

## Separation of Thylakoid Protein Complexes with Two-dimensional Native-PAGE

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**[Abstract]** The hierarchical composition and interactions of the labile thylakoid protein complexes can be assessed by sequential 2D-native gel-electrophoresis system. Mild non-ionic detergent digitonin is used to solubilize labile protein super- and megacomplexes, which are then separated with first-dimension blue native polyacrylamide gel electrophoresis (1D-BN-PAGE). The digitonin derived protein complexes are further solubilized with stronger detergent,  $\beta$ -DM, and subsequently separated on an orthogonal 2D-BN-PAGE to release smaller protein subcomplexes from the higher-order supercomplexes. Here we describe a detailed method for 2D-BN-PAGE analysis of thylakoid protein complexes from *Arabidopsis thaliana*.

**Keywords:** Native gel electrophoresis, Thylakoid membrane, Thylakoid protein complexes, 2D-BN-PAGE, Light harvesting complex, Photosystem, Photosynthesis

**[Background]** Photosynthetic light reactions take place in the thylakoid membrane, which in higher plants is composed of appressed grana thylakoids and non-appressed stroma thylakoids. The light reactions are catalyzed by multi-subunit protein complexes photosystem (PS) I and II, cytochrome *b<sub>6</sub>f* and ATPase. PSII together with its light harvesting antenna complex (LHCII) is most abundant in grana-thylakoids and therefore spatially segregated from stroma thylakoid -located PSI-LHCI complexes (Andersson and Anderson, 1980). The interphase between the grana and stroma thylakoid is enriched in both photosystems (Albertsson, 2001; Suorsa *et al.*, 2015). Mediated by light dependent reversible phosphorylation of LHCII and PSII proteins, the photosystems together with LHCII assemble into larger super- and megacomplexes. PSII core dimer together with two strongly and two moderately bound LHCII-trimers form large C2S2M2 supercomplexes, PSI together with loosely bound LHCII form PSI-LHCII supercomplexes, and finally, PSII and PSI together with L-LHCII form large PSII-LHCII-PSI megacomplexes (Caffarri *et al.*, 2009; Pesaresi *et al.*, 2009; Rantala *et al.*, 2017).

Non-ionic detergents are generally used for isolation of native protein complexes from biological membranes. Mild detergent digitonin maintains weak interactions between protein complexes, but due to its bulky structure, it selectively solubilizes only the non-appressed regions of the thylakoids. However, when supplemented with low ionic strength salt, aminocaproic acid (ACA), digitonin gets access to the partition gap between two grana appressions and solubilizes the entire thylakoid membrane allowing the analysis of the overall organization of labile thylakoid protein complexes (Rantala *et al.*, 2017). Solubilized protein complexes are supplemented with anionic Coomassie G-250 (CBB) dye that binds to the hydrophobic domains of protein complexes providing them with negative charge, and allows their

electrophoretic separation according to molecular mass. Since the negative surface charges repel each other, CBB also prevents random protein complex aggregation.

Blue native PAGE (BN-PAGE) enables membrane protein complex separation in their native and functionally active form (Schägger and von Jagow, 1991). Coupling the 1D-BN-PAGE with a second (2D)-BN-PAGE allows the analysis of the subcomplex composition of the digitonin derived large protein super- and megacomplexes: The 1D-BN-gel lane containing the separated protein complexes is treated with slightly stronger detergent, *n*-dodecyl- $\beta$ -D-maltoside ( $\beta$ -DM), which more effectively interferes with the interactions between protein complexes, particularly destroying the interaction of LHCII with the two photosystems. The lane is then subjected to the 2D-BN-PAGE for the separation of the dissociated subcomplexes. The composition of the subcomplexes can be further analyzed by electroblotting the 2D-gel and detecting specific proteins with antibodies or by cutting the lanes and subjecting the subcomplexes to denaturing 3D-SDS-PAGE.

This protocol describes optimized thylakoid protein complex isolation method and the analysis of the subcomplex composition of large protein super- and megacomplexes of *Arabidopsis thaliana* by 2D-BN-BN-PAGE. The method can be used for the analysis of the organization of the photosynthetic protein complexes and for the analysis of subcomplex composition of higher-order protein super- and megacomplexes.

