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Identifying compounds that prevent the binding of the SARS-CoV-2 S-protein to ACE2

Claudia Guadalupe Benítez-Cardoza a, José Luis Vique-Sánchez b,c,*

- a Laboratorio de Investigación Bioguímica. ENMvH-Instituto Politécnico Nacional. Ciudad de México. Mexico
- ^b Facultad de Medicina Mexicali, Universidad Autónoma de Baja California, Mexicali, BC, Mexico
- ^c Ciencias de La Salud Mexicali, Universidad Autónoma de Baja California, Mexicali, BC, Mexico

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ABSTRACT

We investigated compounds selected by molecular docking to identify a specific treatment for COVID-19 that decreases the interaction between angiotensin-converting enzyme 2 (ACE2) and the receptor-binding domain (RBD) of SARS-CoV-2. Five compounds that interact with ACE2 amino acids Gln24, Asp30, His34, Tyr41, Gln42, Met82, Lys353, and Arg357 were evaluated using specific binding assays for their effects on the interaction between ACE2 with RBD. The compound labeled ED demonstrated favorable ACE2-binding, with an IC_{50} of 31.95 μ M. ED cytotoxicity, evaluated using PC3 cells in an MTT assay, was consistent with the low theoretical toxicity previously reported. We propose that ED mainly interacts with His34, Glu37, and Lys353 in ACE2 and that it has an inhibitory effect on the interaction of ACE2 with the RBD of the S-protein. We recommend further investigation to develop ED into a potential drug or adjuvant in COVID-19 treatment.

1. Introduction

The COVID-19 pandemic continues to cause morbidity and mortality without specific treatment for the wide range of symptoms [1–3]. Different treatments have been proposed during this pandemic [4,5]. Moreover, new antivirals with different therapeutic targets have been developed, focusing on RNA-dependent RNA polymerase (RdRp), polyproteins (3CLpro and PLpro) [6–8], spike protein (S-protein) [9,10], and the ACE2 region that interacts with RBD [11–15]. Many works have focused on developing new drugs against COVID-19, but no results have provided a therapeutic advantage to date.

ACE2 plays an important role, facilitating the movement of SARS-CoV-2 through the cell membrane. It was recently reported (December 2020) that observed mutations in RBD could increase the infection process and reduce the effect of vaccines [16–19]. Furthermore, mutations in positions 452 and 501 have been reported to increase SARS-CoV-2 infectivity and the interaction of RBD with ACE2 [20,21]. Targeting the ACE2 interaction site allows researchers to choose this region as a therapeutic target to inhibit the interaction between ACE2 and the RBD. Several studies have targeted the interaction regions between the RBD and ACE2, performing docking for drug repositioning or using chemotherapeutic libraries (or using both methods) to search for

drugs or compounds that can inhibit the interaction between RBD with ACE2 [22–25]. In this study, we selected five compounds using molecular docking [11] and tested their interaction with ACE2 to determine their potential to inhibit its binding with RBD of the S-protein.

2. Materials and methods

2.1. Synthetic compounds

Five compounds were tested using ID-Chembridge: 7782787 (EA), 7676800 (EB), 7956590 (EC), 7781334 (ED), and 7783270 (EE) [26] (Table 1). These compounds were previously reported as potential inhibitors of ACE2 and RBD binding [11]. The purity of the compounds reported by the manufacturer was greater than 95 %, which we verified by LC/MS. Stock solutions (1–10 mM) of each compound were prepared in DMSO.

2.2. ACE2/RBD-SARS-CoV-2 inhibitor screening assay

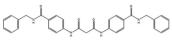
We tested the selected compounds (ID-Chembridge: 7782787, 7676800, 7956590, 7781334, and 7783270) using the COVID-19 Spike-ACE2 Binding Assay Kit (RayBiotech cat: COV-SACE2) according to the

^{*} Corresponding author. Facultad de Medicina Mexicali, Universidad Autónoma de Baja California, BC, Mexico. E-mail address: jvique@uabc.edu.mx (J.L. Vique-Sánchez).

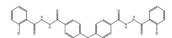
Table 1Study ID, PubChem CID, ID-Chembridge Corp., Chemical name and structure of compounds tested.

