-13413_2982-344del [24]; two were compound heterozygous for L750V and c.2810+2T>C [24]; one was compound heterozygous for L750V and c.2471+1G>A, p.L825*; and one was compound heterozygous for p.Tyr872* (Y872*) and c.2810+2T>C.

The states of origin of 44 unrelated patients analyzed on DNA from blood or molar tissues in this study or in Estrada et al. [16] with at least one copy of the L750V were available and are provided on the Mexican map in Fig. 1, which shows an important clustering of these patients in the state of Mexico City where they were recruited and also in some neighboring states. Haplotype analysis of all the SNPs and variants that are

covered by our Sanger sequencing demonstrated the inheritance of the L750V mutation on a shared haplotype between patients from various Mexican states (Table 2), from rs775886 to rs269933 spanning 18,296 bp. We note that the shared haplotype is certainly larger; however, in Table 2, we included only the single nucleotide polymorphisms that are covered by our Sanger sequencing.

Since *NLRP7* is highly rich in Alu repeats and so far, nine of its 80 reported mutations are mediated by Alu recombination (<https://infervers.umai-montpellier.fr/web/>), which can be easily missed when using only Sanger sequencing, we attempted to retrieve archived FFPE tissues from patients with no mutations to re-evaluate the diagnosis of their HMs and determine whether they are diploid biparental. Among the patients who were recruited in Mexico, we were able to retrieve four products of conception (POCs), two from each of patients 29 and 30. Morphological and genotypic evaluation of two POCs from patient 29 demonstrated that one is a triploid dispermic PHM and the other lacked morphological features of molar pregnancies and we revised its diagnosis to miscarriage (Table 1). Multiplex microsatellite genotyping of this miscarriage demonstrated its diploid biparental genome and SNP microarray confirmed the diagnosed and demonstrated the absence of aneuploidy [22]. Therefore, this patient did not have RHMs (Table 1). The two POCs from patient 30 fulfilled the morphological diagnosis of CHM and both were found diploid androgenetic monospermic by multiplex microsatellite genotyping. From a third patient, 31, no tissues could be retrieved, but one of her POCs had been karyotyped and found to be tetraploid 92,XXYY. Of the patients referred to the RI-MUHC, only one patient was negative for *NLRP7* mutations and four of her molar conceptions were available for genotype analysis and were found diploid androgenetic monospermic. This patient was later analyzed by exome sequencing and found to have biallelic mutation in *MEI1* [15]. Therefore, the data on the POCs of these four patients explain the absence of *NLRP7* mutations in them since biallelic *NLRP7* mutations are associated with RHMs that are diploid biparental (Table 1). In conclusion, of the five patients with no *NLRP7* mutations, only four had RHMs, which brings the number of patients with RHMs recruited in Mexico to thirty and the total number of analyzed and reviewed patients in this study to forty-three.

Discussion

Recurrent molar pregnancy is a rare disease. However, in the current study along with that of Estrada et al. [16], we report on a total of 70 unrelated patients with RHMs of Mexican origin (30 recruited in Mexico, 13 referred to the RI-MUHC, 7 molar tissues, and 20 reported in Estrada et al.). To our knowledge, this is the largest series from a single country