## **Materials and Reagents**

### A. Consumables

1. Eppendorf microcentrifuge tubes 1.5 ml (Eppendorf, catalog number: 0030121694)
2. Falcon, Conical Centrifuge Tubes 15 ml (Corning, catalog number: 352096)
3. Culture tubes 5 ml (Lab Depot, catalog number: TLDT8301)
4. Finntip Pipette Tips (Finntip Flex 10, 250, 1,000 and 5 ml)
5. Whatman chromatographic paper (GE Healthcare, catalog number: 1001-931)
6. PVDF membrane (Merck, catalog number: IPVH304F0)

### B. Plant material

Isolated thylakoids from 5-week old *Arabidopsis thaliana* (isolate thylakoids in the presence of 10 mM NaF in all buffers as described in Järvi *et al.*, 2011)

### C. Reagents

1. 6-aminocaproic acid (Sigma-Aldrich, catalog number: A2504)
2. Bis-Tris (Sigma-Aldrich, catalog number: B4429)
3. Glycerol (Avantor Performance Materials, J.T. Baker, catalog number: 7044)
4. Pefabloc SC (4-(2-Aminoethyl) benzenesulfonyl fluoride hydrochloride) (Roche Diagnostics, catalog number: 11585916001)
5. Sodium Fluoride (NaF) (Avantor Performance Materials, J.T. Baker, catalog number: 3688)

6. EDTA disodium salt (Avantor Performance Materials, J.T. Baker, catalog number: 1073)
7. Digitonin (Merck, Calbiochem, catalog number: 300410)
8. *n*-dodecyl- $\beta$ -D-maltoside (Sigma-Aldrich, catalog number: D4641)
9. Serva Coomassie Blue G (SERVA Electrophoresis, catalog number: 35050)
10. Sucrose (Sigma-Aldrich, catalog number: S0389)
11. Tricine (Sigma-Aldrich, catalog number: T0377)
12. TEMED (Tetramethylethylenediamine) (Bio-Rad Laboratories, catalog number: 1610801)
13. Liquid nitrogen
14. Methanol (VWR)
15. Bovine Serum Albumin (BSA) (Sigma-Aldrich, catalog number: A7030)
16. Acrylamide (AA) (Sigma-Aldrich, catalog number: A9099)
17. (N,N'-Methylene)-Bis-Acrylamide (Bis-AA) (Merck, Omnipur, catalog number: 2610)
18. APS (Ammonium persulfate) (Bio-Rad Laboratories, catalog number: 1610700)
19. Tris (Sigma-Aldrich, catalog number: T1503)
20. Glycine (Fisher Scientific, catalog number: 10070150)
21. SDS (VWR, catalog number: 442444H)

#### D. Antibodies

1. Lhcb1 (Agrisera, catalog number: AS01 004)
2. Phospho (P)-Lhcb1 (Agrisera, catalog number: AS13 2704)
3. Lhcb2 (Agrisera, catalog number: AS01 003)
4. Phospho (P)-Lhcb2 (Agrisera, catalog number: AS13 2705)

#### E. Stock solutions

1. Acrylamide solution A: 48% (w/v), 1.5% (w/v) Bis-acrylamide in MQ-water, store at 4 °C
2. Acrylamide solution B: 20% (w/v), 5% (w/v) Bis-acrylamide in MQ-water, store at 4 °C
3. APS (Ammonium persulfate): 5% (w/v) solution in MQ-water, store at 4 °C
4. Digitonin stock solution: 10% (w/v) in MQ-water, store at -20 °C
5.  $\beta$ -dodecyl maltoside: 10% (w/v) in MQ-water, store at -20 °C
6. Glycerol: 75% (w/v) solution in MQ-water, store at 4 °C
7. Pefabloc: 10 mg/ml (w/v) in MQ-water, store at -20 °C
8. Sucrose: 75% (w/v) in MQ-water, store at -20 °C

#### F. Buffers

1. 3x Gel Buffer, store at 4 °C (see Recipes)
2. 25BTH20G resuspension buffer, prepare fresh (see Recipes)
3. ACA buffer (see Recipes)
4. Detergent buffer A (4% Digitonin), prepare fresh (see Recipes)
5. Detergent buffer B (1%  $\beta$ -DM), prepare fresh (see Recipes)

6. CBB buffer, -20 °C (see Recipes)
7. Anode buffer for BN, store at 4 °C (see Recipes)
8. Blue cathode buffer for BN, store at 4 °C (see Recipes)
9. Clear cathode buffer for BN, store at 4 °C (see Recipes)
10. Transfer buffer, store at 4 °C (see Recipes)
11. BN-PA: 3.5-12.5% separation gel, 3% stacking gel (see Recipes)