EA: 2953970, 7782787, N,N'-[methylenebis (2-hydroxy-4,1-phenylene)]bis [2-(3,4-dimethoxyphenyl)acetamide]

EC, 2973584, 7956590, N,N'-bis{4-[(benzylamino)carbonyl]phenyl}malonamide



EE, 2954039, 7783270, N',N''-[oxybis (4,1-phenylenecarbonyl)]bis (2-chlorobenzohydrazide)



EB, 2947005, 7676800, 2,2'-{1,4-butanediylbis [(4-ethyl-4H-1,2,4-triazole-5,3-diyl) thio]}bis (1-phenylethanone)

ED, 2953854, 7781334, N',N''-[oxybis (4,1-phenylenecarbonyl)]bis (3-methoxybenzohydrazide)

manufacturer's instructions. All compounds were dissolved in DMSO and tested in triplicate at the following concentrations: 100, 50, 30, 15, 5, and 2 μM . Briefly, the ACE2 protein was incubated with 250 μL of each compound in solution for 1 h with shaking. To RBD-coated 96-well plates, we added 100 μL of ACE2-compound solution. The plates were incubated at 2.5 h at room temperature with shaking, and then the plates were washed four times and incubated for 60 min with 100 μL of HRP-conjugated IgG. The plates were washed four times and incubated for 30 min with 100 μL of TMB One-Step Substrate Reagent, then we added 50 μL of stop solution to each well and read the absorbance of all wells at 450 nm immediately in a microplate spectrophotometer (Epoch; BioTek, Winooski, VT, USA). The positive control was ACE2 in DMSO. Decreased absorbance indicates that the ACE2–RBD interaction is inhibited.

2.3. Description of the molecular interaction of ED-compounds with ACE2 by docking

The atomic coordinates of ACE2 were obtained from the Protein Data Bank (PDB: 1R42 and 7DF4). Molecular Operating Environment (MOE) software was used to analyze the docking results [11]. We used the conformers of compounds with an inhibitor effect to describe the potential interaction in ACE2.

2.4. Cytotoxicity assay in PC3 cells cultures determined by MTT

The MTT viability assay was performed as described previously [27]. This assay is based on reducing yellow MTT (3-(4,5-diMethyl-2-Thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide, Sigma-Aldrich, St. Louis, MO, USA) to formazan by mitochondrial succinate dehydrogenase, which is present only in living cells. The absorbance value of formazan is directly proportional to the number of viable cells [28].

PC3 cells in 96-well microplates (8000 cells per well) were treated for 48 h with concentrations of 50–300 μM of the ED compound. DMSO at a final concentration of 0.1 % was used as vehicle control. After treatment, the media was replaced with media containing MTT (0.5 mg/mL), and cells were incubated for an additional 4 h. The medium was removed, and formazan crystals were dissolved in DMSO.

Finally, the plate was read at 570 nm. The assays were performed in triplicate and in three independent studies. The percentage viability was calculated according to the formula: % viability = [mean optical density (O.D.) treated cells \times 100]/(mean O.D. control cells). The concentration corresponding to 50 % inhibition viability (IC $_{50}$) was calculated by non-

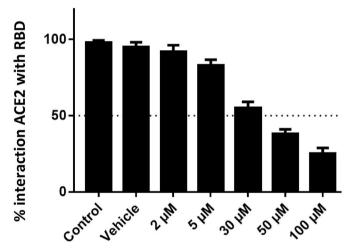


Fig. 1. ACE2-RBD binding assay results, showing the percentage of interaction between ACE2 and RBD when exposed to 2– $100~\mu M$ of the ED compound.

linear regression analysis (percentage of viability vs. log concentration) using GraphPad Prism ver. 7.0 software (GraphPad, San Diego, CA, USA).

2.5. Statistical analysis

Data were expressed as mean \pm standard deviation (SD) of a minimum of three independent experiments. Averages and standard deviations were calculated in Excel (Microsoft), and the IC50 of the ED compound was calculated using GraphPad Prism 7 (GraphPad Software, La Jolla, CA, USA).

3. Results

3.1. Inhibition of ACE2-RBD binding

The ED compound (PubChem CID: 2953854) demonstrated an IC $_{50}$ of 31.95 \pm 3.98 μ M (Fig. 1). The other compounds were not determined some inhibitor effect below 100 μ M.