Table 1 Recapitulation of data on 71 analyzed patients with RHMs from Mexico

Case N.	Patient ID	Reproductive history (complications)	<i>NLRP7</i> mutations	Complication	References
Mexican patients recruited in Mexico between 2013-2020					
1	ACC	6 PHM	L750V hom		
2	CEA	4 HM	L750V hom		
3	BBL	2 HM, END (preeclampsia), LB	L750V hom	GTD	
4	MMN (consanguinity)	4 HM	L750Vhom		
5	GHR*	3 HM, MC	L750V hom	GTD	
6	VGDE*	2 HM	L750V hom		
	VGLE (sister)	HM	L750V hom		
7	DJEY*	3 HM	L750V hom		
	DJER (sister)	2 HM	L750V hom		
	DJEG (sister)	2 HM	L750V hom		
8	OLO*	2 HM, MC	L750V hom		
9	CLL	2 HM	L750V hom	GTD	
10	RLMC	2 HM	L750V hom		
11	PQRM	3 HM	L750V hom		
12	PAF	4 HM, MC	L750V hom		
13	GGE*	4 HM	L750V hom	GTD	
14	DSL	3 PHM, MC	L750V hom		
15	VOM	3 HM	L750V hom		
16	CRA	HM, 2 MC	L750V hom	GTD	
17	ABH	5 CHM	L750V hom	GTD	
18	GEM	2 PHM	L750V, c.2810+2T>G		
19	MADM	HM, CHM, 2 MC	L750V, c.2810+2T>G		
20	CR	5 HM, LB	L750V, c.2810+2T>G	GTD	
21	PVI	2 HM, MC	L750V, c.2810+2T>G		
22	RJG	2 PHM, MC	L750V, c.2810+2T>G		
23	HME	HM, CHM, 2 PHM	L750V, c.-6831_-39-1586del	GTD	Rezaei et al. [23]
24	RGR*	HM, 2 CHM, MC	L750V, c.2471+1G>A		
25	LCMV	HM, 2 PHM	L750V, W653*		
26	TGR*	2 HM	R390Afs*26 hom		Nguyen et al. [9]
27	GBNA	CHM, PHM	No mutation		
28	VPA	2 HM, LB	No mutation		
29	QVSL	PHM (triploid dispermic), PHM revised to MC	No mutation		
30	MCV	2 CHM (2 androgenetic monospermic), MC	No mutation	GTD	
31	MTMC	2 PHM, 3 MC, MC (92,XXYY)	No mutation		
Screening for L750V in HM tissues from patients with RHMs received between 2003 and 2019					
32	MTO	2 HM (1 diploid biparental)	L750V hom		
33	PFME*	3 HM (1 diploid biparental)	L750V het		
34	PSJ	5 HM (1 diploid biparental), MC	L750V het		
35	MGMJ	3 HM (1 diploid biparental)	L750V het		
36	CPE	MC, 3 HM (1 diploid biparental)	Negative for L750V		
37	DCRN	MC, 3 HM (1 diploid biparental)	Negative for L750V		
38	MAD	2HM (1 androgenetic monospermic)	not screened		
Patients of Mexican origin referred from various clinics and hospitals to the MUHC-RI between 2006 and 2020					
39	655	2 PHM, MC, PHM	L750V hom		Deveault et al. 2009;
	657 (sister)	PHM, CHM, HM	L750V hom		Nguyen et al. [29]

Table 1 (continued)

Case N.	Patient ID	Reproductive history (complications)	<i>NLRP7</i> mutations	Complication	References
30	733	2 HM, MC, 2 HM, IVF-PGT-HM, 5 HM, donated ovum-LB	L750V hom		Nguyen et al. [29]
41	908*	4 HM (with 3 partners)	L750V hom		
42	1220*	3 HM	L750V hom		
	1224 (sister)	3 HM, donated ova-2 LB	L750V hom		
	1227 (sister)	HM	L750V hom		
43	1352	2 PHM, 3 HM	L750V hom		
44	1371*	2 HM, CHM, 2 HM (with 3 partners)	L750V hom		
45	1878	5 HM, BO, donated ovum-LB	L750V hom		
46	1359	4 HM	L750V, c.-13413_2982-344del		Reddy et al. [24]
47	1243	PHM, 8 MC, MC, PHM	L750V, c.2810+2T>G		Reddy et al. [24]
48	1674	2 HM, MC, HM, MC, PHM, MC	L750V, c.2810+2T>G		
49	1888*	MC, 2 HM, MC, HM	L750V, c.2471+1G>A		
	1889 (sister)	2 HM, HAT	L750V, c.2471+1G>A		
50	1074	MC, PHM, HM, 5 MC (2 after clomide), HM, CHM	Y872X, c.2810+2T>G		Nguyen et al. [29]; Reddy et al. [24]
51	1333	4 MC, 4 CHM (4 androgenetic monospermic)	Biallelic <i>MEI1</i> mutations		Nguyen et al. [9]