## **Equipment**

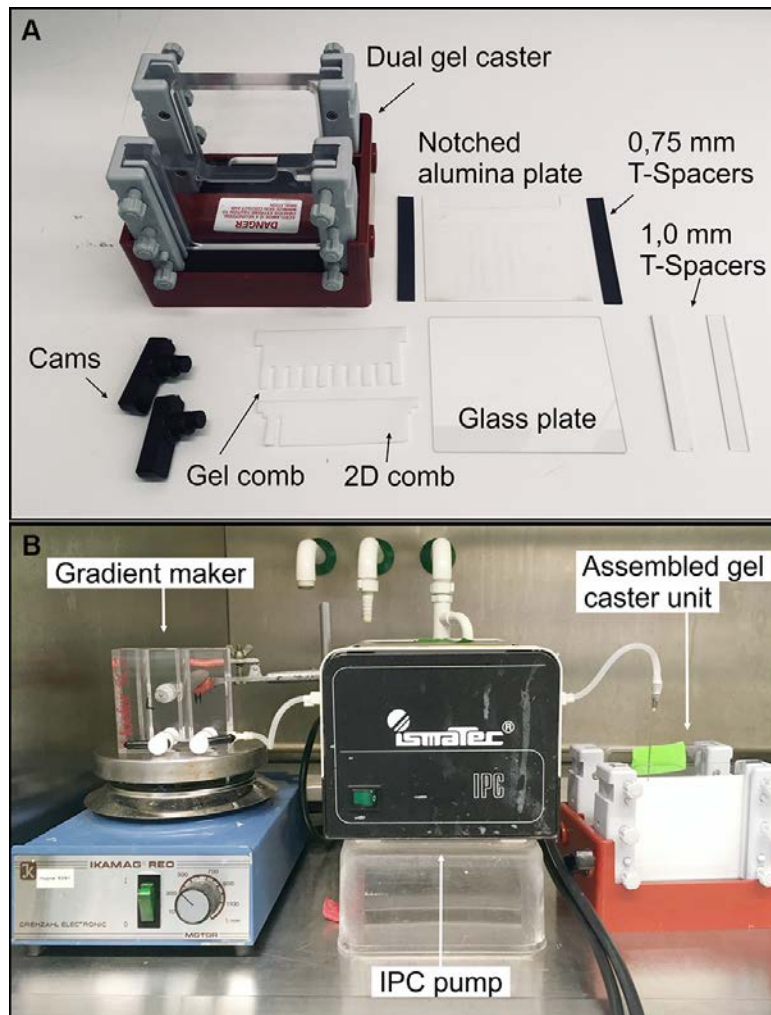
1. Dual gel caster with 10 x 8 cm plates (Hoefer, catalog number: SE215)
2. Hoefer gradient maker SG5 (or any gradient maker containing two 5 ml chambers)
3. 0.75 mm T-spacers (Hoefer, catalog number: SE2119T-2-.75)
4. 1 mm T-spacers (Hoefer, catalog number: SE2119T-2-1.0)
5. Sample gel comb, 0.75 mm, 10 wells (Hoefer, catalog number: SE211A-10-.75)
6. 2D comb (flat) with a reference well, 1.0 mm thick
7. Mighty Small SE250 vertical electrophoresis system (Hoefer, catalog number: SE250)
8. Ismatec IPC-pump
9. Power supply, PowerPac HV (Bio-Rad Laboratories, catalog number: 164-5056)
10. Centrifuge (Eppendorf, model: 5424R)
11. Finnpiquette™ F2 Variable Volume Single-Channel Pipettes
12. Cold room (4 °C)
13. Freezer (-20 °C)
14. Photo scanner (e.g., Perfection V300 Photo, Epson, model: V300)
15. Rocker-Shaker (Biosan, model: MR-12, catalog number: BS-010130-AAI)
16. Semi-dry blotting system (Hoefer, catalog number: TE77X)

## **Procedure**

- A. Casting the BN-PA gel (linear gradient 3.5%-12.5% separation gel; 3% stacking gel)
  1. Assemble gel caster (e.g., Hoefer) with 10 x 8 cm plates (Figure 1A) according to manufacturer's instructions. Any vertical electrophoresis system with any compatible casting unit can be used.
  2. Prepare the light (3.5% AA) and heavy (12.5% AA) buffers from the acrylamide stock solution A (48% acrylamide, 1.5% Bis-acrylamide) (see Recipes). Keep the solutions on ice to prevent untimely polymerization. Cast the gel between a glass plate and a notched aluminum oxide plate (10 x 8 cm) with a gradient maker (Figure 1B). Use 0.75 mm T-spacer for the 1D-BN-PAGE and 1 mm T-spacer for 2D-BN-PAGE. Allow the gradient gel to polymerize for 1-2 h at RT.
  3. Cast the stacking gel (3% acrylamide) using the acrylamide stock solution B (20% acrylamide, 5% Bis-acrylamide). Use an appropriate sample gel comb (1D-BN-PA gel: standard comb with

10 wells, 2D-BN-PA gel: 2D-comb). Allow the stacking gel to polymerize for 30-60 min. Remove the sample gel comb under MQ-water. Store the gel at 4 °C.