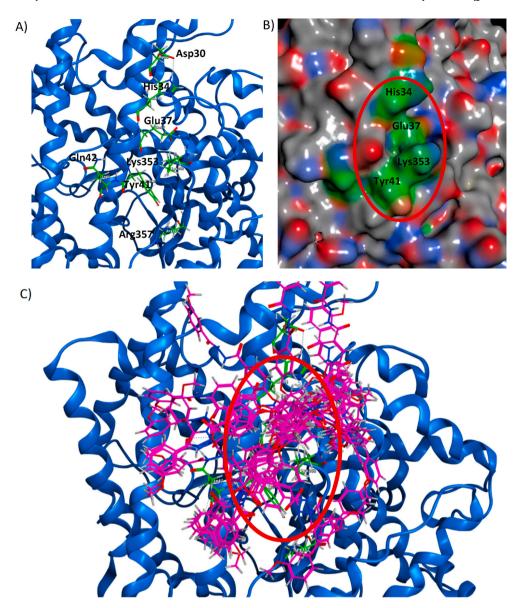


Fig. 2. Potential site in ACE2 (blue) with main amino acids (green). A) The ACE2 Asp30, His34, Tyr41, Gln42, Met82, Lys353, and Arg357 amino acids as the potential docking site; the Lys353 is essential for interaction between ACE2 and RBD of the S-protein, B) Pocket of the potential site (red circle) and C) Pocket of potential site with 18 conformers of ED-compound (pink).

3.2. Description of the molecular interaction of the ED compound with ACE2 by molecular docking

From the docking results [11], we described the probable interactions of the ED compound, and 18 conformers of the ED compound were used. The graphical interaction of each conformer is provided in the Supplementary Material (Table S1). We propose Asp30, His34, Glu37, Asp38, Tyr41, Gln42, Lys68, Asn330, Lys353, Asp355, and Ala386 ACE2 amino acids are the most important in interacting with the ED compound. However, 13 conformers interacted with the hydrogen bridge bond at His34, Glu37, and Lys353 amino acids (Fig. 2, Table S2). The interactions of the ED compound were described previously (shown in Table S3) [11].

3.3. Cytotoxic effect of ED-compound by MTT viability assay

Mitochondrial activity, measured by MTT assay, indicates cell viability. We determined the cytotoxic effect of the ED compound in concentrations between 50 and 300 μ M to determine culture viability.

After 48 h of incubation, the 100 μM solution demonstrated no cytotoxic effect (similar results to the negative control and vehicle/DMSO). At 150 μM , the approximate cytotoxic effect was 12 %, and at a concentration of 300 μM , the cytotoxic effect was 25 % (Fig. 3). These results indicate that the ED compound may be safe in human cells, and other toxicity assays are needed to validate these results.

4. Discussion

Effective treatments against COVID-19 are still under development, and there are multiple potential therapeutic targets against SARS-CoV-2 [5,29–34].

In this study, we tested five compounds reported to inhibit the interaction between ACE2 and RBD in the S-protein [11]. These compounds were selected based on the docking directed to the amino acids in the crystallographic structure of the interaction between ACE2 (Gln24, Asp30, His34, Tyr41, Gln42, Met82, Lys353, and Arg357) and the RBD in the S-protein of SARS-CoV-2 [35–37].

In the inhibition tests, we identified that only the ED compound had

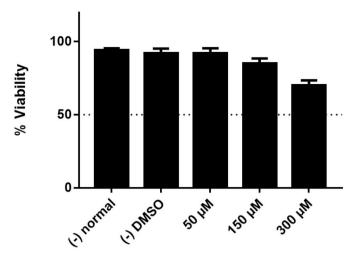


Fig. 3. Percentage cytotoxicity in PC3 cultures after 48 h of incubation with ED compound concentrations between 50 and 300 $\mu M.$

an inhibitory effect up to 100 μM , and we determined an IC_{50} of 31.95 \pm 3.98 μM . This result is notable as compounds or drugs repurposed for use against COVID-19 have demonstrated inhibitory effects of 10–40 % at concentrations between 10 and 30 μM [38]. These results suggest that those drugs participate in specific interactions.

To support the inhibitory effect of the ED compound, it is necessary to analyze the docking results, which showed that the ED compound has 18 conformers, each of which interacts differently with a potential ACE2 site. These results allowed us to identify the main amino acids that are important in the interaction of the 18 conformers of the ED compound with ACE2 (Asp30, His34, Glu37, Asp38, Tyr41, Gln42, Lys68, Asn330, Lys353, Asp355, and Ala386 amino acids, Tables S1 and S2). It is essential that 13 conformers interact with the hydrogen bridge bond with His 34, Glu37, and Lys353 in ACE2 (Fig. 2 and Table S2). We highlight the interactions generated close to His34 and Lys353, as these amino acids interact with Tyr453 and Asn501, respectively, in the RBD in the S-protein [35,39]. Currently, some variants of SARS-CoV-2 [21] have different consequences in the infection process. The ED compound may hinder the interaction with the region between Tyr453 and Asn501 in the S-protein, in which it is reported that the mutations in the positions 452 and 501 increase the Infectivity and the interaction between