HM, hydatidiform mole, which is used when the pathology report did not specify the classification; *CHM*, complete hydatidiform mole; *PHM*, partial hydatidiform mole; *MC*, miscarriage; *END*, early neonatal death; *LB*, live birth; *GTD*, gestational trophoblastic disease; *BO*, blighted ovum; *IVF*, *in vitro* fertilization; *PGD*, preimplantation genetic testing; *HAT*, total hysterectomy; *hom*, homozygous; *het*, heterozygous

and suggests a higher frequency of RHMs in Mexico than in other countries. This finding is in line with a previous report describing a higher frequency of RHMs in Mexico as compared to western countries.

Here, we describe the results of mutation analysis on 30 new unrelated patients with RHMs recruited in Mexico, seven molar tissues from seven unrelated patients with RHMs, and review mutation analysis on 13 unrelated patients of Mexican origin referred to the RI-MUHC. Of the 43 analyzed patients, excluding the molar tissues, L750V was present in homozygous or heterozygous state in 37 (86%) of them (Table 3). These data make the L750V the most frequent *NLRP7* mutation reported to date and are in agreement with its presence at a minor allele frequency (MAF) of 0.025 in control subjects from Mexico [16] and 0.00310 in Latino population reported in gnomAD v2.1.1 (135 out of 35,430) (gnomAD (broadinstitute.org)) and Varsome (3 out of 848) (Varsome The Human Genomics Community).

In addition, this study revealed a second founder variant, c.2810+2T>G in the Mexican population that was present in eight unrelated patients (Table 3). This mutation is also reported in databases with a MAF in Latino population of 0.0002892 (10 out of 34,574) in gnomAD v.2.1.1 and 0.0004 in Varsome. Of note, that L750V and c.2810+2T>G both appear to be specific for Mexican/Latino population (Varsome) and have never been

reported in patients with RHMs or healthy subjects from other populations. However, the c.2471+1G>A mutation has been reported in patients of Pakistani, Indian, and Chinese origin, and this study revealed its presence for the first time in two unrelated Mexican patients, which is not unexpected since the Mexican population consists of a mixture of Native American inhabitants (56.4%), European migrants (41.8%), and West Africans (1.8%) [25]. Ruiz-Linares et al. [26] estimated individual ancestry proportions in different countries from Latin America and found that in the Mexican population, Native American ancestry is highest in the center/south of the country where the highest number of patients with L750V was observed. This suggests that L750V may have been inherited from the Native American population that remains to be demonstrated in future studies.

Two patients [3 and 20], the first with a homozygous L750V and the second with L750V and c.2810+2T>G, had each a live birth from a spontaneous conception that led to healthy children. These observations are in agreement with previous ones documenting the occurrence of a total of 13 live births [8, 19, 23, 24, 27, 28], observed mostly in patients with mutations that have mild functional consequences on the protein such as missense, splice, or sometimes protein-truncating mutations at the end of the protein [27]. Among the 13 reported live births, 12 children were reported to be healthy and only