*PAUSE POINT: The gel can be stored at 4 °C at least for few days. The gel must not dry.*



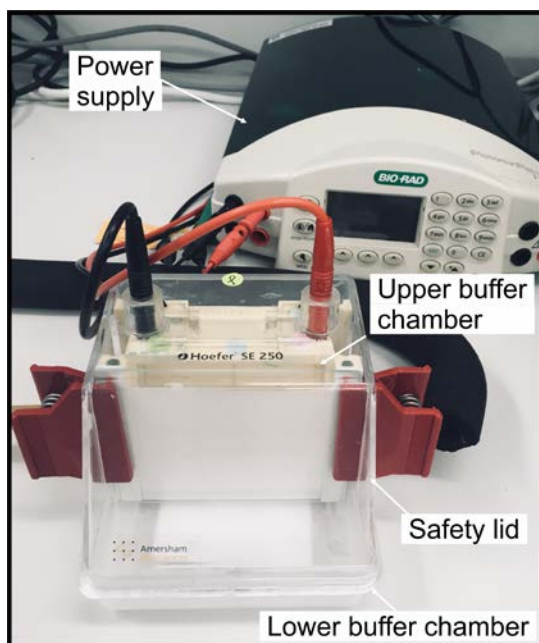
**Figure 1. Equipment for casting gradient gels.** A. Hoefer dual gel caster with all the required equipment; B. Gradient maker, IPC-pump and gel assembled and ready for gel casting.

**B. Solubilization of thylakoid membranes (Perform the solubilization under dim light)**

1. Resuspend thylakoids equivalent to 3-7  $\mu\text{g}$  chlorophyll in ACA buffer to a chlorophyll concentration of 1 mg/ml.
2. Add an equal volume of detergent buffer A (4% digitonin/ACA) on the sample and solubilize at RT with continuous gentle mixing for 10 min, e.g., in a shaker. The final chlorophyll concentration is 0.5 mg/ml.
3. Pellet the insolubilized material by centrifugation at 18,000  $\times g$ , at 4 °C, for 20 min, remove the supernatant to a new Eppendorf tube.
4. Add 1/10 of CBB-buffer to the sample and carefully mix with a pipet. Avoid making air bubbles.

C. Separation of the protein complexes with 1D-BN-PAGE

1. Assemble the gel to the vertical electrophoresis system and pour ~70 ml of blue cathode buffer to the gel tank (upper chamber), and ~100 ml of anode buffer to the lower chamber (Figure 2).



**Figure 2. Example of the gradient gel assembly.** The gel is assembled in the SE 250 Mini-Vertical electrophoresis unit.

2. Load thylakoids equivalent to 2-5  $\mu\text{g}$  of chlorophyll to the sample well.
3. Start the electrophoretic run by gradually increasing the voltage as follows:
  - a. 75 V 30 min
  - b. 100 V 30 min
  - c. 125 V 30 min
  - d. 150 V 60 min
  - e. 175 V 30 min

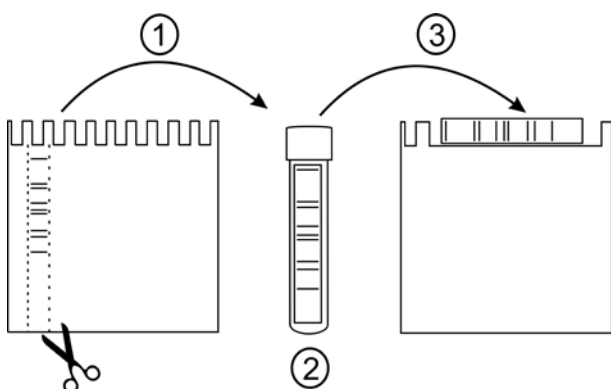
Perform the electrophoresis at 4 °C

4. As the blue running front has moved about one third (at ~100-125 V) of the desired running distance, remove the blue cathode buffer from the upper chamber and replace it with a clear cathode buffer.
5. Stop the electrophoresis, when the protein complexes (B1-B9) (see Figure 6A) are separated and the gel is clear from the blue cathode.
6. Scan the gel with a photo scanner (keep the gel between the glass and alumina plates while scanning), remove the glass plate and carefully cut out the lane containing the separated protein complexes using a spacer (see sketch in Figure 3). Do not use scalpel or any other sharp object for cutting to avoid scratching the alumina plate.