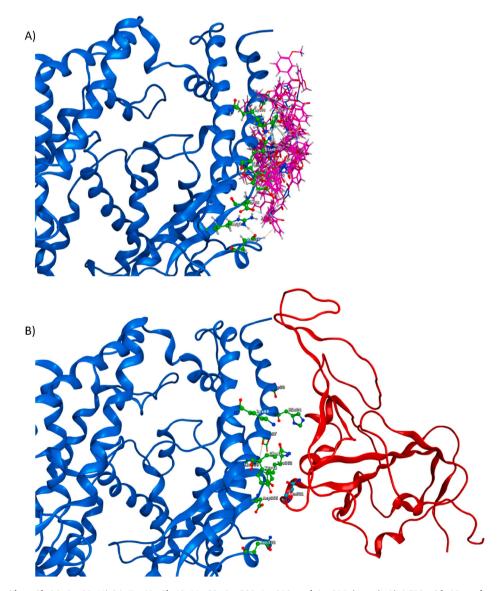


Fig. 4. ACE2 (blue) residues Gln24, Asp30, His34, Tyr41, Gln42, Met82, Asp330, Lys353, and Asp355 (green). A) ACE2 with 18 conformers of the ED compound (pink), B) ACE2 interacting with RBD of the S-protein (red) and the amino acid in position 501 (Asn, Cyan).

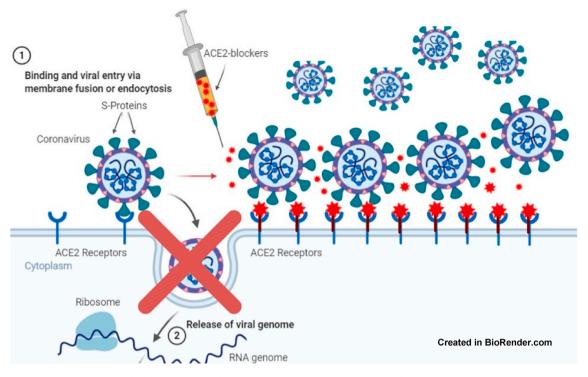


Fig. 5. Blocking effect of a selective compound in ACE2, preventing cell entry of SARS-CoV-2.

the RBD with ACE2 and decrease the effectiveness of vaccines [16–18, 20,21]. Therefore, our results indicate that this interaction site in ACE2 is significant as a therapeutic target to inhibit the interaction between ACE2 and the RBD (Fig. 4).

The cytotoxicity results for the ED compound in PC3 cells are favorable, with no effect at a concentration of $100~\mu M.$ These results are consistent with previously reported theoretical toxicity data, so the ED compound has a high probability of being safe in humans, and the development of this drug against COVID-19 may continue.

The ED compound was chosen by a docking directed to the specific region in ACE2 that is important in interacting with RBD, so at the molecular level, ED is highly specific in ACE2, providing a specific drug that prevents SARS-CoV-2 from interacting with human cells. This drug would not be antiviral since it would interact with ACE2 and might generate an inhibitory effect on the interaction with the S-protein of SARS-CoV-2 (Fig. 4). In addition, the ED compound might be used preventively, prophylactically, and during the COVID-19 infection process (Fig. 5).

The ED compound does not have any specific use or patent registered (PubChem CID: 2953854 [40]), and it is available for *in vitro* and other ACE2/RBD interaction assays. We propose that these results will contribute to developing a new drug that helps combat this pandemic.

5. Conclusions

We propose that the ED compound will provide the basis for developing a new drug that interacts with the angiotensin-converting enzyme 2 (ACE2). Drugs based on ED could hinder or prevent the interaction of ACE2 with SARS-CoV-2. The ED compound has a good preliminary IC $_{50}$ that significantly reduces the interaction of ACE2 with RBD and thus could hinder the COVID-19 infection process in humans. Cytotoxicity testing of the ED compound revealed it would likely be safe in humans. Therefore, it is necessary to continue the development of this drug against COVID-19.

Declaration of competing interest

The authors declare that they have no conflict of interest.

Acknowledgments

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.compbiomed.2021.104719.

Ethical approval

Not required.

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