Table 2 Shared haplotype between patients

cDNA	rs number	MAF	Haplotype																						
			908	CRA	VOM	ML	655	CEA	1220	733	BHA	1371	1352	1888	CRL	1674	PVI	1074	MADM	GEM	1359	RGR	BBL	HME	
c-13413_2982-344del	Novel	Novel	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT
c-6831_-39-1586	Novel	Novel	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT	WT
c-39-16C-T	rs775886	0.3906	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c-353-56A-C	rs775884	0.38	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c-390G>A,Q130Q	rs775883	0.345	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c-955G>A,V319I	rs775882	0.245	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.1137G>C,K379N	rs10418277	0.00513	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.1932-146C>T	rs75879	0.7075	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.1959G>A,W653*	Novel	Novel	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2130-41T>G	rs4806626	0.2384	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2248C>G,L750V	rs104895512	0.0004809	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2300-34T>C	rs7359929	0.2268	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2300-57T>C	rs775876	0.7056	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2471T>G>A,L1825*	rs104895505	0.00005169	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2472-67A>G	rs260957	0.2096	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2616C>A,V872*	Novel	Novel	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2822T>C,V884Y	rs260951	0.5843	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2755A>G,A925A	rs260950	0.5845	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2810-21G	rs104895513	0.00004781	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2810-98C>T	rs260949	0.449	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2810-123G>A	rs647845	0.446	G	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2810-126T>C	rs647844	0.5901	T	T	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2810-224G>A	rs260948	0.5577	G	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2811-523C>T	rs775872	0.0559	T	T	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2811-496T>C	rs175092	0.55736	T	T	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2811-402C>T	rs617543	0.55508	C	C	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2811-390A>G	rs25912435	0.55535	A	A	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2811-394G>T	rs2534059	0.55528	G	G	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	T	
c.2811-329A>G	rs260937	0.55674	A	A	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2811-312C>A	rs260936	0.5929	C	C	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2811-228T>C	rs260935	0.55744	T	T	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2811-178G>A	rs12079871	0.06807	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2811-54T>G	rs260934	0.5575	T	T	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2811-25G>C	rs775870	0.04071	C	C	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.2811-23A>G	rs260933	0.5841	A	A	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c-3981-29_32delGTTT	rs104895542	0.5823	WT	WT	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	Del	
c.2981-123T>C	rs260932	0.55579	T	T	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	
c.2981-142C>A	rs260933	0.587446	C	C	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
c.2982-28delG	rs34438464	0.1229	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	
c.*290T>C	rs634742	0.63906	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	C	

MAF minor allele frequency in gnomAD database. In column cDNA, bold character indicates pathogenic variants

one was reported with various morphological abnormalities [28]. Despite these relatively encouraging outcomes, spontaneous live births from such patients are extremely rare and

account for approximately 1.5% of all their conceptions [29]. Because the primary defect in patients with biallelic NLRP7 mutations is in their oocytes, ovum donation has been



Fig. 1 Geographical distribution of patients carrying L750V in Mexican states. The numbers refer to unrelated patients

Table 3 Distribution of mutations in 43 Mexican patients with RHMs

Biallelic <i>NLRP7</i> mutations	Number of patients
L750V homozygous	24 (55.8%)
L750V, c.2810+2T>G	8 (18.6%)
L750V, c.2471+1G>A	2 (4.6%)
L750V, c.-13413_2982-344del	1 (2%)
L750V, c.-6831_-39-1586del	1 (2%)
L750V,W653*	1 (2%)
Number of patients with ≥ 1 L750V	37 (86%)
Y872X, c.2810+2T>G	1 (2%)
R390Afs*26 homozygous	1 (2%)
RHMs and no mutations in <i>NLRP7</i>	4 (9.3%)
Biallelic <i>MEI1</i> mutations	1 (2%)
Number of patients with RHMs	43

proposed to these patients as their best reproductive option. To date, eight such patients, including three reported in this study, patients 733, 1224, and 1878, and another in a patient that we previously reported in Estrada et al. [16], have achieved successful pregnancies from donated ova and conceived ten healthy live births [27, 30, 31].