**PAUSE POINT:** It is possible to store the BN-gel strip at  $-80\text{ }^{\circ}\text{C}$ , but do not shock-freeze the strip with liquid nitrogen.

Notes:

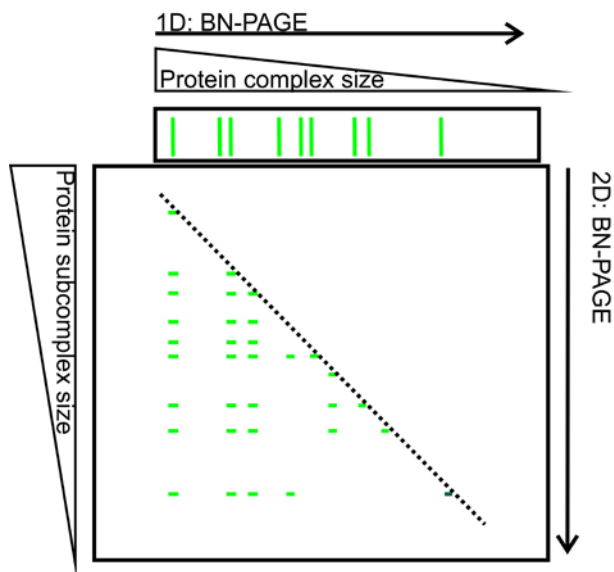
- a. The amount of chlorophyll loaded on the gel depends on the subsequent analysis: for immunoblotting, load  $2\text{ }\mu\text{g}$  (or less) of chlorophyll and for the 3D-SDS-PAGE, load  $5\text{ }\mu\text{g}$  or more chlorophyll.
- b. It is possible to also excise individual bands (complexes B1-B9) from the BN-PA-gel, re-solubilize the bands in Eppendorf-tubes with  $1\%\text{ }\beta\text{-DM}$  and subject them to 2D-BN-PAGE.



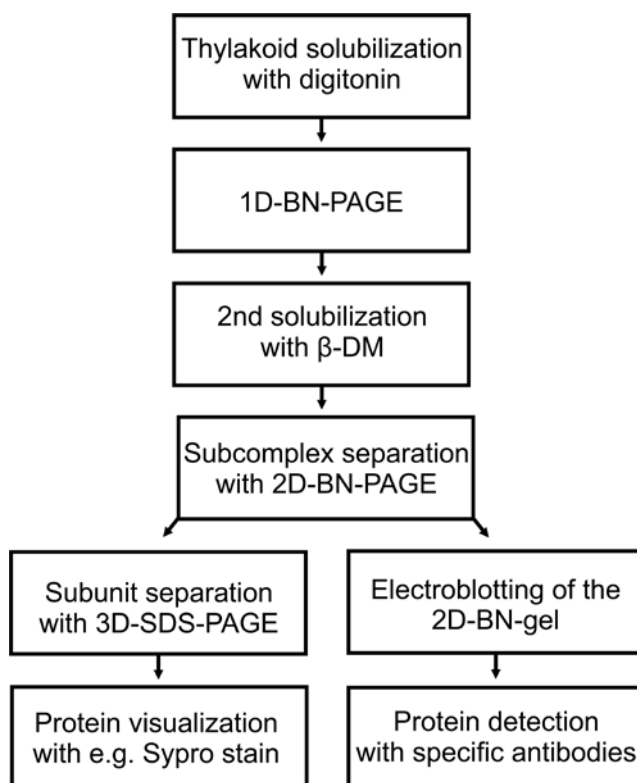
**Figure 3. Preparation of 1D-BN-strip for 2D-BN-PAGE.** (1) After finishing 1D-BN-PAGE, remove the glass plate and excise the lane containing the protein complexes (B1-B9) using e.g., the T-spacer. (2) Carefully place the gel strip to a 5 ml culture tube containing 2 ml of detergent buffer B ( $1\%\text{ }\beta\text{-DM}$ ). Solubilize in a shaker for 40 min. (3) Place the gel strip on the second BN-PA gel (the large well) and assemble the gel to the electrophoresis tank. Perform the 2D-electrophoresis as described for 1D-BN-PAGE.