Based on the above data and the replicated strong founder effect for L750V, if Sanger sequencing were to be used for mutation analysis, we propose to begin the analysis by sequencing exon 6 of *NLRP7*. If the patient is negative for the common mutation, completing the gene sequencing is then recommended. Genetic counseling of patients with biallelic *NLRP7* mutations must consider the age of the patients, the risk of neoplastic degeneration, which occurred in 29% of the 31 patients recruited in Mexico, the scarcity of spontaneous live births in these patients, and the benefit of oocyte donation. Spontaneous live births have been observed in 13 patients; however, we still do not know if these children are at a higher risk for imprinting disorders. It is therefore important to keep in mind that the earliest known defect in patients with biallelic *NLRP7* mutations is the impaired establishment of maternal methylation marks in their oocytes. In addition, biallelic mutations in another member of the subcortical maternal complex, *PADI6*, which have been shown to cause female infertility, early embryonic arrest during preimplantation development [32], and miscarriages and HM [23, 33] were recently documented in patients with Beckwith-Wiedemann and Silver-Russell syndromes [34, 35]. Therefore, a close follow-up of the pregnancies of patients with biallelic *NLRP7* mutations is highly recommended and may help monitoring for imprinting

disorders which may lead to a broad spectrum of clinical manifestations.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10815-021-02132-1>.

Author contribution All authors contributed to the study conception and design. Material preparation was performed by Maryam Rezaei, Irma Monroy, Mechtouf Nawel, Javier Pérez, Elsa Moreno, Yolotzin Valdespino, Carolina Galaz, and Guadalupe Razo. Data collection was performed by Monica Aguinaga, Carolina Galaz, Daniela Medina D, Raúl Piña, and Rima Slim. Analyses were performed by Maryam Rezaei, Irma Monroy, Mechtouf Nawel, Javier Pérez, and Guadalupe Razo. The first draft of the manuscript was written by Mónica Aguinaga and all authors commented on previous versions of the manuscript. Rima Slim revised the work critically for important intellectual content and approved the version to be published. All authors read and approved the final manuscript.

Funding This work was supported by the Canadian Institute of Health Research MOP130364 and by the Instituto Nacional de Perinatología, Mexico City.

Data Availability All data and materials are available upon request.

Code availability Not applicable.

Declarations

Ethics approval The study was approved by the Instituto Nacional de Perinatología (INPer) Review Board, study number: 212250-3220-11108-01-14 and the McGill Institutional Review Board (IRB# A01-M07-03A). This study was performed in line with the principles of the Declaration of Helsinki.

Consent to participate All patients provided written consent to participate in our study.

Conflict of interest The authors declare no competing interests.

References

1. Savage PM, Sita-Lumsden A, Dickson S, Iyer R, Everard J, Coleman R, et al. The relationship of maternal age to molar pregnancy incidence, risks for chemotherapy and subsequent pregnancy outcome. *J Obstet Gynaecol*. 2013;33(4):406–11.
2. Hui P, Buza N, Murphy KM, Ronnett BM. Hydatidiform moles: genetic basis and precision diagnosis. *Annu Rev Pathol*. 2017;12: 449–85.
3. Lurain JR, Sand PK, Carson SA, Brewer JI. Pregnancy outcome subsequent to consecutive hydatidiform moles. *Am J Obstet Gynecol*. 1982;142(8):1060–1.
4. Sebire NJ, Fisher RA, Foskett M, Rees H, Seckl MJ, Newlands ES. Risk of recurrent hydatidiform mole and subsequent pregnancy outcome following complete or partial hydatidiform molar pregnancy. *BJOG*. 2003;110(1):22–6.

5. Kim JH, Park DC, Bae SN, Namkoong SE, Kim SJ. Subsequent reproductive experience after treatment for gestational trophoblastic disease. *Gynecol Oncol.* 1998;71(1):108–12.
6. Yapar EG, Ayhan A, Ergeneli MH. Pregnancy outcome after hydatidiform mole, initial and recurrent. *J Reprod Med.* 1994;39(4):297–9.
7. Eagles N, Sebire NJ, Short D, Savage PM, Seckl MJ, Fisher RA. Risk of recurrent molar pregnancies following complete and partial hydatidiform moles. *Hum Reprod.* 2015;30(9):2055–63.
8. Murdoch S, Djuric U, Mazhar B, Seoud M, Khan R, Kuick R, et al. Mutations in NALP7 cause recurrent hydatidiform moles and reproductive wastage in humans. *Nat Genet.* 2006;38(3):300–2.
9. Nguyen NMP, Khawajkie Y, Mechtouf N, Rezaei M, Breguet M, Kurvinen E, et al. The genetics of recurrent hydatidiform moles: new insights and lessons from a comprehensive analysis of 113 patients. *Mod Pathol.* 2018;31(7):1116–30.
10. Parry DA, Logan CV, Hayward BE, Shires M, Landolsi H, Diggle C, et al. Mutations causing familial biparental hydatidiform mole implicate c6orf221 as a possible regulator of genomic imprinting in the human oocyte. *Am J Hum Genet.* 2011;89(3):451–8.
11. Sebire NJ, Savage PM, Seckl MJ, Fisher RA. Histopathological features of biparental complete hydatidiform moles in women with NLRP7 mutations. *Placenta.* 2013;34(1):50–6.
12. Li L, Baibakov B, Dean J. A subcortical maternal complex essential for preimplantation mouse embryogenesis. *Dev Cell.* 2008;15(3):416–25.
13. Akoury E, Zhang L, Ao A, Slim R. NLRP7 and KHDC3L, the two maternal-effect proteins responsible for recurrent hydatidiform moles, co-localize to the oocyte cytoskeleton. *Hum Reprod.* 2015;30(1):159–69.
14. Demond H, Anvar Z, Jahromi BN, Sparago A, Verma A, Davari M, et al. A KHDC3L mutation resulting in recurrent hydatidiform mole causes genome-wide DNA methylation loss in oocytes and persistent imprinting defects post-fertilisation. *Genome Med.* 2019;11(1):84.
15. Nguyen NMP, Ge ZJ, Reddy R, Fahiminiya S, Sauthier P, Bagga R, et al. Causative mutations and mechanism of androgenetic hydatidiform moles. *Am J Hum Genet.* 2018;103(5):740–51.
16. Estrada H, Buentello B, Zenteno JC, Fiszman R, Aguinaga M. The p.L750V mutation in the NLRP7 gene is frequent in Mexican patients with recurrent molar pregnancies and is not associated with recurrent pregnancy loss. *Prenat Diagn.* 2013;33(3):205–8.
17. Kou YC, Shao L, Peng HH, Rosetta R, del Gaudio D, Wagner AF, et al. A recurrent intragenic genomic duplication, other novel mutations in NLRP7 and imprinting defects in recurrent biparental hydatidiform moles. *Mol Hum Reprod.* 2008;14(1):33–40.
18. Slim R, Bagga R, Chebaro W, Srinivasan R, Agarwal N. A strong founder effect for two NLRP7 mutations in the Indian population: an intriguing observation. *Clin Genet.* 2009;76(3):292–5.
19. Mahadevan S, Wen S, Balasa A, Fruhman G, Mateus J, Wagner A, et al. No evidence for mutations in NLRP7 and KHDC3L in women with androgenetic hydatidiform moles. *Prenat Diagn.* 2013;33(13):1242–7.
20. Qian J, Deveault C, Bagga R, Xie X, Slim R. Women heterozygous for NALP7/NLRP7 mutations are at risk for reproductive wastage: report of two novel mutations. *Hum Mutat.* 2007;28(7):741.