#### D. 2D-BN-PAGE

1. Place the gel strip in a culture tube (5 ml) and add 2 ml of detergent buffer B ( $1\%\text{ }\beta\text{-DM}$ ).
2. Solubilize the strip for 40 min at  $4\text{ }^{\circ}\text{C}$  in gentle rocking to allow even solubilization of the strip (e.g., 20 rpm).
3. Place the strip on the second BN-PA gel (see sketch in Figure 3) and assemble the gel to the electrophoresis tank.
4. Perform the electrophoresis as described for the 1D-BN-PAGE.
5. Scan the gel after the electrophoresis with a photo scanner (see sketch of the 2D-BN-PAGE in Figure 4, the flowchart demonstrating the follow-up paths in Figure 5 and scanned image of the representative gel in Figure 6B).



**Figure 4. 2D-BN-PAGE separation of thylakoid protein complexes.** Protein complexes B1-B9 (green bands on the upper horizontal gel slice) were first separated according to their mass and shape on 1D-BN-PAGE. After separation the gel slice was subjected to 2D-BN-PAGE during which the complexes (B1-B9) are fractionated into subcomplexes (narrow green bands on the 2D-BN-gel). The complexes on the diagonal (dashed line) represent complexes that have preserved their mass, whereas the complexes below the diagonal are subcomplexes (of B1-B9) that have been disconnected during second solubilization and 2D-BN-PAGE.



**Figure 5. Flowchart of the experimental procedure**



#### E. Western blotting of 2D-native gels

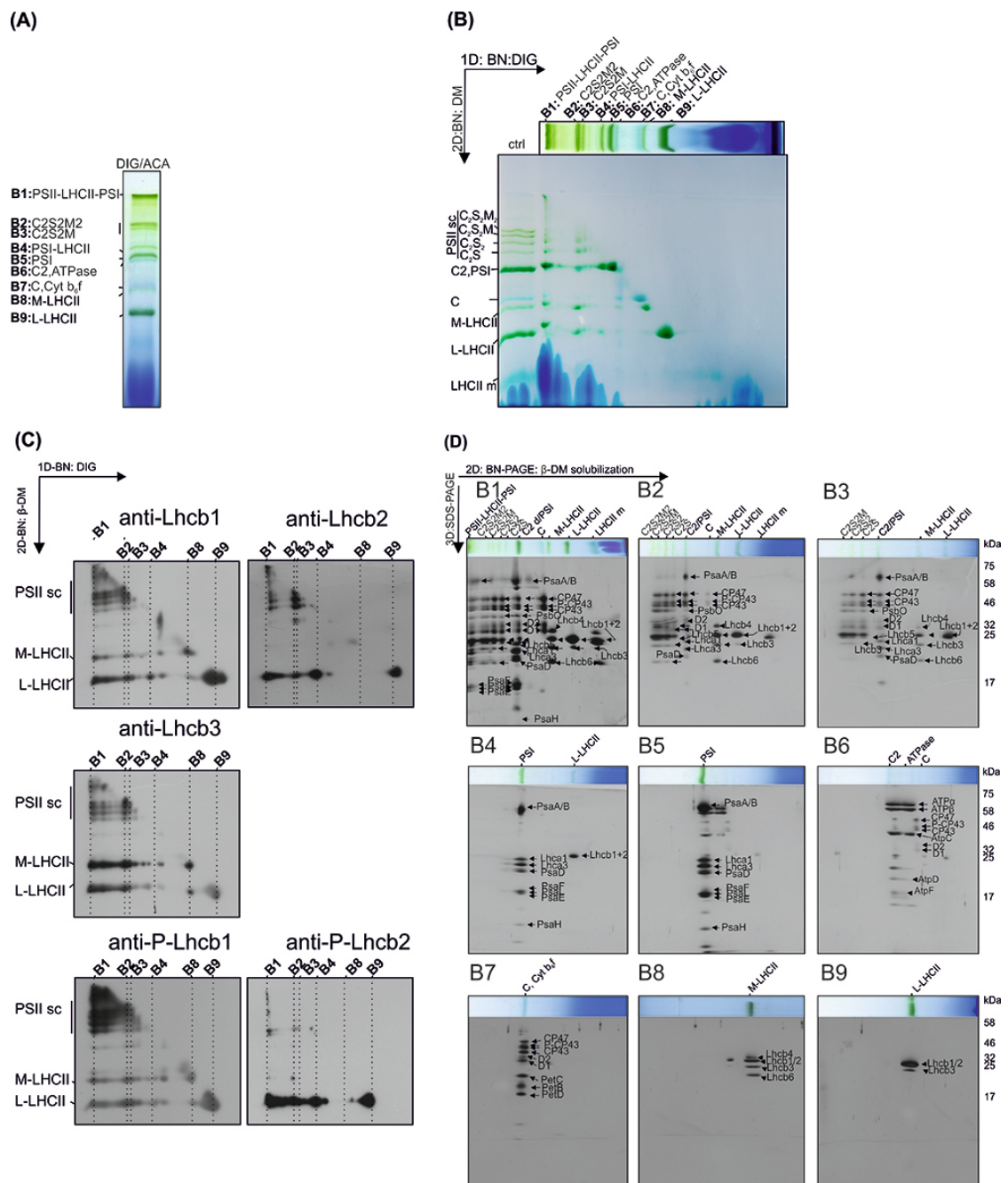
1. Place the 2D-BN-PA-gel to transfer buffer and incubate for 30 min.
2. Activate the PVDF membrane with 100% methanol and place the membrane to transfer buffer. Soak six Whatman filter papers in the transfer buffer.
3. Assemble three papers on the electrode (anode). Place the membrane on top of the papers and the gel on top of the membrane. Finally, place the three papers on top of the gel and mount the cathode on top. Set the current for 1 mA/cm<sup>2</sup> gel area, and transfer for at least 1 h.
4. Destain the membrane with 100% methanol.
5. Block the membrane with 5% BSA for 1 h.
6. Perform immunostaining with specific antibodies (see representative blots from Figure 6C).
  - a. Lhcb1, Lhcb2, Lhcb3
  - b. P-Lhcb1 and P-Lhcb2