21. Reddy R, Akoury E, Phuong Nguyen NM, Abdul-Rahman OA, Dery C, Gupta N, et al. Report of four new patients with protein-truncating mutations in C6orf221/KHDC3L and colocalization with NLRP7. *Eur J Hum Genet.* 2013;21(9):957–64.
22. Khawajkie Y, Mechtouf N, Nguyen NMP, Rahimi K, Breguet M, Arseneau J, et al. Comprehensive analysis of 204 sporadic hydatidiform moles: revisiting risk factors and their correlations with the molar genotypes. *Mod Pathol.* 2020;33(5):880–92.
23. Rezaei M, Jagadeesh/Beena S, Bereke E, Aguinaga M, Qian J, Hadipour Z, et al. Novel pathogenic variants in PADI6, NLRP5, and NLRP7 in patients with hydatidiform moles and reproductive failure. *Clin Genet.* 2021. <https://doi.org/10.1111/cge.13941>.
24. Reddy R, Nguyen NM, Sarabay G, Rezaei M, Rivas MC, Kavasoglu A, et al. The genomic architecture of NLRP7 is Alu rich and predisposes to disease-associated large deletions. *Eur J Hum Genet.* 2016;24(10):1516.
25. Silva-Zolezzi I, Hidalgo-Miranda A, Estrada-Gil J, Fernandez-Lopez JC, Uribe-Figueroa L, Contreras A, et al. Analysis of genomic diversity in Mexican Mestizo populations to develop genomic medicine in Mexico. *Proc Natl Acad Sci U S A.* 2009;106(21):8611–6.
26. Ruiz-Linares A, Adhikari K, Acuna-Alonzo V, Quinto-Sanchez M, Jaramillo C, Arias W, et al. Admixture in Latin America: geographic structure, phenotypic diversity and self-perception of ancestry based on 7,342 individuals. *PLoS Genet.* 2014;10(9):e1004572.
27. Akoury E, Gupta N, Bagga R, Brown S, Dery C, Kabra M, et al. Live births in women with recurrent hydatidiform mole and two NLRP7 mutations. *Reprod BioMed Online.* 2015;31(1):120–4.
28. Sunde L, Vejerslev LO, Jensen MP, Pedersen S, Hertz JM, Bolund L. Genetic analysis of repeated, biparental, diploid, hydatidiform moles. *Cancer Genet Cytogenet.* 1993;66(1):16–22.
29. Nguyen NM, Slim R. Genetics and Epigenetics of recurrent hydatidiform moles: basic science and genetic counselling. *Curr Obstet Gynecol Rep.* 2014;3:55–64.
30. Fisher RA, Lavery SA, Carby A, Abu-Hayyeh S, Swingler R, Sebire NJ, et al. What a difference an egg makes. *Lancet.* 2011;378(9807):1974.
31. Fallahi J, Razban V, Momtahan M, Akbarzadeh-Jahromi M, Namavar-Jahromi B, Anvar Z, et al. A novel mutation in NLRP7 related to recurrent hydatidiform mole and reproductive failure. *Int J Fertil Steril.* 2019;13(2):135–8.
32. Xu Y, Shi Y, Fu J, Yu M, Feng R, Sang Q, et al. Mutations in PADI6 cause female infertility characterized by early embryonic arrest. *Am J Hum Genet.* 2016;99(3):744–52.
33. Qian J, Nguyen NMP, Rezaei M, Huang B, Tao Y, Zhang X, et al. Biallelic PADI6 variants linking infertility, miscarriages, and hydatidiform moles. *Eur J Hum Genet.* 2018;26(7):1007–13.
34. Cubellis MV, Pignata L, Verma A, Sparago A, Del Prete R, Monticelli M, et al. Loss-of-function maternal-effect mutations of PADI6 are associated with familial and sporadic Beckwith-Wiedemann syndrome with multi-locus imprinting disturbance. *Clin Epigenetics.* 2020;12(1):139.
35. Eggermann T, Kadgien G, Begemann M, Elbracht M. Biallelic PADI6 variants cause multilocus imprinting disturbances and miscarriages in the same family. *Eur J Hum Genet.* 2020.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.