#### F. 3D-SDS-PAGE

1. Cut the lanes containing the subcomplexes (corresponding to complexes B1-B9).
2. Protein solubilization and separation on SDS-PAGE are as described for the 2D-SDS-PAGE analysis in Järvi *et al.*, 2011.
3. After electrophoresis, visualize the protein with Sypro Ruby Stain or with silver staining according to Blum *et al.*, 1987 (see representative 3D gels from Figure 6D, the nine gels correspond to complexes B1-B9).

#### **Data analysis**

1. For all the experiments (2D-BN-PAGE, 3D-SDS-PAGE, Western blotting) we use three biological replicates.
2. No image processing was performed. The identification of the protein complexes (on native gels) and individual proteins (on 3D-SDS gels) is based on mass spectrometry analysis according to (Aro *et al.*, 2005; Suorsa *et al.*, 2015).
3. The representative data (Figure 6) is from the original research article (Rantala *et al.*, 2017).



**Figure 6. The representative results from the analysis of the subcomplex and subunit composition of thylakoid protein complexes.** A. Protein complexes (B1-B9) of digitonin solubilized thylakoids after 1D-BN-PAGE separation. B. 2D-BN-PAGE separation of the subcomplexes obtained by re-solubilizing the 1D-strip with β-DM. Ctrl represents thylakoids directly solubilized with 1% DM. C. The Lhcb-protein localization in protein subcomplexes was analyzed by electroblotting the 2D-BN-gels and immunodecorating the blots with Lhcb1, Lhcb2, Lhcb3 protein specific antibodies and with Phospho-Lhcb1-2 protein specific antibodies. D. The protein composition of the subcomplexes (derived from complexes B1-B9) was analyzed by cutting the lanes from 2D-BN-gel and subjecting the gel strips to 3D-BN-PAGE. The proteins

are visualized with Sypro Ruby stain. This figure has been originally published in Rantala *et al.* (2017).

### **Notes**

1. The protocol is highly reproducible in our hands.
2. Digitonin must be purified according to the manufacturer's instructions. We had no problems with digitonin, but the detergent stock may precipitate easily when melted.
3. The thylakoid isolation (protocol in Järvi *et al.*, 2011) must be done from fresh leaves to obtain high-quality thylakoid protein complexes.
4. Freezing and thawing the thylakoid sample several times will affect the stability of the protein complexes.

### **Recipes**

*Note: All the solutions are prepared in MQ water.*

1. 3x Gel buffer
  - 1.5 M 6-Aminocaproic acid
  - 150 mM Bis-Tris
2. 25BTH20G
  - 25 mM Bis-Tris/HCl (pH 7.0)
  - 20% (w/v) glycerol
  - 0.25 mg/ml Pefabloc (add freshly)
  - 10 mM NaF (add freshly)
3. ACA buffer
  - 25 mM Bis-Tris/HCl (pH 7.0)
  - 375 mM Aminocaproic acid
  - 1 mM EDTA
  - 0.25 mg/ml Pefabloc (add freshly from the stock solution)
  - 10 mM NaF (add freshly)
4. Detergent buffer A
  - 4 % digitonin (w/v)
  - 25 mM Bis-Tris/HCl (pH 7.0)
  - 375 mM Aminocaproic acid
  - 1 mM EDTA
  - 0.25 mg/ml Pefabloc (add freshly from the stock solution)
  - 10 mM NaF (add freshly)
5. Detergent buffer B
  - 1%  $\beta$ -dodecyl maltoside (w/v)

- 25 mM Bis-Tris/HCl (pH 7.0)  
 20% (w/v) glycerol and  
 0.25 mg/ml Pefabloc (add freshly from the stock solution)  
 10 mM NaF (add freshly)
6. CBB buffer  
 100 mM Bis-Tris/HCl (pH 7.0)  
 0.5 M ACA  
 30% (w/v) sucrose  
 50 mg/ml Serva Blue G
7. Anode buffer  
 50 mM Bis-Tris/HCl (pH 7.0)
8. Cathode buffer  
 50 mM Tricine  
 15 mM Bis-Tris/HCl (pH 7.0)  
 0.01% Serva Blue G
9. Transfer buffer  
 39 mM glycine  
 48 mM Tris  
 0.0375% SDS  
 20% methanol
10. BN-PA: 3.5-12.5% separation gel, 3% stacking gel

	Light	Heavy	Stack
48% AA, 1.5% bis-AA	148 µl	530 µl	-
20% AA, 5% bis-AA	-	-	180 µl
3x gel buffer	700 µl	700 µl	500 µl
75% glycerol	140 µl	560 µl	-
H <sub>2</sub> O	1,092 µl	290 µl	800 µl
5% APS	15 µl	11 µl	30 µl
TEMED	3 µl	2 µl	3 µl
Pipet to the chamber	1,830 µl	1,880 µl	

Notes:

- a. 10% aqueous APS must be made up fresh or stored frozen.
- b. The recipe is suitable for 0.75 mm gel, multiply the recipe by 1.5 for 1 mm gel.
- c. Prepare the solutions on ice to prevent untimely polymerization.

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## References

1. Albertsson, P. (2001). [A quantitative model of the domain structure of the photosynthetic membrane](#). *Trends Plant Sci* 6: 349-358.
2. Andersson, B. and Anderson, J. M. (1980). [Lateral heterogeneity in the distribution of chlorophyll-protein complexes of the thylakoid membranes of spinach chloroplasts](#). *Biochim Biophys Acta* 593: 427-440.
3. Aro, E. M., Suorsa, M., Rokka, A., Allahverdiyeva, Y., Paakkarinen, V., Saleem, A., Battchikova, N. and Rintamäki, E. (2005). [Dynamics of photosystem II: a proteomic approach to thylakoid protein complexes](#). *J Exp Bot* 56(411): 347-356.
4. Blum, H., Beier, H. and Gross, H. J. (1987). [Improved silver staining of plant proteins, RNA and DNA in polyacrylamide gels](#). *Electrophoresis* 8: 93-99.
5. Caffarri, S., Kouřil, R., Kereiče, S., Boekema, E. J. and Croce, R. (2009). [Functional architecture of higher plant photosystem II supercomplexes](#). *EMBO J* 28(19): 3052-3063.
6. Järvi, S., Suorsa, M., Paakkarinen, V. and Aro, E. M. (2011). [Optimized native gel systems for separation of thylakoid protein complexes: novel super- and mega-complexes](#). *Biochem J* 439: 207-214.
7. Pesaresi, P., Hertle, A., Pribil, M., Kleine, T., Wagner, R., Strissel, H., Ihnatowicz, A., Bonardi, V., Scharfenberg, M., Schneider, A., Pfannschmidt, T. and Leister, D. (2009). [Arabidopsis STN7 kinase provides a link between short- and long-term photosynthetic acclimation](#). *Plant Cell* 21(8): 2402-2423.
8. Rantala, M., Tikkanen, M. and Aro, E. M. (2017). [Proteomic characterization of hierarchical megacomplex formation in Arabidopsis thylakoid membrane](#). *Plant J* 92: 951-962.
9. Schägger, H. and von Jagow, G. (1991). [Blue native electrophoresis for isolation of membrane protein complexes in enzymatically active form](#). *Anal Biochem* 199: 223-231.
10. Suorsa, M., Rantala, M., Mamedov, F., Lespinasse, M., Trotta, A., Grieco, M., Vuorio, E., Tikkanen, M., Jarvi, S. and Aro, E. M. (2015). [Light acclimation involves dynamic re-organization of the pigment-protein megacomplexes in non-appressed thylakoid domains](#). *Plant J* 84: 360-373.